

BASIC SCIENCE

OPEN

Cobalt-Chromium Alloy Has Superior Antibacterial Effect Than Titanium Alloy

In Vitro and In Vivo Studies

Kota Watanabe, MD, PhD,^a Satoshi Fukuzaki, PhD,^b Atsushi Sugino, PhD,^c Nicholas Benson, PhD,^d Newt Metcalf, BS,^d Masaya Nakamura, MD, PhD,^a and Morio Matsumoto, MD^a

Study Design. *In vitro* and *in vivo* laboratory studies.

Objective. This study aimed to compare bacterial survival on titanium alloy (Ti) and cobalt–chromium alloy (CC) using *in vitro* and *in vivo* experiments.

Summary of Background Data. Spinal implants are frequently manufactured from Ti and CC. These foreign materials are thought to be susceptible to biofilm formation that contributes to the development of surgical site infections. Certain metals (*i.e.*, silver, cobalt) are known to have antibacterial properties.

Methods. In the *in vitro* study, discs made of Ti or CC were incubated with one of two common bacteria: Staphylococcus aureus (S. aureus) and Propionibacterium acnes (P. acnes). After incubation, discs were assessed to determine the number of viable bacterial cells. In the *in vivo* study, the discs that were made of CC or Ti were implanted into the subcutaneous layer of BALB/c mice. After skin closure, a suspension including either S. aureus or P. acnes was directly inoculated on the implanted discs. The discs were retrieved and analyzed to determine the number of viable bacteria at 0.5, 1, and 3 days after inoculation. **Results.** The number of viable S. aureus cultured from the CC discs was $0.9 \pm 0.2 \times 10^3$ CFU/disc, which was significantly

From the ^aDepartment of Orthopaedic Surgery, Keio University School of Medicine, Tokyo, Japan; ^bGraduate School of Bioresources, Mie University, Mie, Japan; ^cMedtronic Sofamor Danek Co., Ltd., Osaka, Japan; and ^dMedtronic, Memphis, TN

Acknowledgment date: June 23, 2020. First revision date: September 1, 2020. Second revision date: November 9, 2020. Acceptance date: December 11, 2020.

The manuscript submitted does not contain information about medical device(s)/drug(s).

Medtronic Sofamor Danek Co, Ltd research funds were received in support of this work.

Relevant financial activities outside the submitted work: employment.

This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial-No Derivatives License 4.0 (CCBY-NC-ND), where it is permissible to download and share the work provided it is properly cited. The work cannot be changed in any way or used commercially without permission from the journal.

Address correspondence and reprint requests to Kota Watanabe, MD, PhD, Department of Orthopaedic Surgery, Keio University School of Medicine, 35 Shinanomachi, Shinjuku-ku, Tokyo 160-8582, Japan; E-mail: kw197251@keio.jp

DOI: 10.1097/BRS.0000000000003970

lower than the cultured Ti discs $(114.8\pm18.3\times10^3\,\text{CFU/disc})$. Moreover, a significantly lower mean number of *P. acnes* were cultured with CC $(1.9\pm1.2\times10^3\,\text{CFU/disc})$ compared with the Ti $(180.0\pm72.1\times10^3\,\text{CFU/disc})$. The *in vivo* infection model testing against *S. aureus* or *P. acnes* showed a significantly lower number of viable *S. aureus* or *P. acnes* on CC discs than Ti discs. The result was seen at all measured time points.

Conclusion. CC suppressed *S. aureus* and *P. acnes* proliferation compared with Ti *in vitro* and in an *in vivo* infection model.

Key words: antibacterial, antibacterial testing, biomaterial, cobalt–chromium alloy, infection model, *Propionibacterium acnes*, spinal instrumentation, *Staphylococcus aureus*, surgical site infection.

Level of Evidence: N/A Spine 2021;46:E911–E915

pinal instrumentation, such as pedicle screws or rods made of titanium (Ti) alloy or cobalt—chromium (CC) alloy, is an important tool for correcting adult deformity and adolescent idiopathic scoliosis and for providing stabilization until a definitive fusion for many spinal disorders. Although successful clinical results of instrumented spinal surgery have been extensively published, the incidence of surgical site infection (SSI) remains a concern. This concern is consistent across all orthopedic surgeries including joint arthroplasty and fracture surgery. The incidence of SSI in spinal surgery is reported to be between 0.6% and 11.9%. The incidence of SSI in spinal surgery is reported to be between 0.6% and 11.9%. The incidence of surgeries including joint arthroplasty and fracture surgery. The incidence of SSI in spinal surgery is reported to be between 0.6% and 11.9%. The incidence of surgeries in prolonged hospitalization, increased morbidity, reduced outcomes, and increased economic costs for patients and hospitals.

SSI risk factors are multifactorial including patient characteristics and pre-, intra-, and postoperative characteristics. Among the intraoperative risk factors, foreign materials, such as suture or spinal instruments at the surgery site, are known to increase the risk of SSI in surgery. Surface modifications to give an antibacterial effect to reduce the risk of foreign materials have been reported. Coatings including silver are common to provide materials with an antibacterial effect, and some researchers have reported experimental results. 11–13 Other promising approaches to add an antibacterial effect are polymer coatings including antibacterial agents, 4 such as an iodine-incorporated oxidized layer. 5,16 Other materials

combined with metallic cations, such as copper, zinc, cobalt, and gallium, and silver ions have also shown an antibacterial effect. ^{17,18} To test the hypothesis that CC is expected to show antibacterial effect according to previous reports, we conducted a comparative study of the antibacterial effect of common materials used for spinal implants.

This study aimed to compare the *Staphylococcus aureus* (*S. aureus*) and *Propionibacterium acnes* (*P. acnes*) survival on Ti and CC used as common spinal implant materials in *in vitro* and *in vivo* experiments. *S. aureus* and *P. acnes* are common SSI-causing pathogens and have been linked to late-onset screw loosening. ^{19–21}

MATERIALS AND METHODS

Specimen

For the *in vitro* antibacterial testing, discs (15 mm in diameter and 3 mm in thickness) made of either Ti (TI-6Al-4 V, ASTM F136) or CC (Co-28Cr-6Mo, ASTM F1537) were used. A disc (10 mm in diameter) made of polyethylene (Organo Corp, Tokyo, Japan) was used as a control. For *in vivo* testing, discs (7 mm in diameter and 1 mm in thickness) made from either from Ti or CC were used. To reproduce and validate the widely used implant materials, discs were provided from Medtronic Sofamor Danek Co, Ltd, in similar alloys to currently available spinal implants.

In Vitro Antibacterial Testing

Two types of bacteria, *S. aureus* (NBRC 12732, National Institute of Technology and Evaluation, Tokyo, Japan) and *P. acnes* (NBRC 107605, National Institute of Technology and Evaluation, Tokyo, Japan), were used in the *in vitro* experiments. *S. aureus* was cultured in a liquid nutrient broth (NB) medium (Nissui Pharmaceutical Co, Ltd, Tokyo, Japan) at 35°C for 48 hours under aerobic conditions. *P. acnes* was cultured in a liquid Gifu Anaerobic Medium (GAM) broth (Nissui Pharmaceutical Co, Ltd, Tokyo, Japan) at 35°C for 5 days under anaerobic conditions. The cells harvested by centrifugation were washed three times with 0.9% NaCl solution and then suspended in 0.9% NaCl solution. Cell suspensions were diluted with 1/100 NB medium or 1/100 GAM at appropriate dilution and used as the test inoculums.

The *in vitro* antibacterial testing was conducted according to the Japanese Industrial Method Z 2801:2010 "Antibacterial products–Test for antibacterial activity and efficacy" with minor modifications. To spread the inoculum on the surface and ensure close contact with the material surface, 40 μL of cell suspension was injected onto the discs and then covered with polyethylene film. The discs contacted with *S. aureus* and *P. acnes* were incubated at 35°C for 24 hours and then washed out with 4 mL of 0.9% NaCl solution containing 0.7% Tween 80 (Kanto Chemical Co, Inc, Tokyo, Japan). To determine the number of viable cells, serial decimal dilutions of 0.9% NaCl solution were prepared, and 0.1 mL was uniformly spread on an agar plate including appropriate culture mediums. The plates for *S.*

aureus and *P. acnes* were incubated at 35°C for 24 hours and 5 days under aerobic and anaerobic conditions, respectively. The data of viable cell numbers were counted as colonyforming units per disc (CFU/disc) and calculated antibacterial activity as a log reduction by comparing the number of viable cells on polyethylene with that on each alloy. Mann—Whitney *U* test with a 5% significance threshold was used to determine the difference of viable cells between CC and Ti. A sample size of 5 was used in the experiments.

In Vivo Infection Model

Two types of bacteria, *S. aureus* (ATCC 25923, American Type Culture Collection, VA) and *P. acnes* (NBRC 107605), were used in the *in vivo* experiments. *S. aureus* was cultured on *Staphylococcus* agar (Becton, Dickinson and Company, NJ) at 37° C for 2 days under aerobic condition. *P. acnes* was cultured on GAM agar (Nissui Pharmaceutical Co, Ltd, Tokyo, Japan) at 37° C for 3 days under the anaerobic condition. Then, the prepared cell suspensions were diluted with a concentration of 2×10^{7} CFU/mL by using physiological saline (Otsuka Pharmaceutical Factory, Inc, Tokyo, Japan).

Six-week-old, specific-pathogen-free, female BALB/c mice (Japan SLC Inc, Shizuoka, Japan) were used. A sample size of 10 animals was measured at each time period and study arms. The mice were anesthetized by an intramuscular injection of the mixture of 80 mg/kg ketamine (Daiichi Sankyo Propharma Co, Ltd, Tokyo, Japan) and 10 mg/kg xylazine (Bayer Yakuhin, Ltd, Osaka, Japan). After the back of each animal was shaved and disinfected with alcohol, an approximately 10-mm skin incision was made at the midline of the back, followed by the creation of a subcutaneous pocket for the discs. The discs made of CC and Ti were implanted onto the pocket, and then the surgical site was sutured. Then, 0.05 mL of each of cell suspensions $(1 \times 10^6 \text{ CFU})$ was directly injected on the surface of implanted disc. The mice were sacrificed at 0.5, 1, or 3 days with direct inoculation of excessive isoflurane anesthesia of isoflurane. Each removed disc was transferred to a test tube containing 1 mL of Hank's balanced salt solution (Thermo Fisher Scientific, MA), sonicated, and vigorously mixed to collect the S. aureus or P. acnes. The serial diluted suspension of S. aureus was then cultured on Staphylococcus agar plates at 37°C for 2 days under aerobic condition. The suspension of P. acnes was cultured on GAM agar plates at 37°C for 3 days under anaerobic conditions. The number of colonies on the plates was measured. Wilcoxon rank-sum test with a 1% significance threshold was used to determine the difference of viable cells between CC and Ti. A sample size of 10 was examined at each time point for the examined alloys. All experiments were approved by the Committee for the Ethical Treatment of Animals of Nihon Bioresearch Inc (Gifu, Japan) and conducted in accordance with the guidelines published from the Ministry of Health, Labor and Welfare and Nihon Bioresearch Inc.

TABLE 1. Results of the *In Vitro* Antibacterial Testing Against *Staphylococcus aureus* and *Propionibacterium acnes* on Titanium Alloy and Cobalt-Chromium Alloy

	Staphylococcus aureus		Propionibacterium acnes	
Material	Viable Cell /×10 ³ CFU/Disc (Mean ± SD)	Antibacterial Activity	Viable Cell $/ \times 10^3$ CFU/Disc (Mean \pm SD)	Antibacterial Activity
Cobalt-chromium alloy	$0.9 \pm 0.2^*$	2.9	$1.9 \pm 1.2^*$	2.2
Titanium alloy	114.8 ± 18.3	0.8	180.0 ± 72.1	0.2
Polyethylene	660.0 ± 143.4	N/A	318.0 ± 67.6	N/A

*P < 0.05 (Mann–Whitney U test). A sample size of 5 was used in the experiments. The number of viable S. aureus cultured from the cobalt–chromium (CC) discs was $0.9 \pm 0.2 \times 10^3$ CFU/disc, which was significantly lower than the cultured Ti discs (114.8 \pm 18.3 \times 10³ CFU/disc). In addition, a significantly lower mean number of P. acnes were cultured with CC (1.9 \pm 1.2 \times 10³ CFU/disc) compared with the Ti (180.0 \pm 72.1 \times 10³ CFU/disc).

RESULTS

In Vitro Antibacterial Efficacy

The results of the *in vitro* antibacterial testing against *S*. aureus and P. acnes on Ti and CC are presented in Table 1. The number of viable *S. aureus* on CC $(0.9 \pm 0.2 \times 10^3)$ CFU/ was significantly lower than that on Ti $(114.8 \pm 18.3 \times 10^3 \text{ CFU/disc})$ 24 hours after inoculation (P < 0.05). The reduction ratio of CC alloy and Ti alloy (vs. polyethylene) against S. aureus was 99.9% and 82.6% respectively. The antibacterial activity of CC against S. aureus was calculated using a log reduction comparison polyethylene. The log reduction was measured at 2.9, which means a 99.9% reduction in viable S. aureus on CC surface. The numbers of viable P. acnes on CC and Ti after inoculation for 24 hours were $1.9 \pm 1.2 \times 10^3$ CFU/disc and $180.0 \pm 72.1 \times 10^3$ CFU/disc, respectively. Statistical difference in the number of viable P. acnes between Ti and CC was observed (P < 0.05). The reduction ratio of CC alloy

and Ti alloy (*vs.* polyethylene) against *P. acnes* was 99.4% and 43.4% respectively. The antibacterial activity of CC against *P. acnes* was 2.2, which is able to reduce to approximately 100th of viable *P. acnes* in comparison with Ti.

In Vivo Antibacterial Efficacy

The number of viable S. aureus on Ti and CC in the in vivo infection model is presented in Figure 1. The number of viable S. aureus on the Ti was increased with extended implantation periods. In contrast, the number of viable S. aureus on the CC was significantly decreased from the 0.5 days after inoculation. The significant reduction of the viable S. aureus on the CC was kept for all implantation periods (P < 0.01).

The number of viable *P. acnes* on Ti and CC in the *in vivo* infection model is presented in Figure 2. The number of viable *P. acnes* on Ti and CC for 0.5 days after inoculation was $2.8 \pm 1.1 \times 10^{1}$ CFU/mm² and $0.5 \pm 0.1 \times 10^{1}$ CFU/mm², respectively. Significant differences were observed

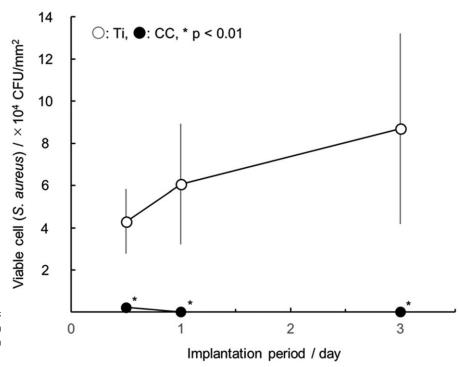


Figure 1. The number of viable *Staphylococcus aureus* on titanium alloy and cobalt–chromium alloy in the *in vivo* infection model (data shown as mean \pm 2SE).

Spine www.spinejournal.com E913

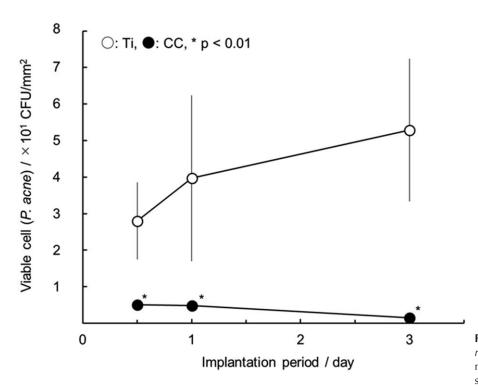


Figure 2. The number of viable *Propionibacte-rium acnes* on titanium alloy and cobalt-chromium alloy in the *in vivo* infection model (data shown as mean \pm 2SE).

in the number of viable P. acnes on CC in comparison with Ti for 0.5 days after inoculation (P < 0.01). The proliferation of P. acnes on the Ti was continued for all implantation time points, but not on CC. Significant difference was also observed in the number of the viable P. acnes on 1 and 3 days after inoculation on the CC in comparison with the Ti.

DISCUSSION

Because CC alloys has high corrosion resistance, wear resistance, fatigue strength, and biocompatibility, they have been widely used for currently available spinal implants. In this study, CC alloy significantly reduced *S. aureus* and *P. acnes* proliferation compared with Ti alloy in the both *in vitro* and *in vivo* experiments.

The experiments were conducted with two common SSI pathogens, S. aureus and P. acnes. S. aureus, which has been reported as the most common pathogen, is an aerobic bacterium that causes superficial SSI (64%)¹⁹ and deep incisional SSI (63%)4 in orthopedic and spinal surgeries, respectively, whereas P. acnes is an anaerobic bacterium that has been found in delayed postoperative spinal infections and late-onset screw loosening for spinal instrumentation. 19,20 Shiono et al. reported that P. acnes and P. species are the most frequent pathogens from intraoperative specimens collected during posterior correction surgery.²² They also reported that delayed infection of P. acnes occurred only in the presence of implants in the *in vivo* experiments.²³ Bacteria are initially attached to the material surface in a reversible adhesion. After changes to the irreversible adhesion on the material surface, the bacteria start to proliferate and produce extracellular polysaccharides as biofilms. The formed biofilms prevent the penetration of antimicrobial agents and disable their efficacy. The antibacterial effect of the CC alloy might be expected to suppress the *S. aureus* and *P. acnes* proliferation on the surface during initial phases of SSI development and may reduce one of the risks of SSI and delayed postoperative spinal infection and late-onset screw loosening in the spinal instrumentation. Only one clinical report can be found that assessed the effect of implant materials on SSI in pediatric scoliosis surgery, and there are no significant differences in SSI with stainless steel, Ti, or CC alloy/Ti alloy instrumentation.²⁴

Similar studies have been reported with different antibacterial testing methods and/or different species. Patel et al.²⁵ reported that the biofilm formation rate on CC against S. aureus was significantly higher than that on Ti. The testing methodology used small cut rods made of CC or Ti, and the rods were soaked in a culture medium with S. aureus for 24 hours. Optical density values and CFUs were measured to find the results. Conversely, Koseki et al.26 found a lower biofilm coverage rate of *S. epidermidis* on CC than Ti after culturing for 6 hours. In their experimental method, the bacteria suspension was directly poured on the discs made from various materials. There was no statistical difference between CC and Ti after 2 and 4 hours of culture. Although the experimental methods of these two papers and our in vitro testing methods seem similar, the testing methods were quite different. Direct comparisons are difficult to make leading to the need for an agreed-upon testing standard.

In conclusion, this study demonstrates that CC alloy suppresses *S. aureus* and *P. acnes* growth and survival compared with Ti alloy in both standardized *in vitro* antibacterial testing and in an *in vivo* infection model. This

study has many limitations, such as a small sample size, the use of a single small animal type, the use of CFUs as a proxy for infection strength, and an absence of clinical data. Adequately powered clinical studies are required to investigate whether differences in SSI incidence and screw loosening related to the examined pathogens exist.

> Key Points

- ☐ The antibacterial effects of CC alloy and Ti alloy against Staphylococcus aureus and Propionibacterium acnes were measured.
- □ CC alloy significantly suppressed *Staphylococcus* aureus and Propionibacterium acnes proliferation and inhibited survival compared with Ti alloy using both in vitro and in vivo experiments.
- Clinical studies are required to determine whether differences in surgical site infections exist between Ti and CC implant materials.

References

- 1. National Nosocomial Infections Surveillance (NNIS) system report, data summary from October 1986-April 1996, issued May 1996. A report from the National Nosocomial Infections Surveillance (NNIS) System. Am J Infect Control 1996;24:380-
- 2. National Nosocomial Infections Surveillance (NNIS) system report, data summary from January 1992 through June 2004 issued October 2004. A report from the National Nosocomial Infections Surveillance (NNIS) System. Am J Infect Control 2004;32:470-85.
- 3. Dave SH, Myers DL. Complications of lumbar spinal fusion with transpedicular instrumentation. Spine (Phila Pa 1976) 1992;17:
- 4. Weinstein MA, McCabe JP, Cammisa FP Jr. Postoperative spinal wound infection: a review of 2,391 consecutive index procedures. J Spinal Disord 2000;13:422-6.
- 5. Sponseller PD, LaPorte DM, Hungerford MW, et al. Deep wound infections after neuromuscular scoliosis surgery: a multicenter study of risk factors and treatment outcomes. Spine (Phila Pa 1976) 2000;25:2461-6.
- 6. Kirkland KB, Briggs JP, Trivette SL, et al. The impact of surgicalsite infections in the 1990 s: attributable mortality, excess length of hospitalization, and extra costs. Infect Control Hosp Epidemiol 1999;20:725-30.
- 7. Mangram AJ, Horan TC, Pearson ML, et al. Guideline for prevention of surgical site infection, 1999. Centers for Disease Control and Prevention (CDC) Hospital Infection Control Practices Advisory Committee. Am J Infect Control 1999;27:97-132.

- 8. Berard F, Gandon J. Postoperative wound infections: the influence of ultraviolet irradiation of the operating room and of various other factors. Ann Surg 1964;160:1-192.
- 9. Chu CC, Williams DF. Effects of physical configuration and chemical structure of suture materials on bacterial adhesion. A possible link to wound infection. Am J Surg 1984;147:197-204.
- 10. Portillo ME, Corvec S, Bores O, et al. Propionibacterium acnes: an underestimated pathogen in implant-associated infections. Biomed Res Int 2013;2013:804391.
- 11. Noda I, Miyaji F, Ando Y, et al. Development of novel thermal sprayed antibacterial coating and evaluation of release properties of silver ions. J Biomed Mater Res B Appl Biomater 2009;89:456-65.
- 12. Akiyama T, Miyamoto H, Yonekura Y, et al. Silver oxide-containing hydroxyapatite coating has in vivo antibacterial activity in the rat tibia. J Orthop Res 2013;31:1195-200.
- 13. Funao H, Nagai S, Sasaki A, et al. A novel hydroxyapatite film coated with ionic silver via inositol hexaphosphate chelation prevents implant-associated infection. Sci Rep 2016;6:23238.
- 14. Hart E, Azzopardi K, Taing H, et al. Efficacy of antimicrobial polymer coatings in an animal model of bacterial infection associated with foreign body implants. J Antimicrob Chemother 2010;65:974-80.
- 15. Shirai T, Shimizu T, Ohtani K, et al. Antibacterial iodine-supported titanium implants. Acta Biomater 2011;7:1928-33.
- 16. Tsuchiya H, Shirai T, Nishida H, et al. Innovative antimicrobial coating of titanium implants with iodine. J Orthop Sci 2012;17:595-604.
- 17. Saugo M, Brugnoni LI, Flamini DO, et al. Immobilization of antibacterial metallic cations (Ga3+, Zn2+ and Co2+) in a polypyrrole coating formed on Nitinol. Mater Scie Eng C Mater Biol Appl 2018;86:62-9.
- 18. Yousefshahi H, Aminsobhani M, Shokri M, et al. Anti-bacterial properties of calcium hydroxide in combination with silver, copper, zinc oxide or magnesium oxide. Eur J Transl Myol 2018;28:7545.
- 19. Arciola CR, Cervellati M, Pirini V, et al. Staphylococci in orthopaedic surgical wounds. New Microbiol 2001;24:365-9.
- 20. Haidar R, Najjar M, Der Boghossian A, et al. Propionibacterium acnes causing delayed postoperative spine infection: review. Scand I Infect Dis 2010;42:405-11.
- 21. Leitner L, Malaj I, Sadoghi P, et al. Pedicle screw loosening is correlated to chronic subclinical deep implant infection: a retrospective database analysis. Eur Spine J 2018;27:2529-35.
- 22. Shiono Y, Watanabe K, Hosogane N, et al. Sterility of posterior elements of the spine in posterior correction surgery. Spine (Phila Pa 1976) 2012;37:523-6.
- 23. Shiono Y, Ishii K, Nagai S, et al. Delayed Propionibacterium acnes surgical site infections occur only in the presence of an implant. Sci Rep 2016;6:32758.
- 24. Wright ML, Skaggs DL, Matsumoto H, et al. Does the type of metal instrumentation affect the risk of surgical site infection in pediatric scoliosis surgery?. Spine Deform 2016;4:206-10.
- 25. Patel SS, Aruni W, Inceoglu S, et al. A comparison of Staphylococcus aureus biofilm formation on cobalt-chrome and titaniumalloy spinal implants. J Clin Neurosci 2016;31:219-23.
- 26. Koseki H, Yonekura A, Shida T, et al. Early staphylococcal biofilm formation on solid orthopaedic implant materials: in vitro study. PLoS One 2014;9:e107588.

E915 Spine www.spinejournal.com