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Scaling of thermal tolerance with body mass and genome size in ectotherms: a comparison between water- and air-breathers

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Global warming appears to favour smaller-bodied organisms, but whether larger species are also more vulnerable to thermal extremes, as suggested for past mass-extinction events, is still an open question. Here, we tested whether interspecific differences in thermal tolerance (heat and cold) of ectotherm organisms are linked to differences in their body mass and genome size (as a proxy for cell size). Since the vulnerability of larger, aquatic taxa to warming has been attributed to the oxygen limitation hypothesis, we also assessed how body mass and genome size modulate thermal tolerance in species with contrasting breathing modes, habitats and life stages. A database with the upper (CTmax) and lower (CTmin) critical thermal limits and their methodological aspects was assembled comprising more than 500 species of ectotherms. Our results demonstrate that thermal tolerance in ectotherms is dependent on body mass and genome size and these relationships became especially evident in prolonged experimental trials where energy efficiency gains importance. During long-term trials, CTmax was impaired in larger-bodied water-breathers, consistent with a role for oxygen limitation. Variation in CTmin was mostly explained by the combined effects of body mass and genome size and it was enhanced in larger-celled, air-breathing species during long-term trials, consistent with a role for depolarization of cell membranes. Our results also highlight the importance of accounting for phylogeny and exposure duration. Especially when considering long-term trials, the observed effects on thermal limits are more in line with the warming-induced reduction in body mass observed during long-term rearing experiments.

This article is part of the theme issue 'Physiological diversity, biodiversity patterns and global climate change: testing key hypotheses involving temperature and oxygen'.

1. Introduction

The capacity of organisms to take up and transform resources from their environment is a key attribute governing growth and reproduction, and subsequently affecting population dynamics, community composition and ecosystem functioning [1,2]. Such capacity seems to be mainly dictated by species' body mass [3]. Macroecological and palaeoecological data show spatial (e.g. *Bergmann's rule* [4,5]) and temporal (*Lilliput effect* [6]) variation in body mass, which share a common point related to the environmental temperature: at warmer, tropical latitudes and during the past mass extinctions, warming appears to select for smaller-bodied species [5,7–9]. Body size reductions with warming appear to be stronger in aquatic taxa than in terrestrial taxa

[5]. In tandem with body size reductions, both aquatic and terrestrial species are shifting their distribution towards cooler habitats and their phenology to earlier and hence cooler conditions [10,11]. One approach that has been taken to clarify the extent and variation in species redistributions, and to determine which taxonomic groups are potentially more vulnerable to the effects of climate change, is that of comparative studies that analyse thermal tolerance limits (upper and lower) synthesized from the literature [12–15]. These studies also highlight key differences in thermal responses between aquatic and terrestrial taxa, likely related to their breathing mode [16]. The physiological mechanisms underpinning size adjustments and thermal limits are actively debated [17–20], but oxygen limitation has been implicated for both thermal limits [21–23], and size adjustments [24–29] and hypoxia possibly also contributed to mass extinctions [8,30].

By affecting both oxygen demand [31] and the availability of oxygen in water [32,33], warming is hypothesized to result in oxygen limitation, which then causes reductions in thermal limits [22,34] and/or body mass [24,29]. As breathing underwater is more challenging than breathing in air, this oxygen-based mechanism could explain the divergent responses observed in air- and water-breathers [25]. While studies to date hint at a possible size-dependence of thermal limits, no studies have tested this possibility comprehensively. In fact, most studies have focused on one or a few species and although these studies often find no effect of body mass when included as a covariate in analyses, thermal tolerance limits (heat tolerance rather than cold tolerance) are more frequently reported to decrease rather than increase with increasing body mass [35–38]. In an effort to address this knowledge gap regarding how body mass modulates the response to the temperature in ectotherms, we took advantage of the large body of literature and created a database of upper and lower thermal limits supplemented with biological information of 510 species.

Larger-bodied species may be more susceptible to oxygen limitation because of their lower surface area to volume ratio, which (all else being equal) constrains their capacity to extract oxygen from their environment and deliver it to their metabolizing tissues [24,27,32], or because transport distances increase, which may be especially a problem if oxygen transport is based on diffusion [28]. If oxygen limitation plays a role in setting thermal limits, one prediction would be that thermal limits vary across organisms with distinct capacities to supply oxygen, including differences between water- and air-breathers, or between gas exchange systems across life stages. As body mass is intimately connected to a suite of other traits, size-dependency of thermal limits may be driven by traits related to body mass, rather than body mass *per se*. For example, relative to the larger adults, smaller life stages also may experience relatively cool early-season conditions, especially in temperate and polar regions with a clear seasonality, such that their thermal limits are shifted to lower temperatures, i.e. improved cold tolerance and impaired heat tolerance. In addition, organisms living in aquatic habitats will experience different thermal regimes from those living on land [15].

Variation in body mass can result from changes in cell number, cell size or a combination of both [39,40], but usually larger-bodied species tend to have larger cells, as documented in arthropods [40,41], fish [42] and birds and mammals [43]. The theory of optimal cell size [44] highlights how

differences in cell size have repercussions for oxygen uptake at the cellular level. In the same way, a diversity of cellular physiological functions scales with the cell size [45]. Therefore, differences in thermal tolerance between animals of different body mass may be mechanistically linked to differences in cell size, rather than body mass. In contrast to the hypothesized influence of oxygen limitation on heat tolerance, the evidence for such an influence on cold tolerance is rather limited [16], and these limits are thought to arise from membrane depolarization and subsequent cell dysfunction due to energy deficits or—in the case of extreme cold tolerance—the freezing of body fluids [46]. Thus, for cold tolerance, a cellular perspective may be more informative, although the correlation between cell size and body mass may result in size-dependency for CTmin.

In the present study, we use a global database of lower (CTmin) and upper (CTmax) critical thermal limits supplemented with information on other biological traits of ectotherm species and their phylogenetic relationships, to investigate whether and how the tolerance to high and low temperatures is modulated by the body mass and genome size (proxy for cell size) across arthropod and vertebrate (amphibian, fish and reptile) species have distinct breathing modes, life stages and habitats. We hypothesize that: (1) both CTmax and CTmin will be related to the body mass and genome size of the species, with thermal limits decreasing with increasing body mass (for CTmax) and with increasing genome size (for CTmin); (2) both CTmax and CTmin will differ across breathing modes and a species' habitat, and such differences will become more pronounced in large-bodied organisms or those with larger genomes; and (3) early life stages will be more susceptible to heat stress than their adult counterparts, and more resistant to cold stress.

2. Material and methods

(a) Data search

We created a global database of body mass-related traits (body mass and genome size), life stage (adult, juvenile and larva) and breathing mode (air-, bimodal- and water-breathers) of aquatic and terrestrial species belonging to four taxonomic groups (amphibians, arthropods, fish and reptiles) for which the critical thermal limits (upper and lower) have been evaluated using dynamic methods (i.e. CTmax or CTmin, *sensu* [47]). The chosen groups comprise taxa for which the determination of body mass was expected to be straightforward. We started by retrieving information from articles on body mass and thermal limits from existing quantitative reviews whose aim has been to explore global patterns of thermal tolerance in ectotherms [12,13,15]. We then added information from recently published references, from January 2015 to October 2018, which were found by using the following keyword combinations of Boolean terms through ISI Web of Science: (thermal tolerance OR heat tolerance OR cold tolerance OR upper thermal limit OR lower thermal limit OR thermal range OR CTmax OR CTmin) AND (body mass OR body size OR length) AND (amphib* OR arthrop* OR crustacea* OR fish* OR insect* OR reptil*). Searches were limited by research area (ecology, evolutionary biology, biodiversity conservation, environmental sciences, marine freshwater biology, physiology, entomology, zoology, biology, oceanography, fisheries, limnology, environmental studies, behavioural sciences, toxicology, water resources and multidisciplinary sciences) and research articles. To supplement our searches, we delved into the reference list of each paper to identify additional studies

missed in the initial search and, if necessary, asked corresponding authors for additional data not provided in the main text or electronic supplementary material.

(b) Inclusion criteria

CT_{max} and CT_{min} data established by a dynamic (or ramping) method were included in our database, which represents the most common metrics used to assess thermal tolerances in chosen taxa [48]. To account for methodological variation related to differences in starting temperatures and heating/cooling rates across species or studies, we calculated the exposure duration as a single metric that takes into account how long animals are exposed to thermal stress during the heating and cooling trials. After having merged the already published databases with the articles resulting from our search, all duplicates were removed and each article was screened and filtered to build our dataset based only on experimental studies on the basis of three main inclusion criteria: (i) mention of species name belonging to at least one of the four taxa selected (amphibians, arthropods, fish and reptiles), (ii) mention of body mass estimates as mass (wet or dry), width (carapace) or length (carapace, fork, intertegular, snout–vent, standard and total), and (iii) species candidates should be enlisted in the Open Tree of Life (<https://ot39.opentreeoflife.org>) for subsequent phylogenetic analyses (see §2c, Data analyses). Despite the restrictive nature of our criteria, just in a few cases, multiple articles reported data on thermal limits for the same species. For this, we prioritized the articles with the most information available, covering the largest number of entries in our database. Even so, if there were duplicates per species, we favoured those studies that (i) give both CT_{max} and CT_{min} estimations, over studies reporting only one thermal limit, (ii) mention the life stage used during the experiments, and (iii) mention methodological information such as cooling/heating rates, starting temperatures and geographical coordinates of collection. In the end, all these criteria allowed us to identify 510 species from 174 research articles providing thermal limits and body mass and phylogenetic information (electronic supplementary material, figure S1). For each species, we compiled taxonomic and biological information (life-stage, habitat, breathing mode, body mass and genome size), data on the site where a species was collected (geographical coordinates: latitude and longitude, and origin: laboratory or field), methodological information related to the estimation of the thermal limits (starting temperature, heating/cooling rates and acclimation time) and, finally, the CT_{max} and CT_{min} values.

All body size data collected in units other than mass were transformed using appropriate allometric relationships at the species level [49]; if this was not possible, we moved up to a higher taxonomic level (e.g. genus or family [50,51]), aiming to obtain a more representative unit of size for all species in the database, in this case the body mass in grams (g). As a proxy of cell size, we collected genome size data (in picograms, pg) from the Animal Genome Size Database [52] if it was available. The breathing mode was established on the basis of the species used in each experiment, through ‘expert judgement’ or consulting secondary references if necessary (e.g. [53]). Bimodal-breathers were classified either as water-breathers (for trials where they relied on underwater gas exchange) or air-breathers (for trials where they relied on aerial gas exchange). As most data concerned adults, with only few data for larvae and juveniles, these two categories were grouped as non-adults. Data from publications where CT_{max} or CT_{min} was not reported in the text or tables (i.e. presented only as figures) were extracted using the LibreOffice extension ‘OOdigitizer v1.2.1’.

(c) Data analyses

All the results presented in the paper, both in the main text and in the electronic supplementary material were based on linear

versions of phylogenetic generalized least-squares (PGLS) models. The correlation structure of these models was given by the potential similarity of species’ traits resulting from the shared evolutionary history and described by their phylogenetic signal using the Pagel’s lambda (λ) [54]. For this index, a value closer to 0 indicates non-phylogenetic signal (phylogenetic independence between species, or a star phylogeny) while a value closer to 1 indicates that species’ traits evolved randomly through evolutionary timescales (Brownian phylogeny) [55]. Such information, available as phylogenetic trees, was accessed following [56] and pruned to include only species present in our database. In addition to the estimation of phylogenetic signal in the model residuals, we tested for phylogenetic signal both in the dependent variables (i.e. the thermal limits) as well as in the independent variables of interest included in the main models following [57] (see electronic supplementary material, table S11).

Before the main analyses, we first performed preliminary PGLSs in order to determine whether methodological variables influence thermal limits within this dataset and needed to be included in the main analyses. For this, we tested whether the (1) species origin (laboratory or field), or (2) latitude of collection, or (3) acclimation time in the laboratory and the (4) time necessary to reach the CT_{max} and CT_{min} affected these thermal limits. The time was calculated after [58,59], as the relation between ramping rate (ΔT , in $^{\circ}\text{C min}^{-1}$) and the starting temperature (T_0) for CT_{max} as: $\text{time} = [\text{CT}_{\text{max}} - T_0] \times \Delta T^{-1}$, and for CT_{min} trials as: $\text{time} = [T_0 - \text{CT}_{\text{min}}] \times \Delta T^{-1}$. Out of these four methodological variables, only time and/or latitude showed the highest support and also had significant effects on the thermal limits (for CT_{max}: latitude and time, and for CT_{min}: only latitude) and these two were subsequently included as covariates in the main models (see electronic supplementary material, tables S1 and S2 and figure S7).

For the main analyses, we fitted PGLSs models to each of CT_{max} and CT_{min}, first with body mass (\log_{10} -transformed body mass) as an independent numerical variable, and breathing mode (air and water), life stage (adult and non-adult) and habitat (aquatic, intertidal and terrestrial) as categorical variables. We also ran models that included all possible interactions of these categorical variables and body mass. In a similar, second set of models, we used genome size (\log_{10} -transformed genome size) instead of body mass. Since we did not have a reliable estimate of genome size for all 510 taxa, the models using genome size were based on a smaller set of species, and hence model performance cannot be compared directly between models based on body mass and those based on genome size.

For each model, we explored the contribution of covariates and phylogeny by fitting models that excluded the effects of phylogeny (i.e. with $\lambda = 0$), latitude or exposure duration. Within each variable and covariate combination, we selected the most informative model using a multimodel inference approach by means of the lowest Akaike’s weights (w_i), which provide the relative weight of the evidence towards one of all tested models, and therefore must add up to 1 [60]. After fitting the models by maximum likelihood, hypothesis testing was performed in models with the highest support using an analysis of deviance with a significance level of $p \leq 0.05$. All analyses and figures presented in this paper were implemented and generated in R version 3.5.1 [61] using the packages ‘AICcmodavg’ [62], ‘APE’ [63], ‘nlme’ [64], ‘phytools’ [65], ‘rotl’ [56] and ‘visreg’ [66].

3. Results

We present results of empirical observations on critical thermal limits for 510 (CT_{max}) and 232 (CT_{min}) species (electronic supplementary material, figure S2a,b). For each

Table 1. Results for phylogenetic generalized least-squares (PGLSs) models to explain variation in ectotherms' CTmax ($N = 510$ species) as a function of \log_{10} -transformed body mass, breathing mode (air and water), life stage (adult and non-adult), habitat (aquatic, intertidal and terrestrial) and their interactions. All models were assessed using exposure duration (Time) and/or absolute latitude (Lat) of the animal collection as covariates. The number of parameters (k), corrected Akaike's information criterion (AICc), difference in AICc with respect to the model with highest support (ΔAICc) and Akaike's weight (w_i) are given for each model. Pagel's lambda (λ) denotes correlation structure used ($\lambda = 0$, star phylogeny, and $\lambda = 1$, Brownian phylogeny). Bold font indicates model with highest support.

model		k	AICc	ΔAICc	w_i
0. Covariates only	$\lambda = 1 + \text{Lat} + \text{Time}$	4	3013.58	54.43	0.00
	$\lambda = 0 + \text{Lat} + \text{Time}$	4	3306.34	347.19	0.00
	$\lambda = 1 + \text{Lat}$	3	3040.98	81.83	0.00
	$\lambda = 1 + \text{Time}$	3	3066.19	107.04	0.00
1. Body mass	$\lambda = 1 + \text{Lat} + \text{Time}$	5	3014.70	55.55	0.00
	$\lambda = 0 + \text{Lat} + \text{Time}$	5	3200.72	241.57	0.00
	$\lambda = 1 + \text{Lat}$	4	3042.19	83.04	0.00
	$\lambda = 1 + \text{Time}$	4	3066.02	106.87	0.00
2. Breathing mode	$\lambda = 1 + \text{Lat} + \text{Time}$	5	2984.49	25.34	0.00
	$\lambda = 0 + \text{Lat} + \text{Time}$	5	3232.59	273.43	0.00
	$\lambda = 1 + \text{Lat}$	4	3005.53	46.38	0.00
	$\lambda = 1 + \text{Time}$	4	3031.19	72.03	0.00
3. Life stage	$\lambda = 1 + \text{Lat} + \text{Time}$	5	3015.52	56.37	0.00
	$\lambda = 0 + \text{Lat} + \text{Time}$	5	3307.18	348.02	0.00
	$\lambda = 1 + \text{Lat}$	4	3039.31	80.16	0.00
	$\lambda = 1 + \text{Time}$	4	3067.71	108.56	0.00
4. Habitat	$\lambda = 1 + \text{Lat} + \text{Time}$	6	3007.04	47.89	0.00
	$\lambda = 0 + \text{Lat} + \text{Time}$	6	3246.43	287.27	0.00
	$\lambda = 1 + \text{Lat}$	5	3032.57	73.42	0.00
	$\lambda = 1 + \text{Time}$	5	3063.24	104.09	0.00
5. Body mass \times Breathing mode	$\lambda = 1 + \text{Lat} + \text{Time}$	7	2980.31	21.16	0.00
	$\lambda = 0 + \text{Lat} + \text{Time}$	7	3170.76	211.61	0.00
	$\lambda = 1 + \text{Lat}$	6	3002.48	43.33	0.00
	$\lambda = 1 + \text{Time}$	6	3030.55	71.40	0.00
6. Body mass \times Life stage	$\lambda = 1 + \text{Lat} + \text{Time}$	7	3004.43	45.27	0.00
	$\lambda = 0 + \text{Lat} + \text{Time}$	7	3190.02	230.87	0.00
	$\lambda = 1 + \text{Lat}$	6	3031.97	72.82	0.00
	$\lambda = 1 + \text{Time}$	6	3056.17	97.02	0.00
7. Body mass \times Habitat	$\lambda = 1 + \text{Lat} + \text{Time}$	9	3003.94	44.79	0.00
	$\lambda = 0 + \text{Lat} + \text{Time}$	9	3167.62	208.47	0.00
	$\lambda = 1 + \text{Lat}$	8	3027.88	68.73	0.00
	$\lambda = 1 + \text{Time}$	8	3052.40	93.25	0.00
8. Body mass \times Breathing mode \times Time	$\lambda = 1 + \text{Lat}$	10	2959.15	0.00	1.00
	$\lambda = 0 + \text{Lat}$	10	3148.75	189.60	0.00
9. Body mass \times Life stage \times Time	$\lambda = 1 + \text{Lat}$	10	2991.95	32.80	0.00
	$\lambda = 0 + \text{Lat}$	10	3189.60	230.44	0.00
10. Body mass \times Habitat \times Time	$\lambda = 1 + \text{Lat}$	14	2976.66	17.51	0.00
	$\lambda = 0 + \text{Lat}$	14	3174.65	215.49	0.00

species, we also included information on the body mass of the experimental animals (electronic supplementary material, figure S2c) used during the tests and their phylogenetic relationships (electronic supplementary material, figures S3–S6). The smallest species (red fire ant, *Solenopsis invicta*, 0.0000314 g) is separated from the largest (bonefish, *Albula*

vulpes, 1235.42 g) by 3.93×10^7 orders of magnitude (or 7.5 on \log_{10} -scale). For most of these species, we also included information on their genome size (electronic supplementary material, figure S2d), and this ranged from 0.14 pg, for the aphid *Aphidius avenae*, to 66.6 pg for the southern torrent salamander, *Rhyacotriton variegatus*. Breathing mode was

Table 2. Results for phylogenetic generalized least-squares (PGLSs) models to explain variation in ectotherms' CTmin ($N = 232$ species) as a function of \log_{10} -transformed body mass, breathing mode (air and water), life stage (adult and non-adult), habitat (aquatic, intertidal and terrestrial), exposure duration (Time) and their interactions. All models were assessed using absolute latitude (Lat) of the animal collection as a covariate. The number of parameters (k), corrected Akaike's information criterion (AICc), difference in AICc with respect to the model with highest support (Δ AICc) and Akaike's weight (w_i) are given for each model. Pagel's lambda (λ) denotes correlation structure used ($\lambda = 0$, star phylogeny, and $\lambda = 1$, Brownian phylogeny). Bold font indicates model with highest support.

model		k	AICc	Δ AICc	w_i
0. Covariates only	$\lambda = 1 + \text{Lat}$	3	1304.72	69.18	0.00
	$\lambda = 0 + \text{Lat}$	3	1399.46	163.92	0.00
1. Body mass	$\lambda = 1 + \text{Lat}$	4	1306.73	71.19	0.00
	$\lambda = 0 + \text{Lat}$	4	1289.78	54.24	0.00
2. Breathing mode	$\lambda = 1 + \text{Lat}$	4	1305.39	69.85	0.00
	$\lambda = 0 + \text{Lat}$	4	1384.55	149.01	0.00
3. Life stage	$\lambda = 1 + \text{Lat}$	4	1306.57	71.03	0.00
	$\lambda = 0 + \text{Lat}$	4	1398.91	163.37	0.00
4. Habitat	$\lambda = 1 + \text{Lat}$	5	1292.00	56.46	0.00
	$\lambda = 0 + \text{Lat}$	5	1396.35	160.81	0.00
5. Body mass \times Breathing mode	$\lambda = 1 + \text{Lat}$	6	1307.00	71.46	0.00
	$\lambda = 0 + \text{Lat}$	6	1273.49	37.95	0.00
6. Body mass \times Life stage	$\lambda = 1 + \text{Lat}$	6	1306.84	71.30	0.00
	$\lambda = 0 + \text{Lat}$	6	1285.09	49.55	0.00
7. Body mass \times Habitat	$\lambda = 1 + \text{Lat}$	8	1297.78	62.24	0.00
	$\lambda = 0 + \text{Lat}$	8	1274.84	39.30	0.00
8. Body mass \times Breathing mode \times Time	$\lambda = 1 + \text{Lat}$	10	1288.19	52.65	0.00
	$\lambda = 0 + \text{Lat}$	10	1235.54	0.00	1.00
9. Body mass \times Life stage \times Time	$\lambda = 1 + \text{Lat}$	10	1290.22	54.68	0.00
	$\lambda = 0 + \text{Lat}$	10	1249.66	14.12	0.00
10. Body mass \times Habitat \times Time	$\lambda = 1 + \text{Lat}$	14	1269.39	33.85	0.00
	$\lambda = 0 + \text{Lat}$	14	1264.29	28.75	0.00

represented by 225 and 285 species, corresponding to air- and water-breathing species, respectively. On the other hand, most data concerned adults ($N = 402$), while the remaining larvae and juveniles were grouped as non-adults ($N = 108$). In terms of habitat, the majority of species were aquatic (316 species), or terrestrial (181 species), with only a few species being intertidal (13 species).

Both CTmax and CTmin showed a clear phylogenetic signal (electronic supplementary material, table S11), indicating that thermal tolerance among the studied species has been largely conserved across evolutionary lineages. A comparison between PGLSs models under a Brownian mode of evolution ($\lambda = 1$) and non-phylogenetic models ($\lambda = 0$) showed, in most cases, that accounting for phylogenetic relationships among the studied species improved the model fit for both CTmax (tables 1 and 3) and CTmin (tables 2 and 4). The two covariates (i.e. absolute latitude and exposure duration) were always included in the best-supported model, indicating their importance in explaining variation in thermal tolerance. For all CTmax models, greater support and the lowest corrected Akaike information criterion (AICc) were observed when phylogeny was taken into account ($\lambda = 1$). By contrast, for CTmin, accounting for the shared evolutionary history of species was less important for those models that already included body mass as an explanatory variable, possibly because body mass is strongly

phylogenetically structured and may, therefore, obviate the need to include phylogeny (table 2, model 5 to model 10).

Modelled effects of body mass and genome size on both thermal limits differed according to whether the model included phylogeny or not. For CTmax, a negative relationship with body mass was most apparent in the model that did not include phylogeny ($\lambda = 0$), likely because extreme values of both CTmax and body mass were phylogenetically clustered (tables 1 and 3 and figure 1a,b; electronic supplementary material, figures S3 and S4). Effects of both body mass and genome size on CTmax differed with breathing mode and exposure duration (electronic supplementary material, tables S3 and S5; see below). For CTmin, the best-supported models indicated that cold tolerance declined (i.e. higher CTmin values) with increasing body mass (table 2 and figure 1c) and with increasing genome size (table 4 and figure 1d). Effects of body mass on CTmin differed with breathing mode and exposure duration (electronic supplementary material, table S4), whereas those of genome size differed with habitat and exposure duration (electronic supplementary material, table S6).

Consistent with the expectation that both CTmax and CTmin differ in species with different breathing modes and across habitats, our results indicate a generally lower tolerance for water-breathers compared with air-breathers, suggesting that water-breathers are more vulnerable to both

Table 3. Results for phylogenetic generalized least-squares (PGLSs) models to explain variation in ectotherms' CTmax ($N = 433$ species) as a function of \log_{10} -transformed genome size, breathing mode (air and water), life stage (adult and non-adult), habitat (aquatic, intertidal and terrestrial) and their interactions. All models were assessed using exposure duration (Time) and/or absolute latitude (Lat) of the animal collection as covariates. The number of parameters (k), corrected Akaike's information criterion (AICc), difference in AICc with respect to the model with highest support (Δ AICc) and Akaike's weight (w_i) are given for each model. Pagel's lambda (λ) denotes correlation structure used ($\lambda = 0$, star phylogeny, and $\lambda = 1$, Brownian phylogeny). Bold font indicates models with highest support.

model		k	AICc	Δ AICc	w_i
0. Covariates only	$\lambda = 1 + \text{Lat} + \text{Time}$	4	2505.59	3.24	0.07
	$\lambda = 0 + \text{Lat} + \text{Time}$	4	2759.27	256.92	0.00
	$\lambda = 1 + \text{Lat}$	3	2521.96	19.60	0.00
	$\lambda = 1 + \text{Time}$	3	2555.30	52.94	0.00
1. Genome size	$\lambda = 1 + \text{Lat} + \text{Time}$	5	2506.18	3.82	0.05
	$\lambda = 0 + \text{Lat} + \text{Time}$	5	2731.05	228.69	0.00
	$\lambda = 1 + \text{Lat}$	4	2521.94	19.58	0.00
	$\lambda = 1 + \text{Time}$	4	2554.25	51.89	0.00
2. Breathing mode	$\lambda = 1 + \text{Lat} + \text{Time}$	5	2504.30	1.95	0.13
	$\lambda = 0 + \text{Lat} + \text{Time}$	5	2644.82	142.46	0.00
	$\lambda = 1 + \text{Lat}$	4	2519.78	17.43	0.00
	$\lambda = 1 + \text{Time}$	4	2551.15	48.79	0.00
3. Life stage	$\lambda = 1 + \text{Lat} + \text{Time}$	5	2503.91	1.55	0.16
	$\lambda = 0 + \text{Lat} + \text{Time}$	5	2758.50	256.14	0.00
	$\lambda = 1 + \text{Lat}$	4	2513.96	11.60	0.00
	$\lambda = 1 + \text{Time}$	4	2551.56	49.20	0.00
4. Habitat	$\lambda = 1 + \text{Lat} + \text{Time}$	6	2509.48	7.12	0.01
	$\lambda = 0 + \text{Lat} + \text{Time}$	6	2718.37	216.01	0.00
	$\lambda = 1 + \text{Lat}$	5	2525.79	23.43	0.00
	$\lambda = 1 + \text{Time}$	5	2558.64	56.28	0.00
5. Genome size \times Breathing mode	$\lambda = 1 + \text{Lat} + \text{Time}$	7	2506.43	4.07	0.05
	$\lambda = 0 + \text{Lat} + \text{Time}$	7	2599.70	97.34	0.00
	$\lambda = 1 + \text{Lat}$	6	2521.26	18.90	0.00
	$\lambda = 1 + \text{Time}$	6	2551.27	48.91	0.00
6. Genome size \times Life stage	$\lambda = 1 + \text{Lat} + \text{Time}$	7	2506.65	4.30	0.04
	$\lambda = 0 + \text{Lat} + \text{Time}$	7	2715.88	213.52	0.00
	$\lambda = 1 + \text{Lat}$	6	2515.68	13.32	0.00
	$\lambda = 1 + \text{Time}$	6	2552.49	50.13	0.00
7. Genome size \times Habitat	$\lambda = 1 + \text{Lat} + \text{Time}$	9	2511.67	9.31	0.00
	$\lambda = 0 + \text{Lat} + \text{Time}$	9	2669.46	167.10	0.00
	$\lambda = 1 + \text{Lat}$	8	2527.59	25.23	0.00
	$\lambda = 1 + \text{Time}$	8	2553.99	51.64	0.00
8. Genome size \times Breathing mode \times Time	$\lambda = 1 + \text{Lat}$	10	2502.36	0.00	0.36
	$\lambda = 0 + \text{Lat}$	10	2590.26	87.90	0.00
9. Genome size \times Life stage \times Time	$\lambda = 1 + \text{Lat}$	10	2510.08	7.72	0.01
	$\lambda = 0 + \text{Lat}$	10	2718.49	216.13	0.00
10. Genome size \times Habitat \times Time	$\lambda = 1 + \text{Lat}$	14	2504.72	2.36	0.11
	$\lambda = 0 + \text{Lat}$	14	2669.50	167.14	0.00

heat and cold (figure 2*a,d*). Contrary to our expectation, we found no differences in thermal limits between different life stages (figure 2*b,e*). Intertidal species were shown to be more tolerant to the effects of cold (figure 2*f*). However, these results should be interpreted with caution in light of low representation of intertidal species in our analyses (five

species for CTmin). Also, this difference for intertidal species was not present when phylogenetic relationships were not accounted for (electronic supplementary material, figure S8). Although breathing mode and habitat strongly covaried (most aquatic species are water-breathers and most terrestrial species are air-breathers), variation in CTmax was best

Table 4. Results for phylogenetic generalized least-squares (PGLSs) models to explain variation in ectotherms' CTmin ($N = 190$ species) as a function of \log_{10} -transformed genome size, breathing mode (air and water), life stage (adult and non-adult), habitat (aquatic, intertidal and terrestrial) and their interactions. All models were assessed using the absolute latitude (Lat) of the animal collection as a covariate. The number of parameters (k), corrected Akaike's information criterion (AICc), difference in AICc with respect to the model with highest support (Δ AICc) and Akaike's weight (w_i) are given for each model. Pagel's lambda (λ) denotes correlation structure used ($\lambda = 0$, star phylogeny, and $\lambda = 1$, Brownian phylogeny). Bold font indicates model with highest support.

model		k	AICc	Δ AICc	w_i
0. Covariates only	$\lambda = 1 + \text{Lat}$	3	1076.21	37.66	0.00
	$\lambda = 0 + \text{Lat}$	3	1157.44	118.88	0.00
1. Genome size	$\lambda = 1 + \text{Lat}$	4	1074.03	35.47	0.00
	$\lambda = 0 + \text{Lat}$	4	1159.53	120.97	0.00
2. Breathing mode	$\lambda = 1 + \text{Lat}$	4	1076.83	38.27	0.00
	$\lambda = 0 + \text{Lat}$	4	1150.57	112.01	0.00
3. Life stage	$\lambda = 1 + \text{Lat}$	4	1077.85	39.29	0.00
	$\lambda = 0 + \text{Lat}$	4	1158.68	120.13	0.00
4. Habitat	$\lambda = 1 + \text{Lat}$	5	1063.79	25.23	0.00
	$\lambda = 0 + \text{Lat}$	5	1148.50	109.94	0.00
5. Genome size \times Breathing mode	$\lambda = 1 + \text{Lat}$	6	1070.58	32.02	0.00
	$\lambda = 0 + \text{Lat}$	6	1144.42	105.87	0.00
6. Genome size \times Life stage	$\lambda = 1 + \text{Lat}$	6	1075.65	37.09	0.00
	$\lambda = 0 + \text{Lat}$	6	1154.62	116.06	0.00
7. Genome size \times Habitat	$\lambda = 1 + \text{Lat}$	8	1046.08	7.53	0.02
	$\lambda = 0 + \text{Lat}$	8	1153.09	114.53	0.00
8. Genome size \times Breathing mode \times Time	$\lambda = 1 + \text{Lat}$	10	1047.97	9.42	0.01
	$\lambda = 0 + \text{Lat}$	10	1136.59	98.03	0.00
9. Genome size \times Life stage \times Time	$\lambda = 1 + \text{Lat}$	10	1045.99	7.43	0.02
	$\lambda = 0 + \text{Lat}$	10	1156.36	117.80	0.00
10. Genome size \times Habitat \times Time	$\lambda = 1 + \text{Lat}$	14	1038.56	0.00	0.95
	$\lambda = 0 + \text{Lat}$	14	1153.79	115.23	0.00

explained by models based on breathing mode (table 1, model 8), rather than on habitat (table 1, model 10). Variation in CTmin was best explained by models based on breathing mode (when including body mass; table 2) and habitat (when including genome size; table 4). Cold tolerance declined (i.e. higher CTmin values) with increasing body mass (figure 1c).

More complex models, testing three interactions (body mass \times breathing mode \times exposure duration), showed the highest support to explain variations both in CTmax (tables 1 and 3) and, with some exceptions, in CTmin (tables 2 and 4). In general, these models indicate that exposure duration modulates the intensity or even reverses the direction of the effects of body mass (figure 3) or genome size (figure 4). For water-breathers, larger species were found to have a lower CTmax in long-term experimental trials, while the model indicates an opposite effect in short-term trials (figure 3*a,b*). For cold tolerance, the three-way interaction with exposure duration was also important for models including body mass and genome size. Here, air-breathers showed improved cold tolerance (lower CTmin values) with increasing genome size, but only for long-term trials (figure 4*d*).

Since different numbers of species were included in our analyses on body mass and genome size, the performance of the models cannot be compared directly. We therefore

repeated the analyses on a smaller set of species for which information on both body mass and genome size was available. This smaller set included 433 species for CTmax and 190 species for CTmin. These analyses allowed us not only to compare the contributions of body mass and genome size but also to test for possible interactions between body mass and genome size. The results of these analyses were highly consistent with those presented above, that is, models with the highest support, for both the CTmax and CTmin, were those that incorporated body mass, genome size, breathing mode and exposure duration. Interestingly, variations in CTmax were mainly driven by those models that considered body mass instead of genome size (electronic supplementary material, table S7). On the contrary, for the CTmin, the model with the highest support ($w_i = 0.99$) was that which considered the three-way interaction of body mass and genome size and exposure duration (electronic supplementary material, table S9).

4. Discussion

Body mass is of fundamental importance for the ecology of ectotherms, governing the rates of energy uptake and energy transformation at the organismal level, with subsequent consequences for species interactions and to the

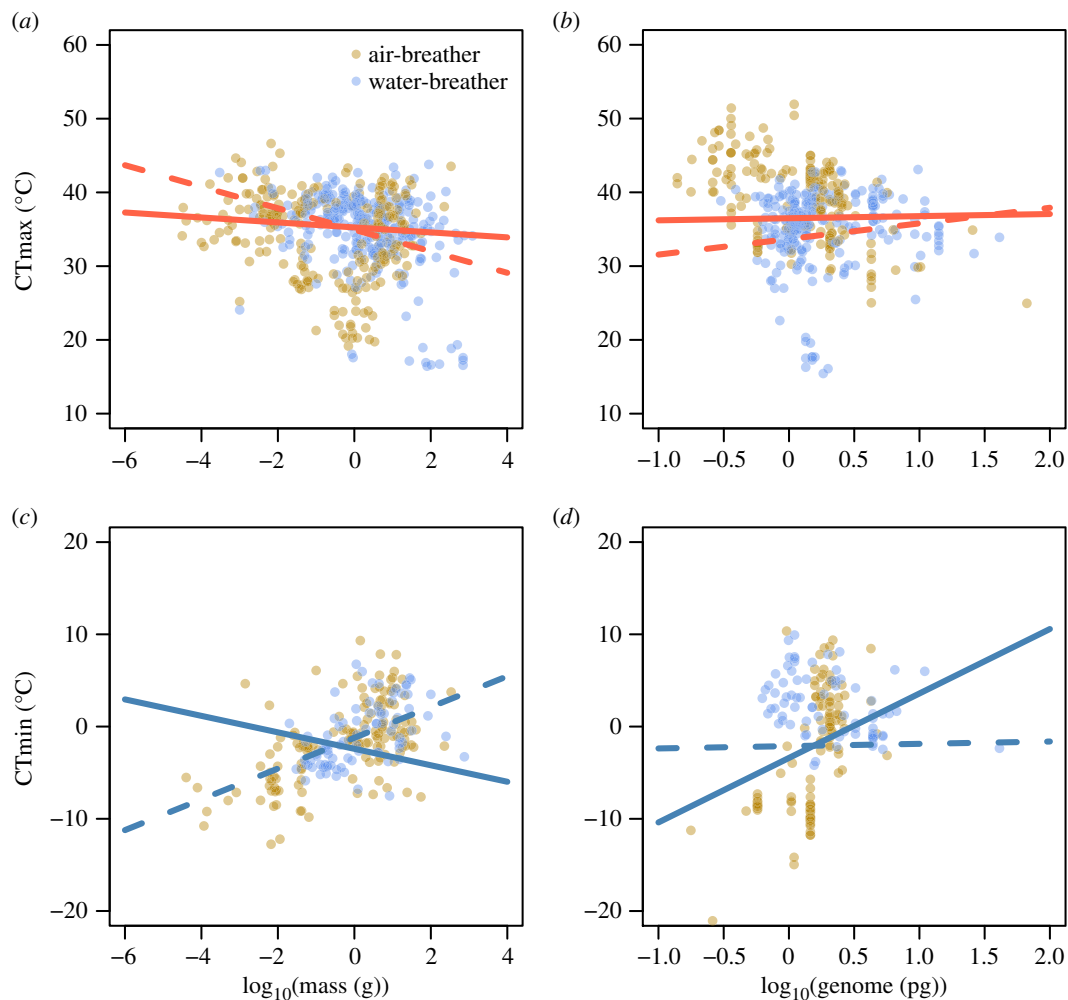


Figure 1. Partial residuals plots showing the predicted effects of \log_{10} -transformed body mass and \log_{10} -transformed genome size in ectotherms' CT_{\max} (top, red) and CT_{\min} (bottom, blue). CT_{\max} (a,b) and CT_{\min} plots (c,d) were based on the model with the highest support, shown in electronic supplementary material, tables S1–S4. Solid lines indicate predictions of models that included all covariates (latitude, time) and phylogenetic relationships, whereas dashed lines indicate predictions of models that included all covariates, but do not account for phylogenetic relationships ($\lambda = 0$). For details on model estimates and significance, see electronic supplementary material, tables S3–S6. (Online version in colour.)

ecosystem functioning. Knowing whether the consequences of global warming are size-dependent is therefore central, particularly in light of the ongoing global climatic warming. Here, we have taken a comparative approach to shed light on the relationship between thermal tolerance levels and body mass and genome size in ectotherms. A challenge in such large-scale, comparative studies lies in dealing with the unique evolutionary history of the various species [67], as well as dealing with differences in methodology across studies [58,68,69]. Our results show that effects of body mass and genome size on thermal limits (CT_{\max} and CT_{\min}) are context-dependent, covarying to some extent with the evolutionary relationships across species and differing mainly with breathing mode of species. The methodology was also influential (see also [70]), as size-dependent differences in thermal limits were magnified in long-term trials.

(a) Do body mass and genome size relate to thermal limits?

Our results indicate that there is not a simple, straightforward answer as to whether body mass and genome size matter in defining a species' thermal tolerance or not. If heat tolerance limits arise from insufficient oxygen provisioning to meet

demand, and if such oxygen limitation is more likely to occur in larger ectotherms, we would expect heat tolerance to be more impaired in larger-bodied animals. We found such a relationship, but only in our analyses that did not include phylogenetic relationships. Accounting for phylogeny appears to be a more parsimonious explanation for variation in heat tolerance. Still, even when accounting for phylogenetic relationships, we found size-dependence of heat tolerance, but this was contingent upon exposure duration and breathing mode, with impaired heat tolerance being more apparent in larger, water-breather animals during longer trials. Owing to the challenge of underwater gas exchange, water-breathers have been argued to be more susceptible to oxygen-limited heat tolerance [16,21]. The timescale is also important here as stress relates to both its intensity and duration [68]. Heat stress may result in energy deficits, and while energy can be generated either aerobically or anaerobically, anaerobic metabolism is much less efficient and more suitable to deal with acute, short-term energy deficits [71]. For fish, it has been suggested that larger species rely more on anaerobic metabolism when faced with energy deficits [72–74] and if this mass scaling generalizes, this could explain why larger species may be better in coping with short, acute heat stress, but not with prolonged heat stress.

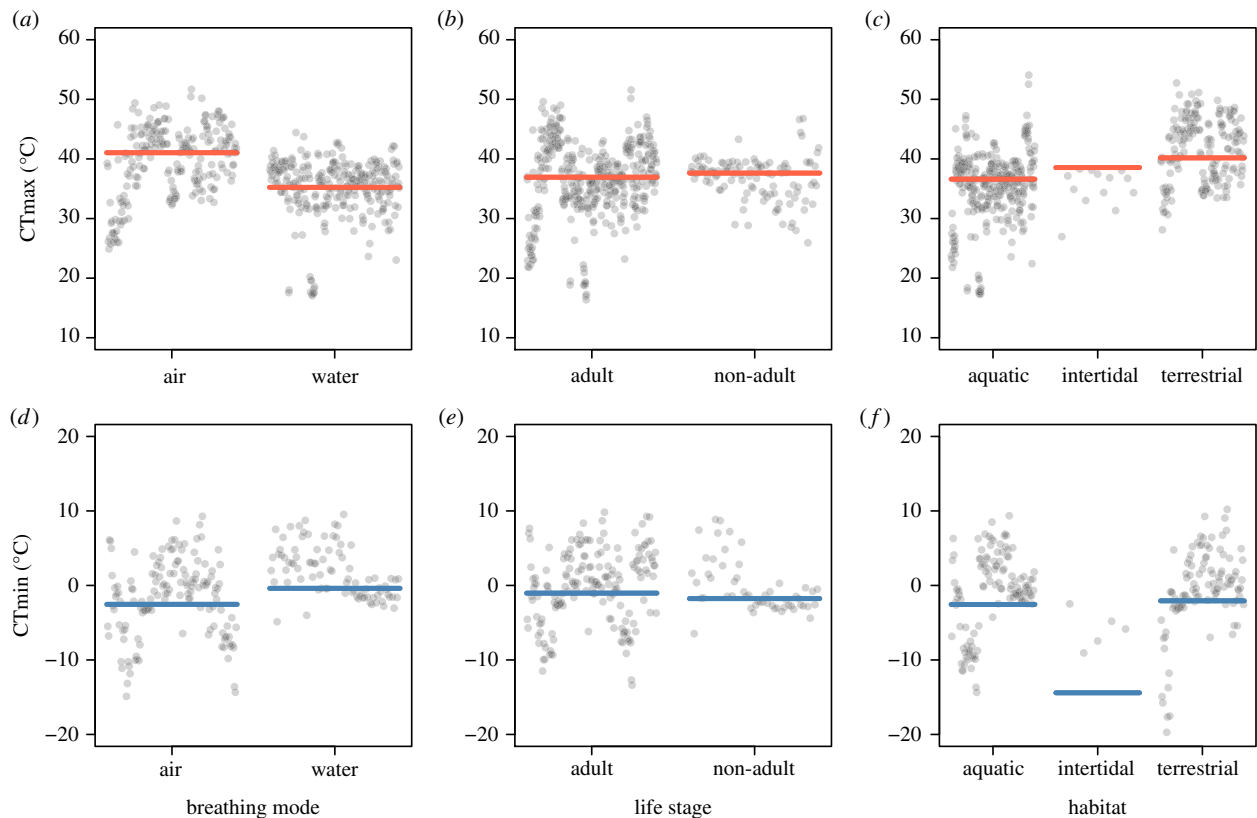


Figure 2. Partial residuals plots showing the predicted effects of breathing mode (*a,d*), life stage (*b,e*) and habitat (*c,f*) in ectotherms' CT_{max} (top, red) and CT_{min} (bottom, blue). CT_{max} (*a–c*) and CT_{min} plots (*d–f*) were based on the model with the highest support shown in electronic supplementary material, tables S1 and S3 that included either breathing mode, life stage or habitat. Horizontal solid lines are the predicted medians of the thermal limits within factor level, conditioned on 2 h of exposure duration (Time) and 45° of absolute latitude. For details on model estimates and significance, see electronic supplementary material, tables S3–S6. (Online version in colour.)

Given these considerations, it is perhaps not surprising to find the strongest effects of body mass in prolonged trials on water-breathers.

Heat tolerance was lower in water-breathers compared with air-breathers during prolonged trials when they had larger body sizes, but also when they had larger cell size. The observed effects of genome size can also be interpreted from an oxygen perspective, as smaller genome size is coupled to smaller cell size [75], which can promote a more efficient diffusion of oxygen towards the mitochondria owing to increased membrane surface area to cell volume ratios and shorter diffusion distances [24,44,76,77]. Studies on flies and isopods have shown that warming-induced size reductions are more pronounced under hypoxia [29,76,78], supporting the idea of oxygen shortage setting limits to the size that an animal can attain. This idea also implies that oxygen is unlikely to be limiting in animals that have not yet approached their maximum species-specific size. As the body mass used here is that of the experimental species, in most cases the specimens used in the experiments will not have represented the upper size classes. This may explain why phylogeny better explains the variation in CT_{max}: phylogeny is more likely to covary with the maximum size that a species can attain, than with the size of the individuals used in the experiments. Indeed, juveniles and larvae, which by definition are not yet fully developed, both show improved heat tolerance with increasing body mass, contrasting with impaired heat tolerance in adults (electronic supplementary material, figure S9). Along the same lines, in a study looking at intraspecific variation in body mass, CT_{max} improved

with body mass in juvenile spiders (family Linyphiidae), but deteriorated with size of adults in Hemiptera (*Heterocerus* sp.) and Collembola (*Isotoma riparia*) [38]. Thus, an oxygen-based mechanism could play a role in heat tolerance but appears to be more relevant for water-breathers and on longer timescales: i.e. exactly those conditions for which a strong warming-induced reduction in body mass has been observed [5].

Unlike heat tolerance, cold tolerance has been suggested to result from depolarization of cell membranes and subsequent cell death [46,79–82], and not from oxygen limitation [83]. Our results also suggest that the mechanisms underpinning CT_{min} differ from those underpinning CT_{max} as the contribution of phylogeny, body mass and genome size to explaining variation in CT_{min} differed from that to explaining variation in CT_{max} (electronic supplementary material, tables S3–S6). Models that consider combined effects of body mass and genome size indicate that this combination better explains variation in CT_{min}, than variation in CT_{max} (see electronic supplementary material, tables S7–S10). While a small genome size (or small cell size) may enhance oxygen diffusion, it also entails greater costs in keeping membranes polarized [44,84]. Thus, larger cells may be more cost efficient and this could explain why larger genomes can improve cold tolerance. The effect of such an efficiency-based mechanism would likely be more apparent during prolonged trials, and indeed we found that including the interaction between genome size, habitat and exposure duration showed the highest support across all models (table 4), showing improved cold tolerance in terrestrial

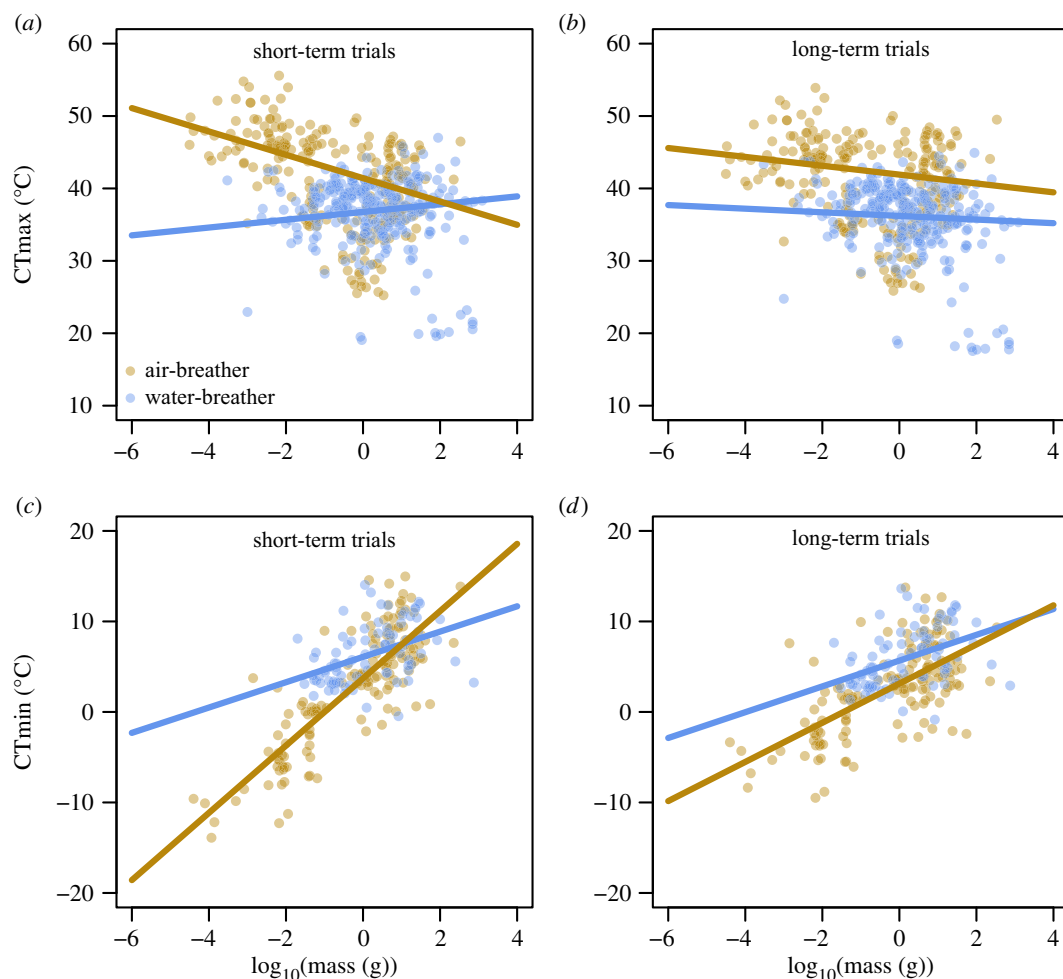


Figure 3. Partial residuals plots showing the interactive effects of body mass and breathing mode (brown and blue for air- and water-breathers, respectively) in ectotherms' CTmax (top) and CTmin (bottom), for short- (1st quartile) and long-term trials (3rd quartile). Predicted lines were based on models with the highest support shown in tables 1 and 2 and based on the median of absolute latitude. For details on model estimates and significance, see electronic supplementary material, tables S3 and S4. (Online version in colour.)

animals with larger genome during prolonged trials (figure 4). In line with these findings, results on the larvae of the pipevine swallowtail *Battus philenor* (Linnaeus, 1771) suggested that larger species may have more energy reserves for maintaining metabolism during chill coma, thus explaining their improved cold tolerance [85]. When coupled to lower mass-specific metabolism in larger animals, such an efficiency mechanism would be generally applicable to the whole size range and not only restricted to the largest size classes within a species. This may explain why cold tolerance is most parsimoniously explained by differences in body mass, since phylogeny is more likely to covary with maximum size than with the size of the animals used in the experiment). Interestingly, these patterns for CTmin were more apparent for air-breathers, perhaps because cold tolerance limits in water-breathers are more related to the freezing of water.

(b) Model fit, phylogenetic correlation structure and covariates

We found evidence of the influence of phylogeny on two fronts. First, both the CTmax and CTmin are phylogenetically structured, displaying high Pagel's λ (electronic supplementary material, table S11) and second, those models

incorporating phylogeny generally received greater support (especially for CTmax) compared with those where the evolutionary history of the species was considered independent. Also, the Pagel's λ used in our models ($\lambda = 1$) is highly likely to be a representative value of the shared evolution of species present in our database, since all continuous variables, both independent (body mass, genome size, exposure duration and absolute latitude) and dependent (CTmax and CTmin), showed high phylogenetic signals (all $\lambda > 0.7$) (electronic supplementary material, table S11).

The influence of phylogeny on thermal limits is also evident from the contrasting effects of body mass and genome size between models that considered a Brownian or star phylogeny correlation structure (figure 1). This indicates that body mass and genome size covary with phylogeny, something that is also evident from the high Pagel's λ value for body mass and genome size (electronic supplementary material, table S11). Consequently, incorporating phylogeny already accounts for much of the variation in thermal tolerance, thereby changing the fitted relationship for body mass and genome size. For CTmax, models that included phylogeny always showed greater support, suggesting that the patterns in heat tolerance were more parsimoniously explained by including evolutionary history, possibly because phylogeny better captures the maximum body size,

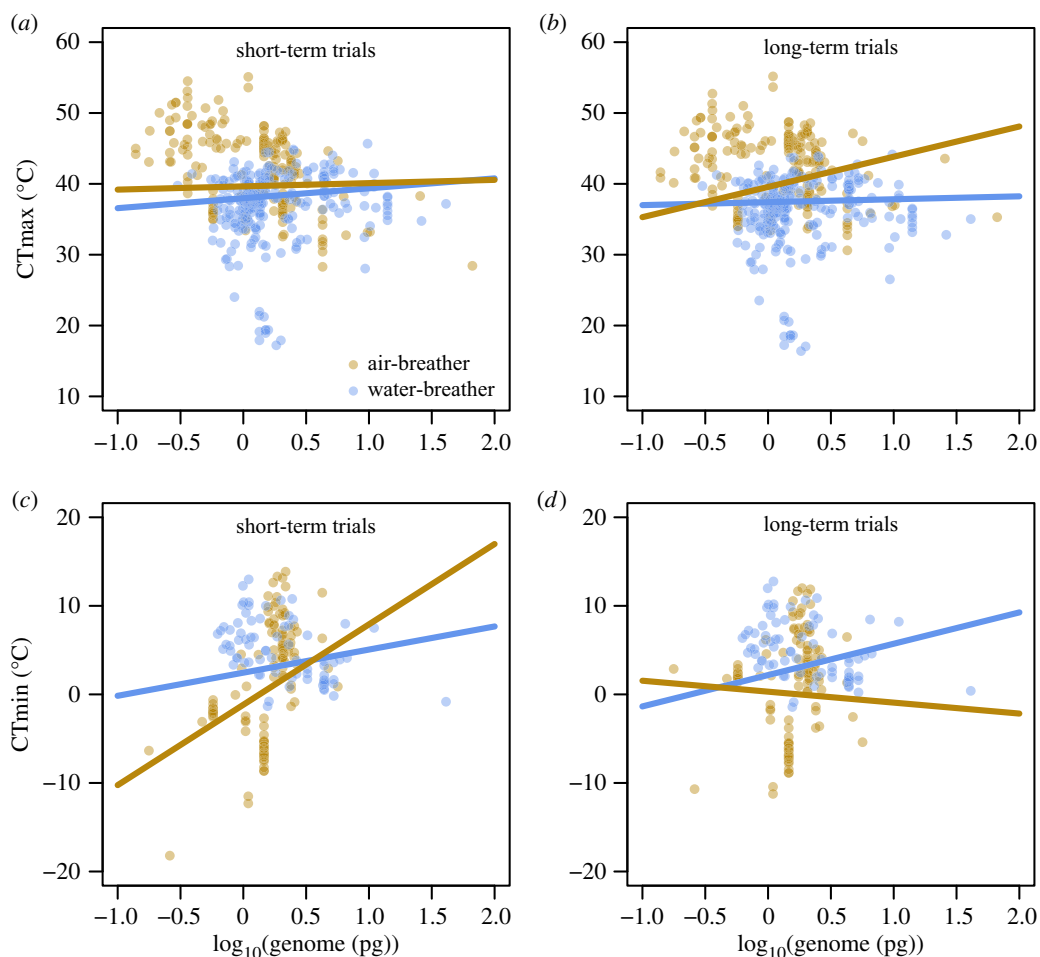


Figure 4. Partial residuals plots showing the interactive effects of genome size and breathing mode (brown and blue for air- and water-breathers, respectively) in ectotherms' CT_{max} (top) and CT_{min} (bottom), for short- (1st quartile) and long-term trials (3rd quartile). Predicted lines were based on models with the highest support shown in tables 3 and 4 and based on the median of absolute latitude. For details on model estimates and significance, see electronic supplementary material, tables S5 and S6. (Online version in colour.)

which may be causally related to CT_{max} (see above). For CT_{min} , models that included the species' body mass as an independent variable showed greater support when the evolution of the species was assumed as independent (i.e. star phylogeny), possibly because here the actual body size of the experimental individuals is causally related to cold tolerance (see above). Overall, our results confirm earlier findings suggesting that evolutionary history matters for thermal tolerance [86–89], especially for heat tolerance [86,90] and also suggest that this applies to the much larger set of ectotherm species, including insects, crustaceans, fish, amphibians and reptiles. Our results also point out the importance of including mainly methodological aspects as covariates in modelling (see also [70]). Of the four methodological aspects evaluated in the preliminary models (absolute latitude, exposure duration, acclimation time and origin), latitude and exposure duration were consistently included in the best-fitted models. The absolute latitude of the site where animals were collected consistently shifted thermal windows, impairing the heat tolerance and improving the cold tolerance at higher latitudes (electronic supplementary material, figure S7). While the effects of latitude were not the focus of our analyses, these results reinforce the clear patterns of thermal tolerance across latitudinal gradients documented in the literature [12] (see also [70]). The exposure duration was also consistently included in the best-fitted models, with long-term trials consistently reducing CT_{max} (electronic

supplementary material, figure S7). This indicates that methodological variation explains a significant part of the variation in CT_{max} and adding exposure duration as covariate may help to reveal more clearly the effects of other factors, such as that of latitude [68]. Furthermore, the inclusion of exposure duration as a covariate has direct biological meaning as tolerance to high-temperature conditions is time-dependent [68].

5. Conclusion

In conclusion, for CT_{max} we found that support for the oxygen limitation hypothesis was limited to long-term trials in larger-bodied water-breathers. For CT_{min} , we found improved cold tolerance for air-breather animals with larger genomes, again when considering long-term trials. Coping with thermal stress on long timescales requires sustained energy generation. Long-term heat tolerance appears to be enhanced in smaller-bodied, water-breathing species, possibly as this enables a higher capacity to generate energy aerobically and efficiently. On the other hand, long-term cold tolerance appears to be enhanced in species with a larger body mass and cell size (i.e. more energy reserves and lower energetic costs), which appeared especially important for air-breathers. Incorporating the exposure duration of the experimental trials can reveal body and genome size-dependence of thermal tolerance, with body size being

more important for CT_{max} and water-breathers and genome size being more important for CT_{min} and air-breathers. Our results highlight the importance of accounting for phylogeny and exposure duration. Especially when considering long-term trials, these effects are more in line with the warming-induced reduction in body mass observed during long-term rearing experiments [5] and over past extinctions [8]. Explicitly incorporating timescale may thus hold the key to resolve discrepancies between short-term trials, which do not always find evidence for oxygen limitation, and the results of long-term laboratory and field studies, which do suggest a role for oxygen limitation.

Data accessibility. Data available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.878vn25> [91].

Authors' contributions. F.P.L. extracted the data from the articles; F.P.L. conducted the statistical analyses and prepared figures with inputs from W.C.E.P.V. and P.C.; F.P.L. and W.C.E.P.V. wrote the first

version of the manuscript with inputs from P.C. All authors contributed and approved the final version of the manuscript.

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References

- Atkinson D, Hirst AG. 2007 Life histories and body size. In *Body size: the structure and function of aquatic ecosystems* (eds AG Hildrew, DG Raffaelli, R Edmonds-Brown), pp. 33–54. Cambridge, UK: Cambridge University Press.
- Enquist BJ, Economo EP, Huxman TE, Allen AP, Ignace DD, Gillooly JF. 2003 Scaling metabolism from organisms to ecosystems. *Nature* **423**, 639–642. (doi:10.1038/nature01671)
- Brown JH, Gillooly JF, Allen AP, Savage VM, West GB. 2004 Toward a metabolic theory of ecology. *Ecology* **85**, 1771–1789. (doi:10.1890/03-9000)
- Bergmann C. 1848 Über die Verhältnisse der Wärmeökonomie der Thiere zu ihrer Grösse. *Göttinger Studien* **1**, 595–708 [in German].
- Horne CR, Hirst A, Atkinson D. 2015 Temperature-size responses match latitudinal-size clines in arthropods, revealing critical differences between aquatic and terrestrial species. *Ecol. Lett.* **18**, 327–335. (doi:10.1111/ele.12413)
- Sallan L, Galimberti AK. 2015 Body-size reduction in vertebrates following the end-Devonian mass extinction. *Science* **350**, 812–815. (doi:10.1126/science.aac7373)
- Angilletta MJ, Dunham AE. 2003 The temperature-size rule in ectotherms: simple evolutionary explanations may not be general. *Am. Nat.* **162**, 332–342. (doi:10.1086/377187)
- Calosi P, Putnam HM, Twitchett RJ, Vermandele F. 2019 Marine Metazoan modern mass extinction: improving predictions by integrating fossil, modern, and physiological data. *Annu. Rev. Mar. Sci.* **11**, 369–390. (doi:10.1146/annurev-marine-010318-095106)
- Daufresne M, Lengfellner K, Sommer U. 2009 Global warming benefits the small in aquatic ecosystems. *Proc. Natl Acad. Sci. USA* **106**, 12 788–12 793. (doi:10.1073/pnas.0902080106)
- Root TL, Price JT, Hall KR, Schneider SH, Rosenzweig C, Pounds JA. 2003 Fingerprints of global warming on wild animals and plants. *Nature* **421**, 57–60. (doi:10.1038/nature01333)
- Chen I-C, Hill JK, Ohlemüller R, Roy DB, Thomas CD. 2011 Rapid range shifts of species associated with high levels of climate warming. *Science* **333**, 1024–1026. (doi:10.1126/science.1206432)
- Sunday JM, Bates AE, Dulvy NK. 2011 Global analysis of thermal tolerance and latitude in ectotherms. *Proc. R. Soc. B* **278**, 1823–1830. (doi:10.1098/rspb.2010.1295)
- Gunderson AR, Stillman JH. 2015 Plasticity in thermal tolerance has limited potential to buffer ectotherms from global warming. *Proc. R. Soc. B* **282**, 20150401. (doi:10.1098/rspb.2015.0401)
- Araújo MB, Ferri-Yáñez F, Bozinovic F, Marquet PA, Valladares F, Chown SL. 2013 Heat freezes niche evolution. *Ecol. Lett.* **16**, 1206–1219. (doi:10.1111/ele.12155)
- Addo-Bediako A, Chown SL, Gaston KJ. 2000 Thermal tolerance, climatic variability and latitude. *Proc. R. Soc. Lond. B* **267**, 739–745. (doi:10.1098/rspb.2000.1065)
- Verberk WCEP, Overgaard J, Ern R, Bayley M, Wang T, Boardman L, Terblanche JS. 2016 Does oxygen limit thermal tolerance in arthropods? A critical review of current evidence. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **192**, 64–78. (doi:10.1016/j.cbpa.2015.10.020)
- Lefevre S. 2016 Are global warming and ocean acidification conspiring against marine ectotherms? A meta-analysis of the respiratory effects of elevated temperature, high CO₂ and their interaction. *Conserv. Physiol.* **4**, cow009. (doi:10.1093/conphys/cow009)
- Pauly D, Cheung WW. 2018 Sound physiological knowledge and principles in modeling shrinking of fishes under climate change. *Glob. Change Biol.* **24**, 13831. (doi:10.1111/gcb.13831)
- Audzijonyte A, Barneche DR, Baudron AR, Belmaker J, Clark TD, Marshall CT, Morrongiello JR, van Rijn I. 2018 Is oxygen limitation in warming waters a valid mechanism to explain decreased body sizes in aquatic ectotherms? *Glob. Ecol. Biogeogr.* **28**, 64–77. (doi:10.1111/gcb.12847)
- Pörtner HO, Bock C, Mark FC. 2017 Oxygen- and capacity-limited thermal tolerance: bridging ecology and physiology. *J. Exp. Biol.* **220**, 2685–2696. (doi:10.1242/jeb.134585)
- Verberk WCEP, Bilton DT. 2013 Respiratory control in aquatic insects dictates their vulnerability to global warming. *Biol. Lett.* **9**, 20130473. (doi:10.1098/rsbl.2013.0473)
- Pörtner HO. 2001 Climate change and temperature-dependent biogeography: oxygen limitation of thermal tolerance in animals. *Naturwissenschaften* **88**, 137–146. (doi:10.1007/s001140100216)
- Verberk WCEP, Sommer U, Davidson RL, Viant MR. 2013 Anaerobic metabolism at thermal extremes: a metabolomic test of the oxygen limitation hypothesis in an aquatic insect. *Integr. Comp. Biol.* **53**, 609–619. (doi:10.1093/icb/ict015)
- Atkinson D, Morley SA, Hughes RN. 2006 From cells to colonies: at what levels of body organization does the 'temperature-size rule' apply? *Evol. Dev.* **8**, 202–214. (doi:10.1111/j.1525-142X.2006.00090.x)
- Verberk WCEP, Atkinson D. 2013 Why polar gigantism and Palaeozoic gigantism are not equivalent: effects of oxygen and temperature on the body size of ectotherms. *Funct. Ecol.* **27**, 1275–1285. (doi:10.1111/1365-2435.12152)
- Atkinson D. 1994 Temperature and organism size—a biological law for ectotherms? *Adv. Ecol. Res.* **25**, 1–58. (doi:10.1016/S0065-2504(08)60212-3)
- Pauly D. 2010 *Gasping fish and panting squids: oxygen, temperature and the growth of water-breathing animals*. Oldendorf (Luhe), Germany: International Ecology Institute.
- Lane SJ, Shishido CM, Moran AL, Tobalske BW, Arango CP, Woods HA. 2017 Upper limits to body size imposed by respiratory–structural trade-offs in Antarctic pycnogonids. *Proc. R. Soc. B* **284**, 20171779. (doi:10.1098/rspb.2017.1779)
- Hoefnagel KN, Verberk WCEP. 2015 Is the temperature-size rule mediated by oxygen in

- aquatic ectotherms? *J. Therm. Biol.* **54**, 56–65. (doi:10.1016/j.jtherbio.2014.12.003)
30. Penn JL, Deutsch C, Payne JL, Sperling EA. 2018 Temperature-dependent hypoxia explains biogeography and severity of end-Permian marine mass extinction. *Science* **362**, eaat1327. (doi:10.1126/science.aat1327)
 31. Gillooly JF, Brown JH, West GB, Savage VM, Charnov EL. 2001 Effects of size and temperature on metabolic rate. *Science* **293**, 2248–2251. (doi:10.1126/science.1061967)
 32. Verberk WCEP, Bilton DT, Calosi P, Spicer JI. 2011 Oxygen supply in aquatic ectotherms: partial pressure and solubility together explain biodiversity and size patterns. *Ecology* **92**, 1565–1572. (doi:10.1890/10-2369.1)
 33. Dejours P. 1981 *Principles of comparative respiratory physiology*. Amsterdam, The Netherlands: Elsevier North-Holland Biomedical Press.
 34. Pörtner HO, Knust R. 2007 Climate change affects marine fishes through the oxygen limitation of thermal tolerance. *Science* **315**, 95–97. (doi:10.1126/science.1135471)
 35. Recsetar MS, Zeigler MP, Ward DL, Bonar SA, Caldwell CA. 2012 Relationship between fish size and upper thermal tolerance. *Trans. Am. Fish. Soc.* **141**, 1433–1438. (doi:10.1080/00028487.2012.694830)
 36. Verberk WCEP, Leuven RS, van der Velde G, Gabel F. 2018 Thermal limits in native and alien freshwater peracarid Crustacea: the role of habitat use and oxygen limitation. *Funct. Ecol.* **32**, 926–936. (doi:10.1111/1365-2435.13050)
 37. Peck LS, Clark MS, Morley SA, Massey A, Rossetti H. 2009 Animal temperature limits and ecological relevance: effects of size, activity and rates of change. *Funct. Ecol.* **23**, 248–256. (doi:10.1111/j.1365-2435.2008.01537.x)
 38. Franken O, Huizinga M, Ellers J, Berg M. 2018 Heated communities: large inter- and intraspecific variation in heat tolerance across trophic levels of a soil arthropod community. *Oecologia* **186**, 311–322. (doi:10.1007/s00442-017-4032-z)
 39. Hessen DO, Daufresne M, Leinaas HP. 2013 Temperature-size relations from the cellular-genomic perspective. *Biol. Rev.* **88**, 476–489. (doi:10.1111/brv.12006)
 40. Partridge L, Barrie B, Fowler K, French V. 1994 Evolution and development of body size and cell size in *Drosophila melanogaster* in response to temperature. *Evolution* **48**, 1269–1276. (doi:10.1111/j.1558-5646.1994.tb05311.x)
 41. Jeffery NW. 2015 Genome size diversity and evolution in the Crustacea. PhD thesis: University of Guelph. See <https://atrium.lib.uoguelph.ca/xmlui/handle/10214/9216>.
 42. Maciak S, Janko K, Kotusz J, Choleva L, Boroń A, Juchno D, Kujawa R, Kozłowski J, Konarzewski M. 2011 Standard metabolic rate (SMR) is inversely related to erythrocyte and genome size in allopolyploid fish of the *Cobitis taenia* hybrid complex. *Funct. Ecol.* **25**, 1072–1078. (doi:10.1111/j.1365-2435.2011.01870.x)
 43. Czarnoleski M, Labecka AM, Dragosz-Kluska D, Pis T, Pawlik K, Kapustka F, Kilarski WM, Kozłowski J. 2018 Concerted evolution of body mass and cell size: similar patterns among species of birds (Galliformes) and mammals (Rodentia). *Biol. Open* **7**, bio-029603. (doi:10.1242/bio.029603)
 44. Kozłowski J, Konarzewski M, Gawelczyk AT. 2003 Cell size as a link between noncoding DNA and metabolic rate scaling. *Proc. Natl Acad. Sci. USA* **100**, 14 080–14 085. (doi:10.1073/pnas.2334605100)
 45. Miettinen TP, Björklund M. 2017 Mitochondrial function and cell size: an allometric relationship. *Trends Cell Biol.* **27**, 393–402. (doi:10.1016/j.tcb.2017.02.006)
 46. Overgaard J, MacMillan HA. 2017 The integrative physiology of insect chill tolerance. *Annu. Rev. Physiol.* **79**, 187–208. (doi:10.1146/annurev-physiol-022516-034142)
 47. Lutterschmidt WI, Hutchison VH. 1997 The critical thermal maximum: history and critique. *Can. J. Zool.* **75**, 1561–1574. (doi:10.1139/z97-783)
 48. Bennett JM *et al.* 2018 GlobTherm, a global database on thermal tolerances for aquatic and terrestrial organisms. *Scient. Data* **5**, 180022. (doi:10.1038/sdata.2018.22)
 49. Froese R, Pauly D. 2018 *FishBase ver. (06/2018)*. See www.fishbase.org.
 50. Tschinkel WR. 2013 The morphometry of *Solenopsis* fire ants. *PLoS ONE* **8**, e79559. (doi:10.1371/journal.pone.0079559)
 51. Cane JH. 1987 Estimation of bee size using intertegular span (Apoidea). *J. Kansas Entomol. Soc.* **60**, 145–147.
 52. Gregory TR. 2019 *Animal genome size database*. See <http://www.genomesize.com/>.
 53. Rollinson N, Rowe L. 2018 Temperature-dependent oxygen limitation and the rise of Bergmann's rule in species with aquatic respiration. *Evolution* **72**, 977–988. (doi:10.1111/evo.13458)
 54. Pagel M. 1994 Detecting correlated evolution on phylogenies: a general method for the comparative analysis of discrete characters. *Proc. R. Soc. Lond. B* **255**, 37–45. (doi:10.1098/rspb.1994.0006)
 55. Hernandez CE, Rodríguez-Serrano E, Avaria-Llautereo J, Inostroza-Michael O, Morales-Pallero B, Boric-Bargetto D, Canales-Aguirre CB, Marquet PA, Meade A. 2013 Using phylogenetic information and the comparative method to evaluate hypotheses in macroecology. *Methods Ecol. Evol.* **4**, 401–415. (doi:10.1111/2041-210X.12033)
 56. Michonneau F, Brown JW, Winter DJ. 2016 rotl: an R package to interact with the Open Tree of Life data. *Methods Ecol. Evol.* **7**, 1476–1481. (doi:10.1111/2041-210X.12593)
 57. Revell LJ. 2010 Phylogenetic signal and linear regression on species data. *Methods Ecol. Evol.* **1**, 319–329. (doi:10.1111/j.2041-210X.2010.00044.x)
 58. Santos M, Castañeda LE, Rezende EL. 2011 Making sense of heat tolerance estimates in ectotherms: lessons from *Drosophila*. *Funct. Ecol.* **25**, 1169–1180. (doi:10.1111/j.1365-2435.2011.01908.x)
 59. Wang S, Tang J, Hansen JD. 2007 Experimental and simulation methods of insect thermal death kinetics. In *Heat treatments for postharvest pest control: theory and practice* (eds J Tang, E Mitcham, S Wang, S Lurie), pp. 105–132. Oxford, UK: CABI Publishing.
 60. Burnham KP, Anderson DR. 2003 *Model selection and multimodel inference: a practical information-theoretic approach*. Berlin, Germany: Springer Science & Business Media.
 61. R Core Team. 2018 *R: a language and environment for statistical computing*. Vienna, Austria: R Foundation for Statistical Computing. See <http://www.R-project.org/>.
 62. Mazerolle MJ. 2019 AICcmodavg: model selection and multimodel inference based on (Q)AIC(c). R package version v2.2-1. See <https://cran.r-project.org/package=AICcmodavg>.
 63. Paradis E, Claude J, Strimmer K. 2004 APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290. (doi:10.1093/bioinformatics/btg412)
 64. Pinheiro J, Bates D, DebRoy S, Sarkar D. 2019 nlme: linear and nonlinear mixed effects models. R package version 3.1-140. See <https://CRAN.R-project.org/package=nlme>.
 65. Revell LJ. 2012 phytools: an R package for phylogenetic comparative biology (and other things). *Methods Ecol. Evol.* **3**, 217–223. (doi:10.1111/j.2041-210X.2011.00169.x)
 66. Breheny P, Burchett W. 2017 Visualization of regression models using visreg. *R Journal* **9**, 56–71. (doi:10.32614/RJ-2017-046)
 67. Spicer JI, Morley SA, Bozinovic F. 2019 Physiological diversity, biodiversity patterns and global climate change: testing key hypotheses involving temperature and oxygen. *Phil. Trans. R. Soc. B* **374**, 20190032. (doi:10.1098/rstb.2019.0032)
 68. Rezende EL, Castañeda LE, Santos M. 2014 Tolerance landscapes in thermal ecology. *Funct. Ecol.* **28**, 799–809. (doi:10.1111/1365-2435.12268)
 69. Terblanche JS, Deere JA, Clusella-Trullas S, Janion C, Chown SL. 2007 Critical thermal limits depend on methodological context. *Proc. R. Soc. B* **274**, 2935–2943. (doi:10.1098/rspb.2007.0985)
 70. Sunday J *et al.* 2019 Thermal tolerance patterns across latitude and elevation. *Phil. Trans. R. Soc. B* **374**, 20190036. (doi:10.1098/rstb.2019.0036)
 71. Hagerman L. 1998 Physiological flexibility; a necessity for life in anoxic and sulphidic habitats. *Hydrobiologia* **375**, 241–254. (doi:10.1023/A:1017033711985)
 72. Urbina MA, Glover CN. 2013 Relationship between fish size and metabolic rate in the oxyconforming inanga *Galaxias maculatus* reveals size-dependent strategies to withstand hypoxia. *Physiol. Biochem. Zool.* **86**, 740–749. (doi:10.1086/673727)
 73. Sloman KA, Wood CM, Scott GR, Wood S, Kajimura M, Johannsson OE, Almeida-Val VM, Val AL. 2006 Tribute to RG Boutilier: the effect of size on the physiological and behavioural responses of oscar, *Astronotus ocellatus*, to hypoxia. *J. Exp. Biol.* **209**, 1197–1205. (doi:10.1242/jeb.02090)

74. Lv X, Xie H, Xia D, Shen C, Li J, Luo Y. 2018 Mass scaling of the resting and maximum metabolic rates of the black carp. *J. Comp. Physiol. B* **188**, 591–598. (doi:10.1007/s00360-018-1154-5)
75. Gregory TR. 2001 The bigger the C-value, the larger the cell: genome size and red blood cell size in vertebrates. *Blood Cells Mol. Dis.* **27**, 830–843. (doi:10.1006/bcmd.2001.0457)
76. Czarnoleski M, Cooper BS, Kierat J, Angilletta MJ. 2013 Flies developed small bodies and small cells in warm and in thermally fluctuating environments. *J. Exp. Biol.* **216**, 2896–2901. (doi:10.1242/jeb.083535)
77. Szarski H. 1983 Cell size and the concept of wasteful and frugal evolutionary strategies. *J. Theor. Biol.* **105**, 201–209. (doi:10.1016/S0022-5193(83)80002-2)
78. Frazier MR, Woods HA, Harrison JF. 2001 Interactive effects of rearing temperature and oxygen on the development of *Drosophila melanogaster*. *Physiol. Biochem. Zool.* **74**, 641–650. (doi:10.1086/322172)
79. Overgaard J, Tomčala A, Sørensen JG, Holmstrup M, Krogh PH, Šimek P, Košťál V. 2008 Effects of acclimation temperature on thermal tolerance and membrane phospholipid composition in the fruit fly *Drosophila melanogaster*. *J. Insect Physiol.* **54**, 619–629. (doi:10.1016/j.jinsphys.2007.12.011)
80. Ronges D, Walsh JP, Sinclair BJ, Stillman JH. 2012 Changes in extreme cold tolerance, membrane composition and cardiac transcriptome during the first day of thermal acclimation in the porcelain crab *Petrolisthes cinctipes*. *J. Exp. Biol.* **215**, 1824–1836. (doi:10.1242/jeb.069658)
81. Overgaard J, Sørensen JG, Petersen SO, Loeschke V, Holmstrup M. 2005 Changes in membrane lipid composition following rapid cold hardening in *Drosophila melanogaster*. *J. Insect Physiol.* **51**, 1173–1182. (doi:10.1016/j.jinsphys.2005.06.007)
82. Slotsbo S, Sørensen JG, Holmstrup M, Kostal V, Kellermann V, Overgaard J. 2015 Tropical to subpolar gradient in phospholipid composition suggests adaptive tuning of biological membrane function in drosophilids. *Funct. Ecol.* **30**, 759–768. (doi:10.1111/1365-2435.12568)
83. Stevens MM, Jackson S, Bester SA, Terblanche JS, Chown SL. 2010 Oxygen limitation and thermal tolerance in two terrestrial arthropod species. *J. Exp. Biol.* **213**, 2209–2218. (doi:10.1242/jeb.040170)
84. Hulbert AJ, Else PL. 2005 Membranes and the setting of energy demand. *J. Exp. Biol.* **208**, 1593–1599. (doi:10.1242/jeb.01482)
85. Fordyce JA, Shapiro AM. 2003 Another perspective on the slow-growth/high-mortality hypothesis: chilling effects on swallowtail larvae. *Ecology* **84**, 263–268. (doi:10.1890/0012-9658(2003)084[0263:APOTSG]2.0.CO;2)
86. Kellermann V, Overgaard J, Hoffmann AA, Fløjgaard C, Svenning J-C, Loeschke V. 2012 Upper thermal limits of *Drosophila* are linked to species distributions and strongly constrained phylogenetically. *Proc. Natl Acad. Sci. USA* **109**, 16 228–16 233. (doi:10.1073/pnas.1207553109)
87. Marshall DJ, Rezende EL, Baharuddin N, Choi F, Helmuth B. 2015 Thermal tolerance and climate warming sensitivity in tropical snails. *Ecol. Evol.* **5**, 5905–5919. (doi:10.1002/ece3.1785)
88. Gutiérrez-Pesquera LM, Tejedo M, Olalla-Tárraga M, Duarte H, Nicieza A, Solé M. 2016 Testing the climate variability hypothesis in thermal tolerance limits of tropical and temperate tadpoles. *J. Biogeogr.* **43**, 1166–1178. (doi:10.1111/jbi.12700)
89. Calosi P, Bilton DT, Spicer JJ, Votier SC, Atfield A. 2010 What determines a species' geographical range? Thermal biology and latitudinal range size relationships in European diving beetles (Coleoptera: Dytiscidae). *J. Anim. Ecol.* **79**, 194–204. (doi:10.1111/j.1365-2656.2009.01611.x)
90. Diamond SE, Chick LD. 2017 The Janus of macrophysiology: stronger effects of evolutionary history, but weaker effects of climate on upper thermal limits are reversed for lower thermal limits in ants. *Curr. Zool.* **64**, 223–230. (doi:10.1093/cz/zox072)
91. Leiva FP, Calosi P, Verberk WCEP. 2019 Data from: Scaling of thermal tolerance with body mass and genome size in ectotherms: a comparison between water- and air-breathers. Dryad Digital Repository. (doi:10.5061/dryad.878vn25)