### **ILLUSTRATED REVIEW**



## Polyphosphate in thrombosis, hemostasis, and inflammation

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#### **Funding information**

National Heart, Lung, and Blood Institute, Grant/Award Number: R35 HL135823

#### **Abstract**

This illustrated review focuses on polyphosphate as a potent modulator of the plasma clotting cascade, with possible roles in hemostasis, thrombosis, and inflammation. Polyphosphates are highly anionic, linear polymers of inorganic phosphates that are widespread throughout biology. Infectious microorganisms accumulate polyphosphates with widely varying polymer lengths (from a few phosphates to over a thousand phosphates long), while activated human platelets secrete polyphosphate with a very narrow size distribution (about 60-100 phosphates long). Work from our lab and others has shown that long-chain polyphosphate is a potent trigger of clotting via the contact pathway, while polyphosphate of the size secreted by platelets accelerates factor V activation, blocks the anticoagulant activity of tissue factor pathway inhibitor, promotes factor XI activation by thrombin, and makes fibrin fibrils thicker and more resistant to fibrinolysis. Polyphosphate also modulates inflammation by triggering bradykinin release, inhibiting the complement system, and modulating endothelial function. Polyphosphate and nucleic acids have similar physical properties and both will trigger the contact pathway—although polyphosphate is orders of magnitude more procoagulant than either DNA or RNA. Important caveats in these studies include observations that nucleic acids and polyphosphate may co-purify, and that these preparations can be contaminated with highly procoagulant microparticles if silica-based purification methods are employed. Polyphosphate has received attention as a possible therapeutic, with some recent studies exploring the use of polyphosphate in a variety of formulations to control bleeding. Other studies are investigating treatments that block polyphosphate function as novel antithrombotics with the possibility of reduced bleeding side effects.

### KEYWORDS

blood coagulation, contact pathway, DNA, nucleic acids, polyphosphate, RNA

### **Essentials**

- Polyphosphate is present in microorganisms and human cells such as platelets.
- Polyphosphate modulates coagulation via interactions with multiple proteins.
- Polyphosphate modulates inflammation by triggering bradykinin release and inhibiting complement.
- · Nucleic acids and polyphosphate co-purify and may be contaminated with silica-based methods.

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# Where is polyP?

In microbes, polyP size is heterogeneous, ranging from a few phosphates to over a thousand phosphates in length.¹

Trypanosome

Trypanosome

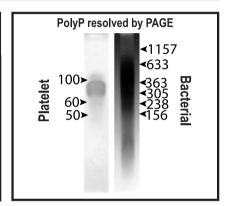
I have it too!

It's stored in acidocalcisomes (eukaryotes) or volutin granules (bacteria), along with divalent metals and amines.<sup>2,3</sup>

Platelet dense granules are similar in composition to acidocalcisomes and have abundant polyP. Its polyP length is tightly regulated (60-100 phosphates long).

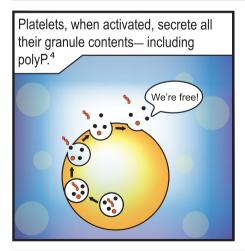
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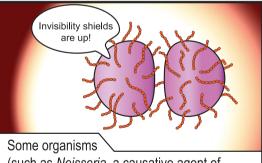
image courtesy of Roberto Docampo



PolyP is also reported in mast cells,<sup>5</sup> prostasomes,<sup>6</sup> cardiac muscle,<sup>7,8</sup> brain,<sup>9</sup> and nervous tissue.<sup>10</sup>

## How does it come in contact with blood?





(such as *Neisseria*, a causative agent of meningitis) express polyP on their capsule surface.<sup>11</sup> It is a virulence factor that may allow them to evade complement killing.<sup>12,13</sup>

PolyP may be released following tissue damage or cell necrosis.

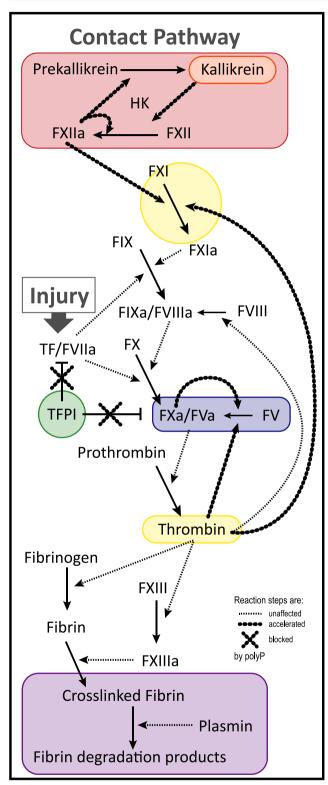
PolyP is very short (25mer) in cardiac muscle,<sup>7-8</sup> slightly longer in platelets<sup>4</sup> and mast cells<sup>5</sup> (60-100mer). Some tissues, such as brain,contain very long polymers (800mer).<sup>9</sup>

PolyP is widespread in infectious microorganisms and is released by activated platelets and mast cells. It may also be released following tissue damage. In these settings, it may act as a pathogen-associated molecular pattern (PAMP) or a damage-associated molecular pattern (DAMP) to help trigger host defenses.

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# Roles of polyP in coagulation & kinin generation



## PolyP is a contact activator

Long-chain (microbial) polyP triggers clotting by providing a template for autoactivation and reciprocal activation of the proteins in the contact pathway. The enzymes, FXIIa and kallikrein, are generated. Platelet-size polyP supports this reaction poorly. The contact pathway is a polyP support to the contact pathway.

### PolyP causes bradykinin release

Newly generated kallikrein cleaves high molecular weight kininogen to release bradykinin, which is a potent vasodilator and proinflammatory mediator.<sup>16</sup>

## PolyP greatly accelerates FXI activation

FXI deficiency is associated with bleeding, indicating that FXI activation plays a role in hemostasis.<sup>17</sup> While thrombin activation of FXI is slow, <sup>18,19</sup> platelet-size polyP enhances its rate 3000-fold, making this back-activation reaction a physiologically relevant contributor to sustained thrombin generation.<sup>20,21</sup>

## PolyP blocks TFPI activity

Tissue Factor Pathway Inhibitor (TFPI) antagonizes tissue factor-dependent initiation of coagulation by inhibiting FXa and FVIIa. Platelet-size polyP potently abrogates TFPI's anticoagulant function<sup>14,22</sup> and enhances the inactivation of TFPI by FXIa.<sup>23</sup>

## PolyP accelerates thrombin generation

FV activation is a rate-limiting step in thrombin generation. Because platelet-size polyP enhances the rate of FV cleavage by FXIa, FXa and thrombin, the kinetics of thrombin generation are improved.<sup>22,24</sup>

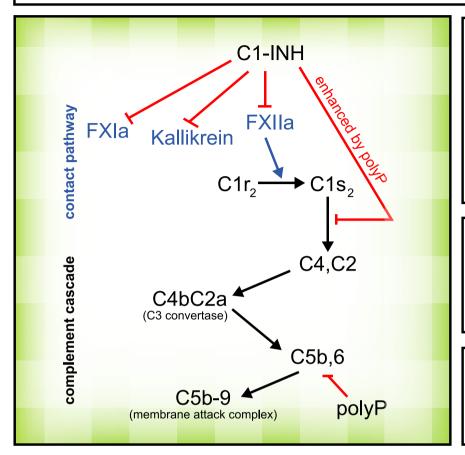
## PolyP strengthens clots and delays lysis

PolyP is incorporated into fibrin, leading to thicker fibrin fibrils that are more resistant to fibrinolysis. 14,25,26

PolyP accelerates blood clotting by targeting a few specific points in the clotting cascade, always in a procoagulant manner. The exact steps modulated by polyP depend on its polymer length.



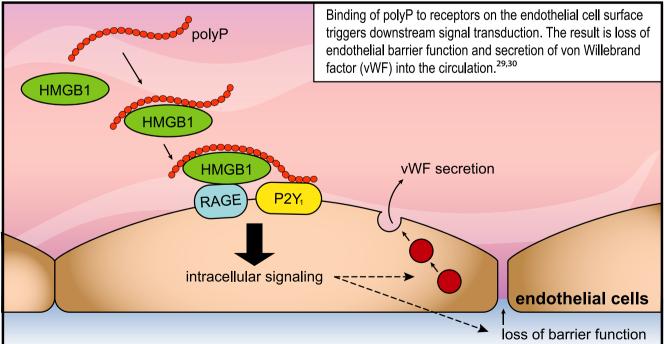
# Roles of polyP outside of coagulation



Although C1 esterase inhibitor (C1-INH) is a promiscuous serpin which can inhibit members of both the contact pathway (FXIa, FXIIa, and kallikrein) and the complement cascade (C1s), polyP only enhances the inhibitory effect of C1-INH toward complement,<sup>27</sup> not toward clotting factors.

PolyP also destabilizes the C5b,6 complex, thereby reducing the lytic capacity of the membrane attack complex of the complement system.<sup>28</sup>

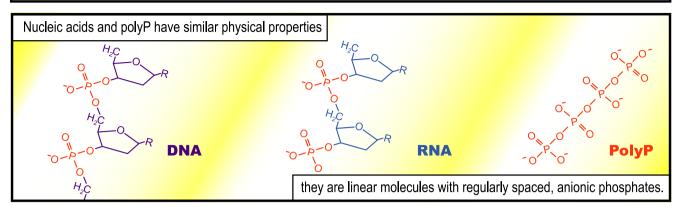
The overall effect of polyP is downregulation of the complement system, which is opposite to the effect it has on the clotting cascade.<sup>28</sup>

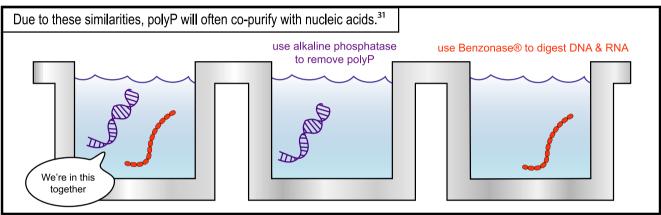


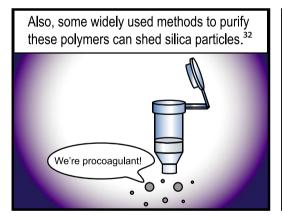
It's likely that additional roles of polyP in inflammation and vascular function are yet to be found.

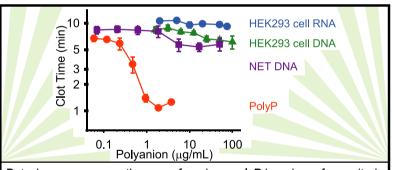


# Comparison between DNA, RNA, and polyP

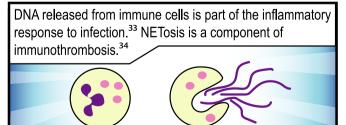


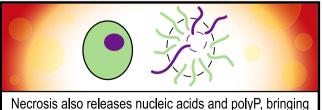






But when you remove those confounders, polyP is orders of magnitude more active than DNA or RNA at activating the contact system.<sup>31</sup>



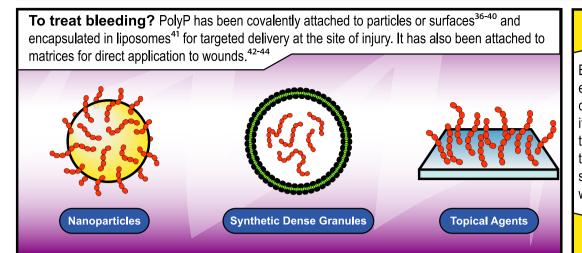


Necrosis also releases nucleic acids and polyP, bringing them into contact with clotting proteins in plasma.<sup>35</sup>

Extracellular polyP, DNA, and RNA can be procoagulant and proinflammatory, but their contribution to these processes requires much further investigation!

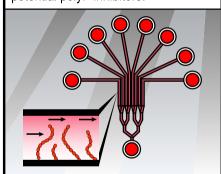


# PolyP as a possible therapeutic or target

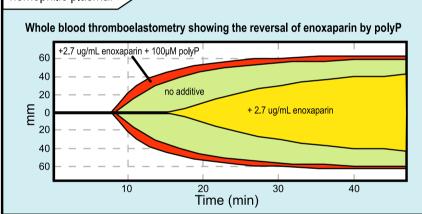


Because polyP enhances fibrin clot structure,<sup>25</sup> it could be used to augment treatment of surgical bleeding with fibrin glue.

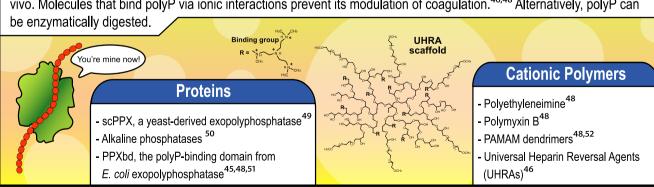
Ex vivo investigations of whole blood using microfluidics are helping define polyP's contributions to hemostasis and thrombosis under more realistic conditions.<sup>37,45</sup> These studies also test potential polyP inhibitors.<sup>45-46</sup>



Adding polyP to blood in vitro reverses the anticoagulant effects of heparins and DOACS (direct FXa or thrombin inhibitors). PolyP also mitigates the anticoagulant effects of vitamin K antagonists and enhances clotting in hemophilic plasma.<sup>47</sup>



**To treat or prevent thrombosis?** Various compounds antagonize the procoagulant effects of polyP, both in vitro and in vivo. Molecules that bind polyP via ionic interactions prevent its modulation of coagulation. Alternatively, polyP can be appropriately disperted.



Some researchers are incorporating polyP into novel hemostatic agents to control bleeding. Also, because polyP enhances, but is not essential for, coagulation, it represents an attractive target for thrombosis prevention and treatment.

#### **ACKNOWLEDGMENTS**

The authors thank Roberto Docampo for sharing his platelet images.

#### **RELATIONSHIP DISCLOSURES**

SAS and JHM hold patents related to the potential medical uses of polyphosphate and polyphosphate inhibitors. JHM has equity ownership in PrevThro Pharmaceuticals and consults for Cayuga Pharmaceuticals.

#### **AUTHOR CONTRIBUTIONS**

CJB created the graphics; SAS contributed images and wrote the text; CJB, SAS, and JHM contributed to the conceptual design and edited the manuscript.

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### REFERENCES

- Rao NN, Gómez-García MR, Kornberg A. Inorganic polyphosphate: essential for growth and survival. Annu Rev Biochem. 2009;78:605-47.
- Ruiz FA, Rodrigues CO, Docampo R. Rapid changes in polyphosphate content within acidocalcisomes in response to cell growth, differentiation, and environmental stress in *Trypanosoma cruzi*. J Biol Chem. 2001;276:26114–21.
- 3. Docampo R, Moreno SN. Acidocalcisomes. Cell Calcium. 2011; 50:113-9.
- Ruiz FA, Lea CR, Oldfield E, Docampo R. Human platelet dense granules contain polyphosphate and are similar to acidocalcisomes of bacteria and unicellular eukaryotes. J Biol Chem. 2004;279:44250-7.
- Moreno-Sanchez D, Hernandez-Ruiz L, Ruiz FA, Docampo R. Polyphosphate is a novel pro-inflammatory regulator of mast cells and is located in acidocalcisomes. J Biol Chem. 2012;287:28435–44.
- Nickel KF, Ronquist G, Langer F, et al. The polyphosphate-factor XII pathway drives coagulation in prostate cancer-associated thrombosis. Blood. 2015;126:1379–89.
- Dedkova EN. Inorganic polyphosphate in cardiac myocytes: from bioenergetics to the permeability transition pore and cell survival. Biochem Soc Trans. 2016;44:25–34.
- Seidlmayer LK, Gómez-García MR, Blatter LA, Pavlov E, Dedkova EN. Inorganic polyphosphate is a potent activator of the mitochondrial permeability transition pore in cardiac myocytes. J Gen Physiol. 2012;139:321–31.
- Kumble KD, Kornberg A. Inorganic polyphosphate in mammalian cells and tissues. J Biol Chem. 1995;270:5818–22.
- Stotz SC, Scott LO, Drummond-Main C, et al. Inorganic polyphosphate regulates neuronal excitability through modulation of voltage-gated channels. Mol Brain. 2014;7:42.
- Noegel A, Gotschlich EC. Isolation of a high molecular weight polyphosphate from *Neisseria gonorrhoeae*. J Exp Med. 1983;157: 2049–60.

- Zhang Q, Li Y, Tang CM. The role of the exopolyphosphatase PPX in avoidance by *Neisseria meningitidis* of complement-mediated killing. J Biol Chem. 2010;285:34259–68.
- Tinsley CR, Gotschlich EC. Cloning and characterization of the meningococcal polyphosphate kinase gene: production of polyphosphate synthesis mutants. Infect Immun. 1995;63:1624–30.
- 14. Smith SA, Choi SH, Davis-Harrison R, et al. Polyphosphate exerts differential effects on blood clotting, depending on polymer size. Blood. 2010;116:4353-9.
- Engel R, Brain CM, Paget J, Lionikiene AS, Mutch NJ. Single-chain factor XII exhibits activity when complexed to polyphosphate. J Thromb Haemost. 2014;12:1513–22.
- Müller F, Mutch NJ, Schenk WA, et al. Platelet polyphosphates are proinflammatory and procoagulant mediators in vivo. Cell. 2009;139:1143-56.
- Seligsohn U. Factor XI deficiency in humans. J Thromb Haemost. 2009;7(Suppl 1):84-7.
- Naito K, Fujikawa K. Activation of human blood coagulation factor XI independent of factor XII. Factor XI is activated by thrombin and factor XIa in the presence of negatively charged surfaces. J Biol Chem. 1991;266:7353-8.
- Gailani D, Broze GJ Jr. Factor XI activation in a revised model of blood coagulation. Science. 1991;253:909-12.
- Choi SH, Smith SA, Morrissey JH. Polyphosphate is a cofactor for the activation of factor XI by thrombin. Blood. 2011;118:6963-70.
- 21. Geng Y, Verhamme IM, Smith SB, et al. The dimeric structure of factor XI and zymogen activation. Blood. 2013;121:3962–3669.
- Smith SA, Mutch NJ, Baskar D, Rohloff P, Docampo R, Morrissey JH. Polyphosphate modulates blood coagulation and fibrinolysis. Proc Natl Acad Sci USA. 2006;103:903–8.
- 23. Ivanov I, Shakhawat R, Sun MF, et al. Nucleic acids as cofactors for factor XI and prekallikrein activation: different roles for high-molecular-weight kininogen. Thromb Haemost. 2017; 117:471-81
- Choi SH, Smith SA, Morrissey JH. Polyphosphate accelerates factor V activation by factor XIa. Thromb Haemost. 2015;113:599–604.
- 25. Smith SA, Morrissey JH. Polyphosphate enhances fibrin clot structure. Blood. 2008;112:2810-6.
- Mutch NJ, Engel R, Uitte de Willige S, Philippou H, Ariens RA. Polyphosphate modifies the fibrin network and down-regulates fibrinolysis by attenuating binding of tPA and plasminogen to fibrin. Blood. 2010:115:3980-8.
- 27. Wijeyewickrema LC, Lameignere E, Hor L, et al. Polyphosphate is a novel cofactor for regulation of complement by a serpin, C1 inhibitor. Blood. 2016;128:1766–76.
- 28. Wat JM, Foley JH, Krisinger MJ, et al. Polyphosphate suppresses complement via the terminal pathway. Blood. 2014;123:768–76.
- Hassanian SM, Dinarvand P, Smith SA, Rezaie AR. Inorganic polyphosphate elicits proinflammatory responses through activation of mTOR complexes 1 and 2 in vascular endothelial cells. J Thromb Haemost. 2015;13:860–71.
- 30. Dinarvand P, Hassanian SM, Qureshi SH, et al. Polyphosphate amplifies proinflammatory responses of nuclear proteins through interaction with receptor for advanced glycation end products and P2Y<sub>1</sub> purinergic receptor. Blood. 2014;123:935–45.
- Smith SA, Gajsiewicz JM, Morrissey JH. Ability of polyphosphate and nucleic acids to trigger blood clotting: some observations and caveats. Front Med. 2018;5:107.
- Smith SA, Baker CJ, Gajsiewicz JM, Morrissey JH. Silica particles contribute to the procoagulant activity of DNA and polyphosphate isolated using commercial kits. Blood. 2017;130:88–91.
- Branzk N, Papayannopoulos V. Molecular mechanisms regulating NETosis in infection and disease. Semin Immunopathol. 2013; 35:513–30.

- Martinod K, Wagner DD. Thrombosis: tangled up in NETs. Blood. 2014;123:2768-76.
- 35. Esmon CT. Molecular circuits in thrombosis and inflammation. Thromb Haemost. 2013;109:416–20.
- Kudela D, Smith SA, May-Masnou A, et al. Clotting activity of polyphosphate-functionalized silica nanoparticles. Angew Chem Int Ed. 2015:54:4018–22.
- Yeon JH, Mazinani N, Schlappi TS, et al. Localization of short-chain polyphosphate enhances its ability to clot flowing blood plasma. Sci Rep. 2017;7:42119.
- Donovan AJ, Kalkowski J, Smith SA, Morrissey JH, Liu Y. Sizecontrolled synthesis of granular polyphosphate nanoparticles at physiologic salt concentrations for blood clotting. Biomacromol. 2014;15:3976–84.
- 39. Szymusiak M, Donovan AJ, Smith SA, et al. Colloidal confinement of polyphosphate on gold nanoparticles robustly activates the contact pathway of blood coagulation. Bioconjug Chem. 2016;27:102–9.
- Schröder HC, Tolba E, Diehl-Seifert B, Wang X, Müller WE. Electrospinning of bioactive wound-healing nets. Prog Mol Subcell Biol. 2017;55:259–90.
- Donovan AJ, Kalkowski J, Szymusiak M, et al. Artificial dense granules: a procoagulant liposomal formulation modeled after platelet polyphosphate storage pools. Biomacromol. 2016;17:2572–81.
- 42. Sakoda M, Kaneko M, Ohta S, et al. Injectable hemostat composed of a polyphosphate-conjugated hyaluronan hydrogel. Biomacromol. 2018;19:3280–90.
- Wang Y, Kim K, Lee MS, Lee H. Hemostatic ability of chitosanphosphate inspired by coagulation mechanisms of platelet polyphosphates. Macromol Biosci. 2018;18:e1700378.
- Ong SY, Wu J, Moochhala SM, Tan MH, Lu J. Development of a chitosan-based wound dressing with improved hemostatic and antimicrobial properties. Biomaterials. 2008;29:4323–32.

- Zhu S, Travers RJ, Morrissey JH, Diamond SL. FXIa and platelet polyphosphate as therapeutic targets during human blood clotting on collagen/tissue factor surfaces under flow. Blood. 2015;126:1494–502.
- Travers RJ, Shenoi RA, Kalathottukaren MT, Kizhakkedathu JN, Morrissey JH. Nontoxic polyphosphate inhibitors reduce thrombosis while sparing hemostasis. Blood. 2014;124:3183–90.
- 47. Smith SA, Morrissey JH. Polyphosphate as a general procoagulant agent. J Thromb Haemost. 2008;6:1750-6.
- 48. Smith SA, Choi SH, Collins JN, Travers RJ, Cooley BC, Morrissey JH. Inhibition of polyphosphate as a novel strategy for preventing thrombosis and inflammation. Blood. 2012;120:5103–10.
- Wurst H, Kornberg A. A soluble exopolyphosphatase of Saccharomyces cerevisiae. Purification and characterization. J Biol Chem. 1994;269:10996–1001.
- Lorenz B, Schröder HC. Mammalian intestinal alkaline phosphatase acts as highly active exopolyphosphatase. Biochim Biophys Acta. 2001;1547:254-61.
- Labberton L, Kenne E, Long AT, et al. Neutralizing blood-borne polyphosphate in vivo provides safe thromboprotection. Nat Commun. 2016;7:12616.
- Jain S, Pitoc GA, Holl EK, et al. Nucleic acid scavengers inhibit thrombosis without increasing bleeding. Proc Natl Acad Sci USA. 2012;109:12938-43.

How to cite this article: Baker CJ, Smith SA, Morrissey JH. Polyphosphate in thrombosis, hemostasis, and inflammation. *Res Pract Thromb Haemost*. 2019;3:18–25. https://doi.

org/10.1002/rth2.12162