




In Vitro and In Vivo Antimalarial Activities and Toxicological Assessment of *Pogostemon Cablin* (Blanco) Benth

Arisara Phuwajaroanpong, BSc¹, Prapaporn Chaniad, PhD¹,
 Natharinee Horata, PhD², Saowanee Muangchanburee, MSc³,
 Kantarakorn Kaewdana, BSc¹, and Chuchard Punsawad, PhD¹ 

Abstract

The aim of this study was to investigate the antimalarial activities and toxicity of *Pogostemon cablin* extracts. *In vitro* activities against the chloroquine-resistant *Plasmodium falciparum* K1 strain were assessed by using the *Plasmodium* lactate dehydrogenase enzyme (pLDH) assay, while *in vivo* activity against the *Plasmodium berghei* ANKA strain in mice was investigated using a 4-day suppressive test. The *in vitro* and *in vivo* toxicity were determined in Vero cells and mice, respectively. The ethanolic extract possessed antimalarial activity with an IC₅₀ of 24.49 ± 0.01 µg/ml, whereas the aqueous extract showed an IC₅₀ of 549.30 ± 0.07 µg/ml. Cytotoxic analyses of the ethanolic and aqueous extracts revealed a nontoxic effect on Vero cells at a concentration of 80 µg/ml. Based on a preliminary study of *in vitro* antimalarial activity, the ethanolic extract was chosen as a potential agent for further *in vivo* antimalarial activity analysis in mice. The ethanolic extract, which showed no toxic effect on mice at a dose of 2000 mg/kg body weight, significantly suppressed parasitemia in mice by 38.41%, 45.12% and 89.00% at doses of 200, 400 and 600 mg/kg body weight, respectively. In conclusion, this study shows that the ethanolic *P. cablin* extract possesses *in vitro* and *in vivo* antimalarial activity without toxic effects.

Keywords

malaria, antimalarial activity, *Pogostemon cablin* (Blanco) benth, *Plasmodium falciparum*, *Plasmodium berghei*, toxicity

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Introduction

Malaria is an important parasite-induced tropical disease that threatens many people globally.^{1,2} The malaria parasites, namely, *Plasmodium falciparum*, *Plasmodium vivax*, *Plasmodium malariae*, *Plasmodium ovale* and *Plasmodium knowlesi*, are the 5 species that cause malaria in humans. According to a report published in 2019, approximately 228 million malaria cases occurred worldwide, resulting in 405,000 deaths.^{3,4} The severe complications associated with malaria are related to mortality incidence and are a significant public health concern.⁵ At present, treatment of malaria involves mainly chemotherapy. However, emerging resistance to antimalarial drugs of different classes has become an urgent problem.⁶⁻¹⁰ The treatment failure rates of the first-line drug artemisinin-based combination therapy against *P. falciparum* were above 10% in the WHO Southeast Asia region and were as high as 93% in Thailand,³ indicating that new therapeutic options are urgently needed. Natural medicinal products serve as potential

sources of diverse pharmacological activities based on types of bioactive compounds with differences in chemical structures. *Pogostemon cablin* (Blanco) Benth., commonly recognized as “Patchouli” or “Phimsen ton” in Thailand, belongs to the Lamiaceae family,¹¹ which is commonly found in South Asian countries.¹² This plant is a short-lived perennial herb that grows to 1.2 m tall and is branched with an erect and quadrangular stem.¹³ It is traditionally used as remedies for colds, fever,

¹ School of Medicine, Walailak University, Nakhon Si Thammarat, Thailand

² Faculty of Medical Technology, Huachiew Chalermprakiet University, Samutprakan, Thailand

³ Department of Biomedical Sciences, Faculty of Medicine, Prince of Songkla University, Songkhla, Thailand

Corresponding Author:

Chuchard Punsawad, PhD, School of Medicine, Walailak University, Nakhon Si Thammarat 80160, Thailand.
 Email: chuchard.pu@wu.ac.th



abdominal pain, diarrhea, nausea, vomiting, and insect bites and possesses many pharmacological activities, such as antimicrobial, gastroprotective, antioxidant, antiemetic, antithrombotic, blood coagulative and fibrinolytic activities.¹⁴⁻¹⁷ However, neither the antimalarial activity nor the toxicity of *P. cablin* had been studied until this study.

Experimental Section

Plant Material

The dried aerial part of *P. cablin* was obtained from a traditional Thai drug store in Nakhon Si Thammarat Province, Thailand. The plant was botanically identified by Assoc. Prof. Tanomjit Supavita, an expert botanist, and has a voucher number of SMD143228 at the School of Medicine, Walailak University. The plant material was cleaned to remove dust or dead material and dried at 60°C by using hot air oven. The dried plant material was coarsely chopped using an herb grinder (Jincheng, Model; SF, China) and stored at room temperature until extraction.

Plant Extraction

P. cablin was extracted with ethanol by maceration and with water extraction by the reflux method. Sixty grams of plant powder was soaked in 600 ml of ethanol at room temperature for 72 h, and another 60 grams was extracted with 600 ml of distilled water under reflux for 2 h. These were then filtered and evaporated using a rotary evaporator (Rotavapor®, Buchi, China) at 60°C. The dried crude extracts were collected into screw cap containers and stored in a refrigerator at 4°C.

Phytochemical Analysis

Phytochemical analysis of ethanolic and aqueous extracts of *P. cablin* was conducted to determine their phytoconstituents, including flavonoids, terpenoids, alkaloids, tannins, anthraquinone, cardiac glycosides, saponins and coumarins. This screening was performed according to methods described by Ngbolua et al and Malar et al.^{18,19}

In vitro Plasmodium Falciparum Cultivation

The culture of *P. falciparum* was performed using the method demonstrated by Trager and Jensen with some modifications.²⁰ The chloroquine-resistant *P. falciparum* K1 strain, obtained from Dr. Rapatbhorn Patrapuvich, Department of Drug Research Unit for Malaria, Faculty of Tropical Medicine, Mahidol University, Thailand, was cultivated with noninfected type O-positive red blood cells (2% hematocrit) using RPMI-1640 supplemented with 2 mg/ml sodium bicarbonate, 10 µg/ml hypoxanthine (Sigma-Aldrich, New Delhi, India), 4.8 mg/ml HEPES (Himedia, Mumbai, India), 0.5% AlbuMAX II (Gibco, MA, USA) and 2.5 µg/ml gentamicin (Sigma-Aldrich, New Delhi, India). *P. falciparum* culture was maintained in 5% CO₂ at 37°C. The culture medium was changed daily after assessing parasite density by Giemsa staining.

In vitro Assessment of Antimalarial Activity

The antimalarial activity of ethanolic and aqueous extracts of *P. cablin* was carried out using an *in vitro* *Plasmodium* lactate dehydrogenase (pLDH) assay as described by Makler with some modifications.²¹ Briefly, the crude extracts (500 mg) were dissolved in 1 ml of dimethyl sulfoxide (DMSO) (Merck, Darmstadt, Germany) and adjusted to obtain a 2-fold serial dilution in the range of 9.77 to 2,500 µg/ml. The parasitized red blood cells (2% hematocrit and 2% parasitemia) were added to a 96-well cell culture plate, and then, the infected red cells were exposed to the ethanolic and aqueous *P. cablin* extracts (final concentration of each extract: 9.77 µg/ml). Wells containing artesunate (Sigma-Aldrich, New Delhi, India) served as a positive control (final concentration ranging from 0.31 to 10 µg/ml). The plates were incubated in a CO₂ incubator at 37°C for 72 h, frozen at -20°C and thawed at 37°C 3 times to achieve complete hemolysis. The lysed cells were transferred to a new 96-well plate containing 100 µl of Malstat reagent and 20 µl of nitroblue tetrazolium/phenazine ethosulfate solution (Calbiochem®, Sigma-Aldrich, New Delhi, India) and kept in the dark for 1 h. Optical density was determined at a 650 nm wavelength using a microplate reader. The percentage of inhibition and the half maximal inhibitory concentration (IC₅₀) were determined from a nonlinear dose dependence curve with concentrations ranging from 9.77 to 2,500 µg/ml.

In vitro Assessment of Cytotoxicity

The 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay was used to assess the toxicity of *P. cablin*. Vero cells (Elabscience, Wuhan, Hubei, China) were cultured in Dulbecco's modified Eagle's medium (CaissonLab, Smithfield, UT) supplemented with 10% fetal bovine serum (CaissonLab, Smithfield, UT). Vero cells (10⁴ cells/ml) were added to a 96-well plate and incubated for 24 h at 37°C and 5% CO₂. *P. cablin* ethanolic and aqueous extracts were added to the plates at 5 concentrations ranging from 5 to 80 µg/ml. The wells containing doxorubicin (Sigma-Aldrich, New Delhi, India) were used as a positive control (final concentration: 0.63 µg/ml). The plates were incubated for 48 h at 37°C and 5% CO₂. After the incubation period, the MTT solution was added to the plates, which were then incubated in a CO₂ incubator for 2 h. The supernatant was removed, and 100 µl of DMSO was added to each well. Finally, the optical density was determined at a wavelength of 590 nm using a microplate reader. The percentage of survival and 50% cytotoxicity concentration (CC₅₀) were determined by using the dose response curve with concentrations ranging from 0.31 to 10 µg/ml.

Animal Experiments

Healthy ICR mice weighing 20-30 g (body weight) were bought from Nomura Siam International Co., Ltd., Pathumwan, Bangkok, Thailand. The mice were randomly grouped and acclimatized in cages for 1 week prior to the experiment. The

experimental animal room temperature was 22°C (\pm 3°C), and the relative humidity was 50-60%. The lighting environment was set to 12-h light/dark cycles, and the mice had free access to food pellets and clean drinking water provided by the animal care staff.

Four-Day Suppressive Test (Peter's Test)

The assessment of *in vivo* antimalarial activity was performed using a 4-day suppressive test to evaluate the schizonticidal activity of the extracts.²² Rodent parasites of *Plasmodium berghei* strain ANKA were obtained from BEI Resources, NIAID, NIH: *P. berghei*, strain ANKA, MRA-311, contributed by Thomas F. McCutchan. Five groups of ICR mice (5 mice each) were randomly created, including 3 groups of mice treated with 200, 400 and 600 mg/kg body weight ethanolic *P. cablin* extract according to previous studies²³⁻²⁵ and positive and negative control groups. Briefly, mice were inoculated intraperitoneally with 200 μ l of *P. berghei* ANKA-infected red cells containing 10⁷ cells/ml on day 0. At 4, 24, 48 and 72 h post-infection, each treated group received 3 doses of the extract via oral gavage. Artesunate at a dose of 6 mg/kg body weight and 7% Tween 80 were administered by oral gavage to the positive and negative control groups, respectively. Each mouse was weighed on the first and last days of the experiment by a sensitive digital balance (Mettler Toledo, Switzerland). At 96 h postinfection, thin blood films were prepared and stained with 10% Giemsa solution and then observed under a light microscope at 100 \times magnification to determine the percentage of parasitemia. The mean parasitemia was calculated from 10 different fields on each slide. The % parasitemia was calculated as follows:

$$\% \text{ parasitemia} = \frac{\text{number of parasitized red blood cell}}{\text{number of total red blood cell count}} \times 100$$

Finally, the following equation was used to calculate the mean percentage of parasitemia suppression:

$$\% \text{ suppression} = \frac{[A - B]}{A} \times 100$$

where A and B represent the mean percentages of negative parasitemia in the control group and the *P. cablin*-treated group, respectively.

Acute Toxicity Test

Fifteen ICR mice were randomly divided into 3 groups, including mice treated with 2000 mg/kg *P. cablin* extract and the negative control and untreated groups. The protocol of acute toxicity followed the 2008 OECD guidelines.²⁶ Before the experiment began, the mice were fasted for 3 h and had free access to only clean drinking water. The ethanolic *P. cablin* extract was dissolved in 7% Tween 80 to make a stock solution with a concentration of 2000 mg/kg. Mice in the experimental group were treated orally with a single dose of the extract at 2000 mg/kg body weight, whereas mice in the control group

were given a 7% Tween 80 solution. Mice were observed during the first 30 min after administration. Physical and behavioral changes, including changes in appetite, sleep, abnormal secretion, diarrhea, rigidity and hair erection, were noted for 14 days after administration. On day 14, 60 mg/kg pentobarbital (Nembutal, Ceva, France) was injected intraperitoneally as an anesthetic. Then, blood was collected using the cardiac puncture technique for biochemical analysis. In addition, kidney and liver tissues were processed and fixed in 10% formalin solution for histopathological examination.

Biochemical Analysis

Heparinized blood was centrifuged at 3,000 \times g for 5 min to obtain plasma samples. The samples were analyzed for biochemical parameters of kidney function (including blood urea nitrogen (BUN) and creatinine (Cr)) and liver function (including alanine aminotransferase (ALT), aspartate aminotransferase (AST), and alkaline phosphatase (ALP)) by standard techniques (kinetic methods for AST, ALT, ALP and BUN and a fixed time method for Cr) using an AU480 chemistry analyzer (Beckman Coulter, USA).

Histopathological Examination

Histopathological examination of formalin-fixed kidney and liver tissues was carried out according to a previous study.²⁷ Briefly, kidney and liver tissues were dehydrated with a gradient series of ethanol solutions and rinsed with xylene 3 times. Then, fixed tissues were placed in a mold that contained a small amount of molten paraffin. In the sectioning process, a 5- μ m section was sliced from the paraffin blocks of each tissue sample by a manual rotary microtome (Model RM2235, Leica Biosystems, Germany). Then, the sections were floated in a water bath and then transferred to glass slides. For the staining process, the sections were deparaffinized with xylene 3 times for 10 min each, rehydrated with a descending ethanol series of 100%, 96%, 90%, 80%, and 70% for 10 min each and stained with a hematoxylin and eosin (H&E) solution. Then, the sections were dehydrated by increasing the concentrations of ethanol as follows: 70%, 80%, 90%, 96%, and 100%. The sections were then rinsed with xylene and mounted with glass cover slips. Finally, 2 independent observers who were blinded to the group assignments examined the slides under a light microscope.

Statistical Analysis

The data were analyzed using SPSS for Microsoft Windows version 17.0 (SPSS, IL, USA). Normal distribution was determined by the Koimogorov-Smirnov goodness-of-fit test. The statistical significance of parasitemia suppression and the effect of the extract on kidney and liver functions in the acute toxicity test were tested by 1-way ANOVA followed by post hoc Tukey's multiple comparison test.

Table 1. Phytochemical Constituents of *P. cablin* in Different Solvent Extracts.

Phytochemical constituents	Ethanollic extract	Aqueous extract
Flavonoids	+	–
Terpenoids	–	–
Alkaloids	+	+
Tannins	+	+
Anthraquinones	–	–
Cardiac glycosides	–	–
Saponins	–	+
Coumarins	+	+

+: detected; –: not detected.

Results

Phytochemical Constituents

The ethanolic *P. cablin* extract exhibited alkaloids, flavonoids, coumarins and tannins, whereas the aqueous extract exhibited alkaloids, tannins, saponins and coumarins (Table 1).

In vitro Antimalarial Activity and Cytotoxicity

Quantification of antimalarial activity and cytotoxicity was performed by estimating the concentration of extracts that inhibited 50% of the growth of parasites (IC_{50}) and 50% of the viable cells. The ethanolic and aqueous extracts of *P. cablin* demonstrated antimalarial activity, with IC_{50} values of 24.49 ± 0.01 and 549.30 ± 0.07 $\mu\text{g/ml}$, respectively. Artesunate exhibited antimalarial activity with an IC_{50} value of 1.28 ± 0.71 $\mu\text{g/ml}$. The cytotoxicity of the extracts was determined with the highest concentration of 80 $\mu\text{g/ml}$. According to the classification of cytotoxicity,²⁸ the ethanolic and aqueous extracts of *P. cablin* exhibited no inhibitory effect at a concentration of 80 $\mu\text{g/ml}$, indicating no cytotoxic effects on Vero cells.

The 4-day Suppressive Test

The ethanolic *P. cablin* extract exhibited significant suppressive activity against *P. berghei* in a dose-dependent manner, with 38.41, 45.12 and 89.00% suppression at doses of 200, 400, and 600 mg/kg, respectively, compared to the activity in the negative control group ($p < 0.001$) (Table 2). The standard drug artesunate at a 6 mg/kg dose exhibited 96.69% suppression of *P. berghei* parasitemia compared to the activity in the negative control group, as well as significantly greater suppression than that in the other treated groups ($p < 0.001$) (Table 2).

In vivo Acute Toxicity Test

Physical and behavioral changes in mice treated with a dose of 2000 mg/kg extract were observed daily during the 14-day follow-up period. Extract-treated mice showed no signs or symptoms of toxicity, such as hair erection, changed feeding activities, vomiting, diarrhea, abnormal secretion, abnormal

sleep and excitement, during the experiment. The mean body weight and percent change in body weight of mice treated with 2000 mg/kg extract and 7% Tween 80 were not significantly different compared to those of the untreated control group ($p > 0.05$). There was no mortality at all dose levels tested. Based on these results, the lethal dose of the extract was found to be greater than 2000 mg/kg body weight.

Effects of Ethanolic *P. cablin* Extract on Kidney and Liver Functions

The biochemical parameters of creatinine and ALT in mice treated with 2000 mg/kg extract and 7% Tween 80 did not significantly differ from those in the untreated control group ($p < 0.05$) (Table 3). The levels of AST in mice treated with 7% Tween 80 and 2000 mg/kg extract were significantly increased compared to those in the untreated mice ($p < 0.05$). The levels of BUN in mice treated with the 2000 mg/kg extract were significantly less than those in the untreated mice ($p < 0.05$). The levels of AST and BUN in mice treated with 7% Tween 80 were significantly increased compared with those in untreated mice ($p < 0.05$) (Table 3).

Histopathological Examination of the Kidney and Liver in the Acute Toxicity Test

Figure 1 shows the results of the histological examinations of the kidney and liver tissues from mice in the negative control group and from mice treated with 2000 mg/kg body weight ethanolic *P. cablin* extract. The results demonstrated that mice treated with the 2000 mg/kg body weight extract had normal histological kidney and liver functions compared with those of the negative controls (Figure 1).

Discussion

In the extraction processes, the percentage yield was higher for the aqueous extract (13.83%) than for the ethanolic extract (3.45%). The higher yield may be due to the phytoconstituents in the extract being easier to extract in polar solvents such as aqueous solvents.^{29,30} The *in vitro* antimalarial activity of ethanolic and aqueous extracts of *P. cablin* was tested using enzymatic detection of the pLDH enzyme, which is the terminal enzyme in the glycolytic pathway.³¹ According to a previous study, the *in vitro* antimalarial activity of extracts was classified as follows: high extract activity at $IC_{50} < 5$ $\mu\text{g/ml}$, promising activity at 5–15 $\mu\text{g/ml}$, moderate activity at 15–50 $\mu\text{g/ml}$ and inactivity at > 50 $\mu\text{g/ml}$.³² Our results showed that the ethanolic extract possessed moderate antimalarial activity against the *P. falciparum* K1 strain with an IC_{50} value of 24.49 ± 0.01 $\mu\text{g/ml}$. Phytochemical examination was carried out for the 2 different solvent extracts. The phytochemical constituent analysis of *P. cablin* revealed alkaloids, tannins and coumarins in both extracts. In contrast, flavonoids were present only in the ethanolic extract, and saponins were present only in the aqueous extract. The absence of terpenoids, anthraquinones and cardiac

Table 2. Parasitemia Suppressive Activity of the Ethanolic *P. cablin* Extract Against *P. berghei*.

Group	Dose (mg/ml)	% Parasitemia	% Suppression
Negative control	-	52.39 ± 0.39 ^{b, c, d, e}	-
Artesunate	6	1.73 ± 0.67 ^{a, c, d, e}	96.69 ± 0.13 ^{a, c, d, e}
Ethanolic extract	200	32.27 ± 0.00 ^{a, b, d, e}	38.41 ± 0.00 ^{a, b, d, e}
	400	28.75 ± 0.14 ^{a, b, c, e}	45.12 ± 0.27 ^{a, b, c, e}
	600	5.76 ± 0.17 ^{a, b, c, d}	89.00 ± 0.32 ^{a, b, c, d}

Data are presented as the mean ± SEM (n = 5 per group).

^aCompared to negative control, ^bcompared to artesunate, ^ccompared to 200 mg/kg extract, ^dcompared to 400 mg/kg extract, ^ecompared to 600 mg/kg extract, *p* < 0.001.

Table 3. Effect of Ethanolic *P. cablin* Extract on Kidney and Liver Functions.

Parameters	Untreated control	7% Tween 80	2000 mg/kg extract
Liver function test			
AST (U/L)	127.50 ± 28.99	200.50 ± 3.53 ^a	172.50 ± 46.67 ^a
ALT (U/L)	36.00 ± 14.14	36.50 ± 9.19	39.5 ± 19.09
ALP (U/L)	84.50 ± 0.70	138.00 ± 31.11 ^{a, c}	82.50 ± 31.11 ^b
Kidney function test			
BUN (mg/dL)	23.55 ± 0.21	21.35 ± 0.49 ^{a, c}	20.00 ± 0.14 ^{a, b}
Creatinine (mg/dL)	0.55 ± 0.06	0.54 ± 0.01	0.59 ± 0.02

Data are presented as the mean ± SEM (n = 5 per group).

^aCompared to untreated control, ^bcompared to 7% Tween 80, ^ccompared to 2000 mg/kg extract, *p* < 0.05.

glycosides was found in both extracts in this study. Our results were similar to those of a previous study that reported chemical constituents from *P. cablin*.¹⁷ Moreover, the ethanolic *P. cablin* extract has been reported to have 12 secondary metabolites with one important flavonoid compound, tilianin.³³ Tilianin is a polyphenol flavonoid compound, and it was previously reported to have pharmacological activities such as antidiabetic, antihyperlipidemic, cardioprotective and anti-inflammatory activities.³⁴⁻³⁶ Secondary metabolites with pharmacological activity, such as flavonoids and alkaloids, could be accountable for the antimalarial activity.^{37,38} The potential toxicity of *P. cablin* extracts was evaluated in Vero cells. According to the cell cytotoxic classification, the CC₅₀ value was used to define the potency of cytotoxicity. A nontoxic effect could be classified as a CC₅₀ value greater than 50 µg/ml.²⁸ Ethanolic and aqueous extracts of *P. cablin* exhibited a Vero cell survival rate greater than 50% at 80 µg/ml, indicating no cytotoxic effects on Vero cells. This result was consistent with that of a previous study, which reported that the ethanolic *P. cablin* extract exhibited a noncytotoxic effect, with more than 70% cell viability.³⁹ The selection of the extract with the best antimalarial activity in an animal model that is widely used to predict drug activity was based on the greatest antimalarial activity and the lowest toxicity. According to the results of *in vitro* studies, the ethanolic *P. cablin* extract exhibited better antimalarial activity against the *P. falciparum* K1 strain and had no toxicity toward Vero cells. Therefore, the ethanolic extract was selected for further *in vivo* antimalarial activity and toxicological assessment in a mouse model. The highest

suppression occurred in mice treated with the extract at 600 mg/kg body weight (89.00%), followed by those treated with 400 and 200 mg/kg body weight (45.12 and 38.41%, respectively). These results were consistent with those of a previous study regarding the *in vivo* antimalarial activity of the *Betula alnoides* aqueous stem extract, which also exhibited significant suppressive activity against *P. berghei* in a dose-dependent manner.²³ To confirm extract safety, an acute toxicity test was performed by orally administering the extract at a dose of 2000 mg/kg body weight. Manifestations of toxicity or mortality were not observed. Furthermore, biochemical parameters and histopathological changes in the kidney and liver were determined to verify nonvisible toxicity. Biochemical analysis of kidney and liver function is crucial for evaluating the toxicological effects of xenobiotics.^{40,41} Plasma levels of AST in mice treated with the extract were higher than those in untreated mice, but the levels of ALT and ALP were not different from those in the control mice. Based on these results, ALT is a liver-specific enzyme, rather than AST, and the AST enzyme can be found in many other tissues, such as muscles, and in red blood cells.⁴² Thus, elevation of AST may occur due to other tissue injuries. This suggestion was confirmed by histopathological changes with normal histological features of the liver. However, the levels of AST and ALP in mice treated with 7% Tween 80 were increased compared with those in the other groups. This evidence might suggest that this vehicle should be used to dissolve crude extracts. For the kidney function test, BUN levels in mice treated with the extract and 7% Tween 80 were statistically less than those of untreated mice; however,

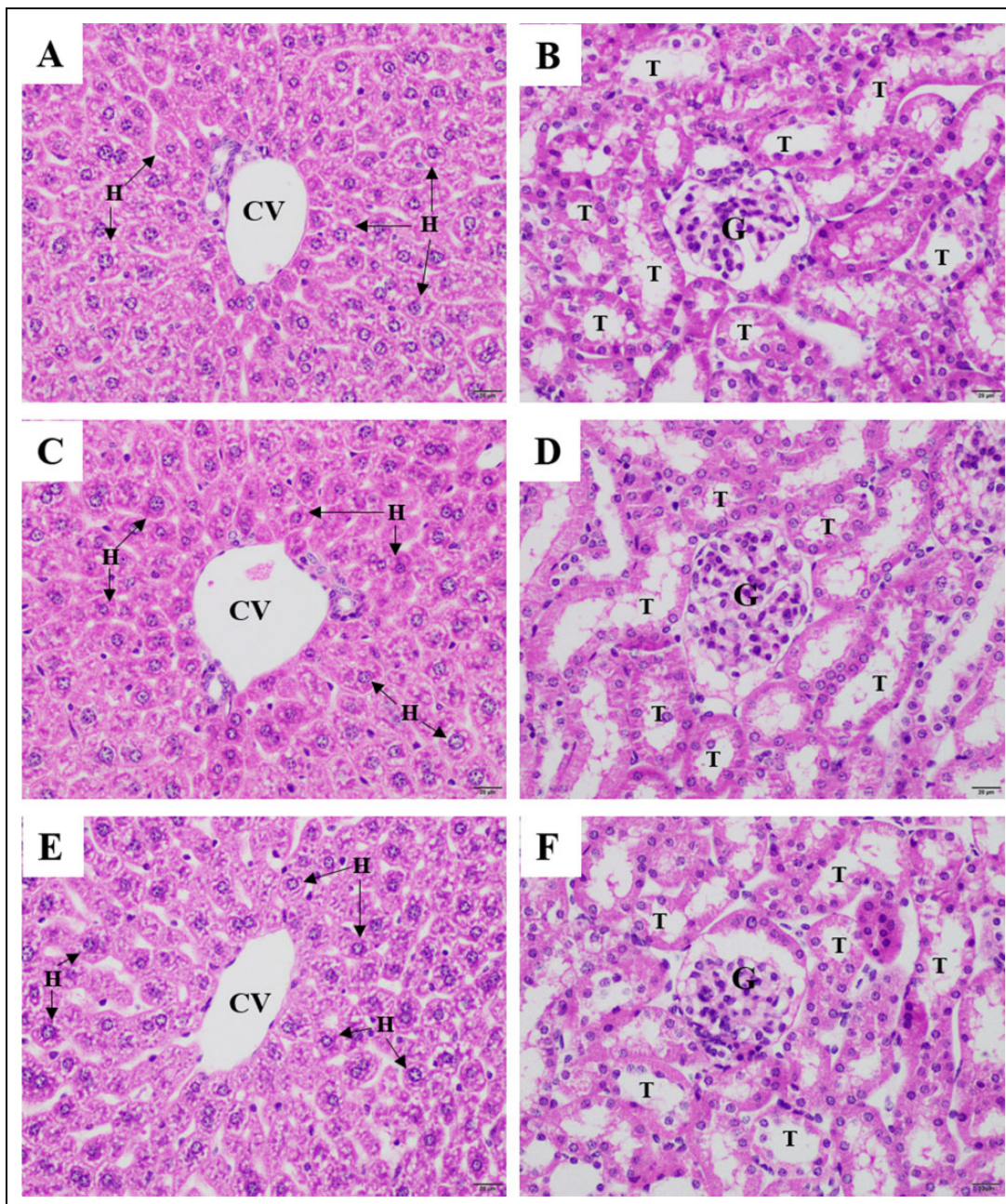


Figure 1. Representative image of H&E staining of the liver and kidneys from the untreated group (A) and (B), the negative control group (C) and (D), and the group treated with 2,000 mg/kg of the ethanolic *P. cablin* extract (E) and (F). All images were obtained at 400× magnification. Bars = 20 µm. T: tubules, G: glomerulus, CV: central vein, H: hepatocytes.

this difference may not be clinically significant. In addition, analysis of the histological changes in the kidneys and livers of mice treated with the extract and 7% Tween 80 showed normal features compared to those of control mice. Moreover, the body weights of the treated mice were not different from those of the untreated mice. The observed safety of *P. cablin* was consistent with a previous study reporting the safety of *P. cablin* administration with a maximal dose tolerance of 12.5 g/kg.⁴³ Our study confirmed that the ethanolic *P. cablin* extract exhibited antimalarial activity and no toxic effect on mice at a dose of

2000 mg/kg body weight. Therefore, our study suggests that further work is required to characterize the bioactive molecules in the ethanolic *P. cablin* extract that are responsible for the activity against malaria parasites.

Conclusions

This is the first report of the *in vitro* and *in vivo* antimalarial activities of *P. cablin* extracts. The ethanolic *P. cablin* extract possessed antimalarial activity against the malaria parasites *P.*

falciparum K1 and *P. berghei* ANKA. This study supports the safety of *P. cablin* as a medicinal plant with no toxicity *in vitro* and *in vivo*. Further phytochemical studies on the ethanolic *P. cablin* extract are recommended to identify the bioactive constituents that can be developed as antimalarial agents.

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Author Contributions

AP, PC, NH and CP designed the study; AP, PC, NH, SM, KK and CP carried out the experiments; AP drafted the manuscript; PC, NH and CP conducted experiments; AP, SM and KK analyzed the data; PC and CP revised the manuscript. All authors read and approved the final manuscript.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Ethical Approval

Ethical clearance was obtained from the Ethics Committee on Human Rights Related to Research Involving Human Subjects, Walailak University, prior to the recruitment of any participants (Approval number: WUEC-19-122-01). Written informed consent was obtained from all participants before data and sample collection. The animal use protocol was reviewed and approved by the Animal Ethics Committee, Walailak University (Protocol number: WU-AICUC-63001).

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ORCID iD

Chuchard Punsawad, PhD  <https://orcid.org/0000-0003-4826-9474>

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