

Identification of long non-coding RNA signature for paclitaxel-resistant patients with advanced ovarian cancer

Luqing Wang^{1,*}, Yanjun Hu^{2,*}, Xiaohong Xiang³, Kai Qu³ and Yue Teng⁴

¹Department of Nuclear Medicine, Liaocheng People's Hospital, Taishan Medical College, Liaocheng 252000, China

²Department of Clinical Laboratory, Liaocheng People's Hospital, Taishan Medical College, Liaocheng 252000, China

³Department of Hepatobiliary Surgery, The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710061, China

⁴Department of Obstetrics and Gynecology, The First Affiliated Hospital of Xi'an Jiaotong University, Xi'an 710061, China

*These authors have contributed equally to this work

Correspondence to: Yue Teng, email: navimoon@126.com

Kai Qu, email: joanne8601@163.com

Keywords: long non-coding RNA, epithelial ovarian cancer, paclitaxel, chemoresistance

Received: May 07, 2017

Accepted: June 19, 2017

Published: August 02, 2017

Copyright: Wang et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License 3.0 (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Ovarian cancer is the most lethal gynecologic malignancy, characterized by late diagnosis, frequent relapse, and easy development of chemoresistance. Recent studies suggest that lncRNAs are involved in ovarian cancer onset and progression, as well as the resistance in paclitaxel-containing chemotherapy. However, the genome-wide expression pattern and associated functional implications of lncRNAs in paclitaxel-resistant ovarian cancer cells remain undetermined. In the present study, we identified a panel of lncRNAs aberrantly expressed in both paclitaxel resistant ovarian cancer tissues and cell lines, including XR_948297, XR_947831, XR_938728, XR_938392, NR_103801, NR_073113, and NR_036503. Moreover, the seven-lncRNA signature showed a relatively high predictive accuracy of chemoresistance with an area under the ROC curve (AUC) of 0.93, and was associated with progression-free survival in ovarian cancer patients (HR=2.05, $p=0.015$). Our function prediction demonstrated that the seven-lncRNA signature was positively correlated with a cluster containing 129 genes enriched in insulin secretion-related pathway. Our findings suggest that the seven-lncRNA signature may be utilized as potent biomarkers for predicting chemoresistance for ovarian cancer patients with paclitaxel-containing chemotherapy.

INTRODUCTION

Epithelial ovarian cancer (EOC), accounting for the first place in mortality rate of all gynecologic cancers [1], still has a poor prognosis, with a 5-year survival rate lower than 30%, despite the amelioration of modern therapeutic methods [2]. The high mortality rate of EOC is mainly on account of the difficulty in early diagnosis, metastasis, and chemoresistance [3]. Combination of cytoreductive surgery and post-operational chemotherapy is the current standard treatment for advanced EOC.

However, more than 70% patients exhibit chemoresistance after surgery, leading to adverse effects to subsequent curative results and survival [4]. The mechanistic research on chemoresistance remains clinically challenging, simply because it holds back a successful long-term efficacy of EOC [5, 6]. Currently, the paclitaxel and platinum-containing chemotherapy is the main scheme for dealing with advanced EOC. Compared with platinum resistance which has been widely studied in EOC, much less mechanism of paclitaxel resistance has been clarified. Efforts in exploring molecular mechanism of paclitaxel

resistance will definitely enhance the clinical treatment as well as improve the survival rate of EOC patients.

Non-coding RNAs are drawing increasing attention in recent years for their pivotal regulating functions in various biological and pathological processes. As an important member of the non-coding RNAs, long non-coding RNAs (lncRNAs) have been identified as versatile players in multiple diseases by regulating target genes expression, directly either on transcripts originating from the same locus as the lncRNA itself (cis-acting) or on target transcripts originating on other loci (trans-acting) [7]. Numerous evidences reveal that they are significant regulators of various biological processes including immunoreaction, tumorigenesis, genetic modification, and metabolism regulation [8-11], and play pivotal roles in various cellular functions, disease occurrence and progression, including drug resistance [12]. To date, several lncRNAs have been found to mediate drug resistance. For example, studies revealed the harmful effects of a specific lncRNA, named homeobox transcript anti-sense RNA (HOTAIR), with an increasing prevalence of multiple neoplasm chemoresistance phenotypes [13-15]. Huang et al. identified that certain lncRNA could inhibit the transcription of tumor-suppressor genes by regulating protein activity to promote drug resistance in tumor cells [16]. In addition, an eight-lncRNA signature was found to be eligible for classifying patients with poor and improved overall survival rate [17]. Noticeably, two lncRNAs, RP11-284N8.3.1 and AC104699.1.1, have been recently demonstrated to be potential prognosis indicators of EOC patients by lncRNA-mRNA co-regulation network [18]. Collectively, these results confirmed the involvement of lncRNAs in the occurrence and progression of cancer and raised the possibility that lncRNAs could be used as biomarkers for diagnosis and prognosis as well as novel treatment targets [19, 20]. However, so far there is no research available on whether lncRNAs play a role in paclitaxel resistance of EOC. We believe that identifying a paclitaxel resistance-specific signature of lncRNAs in EOC and elaborating the latent molecular mechanisms will broaden our understanding of human EOC and provide future clinical approaches to treating this disease.

The present study aims to demonstrate whether lncRNAs play a vital role in paclitaxel-containing chemotherapies of EOC patients and to assess the prognostic values of identified lncRNAs. Furthermore, we also attempt to explore the underlying molecular mechanisms and involved pathways in paclitaxel resistance of EOC patients.

RESULTS

Identification of differentially expressed lncRNAs from the datasets

The GSE30161, a gene expression profile consisting of 46 patient samples, was used for the detection of lncRNAs related to incomplete response (IR) and complete response

(CR) of EOC patients. With analyzing package GEO2R, we identified a total of 50 up-regulated genes and 181 down-regulated genes ($p < 0.05$ and $FC > 1.2$) (Figure 1A), including both mRNAs (35 up-regulated mRNAs and 114 down-regulated mRNAs) and lncRNAs (15 up-regulated lncRNAs and 67 down-regulated lncRNAs) from GSE30161 (Figure 1B and 1C). Then, we analyzed GSE54772, another gene expression profile consisting of samples of 2 paclitaxel-resistant ovarian cancer cell lines as well as 2 paclitaxel-sensitive ovarian cancer cell lines, and found approximately 400 differentially expressed genes between the two groups ($p < 0.05$ and $FC > 2$ were considered statistically significant) (Figure 2A). We further analyzed the combination of GSE30161 and GSE54772 and identified a panel of seven lncRNAs (XR_948297, XR_947831, XR_938728, XR_938392, NR_103801, NR_073113, and NR_036503) that were significantly correlated with EOC patients' chemotherapeutic sensitivity and efficacy (Figure 2B). Distribution patterns of the seven enrolled lncRNAs in different sample groups were presented (Figure 2C).

Risk assessment and predictive value

We assessed the risk score on the basis of the expression levels of seven enrolled lncRNAs between the two patient groups (IR vs. CR) (Figure 3). Besides the relevance of sensitivity to EOC chemotherapy, the relationships between the identified seven lncRNAs and chemotherapeutic effects were also explored. ROC analysis was performed to assess the predictive accuracy of each individual lncRNA, respectively (Figure 4). Furthermore, we also analyzed the predictive accuracy of the combined seven-lncRNA signature. The seven lncRNAs were integrated into risk score to predict the risk of tumor recurrence of EOC patients receiving paclitaxel-containing chemotherapy. The IR group showed a higher risk score than the CR group ($p < 0.0001$) (Figure 5A). Compared with each individual lncRNA signature, the combination of all 7-identified-lncRNA signature showed a much better performance testified by a significantly higher AUC value (AUC = 0.93, 95% CI = 0.86 - 1.00) (Figure 5B).

Prognostic value of the combined seven-lncRNA signature

According to the risk score, patients in the dataset GSE30161 were further divided into low-score and high-score groups, with the median risk score taken as the cut-off point (Figure 5C). As presented in Figure 5C, our results showed that patients with low risk scores always had a longer progression-free survival (PFS) time. In contrast, the overall survival (OS) time between high- and low- risk groups did not reach a significant difference. In order to confirm whether the seven-lncRNA expression signature was an independent predictor of ovarian patients' sensitivity to platinum-based paclitaxel chemotherapy, the Kaplan-Meier plotter analysis

was conducted. The patients with a high risk score showed a shorter OS time, compared with their counterparts in the low risk group (MST = 40.40 vs 50.03 months). However, the association between seven-lncRNA expression signature and OS did not reach a significant difference (HR = 1.29, 95% CI = 0.63-2.64, $p = 0.490$). Interestingly, when analyzing the association between seven-lncRNA expression signature

and PFS, we found that patients with higher risk score had a shorter PFS time (MST = 10.30 vs 15.13 months) with a significant LogRank p -value ($R = 2.05$, 95% CI = 1.20-4.23, $p = 0.015$) (Figure 5D). Together, these results showed that the combined seven-lncRNA signature is an independent predictor of ovarian patients' sensitivity to platinum-based/paclitaxel-containing chemotherapy.

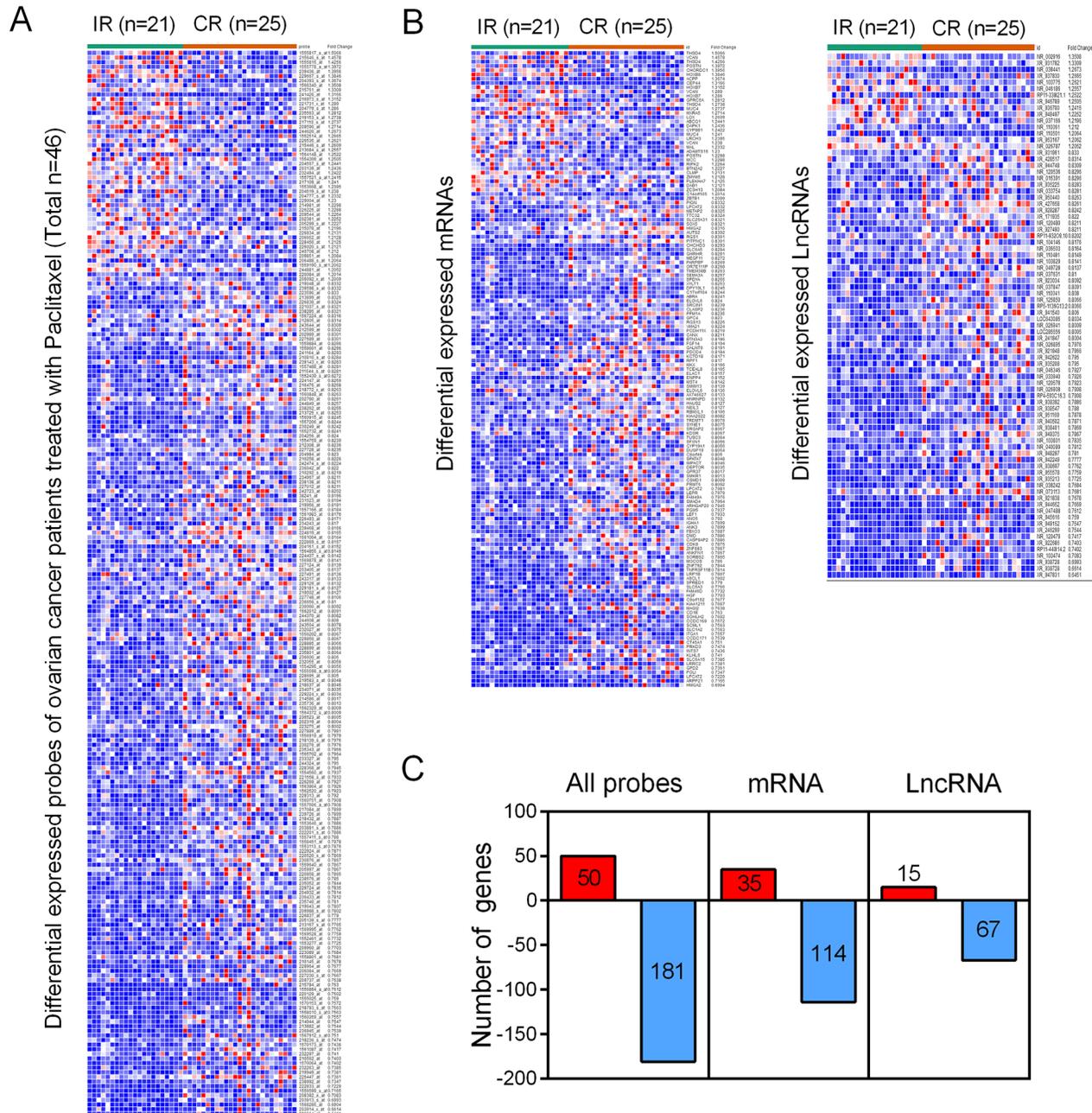


Figure 1: Identification of differentially expressed genes from the EOC dataset GSE30161. (A) A total of 231 differentially expressed genes were picked up from EOC patients treated with paclitaxel. (B) Within the 231 differentially expressed genes, there were both mRNAs (35 up-regulated mRNAs and 114 down-regulated mRNAs) and lncRNAs (15 up-regulated lncRNAs and 67 down-regulated lncRNAs). (C) A diagrammatic sketch demonstrating the specific pattern of differentially expressed genes from GSE30161. IR: incomplete response; CR: complete response.

Function annotation of seven-lncRNA signature

To predict the biological function of the seven identified lncRNAs, we performed WGCNA based on the mRNA and lncRNA profiles derived from GSE30161 (Figure 6A). The co-expressed relationships between the expression levels of seven lncRNAs and genome-wide genes were investigated by module-trait relationship analysis (Figure 6B), and we identified 129 genes (pink module) with which the expression of seven lncRNAs were all significantly and positively correlated (Figures 6B and 7A). Then we made use of Gene Ontology (GO) analysis and KEGG analysis to analyze the potential biological functions of the 129 picked-up genes. The results revealed these genes are closely involved in biological processes including insulin secretion, regulation of exocytosis, cyclic Adenosine monophosphate (cAMP)-mediated signaling, synapse assembly, calcium ion regulated exocytosis (Figure 7B), and cellular components such as receptor complex, nucleoplasm and membrane (Figure 7C). KEGG pathway enrichment analysis further suggested that those genes were associated with cancer and insulin secretion (Figure 7D). Interestingly, both GO and KEGG pathway enrichment analyses suggested the insulin secretion related pathways were involved in the predicted functions of the seven lncRNAs. Above findings provide evidence to explore molecular mechanism of paclitaxel resistance in EOC patients.

DISCUSSION

Most EOC patients respond well at the initial of chemotherapy, but quickly develop drug resistance, which finally leads to treatment failure. To date, there are still no effective methods to prevent drug resistance in EOC. To design new and more effective treatments, it is essential to understand the underlying molecular mechanism inducing drug resistance. In this study, a comprehensive analysis of lncRNA expression profiles in EOC patients was conducted. To the best of our knowledge, our study is the first to explore the association between lncRNAs expression signature and paclitaxel treatment in EOC patients.

So far, although increasing numbers of lncRNAs have been discovered and recorded in biological databases, most of their functions still remain unclear. Previous studies found aberrant lncRNA expressions could indicate the range of tumor progression and have a great power in the diagnosis and prognosis of cancer as novel independent molecular biomarkers [26, 27]. Certain lncRNA, such as HOTAIR, has been identified as the survival biomarker of EOC patients. However, whether lncRNA expression signatures could be used as predictors of paclitaxel chemo-sensitivity in EOC patients have not been determined. Qiao. et al. identified a lncRNA, CCAT1, regulates paclitaxel chemo-sensitivity in nasopharynx cancer cells by manipulating the miR-

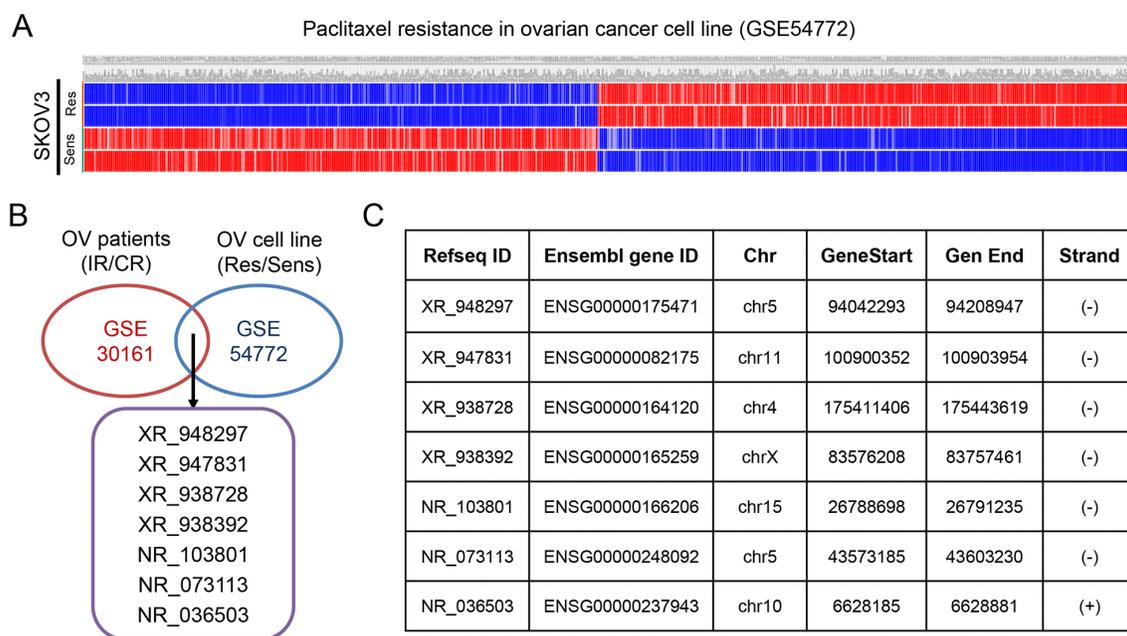


Figure 2: Identification of differentially expressed genes from the EOC dataset GSE54772. (A) Approximately 400 differentially expressed genes were picked up from paclitaxel-sensitive and paclitaxel-resistant EOC cell lines. (B) A panel of seven lncRNAs (XR_948297, XR_947831, XR_938728, XR_938392, NR_103801, NR_073113, and NR_036503) that were significantly correlated with EOC patients' chemotherapeutic sensitivity and efficacy were picked up when EOC dataset GSE30161 and GSE54772 were analyzed simultaneously. (C) Distribution patterns of the seven enrolled lncRNAs. OV: ovarian cancer; IR: incomplete response; CR: complete response; Res: resistant; Sens: sensitive.

Sample dendrogram and trait heatmap

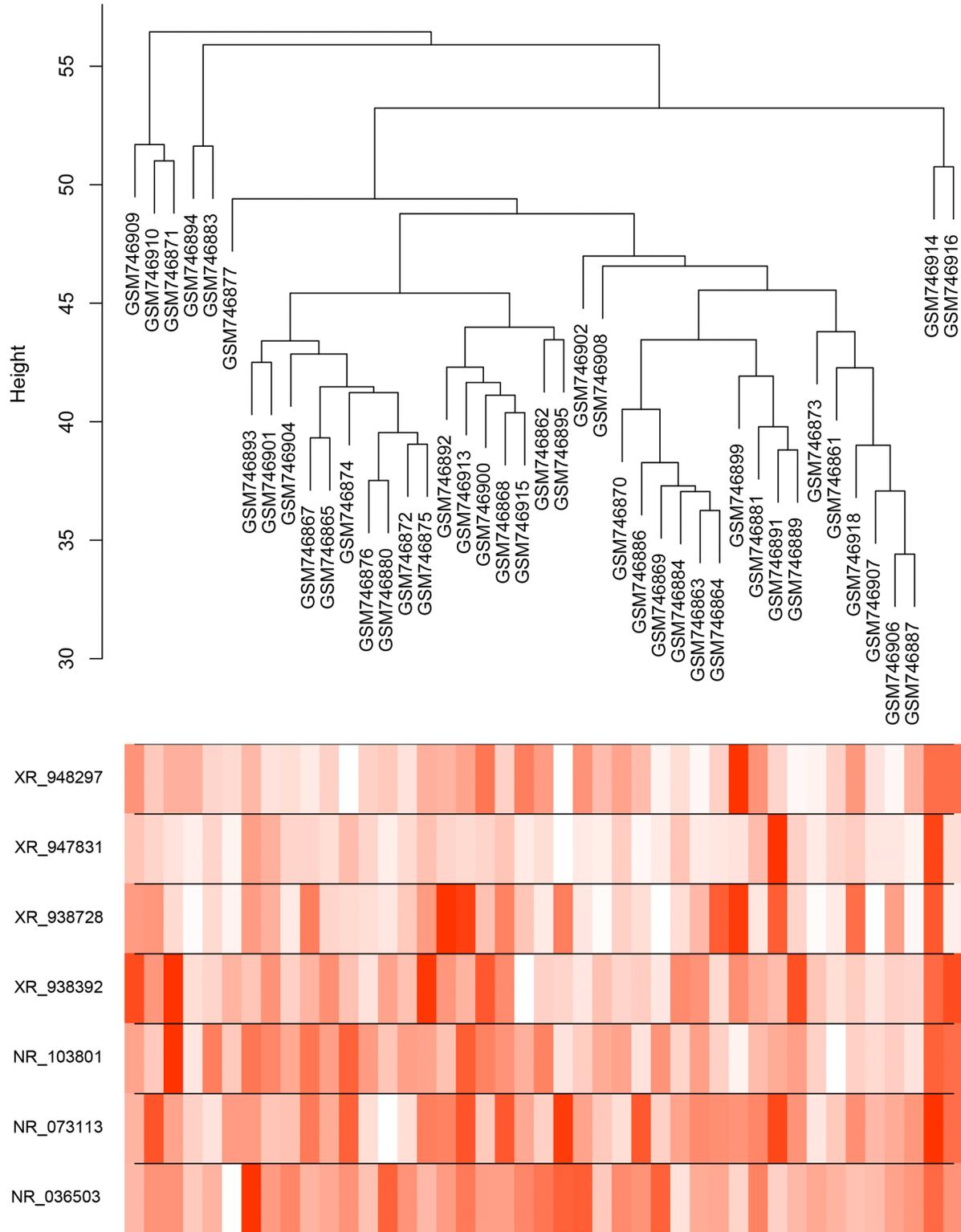


Figure 3: Dendrogram and trait heatmap of seven enrolled lncRNAs between IR and CR EOC patient groups. Risk scores were assessed on the basis of the expression levels of seven enrolled lncRNAs between IR and CR EOC patient groups.

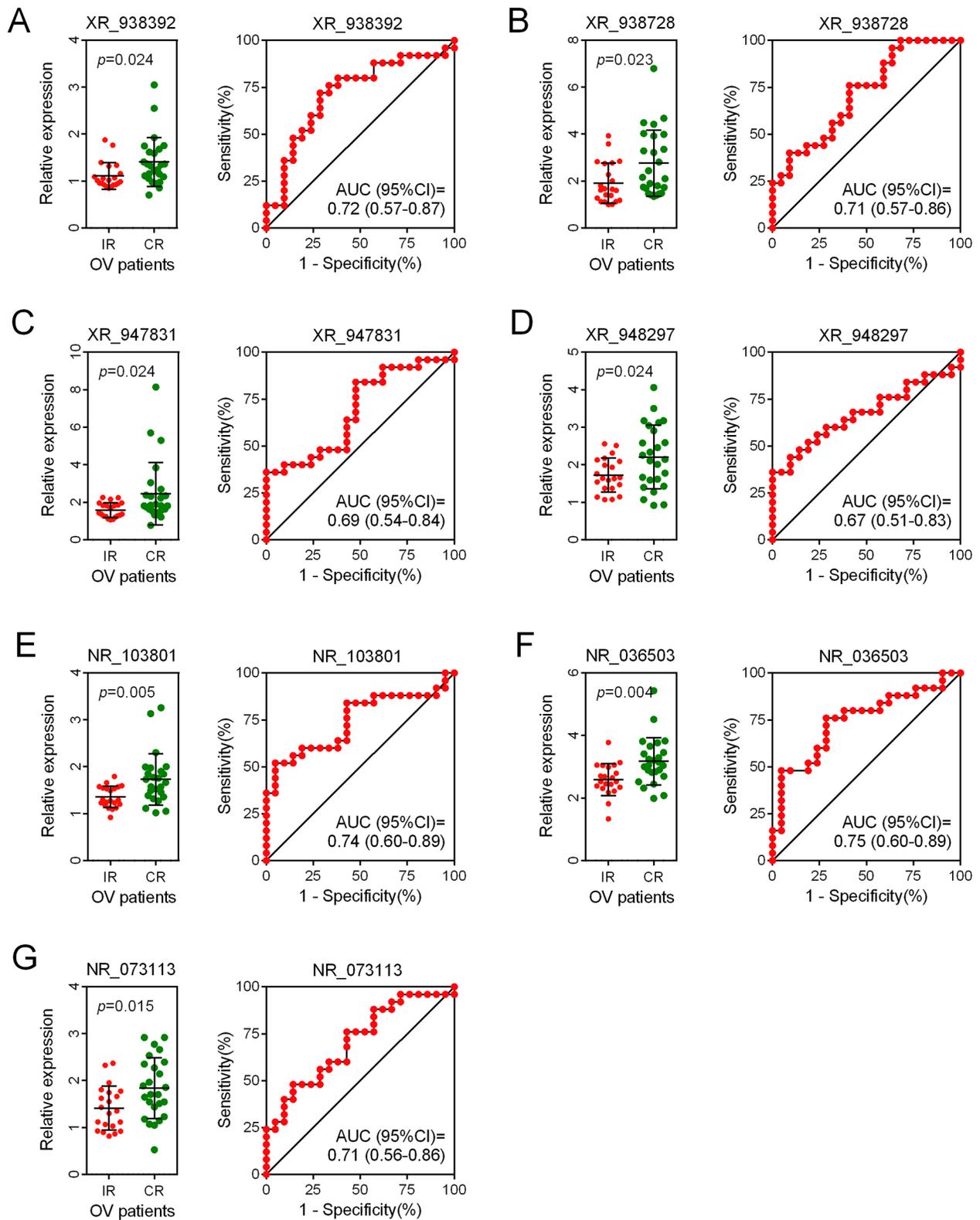


Figure 4: ROC analysis of the predictive accuracy of each individual lncRNA. (A-G) The predictive accuracy of each individual lncRNA of the seven enrolled lncRNAs was assessed by ROC analysis, respectively. OV: ovarian cancer.

181a/CPEB2 axis [28]. It is demonstrated that lncRNA H19 is an important element related to paclitaxel chemoresistance in the ER-positive breast carcinoma cells [29]. In our study, informatics prediction results showed that the genes associated with the seven lncRNAs (XR_948297,

XR_947831, XR_938728, XR_938392, NR_103801, NR_073113, and NR_036503) significantly participated in cancer pathways. The predictive capacity of the seven-lncRNA signature in chemotherapeutic sensitivity was identified via ROC analysis. Our findings revealed

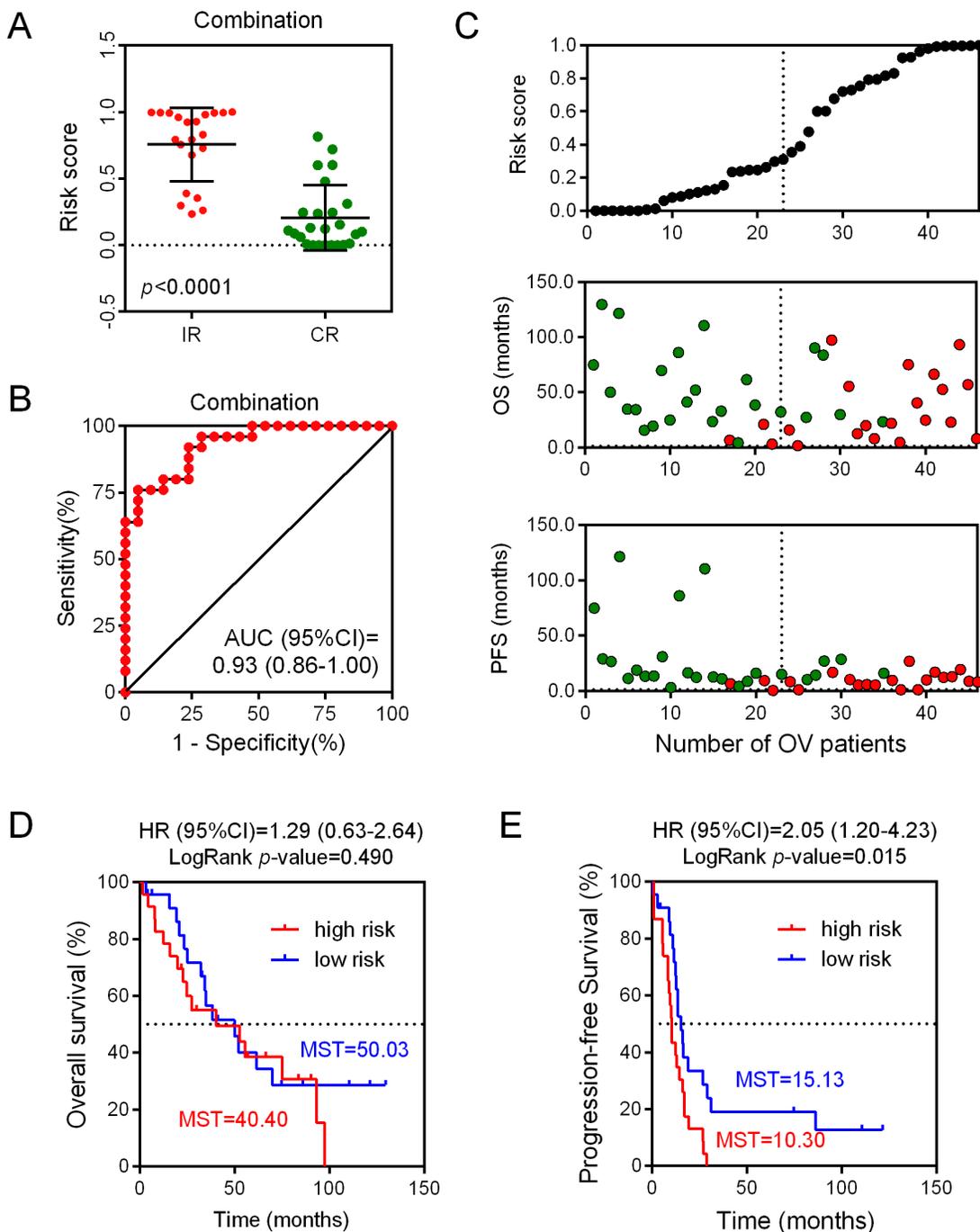


Figure 5: Predictive and prognostic value of each individual lncRNA as well as the combined seven-lncRNA signature. (A) Risk scores of the combined seven-lncRNA signature between IR and CR EOC patient groups. (B) Sensitivity analysis of the combined seven-lncRNA signature between IR and CR EOC patient groups. (C) Correlation between risk scores and progression-free survival, as well as overall survival. (D) The Kaplan-Meier plotter analysis of overall survival between low risk and high risk EOC patient groups on the basis of the combined seven-lncRNA signature. (E) The Kaplan-Meier plotter analysis of progression-free survival between low risk and high risk EOC patient groups on the basis of the combined seven-lncRNA signature. IR: incomplete response; CR: complete response; OS: overall survival; OV: ovarian cancer; PFS: progression-free survival; MST: mean survival time.

that the seven-lncRNA signature had a relatively high predictive accuracy of chemoresistance in EOC patients with paclitaxel-containing chemotherapy and was associated with patients' PFS time. Therefore, based on those findings as well as our analysis, we concluded the seven identified lncRNAs might be potential biomarkers for chemoresistance in EOC patients. Admittedly, more and further studies are in urgent need to confirm the connection between the seven-lncRNA signature and relevant pathways.

Biologically diverse mechanisms are involved in the development of chemoresistant phenotype in EOC. Among the various biological functions predicted by bioinformatics, insulin and Insulin-like growth factor (IGF) signaling pathways stood out from the crowd to be the top of the list. Actually, insulin and IGF signaling have long been discovered to regulate cellular growth, proliferation, metabolism, and survival [30]. Interestingly, chemo-resistant cells also displayed an enhanced proliferative response to insulin [31]. Insulin and insulin analogs are reported to be associated with a high proliferation rate and chemoresistance in patients with acute lymphoblastic leukemia [32]. Previous study found that high levels of insulin conferred AKT signaling activation and resistance to oxaliplatin in colon cancer

cell lines [33]. A recently published research suggests that stroma-derived IGFs can blunt the response to chemotherapy in pancreatic cancer via an IGF-insulin/IGF1R paracrine signaling axis [34]. A recent study also demonstrated that exogenous IGF-I increased cell growth in the ovarian cancer cell line OVCAR-3 in a manner equivalent to 10% FCS [35]. Hyperactivation of the IGF-IR and PI3K signaling pathways has been revealed as an essential event for cisplatin resistance in ovarian cancer cells [36]. Furthermore, PI3KCA is regarded as an oncogene in EOC onset and development, while PIK3R1 is constitutively activated via mutations [37, 38]. Together, these findings indicate a close interaction between chemoresistance and insulin and IGF signaling pathways in EOC.

Considering the regulatory relationship between the seven lncRNAs and prognosis of EOC patients, we further studied the prognostic value of the combination of these seven lncRNAs in EOC patients. As expected, survival analysis confirmed the significant association between the seven-lncRNA signature and EOC patients' PFS. However, we did not find a significant association between the seven-lncRNA signature and OS. The possible explanation might be that the seven lncRNAs were identified as paclitaxel-resistance-related biomarkers. Their expression levels

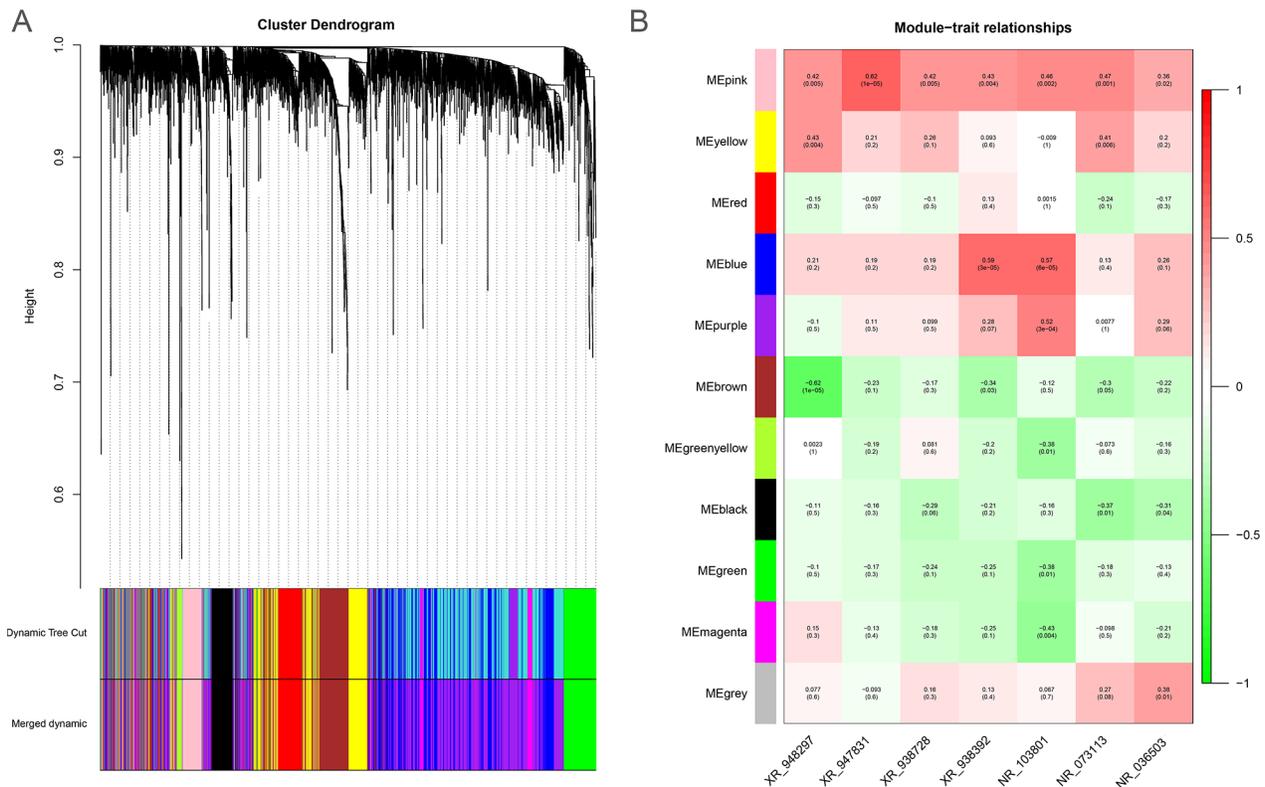


Figure 6: The WGCNA gene-coexpression analysis of differentially expressed genes from the EOC dataset GSE30161. (A) The cluster dendrogram differentially expressed mRNAs and lncRNAs derived from the EOC dataset GSE30161. **(B)** The module-trait relationship analysis of the co-expressed patterns between the seven lncRNAs and genome-wide genes. ME: module eigengenes.

could be used to predict chemoresistance but not death. Admittedly, the moderate sample size in our study limited the validity of some stratified analyses to some extent. Although the diagnostic power for these seven lncRNAs in combination is rather modest, our present study offers evidence for the future investigation on whether these seven lncRNAs could be used as prognostic markers in EOC management. Thereby, numerous prospective validation studies need to be performed before they are applied to clinical practice. Besides, another limitation of

our present study is the lack of serum levels of those seven lncRNAs. Since blood-based biomarkers are minimally invasive and more important for monitoring therapeutic effects of chemotherapy, We plan to explore the clinical values of serum lncRNAs in the next step.

In summary, we identified seven lncRNAs which were shown to be distinctively expressed in most ovarian tumor samples and significantly correlated with a poor chemotherapeutic response of EOC patients. Mechanistically, lncRNA promotes the malignant activities

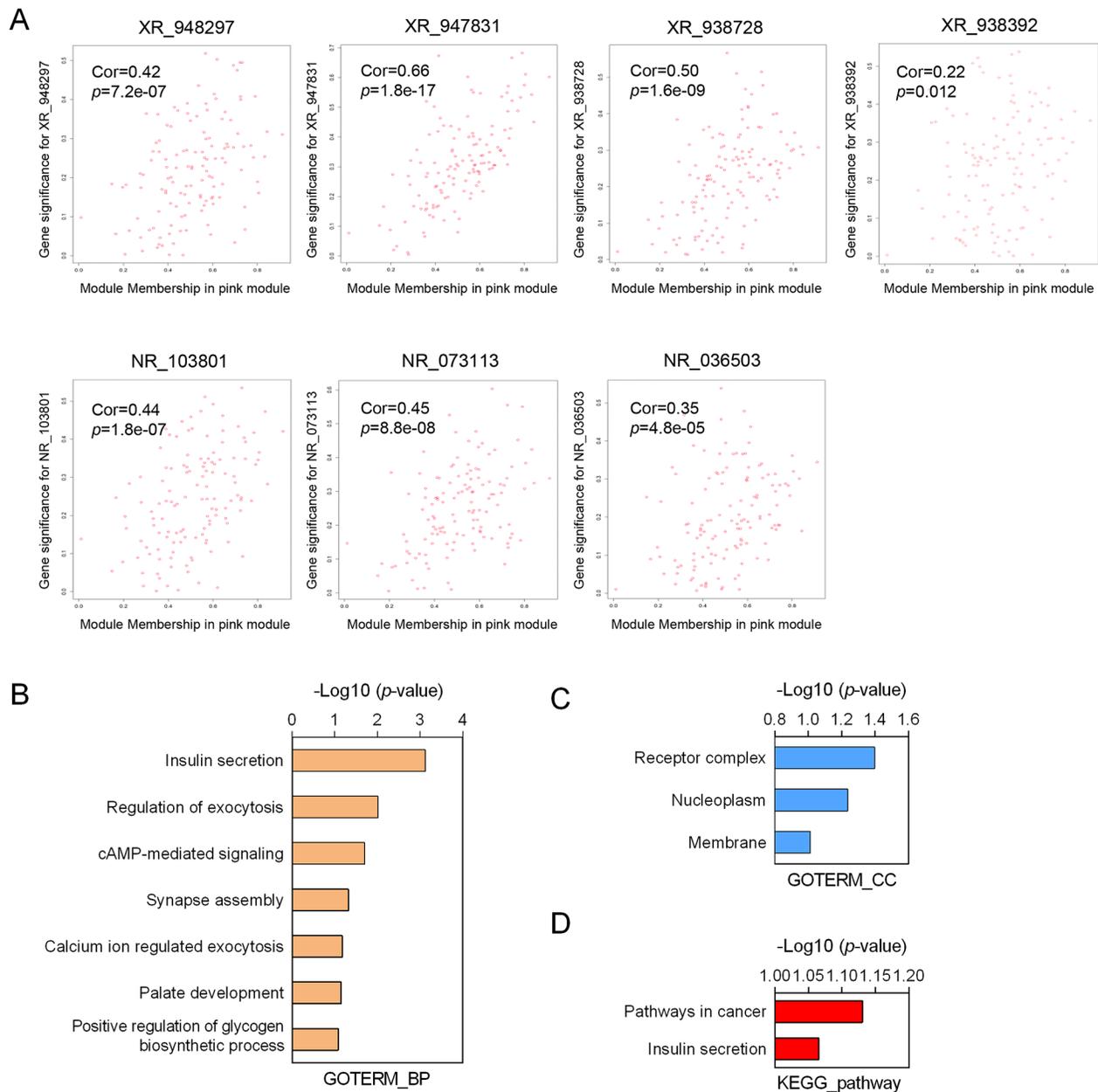


Figure 7: Function annotation of seven-lncRNA signature. (A) Correlation of each individual lncRNA with the differentially expressed genes from the EOC dataset GSE30161. (B-C) Gene Ontology analysis of biological processes (B) and cellular components (C) related to the 129-seven lncRNAs-co-expressed genes. (D) The KEGG pathway enrichment analysis of the predicted functions of the seven lncRNAs. GO: Gene Ontology.

of ovarian cancer cells via cancer-related pathways and insulin secretion. In addition, our study indicates a joint effect of lncRNA-gene markers in predicting the prognosis of EOC patients. Our data also shed light on the potential of the seven lncRNAs as novel therapeutic targets for future treatment of EOC.

MATERIALS AND METHODS

Microarray data

The Gene Expression Omnibus (GEO, <http://www.ncbi.nlm.nih.gov/geo>) is a public database available. Two gene expression profiles (GSE54772 and GSE30161) were obtained from GEO database. GSE54772 consisted of 2 paclitaxel-resistant ovarian cancer cell line samples and 2 paclitaxel-sensitive ovarian cancer cell line samples [21]. GSE30161 included 21 IR (incomplete response) ovarian cancer samples and 25 CR (complete response) ovarian cancer samples [22].

Weighted correlation network analysis (WGCNA)

A total of 3187 probe sets with 2730 corresponding differentially expressed genes were annotated to construct a co-expression network using the R package “WGCNA” according to previous reports [23]. The soft thresholding power was selected to 6 to produce a weighted network. The enrolled genes were hierarchically clustered based on their topological overlap (TO). Modules of clustered genes were then selected using the Dynamic Tree Cut algorithm. The module eigengenes (ME) were calculated by principal component analysis and clustered according to their correlation to quantify the co-expression similarity of entire modules.

Receiver operating characteristic analysis

In order to explore the possibility whether lncRNAs could be potential biomarkers for EOC diagnosis and prognosis prediction and assess the risk between two groups of patients, we did receiver operating characteristic (ROC) analysis. The area under the ROC curve (AUC) was used to evaluate the classification performance of the lncRNA signature according to their capability to distinguish IR patients from CR patients.

Pathway enrichment analysis

Gene ontology analysis (GO) is the most common useful method of annotating genes and gene products and of identifying characteristic biological properties for high-throughput genome or transcriptome data [24]. Kyoto

Encyclopedia of Genes and Genomes (KEGG) is a tool about systematic analysis of gene functions, connecting genomic information with higher-order functional information [25]. We used GO analysis and KEGG analysis to identify the biology processes, cellular components and related pathways of the seven candidate lncRNAs-associated genes. $p < 0.05$ was considered as statistically significant.

Survival analysis

Survival analyses for high- and low- risk EOC patients were carried out based on prognostic data derived from GSE30161, respectively. The prognosis survival and progression free survival analysis for each eligible lncRNA and also the combination of all the seven candidate lncRNAs were performed, respectively.

Author contributions

Kai Qu and Yue Teng: Designed the research and wrote the paper; Yanjun Hu and Luqing Wang: Collected and analyzed data; Kai Qu: Constructed figures; Xiaohong Xiang and Yue Teng: Drafted and revised the manuscript.

CONFLICTS OF INTEREST

The authors declare no potential conflicts of interest.

GRANT SUPPORT

This work was supported by National Science Foundation of China (Nos. 81201549, 81402313 and 81602374); the Fundamental Research Funds for the Central Universities (Nos. 2016qngz05 and xjj2016111); the Clinical Research Award of the First Affiliated Hospital of Xi'an Jiaotong University, China (No. XJTU1AF-CRF-2015-011).

REFERENCES

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. *CA Cancer J Clin.* 2016; 66:7-30.
2. Harter P, Gershenson D, Lhomme C, Lecuru F, Ledermann J, Provencher DM, Mezzanzanica D, Quinn M, Maenpaa J, Kim JW, Mahner S, Hilpert F, Baumann K, et al. Gynecologic Cancer InterGroup (GCIg) consensus review for ovarian tumors of low malignant potential (borderline ovarian tumors). *Int J Gynecol Cancer.* 2014; 24:S5-S8.
3. Jayson GC, Kohn EC, Kitchener HC, Ledermann JA. Ovarian cancer. *Lancet.* 2014; 384:1376-1388.
4. Huang KC, Yang J, Ng MC, Ng SK, Welch WR, Muto MG, Berkowitz RS, Ng SW. Cyclin A1 expression and paclitaxel resistance in human ovarian cancer cells. *Eur J Cancer.* 2016; 67:152-163.

5. Zhan Q, Wang C, Ngai S. Ovarian cancer stem cells: a new target for cancer therapy. *Biomed Res Int.* 2013; 2013:916819.
6. Vergote I, Trope CG, Amant F, Kristensen GB, Ehlen T, Johnson N, Verheijen RH, van der Burg ME, Lacave AJ, Panici PB, Kenter GG, Casado A, Mendiola C, et al. Neoadjuvant chemotherapy or primary surgery in stage IIIC or IV ovarian cancer. *N Engl J Med.* 2010; 363:943-953.
7. Ayers D. Long non-coding RNAs: novel emergent biomarkers for cancer diagnostics. *J Cancer Res Treat.* 2013; 1:31-35.
8. Alvarez-Dominguez JR, Hu W, Gromatzky AA, Lodish HF. Long noncoding RNAs during normal and malignant hematopoiesis. *Int J Hematol.* 2014; 99:531-541.
9. Ballarino M, Cazzella V, D'Andrea D, Grassi L, Bisceglie L, Cipriano A, Santini T, Pinnaro C, Morlando M, Tramontano A, Bozzoni I. Novel long noncoding RNAs (lncRNAs) in myogenesis: a miR-31 overlapping lncRNA transcript controls myoblast differentiation. *Mol Cell Biol.* 2015; 35:728-736.
10. Clark MB, Mattick JS. Long noncoding RNAs in cell biology. *Semin Cell Dev Biol.* 2011; 22:366-376.
11. Gallagher PG. Long noncoding RNAs in erythropoiesis. *Blood.* 2014; 123:465-466.
12. Gupta RA, Shah N, Wang KC, Kim J, Horlings HM, Wong DJ, Tsai MC, Hung T, Argani P, Rinn JL, Wang Y, Brzoska P, Kong B, et al. Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. *Nature.* 2010; 464:1071-1076.
13. Fang S, Gao H, Tong Y, Yang J, Tang R, Niu Y, Li M, Guo L. Long noncoding RNA-HOTAIR affects chemoresistance by regulating HOXA1 methylation in small cell lung cancer cells. *Lab Invest.* 2016; 96:60-68.
14. Liu Z, Sun M, Lu K, Liu J, Zhang M, Wu W, De W, Wang Z, Wang R. The long noncoding RNA HOTAIR contributes to cisplatin resistance of human lung adenocarcinoma cells via downregulation of p21(WAF1/CIP1) expression. *PLoS One.* 2013; 8:e77293.
15. Ozes AR, Miller DF, Ozes ON, Fang F, Liu Y, Matei D, Huang T, Nephew KP. NF- κ B-HOTAIR axis links DNA damage response, chemoresistance and cellular senescence in ovarian cancer. *Oncogene.* 2016; 35:5350-5361.
16. Huang JF, Guo YJ, Zhao CX, Yuan SX, Wang Y, Tang GN, Zhou WP, Sun SH. Hepatitis B virus X protein (HBx)-related long noncoding RNA (lncRNA) down-regulated expression by HBx (Dreh) inhibits hepatocellular carcinoma metastasis by targeting the intermediate filament protein vimentin. *Hepatology.* 2013; 57:1882-1892.
17. Zhou M, Sun Y, Sun Y, Xu W, Zhang Z, Zhao H, Zhong Z, Sun J. Comprehensive analysis of lncRNA expression profiles reveals a novel lncRNA signature to discriminate nonequivalent outcomes in patients with ovarian cancer. *Oncotarget.* 2016; 7:32433-32448. <https://doi.org/10.18632/oncotarget.8653>.
18. Guo Q, Cheng Y, Liang T, He Y, Ren C, Sun L, Zhang G. Comprehensive analysis of lncRNA-mRNA co-expression patterns identifies immune-associated lncRNA biomarkers in ovarian cancer malignant progression. *Sci Rep.* 2015; 5:17683.
19. Li P, Xue WJ, Feng Y, Mao QS. Long non-coding RNA CASC2 suppresses the proliferation of gastric cancer cells by regulating the MAPK signaling pathway. *Am J Transl Res.* 2016; 8:3522-3529.
20. Meseure D, Drak Alsibai K, Nicolas A, Bieche I, Morillon A. Long noncoding RNAs as new architects in cancer epigenetics, prognostic biomarkers, and potential therapeutic targets. *Biomed Res Int.* 2015; 2015:320214.
21. Ingemarsdotter CK, Tookman LA, Browne A, Pirlo K, Cutts R, Chelela C, Khurram KF, Leung EY, Dowson S, Webber L, Khan I, Ennis D, Syed N, et al. Paclitaxel resistance increases oncolytic adenovirus efficacy via upregulated CAR expression and dysfunctional cell cycle control. *Mol Oncol.* 2015; 9:791-805.
22. Ferriss JS, Kim Y, Duska L, Birrer M, Levine DA, Moskaluk C, Theodorescu D, Lee JK. Multi-gene expression predictors of single drug responses to adjuvant chemotherapy in ovarian carcinoma: predicting platinum resistance. *PLoS One.* 2012; 7:e30550.
23. Xu X, Zhou Y, Miao R, Chen W, Qu K, Pang Q, Liu C. Transcriptional modules related to hepatocellular carcinoma survival: coexpression network analysis. *Front Med.* 2016; 10:183-190.
24. Gene Ontology Consortium. The Gene Ontology (GO) project in 2006. *Nucleic Acids Res.* 2006; 34:D322-D326.
25. Ogata H, Goto S, Sato K, Fujibuchi W, Bono H, Kanehisa M. KEGG: Kyoto encyclopedia of genes and genomes. *Nucleic Acids Res.* 1999; 27:29-34.
26. Meng J, Li P, Zhang Q, Yang Z, Fu S. A four-long non-coding RNA signature in predicting breast cancer survival. *J Exp Clin Cancer Res.* 2014; 33:84.
27. Hu Y, Chen HY, Yu CY, Xu J, Wang JL, Qian J, Zhang X, Fang JY. A long non-coding RNA signature to improve prognosis prediction of colorectal cancer. *Oncotarget.* 2014; 5:2230-2242. <https://doi.org/10.18632/oncotarget.1895>.
28. Wang Q, Zhang W, Hao S. lncRNA CCAT1 modulates the sensitivity of paclitaxel in nasopharynx cancers cells via miR-181a/CPEB2 axis. *Cell Cycle.* 2017; 16:795-801.
29. Si X, Zang R, Zhang E, Liu Y, Shi X, Zhang E, Shao L, Li A, Yang N, Han X, Pan B, Zhang Z, Sun L, Sun Y. lncRNA H19 confers chemoresistance in ER α -positive breast cancer through epigenetic silencing of the pro-apoptotic gene BIK. *Oncotarget.* 2016; 7:81452-81462. <https://doi.org/10.18632/oncotarget.13263>.
30. Lashinger LM, Rossi EL, Hursting SD. Obesity and resistance to cancer chemotherapy: interacting roles of inflammation and metabolic dysregulation. *Clin Pharmacol Ther.* 2014; 96:458-463.

31. Djiogue S, Nwabo Kamdje AH, Vecchio L, Kipanyula MJ, Farahna M, Aldebasi Y, Seke Etet PF. Insulin resistance and cancer: the role of insulin and IGFs. *Endocr Relat Cancer*. 2013; 20:R1-R17.
32. Pan J, Chen C, Jin Y, Fuentes-Mattei E, Velazquez-Tores G, Benito JM, Konopleva M, Andreeff M, Lee MH, Yeung SC. Differential impact of structurally different anti-diabetic drugs on proliferation and chemosensitivity of acute lymphoblastic leukemia cells. *Cell Cycle*. 2012; 11:2314-2326.
33. Chen J, Huang XF, Qiao L, Katsifis A. Insulin caused drug resistance to oxaliplatin in colon cancer cell line HT29. *J Gastrointest Oncol*. 2011; 2:27-33.
34. Ireland L, Santos A, Ahmed MS, Rainer C, Nielsen SR, Quaranta V, Weyer-Czernilofsky U, Engle DD, Perez-Mancera PA, Coupland SE, Taktak A, Bogenrieder T, Tuveson DA, et al. Chemoresistance in pancreatic cancer is driven by stroma-derived insulin-like growth factors. *Cancer Res*. 2016; 76:6851-6863.
35. Yee D, Morales FR, Hamilton TC, Von Hoff DD. Expression of insulin-like growth factor I, its binding proteins, and its receptor in ovarian cancer. *Cancer Res*. 1991; 51:5107-5112.
36. Eckstein N, Servan K, Hildebrandt B, Politz A, von Jonquieres G, Wolf-Kummeth S, Napierski I, Hamacher A, Kassack MU, Budczies J, Beier M, Dietel M, Royer-Pokora B, et al. Hyperactivation of the insulin-like growth factor receptor I signaling pathway is an essential event for cisplatin resistance of ovarian cancer cells. *Cancer Res*. 2009; 69:2996-3003.
37. Levine DA, Bogomolny F, Yee CJ, Lash A, Barakat RR, Borgen PI, Boyd J. Frequent mutation of the PIK3CA gene in ovarian and breast cancers. *Clin Cancer Res*. 2005; 11:2875-2878.
38. Shayesteh L, Lu Y, Kuo WL, Baldocchi R, Godfrey T, Collins C, Pinkel D, Powell B, Mills GB, Gray JW. PIK3CA is implicated as an oncogene in ovarian cancer. *Nat Genet*. 1999; 21:99-102.