Candidal colonization, strain diversity, and antifungal susceptibility among adult diabetic patients

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Ann Saudi Med 2010; 30(2): 101-108

DOI: 10.4103/0256-4947.60514

BACKGROUND AND OBJECTIVES: Candidal colonization in diabetics is a matter of debate. The aim of this study is to investigate oral candidal colonization, strain diversity, antifungal susceptibility, and the influence of local and systemic host factors on candidal colonization in adult diabetics.

METHODS: We conducted a case-control study that compared 150 diabetics (49 type 1, 101 type 2) with 50 healthy controls. Two salivary samples were collected, using the oral rinse sampling method: one for salivary flow rate and pH determination, and the other for candidal colonization assessment. The candidal isolates were identified and tested in vitro for antifungal susceptibility using the commercial kit, Candifast. The relationship between specific host factors and candidal colonization was also investigated.

RESULTS: Diabetics had a higher candidal carriage rate compared to controls, but not density. *Candida albicans* was the most frequently isolated species, but diabetics had a variety of other candidal species present. None of the control samples were resistant to any tested antifungal, while the diabetic samples had differing resistances to azole antifungals. Although there was a significant positive correlation between glycemic control and candidal colonization in type 2 diabetics, there was a negative correlation between salivary pH and candidal carriage in the controls versus density in type 2 diabetics.

CONCLUSIONS: Diabetic patients not only had a higher candidal carriage rate, but also a variety of candidal species that were resistant to azole antifungals. Oral candidal colonization was significantly associated with glycemic control, type of diabetes, and salivary pH.

ral candidiasis is a common opportunistic infection of the oral cavity caused by an overgrowth of candidal species, the most common being *Candida albicans.*¹ The underlying causes of oral candidiasis include extremes of age, xerostomia, antibiotic therapy, dentures, smoking, Cushing syndrome, malignancies, immune deficiencies, and diabetes mellitus (DM).^{1,2} The presence of *Candida* species and the density of candidal growth in the oral cavity is often said to be increased in patients with DM.^{3,4} However, these observations remain controversial.⁵ Similarly, there are conflicting reports on the identity of the underlying risk factors.³⁻⁵ These uncertainties are thought to be related to the different pathophysiological behaviors of the two clinical types of

DM, to different patients and disease data such as, age, duration or control of diabetes or, at least in part, are thought to reflect racial and environmental differences among diabetic populations worldwide.

Diabetes mellitus, specifically type 2 diabetes, is a major public health issue. The diabetic population is expected to increase from 171 million in 2000 to 366 million by 2030.⁶ World Health Organization statistics show that, worldwide, almost three million deaths per year are attributed to diabetes, equivalent to 5.2% of all deaths.⁷ DM is emerging as a major public health problem in Saudi Arabia, parallel with the worldwide diabetes pandemic. Early reports indicated that nearly one Saudi in five of age 30 years or older had DM,⁸ while the latest report showed that the overall prevalence of

DM among adult Saudis of both sexes in rural as well as urban communities is 23.7%.⁹ Despite these recognized high rates of DM among the Saudi population, there is an obvious lack of published studies evaluating the prevalence and characteristics of oral fungal infections, or investigating the underlying risk factors associated with DM, in Saudi Arabia. The aim of this study is to investigate oral yeast colonization (rate and density of acquisition of *Candida* species colonies), strain diversity, and antifungal susceptibility in adult diabetics, and to evaluate the influence of some local and systemic host factors on candidal colonization.

METHODS

This case-control study was conducted on 150 diabetic patients (49 type 1, 101 type 2) and 50 healthy controls. The diabetic patients were recruited from the diabetic clinics of King Abdul-Aziz University Hospital (KAAUH), Jeddah, Saudi Arabia, during routine diabetic follow-up appointments. Consecutive eligible patients identified on a specific sampling day were asked to participate. The inclusion criteria were a diagnosis of either type 1 or 2 DM and age of 18 years or older. The control group included age-grouped and sex-matched healthy volunteers (companions of the diabetic patients as well as dental auxiliaries) with no history of diabetes, who were selected from KAAUH. All subjects in both groups gave a signed informed consent to participate in the study. Individuals who had received antibiotics or steroid therapy or had been using antiseptic mouthwash during the prior three weeks were excluded from the study. The study protocol was approved by the ethics committee of the KAAUH medical faculty.

A structured questionnaire was developed for collecting information on demographics (age and gender), medical variables (diabetes type, duration, and presence of diabetes-related systemic diseases), and local factors (denture status, oral hygiene, and smoking). The investigators supervised the completion of the questionnaire, which had close-ended questions including options. The patient's medical records were used to gather information on diabetes type and duration, as well as, the presence of diabetes-related systemic complications (retinopathy, nephropathy, neuropathy, and peripheral vascular diseases). The presence of any of these complications was considered a positive finding and dichotomized on a scale of yes or no. All subjects provided three samples, two salivary and one blood sample, which were sent immediately to the KAAUH Microbiology and Hematology Laboratories, respectively. To prevent circadian variations, the samples were collected between 9 a.m. and 12 noon.

The subject was typically instructed not to eat, drink or smoke two hours prior to sample collection. They were asked to lean their upper body forward, and allow oral fluid to drip into a graduated collection vial, over a fiveminute period, without swallowing.¹⁰ This sample was used to calculate the salivary flow rate (mL/min) and to determine the saliva pH using the Combur test (Roche Diagnostics Ltd., UK), according to the manufacturer's instructions. Oral yeast colonization was assessed microbiologically in all participants, regardless of the presence of clinical infection. An oral rinse technique described by Samaranayake and colleagues¹¹ was used for sample collections. Each subject was supplied with 10 mL sterile phosphate-buffered saline (0.1 M PBS, pH 7.2) in a universal container, requested to remove dentures if worn, and to swirl the 10 mL of PBS around the mouth for 60 seconds before expectorating the saliva-buffer mixture back into the container. The sample was sent immediately to the microbiology laboratory and inoculated onto a Sabouraud's dextrose agar (SDA) plate, and incubated aerobically at 37°C for 48 hours. The growth of any candidal colonies was recorded as a positive growth and the subject as a candidal carrier. The number of colonies on each plate was counted and the number of colony-forming units (CFU) per mL calculated, to indicate candidal density. The Candida strains in the isolated colonies were identified and tested for susceptibility to amphotericin B, fluconazole, nystatin, flucytosine, econazole, ketoconazole, and miconazole antifungals, using the commercial Candifast kit (International Micobio, France) according to the manufacturer's instructions. Basically, the Candifast test tray consisted of two rows with eight wells. The first row (the identification row) contained seven different sugars, the fermentation of which produced a color change in the phenol red indicator. The first well of this row contained phenol red, actidione, and glucose. Interpretation help was given by a colored chart included in the kit. The second row was the susceptibility row. The first well of this row was a growth control well and contained glucose. Wells two to eight contained glucose and each with contained an antifungal agent. The wells were inoculated with standardized inocula and covered with two drops of paraffin oil. The tray was incubated at 37°C. After 24 and 48 hours, the wells containing antifungal drugs (row 2) were examined, and the isolate was classified as 'resistant' (medium color: yellow and/or visible turbidity and/or a pellet), 'intermediate' (medium color: yellow-orange) or 'susceptible' (medium color:

red). The minimal inhibitory concentrations (MICs) corresponding to these three categories were not provided. Additionally, the identification of *Candida* species was confirmed by the conventional methods of germ tube production in horse serum,¹² following incubation at 35°C for two hours.

For diabetic patients, the blood sample was used to measure glycosylated hemoglobin concentrations (HbA_{1c}), which assessed the long-term glycemic control of that patient. For control subjects, the blood samples were used to measure the fasting plasma glucose level. Those with values of 7 mmol/mL or higher were excluded from the study, according to the WHO definition of diabetes.¹³

RESULTS

The diabetic and control groups were homogenous in terms of age, sex, dental status, and smoking habits (P>.05; Table 1). However, the control group showed a

higher tooth brushing frequency than the diabetic group (P=.001). Also, there were no statistically significant differences between the two types of diabetic patients in any of the above-mentioned variables, nor in disease history, salivary flow rate or pH, except for the age, brushing frequency, and glycemic control (Tables 1 and 2). Type 2 diabetics were older than type 1 diabetics and had both a lower brushing frequency and glycosylated hemoglobin levels in comparison to type 1 diabetics [mean (SD) HbA_{1c}=8.95 (1.78) compared to 10.06 (2.06), respectively]. The diabetics had statistically, significantly lower salivary pH values compared to the controls (P=.015). However, there were statistically no significant differences in the levels of the salivary flow rates among the groups (Table 2, Figure 1).

The carriage rate or the frequency of detecting positive candidal growth was significantly higher in the diabetic patients than in the controls (P=.028; Table 2, Figure 2). The diabetic patients who were candidal

	Diabetics	Controls		Diabetics		
	(n=150)	(n=49)	Р*	Type 1 (n=49)	Type 2 (n=101)	Р*
Age (years)						
<30	12.7	14.0		32.7	3.0	
30-50	37.3	52.0	.128	30.6	40.6	.001
>50	50.0	34 .0		36.7	56.4	
Sex						
Male	41.3	38.0	.678	34.7	44.6	.250
Female	58.7	62.0		65.3	55.4	
Brushing frequency						
<1/day	16.0	4.1		12.8	17.5	
Once/day	37.5	18.4	.001	36.2	38.1	.673
>2/day	46.5	77.6		51.1	44.3	
Dental status						
Dentate	80.0	86.0	.346	84.8	77.8	220
Denture wearer	20.0	14.0		15.2	22.2	.326
Smoker						
Yes	10.1	10.0	.98	12.8	8.9	47
No	89.1	90.0	.90	87.2	91.1	.47

Table 1. Characteristics of the study population

Values are percentages; *Chi-square test

carriers harbored the yeast in higher densities than the control carriers, but the difference was not statistically significant (Figure 3). The mean candidal density for diabetic patients in this study was 3140.74 CFU/mL (SD=11388.76). The number of patients with candidal carriage from the oral cavity was higher in patients with type 1 diabetes than in type 2 (P=.003). Similarly, the yeast density was higher in type 1 diabetics than in samples from type 2 diabetics, and the differences between the two groups were statistically significant (P=.002).

In both diabetic patients and healthy controls, the

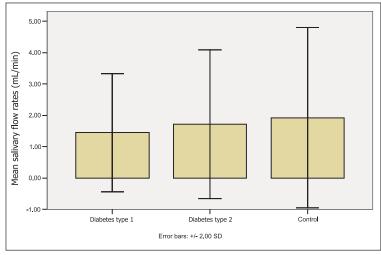


Figure 1. Salivary flow rates among the groups.

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most frequently isolated yeast was *Candida albicans* (68.9% and 40%, respectively). The remaining isolates were found more frequently in DM patients as compared to the controls, including *C. glabrata* (11.1%), *C parapsilosis* (6.7%), *C krusei* (4.4%), *C tropicalis* (2.2%), and other yeast species (6.7%) (Table 3). The in vitro antifungal susceptibility testing revealed that the yeast isolated from the diabetic patients had different rates of resistance to the seven tested antifungal drugs, except amophotericin B and nystatin, against which they had no resistance (Table 4). In contrast, in the healthy controls, none of the isolated yeast showed any resistance to the tested antifungal agents. When patients with different types of DM were compared,

 Table 3. Distribution of candidal isolates in diabetics and healthy controls.

Oradidal ana	Disketise	Ocutuala	Diabetic patients		
Candidal spp.	Diabetics	Controls	Type 1	Type 2	
C albicans	31 (68.9)	2 (40.0)	14 (63.6)	17 (73.9)	
C parapsilosis	3 (6.7)	0 (0.0)	1 (4.5)	2 (8.7)	
C glabrata	5 (11.1)	1 (20.0)	3 (13.6)	2 (8.7)	
C krusei	2 (4.4)	0 (0.0)	2 (9.1)	0 (0)	
C tropicalis	1 (2.2)	0 (0.0)	1 (4.5)	0 (0)	
Other yeast spp	3 (6.7)	2 (40.0)	1 (4.5)	2 (8.7)	

Values are numbers and percentages

	Diabetics	Controls	Diabetic P		patients	Р
	Diabetics	CUIITUIS	r	Type 1	Type 2	r
Candidal carriage						
Positive %	33.3	14.3	.028	51.2	25	.003
Negative %	66.7	85.7		48.8	75	
Candidal density (CFU/mL)						
Median (IQR)	0.0 (2000)	0.0 (0.0)	.200	1000 (3000)	0.0 (750)	.002
Salivary flow rates (mL/ mints)						
Median (IQR)	1.5 (1)	1.5 (1.13)	.479	1.0 (1.3)	1.5 (1)	.195
pH						
Median (IQR)	7 (2)	7 (1.0)	.015	7 (1)	7 (2)	.059

IQR: Interquartile range

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there was statistically no significant difference in the antifungal susceptibility (P>.05).

Statistical analysis showed no significant relationship between either the rate or density of candidal carriage and age, sex, brushing frequency, denture status, or smoking habits in any of the studied groups (P>.05). Similarly, medical and dental parameters such as disease duration, presence of diabetes complications, and salivary flow rates did not correlate significantly with the Candida carriage rate or density (P>.05). However, the rate of candidal carriage correlated significantly with the degree of glycemic control (as determined by the HbA1c values) in type 2 diabetics and the salivary pH levels in healthy controls (Table 5). The candidal carriage rate correlated positively with the Hb_{A1c} values in type 2 diabetics, but negatively with the pH values in the healthy controls. In the type 2 diabetics, the candidal density correlated positively with the HbA₁ values (Pearson correlation=0.212; P=.047), and negatively with the salivary pH values (Pearson correlation=-0.338; *P*=.001).

DISCUSSION

A review of the literature published to date, on the relationship between DM and oral candidal infection reveals a continuing debate. Studies on candidal colonization are often contradictory, which may be a result of the variety of sampling techniques employed.¹⁴⁻¹⁸ The present study comprehensively investigated microbiologically oral yeast colonization and evaluated the effect of some local and systemic factors, which could potentially influence the candidal carriage rate and density in diabetic patients. The reason for not conducting an oral examination for the clinical diagnosis of Candida was that it could be counted microbiologically without manifesting clinically and this was mentioned in the literature. A significant proportion of patients with no clinical evidence of oral candidiasis had >100 CFU/mL of yeast in their mouth rinses, suggesting that even an abundance of organisms in saliva might not initiate candidal infection.⁵ The oral rinse sampling method was used as it was the most appropriate and sensitive technique for evaluating the overall yeast carriage compared to imprint culture, swab or saliva sampling.¹¹ The percentage of diabetic candidal carriers was nearly double that of healthy controls, confirming previous reports that candidal species were more prevalent in the oral cavity of diabetic patients than in healthy individuals.^{4,14,17} In addition, the prevalence rates for candidal species in the present study, of 33.3% in diabetics compared with only 14.3% in the healthy controls, are similar to those in the previous studies,

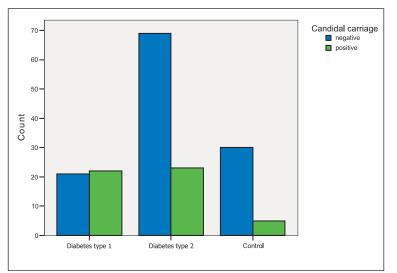


Figure 2. Candidal carriage among the groups.

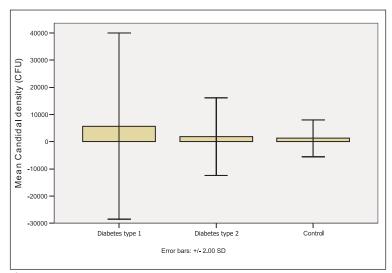


Figure 3. Candidal density among the groups.

which reported a range of 18 to 80% for diabetic patients and 3 to 47% for healthy individuals.^{5,14} The mean candidal density for the current diabetic isolates was 3140.74 CFU/mL, a value also within the range of candidal density reported among diabetic adult populations.^{3,19}

Overall, our results herein agree with the other published results using the same sampling techniques. Diabetic patients have a higher prevalence of oral candidal carriage rate, but not density, compared to non-diabetics.^{14,19,20} Although candidal density seems to be a useful predictor for the development of oral candidiasis in diabetic patients, previous studies have shown that candidal density, regardless of the methods

 Table 4. In vitro susceptibility to antifungal agents.

Antifungol	Diabetics	Controlo	Diabetic patients		
Antifungal	Dianetics	Controls	Type 1	Type 2	
Amphotericin B					
S	37 (100)	2 (100)	21 (100)	16 (100)	
R	0 (0)	0 (0)	0 (0)	0 (0)	
Nystatin					
S	37 (100)	2 (100)	21 (100)	16 (100)	
R	0 (0)	0 (0)	0 (0)	0 (0)	
Flucytosine					
S	31 (86.1)	2 (100)	17 (81)	14 (93.3)	
R	5 (13.9)	0 (0)	4 (19)	1 (6.7)	
Fluconazole					
S	25 (71.4)	2 (100)	14 (73.7)	11 (68.8)	
R	10 (28.6)	0 (0)	5 (26.3)	5 (31.3)	
Ketoconazole					
S	30 (85.7)	2 (100)	17 (85.)	13 (86.7)	
R	5 (14.3)	0 (0)	3 (15)	2 (13.3)	
Miconazole					
S	24 (68.6)	2 (100)	14 (73.7)	10 (62.5)	
R	11 (31.4)	0 (0)	5 (26.3)	6 (37.5)	
Econazole					
S	19 (73.1)	1 (100)	11 (73.3)	8 (72.7)	
R	7 (26.9)	0 (0)	4 (26.3)	3 (27.3)	

Values are numbers and percentages; S: sensitive, R: resistance

 Table 5. Factors potentially influencing candidal growth in type 2 diabetics.

	Positive candid	al growth	Negative candidal growth		
	Type 2 diabetics	Controls	Type 2 diabetics	Controls	
HbA _{1c}	9.90 (1.88) * <i>P</i> =.006	NA	8.72 (1.65) * <i>P</i> =.006	NA	
Salivary pH	6.8 (0.91) * <i>P</i> =.139	6.40 (0.55) * <i>P</i> =.003	7.10 (0.82) * <i>P</i> =.139	7.37 (0.63) * <i>P</i> =.003	

Data are mean (SD); NA= not applicable, *ANOVA

used for sampling, does not correlate with the clinical evidence of oral candidiasis.²¹ The higher carriage rate that we found in our study may be a better predictor for candidal infection in diabetics, but the question needs further investigation. The present study also agrees with the previously published reports that oral yeast colonization is higher in type 1 diabetics than in type 2,^{4,15} although some investigators have failed to show a correlation between candidal carriage rates and density or the type of DM and the treatment used.^{14,18,22}

C albicans was the most frequently isolated candidal species among the study groups, even as 31.1% of diabetic patients also carry other yeasts and candidal species. The latter include C glabrata (11.1%1), C parapsilosis (6.7%), C krusei (4.4%), and C tropicalis (2.2%). These findings are not surprising, as these organisms are the most common candidal species isolated from humans.²³ A similar diversity in candidal species has been reported previously among diabetic populations in Thailand,²⁴ Poland,²⁵ and Brazil.²⁶

The most common antifungal drugs in the current clinical use, for treatment of oral candidiasis are polyenes (amphotericin B and nystatin) and azoles (miconazole, fluconazole, ketoconazole, and itraconazole), mainly used topically.²⁷ The therapeutic and prophylactic use of antifungal agents has given rise to alarming cases of antifungal resistance.²⁸ Interestingly, in vitro antifungal susceptibility tests revealed that none of the candidal isolates from the control group showed any resistance to the seven tested antifungals. However, oral yeasts isolated from diabetic patients displayed different resistance rates from the five azole antifungal agents, mainly miconazole and fluconazole. Such findings confirmed the results of other researchers reporting the emergence of fluconazole and, more generally, triazole resistance among different groups of immunocompromized patients in whom these agents were frequently used.²⁹ A recent in vitro study using the commercial Fungitest kit revealed no difference in susceptibility between candidal isolates from diabetics and non-diabetics to the six common antifungal agents tested.²⁷ However, they reported increased miconazole resistance among diabetic patients in London, which they justified as a consequence of the past widespread use of miconazole for the treatment of Candida induced denture stomatitis. In recent times, a number of rapid and easy-to-use commercial products have been introduced that identify yeast, and test antifungal susceptibility. The Candifast kit used in this study proved to be reliable for the identification of yeasts, but it had low reliability in antifungal susceptibility testing,

which was attributed to high levels of subjectivity in the result interpretations.³⁰ However, the objective of this study was not to report accurate and clinically significant susceptibility, but to investigate whether there was an increase in in vitro antifungal resistance in the diabetic isolates. Therefore, this study should be extended to include more precise standardized tests with clear-cut objectives and interpretative criteria. Overall, based on the above findings we can say that the accurate identification of strains isolated from diabetic patients was especially important, because they were more likely to carry species other than C albicans, which might not be sensitive to certain antifungal agents. In addition the culture and sensitivity testing would add to the value of selecting the appropriate antifungal drugs, rather than prescribing any type of antifungal just for the presence of clinical manifestations of candidal infection.

The risk factors for oral candidal infection in diabetic patients are complex. Some authors emphasize the role of local factors such as low salivary flow rates¹⁹ and pH,15 smoking,321 wearing dentures15,22,31 and poor oral hygiene,²² others focus on systemic factors such as patient age²² or disease types,⁴ degree of control,^{15,31} and the presence of complications.³¹ Overall, our results demonstrate that oral yeast carriage rate and density are not influenced significantly by patient age, gender, disease duration or complications. No statistically significant relationship was found between smoking habits, wearing dentures, brushing frequency, salivary flow rates and carriage rates or the density of candidal isolations. However, glycemic control, as determined by the level of glycosylated hemoglobin, positively correlated with the increased carriage rate and density of Candida isolates in type 2 diabetics. This may explain

the higher carriage rates reported among type 1 diabetics compared to type 2 diabetics, because the former had higher mean HbA_{1c} values and poorer glycemic control. Hyperglycemia could contribute to the risk of oral *Candida* infection by increasing salivary glucose levels, which may promote overgrowth of *Candida*.³¹ The results reveal that decreased salivary pH correlates with an increased candidal carriage rate in healthy controls, and increased candidal density in type 2 diabetics. Interestingly, although similar findings regarding salivary pH and glycemic control have been reported,¹⁵ they have failed to show a significant difference in the candidal carriage rates or density between diabetics and non-diabetic controls.

In conclusion, diabetic patients had a higher oral candidal carriage rate, but not density, compared to non-diabetic controls. Although *C albicans* was the predominant isolate, a variety of other candidal species, with less susceptibility to azole antifungals, were identified in diabetics. Oral candidal colonization was significantly associated with diabetic type, glycemic control, and salivary pH, demonstrating a potential role of these factors in controlling candidal infections.

Aknowledgments

The research group acknowledges, with thanks, the scientific research council of King AbdulAziz University, (KAAU) which granted us the financial support during this research project (No. 424/061). Our sincere thanks also goes to the staff in KAAU hospital for their help and assistance in particulars the doctors at diabetic clinics and technicians in the laboratories. Finally special thanks go to Professor Hala Amer for valuable reviewing and editing of the research manuscript.

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