

EDGE ARTICLE

Cite this: *Chem. Sci.*, 2021, 12, 11484

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 24th April 2021
Accepted 16th July 2021

DOI: 10.1039/d1sc02295d

rsc.li/chemical-science

Influence of stereochemistry on the activity of rapadocin, an isoform-specific inhibitor of the nucleoside transporter ENT1†

Yuefan Wang,[‡] Hanjing Peng,[‡] Zufeng Guo,[‡] Brett R. Ullman,^d Kana Yamamoto,^d Sam Y. Hong,[‡] and Jun O. Liu[‡]

Rapadocin is a novel rapamycin-inspired polyketide–tetrapeptide hybrid macrocycle that possesses highly potent and isoform-specific inhibitory activity against the human equilibrative nucleoside transporter 1 (hENT1). Rapadocin contains an epimerizable chiral center in phenylglycine and an olefin group, and can thus exist as a mixture of four stereoisomers. Herein, we report the first total synthesis of the four stereoisomers of rapadocin using two different synthetic strategies and the assignment of their structures. The inhibitory activity of each of the four synthetic isomers on both hENT1 and hENT2 was determined. It was found that the stereochemistry of phenylglycine played a more dominant role than the configuration of the olefin in the activity of rapadocin. These findings will guide the future design and development of rapadocin analogs as new modulators of adenosine signaling.

Introduction

Rapamycin and FK506 are macrocyclic natural products with potent immunosuppressive activity, making them powerful drugs for preventing graft rejection and treating a variety of autoimmune diseases. These drugs work by inducing the formation of ternary complexes between FK506-binding protein (FKBP) and their targets, mTOR and calcineurin, respectively.^{1,2} The ability of these drugs to bind FKBP confers a number of advantages, including superior bioavailability and distribution *in vivo*.³ Inspired by the unique structure and mode of action of these drugs, we recently developed rapamycin-like hybrid macrocycles by replacing the effector domain of rapamycin with tetrapeptides and named them rapafucins.⁴ High-throughput screening led to the discovery of rapadocin, a highly potent, isoform-specific, and FKBP-dependent inhibitor of the human equilibrative nucleoside transporter 1 (hENT1) with an IC₅₀ of 5 nM for adenosine uptake.⁴ ENTs are an important family of nucleoside transporters, which play a major role in the

transport of nucleoside-based small molecular drugs for cancer and several viral diseases in addition to their physiological substrate adenosine.^{5,6} Rapadocin has been shown to protect against ischemia-reperfusion kidney injury by potentiating adenosine signaling through blocking adenosine uptake, making it a promising lead to develop drugs for treating kidney ischemia/reperfusion injury and other diseases.^{7–9}

Rapadocin shares the two-domain architecture of rapamycin and FK506: an FKBP-binding domain (FKBD) and an effector domain consisting of tetrapeptides.^{10–12} In our previous synthesis of the rapafucin library,⁴ the effector domain in rapadocin is generated using solid-phase peptide synthesis before it is coupled to a preassembled FKBD. The resulting FKBD-peptide conjugate is cyclized and released from the solid support *via* ring-closing metathesis (RCM). Under those conditions, we found that the phenylglycine residue can undergo epimerization during solid-phase synthesis^{13,14} and the olefin formed during the

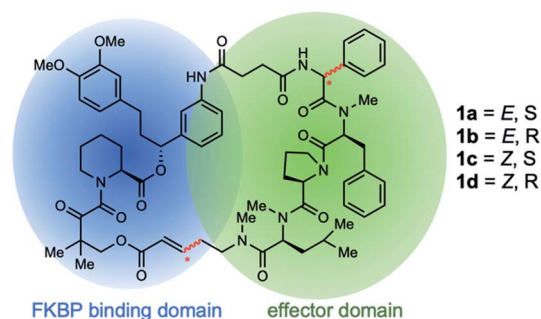


Fig. 1 Possible stereoisomers of rapadocin.

^aDepartment of Pharmacology, Johns Hopkins School of Medicine, 725 North Wolfe Street, Baltimore, MD 21205, USA. E-mail: joliu@jhu.edu

^bSJ Yan and HJ Mao Laboratory of Chemical Biology, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA

^cCenter for Novel Target and Therapeutic Intervention, Institute of Life Sciences, Chongqing Medical University, Chongqing, 400016, China

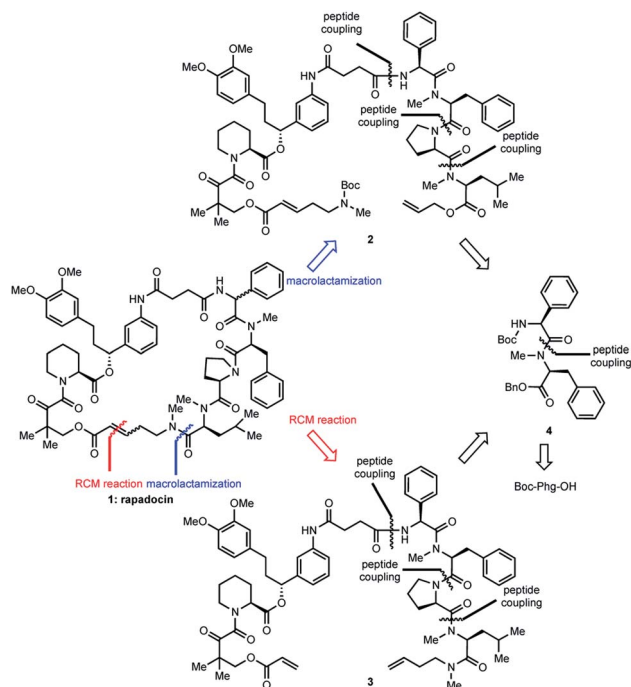
^dRapafusyn Pharmaceuticals Inc., Baltimore, MD 21205, USA

^eSidney Kimmel Comprehensive Cancer Center, Johns Hopkins University School of Medicine, Baltimore, MD 21205, USA

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d1sc02295d

‡ These authors contributed equally to this work.





Scheme 1 Retrosynthetic analyses of rapadocin.

RCM reaction can assume either *Z*- or *E*-configuration (Fig. 1).¹⁵ Due to the complexity of the rapadocin structure, it was difficult to separate and detect each of the four possible stereoisomers for further characterization, leaving open the question of whether the four stereoisomers differ in their inhibitory activities towards hENT1. To address those uncertainties, we embarked on the total synthesis of each of the four possible stereoisomers of rapadocin to determine their biological activity.

Results and discussion

Design of solution phase synthesis of rapadocin stereoisomers

When we did the solid phase synthesis of rapadocin, we found a significant racemization of the phenylglycine moiety. The

configuration of the RCM reaction generated double bond was also difficult to confirm using NMR. We designed two different routes for the total synthesis of rapadocin **1** (Scheme 1). The retrosynthetic planning of macrolactamization began with the disconnection of the macrocycle at the FKBD-mLeu amide bond (**2**) and then on the either side of the *D*-proline residue, leading to an olefin-containing FKBD and a dipeptide (**4**) precursor. This synthetic route required the development of an efficient synthetic approach for the stereoselective synthesis of **4** bearing a highly racemization-prone phenylglycine moiety.^{16–20} The second route is based on RCM for macrocyclization from linearized precursor **3**, which can be broken down on either side of the *D*-proline residue to the precursor dipeptide **4**. Both synthetic routes required the development of an efficient synthesis of dipeptide **4**. The late-stage solution-phase RCM reaction is similar to that used in the solid phase synthesis,⁴ while the macrolactamization strategy could confirm the double bond configuration.

Two synthetic routes of rapadocin isomers

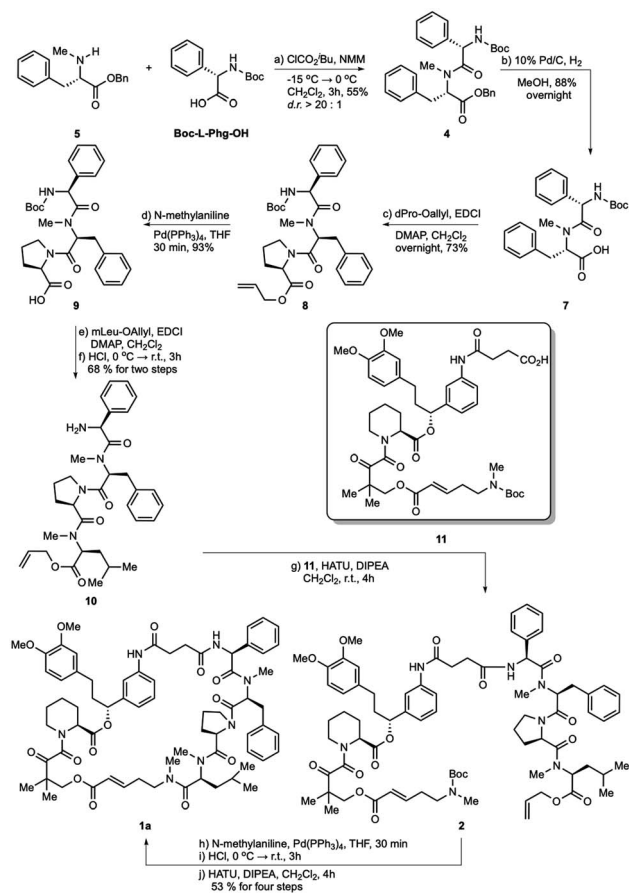
The synthesis of dipeptide **4** commenced with diastereoselective amidation from commercially available Boc-*L*- α -phenylglycine (Boc-Phg-OH). Boc-Phg-OH was activated as a mixed acid anhydride and coupled with *N*-methyl phenylalanine **5** to give the highly diastereoselective dipeptide **4** with an excellent d.r. ratio (>20 : 1),^{20,21} as determined by ¹H-NMR (Table 1).

The hydrogenolysis of benzyl ester **4** with 10% Pd/C gave the corresponding carboxylic acid **7** in 88% yield (Scheme 2). The amidation of **7** with allyl ester of *D*-proline in the presence EDCI and DMAP in CH₂Cl₂ afforded tripeptide **8** in 73% yield. Further allyl deprotection of **8** under Tsuji–Trost allylation conditions in the presence of a catalytic amount of Pd(PPh₃)₄ afforded carboxylic acid **9** in 93% yield.²² The final methyl-leucine residue was attached using coupling agents EDCI and DMAP in CH₂Cl₂ to afford the tetrapeptide, which was then reacted with hydrogen chloride dioxane solution (4 M) to expose the N-terminus of tetrapeptide **10**.

Table 1 Survey of conditions for amidation reaction

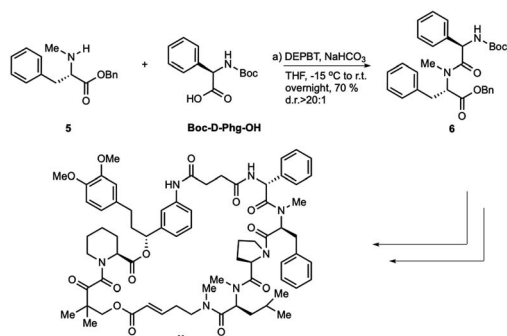
Entry	Reagent (equiv.)	Base (equiv.)	Temperature	Solvent	Ratio (4 : 6) ^a
1	HATU (1.5)	DIPEA (3)	0 °C to r.t.	CH ₂ Cl ₂	2 : 3
2	EDCI (1.6)	DMAP (2.0)	0 °C to r.t.	CH ₂ Cl ₂	1 : 1
3	EDCI/HOAT (1.5/1.5)	NaHCO ₃ (1.5)	0 °C to r.t.	THF	3 : 2
4	BOPCI (1.0)	NMM (2.0)	–15 °C to r.t.	CH ₂ Cl ₂	5 : 1
5	DEPBT (1.6)	NaHCO ₃ (1.6)	–15 °C to r.t.	THF	7 : 1
6	ClCO ₂ ^t Bu (1.0)	NMM (1.5)	–15 °C to 0 °C	CH ₂ Cl ₂	>20 : 1 ^b

^a The ratio of the phenylglycine epimers in the dipeptide was determined by ¹H-NMR. ^b This is the most optimal value attained.



Scheme 2 Total synthesis of rapadocin *via* macrolactamization strategy.

With tetrapeptide fragment **10** in hand, we explored the pathway for the completion of the total synthesis. Tetrapeptide **10** was coupled with Boc-*E*-FKBD fragment **11** upon treatment with HATU and Hünig's base in CH_2Cl_2 to obtain a linear product **2**. The fragment **11** was provided by Affinity Research Chemicals with an established *E*- or *Z*-configuration of the olefin moiety. The deprotection of the allyl ester of **2** with $\text{Pd}(\text{PPh}_3)_4$ was followed by the removal of the Boc group with dioxane HCl solution (4 M). Finally, the lactamization of the acyclic rapadocin with HATU and Hünig's base in CH_2Cl_2 gave rapadocin **1a** in 53% yield over four steps.



Scheme 3 Synthesis of rapadocin isomers.

With the successful synthesis of the first rapadocin isomer **1a** with a $1(S)$ -Phg-(*m*Phe) moiety and *E* configuration of the double bond, we proceeded to prepare **1b** and other isomers. As shown in Scheme 3, **1b** was readily made using the same synthetic procedure as that for **1a** with the dipeptide **6** from Boc-*D*-Phe-OH instead of Boc-*L*-Phe-OH. We found that when DEPBT and NaHCO_3 were used as coupling reagents for the synthesis of **6**, the yield was higher than that for the formation of dipeptide **4** with a 70% yield and the d.r. ratio of the product was greater than 20 : 1, as determined by $^1\text{H-NMR}$. Compounds **1c** and **1d** were synthesized with the same synthetic strategy using the *Z*-FKBD fragment.

For polypeptides and cyclic peptides to which rapafucin belong, $^1\text{H-}$ and $^{13}\text{C-NMR}$ are no longer suited to judge the purity of those compounds, as each amide bond may exist in two rotamer conformations and different rotamer conformations could show distinct spectra, complicating the $^1\text{H-}$ and $^{13}\text{C-NMR}$ spectra. This situation is made worse in the case of rapadocin due to the different rotamer conformations arising from the piperocic ester of FKBD that has been seen in FK506 and rapamycin. We thus resorted to using a combination of NMR, HPLC and high-resolution MS to confirm the purity of the different stereoisomers of rapadocin.^{23–25} Based on the HSQC result of **1a** (see the ESI[†]), we confirmed that the double bond proton is in the range of ppm 6.0–5.5 in the $^1\text{H-NMR}$ spectrum. To provide further support for the successful synthesis of all four isomers **1a–1d**, we compared the four $^1\text{H-NMR}$ spectra for olefin protons. As shown in Fig. 2, the four isomers showed different $^1\text{H-NMR}$, suggestion that four isomers are not the same compound.

After the successful syntheses of the rapadocin isomers *via* the macrolactamization strategy, we investigated the RCM strategy that mimics the solid-phase synthesis to verify the configuration of the double bond formed during the RCM reaction and to explore more efficient synthetic routes to pure rapadocin stereoisomers.

As shown in Scheme 4, tripeptide **9** was coupled with methyl-leucine derivative **12** upon treatment with HATU and Hünig's base in CH_2Cl_2 to obtain tetrapeptide **13** in 80% yield. Intermediate **13** was reacted with hydrogen chloride dioxane

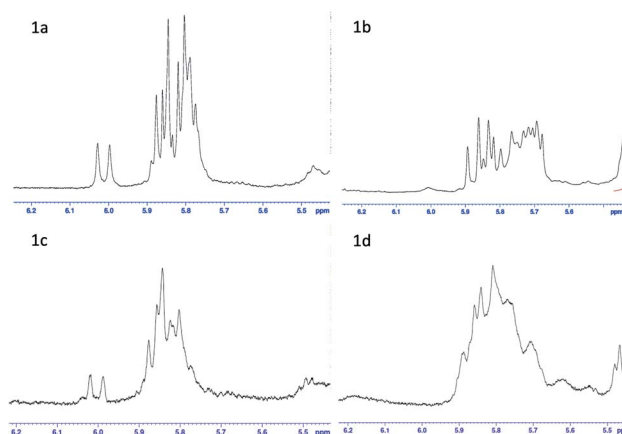
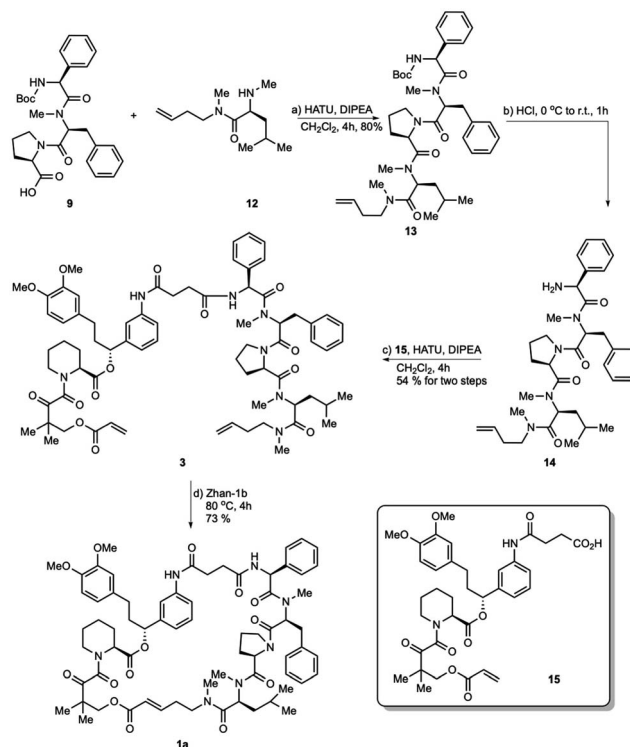


Fig. 2 $^1\text{H-NMR}$ comparison of rapadocin four isomers.



Scheme 4 Total synthesis of rapadocin via the RCM strategy.

solution (4 M) to expose the N-terminus of tetrapeptide **14**, which was then condensed with FKBD **15** to give linear product **3** in 54% yield over two steps. FKBD **15** was made using a reported method.⁴ To complete the total synthesis, **3** was treated with the **Zhan-1b** catalyst to give the final product,^{26,27} which could be the isomer **1a**, **1c** or a mixture of both. By the comparison of the ¹H- and ¹³C-NMR spectra and optical rotations of the product from the RCM reaction with those obtained using the macrolactamization strategy, the RCM product was assigned as **1a** ($[\alpha]_D^{23} = -36.5$ and $[\alpha]_D^{23} = -35.9$). Thus, the double bond configuration of rapadocin from the RCM reaction used in previous study was confirmed to have the *E* configuration.

The configuration of the olefin has a major impact on the stability of the stereoisomers. In contrast to **1a** and **1b**, **1c** and **1d** with the *Z* olefin configuration are unstable in solution. At room temperature, *Z* isomers in DMSO changed to a new undefined compound with a molecular weight of 1270 in one week. At -20 °C, *Z*-isomers underwent the same change in one month. In contrast, *E*-isomers are stable in solution at -20 °C for over a year. This result suggests that the *E*-isomers of rapadocins with the current backbone structures are thermodynamically favored by the RCM reaction.

Biological evaluations of the four isomers

With each of the four isomers of rapadocin in hand, we took advantage of a window of time before the *Z*-isomers of rapadocin underwent appreciable degradation and determined their inhibitory activity toward hENT1-mediated nucleoside uptake

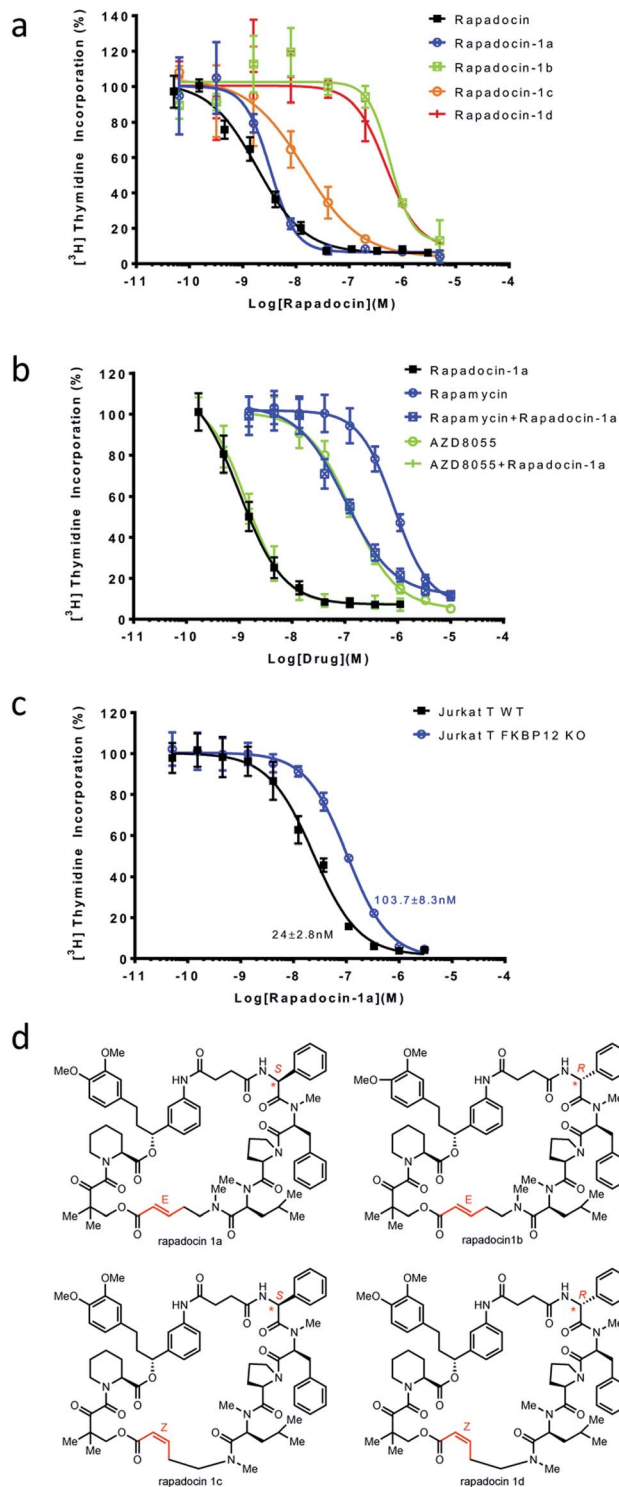


Fig. 3 Inhibition of hENT1 by different stereoisomers of rapadocin. (a) Inhibition of [³H]-thymidine uptake in PK15 cell lines exclusively expressing hENT1 by the four stereoisomers of rapadocin. (b) Rapamycin, but not AZD8055, antagonizes isomer **1a** inhibition of hENT1-mediated thymidine uptake. The dose-response curves were obtained using [³H]-thymidine uptake in PK15-ENT1 cells. (c) Knockout of FKBP12 imparts resistance to isomer **1a**. Dose response curves for the inhibition of [³H]-thymidine uptake by isomer **1a** in wild type (WT) and FKBP12 knockout Jurkat T cells. Error bars represent s.d. Data are presented as mean \pm s.d.; $n = 3$ independent experiments. (d) Structures of the four rapadocin isomers.

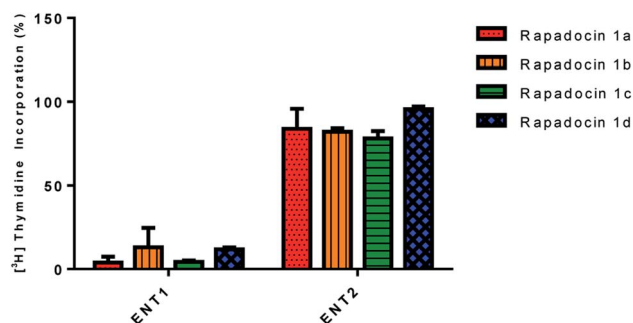


Fig. 4 Effects of rapadocin isomers on hENT1 and hENT2.

using [^3H]-thymidine uptake assay in porcine kidney (PK15) cell lines exclusively expressing hENT1.²⁸ Interestingly, rapadocin isomer **1a**, with a *L*-Phg moiety and *E*-double bond, was the most potent ($\text{IC}_{50} = 3.3 \text{ nM}$) among the four stereoisomers. In contrast, the other three isomers **1b**, **1c**, and **1d** exhibited lower potency against hENT1 with IC_{50} values of 589.2 nM, 14.5 nM, and 469.3 nM, respectively (Fig. 3a). These results suggested that isomer **1a** is the most active component of rapadocin from the solid-phase synthesis. As the potency of the RCM reaction product (rapadocin) is comparable to that of isomer **1a**, it is likely that this stereoisomer is the predominant product from the RCM reaction. The fact that isomers **1a** and **1c** are significantly more active than **1b** and **1d** also suggests that the stereochemistry of the phenylglycine residue has a much greater influence on the activity of rapadocin than the configuration of the olefin moiety.

To further assess the dependence of isomer **1a** on endogenous FKBP, we determined mutual antagonism between two mTOR inhibitors, rapamycin and AZD8055, and rapadocin isomer **1a** as previously described.⁴ As expected, combining rapamycin (5 μM) with isomer **1a** significantly decreased its potency (Fig. 3b). However, the high concentration of AZD8055 (5 μM), an FKBP-independent mTOR inhibitor, had a negligible effect on the activity of isomer **1a**. We also determined the inhibitory activity of isomer **1a** in either a wild type Jurkat T or an FKBP12 knockout cell line using the [^3H]-thymidine incorporation assay. The FKBP12 knockout Jurkat T cells also gained resistance to **1a** (Fig. 3c). These results suggested that, like rapadocin, the inhibitory activities of the four isomers, especially isomer **1a**, are also dependent on endogenous FKBP12.

In addition, we also investigated the inhibition of [^3H]-thymidine uptake by 5 μM of rapadocin isomers in PK15 cell lines exclusively expressing either hENT1 or hENT2. Similar to rapadocin, isomers **1a–1d** had little effect on [^3H]-thymidine uptake in the PK15-ENT2 cell line, suggesting that the four isomers retained high specificity for hENT1 (Fig. 4).

Conclusion

In summary, we designed two synthetic strategies to synthesize the four stereoisomers of rapadocin. We confirmed that the structure of the most active isomer of rapadocin contains *L*-Phg (*S* stereochemistry) with an *E* configuration of the double bond

(**1a**). We also developed an RCM strategy to confirm our structural elucidation result and efficient synthesis of the most active isomer **1a**. We found that isomers with an *E*-configuration of the double bond are much more stable than those with a *Z*-configuration. We observed that the stereochemistry of phenylglycine played a significant role in the observed FKBP12-dependent and isoform-selective inhibition of hENT1. The complete assignment of the phenylglycine stereochemistry and the double bond configuration of the most potent stereoisomer of rapadocin with efficient scalable solution-phase synthetic routes will greatly facilitate the future development of rapadocin as a new molecular probe for hENT1 and as a lead for developing drugs to treat ischemic kidney reperfusion injury among other diseases through the modulation of adenosine signaling.

Data availability

All additional experimental data and procedures are included in the ESI.†

Author contributions

J. L. conceived the project, oversaw the execution of the project and edited the manuscript. Y. W. designed the synthetic routes, completed the total synthesis and wrote the first manuscript. H. P. helped to finish the synthesis of **1c** and **1d**. Z. G. and H. P. performed the biological assays and analysed the data. B. R. U., K. Y. and S. Y. H. assisted the chemical synthesis.

Conflicts of interest

The authors declare conflict of interest. Patent applications covering rapadocin have been filed by Johns Hopkins University and licensed to Rapafusyn Pharmaceuticals, Inc. J. O. L. is a co-founder of, as well as a Scientific Advisory Board Member for, Rapafusyn Pharmaceuticals, Inc. This arrangement has been reviewed and approved by the Johns Hopkins University in accordance with its conflict of interest policies.

Acknowledgements

This work was made possible by the NIH Director's Pioneer Award, the Flight Attendant Medical Research Institute, a generous gift from Mr Shengjun Yan and Ms. Hongju Mao and NCI (P30CA006973) (J. O. L.) and a Damon Runyon Postdoctoral Fellowship (H. P.). We thank Dr Yixin Han in Professor E. J. Corey's lab (Harvard) for help with the optical rotation analysis.

References

- J. Heitman, N. R. Movva and M. N. Hall, *Science*, 1991, **253**, 905–909.
- J. Liu, J. D. Farmer, Jr, W. S. Lane, J. Friedman, I. Weissman and S. L. Schreiber, *Cell*, 1991, **66**, 807–815.

- 3 P. S. Marinec, L. Chen, K. J. Barr, M. W. Mutz, G. R. Crabtree and J. E. Gestiwicki, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 1336–1341.
- 4 Z. Guo, S. Y. Hong, J. Wang, S. Rehan, W. Liu, H. Peng, M. Das, W. Li, S. Bhat, B. Peiffer, B. R. Ullman, C. M. Tse, Z. Tarmakova, C. Schiene-Fischer, G. Fischer, I. Coe, V. O. Paavilainen, Z. Sun and J. O. Liu, *Nat. Chem.*, 2019, **11**, 254–263.
- 5 J. D. Young, S. Y. Yao, J. M. Baldwin, C. E. Cass and S. A. Baldwin, *Mol. Aspects Med.*, 2013, **34**, 529–547.
- 6 R. C. Boswell-Casteel and F. A. Hays, *Nucleosides, Nucleotides Nucleic Acids*, 2017, **36**, 7–30.
- 7 Y. J. Day, L. Huang, H. Ye, J. Linden and M. D. Okusa, *Am. J. Physiol. Ren. Physiol.*, 2005, **288**, F722–F731.
- 8 C. M. Lappas, Y. J. Day, M. A. Marshall, V. H. Engelhard and J. Linden, *J. Exp. Med.*, 2006, **203**, 2639–2648.
- 9 A. Grenz, H. Osswald, T. Eckle, D. Yang, H. Zhang, Z. V. Tran, K. Klingel, K. Ravid and H. K. Eltzschig, *PLoS Med.*, 2008, **5**, e137.
- 10 J. P. Griffith, J. L. Kim, E. E. Kim, M. D. Sintchak, J. A. Thomson, M. J. Fitzgibbon, M. A. Fleming, P. R. Caron, K. Hsiao and M. A. Navia, *Cell*, 1995, **82**, 507–522.
- 11 C. R. Kissinger, *et al.*, *Nature*, 1995, **378**, 641–644.
- 12 J. Choi, J. Chen, S. L. Schreiber and J. Clardy, *Science*, 1996, **273**, 239–242.
- 13 C. Brieke and M. J. Cryle, *Org. Lett.*, 2014, **16**, 2454–2457.
- 14 C. Liang, M. A. M. Behnam, T. R. Sundermann and C. D. Klein, *Tetrahedron Lett.*, 2017, **58**, 2325–2329.
- 15 A. H. Hoveyda, *J. Org. Chem.*, 2014, **79**, 4763–4792.
- 16 G. G. Smith and T. Sivakua, *J. Org. Chem.*, 1983, **48**, 627–634.
- 17 M. Bodanszky and A. Bodanszky, *Chem. Commun.*, 1967, **12**, 591–593.
- 18 K. C. Nicolaou, *et al.*, *Chem.–Eur. J.*, 1999, **5**, 2602–2621.
- 19 D. A. Evans, J. L. Katz, G. S. Peterson and T. Hintermann, *J. Am. Chem. Soc.*, 2001, **123**, 12411–12413.
- 20 L. Tan and D. Ma, *Angew. Chem., Int. Ed.*, 2008, **47**, 3614–3617; *Angew. Chem.*, 2008, **120**, 3670–3673.
- 21 T. Noguchi, S. Jung and N. Imai, *Tetrahedron Lett.*, 2014, **55**, 394–396.
- 22 F. Guibé, *Tetrahedron*, 1998, **54**, 2967–3042.
- 23 P. Karuso, H. Kessler and D. F. Mierke, *J. Am. Chem. Soc.*, 1990, **112**, 9434–9436.
- 24 D. F. Mierke, P. Schmieder, P. Karuso and H. Kessler, *Helv. Chim. Acta*, 1991, **74**, 1027–1047.
- 25 H. Kessler, R. Haessner and W. Schüler, *Helv. Chim. Acta*, 1993, **76**, 117–130.
- 26 A. Kajetanowicz and K. Grela, *Angew. Chem., Int. Ed.*, 2021, **60**, 13738–13756.
- 27 T. K. Olszewski, M. Bieniek, K. Skowerski and K. Grela, *Synlett*, 2013, **24**, 903–919.
- 28 J. L. Ward, A. Serali, Z. P. Mo and C. M. Tse, *J. Biol. Chem.*, 2000, **275**, 8375–8381.