

Colorectal cancer-related mutant *KRAS* alleles function as positive regulators of autophagy

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ABSTRACT

The recent interest to modulate autophagy in cancer therapy has been hampered by the dual roles of this conserved catabolic process in cancer, highlighting the need for tailored approaches. Since RAS isoforms have been implicated in autophagy regulation and mutation of the *KRAS* oncogene is highly frequent in colorectal cancer (CRC), we questioned whether/how mutant *KRAS* alleles regulate autophagy in CRC and its implications. We established two original models, *KRAS*-humanized yeast and *KRAS*-non-cancer colon cells and showed that expression of mutated *KRAS* up-regulates starvation-induced autophagy in both. Accordingly, *KRAS* down-regulation inhibited autophagy in CRC-derived cells harboring *KRAS* mutations. We further show that *KRAS*-induced autophagy proceeds via up-regulation of the MEK/ERK pathway in both colon models and that *KRAS* and autophagy contribute to CRC cell survival during starvation. Since *KRAS* inhibitors have proven difficult to develop, our results suggest using autophagy inhibitors as a combined/alternative therapeutic approach in CRCs with mutant *KRAS*.

INTRODUCTION

Colorectal cancer (CRC) is a leading cause of cancer-related death worldwide [1]. Recent therapies to treat metastatic CRC (mCRC) use EGFR pathway inhibitors in order to block tumor growth and proliferation [2, 3]. However, resistance still occurs, highlighting the need to develop alternative therapeutic approaches. In particular, mutations in *KRAS*, an oncogene with an important role in colon carcinogenesis, reportedly occur in 30% to 50% of CRCs and are frequently associated with resistance of mCRC to anti-EGFR therapies. Therefore, these therapies are only efficient in mCRC patients with wild type *KRAS* [4–8]. Autophagy has also emerged as a potential target for cancer therapy, as it has been implicated in the resistance of tumors to chemotherapy [9]. Autophagy is an evolutionary conserved catabolic process in eukaryotic cells from yeast to mammals

that targets cellular components for lysosomal degradation to maintain cellular homeostasis [10]. Notably, cell lines harboring *RAS* mutations, such as those derived from bladder (T24, *HRAS*^{G12V} mutation), lung (H1299, *NRAS*^{G12C} mutation; H460, *KRAS*^{Q61H} mutation), pancreatic (PANC-1, *KRAS*^{G12V} mutation), and colorectal cancer (HCT116, *KRAS*^{G13D} mutation), have high basal levels of autophagy [11]. Indeed, several reports implicated RAS isoforms in autophagy regulation, with different outcomes in cancer progression depending on the cell type [11–17]. However, little is known about the role of *KRAS* mutations in autophagy regulation in CRC cells and the signaling pathways involved.

In this work, we aimed to understand the precise role of mutant *KRAS* alleles in autophagy. For this purpose, we established two original cell models for a controlled genetic background where the only difference is the *KRAS* variant they express: non-cancer colon cells derived from normal human

colon mucosal epithelium (NCM460) and a “humanized” *Saccharomyces cerevisiae*. The NCM460 cell line background does not contain *KRAS* mutations or deregulated pathways [18]. The yeast *S. cerevisiae*, a unicellular eukaryote highly amenable to manipulation and genetic modifications, is a well-established tool to study molecular aspects of different conserved biological processes, and a particularly advantageous system for heterologous expression [19, 20]. Moreover, autophagy-related (*ATG*) genes, responsible for the formation of the autophagosome, were first discovered in yeast and orthologues of many of these genes have been identified and characterized in higher eukaryotes, indicating that autophagy is a highly conserved pathway through evolution [21]. To transpose our findings to more clinically relevant cancer models, we used the reverse approach by silencing *KRAS* in two CRC-derived cell lines harboring different *KRAS* activating mutations. Here, we find that expression of mutant *KRAS* consistently increases autophagy in all models, which, in the colon cell lines, is associated with up-regulation of the MEK/ERK pathway. We also demonstrate that *KRAS* and autophagy support the survival of CRC cells exposed to stressful conditions like nutrient limitation. This suggests that inhibition of *KRAS* or autophagy may sensitize CRC cells harboring *KRAS* mutations to cancer therapies, reinforcing *KRAS*-induced autophagy inhibition as an emerging target for CRC therapeutic approaches.

RESULTS

***KRAS* mutations up-regulate autophagy during starvation in non-cancer colon cells and in a “*KRAS*-humanized yeast” model**

To elucidate the role of oncogenic *KRAS* in autophagy regulation, we expressed FLAG-tagged wild-type *KRAS* (*KRAS*^{WT}) or mutated *KRAS* (*KRAS*^{G13D}, *KRAS*^{G12D} and *KRAS*^{G12V}) in two model systems: in the human non-cancer colon NCM460 cell line, after stable infection, and in the yeast *S. cerevisiae ras2Δ* strain from the pCM184 plasmid, under the control of a *tet-off* promoter (this strain was chosen to abolish the input of yeast Ras2p, itself involved in autophagy regulation).

We first assessed the autophagic flux in NCM460 cells, by monitoring the conversion of LC3-I into LC3-II, a hallmark of autophagy, in the presence and absence of Bafilomycin A1 (Baf. A1) [22]. Overexpression of FLAG-*KRAS*^{WT}, -*KRAS*^{G13D}, -*KRAS*^{G12D} and -*KRAS*^{G12V} increased the basal level of LC3-II in complete medium, in comparison with parental NCM460 (non infected) cells (Figure 1a). During starvation, overexpression of FLAG-*KRAS*^{G13D} and -*KRAS*^{G12D}, but not of -*KRAS*^{G12V}, increased the autophagic flux in comparison with parental NCM460 and -*KRAS*^{WT}-expressing cells, as shown by the increased accumulation of LC3-II upon inhibition of lysosomal degradation by Baf. A1 (Figures 1a, 1b and Supplementary Figure S1a). Though these differences were consistent, variability was high and did

not reach statistical significance. We therefore quantitatively measured autophagic proteolysis of long-lived proteins radiolabeled with L-[¹⁴C]valine [23, 24]. In accordance with the previous assay, overexpression of FLAG-*KRAS*^{G13D} and -*KRAS*^{G12D}, but not -*KRAS*^{G12V}, increased the level of autophagic proteolysis, in comparison with parental NCM460 and -*KRAS*^{WT}-expressing cells (Figures 1c and Supplementary Figure S1b). This increase was statistically significant in -*KRAS*^{G13D}-expressing cells, but in -*KRAS*^{G12D}- only when comparing with cells expressing -*KRAS*^{WT} (Figure 1c). Our results therefore indicate that some forms of mutated *KRAS* up-regulate autophagy in non-cancer colon cells.

In the *S. cerevisiae* model, autophagy levels were monitored by assessing the levels of the autophagic marker Atg8p, the yeast homologue of LC3, in the presence of the protease inhibitor PMSF to avoid degradation during the subsequent steps of the autophagy process [25]. No significant differences were observed in Atg8p levels between *ras2Δ* cells carrying the empty vector and *ras2Δ* cells expressing *KRAS*^{WT}, after 24 h of nitrogen starvation (Figures 1d and 1e). In contrast, expression of *KRAS*^{G13D}, *KRAS*^{G12D} and *KRAS*^{G12V} increased Atg8p levels under the same conditions (Figures 1d and 1e). These results show that activating *KRAS* mutations increase Atg8p levels more than wild-type *KRAS* in response to nitrogen starvation.

To confirm that the higher Atg8p levels detected were associated with induction of the autophagy flux, we also monitored the amount of Atg8p delivered to the vacuole. We transformed *ras2Δ* strains containing pCM184/*KRAS* plasmids or empty vector control with a GFP-*ATG8* construct and monitored the degradation of GFP-Atg8 to free GFP by vacuolar hydrolases, which reflects the level of autophagy, by immunoblotting [26]. We found no significant differences in the accumulation of free GFP between *ras2Δ* cells carrying the empty vector and *ras2Δ* cells expressing *KRAS*^{WT}, after 24 h of nitrogen starvation (Figures 1f and 1g). On the other hand, expression of *KRAS*^{G13D}, *KRAS*^{G12D} and *KRAS*^{G12V} increased the levels of the free GFP moiety after 24 h of nitrogen starvation (Figures 1f and 1g). *KRAS* protein levels were confirmed by immunoblot against this specific human RAS isoform (Figure 1f). These results indicate that mutated *KRAS* lead to a higher delivery of Atg8p into the yeast vacuole during starvation than wild-type *KRAS*.

Altogether, our results in both non-cancer colon and “*KRAS*-humanized yeast” models show that mutated *KRAS* up-regulate autophagy to a higher extent than wild-type *KRAS* in response to nitrogen starvation, and validate both cell models to study the role of human *KRAS* proteins in the evolutionary conserved autophagic process.

Suppression of *KRAS* by RNA interference inhibits autophagy in HCT116 and SW480 cell lines

In order to confirm the aforementioned results obtained in yeast and NCM460 cells, we used cell lines derived from CRC patients and assessed whether suppression of

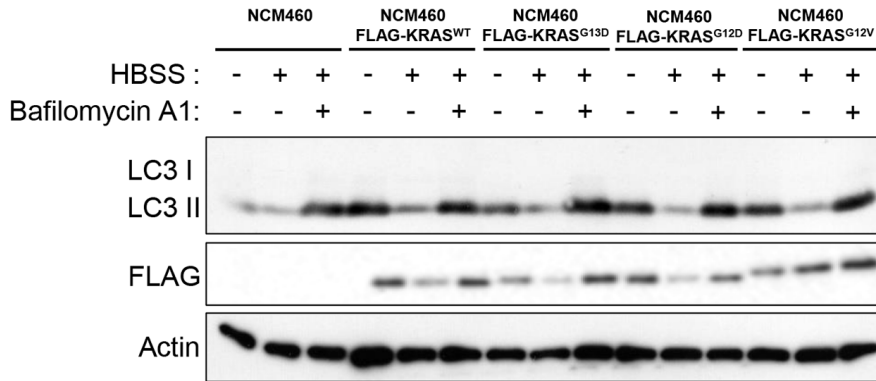
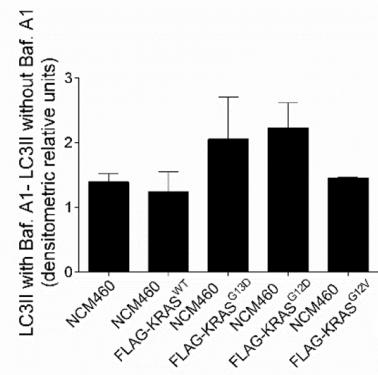
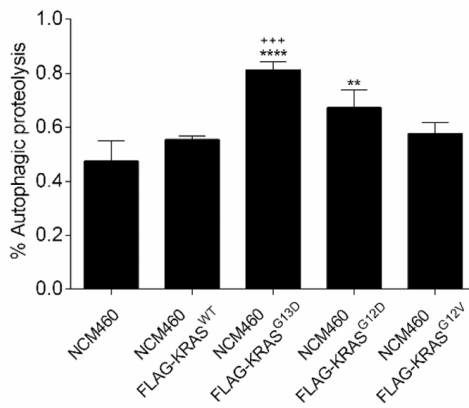
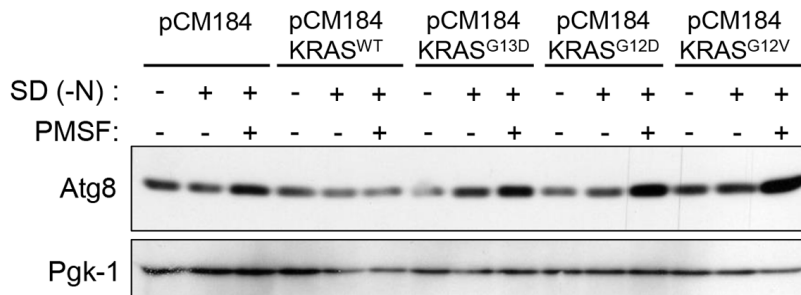
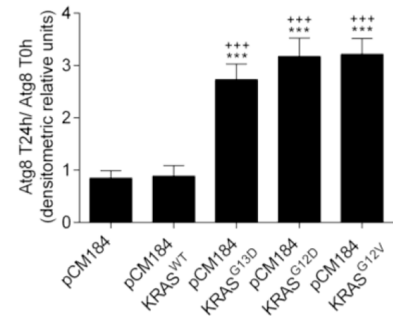
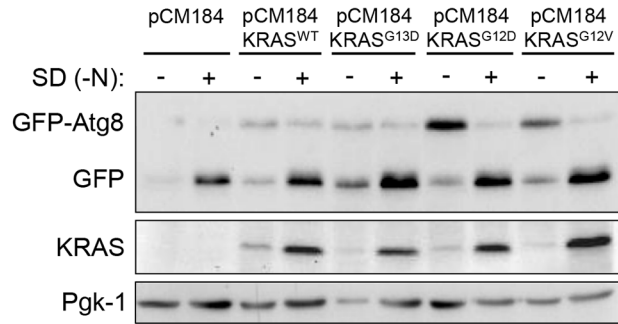
a**b****c****d****e**

Figure 1: Activating *KRAS* mutations increase the autophagic flux during starvation, in NCM460 cells and *S. cerevisiae*. **a.** Immunoblot analysis of LC3-I/II in NCM460 cells and NCM460 cells expressing *FLAG-KRAS^{WT}*, *FLAG-KRAS^{G13D}*, *FLAG-KRAS^{G12D}* or *FLAG-KRAS^{G12V}*. LC3-I expression levels were too low to be detected. FLAG immunoblot was used as a control for expression of FLAG-tagged *KRAS* constructs. **b.** LC3II/Actin ratios were determined and used to calculate the autophagic flux by subtracting the normalized LC3-II levels in the absence of Baf. A1 from the corresponding levels obtained in the presence of Baf. A1, according to guidelines already published. [24] Results are means \pm SEM of three independent experiments. **c.** Level of autophagic proteolysis during starvation in NCM460 cell lines. Cells were incubated with L-[¹⁴C]valine, and chased for 6 h in complete medium, HBSS or HBSS supplemented with 0.1 μ M Baf. A1. The percentage of autophagic proteolysis was determined by subtracting the levels of proteolysis of cells incubated in HBSS in the absence of Baf. A1 from the corresponding levels of proteolysis of cells incubated in HBSS supplemented with Baf. A1. Values represent mean \pm SEM of three independent experiments. *** P < 0.001 vs NCM460; *** P < 0.001, * P < 0.05 vs NCM460 FLAG-KRAS^{WT}; One-way ANOVA. **d.** Detection of Atg8p in the presence and absence of 1mM PMSF in *S. cerevisiae* ras2 Δ cells expressing the empty vector pCM184, pCM184/KRAS^{WT}, pCM184/KRAS^{G13D}, pCM184/KRAS^{G12D} or pCM184/KRAS^{G12V} after 24h of nitrogen starvation (SD -N) and **e.** Atg8 T24 h -N/Atg8 T0 h -N ratio after normalization of Atg8 to Pgk-1. (Continued)

f



g

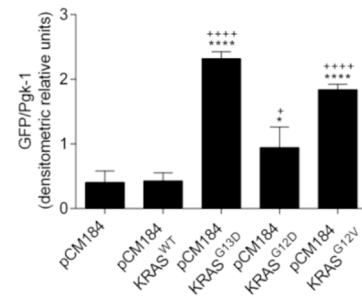


Figure 1: (Continued) Activating *KRAS* mutations increase the autophagic flux during starvation, in NCM460 cells and *S. cerevisiae*. f. Free GFP generated from the GFP-Atg8p fusion protein in *S. cerevisiae ras2Δ* cells co-expressing GFP-Atg8p with either empty vector pCM184, pCM184/*KRAS*^{WT}, pCM184/*KRAS*^{G13D}, pCM184/*KRAS*^{G12D} or pCM184/*KRAS*^{G12V} after 24 h of nitrogen starvation, as shown by immunoblot analysis. *KRAS* immunoblot was used as a control for expression of human *KRAS*. Pgk-1 immunoblot was used as a loading control. g. Free GFP/Pgk-1 ratio at 24 h of nitrogen starvation, using *ImageJ* software. Values are means ± SEM of three independent experiments. ***P* < 0.01; **P* < 0.05 one-way ANOVA followed by Tukey's test vs pCM184/*KRAS*^{WT}.

KRAS affects autophagy levels in HCT116 (*KRAS*^{G13D}) and SW480 (*KRAS*^{G12V}) cells. Indeed, suppression of *KRAS* by targeted small interference RNA (siRNA) in both cell lines significantly impaired the autophagic flux, evidenced by the decreased accumulation of LC3-II upon inhibition of lysosomal degradation by Baf. A1, whereas transfection with control siRNA did not (Figures 2a, 2b, Supplementary Figure S2a, 2c, 2d and Supplementary Figure S2b).

We next determined how depletion of *KRAS* affects the levels of other components of the autophagic machinery: the Atg5-Atg12 conjugate, a critical component of autophagosome formation [27], and Beclin 1, a component of the class III PI3K complex (PI3KC3) that induces autophagosome formation [28]. Depletion of *KRAS* by siRNA in SW480 and HCT116 cells decreased the levels of Atg5-Atg12 conjugate and Beclin 1 in comparison with control siRNA-transfected cells, both in complete medium and starvation conditions (Figures 2e–2j). Taken together, these data show that HCT116 and SW480 cells rely on *KRAS*^{G13D} and *KRAS*^{G12V} mutations, respectively, to maintain autophagy.

We also evaluated how manipulating the levels of mutant *KRAS* alleles in *S. cerevisiae* affects autophagy. For this purpose, cells grown in the absence of doxycycline (expressing *KRAS*) were transferred to nitrogen starvation media without or with doxycycline (to maintain or repress *KRAS* transcription, respectively) and the conversion of GFP-Atg8p to free GFP was monitored. As seen in Figures 2k and Supplementary Figure S3a, activation of *KRAS* transcription during starvation increased the level of wild-type *KRAS*, associated with an increase in the amount of free GFP generated, hence of autophagy induction. In turn, repression of wild-type *KRAS* transcription stabilized the level of *KRAS* protein and consequently the level of free GFP generated (Figures 2k and Supplementary Figure S3a). The same was not observed in yeast cells expressing *KRAS*^{G13D},

KRAS^{G12D} or *KRAS*^{G12V}. In these cells, nitrogen starvation maintaining *KRAS* expression also increased the levels of the respective *KRAS* variant, associated with accumulation of free GFP (Figures 2k, Supplementary Figure S3b, S3c and S3d). However, during repression of *KRAS* transcription, the level of *KRAS* proteins stabilized but the level of free GFP still increased (Figures 2k, Supplementary Figure S3b, S3c and S3d). Therefore, yeast cells harboring wild-type *KRAS* depend on the amount of *KRAS* protein for autophagy induction, whereas those harboring activating *KRAS*^{G13D}, *KRAS*^{G12D} or *KRAS*^{G12V} do not. These results are consistent with constitutive activation of *KRAS* signaling due to the inability of mutated *KRAS* to hydrolyze GTP and consequently to be “switched off” [29–32], thus reproducing in the yeast background the effect of the aforementioned mutations on *KRAS* signaling.

These data also further support that *KRAS* mutations confer cells increased capacity for autophagy induction and that efficiently countering this effect can be achieved only by decreasing mutant *KRAS* protein levels below a certain threshold, as we observed in the CRC cell lines.

***KRAS* mutations induce autophagy through up-regulation of the ERK pathway**

The MEK/ERK and the PI3K/AKT/mTOR pathways are the classical effectors of *RAS*, implicated in the regulation of various cellular responses known to generate resistance to CRC therapy [33]. We therefore analyzed how the *KRAS* variants affect the phosphorylation levels of ERK1/2 and AKT proteins in response to starvation. ERK phosphorylation increased in response to starvation in NCM460 cells overexpressing FLAG-*KRAS*^{G13D}, -*KRAS*^{G12D} and -*KRAS*^{G12V}, but not in parental NCM460 or FLAG-*KRAS*^{WT}-expressing cells (Figures 3a and 3b). In contrast, starvation decreased AKT phosphorylation

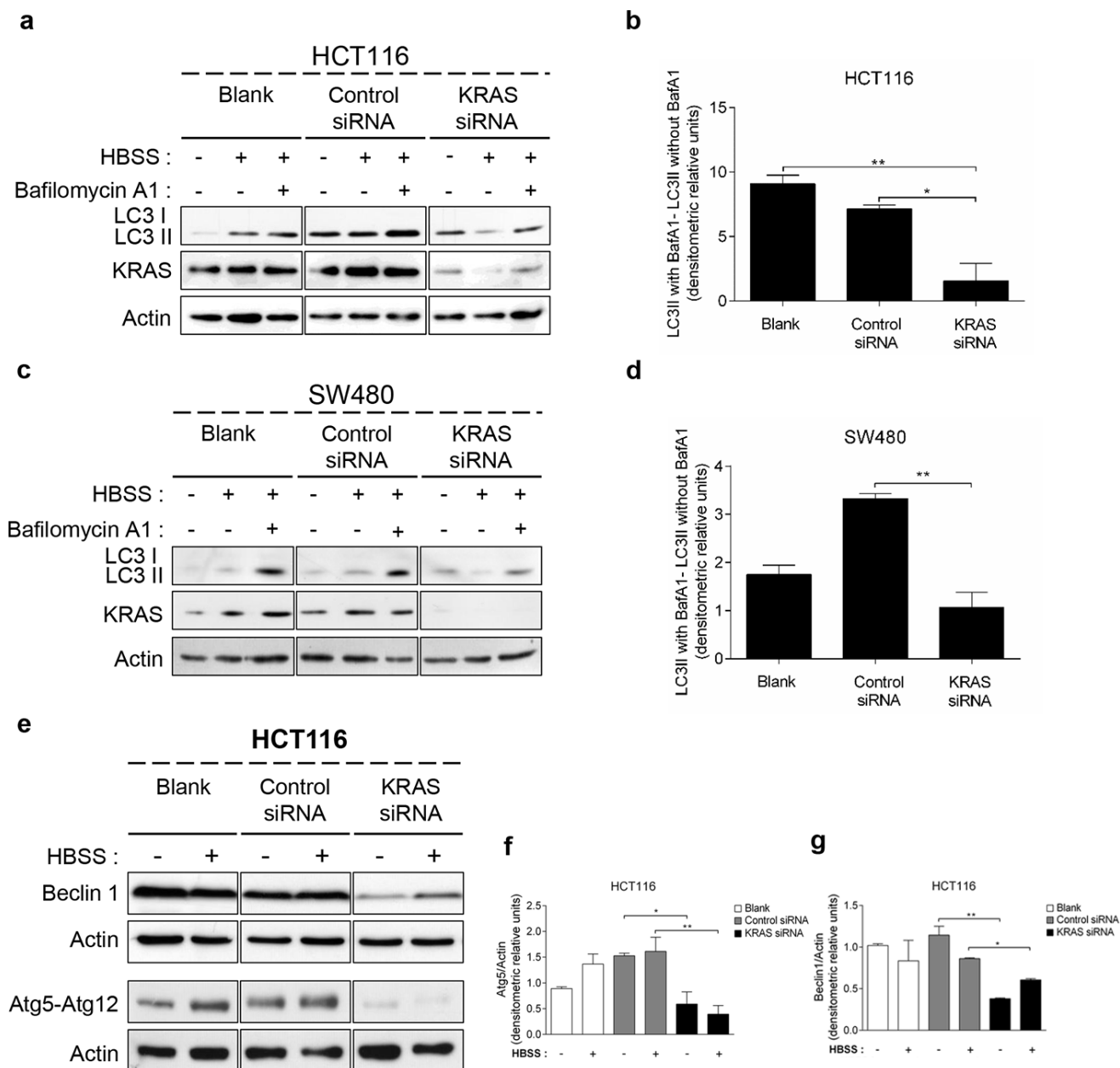


Figure 2: Suppression of *KRAS* in CRC-derived cell lines decreases autophagy. **a. c.** Immunoblot analysis of LC3-I/II, **e.** Atg5-Atg12 conjugate (using anti-Atg5 antibody) and Beclin 1 in HCT116 and SW480 cell lines. Cells were left non transfected (blank) or transfected with either siRNA control or siRNA targeted against *KRAS*. After 48 h of transfection, the cells were maintained in complete medium or incubated in HBSS medium (starvation) for further 6 h in presence and absence of 20 nM Baf A1. LC3-I expression levels were too small to be detected. *KRAS* immunoblot was used to show the efficient silencing by *KRAS* siRNA. Actin immunoblot was used as a loading control. All the protein bands showed in the lanes are from the same immunoblot despite being organized in different boxes. [24] **b. d.** LC3-II/Actin ratios determined using *ImageJ* software were used to determine the autophagic flux ratio of by subtracting the normalized LC3-II levels in the absence of Baf. A1 from the corresponding levels obtained in the presence of Baf. A1, according to guidelines already published. [24] **f.** Atg5-Atg12/Actin ratio and **g.** Beclin 1/Actin ratio of HCT116 cells were determined using *Image J* software. Results are means \pm SEM of three independent experiments. ** $P < 0.01$; * $P < 0.05$ one-way ANOVA followed by Tukey's test. (Continued)

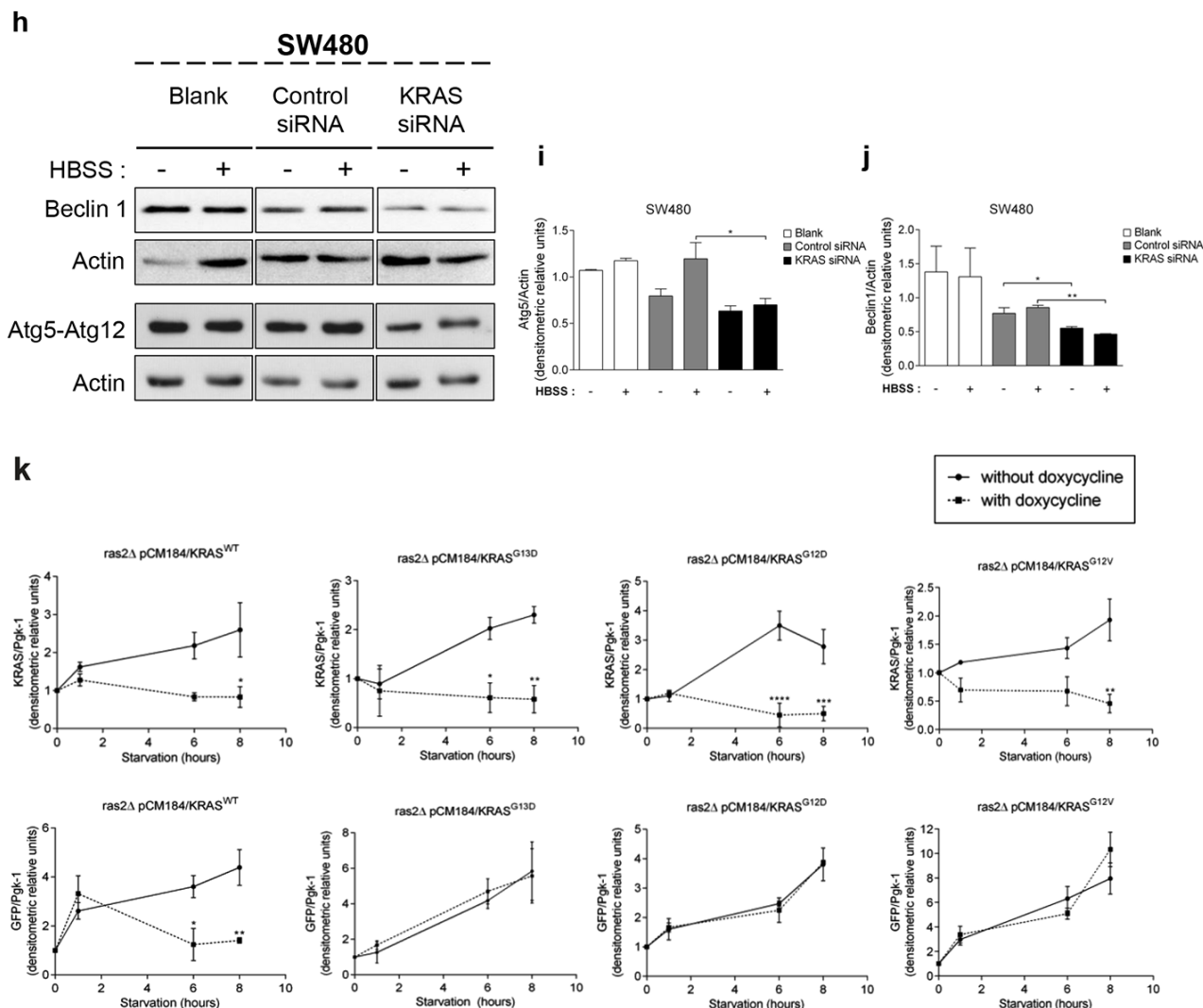


Figure 2: (Continued) Suppression of *KRAS* in CRC- derived cell lines decreases autophagy. **h.** Atg5-Atg12 conjugate (using anti-Atg5 antibody) and Beclin 1 in SW480 cell lines. Cells were left non transfected (blank) or transfected with either siRNA control or siRNA targeted against *KRAS*. After 48 h of transfection, the cells were maintained in complete medium or incubated in HBSS medium (starvation) for further 6 h in presence and absence of 20 nM Baf A1. LC3-I expression levels were too small to be detected. *KRAS* immunoblot was used to show the efficient silencing by *KRAS* siRNA. Actin immunoblot was used as a loading control. All the protein bands showed in the lanes are from the same immunoblot despite being organized in different boxes. **i.** Atg5-Atg12/Actin ratio and **j.** Beclin 1/Actin ratio of SW480 cells were determined using *Image J* software. Results are means \pm SEM of three independent experiments. $**P < 0.01$; $*P < 0.05$ one-way ANOVA followed by Tukey's test. **k.** Autophagy induction depends on the activating status of mutated *KRAS* activation during starvation in *S. cerevisiae*. *KRAS*/Pgk-1 and free GFP/Pgk-1 ratios of *ras2*Δ cells with/without expression of pCM184/*KRAS*^{WT}, pCM184/*KRAS*^{G13D}, pCM184/*KRAS*^{G12D} or pCM184/*KRAS*^{G12V}. Cells were cultured in SC medium without the appropriate aminoacids + 2% glucose until exponential phase. At time 0, the cultures were subdivided: one part was incubated without doxycycline to maintain *KRAS* transcription active and another incubated with 10 μ g/ml doxycycline to repress *KRAS* transcription. The level of free GFP generated and *KRAS* expression were assessed by immunoblot, at the indicated times and normalized for Pgk-1 using *ImageJ* software. Results are mean \pm SEM of three independent experiments. $****P < 0.0001$, $***P < 0.001$, $**P < 0.01$, $*P < 0.05$ compared to the respective time point of cells non-treated with doxycycline.

in NCM460 expressing all *KRAS* variants, but not in parental cells, though not to a significant extent in the case of FLAG-*KRAS*^{G13D} (Figures 3a and 3c).

In agreement with the results obtained in NCM460 cells overexpressing FLAG-*KRAS*^{G13D} and FLAG-*KRAS*^{G12V} (Figure 3a), starvation also increased ERK phosphorylation in both HCT116 (*KRAS*^{G13D} mutation)

and SW480 (*KRAS*^{G12V} mutation) cells (Figures 3d, 3e, 3g and 3h). Accordingly, depletion of *KRAS* not only prevented this increase but also decreased ERK phosphorylation in non-starved cells (Figures 3d, 3e, 3g and 3h).

In HCT116 cells, AKT phosphorylation also mimicked the results obtained in the NCM460

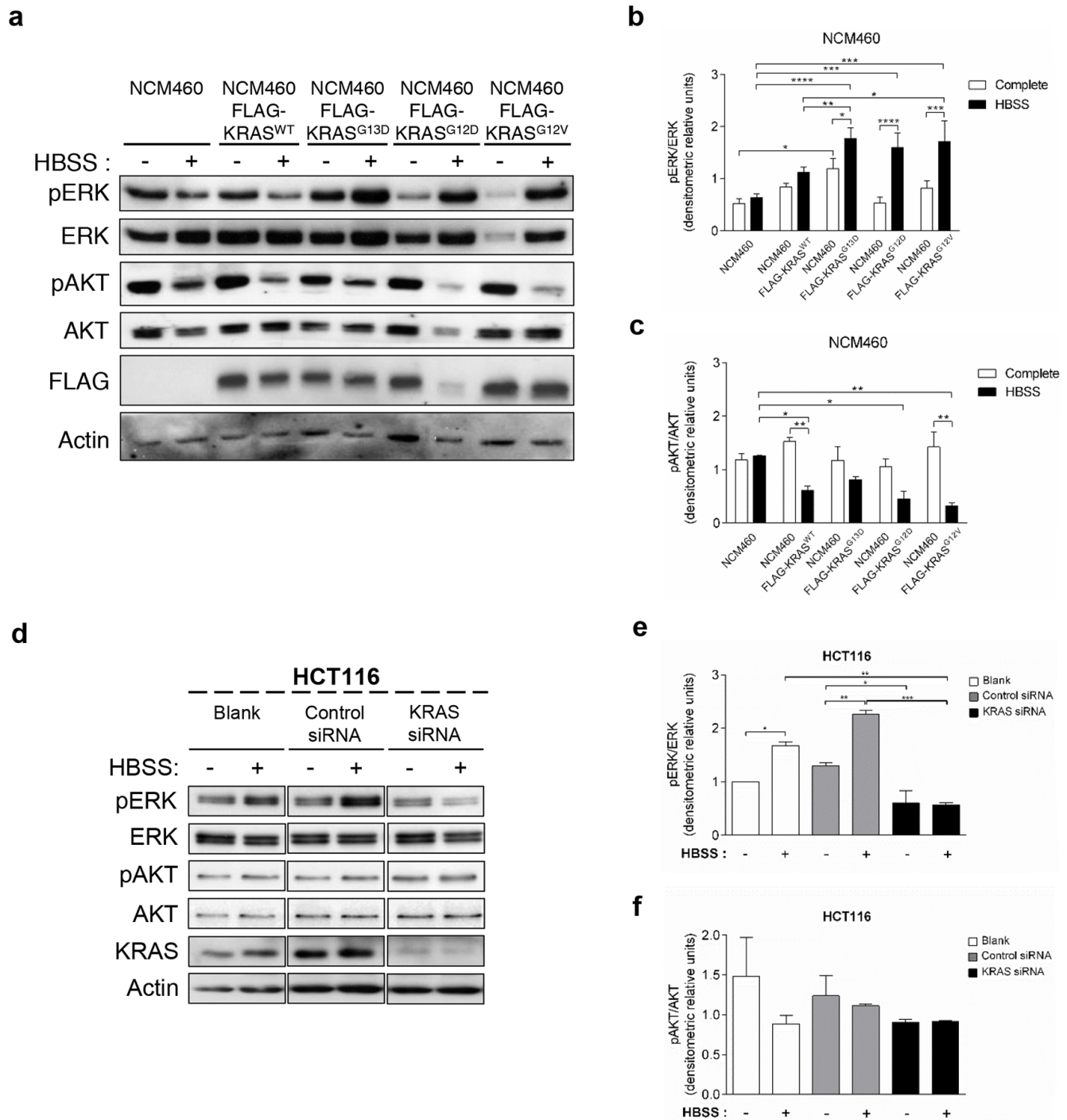


Figure 3: Mutated *KRAS* up-regulates ERK phosphorylation during starvation in normal and CRC- derived cells.
a. Immunoblot analysis of pERK and pAKT in parental NCM460 and NCM460 cells infected with *FLAG-KRAS^{WT}*, *FLAG-KRAS^{G13D}*, *FLAG-KRAS^{G12D}* or *FLAG-KRAS^{G12V}*. Cells were grown at a density of 80% and were maintained in complete conditions or incubated in HBSS (starvation) medium for 6 h. Cell lysates were analyzed by immunoblot for the indicated proteins. FLAG immunoblot was used as a control for expression of FLAG-tagged *KRAS* constructs. Actin immunoblot was used as a loading control. The **b.** pERK/ERK and **c.** pAKT/AKT ratios determined using *ImageJ* software. **** $P < 0.0001$; *** $P < 0.001$; ** $P < 0.01$; * $P < 0.05$ one-way ANOVA followed by Tukey's test. **d.** Immunoblot analysis of pERK and pAKT in HCT116 cells. Cells were left non transfected (blank) or transfected with either siRNA control or siRNA targeted against *KRAS*. 48 h after transfection, the cells were maintained in complete medium or incubated in HBSS medium (starvation) for further 6 h and cell lysates were analyzed by immunoblot for the indicated proteins. *KRAS* immunoblot was used to show the efficient silencing by siRNA *KRAS*. Actin immunoblot was used as a loading control. All the protein bands showed in the lanes are from the same immunoblot despite being organized in different boxes. **e.** pERK/ERK and **f.** pAKT/AKT ratios were determined using *ImageJ* software. Results are means \pm SEM of three independent experiments. *** $P < 0.001$; ** $P < 0.01$; * $P < 0.05$ one-way ANOVA followed by Tukey's test. (Continued)

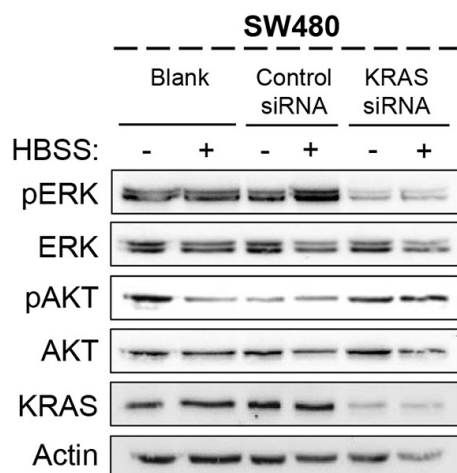
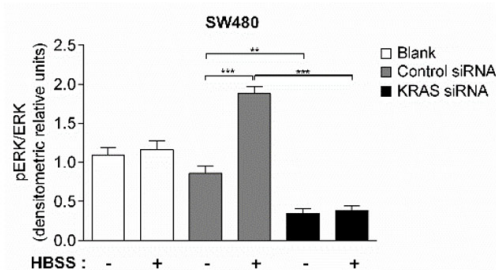
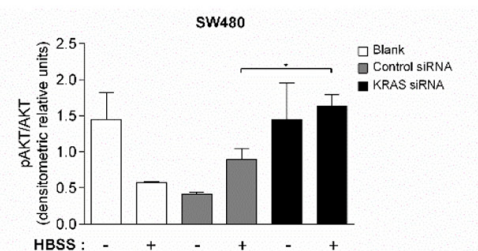
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Figure 3: (Continued) Mutated *KRAS* up-regulates ERK phosphorylation during starvation in normal and CRC-derived cells. g. Immunoblot analysis of pERK and pAKT in SW480 cells. Cells were left non transfected (blank) or transfected with either siRNA control or siRNA targeted against *KRAS*. 48 h after transfection, the cells were maintained in complete medium or incubated in HBSS medium (starvation) for further 6 h and cell lysates were analyzed by immunoblot for the indicated proteins. *KRAS* immunoblot was used to show the efficient silencing by siRNA *KRAS*. Actin immunoblot was used as a loading control. All the protein bands showed in the lanes are from the same immunoblot despite being organized in different boxes. The **h**, pERK/ERK and **i**, pAKT/AKT ratios were determined using *ImageJ* software. Results are means \pm SEM of three independent experiments. *** $P < 0.001$; ** $P < 0.01$; * $P < 0.05$ one-way ANOVA followed by Tukey's test.

FLAG-*KRAS*^{G13D} model: a slight but not significant decrease in control siRNA-transfected cells, and no difference in *KRAS*-depleted cells (Figures 3d and 3f). In SW480 control siRNA-transfected cells, no statistical significance differences were obtained in rich medium. On the other hand, under starvation conditions, depletion of *KRAS* increased AKT phosphorylation in comparison control siRNA-transfected cells also in agreement with the NCM460 FLAG-*KRAS*^{G12V} model (Figures 3g and 3i).

These data indicate that *KRAS*-induced autophagy is mediated through up-regulation of the MEK/ERK pathway and down-regulation of the PI3K/AKT pathway, known to activate the autophagy inhibitor mTOR [34], therefore resulting in autophagy induction. To confirm this hypothesis, we examined if these pathways affect starvation-induced accumulation of LC3-II after inhibition of lysosomal degradation by Baf. A1. In accordance with the aforementioned results, depletion of *MEK1/2* by siRNA decreased the autophagic flux of SW480 cells, whereas depletion of *PI3KCA* had no effect (Figures 4a, 4b, Supplementary Figure S4a, 4c, 4d and Supplementary Figure S4b).

Taken together, our results indicate that oncogenic *KRAS* proteins activate autophagy through up-regulation of the MEK/ERK pathway in non-cancer colon and CRC-derived cells, suggesting that autophagy plays an important role in *KRAS*-driven colorectal carcinogenesis.

Suppression of *KRAS* or autophagy leads to cell death in CRC-derived cells

To understand whether *KRAS* and/or autophagy are required for CRC cell survival, prompting a possible therapeutic approach, we analyzed how depletion of *KRAS*, *ATG5* or *BECN1* by siRNA affected cell death. We found that depletion of *KRAS* in SW480 cells increased the number of cells stained with annexin V (AV+ PI -/+), early apoptosis plus late apoptosis/necrosis) in comparison with control siRNA-transfected cells, particularly evident under starvation conditions (Figure 5). Depletion of *ATG5* or *BECN1* (Supplementary Figure S5) had the same effect, though to a slightly lesser extent (Figure 5).

Taken together, our results show that *KRAS*-induced autophagy allows CRC-derived cells to survive in stressful conditions, such as nutrient limitation, as depletion of *KRAS* or impairment of the autophagic machinery promotes cell death of CRC cells.

DISCUSSION

In the present study, we analyzed the role of activating *KRAS* mutations on autophagy regulation in CRC. We established a non-cancer colon NCM460 cell model to study, in the same genetic background, the effect of overexpressing three of the most common mutant *KRAS* alleles that occur in CRCs: *KRAS*^{G13D}, *KRAS*^{G12D} and *KRAS*^{G12V} [35]. We show that autophagy is induced

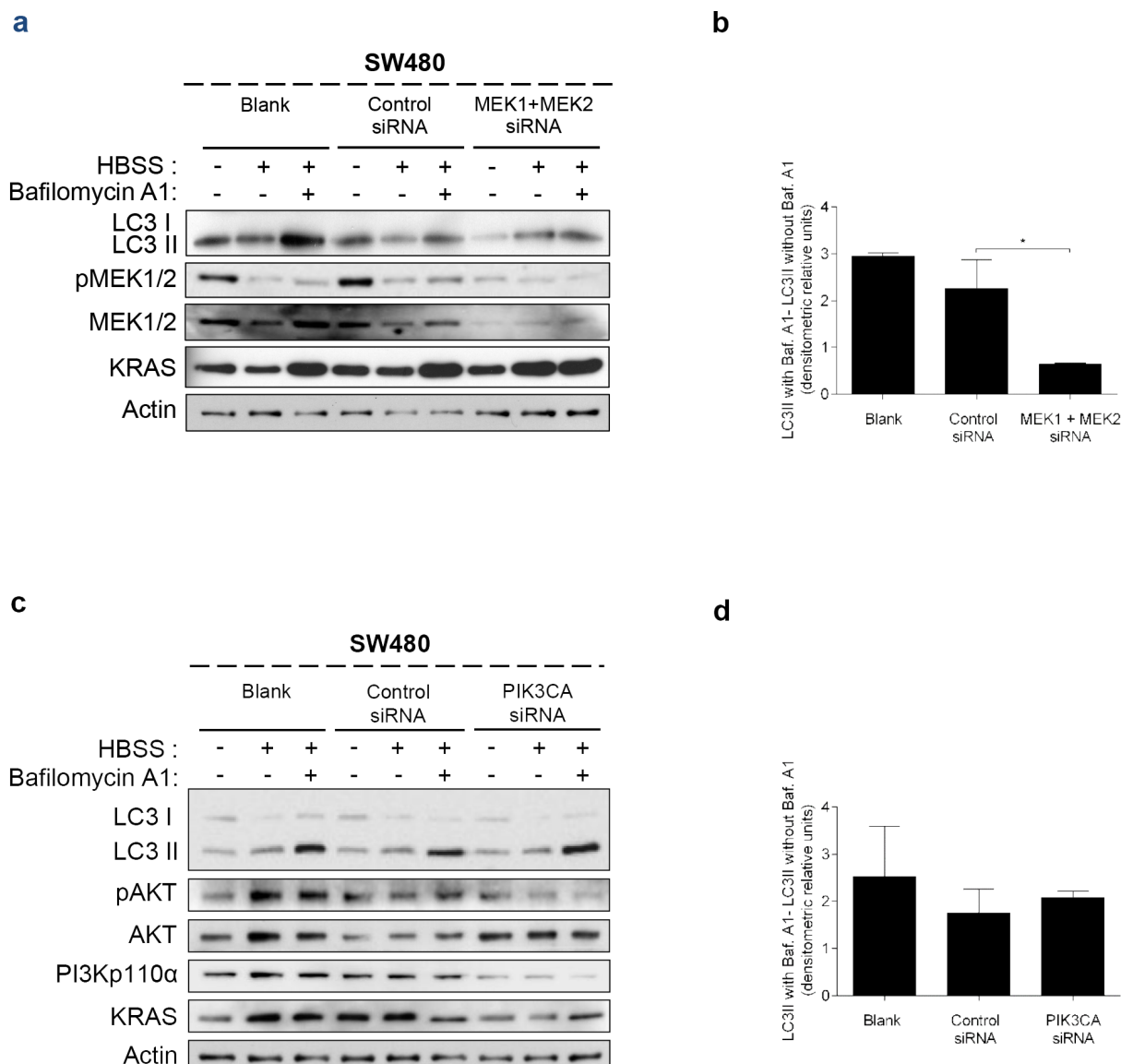


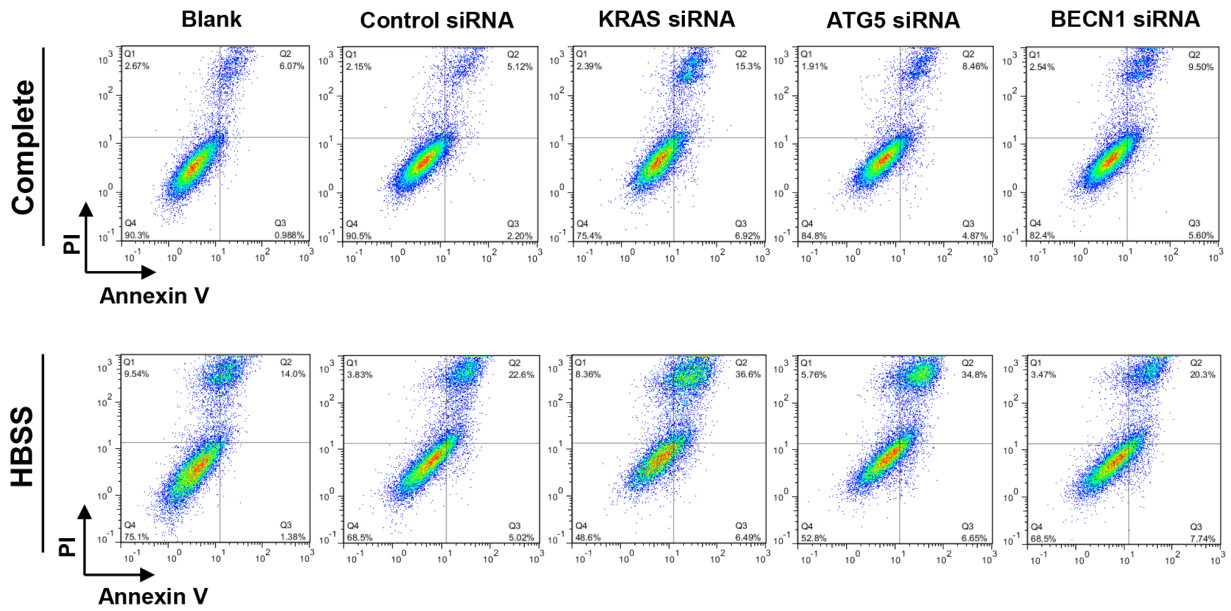
Figure 4: Suppression of MEK1+MEK2, but not of PIK3CA, decreases autophagy in SW480 cells. Immunoblot analysis of LC3-I/II in SW480 cells, after suppression of **a.** *MEK1* and *MEK2* and **c.** *PIK3CA*. Cells were left non transfected (blank) or transfected with either control siRNA, siRNA targeted against *MEK1* and *MEK2* or siRNA targeted against *PIK3CA*. 48 h after transfection, the cells were maintained in complete medium or incubated in HBSS medium (starvation) for further 6 h and cell lysates were analyzed by immunoblot for the indicated proteins. MEK1/2 and PI3Kp110α immunoblot were used to show the efficient silencing by siRNA *MEK1/2* and siRNA *PIK3CA*, respectively. Actin immunoblot was used as a loading control. **b.** **d.** LC3-II/Actin ratios determined using *ImageJ* were used to calculate the autophagic flux by subtracting the normalized LC3-II levels in the absence of Baf A1 from the corresponding levels obtained in the presence of Baf A1. Results are means ± SEM of two independent experiments. **P* < 0.05 one-way ANOVA followed by Tukey's test.

to a different extent depending on the mutant *KRAS* allele, since expression of *KRAS*^{G13D} and *KRAS*^{G12D} mutations up-regulated autophagy in NCM460 cells facing nutrient limitation, whereas wild-type *KRAS* or *KRAS*^{G12V} did not. In our “*KRAS* humanized yeast” system, *KRAS*^{G13D} and *KRAS*^{G12D} also induced autophagy in response to nitrogen starvation while wild-type *KRAS* did not. However, *KRAS*^{G12V} also highly up-regulated autophagy in this cell model. The lack of the genetic complexity found in human regulatory pathways and the lack of interference

from wild-type *KRAS* and other human *RAS* family isoforms may explain the stronger phenotype observed in yeast compared with the human non-cancer colon cell model. Nonetheless, these results show that yeast is a particularly useful organism to address the function of *KRAS* in autophagy and that can be further explored to address other specific cellular functions of *KRAS* or *KRAS* localization within cellular compartments.

The results obtained in the yeast system show that expression of mutant *KRAS per se* can drive autophagy.

a



b

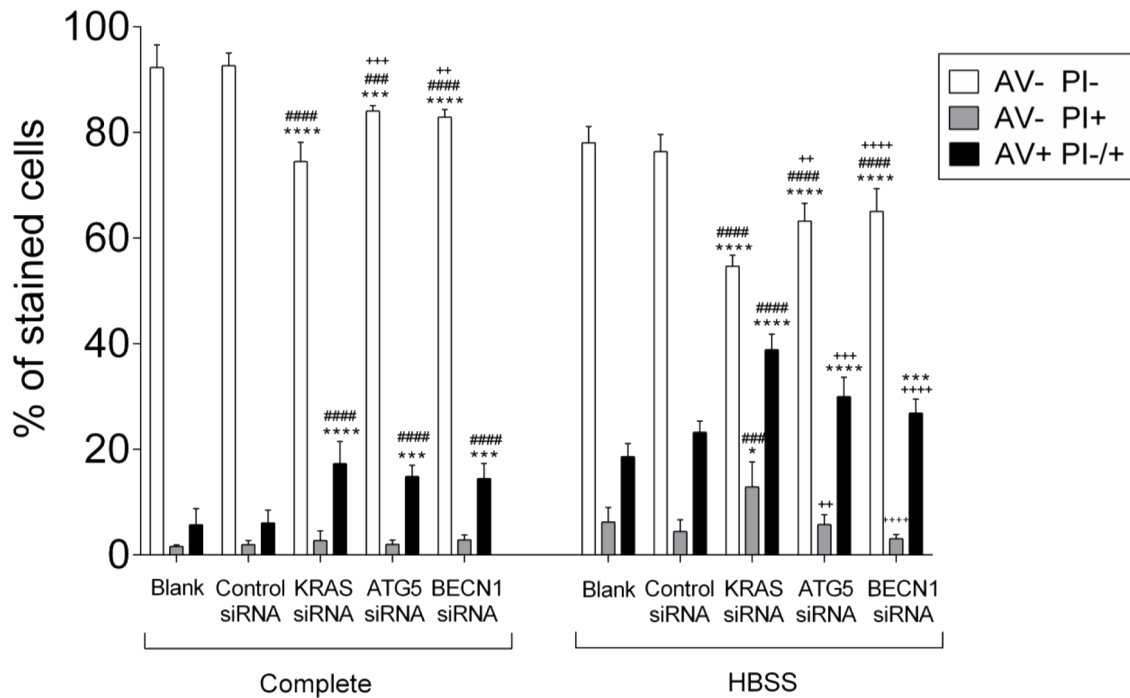


Figure 5: Suppression of *KRAS*, *ATG5* and *BECN1* promotes cell death in SW480. Cell death determined by Annexin V fluorescein isothiocyanate (AV-FITC) and propidium iodide (PI) assay in SW480 cells that were left non transfected (blank) or transfected with control siRNA, siRNA targeted against *KRAS*, *ATG5* or *BECN1*. 24 h after transfection, cells were maintained in complete medium or incubated in HBSS medium (starvation) for another 24 h. Cells were fixed and double stained with Annexin V and PI before analyzed by flow cytometry. **a.** Representative histograms of SW480 cells double-labeled with AV and PI. **b.** Quantitative analysis of AV/PI staining in SW480 cells. Percentages of apoptotic cells (AV+) are the sum of the lower and upper right panels (AV+ PI-/). Values represent mean \pm S.E.M. of at least three independent experiments. * $P < 0.05$; ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$ comparing between each subset of cells (AV-PI-, AV-PI+, AV+ PI-/). * vs Blank; # vs Control siRNA; + vs KRAS siRNA. Two-way ANOVA.

However, the fact that expression of KRAS^{G12V} in NCM460 cells was not sufficient to up-regulate autophagy to the same extent as KRAS^{G13D} and KRAS^{G12D}, suggests other factors may also play a role, depending on the cellular background. Accordingly, recent studies reported that induced expression of HRAS^{G12V} and KRAS^{G12V} can up-regulate autophagy in several cell models, such as mice kidney iBMK [11], breast MCF10A epithelial cells [15, 16], ovarian HOSE epithelial cells, MCF-7 cells and HUVECs cells [12]. However, it had not been addressed whether inhibition of this mechanism has an effect in tumor cells. We determined that depletion of KRAS by siRNA impairs the autophagic flux of CRC- derived cell lines HCT116 and SW480, harboring KRAS^{G13D} and KRAS^{G12V} mutations, respectively, indicating that autophagy of CRC cells harboring KRAS mutations depends on KRAS.

The best-characterized RAS effector pathways involved in autophagy regulation are the MEK/ERK and the PI3K/AKT/mTOR signaling pathways [33], which are commonly deregulated in CRC [36, 37]. However, it is not known if/how KRAS modulates these effectors to regulate autophagy in CRC. Here, we demonstrate that KRAS-induced autophagy is associated with up-regulation of ERK phosphorylation during nutrient limitation conditions in a non-cancer colon NCM460 cell model, consistent with a pro-autophagic role of MEK/ERK signaling [12, 38]. In accordance, depletion of KRAS by siRNA in the two CRC- derived cell lines used decreased activation of ERK. We further show that activation of the MEK/ERK pathway is critical for autophagy in SW480 cells, since inhibiting MEK1/2 also significantly impaired this process. Overall, these results indicate that the MEK/ERK effector pathway is essential for KRAS-induced autophagy in CRC cells. However, activation of this pathway does not seem to be sufficient, since expression of KRAS^{G12V} in NCM460 cells induced phosphorylation of ERK, like KRAS^{G12D} and KRAS^{G13D}, but did not up-regulate autophagy to a higher extent than wild-type KRAS, indicating KRAS may function through additional downstream effectors.

Overexpression of either KRAS variant in NCM460 cells had the opposite effect on phosphorylation of AKT during nutrient limitation conditions, which tended to decrease. However, there was no correlation with autophagy induction; indeed, this decrease was most significant in cells overexpressing KRAS^{WT} and KRAS^{G12V}, which did not up-regulate autophagy. In agreement with the results obtained in the NCM460 model, depletion of KRAS in SW480 cells (KRAS^{G12V} mutation) increased AKT activation during starvation. Since depletion of KRAS in this cell line decreased autophagy, this could suggest an anti-autophagic role of PI3K/AKT/mTOR signaling in CRC [13]. However, the results obtained in the HCT116 cell line, which displays constitutive activation of the AKT pathway due to an activating PIK3CA^{H1047R} mutation, argue against this hypothesis. Indeed, on one hand

autophagy is still induced in these cells; moreover, it is also decreased by depletion of KRAS, which is upstream of PI3K/AKT/mTOR. As expected, depletion of KRAS did not affect AKT activation in the HCT116 cell line, not only because these cells display constitutive activation of the AKT pathway, but also because overexpression of FLAG-KRAS^{G13D} in NCM460 cells did not affect pAKT levels. Overall, our results indicate the PI3K/AKT/mTOR pathway is not relevant in KRAS-induced autophagy. Accordingly, we found that depletion of PIK3CA did not affect autophagy induction of SW480 cells in response to starvation.

The interplay between RAS and autophagy on the survival of cancer cells is complex, as it may have both pro- and anti-tumorigenic roles. Previous studies demonstrated that expression of HRAS^{G12V} in MEFs and HRAS^{G12V} or KRAS^{G12V} in iBMK cells promotes cell proliferation through up-regulation of autophagy [11, 16, 39]. Others showed that HRAS^{G12V}-induced autophagy contributes to cell death in HOSE, MCF-7 and HUVEC cells [12]. In our study, we sought to understand the effect of KRAS/autophagy inhibition on the cell death of CRC cells, which would attest to the clinical relevance of this process. We showed that depletion of KRAS, ATG5 or BECN1 results in cell death during growth of SW480 cells in complete medium, but to a higher extent in starvation conditions. These results indicate that autophagy mediated by activating KRAS mutations has a pro-survival role in CRC cells under stressful conditions, like nutrient deprivation.

EGFR inhibitors have been widely used in anti-cancer therapy; however, they are inefficient in mCRCs harboring activating KRAS mutations, which remains an obstacle to the effective treatment of mCRC [4–8]. Despite continuous efforts, no successful therapy targeting KRAS has been developed and it remains an elusive target for cancer therapy. This is due to the lack of specificity of KRAS inhibitors and the fact that KRAS can overcome inhibition of these targeted therapies [40, 41]. Direct inhibition of KRAS using small interference RNA (siRNA) may be an alternative approach to the use of chemical inhibitors and our results support this hypothesis. However, *in vivo* delivery issues of siRNA have yet to be overcome for widespread clinical applications, although efforts are being made to develop siRNA therapeutic approaches using lipid nanoparticle delivery systems [42, 43]. An alternative approach is to target downstream KRAS signaling. Autophagy is implicated as a potential mechanism of resistance to anticancer agents, as it aids in the response of tumor cells to cellular stress and/or increased metabolic demands related to increased cell proliferation [44]. Our results suggest that inhibition of autophagy can be an alternative or possibly combined therapeutic approach for the treatment of CRCs harboring activating KRAS mutations that depend on autophagy for survival. Indeed, studies in colon cancer xenografts

using autophagy inhibitors in combination with chemotherapeutic agents showed that autophagy inhibition increases apoptosis accompanied by tumor regression [45]. Moreover, current clinical trials use the autophagy inhibitor hydroxychloroquine for treatment of a wide range of cancers, with promising results [46–50].

In summary, the results presented here contribute to a deeper understanding of the survival role of mutant *KRAS* alleles in CRC cells. We show that activating *KRAS* mutations increase autophagy in different cell models, and that this increase is mediated through activation of the MEK/ERK pathway in colon cells. We also demonstrate that inhibition of *KRAS* or autophagy can promote cell death in CRC cells harboring a *KRAS* mutation, highlighting *KRAS*-induced autophagy as an emerging target for CRC therapy.

MATERIALS AND METHODS

Cell lines and culture conditions

Cell lines used in this study were the noncancerous NCM460 cell line [18] and CRC-derived cell lines SW480 (*KRAS*^{G12V} mutation) and HCT116 (*KRAS*^{G13D} and *PIK3CA*^{H1047R} mutations) [51] (ATCC). SW480 cells were grown in RPMI 1640 medium with glutamine (PAA, Austria) supplemented with 1% penicillin–streptomycin and 10% fetal bovine serum (FBS). HCT116 cells were grown in McCoy's 5A (PAA, Austria) supplemented with 2 mM L-glutamine, 1% penicillin–streptomycin and 10% FBS. NCM460 cells were grown in M3:10TM (INCELL, San Antonio, TX). All cell lines were cultured in a humidified incubator with 5% CO₂ at 37°C. To assess basal protein levels, cells were cultured in complete medium and collected at 80% of confluence. For autophagy induction, cells growing in complete medium were washed three times with 1x PBS and incubated for 6 hours in Hank's Buffered Salt Solution (HBSS) buffered with 2.2 g/l NaHCO₃. Bafilomycin A1 was added to inhibit lysosomal degradation in order to analyze p3XFLAG-FLAG-KRAS protein levels (at 20 nM) or autophagic proteolysis (at 100 nM).

Production of lentivirus containing *KRAS* genes and lentiviral infection

The human wild-type *KRAS* mRNA sequence was cloned into a pLenti6/V5 Directional TOPO[®] (Invitrogen), giving rise to the *KRAS*^{WT} expression vector. *KRAS*^{G13D}, *KRAS*^{G12D} or *KRAS*^{G12V} mutant vectors were obtained by site-directed mutagenesis using specific primer sequences for each mutation. These vectors were used as templates to subclone wild-type and *KRAS* mutant sequences into the p3XFLAG-CMV vector to create FLAG-KRAS constructs before generating HIV-1 lentivirus-based vectors. A 5' *NheI* site and a 3' *PstI* site were

introduced in the FLAG-KRAS sequence by PCR using p3XFLAG-CMV KRAS as a template and primers 5'-T TGTAGCATGGACTACAAAAGACCATGACGG-3' and 5'-TTTCTGCAGCTACATAATTACACACTTTGTC TT TGAC-3'. The PCR product was sub-cloned between the *NheI* and *PstI* sites in pRRLSIN.cPPT.PGK.MSC.WPRE lentivector. Correct integration of FLAG-KRAS constructs was confirmed by sequencing. HEK-293FT cells were used as packaging cells, and virus production was as previously described [52]. NCM460 cells were infected with the lentiviral constructs pRR/FLAG-KRAS in complete medium, and cells collected after 5 days of infection. Infection with FLAG-KRAS was confirmed by immunofluorescence and by immunoblot of protein extracts, using an anti-FLAG primary antibody (Supplementary Figures S6a and S6b). All experiments using these cells were performed between the 4th and 8th weeks after infection. Population doubling determination showed a stable growth of cells with constitutive expression of FLAG-KRAS^{WT}, -KRAS^{G13D}, -KRAS^{G12D} or -KRAS^{G12V} (Supplementary Figure S6c).

RNA silencing

HCT116 and SW480 cells were transiently transfected with siRNA using Lipofectamine 2000 (Invitrogen), according to manufacturer recommendations. siRNAs used were: *KRAS* siRNA (Hs_KRAS2_8), *ATG5* siRNA (Hs_APG5L_6) and *BECN1* siRNA (Hs_BECN1_5) designed by Qiagen; and *MAP2K1* siRNA (L-003571-00-0010), *MAP2K2* siRNA (L-003573-00-0010) and *PIK3CA* siRNA (L-003018-00-0010), designed by Dharmacon.

Determination of long-lived protein degradation

Parental NCM460 cells and NCM460 cells infected with FLAG-KRAS constructs were labeled for 24 hours in complete medium containing 0.1 μCi L-[¹⁴C]valine/ml/well. After 24 hours, cells were washed three times with 1x PBS and radioactivity was pre-chased for 1 hour in complete medium in the presence of an excess of L-valine (10 mM) to remove the contribution of short-lived protein degradation. Cells were washed three times with 1x PBS, incubated for 6 hours either in complete medium or in HBSS in the presence or absence of Baf. A1 (0,1 μM) and with an excess of L-valine (10 mM). Supernatants were collected and free amino acids precipitated with 80% trichloroacetic acid (TCA), while proteins in adherent cells were precipitated with 10% TCA. The two fractions were transferred to vials containing 3 ml OptiPhase HiSafe II scintillation liquid (LKB Scintillation Products). Radioactivity was quantified in a liquid scintillation counter Packard Tri-Carb 2200 CA (Packard Instrument). Proteolysis was calculated as a percentage of the radioactivity in the supernatant relative to the total cell radioactivity.

Annexin V/PI assay

SW480 cells were maintained in complete medium or incubated with HBSS medium. After 24 h of treatment, both suspended and attached cells were collected and washed in 1x PBS. 1×10^6 cells were resuspended in 100 μ l 1x binding buffer and incubated with 5 μ l AV-fluorescein isothiocyanate (BD Biosciences, San Jose, CA, USA) and 10 μ l 50 μ g/ml PI for 15 min in the dark. Samples were analyzed in an Epics[®] XL[™] (Beckman Coulter) flow cytometer, equipped with an argon-ion laser emitting a 488-nm beam at 15 mW. Monoparametric detection of red fluorescence was performed using FL-4 (488/675 nm) and detection of green fluorescence was performed using FL-1 (488/525 nm). 20 000 cells were analyzed per sample and data analyzed using FlowJo software (version 7.6, Tree Star Inc., Ashland, OR, USA).

Yeast plasmids

KRAS^{WT}, *KRAS^{G13D}*, *KRAS^{G12D}* or *KRAS^{G12V}* were subcloned as a *Bam*HI-*Apa*I fragments from pLenti6/V5-D-TOPO (Invitrogen) and inserted between the *Bam*HI and *Apa*I sites of pCM184 (*TRP1*) yeast plasmid, under the control of a *tet*-off promoter (repressed by the addition of doxycycline). The GFP-Atg8p (*URA3*) construct, for expression of *ATG8* under the control of its endogenous promoter in the pRS416 plasmid, was previously described [26].

Yeast strains

The *Saccharomyces cerevisiae* mutant strain W303-1B *ras2* Δ *pho8* Δ 60 was generated by transformation of parental strain W303-1B *pho8* Δ 60 [53] with a disruption cassette amplified by PCR from genomic DNA of the BY4741 *ras2* Δ strain (Euroscarf). Correct integration of the cassette was confirmed by colony PCR. W303-1B *ras2* Δ *pho8* Δ 60 was co-transformed with pCM184/*KRAS* variants and pGFP-Atg8, and the resulting transformants grown in selective media lacking the appropriate amino acids. *KRAS* expression was confirmed by immunoblot against *KRAS* (Supplementary Figure S6d). No differences in growth of *S. cerevisiae ras2* Δ cells transformed either with empty vector or pCM184/*KRAS* variants were observed (Supplementary Figure S6e).

Growth conditions

Yeast cells were grown aerobically in synthetic complete medium [SC; 0.17% yeast nitrogen base w/o amino acids and ammonium sulphate (Difco), 0.5% ammonium sulphate, 0.1% potassium phosphate, 0.2% Drop-out Mix and 0.01% auxotrophic requirements, pH 5.5] supplemented with 2% glucose in the presence of 10 μ g ml⁻¹ doxycycline. Cells were harvested, washed three times with water and resuspended in fresh medium without doxycycline to induce *KRAS* expression. For *KRAS* repression assays, 10 μ g/

ml doxycycline was added to the cultures to inhibit *KRAS* transcription. For nitrogen starvation assays, cells were grown in SC media without doxycycline until exponential phase, washed twice with distilled sterile water and suspended in nitrogen starvation media [SD-N; 0.17% yeast nitrogen base w/o amino acids and ammonium sulphate (Difco)] supplemented with 2% glucose for 24 h. All incubations were performed at 30°C, 200 r.p.m.

Immunoblot analysis

Preparation of total protein extracts of human cell lines was performed as described in ref. [54]. Preparation of total protein extracts of yeast cells, SDS-PAGE and Western blots were performed as previously described [55].

Antibodies

Antibodies used were: anti-actin, anti-Atg5, anti-LC3 and anti-FLAG (Sigma); anti-*KRAS*, and anti-yeast Atg8p (Santa-Cruz Biotechnology); anti-Becn1, anti-phospho p44/42 MAPK (Thr202/Tyr204), anti-p44/42 total, anti-phospho Akt (Ser473), anti-Akt total, anti-PI3Kp110 α , anti-phospho MEK1/2 and anti-MEK1/2 total (Cell Signalling); anti-yeast phosphoglycerate kinase (Pgk-1) (Molecular Probes); and mouse monoclonal GFP (1:3000; Roche Applied Science).

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CONFLICTS OF INTEREST

The authors declare no conflict of interest.

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REFERENCES

1. Ferlay J, Shin H.R, Bray F, Forman D, Mathers C, Parkin D.M. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer*. 2010; 127:2893–2917.

2. Merla A, Goel S. Novel drugs targeting the epidermal growth factor receptor and its downstream pathways in the treatment of colorectal cancer: a systematic review. *Chemotherapy research and practice*. 2012; 2012: 387172.
3. Leto S.M, Trusolino L. Primary and acquired resistance to EGFR-targeted therapies in colorectal cancer: impact on future treatment strategies. *J Mol Med (Berl)*. 2014; 92:709–722.
4. Raponi M, Winkler H, Dracopoli N.C. KRAS mutations predict response to EGFR inhibitors. *Curr Opin Pharmacol*. 2008; 8:413–418.
5. Lievre A, Bachet J.B, Boige V, Cayre A, Le Corre D, Buc E, Ychou M, Bouche O, Landi O, Louvet C, Andre T, Bibeau T, Diebold M.D, Rougier P, Ducreux M, Tomasic G, Emile J.F, Penault-Llorca F, Laurent-Puig P. KRAS mutations as an independent prognostic factor in patients with advanced colorectal cancer treated with cetuximab. *Journal of clinical oncology : official journal of the American Society of Clinical Oncology*. 2008; 26:374–379.
6. Di Fiore F, Blanchard F, Charbonnier F, Le Pessot F, Lamy A, Galais M.P, Bastit L, Killian A, Sesboue R, Tuech J.J, Queuniet A.M, Paillot B, Sabourin J.C, Michot F, Michel P, Frebourg T. Clinical relevance of KRAS mutation detection in metastatic colorectal cancer treated by Cetuximab plus chemotherapy. *Br J Cancer*. 2007; 96:1166–1169.
7. Benvenuti S, Sartore-Bianchi A, Di Nicolantonio F, Zanon C, Moroni M, Veronese S, Siena S, Bardelli A. Oncogenic activation of the RAS/RAF signaling pathway impairs the response of metastatic colorectal cancers to anti-epidermal growth factor receptor antibody therapies. *Cancer Res*. 2007; 67:2643–2648.
8. Freeman D.J, Juan T, Reiner M, Hecht J.R, Meropol N.J, Berlin J, Mitchell E, Sarosi I, Radinsky R, Amado R.G. Association of K-ras mutational status and clinical outcomes in patients with metastatic colorectal cancer receiving panitumumab alone. *Clin Colorectal Cancer*. 2008; 7:184–190.
9. Chen N, Karantza V. Autophagy as a therapeutic target in cancer. *Cancer Biol Ther*. 2011; 11:157–168.
10. Yang Z, Klionsky D.J. Mammalian autophagy: core molecular machinery and signaling regulation. *Curr Opin Cell Biol*. 2010; 22:124–131.
11. Guo J.Y, Chen H.Y, Mathew R, Fan J, Strohecker A.M, Karsli-Uzunbas G, Kamphorst J.J, Chen G, Lemons J.M, Karantza V, Coller H.A, Dipaola R.S, Gelinas C, Rabinowitz J.D, White E. Activated Ras requires autophagy to maintain oxidative metabolism and tumorigenesis. *Genes Dev*. 2011; 25:460–470.
12. Elgandy M, Sheridan C, Brumatti G, Martin S.J. Oncogenic Ras-induced expression of Noxa and Beclin-1 promotes autophagic cell death and limits clonogenic survival. *Mol Cell*. 2011; 42:23–35.
13. Furuta S, Hidaka E, Ogata A, Yokota S, Kamata T. Ras is involved in the negative control of autophagy through the class I PI3-kinase. *Oncogene*. 2004; 23:3898–3904.
14. Kim J.H, Kim H.Y, Lee Y.K, Yoon Y.S, Xu W.G, Yoon J.K, Choi S.E, Ko Y.G, Kim M.J, Lee S.J, Wang H.J, Yoon G. Involvement of mitophagy in oncogenic K-Ras-induced transformation: overcoming a cellular energy deficit from glucose deficiency. *Autophagy*. 2011; 7:1187–1198.
15. Kim M.J, Woo S.J, Yoon C.H, Lee J.S, An S, Choi Y.H, Hwang S.G, Yoon G, Lee S.J. Involvement of autophagy in oncogenic K-Ras-induced malignant cell transformation. *J Biol Chem*. 2011; 286:12924–12932.
16. Lock R, Roy S, Kenific C.M, Su J.S, Salas E, Ronen S.M, Debnath J. Autophagy facilitates glycolysis during Ras-mediated oncogenic transformation. *Mol Biol Cell*. 2011; 22:165–178.
17. Guo J.Y, Karsli-Uzunbas G, Mathew R, Aisner S.C, Kamphorst J.J, Strohecker A.M, Chen G, Price S, Lu W, Teng X, Snyder E, Santanam U, Dipaola R.S, Jacks T, Rabinowitz J.D, White E. Autophagy suppresses progression of K-ras-induced lung tumors to oncocytomas and maintains lipid homeostasis. *Genes & development*. 2013; 27:1447–1461.
18. Moyer M.P, Manzano L.A, Merriman R.L, Stauffer J.S, Tanzer L.R. NCM460, a normal human colon mucosal epithelial cell line. *In Vitro Cell Dev Biol Anim*. 1996; 32:315–317.
19. Pereira C, Coutinho I, Soares J, Bessa C, Leao M, Saraiva L. New insights into cancer-related proteins provided by the yeast model. *FEBS, J*. 2012; 279:697–712.
20. Silva R.D, Saraiva L, Coutinho I, Goncalves J, Corte-Real M. Yeast as a powerful model system for the study of apoptosis regulation by protein kinase C isoforms. *Curr Pharm Des*. 2012; 18:2492–2500.
21. He C, Klionsky D.J. Regulation mechanisms and signaling pathways of autophagy. *Annu Rev Genet*. 2009; 43:67–93.
22. Yamamoto A, Tagawa Y, Yoshimori T, Moriyama Y, Masaki R, Tashiro Y. Bafilomycin A1 prevents maturation of autophagic vacuoles by inhibiting fusion between autophagosomes and lysosomes in rat hepatoma cell line. H-4-II-E cells. *Cell Struct Funct*. 1998; 23:33–42.
23. Fuertes G, Martin De Llano J.J, Villarroya A, Rivett A.J, Knecht E. Changes in the proteolytic activities of proteasomes and lysosomes in human fibroblasts produced by serum withdrawal, amino-acid deprivation and confluent conditions. *Biochem, J*. 2003; 375:75–86.
24. Mizushima N, Yoshimori T, Levine B. Methods in mammalian autophagy research. *Cell*. 2010; 140:313–326.
25. Kirisako T, Baba M, Ishihara N, Miyazawa K, Ohsumi M, Yoshimori T, Noda T, Ohsumi Y. Formation process of autophagosome is traced with Apg8/Aut7p in yeast. *J Cell Biol*. 1999; 147:435–446.

26. Shintani T, Klionsky D.J. Cargo proteins facilitate the formation of transport vesicles in the cytoplasm to vacuole targeting pathway. *J Biol Chem.* 2004; 279:29889–29894.
27. Xie Z, Klionsky D.J. Autophagosome formation: core machinery and adaptations. *Nat Cell Biol.* 2007; 9:1102–1109.
28. Mehrpour M, Esclatine A, Beau I, Codogno P. Overview of macroautophagy regulation in mammalian cells. *Cell Res.* 2010; 20:748–762.
29. Schubbert S, Shannon K, Bollag G. Hyperactive Ras in developmental disorders and cancer. *Nat Rev Cancer.* 2007; 7:295–308.
30. Karnoub A.E, Weinberg R.A. Ras oncogenes: split personalities. *Nat Rev Mol Cell Biol.* 2008; 9:517–531.
31. Koeffler H.P, McCormick F, Denny C. Molecular mechanisms of cancer. *West J Med.* 1991; 155:505–514.
32. Pylayeva-Gupta Y, Grabocka E, Bar-Sagi D. RAS oncogenes: weaving a tumorigenic web. *Nat Rev Cancer.* 2011; 11:761–774.
33. Barault L, Veyrie N, Jooste V, Lecorre D, Chapusot C, Ferraz J.M, Lievre A, Cortet M, Bouvier A.M, Rat P, Roignot P, Faivre J, Laurent-Puig P, Piard F. Mutations in the RAS-MAPK, PI(3)K (phosphatidylinositol-3-OH kinase) signaling network correlate with poor survival in a population-based series of colon cancers. *Int J Cancer.* 2008; 122:2255–2259.
34. Jung C.H, Ro S.H, Cao J, Otto N.M, Kim D.H. mTOR regulation of autophagy. *FEBS letters.* 2010; 584:1287–1295.
35. Prior I.A, Lewis P.D, Mattos C. A comprehensive survey of Ras mutations in cancer. *Cancer Res.* 2012; 72:2457–2467.
36. Velho S, Oliveira C, Ferreira A, Ferreira A.C, Suriano G, Schwartz S, Schwartz S Jr, Duval A, Carneiro F, Machado J.C, Hamelin R, Seruca R. The prevalence of PIK3CA mutations in gastric and colon cancer. *Eur J Cancer.* 2005; 41:1649–1654.
37. Oliveira C, Velho S, Moutinho C, Ferreira A, Preto A, Domingo E, Capelinha A.F, Duval A, Hamelin R, Machado J.C, Schwartz S Jr, Carneiro F, Seruca R. KRAS and BRAF oncogenic mutations in MSS colorectal carcinoma progression. *Oncogene.* 2007; 26:158–163.
38. Pattingre S, Bauvy C, Codogno P. Amino acids interfere with the ERK1/2-dependent control of macroautophagy by controlling the activation of Raf-1 in human colon cancer HT-29 cells. *J Biol Chem.* 2003; 278:16667–16674.
39. Yang S, Wang X, Contino G, Liesa M, Sahin E, Ying H, Bause A, Li Y, Stommel J.M, Dell’antonio G, Mautner J, Mautner G, Haigis M, Shirihai O.S, Doglioni C, Bardeesy N, Kimmelman A.C. Pancreatic cancers require autophagy for tumor growth. *Genes Dev.* 2011; 25:717–729.
40. Vasan N, Boyer J.L, Herbst R.S. A RAS renaissance: emerging targeted therapies for KRAS-mutated non-small cell lung cancer. *Clinical cancer research : an official journal of the American Association for Cancer Research.* 2014; 20:3921–3930.
41. de Castro Carpeño J, Belda-Iniesta C. KRAS mutant NSCLC, a new opportunity for the synthetic lethality therapeutic approach. *Translational Lung Cancer Research.* 2013; 2:142–151.
42. Bora R.S, Gupta D, Mukkur T.K, Saini K.S. RNA interference therapeutics for cancer: challenges and opportunities (review). *Molecular medicine reports.* 2012; 6:9–15.
43. Wan C, Allen T.M, Cullis P.R. Lipid nanoparticle delivery systems for siRNA-based therapeutics. *Drug Deliv. and Transl. Res.* 2014; 4:74–83.
44. Degenhardt K, Mathew R, Beaudoin B, Bray K, Anderson D, Chen G, Mukherjee C, Shi Y, Gelinas C, Fan Y, Nelson D.A, Jin S, White S. Autophagy promotes tumor cell survival and restricts necrosis, inflammation, and tumorigenesis. *Cancer Cell.* 2006; 10:51–64.
45. Li J, Hou N, Faried A, Tsutsumi S, Kuwano H. Inhibition of autophagy augments 5-fluorouracil chemotherapy in human colon cancer *in vitro* and *in vivo* model. *Eur J Cancer.* 2010; 46:1900–1909.
46. Rosenfeld M.R, Ye X, Supko J.G, Desideri S, Grossman S.A, Brem S, Mikkelsen T, Wang D, Chang Y.C, Hu J, McAfee Q, Fisher J, Troxel A.B, Piao S, Heitjan D.F, Tan K.S, Pontiggia L, O’Dwyer P.J, Davis L.E, Amaravadi R.K. A phase I/II trial of hydroxychloroquine in conjunction with radiation therapy and concurrent and adjuvant temozolomide in patients with newly diagnosed glioblastoma multiforme. *Autophagy.* 2014; 10:1359–1368.
47. Rangwala R, Leone R, Chang Y.C, Fecher L.A, Schuchter L.M, Kramer A, Tan K.S, Heitjan D.F, Rodgers G, Gallagher M, Piao S, Troxel A.B, Evans T.L, DeMichele A.M, Nathanson K.L, O’Dwyer P.J, Kaiser J, Pontiggia L, Davis L.E, Amaravadi R.K. Phase I trial of hydroxychloroquine with dose-intense temozolomide in patients with advanced solid tumors and melanoma. *Autophagy.* 2014; 10:1369–1379.
48. Vogl D.T, Stadtmauer E.A, Tan K.S, Heitjan D.F, Davis L.E, Pontiggia L, Rangwala R, Piao S, Chang Y.C, Scott E.C, Paul T.M, Nichols C.W, Porter D.L, Kaplan J, Mallon G, Bradner J.E, Amaravadi R.K. Combined autophagy and proteasome inhibition: a phase 1 trial of hydroxychloroquine and bortezomib in patients with relapsed/refractory myeloma. *Autophagy.* 2014; 10:1380–1390.
49. Rangwala R, Chang Y.C, Hu J, Algazy K.M, Evans T.L, Fecher L.A, Schuchter L.M, Torigian D.A, Panosian J.T, Troxel A.B, Tan K.S, Heitjan D.F, DeMichele A.M, Vaughn D.J, Redlinger M, Alavi A, Kaiser J, Pontiggia L, Davis L.E, O’Dwyer P.J, Amaravadi R.K. Combined mTOR and autophagy inhibition: phase I trial of hydroxychloroquine and temsirolimus in patients with advanced solid tumors and melanoma. *Autophagy.* 2014; 10:1391–1402.
50. Mahalingam D, Mita M, Sarantopoulos J, Wood L, Amaravadi R.K, Davis L.E, Mita A.C, Curiel T.J, Espitia C.M, Nawrocki S.T, Giles F.J, Carew J.S. Combined autophagy and HDAC inhibition: a phase I safety, tolerability, pharmacokinetic, and pharmacodynamic analysis of hydroxychloroquine in combination with the HDAC inhibitor vorinostat

- in patients with advanced solid tumors. *Autophagy*. 2014; 10:1403–1414.
51. Ahearn I.M, Haigis K, Bar-Sagi D, Philips M.R. Regulating the regulator: post-translational modification of RAS. *Nat Rev Mol Cell Biol*. 2012; 13:39–51.
 52. Qin X.F, An D.S, Chen I.S, Baltimore D. Inhibiting HIV-1 infection in human T cells by lentiviral-mediated delivery of small interfering RNA against CCR5. *Proceedings of the National Academy of Sciences of the United States of America*. 2003; 100:183–188.
 53. Kissova I, Deffieu M, Manon S, Camougrand N. Uth1p is involved in the autophagic degradation of mitochondria. *J Biol Chem*. 2004; 279:39068–39074.
 54. Preto A, Figueiredo J, Velho S, Ribeiro A.S, Soares P, Oliveira C, Seruca R. BRAF provides proliferation and survival signals in MSI colorectal carcinoma cells displaying BRAF(V600E) but not KRAS mutations. *J Pathol*. 2008; 214:320–327.
 55. Camougrand N, Grelaud-Coq A, Marza E, Priault M, Bessoule J.J, Manon S. The product of the UTH1 gene, required for Bax-induced cell death in yeast, is involved in the response to rapamycin. *Mol Microbiol*. 2003; 47:495–506.