GENOME SEQUENCES





Draft Genome Sequences of Three *Pseudomonas fluorescens* Strains Isolated from Marine Sponges Harvested off the West Coast of Ireland

José A. Gutiérrez-Barranquero,^{a*} María L. Parages,^{a*} Alan D. W. Dobson,^{b,c} F. Jerry Reen,^{a,b} Fergal O'Gara^{a,d,e}

^aBIOMERIT Research Centre, School of Microbiology, University College Cork, Cork, Ireland ^bSchool of Microbiology, University College Cork, Cork, Ireland ^cEnvironmental Research Institute, University College Cork, Cork, Ireland ^dTelethon Kids Institute, Perth Children's Hospital, Perth, WA, Australia ^eSchool of Pharmacy and Biomedical Sciences, Curtin Health Innovation Research Institute, Curtin University, Perth, WA, Australia

ABSTRACT Three *Pseudomonas* sp. strains isolated from marine sponges have shown potential quorum sensing inhibition (QSI) activity. We sequenced the draft genomes of the three strains with the goal of determining which genes or gene cluster(s) could be potentially involved in the QSI activity. Average nucleotide identity (ANI) and phylogenetic analysis classified the three strains as belonging to the *Pseudomonas fluorescens* species.

Pseudomonas is a Gram-negative bacterial genus belonging to the Gammaproteobacteria class, whose members are known to colonize and survive in a wide range of diverse environments, mainly due to inherent broad metabolic diversity (1, 2). This bacterial genus encompasses different species involved in the degradation of xenobiotic compounds (3), human and plant pathogenesis (4, 5), and plant growth promotion and biocontrol (6). Recently, the role of some *Pseudomonas* sp. strains in the inhibition of quorum sensing (QS) signaling systems has been reported (7–9). Three *Pseudomonas* sp. strains (B98C39, B98SK52, and B98SM8) were isolated from 2 different marine sponges, belonging to the *Hexactinellida* class, that were collected off the west coast of Ireland as part of a marine biodiscovery cruise in May 2010. The isolation of these *Pseudomonas* sp. strains and their QS inhibition (QSI) activities were described previously (9). However, the genetic basis of the QSI activities remained unknown. Therefore, in order to identify which genes or gene cluster(s) could potentially be involved in the QSI activity, the draft genome sequencing of these strains was completed.

Overnight shaking cultures grown in lysogeny broth at 23°C for the three *Pseu-domonas* sp. strains were used to perform total DNA isolation using the UltraClean microbial DNA isolation kit (MO BIO Laboratories, Inc., Carlsbad, CA, USA). DNA libraries were prepared using a TruSeq exome library preparation kit. The draft genome sequencing was performed by the Beijing Genomics Institute (China) using the Illumina HiSeq 2000 sequencing platform with paired-end reads and a read length of 90 bp for B98C39. The HiSeq 4000 platform, with paired-end reads and a read length of 150 bp, was used for B98SK52 and B98SM8. In order to obtain high-quality reads for assembly, the FASTA/Q file manipulation tool readfq.v5 (10, 11) was used for quality trimming using the same parameters as described previously (10). Thus, the high-quality-filtered reads were all 90 bp in the case of B98C39 and 150 bp in the cases of B98SK52 and B98SM8. The assembly of high-quality filtered reads was performed using SOPAdenovo v2.04 with default parameters. Genome sequence annotation and gene identification were carried out with the Rapid Annotations using Subsystems Technology (RAST) v2.0 server (using default parameters and RASTtk for the annotation scheme) (12, 13) and

Citation Gutiérrez-Barranquero JA, Parages ML, Dobson ADW, Reen FJ, O'Gara F. 2020. Draft genome sequences of three *Pseudomonas fluorescens* strains isolated from marine sponges harvested off the west coast of Ireland. Microbiol Resour Announc 9:e00457-20. https://doi.org/10.1128/MRA .00457-20.

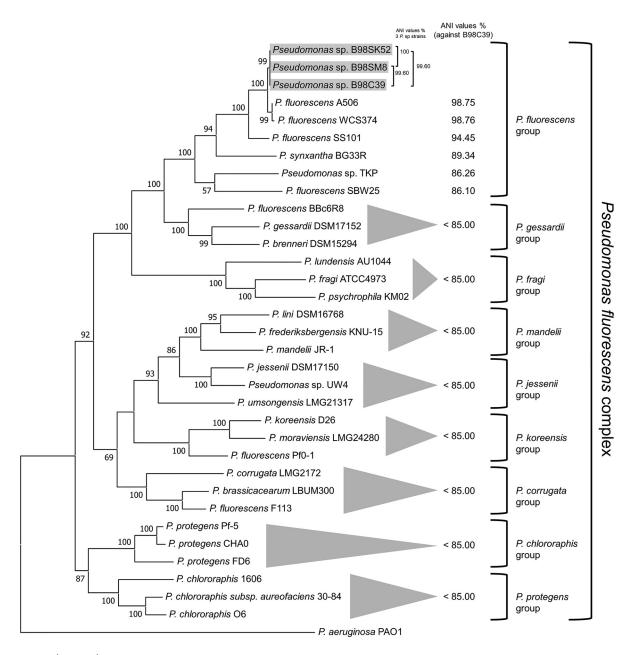
Editor Frank J. Stewart, Georgia Institute of Technology

Copyright © 2020 Gutiérrez-Barranquero et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Fergal O'Gara, f.ogara@ucc.ie.

* Present address: José A. Gutiérrez-Barranquero, Instituto de Hortofruticultura Subtropical y Mediterránea La Mayora, Departamento de Microbiogía, Facultad de Ciencias, Universidad de Málaga, Málaga, Spain; María L. Parages, Departamento de Ecología, Facultad de Ciencias, Universidad de Málaga, Málaga, Spain.

Received 22 April 2020 **Accepted** 1 May 2020 **Published** 21 May 2020



0.020

FIG 1 Phylogenetic analysis using the complete nucleotide sequences of the housekeeping genes *gyrB* and *rpoD*. Thirty-three *Pseudomonas* strains belonging to the eight phylogenetic groups within the *Pseudomonas fluorescens* complex were used. Phylogenetic distribution was determined by the maximum likelihood method and the Tamura-Nei model, with 100 bootstrap replicates, using MEGA7. The three *Pseudomonas fluorescens* strains sequenced in this study are highlighted with gray boxes. The *P. aeruginosa* PAO1 strain was used as an outgroup. ANI values are represented as percentages. According to previous work (20), the ANI value threshold for differentiating species in the *Pseudomonas fluorescens* complex is 85%. Thus, an ANI value over 85% is considered to indicate the same species. The data for the strains used to generate this figure were obtained from the NCBI database, with the following accession numbers: CP003041, CP007638, AHPN00000000, AHPP00000000, CP023466, CP005960, NIWT01000000, NC_019670, LT629767, CP014947, LT629788, CP000094, LT629798, CP012680, CP003150, CP032358, CP003190, CP031396, CP011110, CM001490, CM001559, and AE004091.

the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (using default parameters). The main characteristics of the draft genome sequences obtained, including their accession numbers, are summarized in Table 1.

Genome mining of the three strains identified orthologs of the quiP (14) and pvdQ (15) genes, both of which encode acylase enzymes that were previously shown to

P. fluorescens strain	GenBank accession no.	Genome size (Mb)	SRA accession no.	No. of contigs	No. of scaffolds	N ₅₀ (bp)	Mean coverage (×)	No. of CDS ^a	G+C content (%)
B98C39	JAAXCP000000000	6.21	SRX8046944	282	51	44,154	41.50	5,459	59.9
B98SK52	JAAXCO000000000	6.21	SRX8046945	73	71	259,749	40.90	5,566	59.8
B98SM8	JAAXCN00000000	6.21	SRX8046946	97	97	149,291	42	5,568	59.8

TABLE 1 Accession numbers and genome assembly features

^a CDS, coding DNA sequences.

control the QS system in *Pseudomonas aeruginosa* through the degradation of *N*-3-oxododecanoyl-homoserine lactone. Furthermore, a nonribosomal peptide synthetase secondary metabolite gene cluster was identified using antiSMASH bacterial version 5.0 (16); the cluster was related to the production of salicylic acid and pseudomonine metabolites (17). Interestingly, salicylic acid was reported previously to inhibit the production of *N*-acylhomoserine lactones in bacteria (18). Considering these results, further *in vitro* investigations will be required to elucidate which enzymes or metabolites could be involved in the QSI activity of these *Pseudomonas* sp. strains. Phylogenetic analysis and pairwise ANI analysis using JSpeciesWS were performed (19) (Fig. 1). The sequenced strains were classified as belonging to the species *Pseudomonas fluorescens*.

Data availability. The draft genome sequences discussed in this work have been deposited at DDBJ/ENA/GenBank and the Sequence Read Archive (SRA). The corresponding accession numbers are listed in Table 1.

ACKNOWLEDGMENTS

This research was supported in part by grants awarded to F.J.R. and F.O. by Enterprise Ireland (grant CF-2017-0757-P) and to F.O. by Enterprise Ireland (grant IP-2015-0390), the European Commission (grants FP7-PEOPLE-2013-ITN [project 607786], FP7-KBBE-2012-6 [project CP-TP-312184], FP7-KBBE-2012-6 [project 311975], OCEAN 2011-2 [project 287589], and EU2020-634486-2015), Science Foundation Ireland (grants 12/RC/2275 [SSPC-3], 12/RC/2275 [SSPC-2], 13/TIDA/B2625, 12/TIDA/B2411, 12/ TIDA/B2405, 14/TIDA/2438, 15/TIDA/2977, and SFI09/RFP/BMT2350), the Department of Agriculture and Food (grants FIRM 11/F009/MabS and FIRM 13/F/516), the Irish Research Council for Science, Engineering, and Technology (grant GOIPG/2014/647), the Health Research Board/Irish Thoracic Society (grants MRCG-2014-6 and MRCG-2018-16), The Health Research Board (grant ILP-POR-2019-004), the Department of the Marine (grant BEAU/BIOD/01), and the Cystic Fibrosis Foundation (USA) (grant OG1710).

REFERENCES

- Palleroni NJ. 1992. Present situation of the taxonomy of aerobic pseudomonads, p 105–115. *In* Galli E, Silver S, Witholt B (ed), *Pseudomonas*: molecular biology and biotechnology. ASM Press, Washington, DC.
- Silby MW, Winstanley C, Godfrey SA, Levy SB, Jackson RW. 2011. *Pseudomonas* genomes: diverse and adaptable. FEMS Microbiol Rev 35: 652–680. https://doi.org/10.1111/j.1574-6976.2011.00269.x.
- Poblete-Castro I, Becker J, Dohnt K, dos Santos VM, Wittmann C. 2012. Industrial biotechnology of *Pseudomonas putida* and related species. Appl Microbiol Biotechnol 93:2279–2290. https://doi.org/10.1007/ s00253-012-3928-0.
- Höfte M, De Vos P. 2007. Plant pathogenic *Pseudomonas* species, p 507–533. *In* Gnanamanickam SS (ed), Plant-associated bacteria. Springer, Dordrecht, Netherlands. https://doi.org/10.1007/978-1-4020-4538-7.
- Gellatly SL, Hancock R. 2013. Pseudomonas aeruginosa: new insights into pathogenesis and host defences. Pathog Dis 67:159–173. https://doi .org/10.1111/2049-632X.12033.
- Santoyo G, Orozco-Mosqueda MC, Govindappa M. 2012. Mechanisms of biocontrol and plant growth-promoting activity in soil bacterial species of *Bacillus* and *Pseudomonas*: a review. Biocontrol Sci Technol 22: 855–872. https://doi.org/10.1080/09583157.2012.694413.
- Alymanesh MR, Taheri P, Tarighi S. 2016. *Pseudomonas* as a frequent and important quorum quenching bacterium with biocontrol capability against many phytopathogens. Biocontrol Sci Technol 26:1719–1735. https://doi.org/10.1080/09583157.2016.1239065.

- Chong TM, Koh CL, Sam CK, Choo YM, Yin WF, Chan KG. 2012. Characterization of quorum sensing and quorum quenching soil bacteria isolated from Malaysian tropical montane forest. Sensors (Basel) 12: 4846–4859. https://doi.org/10.3390/s120404846.
- Gutiérrez-Barranquero JA, Reen FJ, Parages ML, McCarthy R, Dobson ADW, O'Gara F. 2019. Disruption of *N*-acyl-homoserine lactone-specific signalling and virulence in clinical pathogens by marine sponge bacteria. Microb Biotechnol 12:1049–1063. https://doi.org/10.1111/1751-7915 .12867.
- Gutiérrez-Barranquero JA, Parages ML, Dobson ADW, Reen FJ, O'Gara F. 2019. Genome sequence of *Paracoccus* sp. JM45, a bacterial strain isolated from a marine sponge with a dual quorum sensing inhibition activity. Microbiol Resour Announc 8:e01496-18. https://doi.org/10 .1128/MRA.01496-18.
- Tong C, Wu Z, Zhao X, Xue H. 2018. Arginine catabolic mobile elements in livestock-associated methicillin-resistant staphylococcal isolates from bovine mastitic milk in China. Front Microbiol 9:1031. https://doi.org/10 .3389/fmicb.2018.01031.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.

- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. https:// doi.org/10.1038/srep08365.
- Huang JJ, Petersen A, Whiteley M, Leadbetter JR. 2006. Identification of QuiP, the product of gene PA1032, as the second acyl-homoserine lactone acylase of *Pseudomonas aeruginosa* PAO1. Appl Environ Microbiol 72:1190–1197. https://doi.org/10.1128/AEM.72.2.1190-1197.2006.
- Sio CF, Otten LG, Cool RH, Diggle SP, Braun PG, Bos R, Daykin M, Camara M, Williams P, Quax WJ. 2006. Quorum quenching by an N-acyl-homoserine lactone acylase from *Pseudomonas aeruginosa* PAO1. Infect Immun 74:1673–1682. https://doi.org/10.1128/IAI.74.3 .1673-1682.2006.
- Blin K, Shaw S, Steinke K, Villebro R, Ziemert N, Lee SY, Medema MH, Weber T. 2019. antiSMASH 5.0: updates to the secondary metabolite genome mining pipeline. Nucleic Acids Res 47:W81–W87. https://doi .org/10.1093/nar/gkz310.

- Mercado-Blanco J, Olsson GM, Van der Drift PE, Thomas-Oates JE, Bakker LC, Van Loon P. 2001. Analysis of the *pmsCEAB* gene cluster involved in biosynthesis of salicylic acid and the siderophore pseudomonine in the biocontrol strain *Pseudomonas fluorescens* WCS374. J Bacteriol 183: 1909–1920. https://doi.org/10.1128/JB.183.6.1909-1920.2001.
- Chang CY, Krishnan T, Wang H, Chen Y, Yin WF, Chong YM, Tan LY, Chong TM, Chan KG. 2014. Non-antibiotic quorum sensing inhibitors acting against *N*-acyl homoserine lactone synthase as druggable target. Sci Rep 4:7245. https://doi.org/10.1038/srep07245.
- Richter M, Rosselló-Móra R, Glöckner FO, Peplies J. 2016. JSpeciesWS: a Web server for prokaryotic species circumscription based on pairwise genome comparison. Bioinformatics 32:929–931. https://doi .org/10.1093/bioinformatics/btv681.
- Garrido-Sanz D, Meier-Kolthoff JP, Göker M, Martín M, Rivilla R, Redondo-Nieto M. 2016. Genomic and genetic diversity within the *Pseudomonas fluorescens* complex. PLoS One 11:e0150183. https://doi.org/10.1371/ journal.pone.0150183.