



Sperm quality is not affected by the BNT162b2 mRNA SARS-CoV-2 vaccine: results of a 6–14 months follow-up

Gilad Karavani^{1,2} · Henry H. Chill³ · Cherut Meirman⁴ · Einat Gutman-Ido² · Shmuel Herzberg^{1,2} · Tachover Tzipora^{1,2} · Tal Imbar^{1,2} · Assaf Ben-Meir^{1,2}

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Abstract

Purpose We aimed to investigate the possible effect of SARS-CoV-2 vaccination on sperm quality by evaluating semen analyses of men prior to vaccination and 6–14 months after vaccination.

Methods This was a retrospective cohort study, conducted in a university-affiliated in vitro fertilization center between October 2021 and March 2022, including men not previously infected with the SARS-CoV-2 virus who received at least 2 doses of the Pfizer-BioNTech (BNT162b2) SARS-CoV-2 vaccine. Semen analyses of samples given pre-vaccination and 6–14 months post-vaccination were analyzed for the parameters of volume, concentration, motility, morphology, and total motile count (TMC) and compared. These parameters were also compared separately for men who received a third (booster) dose and for men with pre-vaccination normal and abnormal sperm. Correlations between time from vaccination and post-vaccination sperm parameters were also assessed.

Results Fifty-eight men were included in the final analysis. Semen volume (2.9 ± 1.4 vs. 2.9 ± 1.6 ml), sperm concentration (42.9 ± 37.9 vs. 51.5 ± 46.2 million/ml), motility (42.5 ± 23.1 vs. 44.3 ± 23.4 percent), morphology (8.8 ± 16.6 vs. 6.6 ± 8.8 percent), and TMC (55.7 ± 57.9 vs. 71.1 ± 77.1 million) were comparable between the pre- and post-vaccination samples. This was true for the entire study cohort, for the subgroup of men who received a third dose and for the subgroups of men with a pre-vaccination normal and abnormal semen samples. No correlation was found between time from vaccination and post-vaccination sperm parameters.

Conclusions The Pfizer-BioNTech (BNT162b2) SARS-CoV-2 vaccine does not impair any of the sperm parameters over a relatively long-time interval of 6 to 14 months from vaccination.

Keywords Fertility · Sperm parameters · COVID-19 · SARS-CoV-2 vaccine · BNT162b2 vaccine

Introduction

The COVID-19 pandemic was declared in March 2020, spreading rapidly throughout the world with devastating consequences, including short- and long-term morbidity and mortality [1]. One of the mechanisms in which the SARS-CoV-2 virus causes infection is binding to the angiotensin-converting enzyme 2 (ACE-2) through the spike glycoprotein [2]. As ACE-2 receptors are located on the blood-testis barrier, the male reproductive system is a potential target for the virus. This has promoted research on the effects of the SARS-CoV-2 virus on the male reproductive system. While majority of studies demonstrated the absence of SARS-CoV-2 RNA in the semen of infected men [3, 4], few studies [5, 6] showed a minor negative effect on sperm parameters within the first 2–3 months after infection.

Gilad Karavani and Henry H. Chill have equal contribution as first authors.

✉ Gilad Karavani
Giladk84@gmail.com

¹ Infertility and IVF Unit, Hadassah Ein-Kerem Medical Center and Faculty of Medicine, Hebrew University of Jerusalem, Jerusalem, Israel

² Department of Obstetrics and Gynecology, Hadassah Ein-Kerem Medical Center and Faculty of Medicine, Hebrew University of Jerusalem, Jerusalem, Israel

³ Division of Urogynecology, University of Chicago Pritzker School of Medicine, NorthShore University HealthSystem, Skokie, IL, USA

⁴ Department of Family Medicine, Rabin Medical Center, Tel Aviv & Dan districts, Clalit, Israel

The mRNA SARS-CoV-2 vaccine, introduced in late 2020, dramatically reduced COVID-19 infection and transmission with consequent decrease in hospitalization and death rates within the general population [7, 8]. Following the implementation of the mRNA SARS-CoV-2 vaccine, several studies were conducted to explore possible adverse outcomes of the vaccine on male fertility, confirming its safety on short-term follow-up [9–11].

Previous data focused on relatively short-term (under 6 months, mostly up to 3 months) outcomes [9, 11], while data on longer intervals between vaccination and semen evaluation remains scarce. Vaccine hesitancy is driven, among other factors, by unfounded fear of possible long-term adverse effects on fertility [12]. Moreover, safety results of long-term follow-up are essential to reassure men that have already been vaccinated. Therefore, long-term outcomes are necessary to encourage vaccine uptake and decrease hesitancy.

The aim of this study was to evaluate sperm parameters of non-infected men who received at least two Pfizer-BioNTech (BNT162b2) SARS-CoV-2 vaccines and had a sperm analysis prior to vaccination and 6–14 months post-vaccination.

Materials and methods

Study population

We performed a retrospective cohort study at a tertiary university teaching hospital between October 2021 and March 2022. Included were men, in the process of IVF treatment with a female partner, who received at least two Pfizer-BioNTech (BNT162b2) SARS-CoV-2 vaccines and had submitted a sperm specimen prior and after vaccination. Both men with normal and abnormal semen analysis were included in the study, to evaluate possible effect in both groups. Excluded were men with known azoospermia (absence of sperm cells in the semen), men who were tested positive for SARS-CoV-2 (rapid antigen or PCR testing) at any point and those for whom date of vaccination was unavailable.

Specimen collection and analysis

Sperm specimens were collected as part of our standard protocol of care for couples evaluated for infertility. Following abstinence for 2–3 days, each man provided a semen sample either at home or in our laboratory in a sterile cup by means of masturbation and delivered it within 60 min to our laboratory for evaluation by a single embryologist. For all patients, the second sperm specimen was collected at least 6 months after the first vaccine dose. Patients for which a third (booster) dose was administered were recorded as well.

Semen analysis included evaluation of the semen sample, including the use of the Mackler chamber (Sefi Medical

Instruments, Israel) for the assessment of sperm motility and concentration. The following semen parameters were evaluated—sample volume (milliliter (ml)), concentration (million/ml), motility (%), normal morphology (%), and total motile count (million). The classification of normal sperm parameters was according to the widely accepted WHO semen analysis reference range guidelines [13]. We analyzed two specimens from each sample to determine the average value of all sperm parameters: concentration, motility, and morphology.

Sperm morphology evaluation was done using Quick Stain (Biological Industries, Sartorius AG, Germany). The stained slides were prepared by the laboratory staff according to the manufacturer's instructions. The evaluation was performed at a final magnification of X400 (PLAN CN 40x/0.65 Ph2 OLYMPUS) by counting 100 cells. Spermatozoa classification as normal or abnormal morphologically was according to WHO criteria (13). A smear of 20 μ l 1:1 mix (stain and semen) was evaluated at a X400 magnification (PLAN CN 40x/0.65 Ph2 OLYMPUS) by counting at least 100 cells (unstained and red-stained cells).

Data collection and analysis

Information was extracted from the electronic medical record software used to manage all data within the IVF unit at our medical center. Data retrieved included demographic, past medical history, indication for IVF treatment, and semen analysis-related data—semen volume, sperm concentration (million/milliliter), motility, morphology, and total motile count pre- and post-vaccination. For patients with several sperm specimens, we used the latest specimen collected after vaccination.

These sperm parameters were compared pre-vaccination and post-vaccination in the entire cohort. A subgroup analysis for men who received a third dose (booster dose) was also performed. Additionally, a subgroup analysis for men with pre-vaccination normal and abnormal sperm was done, to test for possible differences in the vaccination effect according to the initial sperm quality. Linear correlation between the parameter of time from vaccination (days) and post-vaccination sperm parameters was also performed to assess possible gradual continuous effects.

Ethical approval

This study was approved by the Human Investigation Review Board of Hadassah Hebrew University Medical center (IRB 0216–22-HMO) and conforms to the provisions of the declaration of Helsinki.

Power analysis

The sample size power calculation for a paired *t* test was directed to detect an 8 million/ml difference (represents a 20% difference for an average concentration of 40 million/ml) in

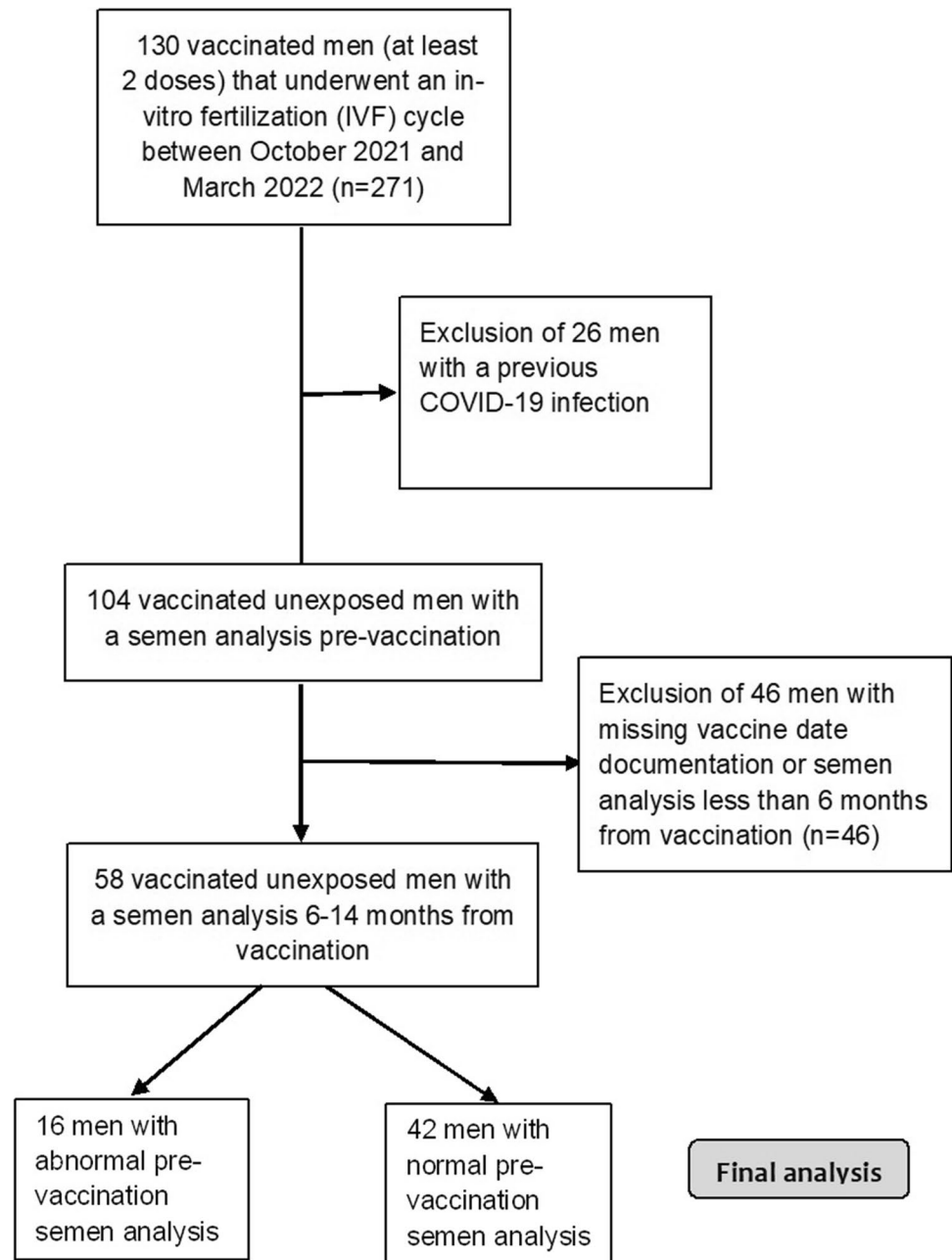
sperm concentration pre- and post-vaccination and achieve a significance level of 5% (two tailed) and power of 80%, with a SD of 20 million/ml. We calculated that a total of 52 patients would provide sufficient power to detect this difference with statistical significance.

Statistical analysis

For quantitative variables, the comparison between independent variables of the study groups was performed using Student's *t* test. Semen parameters pre- and post-vaccination were compared by Student's *t* test for paired samples (as each patient had a semen analysis pre- and post-vaccination).

Testing the association between two categorical variables was carried out using either the Chi-square test or the Fisher's exact test, as indicated. The Fisher's exact test was applied in analyses of small samples, when more than 20% of cells had expected frequencies of less than five. Cross-sectional association of the days from first vaccination to sperm parameters—volume, concentration, motility, morphology, and total motile count—were tested using the Pearson correlation coefficient ("r") in linear regression models. For all comparisons, a *p* value of < 0.05 was considered statistically significant. Statistical analysis was performed using the SPSS software (version 23; IBM Corp).

Fig. 1 Flow chart of study population



Results

In our IVF unit, 130 men underwent an Assisted Reproduction Technology (ART) cycle between October 2021 and March 2022. Out of these men, 104 were uninfected with the SARS-CoV-2 virus, and 85 of them had received at least 2 vaccination doses (3 weeks apart) with documented vaccination dates. Finally, 58 men who had pre- and post-vaccination semen analyses performed at least 6 months from the first vaccination were included in the study (Fig. 1).

The average age at pre-vaccination semen analysis was 38.0 ± 6.1 years (median of 38 years). Sixteen men (27.6%) were diagnosed with male factor infertility, thirteen of them had two abnormal parameters or oligoasthenoteratozoospermia. The average number of days from vaccination to post-vaccination semen analysis was 319.0 ± 61.4 days (median of 317 days). Thirteen men (22.4%) had post-vaccination semen analysis 6–9 months after vaccination, while for 45 men (77.6%), it was performed 9–14 months after vaccination. The basic characteristics and reproductive background of the study population are presented in Table 1. In the entire study population, the post-vaccination TMC parameter was higher in 27 men and lower in 28 men (due to missing data regarding motility in one of the semen analyses, TMC was not calculated for the remaining three men).

When comparing pre- and post-vaccination semen analyses in the entire cohort (Table 2), we found no difference in semen volume (2.9 ± 1.4 vs. 2.9 ± 1.6 ml, respectively; NS), concentration (42.9 ± 37.9 vs. 51.5 ± 46.2 million/ml, respectively; NS), motility (43.2 ± 22.6 vs. 44.1 ± 23.6 percent, respectively; NS), normal morphology (1.5 ± 2.0 vs. 3.8 ± 3.5 percent, respectively; NS), and total motile count (56.7 ± 58.0 vs. 71.1 ± 77.8 million, respectively; NS).

Similar results were demonstrated in a subgroup analysis including men who received three vaccine doses ($n=47$) (Table 2), except for the concentration parameter, which was higher post-vaccination (54.7 ± 47.5 vs. 42.0 ± 38.7 million/ml, $p=0.017$).

Between the pre- and post-vaccination semen analyses, one man had stopped receiving Copaxone treatment for multiple sclerosis (diagnosis was ruled out), one man received antibiotic treatment for possible genitourinary tract infection, and another started taking multivitamin. For all three, sperm parameters were slightly improved in the post-vaccination analysis.

An additional analysis was performed separately for men with normal sperm parameters ($n=42$) and for men ($n=16$) with abnormal sperm with similar results showing comparable semen parameters pre- and post-vaccination (Table 3).

When evaluating the linear correlation (Pearson correlation coefficient) between time from vaccination (days) and the parameters of semen volume, concentration, motility, and total motile count, we found no correlation with any of these parameters (Suppl. Table 1).

Discussion

In this study, we investigated semen analyses of vaccinated men 6–14 months after vaccination. We found no difference in sperm parameters pre- and post-vaccination in IVF patients treated with the Pfizer-BioNTech (BNT162b2) SARS-CoV-2 vaccine. Similar results were found for men who received a third vaccine dose (booster dose). All post-vaccination sperm specimens were collected at least 6 months after vaccination (median 317 days) enabling relatively long-term follow-up for the main outcome measures.

Our findings are in accordance with previous studies. Lifshitz et al. presented data from 75 fertile men 1–2 months after receiving their second vaccine dose showing that sperm parameters were within the WHO normal range, with less than 5% of abnormal parameters [9], though pre-vaccination semen samples were not available for comparison, as the

Table 1 Pre-vaccination characteristics of the study population: men vaccinated with at least two doses of SARS-CoV-2 mRNA vaccine without a previous exposure to the SARS-CoV-2 virus

Number of men	58
Age (years) pre vaccination	38.0 ± 6.1 (38.0)
Smoker	3/58 (5.2%)
Father to children	10/58 (17.2%)
Chronic disease or medication taken	4/58 (%)
Testicular trauma or infection	2/58 (3.4%)
Varicocele	8/58 (13.8%)
Testicular (varicocele/hydrocele/undescended testicle) or inguinal hernia surgeries	7/58 (12.1%)
Third vaccination given	47/58 (81.0%)
Testosterone levels (mIU/ml) ^a	16.3 ± 13.8 (15.1)
FSH levels ^a (mIU/ml) ^a	5.7 ± 2.9 (4.3)
LH levels ^a (mIU/ml) ^a	4.2 ± 1.5 (2.9)
Infertility diagnosis	
Male factor infertility	16/58 (27.6%)
Ovulation disorder	2/58 (3.4%)
Endometriosis	4/58 (6.9%)
Ovarian insufficiency/ age related infertility	11/58 (19.0%)
PGT	6/58 (10.3%)
RPL	2/58 (3.5%)
Female fertility preservation	2/58 (3.5%)
Mechanical factor	3/58 (5.2%)
Unexplained infertility	12/58 (20.7%)
Days from first vaccination to semen analysis	319.0 ± 61.4 (317.5)
Months from first vaccination to semen analysis	
6–9 months	13/58 (22.4%)
9–14 months	45 (77.6%)

Data presented as n/N(%) or mean \pm SD (median)

FSH, follicular stimulating hormone; LH, luteinizing hormone; PGT, preimplantation genetic testing; RPL, recurrent pregnancy loss

^aHormonal profile pre-vaccination was available for 18 patients

Table 2 Comparison of semen analysis results of men vaccinated with at least two doses of SARS-CoV-2 mRNA with semen samples pre-vaccination and post-vaccination

Parameter	Pre-vaccination	Post-vaccination*	P value
Entire cohort (<i>n</i> = 58)			
Semen parameters			
Volume (ml)	2.9 ± 1.4 (3.0)	2.9 ± 1.6 (3.0)	0.956
Concentration (million/ml)	42.9 ± 37.9 (33.5)	51.5 ± 46.2 (48.0)	0.060
Motility (%)	42.5 ± 23.1 (44.5)	44.3 ± 23.4 (43.0)	0.790
Normal morphology ^a (%)	8.8 ± 16.6 (4.0)	6.6 ± 8.8 (3.5)	0.132
Total motile count (million)	55.7 ± 57.9 (30.3)	71.1 ± 77.1 (39.5)	0.187
Men who received 3 vaccinations (<i>n</i> = 47)			
Semen parameters			
Volume (ml)	2.9 ± 1.5 (3.0)	2.9 ± 1.7 (3.0)	0.870
Concentration (million/ml)	42.0 ± 38.7 (30.0)	54.7 ± 47.5 (50.0)	0.017
Motility (%)	41.4 ± 21.9 (44.0)	44.0 ± 25.0 (43.7)	0.706
Normal morphology ^a (%)	6.2 ± 11.9 (3.0)	7.1 ± 9.2 (4.0)	0.054
Total motile count (million)	49.0 ± 53.1 (29.8)	71.0 ± 73.2 (44.3)	0.064

Data presented as mean ± SD (median)

^aMorphology data was available for 8 patients in the entire cohort and for 7 patients in the three vaccination group

authors mentioned. Similar results were noted in a study of 45 healthy volunteers scheduled for mRNA COVID-19 vaccination who provided semen samples prior to vaccination and 75 days after their second vaccine dose [10]. In a more recent study, semen parameters and fertilization rates were compared pre- and post-vaccination in patients undergoing ART. At a median follow-up of 75 days, no differences were noted between groups [11].

The process of spermatogenesis is a complex one which has been estimated to take approximately 74 days [14]. Theoretically, a vaccine administered during this time-period could instigate an immune response affecting

spermatozoa at different stages of maturity. However, the same response may target germ cells which have not yet entered spermatogenesis. In this study, we intentionally excluded patients who received their vaccine less than 6 months prior to semen sample collection. We believe this enabled us to better assess the true effect of vaccination on sperm parameters at all stages.

Our results are unique in that they focus on longer follow-up than previously reported. While most studies reported on outcomes up to 3 months post-vaccination, most of the men included in this study were vaccinated 9–14 months prior to the last semen sample collection. While

Table 3 Comparison of semen analysis results of vaccinated men with normal (*n* = 42) and abnormal (*n* = 16) semen parameters pre-vaccination and post-vaccination

Parameter	Pre-vaccination	Post-vaccination*	P value
Men with normal sperm			
Semen parameters			
Volume (ml)	2.9 ± 1.3 (2.9)	2.9 ± 1.5 (3.0)	0.743
Concentration (million/ml)	56.0 ± 36.6 (50.0)	66.8 ± 44.6 (65.0)	0.084
Motility (%)	51.1 ± 18.5 (52.0)	47.2 ± 22.7 (45.0)	0.274
Normal morphology ^a (%)	10.5 ± 17.8 (5.0)	7.9 ± 10.0 (4.0)	0.296
Total motile count (million)	74.2 ± 57.4 (49.5)	89.8 ± 79.6 (75.0)	0.275
Men with abnormal semen analysis			
Semen parameters			
Volume (ml)	2.8 ± 1.7 (3.3)	3.0 ± 1.8 (2.8)	0.630
Concentration (million/ml)	8.5 ± 8.1 (5.3)	11.5 ± 17.6 (4.4)	0.368
Motility (%)	18.8 ± 17.3 (16.0)	35.6 ± 24.3 (35.0)	0.053
Total motile count (million)	5.1 ± 5.8 (1.7)	15.6 ± 27.3 (3.8)	0.146

Data presented as mean ± SD (median)

^aMorphology data was available for 5 patients in the normal sperm group and for 3 in the abnormal sperm group (therefore not evaluated and not presented)

future studies will surely assess vaccine safety for longer follow-up periods, these results are encouraging for both providers and patients.

In the subgroup of men who received three vaccines, an increase in sperm concentration post-vaccination was detected. This finding might be explained, at least partially, by the three men in this group that received multivitamins or antibiotics or discontinued pharmacological treatment.

This study has several limitations. Firstly, all men participating in this study were part of couples undergoing IVF treatment and included men with male factor infertility, as opposed to studies that evaluated sperm donors. Secondly, data on sperm morphology, progressive motility assessment and DNA integrity and fragmentation were unavailable for part of our cohort. All subjects were vaccinated with the Pfizer BioNTech mRNA vaccine, limiting our conclusions to patients who received this specific formulation. Finally, we included patients with male factor infertility, and though each sperm analysis was paired, this may hamper our ability to draw conclusions to the general population.

Conclusions

Our study shows that the Pfizer-BioNTech (BNT162b2) SARS-CoV-2 vaccine does not negatively affect semen parameters, even in men with male factor infertility. We further found this to be true in patients who received a booster dose. The long-term follow-up presented here is key to establish vaccine safety and address patients' hesitancy towards vaccination and provide reassurance for vaccinated men.

Abbreviations *COVID-19*: Coronavirus disease; *SARS-CoV-2*: Severe acute respiratory syndrome coronavirus 2; *mRNA*: Messenger ribonucleic acid

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s10815-022-02621-x>.

Declarations

Conflict of interest The authors declare no competing interests.

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