Effect of sertraline on proliferation and neurogenic differentiation of human adipose-derived stem cells

Shahnaz Razavi, Maliheh Jahromi, Nushin Amirpour, Zahra Khosravizadeh

Department of Anatomical Sciences and Molecular Biology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, Iran

Abstract Background: Antidepressant drugs are commonly employed for anxiety and mood disorders. Sertraline is extensively used as antidepressant in clinic. In addition, adipose tissue represents an abundant and accessible source of adult stem cells with the ability to differentiate in to multiple lineages. Therefore, human adipose-derived stem cells (hADSCs) may be useful for autologous transplantation.

Materials and Methods: In the present study, we assessed the effect of antidepressant drug Sertraline on the proliferation and neurogenic differentiation of hADSCs using MTT assay and immunofluorescence technique respectively.

Results: MTT assay analysis showed that 0.5 μ M Sertraline significantly increased the proliferation rate of hADSCs induced cells (*P* < 0.05), while immunofluorescent staining indicated that Sertraline treatment during neurogenic differentiation could be decreased the percentage of *glial fibrillary acidic protein* and Nestin-positive cells, but did not significantly effect on the percentage of MAP2 positive cells.

Conclusion: Overall, our data show that Sertraline can be promoting proliferation rate during neurogenic differentiation of hADSCs after 6 days post-induction, while Sertraline inhibits gliogenesis of induced hADSCs.

Key Words: Adipose-derived stem cells, Antidepressant drug, Neurogenic differentiation, proliferation, Sertraline

Address for correspondence:

Dr. Shahnaz Razavi, Department of Anatomical Sciences and Molecular Biology, School of Medicine, Isfahan University of Medical Sciences, Isfahan, 81744-176, Iran. E-mail: razavi@med.mui.ac.ir Received: 06.03.2013, Accepted: 23.05.2013

INTRODUCTION

Depression is the most prevalent mood disorder. It suffers about 21% of the world population^[1] and these disorders treat with antidepressants drugs.^[2]

Access this article online	
Quick Response Code:	
	website: www.advbiores.net
	DOI: 10.4103/2277-9175.129367

Several studies have shown that antidepressants increase hippocampal neurogenesis in both animals and humans^[3-5] and increase neuronal cell proliferation *in vitro*.^[1,6]

Sertraline is an antidepressant drug from selective inhibitor of neuronal serotonin reuptake group.^[7-10] Sertraline in the treatment of major depressive disorders^[11] such as obsessive compulsive disorder^[12] panic disorder^[13] and post-traumatic stress disorder^[14] is effective. This drug acted by elevating the concentration of monoamines,^[15] some dopamine reuptake inhibitors^[16] and inhibition of neuronal serotonin reuptake in the synaptic cleft of the central nervous system. Some reports showed that

Copyright: © 2014 Razavi. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

How to cite this article: Razavi S, Jahromi M, Amirpour N, Khosravizadeh Z. Effect of sertraline on proliferation and neurogenic differentiation of human adiposederived stem cells. Adv Biomed Res 2014;3:97. elevation of serotonin concentration in synaptic cleft cause to maintain mental balance.^[17] Furthermore, rapid increase of monoamines in extracellular levels requires several weeks or months.^[18,19]

Decrease of monoamine resulting a decline in hippocampal neurogenesis while elevation of serotonin and/or norepinephrine lead to increase in hippocampal neurogenesis, so this mechanism lead to consider Sertraline as an antidepressant in clinic.^[2,20]

Mesenchymal stem cells (MSCs) have been isolated from different sources including bone marrow, muscle and adipose tissue, these cells can also differentiated to chondrocytes, adipocytes, myoblasts and osteoblasts *in vitro*.^[21-23] and use for autologous transplantation.^[24,25]

Several factors limit the use of MSCs-derived bone marrow, including: Morbidity, painful procedure and gain a few number cells. HADSCs can be isolated more easily than the other type of MSCs have a significant number of cells from fat tissue and less invasive method; thus, adipose stem cells have important criteria in compare with other source.^[26]

It has been shown that human adipose-derived stem cells (hADSCs) can differentiate into several linage including: Chondrocyte, endothelial, adipocyte, cardiomyocyte^[26-28] and also neurogenic cells that show neuron-like morphology and express neural markers.^[28-31]

On the basis of these findings, it was interest to determine the influence of antidepressant drug sertraline on the proliferation rate and differentiation of hADSCs into neural lineage.

MATERIALS AND METHODS

Tissue collection and isolation of hADSCs

Samples of adipose tissue were obtained from three patients during abdominoplasty surgery, after receiving informed consent. HADSCs were isolated as previously described.^[32]

Briefly, adipose tissues were washed several time with sterile phosphate-buffered saline (PBS) to remove contaminating debris and red blood cells. Then adipose tissue was enzymatically dissociated with 0.075% collagenase in PBS for 30 min at 37°C. The collagenase was neutralized with an equal volume of Dulbecco's modified Eagle's medium: F12 (DMEM: F12)/10% fetal bovine serum (FBS) (Gibco, BRL, Paisley, UK) and centrifuged for 10 min at 1200 g. The cellular pellet was resuspended in growth medium (DMEM: F12/10% FBS and 1% penicillin/streptomycin solution). Cell cultures were maintained in T25 flasks for 4-5 days in a 37°C incubator with 5% $\rm CO_2$ until they confluent. Then, ADSCs were passaged at the ratio of 1:3.^{[32]}

All chemicals, except where specified otherwise, were purchased from Sigma-Aldrich (St. Louis, MO).

In this study, after isolated ADSC from human abdominal fat, these cells differentiated to neuronlike cells and simultaneously treated with Sertraline as an antidepressant drug. MTT assay and immunofluorescent staining were used for assessment of proliferation rate and evaluation of neural markers status using neurogenic differentiation of hADSCs.

Neurogenic differentiation of hADSCs

For neural induction, hADSCs passage 3-6 was used. HADSCs were plated in low attachment plastic tissue culture dishes in culture medium that contained: DMEM: F12, 2% B27, supplemented with 20 ng/ml basic fibroblast growth factor and 20 ng/ml epidermal growth factor. This media renewed every 2 days up to 6-7 days.

After replating the neurospheres dissociated cells on cover slips in a 24-well plate at a density of 2×10^4 /cm², incubated in neurobasal medium supplemented with 5% FBS, 1%l-glutamin, 1% none essential amino acids, 1% N2 supplement and 2% B27 for 1 week (the growth factors and supplements are all from Gibco BRL, Paisley, UK).^[32]

Depending on the purpose of the experiments, the cells were cultured in neural induction medium with or without of 0.5 μ M Sertraline in treated and control groups respectively for 7 days.

MTT assay

For assessment of cell viability and proliferation rate in Sertraline-treated neuronal precursor cells, MTT assay was performed at 2, 4 and 6 days after Sertraline treatment. Differentiated cells with PBS were washed; The MTT assay was performed as previously described.^[32] Briefly, 2×10^3 cells/well, were seeded on 96-well plates and grown in the presence of Sertraline at 0.5 μ M or absence of Sertraline. 100 μ l of DMEM and 10 μ l of a MTT solution (0.5 mg/ml) were added to each well and incubated for a 4 h. The MTT solution was removed from cell cultures and 100 μ l of dimethyl sulfoxide added to extract the MTT formazan. The absorbance of each well was measured by microplate reader at 540 nm.^[32]

Immunocytochemistry staining

The induced cells were fixed in 4% paraformaldehyde for 20 min at room temperature, after rinsed twice with PBS, the cells permeablized using PBS containing 2% Triton X-100 at room temperature for 30 min. primary antibodies diluted in blocking solution consistent 10% goat serum and 1 mg/ml bovine serum albumin (BSA) for 2 h at 37°C. Then, the cells were incubated with primary antibodies overnight at 4°C in the dark. After washing with PBS, the cells were incubated with fluorescein isothiocvanate (FITC) conjugated secondary antibodies for 2 h at 37°C. The differentiated cells were reacted with antibodies against mouse anti-Nestin (1:300, Abcam, Cambridge, MA, USA), mouse anti-microtubule-associated protein 2 ([MAP2], 1:300, Abcam, Cambridge, MA, USA), mouse anti-glial fibrillary acidic protein ([GFAP], 1:600, Abcam, Cambridge, MA, USA) and antimouse FITC-conjugated immunoglobulin antibody (1:500, Abcam, Cambridge, MA, USA). For quantitative analysis, the cells were incubated with 4, 6-diamidino-2-phenylindole ([DAPI] 1:1000) for 2 min at room temperature. Preparations were examined using a fluorescence microscope (Olympus BX51, Japan). The numbers of positive cells for each antigen were counted as percentages of the total DAPI-stained cell population. For quantitative assessment of cell differentiation in control and Sertraline treated cells, the relative numbers of cells expressing different markers like mature neurons (MAP2), astrocytes (GFAP) and neural progenitor cells (Nestin) were counted as percentages of the total DAPIstained cell population. Image J software was used for merging the pictures.^[32]

Statistical analysis

Cell proliferation and neural differentiation data were analyzed using one-way analysis of variance (ANOVA) (SPSS Inc., Chicago, IL). All data were shown as means \pm standard error of the mean. Experiments with two groups were subjected to a one-way ANOVA, P < 0.05 was taken as significant and P < 0.001was taken as highly significant to indicate levels of statistical significance.

RESULTS

Morphologic changes of hADSCs following neurogenic differentiation

To assessment changes in cell morphology following Sertraline treat, we analyzed morphology of neurogenic induced cells for 2, 4 and 6 days. Images were viewed using bright field and phase contrast microscopy (Nikon Eclipse TS100). After two or three passages hADSCs appeared by their spindle-shaped fibroblastic morphology [Figure 1a].

Then, cells were cultured in induction medium for 7 days, the cells floated in suspension as small aggregates (neurospheres) [Figure 1b]. After 9 days treated with Sertraline, the induced cells forming contracted cell bodies with long cytoplasmic processes [Figure 1c] and the cell bodies became bipolar and multipolar appearance on 10 day [Figure 1d].

Effect of Sertraline on proliferation rate

To investigate whether Sertraline affects the proliferation of isolated ADSCs, cell proliferation assay was carried out by MTT assy. The cells exposed to $0.5 \,\mu$ M Sertraline for 2, 4 and 6 days, there were no significant difference mean of absorbance of Sertraline in treated group as compared with the control group for 2 and 4 days post-induction. While, in compared with the control group, exposure to $0.5 \,\mu$ M Sertraline resulted in a significant increase of cell proliferation after 6 days (P = 0.02) [Figure 2].

Effects of Sertraline on neurogenic differentiation of hADSCs

Evaluation of the percentage of neural markers, Nestin, GFAP and MAP2 determined by immunofluorescence technique. [Figure 3] indicated that after treatment with 0.5 μ M sertraline the induced cells were not significant difference for MAP2 positive cells, in comparison with the control group, while the mean percentage of GFAP positive cells was significantly decreased in Sertraline treated group relative to the control group (P < 0.05).

Quantification of immunostaining revealed that $2.6 \pm 1.2\%$ of inducing treated cells were positive for Nestin, which was not significantly different from the control group ($10 \pm 3.7\%$). Immunostaining results showed the mean percentage of GFAP-positive cells was 2 ± 1.1 in Sertraline treated group compared with 32 ± 18 in the control group, which demonstrated that GFAP decreased significantly in treated group. Finally, the mean percentage of MAP 2 positive cells was 36 ± 28 near to the control group (38 ± 31) [Figure 4].

DISCUSSION

Previous studies have shown that antidepressants drugs increase neural cell proliferation and enhance differentiation in neural precursors derived from human embryonic stem cells.^[1,19,33] Antidepressants also increase proliferation and differentiation in hippocampal progenitor cell both *in vivo* and *in vitro*.^[5,17,34-37]

Several studies showed that hippocampus undergoes decrease of size and neuron numbers in stress condition.^[38-42] Therefore, commonly use of antidepressants are useful in depression.^[43,44]



Figure 1: Morphological changes of adipose-derived stem cells following differentiation with 0.5 μ M Sertraline *in vitro*. (a) Undifferentiated adipose-derived stem cells. (b) After 6-7 days, cultured in neurogenic induction medium. The cells were exhibited sphere shape. (c) After 9 days, the induced cells with 0.5 μ M sertraline forming contracted cell bodies with long cytoplasmic processes. (d) After 10 days, the cell bodies became bipolar and multipolar appearance



Figure 3: Immunocytochemical staining for specific markers in neurogenic differentiated cells (Netin, GFAP and MAP2) treated with 0.5 μ M Sertraline. In each experiment, the nuclei were counterstained with6-diamidino-2-phenylindole. Scale bar = 100 μ m

ADSCs have been defined on cells with the capacity to differentiated in to multiple cell lineage, furthermore, these cells can be extracted from donor by a safety procedure.^[45] Therefore, these cells can be the suitable candidate for neurogenesis in neurodegenerative diseases.^[46-48]

Previous investigate showed that Sertraline increased cell proliferation in the human hippocampus, also this study demonstrated that Sertraline could promotes differentiation of neural stem cells (NSCs), which dissociated from hippocampus to neurons, but inhibits its differentiation to glial cells.^[35]



Figure 2: Determination of the absorbance for neuronal precursors treated with 0.5 μ M Sertraline in order to assessment of proliferation rate at 540 nm in 2, 4 and 6 days after induction. Compared with the control group, exposure to 0.5 μ M Sertraline resulted in a significant increase of cell viability after 6 days (* $P \leq 0.05$)



Figure 4: The mean percentage of immunoreaction positive cells for Nestin, microtubule-associated protein2 (MAP2) and glial fibrillary acidic protein (GFAP) in neuronal precursors treated with 0.5 μ M Sertraline in compared with the control group. The mean percentage of MAP2-positive cells was close to control group. While, the mean percentage of Nestin-positive cells was decreased in the Sertraline treated group compared with the control group, but the mean percentage of GFAP positive cells in the treated group significantly decreased relative to the control group (***P \leq 0.001)

In vitro culture of NSCs from the hippocampus of fetal rats was demonstrated that sertraline could not increase proliferation and viability of NSCs. Furthermore, by decreasing the expression of proinflammatory cytokine, Sertraline might provide neuroprotection.^[17]

Sutcigil *et al.* showed that Sertraline treat might have decline in the proinflammatory cytokine *interleukin* (IL)-12 and elevate transforming growth factor beta (TGF- β 1) and anti-inflammatory cytokines IL-4, which can influence neurogenesis widely.^[49]

We showed that 0.5 μ M of Sertraline increased the proliferation rate. Our results were disagreement with the findings of Peng *et al.*,^[17] which showed that 1 μ M of Sertraline cannot effect on proliferation of cells in compare with the control group. They confirm that high concentrations of Sertraline (20 μ M and 50 μ M) inhibit the proliferation of ADSCs.

Previous studies showed that other antidepressants, such as fluoxetine $(1 \ \mu M)$,^[2,19,50] imipramine, venlafaxine $(1 \ \mu M)$ ^[2] and paroxetine $(1-5 \ \mu M)$ ^[37] increased the proliferation rate of with different sources.

Furthermore, Peng *et al.* demonstrated that Sertraline induces NSCs to differentiate into neurons, but inhibits its differentiation to glial cells. These different in findings may have resulted from differences in cell lines, concentration and type of drugs or the duration of treatment.^[2,17,19,37,50]

Our results could be due to Sertraline by decreasing proinflammatory cytokines induced cell proliferation and through TGF- β 1 could be inhibit gliogenesis.

Overall, results of our study show that sertraline can be enhancing proliferation rate during neurogenic differentiation of hADSCs. However, Sertraline treatment can be decreased glial markers (GFAP+) and neural progenitor cells (Nestin+) as compared with the control group, but there were no significant difference in the expression of mature neuron marker (MAP2+) between control and treated groups. However, the molecular mechanisms of Sertraline on ADSCs proliferation and differentiation are not known yet. Moreover, the mechanisms and factors effective of Sertraline function needs to be further investigated.

ACKNOWLEDGMENT

This study was supported by Isfahan University of Medical Sciences.

REFERENCES

- Chang EA, Beyhan Z, Yoo MS, Siripattarapravat K, Ko T, Lookingland KJ, et al. Increased cellular turnover in response to fluoxetine in neuronal precursors derived from human embryonic stem cells. Int J Dev Biol 2010;54:707-15.
- Cabras S, Saba F, Reali C, Scorciapino ML, Sirigu A, Talani G, et al. Antidepressant imipramine induces human astrocytes to differentiate into cells with neuronal phenotype. Int J Neuropsychopharmacol 2010; 13:603-15.
- 3. Duman RS. Role of neurotrophic factors in the etiology and treatment of mood disorders. Neuromolecular Med 2004;5:11-25.
- Wang JW, David DJ, Monckton JE, Battaglia F, Hen R. Chronic fluoxetine stimulates maturation and synaptic plasticity of adult-born hippocampal granule cells. J Neurosci 2008;28:1374-84.
- Boldrini M, Underwood MD, Hen R, Rosoklija GB, Dwork AJ, John Mann J, *et al.* Antidepressants increase neural progenitor cells in the human hippocampus. Neuropsychopharmacology 2009;34:2376-89.

Advanced Biomedical Research | 2014

- Chen SJ, Kao CL, Chang YL, Yen CJ, Shui JW, Chien CS, *et al.* Antidepressant administration modulates neural stem cell survival and serotoninergic differentiation through bcl-2. Curr Neurovasc Res 2007;4:19-2.
- Fava M, Judge R, Hoog SL, Nilsson ME, Koke SC. Fluoxetine versus sertraline and paroxetine in major depressive disorder: Changes in weight with long-term treatment. J Clin Psychiatry 2000;61:863-7.
- Lépine JP, Caillard V, Bisserbe JC, Troy S, Hotton JM, Boyer P. A randomized, placebo-controlled trial of sertraline for prophylactic treatment of highly recurrent major depressive disorder. Am J Psychiatry 2004;161:836-42.
- Schneider LS, Nelson JC, Clary CM, Newhouse P, Krishnan KR, Shiovitz T, *et al.* An 8-week multicenter, parallel-group, double-blind, placebo-controlled study of sertraline in elderly outpatients with major depression. Am J Psychiatry 2003;160:1277-85.
- Swenson JR, O'Connor CM, Barton D, Van Zyl LT, Swedberg K, Forman LM, *et al.* Influence of depression and effect of treatment with sertraline on quality of life after hospitalization for acute coronary syndrome. Am J Cardiol 2003;92:1271-6.
- Fann JR, Uomoto JM, Katon WJ. Sertraline in the treatment of major depression following mild traumatic brain injury. J Neuropsychiatry Clin Neurosci 2000;12:226-32.
- Greist J, Chouinard G, DuBoff E, Halaris A, Kim SW, Koran L, *et al.* Double-blind parallel comparison of three dosages of sertraline and placebo in outpatients with obsessive-compulsive disorder. Arch Gen Psychiatry 1995;52:289-95.
- Pohl RB, Wolkow RM, Clary CM. Sertraline in the treatment of panic disorder: A double-blind multicenter trial. Am J Psychiatry 1998;155:1189-95.
- Brady K, Pearlstein T, Asnis GM, Baker D, Rothbaum B, Sikes CR, *et al.* Efficacy and safety of sertraline treatment of posttraumatic stress disorder: A randomized controlled trial. JAMA 2000;283:1837-44.
- Schloss P, Henn FA. New insights into the mechanisms of antidepressant therapy. Pharmacol Ther 2004;102:47-60.
- 16. Hauser RA, Zesiewicz TA. Sertraline for the treatment of depression in Parkinson's disease. Mov Disord 1997;12:756-9.
- Peng ZW, Xue YY, Wang HN, Wang HH, Xue F, Kuang F, et al. Sertraline promotes hippocampus-derived neural stem cells differentiating into neurons but not glia and attenuates LPS-induced cellular damage. Prog Neuropsychopharmacol Biol Psychiatry 2012;36: 183-8.
- Wong ML, Licinio J. Research and treatment approaches to depression. Nat Rev Neurosci 2001;2:343-51.
- Kusakawa S, Nakamura K, Miyamoto Y, Sanbe A, Torii T, Yamauchi J, et al. Fluoxetine promotes gliogenesis during neural differentiation in mouse embryonic stem cells. J Neurosci Res 2010;88:3479-87.
- Brezun JM, Daszuta A. Depletion in serotonin decreases neurogenesis in the dentate gyrus and the subventricular zone of a dult rats. Neuroscience 1999;89:999-1002.
- Wakitani S, Saito T, Caplan AI. Myogenic cells derived from rat bone marrow mesenchymal stem cells exposed to 5-azacytidine. Muscle Nerve 1995; 18: 1417-26.
- Johnstone B, Hering TM, Caplan AI, Goldberg VM, Yoo JU. *In vitro* chondrogenesis of bone marrow-derived mesenchymal progenitor cells. Exp Cell Res 1998;238:265-72.
- Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, et al. Multilineage potential of adult human mesenchymal stem cells. Science 1999;284:143-7.
- Benayahu D, Kletter Y, Zipori D, Wientroub S. Bone marrow-derived stromal cell line expressing osteoblastic phenotype *in vitro* and osteogenic capacity *in vivo*. J Cell Physiol 1989;140:1-7.
- Bruder SP, Jaiswal N, Ricalton NS, Mosca JD, Kraus KH, Kadiyala S. Mesenchymal stem cells in osteobiology and applied bone regeneration. Clin Orthop Relat Res 1998;355:S247-56.
- Mitchell JB, McIntosh K, Zvonic S, Garrett S, Floyd ZE, Kloster A, et al. Immunophenotype of human adipose-derived cells: Temporal changes in stromal-associated and stem cell-associated markers. Stem Cells 2006;24:376-85.
- Zuk PA, Zhu M, Mizuno H, Huang J, Futrell JW, Katz AJ, et al. Multilineage cells from human adipose tissue: Implications for cellbased therapies. Tissue Eng 2001;7:211-28.

- Zuk PA, Zhu M, Ashjian P, De Ugarte DA, Huang JI, Mizuno H, *et al.* Human adipose tissue is a source of multipotent stem cells. Mol Biol Cell 2002; 13:4279-95.
- Zavan B, Vindigni V, Gardin C, D'Avella D, Della Puppa A, Abatangelo G, *et al*. Neural potential of adipose stem cells. Discov Med 2010;10:37-43.
- Fujimura J, Ogawa R, Mizuno H, Fukunaga Y, Suzuki H. Neural differentiation of adipose-derived stem cells isolated from GFP transgenic mice. Biochem Biophys Res Commun 2005;333:116-21.
- Jang S, Cho HH, Cho YB, Park JS, Jeong HS. Functional neural differentiation of human adiposetissue-derived stem cells using bFGF and forskolin. BMC Cell Biol2010;11:25.
- Ahmadi N, Razavi S, Kazemi M, Oryan S. Stability of neural differentiation in human adipose derived stem cells by two induction protocols. Tissue Cell 2012;44:87-94.
- McHugh PC, Rogers GR, Loudon B, Glubb DM, Joyce PR, Kennedy MA. Proteomic analysis of embryonic stem cell-derived neural cells exposed to the antidepressant paroxetine. J Neurosci Res 2008;86:306-16.
- Dagytė G, Crescente I, Postema F, Seguin L, Gabriel C, Mocaër E, et al. Agomelatine reverses the decrease in hippocampal cell survival induced by chronic mild stress. Behav Brain Res 2011;218:121-8.
- Anacker C, Zunszain PA, Cattaneo A, Carvalho LA, Garabedian MJ, Thuret S, *et al.* Antidepressants increase human hippocampal neurogenesis by activating the glucocorticoid receptor. Mol Psychiatry 2011;16:738-50.
- Liu JX, Pinnock SB, Herbert J. Novel control by the CA3 region of the hippocampus on neurogenesis in the dentate gyrus of the adult rat. PLoS One 2011;6:e 17562.
- Peng ZW, Xue F, Wang HN, Zhang RG, Chen YC, Wang Y, et al. Paroxetine up-regulates neurogenesis in hippocampus-derived neural stem cell from fetal rats. Mol Cell Biochem 2013;375:105-13.
- McEwen BS. The neurobiology of stress: From serendipity to clinical relevance. Brain Res 2000;886:172-89.
- Rajkowska G. Postmortem studies in mood disorders indicate altered numbers of neurons and glial cells. Biol Psychiatry 2000;48:766-77.

- Jin Y, Lim CM, Kim SW, Park JY, Seo JS, Han PL, *et al*. Fluoxetine attenuates kainic acid-induced neuronal cell death in the mouse hippocampus. Brain Res 2009;1281:108-16.
- Navailles S, Hof PR, Schmauss C. Antidepressant drug-induced stimulation of mouse hippocampal neurogenesis is age-dependent and altered by early life stress. J Comp Neurol 2008;509:372-81.
- 42. Boldrini M, Arango V. Antidepressants, age, and neuroprogenitors. Neuropsychopharmacology 2010;35:351-2.
- Manev H, Uz T, Smalheiser NR, Manev R. Antidepressants alter cell proliferation in the adult brain *in vivo* and in neural cultures *in vitro*. Eur J Pharmacol 2001;411:67-70.
- 44. McEwen BS. Stress and hippocampal plasticity. Annu Rev Neurosci 1999;22:105-22.
- 45. Gimble JM. Adipose tissue-derived therapeutics. Expert Opin Biol Ther 2003;3:705-13.
- Shi L, Yang X. Differentiation potential and application of stem cells from adipose tissue. Zhongguo Xiu Fu Chong Jian Wai Ke Za Zhi 2012;26:1007-11.
- Oh JS, Park IS, Kim KN, Yoon do H, Kim SH, Ha Y. Transplantation of an adipose stem cell cluster in a spinal cord injury. Neuroreport 2012;23:277-82.
- 48. Marconi S, Castiglione G, Turano E, Bissolotti G, Angiari S, Farinazzo A, et al. Human a dipose-derived mesenchymal stem cells systemically injected promote peripheral nerve regeneration in the mouse model of sciatic crush. Tissue Eng Part A 2012; 18:1264-72.
- 49. Sutcigil L, Oktenli C, Musabak U, Bozkurt A, Cansever A, Uzun O, *et al* .Pro- and anti-inflammatory cytokine balance in major depression: effect of sertraline therapy. Clin Dev Immunol 2007;76396.
- Zusso M, Debetto P, Guidolin D, Barbierato M, Manev H, Giusti P. Fluoxetine-induced proliferation and differentiation of neural progenitor cells isolated from rat postnatal cerebellum. Biochem Pharmacol 2008;76:391-403.

Source of Support: Isfahan University of Medical Sciences, Conflict of Interest: None declared.