CONTRAST-ENHANCED ULTRASONOGRAPHY OF THE SMALL BOWEL IN HEALTHY CATS

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We characterized the pattern of ultrasonographic contrast enhancement of the small intestinal wall using a commercial contrast medium (Sonove[®]) in 10 healthy awake cats. Subjectively, a rapid intense enhancement of the serosal and submucosal layers was followed by gradual enhancement of the entire wall section during the early phase. At peak enhancement, there was a subjective loss of demarcation between intestinal wall layers. In the late phase, there was a gradual wash out of signal from the intestinal wall. Submucosal wash out occurred last. Time-intensity curves were generated for selected regions in the intestinal wall and multiple perfusion parameters were calculated for each cat. Perfusion parameters included arrival time (7.64 \pm 2.23 s), baseline intensity (1.04 \pm 0.04 a.u.), time to peak from injection (10.74 \pm 2.08 s), time to peak from initial rise (3.1 \pm 1.15), peak intensity (8.92 \pm 3.72 a.u.), wash-in rate (2.06 \pm 0.70 a.u./s) and wash-out rate (-1.07 \pm 0.91 a.u./s). The perfusion pattern of normal feline small bowel may be useful for characterizing feline gastrointestinal disorders that involve the intestinal wall. © 2011 Veterinary Radiology & Ultrasound, Vol. 52, No. 5, 2011, pp 555–559.

Key words: cat, contrast-enhanced ultrasonography, small intestine.

Introduction

CONTRAST-ENHANCED ULTRASONOGRAPHY has been used to evaluate perfusion of normal lymph nodes, liver, spleen, and kidney in the dog¹⁻⁵ and the normal pancreas and kidney in the cat.⁶⁻⁸ This technique has also been used for the characterization of focal splenic,^{9,10} hepatic^{11–13} and renal lesions,¹⁴ and for the diagnosis of congenital portosystemic shunts in small animals.¹⁵

In humans, contrast-enhanced ultrasonography was used to evaluate vascularity of the gastrointestinal wall in neoplastic^{16–18} and inflammatory diseases.^{19–23} Increased bowel vessel density, detected using a second generation ultrasound contrast medium, was associated with disease activity in patients with Crohn's disease.^{21–23} In particular, different patterns of wall enhancement have been described in Crohn's disease patients on the basis of specific mural layer enhancement.²⁴ On the contrary, bowel ischemia is characterized by a decrease or lack of enhancement.²⁵

In veterinary medicine, diagnostic ultrasonography has been used widely for the evaluation of the gastrointestinal wall both in normal and pathological conditions.²⁶ To our knowledge, there are only two reports concerning contrastenhanced ultrasonography of the small intestine in the cat⁸ and dog.²⁷ The first describes the quantitative contrastenhanced ultrasonographic analysis of perfusion in abdominal organs of healthy cats, including small intestine⁸ while the other describes intraoperative contrast-enhanced ultrasonography of normal canine jejunum.²⁷

The first aim of this study was to assess the feasibility of contrast-enhanced ultrasonography in the feline small bowel. The second was to describe the pattern of sonographic contrast enhancement.

Materials and Methods

Ten healthy domestic short haired cats, volunteered by their owner, were studied. There were five males, four females and one neutered female. The mean age was 5.2 years (standard deviation $[SD] \pm 2$ years) and mean bodyweight was 4 kg ($SD \pm 0.7 \text{ kg}$). The cats were healthy on the basis of physical findings and routine laboratory data. The cats were negative for feline leukemia virus antigen, feline immunodeficiency virus antibody, and feline coronavirus antibody. The cats were fasted overnight, for at least 12 h, before imaging. All procedures were conducted by the same sonographer (A.D.), using a real-time ultrasound machine.* Hair over the abdomen was clipped, the skin surface was cleaned with 70% isopropyl alcohol, and coupling gel was applied. The cats were awake and restrained manually during the examination.

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^{*}iU22 ultrasound system, Philips Healthcare, Monza, Italy.

The abdomen was scanned by B-mode ultrasonography using a broadband curved array transducer (5-8 MHz). The gastrointestinal tract was also scanned with a broadband linear array transducer (3-9 MHz). With each cat in right recumbency, one superficial jejunal segment in the left middle abdominal region was selected for color Doppler evaluation and contrast-enhanced ultrasonography. The segment was scanned in a transverse section and the broadband linear array transducer was not moved subsequently. Color Doppler ultrasonography of the intestinal wall was performed using a low wall filter and 300 Hz pulse repetition frequency. Color gain was adjusted dynamically to maximize visualization of the blood vessels while avoiding artifactual color noise. Contrast-specific software (Pulse Inversion Harmonic and Power Modulation combined-PMPI) with a low mechanical index set (0.07) was activated. The gain setting was regulated to obtain an anechoic bowel wall, which was as complete as possible except for the hyperechoic serosal layer and the central hyperechoic line arising from the bowel lumen. Image acquisition was for 180s from the time of onset of contrast medium administration. The contrast medium (Sonovue"^{*}^{*}) was administrated manually through an indwelling cephalic venous 22 G catheter as a rapid bolus dose of 0.5 ml followed immediately by a rapid bolus of 4 ml saline The images were recorded as cine-segments in DICOM format and transferred to a personal computer.

Show Case software‡ was used to view the images and to export selected frames for analysis. Color Doppler images of the intestinal wall were evaluated subjectively for the distribution of mural vessels based on color Doppler signals within the intestinal wall.

The distribution of the contrast medium enhancement within the intestinal wall was evaluated subjectively as satisfactory or unsatisfactory based on the degree of jejunal mural enhancement and homogeneity of mural enhancement at peak intensity (PI). The pattern of contrast medium enhancement was also described.

To assess the degree of the enhancement of the intestinal wall, the E/W ratio was used,²⁴ where E is the major thickness of the enhanced layer and W is the width between the edge of the outer wall and the superficial mucosal interface measured in millimeters on gray-scale images (Fig. 1). The mean value of three measurements, obtained at three different points of the same ultrasonographic image, was used for analysis. The contrast-enhanced ultrasonography findings were evaluated during all three phases of wall perfusion (i.e. early, peak enhancement, and late phase).

A commercial software program§ was used for quantitative computerized analysis of the contrast medium blood pool phase. A region of interest (ROI), drawn to cover the wider portion of the intestinal section, was placed manually in the intestinal wall between the serosal layer and the mucosal interface. The ROI was maintained in the same position by the motion compensation tool of the QLAB software. This tool prevents the displacement of the ROI during respiratory motion. Furthermore, the ROI was adjusted manually on those frames affected severely by respiratory motion.

Artifactual data from adjacent tissue that moved into the ROI during respiratory motion or gas bubbles were removed manually from the final data set to reduce the influence of noise. The raw data obtained from each cat were plotted in time-intensity diagrams.

The following perfusion variables were recorded: arrival time (AT) defined as the time when contrast signal is increasing to greater than double the baseline value in the time-intensity curve, time to peak from injection (TTP_{inj}), time to peak from initial rise (TTP_{inr}), baseline intensity, PI, wash-in rate (W_i) and wash-out rate (W_o) (Fig. 2).

 $W_{\rm i}$ and $W_{\rm o}$ were calculated as the maximal change rate, using data points 20% above baseline and 20% below peak to exclude variability at the toe and shoulder of the time-intensity curve. The data were then regressed for significance of linearity.¶ Significance for analysis was set at P < 0.05. Data calculated for each variable, using a commercial software program, || include means and SD.

Results

There were no Doppler signals within the intestinal wall in any cat. Low Doppler signals were detected in the mesentery adjacent to the intestinal segment in five cats.

Contrast enhancement was judged subjectively to be satisfactory in nine cats. In one cat, inadequate concentration of contrast medium at the level of the intestinal wall was caused by incorrect positioning of the intravenous catheter. During contrast-enhanced ultrasonography, jejunal arteries were identified clearly and there was an initial rapid enhancement of serosal and submucosal layers, followed by a more gradual enhancement of the entire wall section (Fig. 3A). At peak enhancement, there was a subjective lack of demarcation between wall layers (Fig. 3B). In the late phase, there was a gradual wash out of signal from the intestinal wall. The wash out of the submucosal layer occurred last (Fig. 3C). This perfusion pattern of the intestinal wall was seen consistently in all cats. The mean E/W ratio was 0.92 (SD \pm 0.09).

Quantitative computerized analysis of the intestinal wall enhancement was performed in nine cats (Table 1). Data of

^{*}Bracco[®] diagnostic, Milano, Italy.

[‡]Trillium Technology, Ann Arbor, MI.

[§]QLAB quantification software, Philips Healthcare.

[¶]Prism 5[®], GraphPad Software Inc., San Diego, CA.

Microsoft Excel, Microsoft Corporation, Redmond, WA.



FIG. 1. An example of E/W ratio. Contrast enhancement image (A) and gray scale image (B) at peak enhancement. E is the major thickness of the enhanced layer under contrast-enhanced ultrasonography and W is the width between the edge of the outer wall and the superficial mucosal interface. The dotted lines represent the superficial mucosal interface.

 $W_{\rm i}$ and $W_{\rm o}$ were characterized by a significant linear relationship (P < 0.01). The correlation coefficient (r^2) of the regression lines ranged from 0.77 to 0.98.

Discussion

The microvascular architecture of the small intestinal wall is composed of two different parallel capillary beds: the mucous–submucous plexus and the muscular–serous plexus.^{28–30} Under resting conditions, approximately 80% of the blood flow in the feline small intestinal wall is distributed to the mucosa–submucosa and 20% to the muscularis–serosa.^{30–32} In particular, all of the main and large anastomotic vessels (30–80 µm in diameter) are located in the submucosal layer, whereas a plexus of smaller vessels is embedded in the *muscularis mucosae*.³⁰ The mucosal



FIG. 2. Signal intensity in arbitrary units (a.u.) as a function of time in seconds (s) in a normal feline jejunal segment. Contrast enhancement increases until reaching peak intensity. AT, arrival time; BI, baseline intensity; TTP_{inj} , time to peak from injection; PI, peak intensity; TTP_{inr} , time to peak from initial rise; W_i , wash-in rate; W_o , wash-out rate.

arteries divide in a stellate manner into branches which are ramified in the mucosal layer. Each villus is usually supplied by a single arterial vessel that runs in the central villous core. Close to the villous tip, the central vessel arborizes into a dense capillary network. The capillaries collect into veins at the villous base.^{31–33}

We were unable to detect blood flow in intramural vessels in any cat using Doppler ultrasonography and only a few Doppler signals were found in the adjacent mesentery. In human medicine, color Doppler ultrasonography has been used to evaluate the vascularity of the intestinal wall in patients with Crohn's disease.^{34–36} Affected intestinal segments have an increase in the number and caliber of intramural vessels.^{34–36} Increased blood flow in the main splanchnic vessels has also been found in inflammatory bowel disease in both humans^{34–36} and dogs.^{37,38} Diameter and slow blood flow are limiting factors regarding detection of blood vessels in the normal feline bowel wall.³⁹

The contrast-enhanced ultrasonography pattern, which was consistent across all cats, was associated strongly with the anatomic distribution of intramural vessels.^{30,32} The intense and rapid enhancement of the submucosal layer depends on the presence of a rich plexus, draining 80% of the total intramural blood flow.^{30,32,33} The slower wash out of the submucosal layer may be explained by the more tortuous capillary network in this layer that reduces blood flow through the submucosal vessels.

The mean E/W ratio was 0.92 indicating near complete enhancement of the entire intestinal wall from the mucosa to the serosa. Due to the small thickness of the normal feline small intestinal wall (2.41 ± 0.14 mm), a single ROI that included all mural layers was drawn. The results of the quantitative variables represent a mean value obtained from the entire intestinal wall rather than the different vascular distribution in each layer.



FIG. 3. A representative contrast-enhanced ultrasound sequence (A, B, and C) of a normal jejunal segment in a transverse plane after injection of Sonovue[®]. Each image illustrates contrast enhancement on the left and the gray scale image on the right. (A) The contrast medium is present predominantly in the serosal and submucosal layers. (B) Homogeneous enhancement of the entire intestinal wall at peak enhancement. (C) A gradual decrease of enhancement of the intestinal wall. Persistent enhancement is seen in the submucosal layer.

TABLE 1. Results (Mean \pm SD) of Quantitative Contrast Enhanced Ultrasonography of Small Bowel in Nine Healthy Cats

Parameters	Mean \pm SD
AT (s)	7.64 ± 2.23
BI (a.u.)	1.04 ± 0.04
TTP _{ini} (s)	10.74 ± 2.08
PI (a.u.)	8.92 ± 3.72
TTP _{inr} (s)	3.1 ± 1.15
W_i (a.u./s)	2.06 ± 0.70
$W_{\rm o}$ (a.u./s)	-1.07 ± 0.91

AT, arrival time; BI, baseline intensity; TTP_{inj}, time to peak from injection; PI, peak intensity; TTP_{inr}, time to peak from initial rise; W_i , wash in rate; W_o , wash out rate; a.u., arbitrary units.

Good quality time-intensity curves were obtained for nine cats. The upslope and downslope part of the curve was linear. A second small peak of enhancement due to reperfusion was observed later. These values were excluded from the measurement of W_{o} to examine only the first transit of the through the intestinal wall. Some parameters in our cats i.e. AT, TTP_{ini}, and TTP_{inr}, were slightly higher than others reported recently in healthy anesthetized cats.⁸ Sedation and anesthesia were avoided purposely in our cats to obtain baseline values without iatrogenic changes in blood pressure and heart rate.40-42 In feline spleen, anesthesia resulted in a longer AT compared with awake cats.⁴⁰ On the contrary, anesthesia resulted in a shorter time to peak enhancement of normal liver and kidney in the dog.^{2,8} The impact of anesthesia on organ perfusion depends on the characteristics of anesthetic agent

and on the different vascular structures and blood flow of each organ.⁸ Several other factors, such as technical scanning variables, contrast medium characteristics, injection technique, and patient-related factors can influence quantitative variables.^{2,5,8,43–46}

Peak intensity values were lower in comparison to those reported by others.⁸ The use of different contrast media and different imaging system implies a different number of circulating microbubbles and, as a consequence, a different backscatter response from the tissue that influences the degree of contrast enhancement.^{2,27,44,46} Differences in intensity-measuring units and scale among different modalities (dB vs. video-intensity units) can also complicate comparisons.^{2,5} Furthermore, mechanical index influences the amount of microbubble destruction and also affects the amplitude of harmonic signals.^{5,44,46} Because of these reasons, quantitative values obtained from different protocols are not entirely comparable.^{2,58,43-46}

Fasting for at least 12 h before contrast-enhanced ultrasonography of the bowel is suggested to minimize the presence of gas and food particles²⁶ that create strong interference during the activation of the contrast-specific software. Furthermore, fasting is recommended to reduce the peristaltic activity²⁶ to avoid motion artefact that limits quantitative analysis.

In conclusion, we demonstrated that contrast-enhanced ultrasonography can be used for evaluation of intramural blood vessels of the feline small intestine. The perfusion pattern is distinctive. Contrast-enhanced ultrasonography may enable the characterization of bowel wall perfusion in cats affected by various gastrointestinal disorders.

REFERENCES

1. Gaschen L, Angelette N, Stout R. Contrast-enhanced harmonic ultrasonography of medial iliac lymph nodes in healthy dogs. Vet Radiol Ultrasound 2010;51:634–637.

2. Nyman HT, Kristensen AT, Kjelgaard-Hansen M, McEvoy FJ. Contrast-enhanced ultrasonography in normal canine liver. Evaluation of imaging and safety parameters. Vet Radiol Ultrasound 2005;46:243–250.

3. Ohlerth S, Rüefli E, Poirier V, et al. Contrast harmonic imaging of the normal canine spleen. Vet Radiol Ultrasound 2007;48:451–456.

4. Rossi F, Rabbia S, Vignoli M, et al. B-Mode and contrast-enhanced sonographic assessment of accessory spleen in the dog. Vet Radiol Ultrasound 2010;51:173–177.

5. Waller KR, O'Brein RT, Zagzebski JA. Quantitative contrast ultrasound analysis of renal perfusion in normal dogs. Vet Radiol Ultrasound 2007;48:373–377.

6. Kinns J, Aroson L, Hauptman J, Seiler G. Contrast enhanced ultrasound of the feline kidney. Vet Radiol Ultrasound 2010;51:168–172.

7. Rademacher N, Ohlerth S, Scharf G, et al. Contrast-enhanced power and color doppler ultrasonography of the pancreas in healthy and diseased cats. J Vet Intern Med 2008;22:1310–1316.

8. Leinonen MR, Raekallio MR, Vainio OM, Ruohoniemi MO, Biller DS, O'Brien RT. Quantitative contrast-enhanced ultrasonographic analysis of perfusion in the kidneys, liver, pancreas, small intestine, and mesenteric lymph nodes in healthy cats. Am J Vet Res 2010;71:1305–1311.

9. Rossi F, Leone VF, Vignoli M, et al. Use of contrast-enhanced ultrasound for characterization of focal splenic lesions. Vet Radiol Ultrasound 2008;49:154–164.

10. Ohlerth S, Dennler M, Rüefli E, et al. Contrast harmonic imaging characterization of canine splenic lesions. J Vet Intern Med 2008;5:1095–1102.

11. Nakamura K, Takagi S, Sasaki N, et al. Contrast-enhanced ultrasonography for characterization of canine focal liver lesions. Vet Radiol Ultrasound 2010;51:79–85.

12. O'Brien RT, Iani M, Matheson J, et al. Contrast harmonic ultrasound of spontaneous liver nodules in 32 dogs. Vet Radiol Ultrasound 2004;45:547–553.

13. Kutara K, Asano K, Kito A, et al. Contrast harmonic imaging of canine hepatic tumors. J Vet Med Sci 2006;68:433–438.

14. Haers H, Vignoli M, Paes G, et al. Contrast harmonic ultrasonographic appearance of focal space-occupying renal lesions. Vet Radiol Ultrasound 2010;51:516–522.

15. Salwei RM, O'Brein RT, Matheson JS. Use of contrast harmonic ultrasound for the diagnosis of congenital portosystemic shunts in three dogs. Vet Radiol Ultrasound 2003;44:301–305.

16. Lassau N, Lamuraglia M, Chami L, et al. Gastrointestinal stromal tumors treated with imatinib: monitoring response with contrast-enhanced sonography. Am J Roentgenol 2006;187:1267–1273.

17. Lassau N, Chebil M, Chami L, Roche A. A new functional imaging technique for the early functional evaluation of antiangiogenic treatment: dynamic contrast-enhanced ultrasonography (DCE-US). Targ Oncol 2008;3:111–117.

18. Shian L, Pintong H, Zongmin W, et al. The relationship between enhanced intensity and microvessel density of gastric carcinoma using double contrast-enhanced ultrasonography. Ultrasound Med Biol 2009;35:1086–1091.

19. Estebana JM, Aleixandrea A, Hurtadoa MJ, et al. Contrastenhanced power Doppler ultrasound in the diagnosis and follow-up of inflammatory abdominal masses in Crohn's disease. Eur J Gastroen Hepat 2003;15:253–259.

20. Robotti D, Cammarota T, Debani P, et al. Activity of Crohn disease: value of color-power Doppler and contrast-enhanced ultrasonography. Abdom Imaging 2004;29:648–652.

21. Schreyer AG, Finkenzeller T, Gössmann H, et al. Microcirculation and perfusion with contrast enhanced ultrasound (CEUS) in Crohn's disease: first results with linear contrast harmonic imaging (CHI). Clin Hemorheol Microcirc 2008;40:143–155.

22. Girlich C, Jung EM, Iesalnieks I, et al. Quantitative assessment of bowel wall vascularisation in Crohn's disease with contrast-enhanced ultrasound and perfusion analysis. Clin Hemorheol Microcirc 2009;43:141–148.

23. Ripollés T, Martínez MJ, Paredes JM, Blanc E, et al. Crohn disease: correlation of findings at contrast-enhanced US with severity at endoscopy. Radiology 2009;253:241–248.

24. Serra C, Menozzi G, Morselli Labate AM, et al. Ultrasound assessment of vascularization of the thickened terminal ileum wall in Crohn's disease patients using a low-mechanical index real-time scanning technique with a second generation ultrasound contrast agent. Eur J Radiol 2007; 62:114–121.

25. Hata J, Kamada T, Haruma K, Kusunoki H. Evaluation of bowel ischemia with contrast-enhanced US: initial experience. Radiology 2005;236:712–715.

26. Penninck D. Gastrointestinal tract. In: Penninck D, d'Anjou MA (eds): Atlas of small animal ultrasonography. Iowa: Blackwell publishing, 2008;281–318.

27. Jimenez DA, O'Brien RT, Wallace JD, Klocke E. Intraoperative contrast-enhanced ultrasonography of normal canine jejunum. Vet Radiol Ultrasound 2011;52:196–200.

 Piasecki C, Wyatt C. Patterns of blood supply to the gastric mucosa. A comparative study revealing an end-artery model. J Anat 1986;149:21–39.

29. Lundgren O. Studies on blood flow distribution and counter current exchange in the small intestine. Acta Physiol Scand 1967;303:5–42.

30. Kampp M, Lundgren O. Blood flow and flow distribution in the small intestine of the cat as analysed by the Kr85 wash out technique. Acta Physiol Scand 1968;72:282–297.

31. Lundgren O. The circulation of the small bowel mucosa. Gut 1974;15:1005–1013.

32. Granger DN, Taylor AE. Volumetric assessment of the capillary filtration coefficient in the cat small intestine. Pfluegers Arch Eur J Physiol 1979;381:25–33.

33. Premen AJ, Banchs V, Womack WA, et al. Importance of collateral circulation in the vascularly occluded feline intestine. Gastroenterology 1987;92:1215–1219.

34. Spalinger J, Patriquin H, Miron M, et al. Doppler US in patients with Crohn disease: vessel density in the diseased bowel reflects disease activity. Radiology 2000;217:787–791.

35. Esteban JM, Maldonado L, Sanchiz V, et al. Activity of Crohn's disease assessed by color Doppler ultrasound analysis of the affected segments. Eur Radiol 2001;11:1423–1428.

36. Scholbach T, Herrero I, Scholbach J. Dynamic color Doppler sonography of intestinal wall in patients with Crohn disease compared with healthy subjects. J Pediatr Gastr Nutr 2004;39:524–528.

37. Gaschen L, Kircher P. Two-dimensional grayscale ultrasound and spectral Doppler waveform evaluation of dogs with chronic enteropathies. Clin Tech Small Anim Pract 2007;22:122–127.

38. Gaschen L, Kircher P, Stüssi A, et al. Comparison of Ultrasonographic Findings with Clinical Activity Index (CIBDAI) and diagnosis in dogs with chronic enteropathies. Vet Radiol Ultrasound 2008;49:56–64.

39. Browne JE, Watson AJ, Hoskins PR, et al. Validation of a sensitivity performance index test protocol and evaluation of color Doppler sensitivity for a range of ultrasound scanners. Ultrasound Med Biol 2004;30:1475–1483.

40. Leinonen MR, Raekallio MR, Vainio OM, O'Brien RT. Effect of anaesthesia on contrast-enhanced ultrasound of the feline spleen. Vet J, doi:10.1016/j.tvjl.2010.10.013.

41. Brown MJ, McCarthy TJ, Bennett BT. Long term anesthesia using a continuous infusion of guaifenesin, ketamine and xylazine in cats. Lab Anim Sci 1991;41:46–50.

42. Magoon KE, Hsu WH, Hembrough FB. The influence of atropine on the cardiopulmonary effects of a xylazine-ketamine combination in dogs. Arch Int Pharmacodyn Ther 1988;293:143–153.

43. Leinonen MR, Raekallio MR, Vainio OM, et al. The effect of the sample size and location on contrast ultrasound measurement of perfusion parameters. Vet Radiol Utrasound 2010;52:82–87.

44. Mule S, De Cesare A, Lucidarme O, et al. Regularized estimation of contrast agent attenuation to improve the imaging of microbubbles in small animal studies. Ultrasound Med Biol 2008;34:938–948.

45. Li J, Dong B, Yu X, et al. Grey-scale contrast enhancement in rabbit liver with SonoVue at different doses. Ultrasound Med Biol 2005;31: 185–190.

46. Sonne C, Xie F, Lof J, et al. Differences in definity and optison microbubble destruction rates at a similar mechanical index with different real-time perfusion systems. J Am Soc Echocardiogr 2003;16:1178–1185.