

Accumulation of RNA on chromatin disrupts heterochromatic silencing

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Long noncoding RNAs (lncRNAs) play a conserved role in regulating gene expression, chromatin dynamics, and cell differentiation. They serve as a platform for RNA interference (RNAi)-mediated heterochromatin formation or DNA methylation in many eukaryotic organisms. We found in *Schizosaccharomyces pombe* that heterochromatin is lost at transcribed regions in the absence of RNA degradation. We show that heterochromatic RNAs are retained on chromatin, form DNA:RNA hybrids, and need to be degraded by the Ccr4-Not complex or RNAi to maintain heterochromatic silencing. The Ccr4-Not complex is localized to chromatin independently of H3K9me and degrades chromatin-associated transcripts, which is required for transcriptional silencing. Overexpression of heterochromatic RNA, but not euchromatic RNA, leads to chromatin localization and loss of silencing of a distant *ade6* reporter in wild-type cells. Our results demonstrate that chromatin-bound RNAs disrupt heterochromatin organization and need to be degraded in a process of heterochromatin formation.

[Supplemental material is available for this article.]

Regulation of genome expression is required for many cellular processes, including cell proliferation, differentiation, development, and viability. Heterochromatin, or silent chromatin, is essential to maintain genome stability and chromosome segregation. In fission yeast, heterochromatin is established by small RNA pathways that act at the chromatin level (Bond and Baulcombe 2014; Holoch and Moazed 2015). Small RNAs are loaded on Argonaute and interact with target RNAs through base-pairing interactions. The Argonaute-containing RNA-induced transcriptional silencing (RITS) complex is guided by the small RNAs to heterochromatic repeats. To amplify small RNAs, RITS recruits the RNA-dependent RNA Polymerase complex (RDRC), which synthesizes double-stranded RNA that is processed by Dicer into siRNAs. In addition, RITS recruits the H3K9 methyltransferase complex CLRC to chromatin, which leads to repressive histone 3 lysine 9 methylation (H3K9me), recruitment of HP1 proteins, and heterochromatin formation (Volpe et al. 2002; Verdel et al. 2004; Allshire and Ekwall 2015; Holoch and Moazed 2015; Martienssen and Moazed 2015; Zocco et al. 2016).

In fission yeast, constitutive heterochromatin is found at centromeres, subtelomeres, and the mating type (*mat*) locus (Allshire 1995; Cam et al. 2005). At centromeric repeats, RNAi is needed for the establishment and maintenance of heterochromatin. The establishment of centromeric heterochromatin is initiated by a class of single-stranded small RNAs called priRNAs (Halic and Moazed 2010), which are generated by Argonaute and the PARN nuclease Triman (Marasovic et al. 2013). In cells lacking components of the RNAi pathway, heterochromatin is not lost at the *mat* locus and subtelomeres. At those loci, heterochromatin maintenance is RNAi independent, but RNAi is required to establish heterochromatin (Kanoh et al. 2005; Hansen et al. 2006; Allshire and Ekwall 2015). At subtelomeres, the Shelterin complex binds telomeres and recruits the CLRC methyltransferase and the SHREC deacetylase complexes to maintain heterochromatin (Kanoh et al. 2005;

Hansen et al. 2006; Sugiyama et al. 2007; Motamedi et al. 2008; Tadeo et al. 2013; Wang et al. 2016). At the *mat* locus, the *cenH* element shares homology with the centromeric repeats and is essential for RNAi-mediated heterochromatin establishment (Hall et al. 2002). Once established, heterochromatin can be maintained in an RNAi-independent way by the ATF/CREB transcription factors Atf1/Pcr1 (Jia et al. 2004).

We found that the Ccr4-Not complex and RNAi are required for degradation of heterochromatic RNAs, and this is essential for constitutive heterochromatin formation. Ccr4-Not is a conserved complex involved in RNA surveillance, transcription elongation, RNA export, and DNA repair (Collart and Panasenko 2012; Miller and Reese 2012; Wahle and Winkler 2013; Inada and Makino 2014). The Ccr4-Not complex is the primary deadenylase and is essential for downstream decapping and mRNA decay by Xrn1 (*Schizosaccharomyces pombe*: Exo2), a conserved 5'-3' exoribonuclease (Collart and Panasenko 2012; Miller and Reese 2012). Caf1 and Ccr4 build the deadenylase module of Ccr4-Not, while Not1, Not2, Not4 (*S. pombe*: Mot2), and Not5 belong to the Not group of the complex. Caf1 bridges Ccr4 to Not1 and is essential for the deadenylation activity (Basquin et al. 2012).

In RNAi-mediated heterochromatin formation, heterochromatic transcripts serve as a template for siRNA generation, Argonaute targeting, and recruitment of the methyltransferase complex CLRC (Holoch and Moazed 2015). At the same time, heterochromatic transcripts are degraded by RNAi. A link between various RNA degradation machineries and H3K9me at meiotic genes was also recently reported (Hiriart et al. 2012; Zofall et al. 2012; Egan et al. 2014; Chalamcharla et al. 2015; Cotobal et al. 2015; Tucker et al. 2016). Several of those studies suggested a direct recruitment of the H3K9 methyltransferase by different RNA degradation machineries to deposit H3K9me. The aim of our work was

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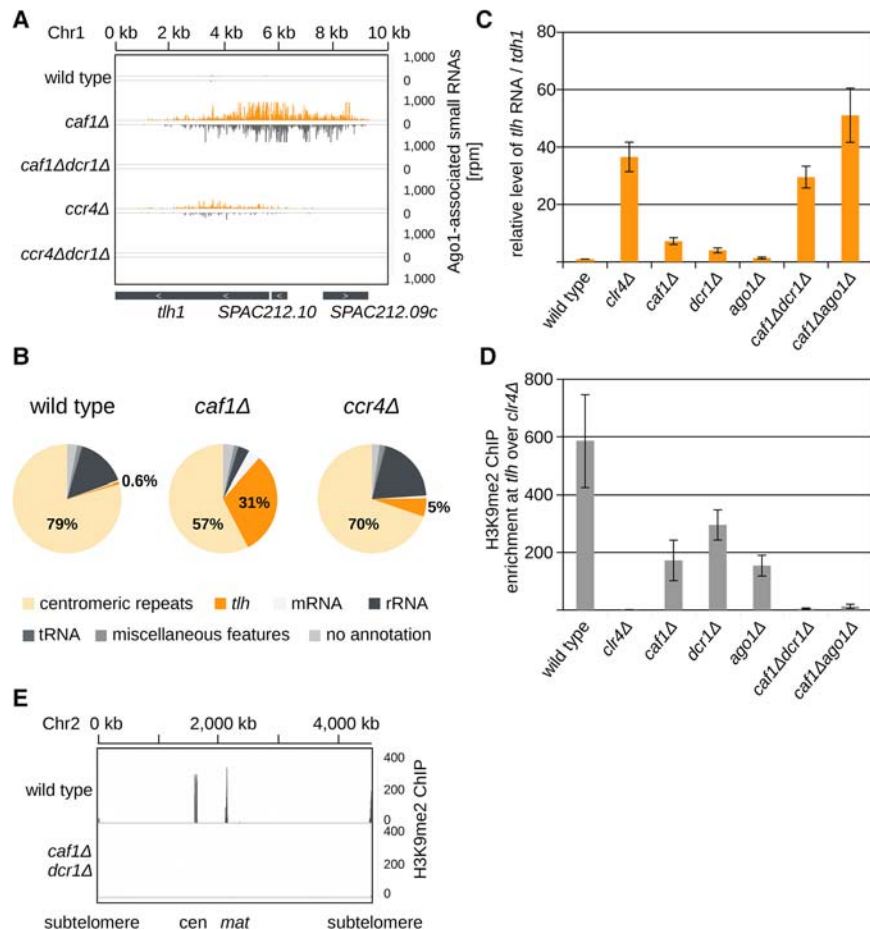


Figure 1. Caf1 and RNAi are required for heterochromatin formation at transcribed regions. (A) Endogenously tagged Argonaute-bound small RNA reads in the indicated cells were plotted over the subtelomeric region. The location of genes is indicated as gray boxes below the small RNA peaks. Reads from + and – strands are depicted in orange and gray, respectively. Scale bars on the right denote small RNA read numbers normalized per one million reads. *tlf* is present at subtelomeres of both arms of Chromosomes 1 and 2 (Mandell et al. 2005; Hansen et al. 2006). (B) Classification of Argonaute-bound small RNAs from wild-type, *caf1Δ*, and *ccr4Δ* cells. Pie charts illustrate percentages for the individual small RNA classes relative to the total number of reads for each strain. Argonaute-bound subtelomeric siRNAs are increased by more than 50-fold in *caf1Δ* cells compared with the wild type. (C) Quantification of subtelomeric *tlf* transcripts in indicated strains by RT-qPCR. In *caf1Δdcr1Δ* and *caf1Δago1Δ* cells, silencing of subtelomeric repeats is lost. Error bars, SE of more than seven independent experiments. For *caf1Δdcr1Δ* and *caf1Δago1Δ*, experiments of two independent colonies were averaged, respectively. Reverse transcription was performed with specific primers; the wild type was set to one. (D) ChIP experiment showing that H3K9me2 is lost at subtelomeric *tlf* repeats in *caf1Δdcr1Δ* cells. Error bars, SE of at least four independent experiments. For *caf1Δdcr1Δ* and *caf1Δago1Δ*, data of two independent colonies were averaged, respectively. *clr4Δ* was set to one. (E) ChIP-seq showing that H3K9me2 is lost at all constitutive heterochromatic loci in *caf1Δdcr1Δ* cells. Scale bars on the right denote read numbers per million reads normalized to the TAS region (Chr 2: 4534 kb–4538 kb) where H3K9me2 is not lost in the mutant.

to analyze if degradation of heterochromatic RNAs is required for functional heterochromatin organization.

Results

Caf1 and RNAi are required for heterochromatin formation

We sequenced Argonaute-bound small RNAs from cells having deletions of different RNA degradation pathways to determine if they might act redundantly with RNAi. In deletion of *caf1*, a deadenylase subunit of the Ccr4-Not complex, we observed a

high amount of small RNAs generated from subtelomeric repeats (Fig. 1A,B). In wild-type cells, <1% of Argonaute-bound small RNAs map to the subtelomeric region, indicating that RNAi is not the major contributor to silencing at the subtelomeric repeats (Fig. 1B). On the contrary, in *caf1Δ* cells >30% of Argonaute-bound small RNAs map to the subtelomeric repeats (Fig. 1B). These small RNAs are Dcr1 dependent and show all features of siRNAs (Fig. 1A; Supplemental Fig. 1A,B). Subtelomeric siRNAs are generated from *tlf1*, *SPAC212.10*, and *SPAC212.09c*, covering a region from 0 to 9 kb on the left arm of Chromosome 1 (Fig. 1A) and the homologous regions on both arms of Chromosomes 1 and 2.

Similar to the subtelomeric region, higher amounts of siRNAs were generated at the *mat* locus in *caf1Δ* cells (Supplemental Fig. 1C). Centromeric siRNAs were generated near wild-type levels at *dg* and *dh* repeats but were strongly reduced at the IRC3 element in *caf1Δ* cells (Supplemental Fig. 1C; Halic and Moazed 2010). A similar siRNA pattern was detected in deletion of *ccr4*, the second deadenylase of the Ccr4-Not complex; however, lower amounts of siRNAs were generated from the subtelomeric and *mat* regions in *ccr4Δ* cells (Fig. 1A,B; Supplemental Fig. 1C). We did not observe a defect in length of Argonaute-bound small RNAs in *caf1Δ* or *ccr4Δ* cells, indicating that Caf1 and Ccr4, in contrast to Triman, are not directly processing small RNAs (Supplemental Fig. S1A,B; Marasovic et al. 2013). Our data suggest that the Caf1 and Ccr4 nucleases degrade subtelomeric, centromeric, and *mat* locus transcripts, and in their absence, RNAi acts as a redundant degradation mechanism.

We observed that centromeric *dg* and subtelomeric *tlf* transcripts were four- to fivefold up-regulated in *caf1Δ* cells, and silencing of a centromeric *ade6* reporter was reduced (Fig. 1C; Supplemental Fig. 1D,E). These data

show a partial loss of heterochromatic silencing at the subtelomeric and centromeric repeats in *caf1Δ* cells. Next, we generated *caf1Δdcr1Δ* and *caf1Δago1Δ* double mutants in several genetic backgrounds to remove both degradation pathways, RNAi and Ccr4-Not. While single deletions of *caf1* and *ago1/dcr1* had a small impact on RNA levels in the subtelomeric region, deletion of both pathways completely de-repressed subtelomeric transcripts (*tlf1*, *SPAC212.10* and *SPAC212.09c*) to the level of deletion of the H3K9 methyltransferase Clr4 (Fig. 1C; Supplemental Fig. S1F,G). We performed chromatin immunoprecipitation (ChIP) with an antibody against H3K9me2 and observed loss of H3K9me2 at the

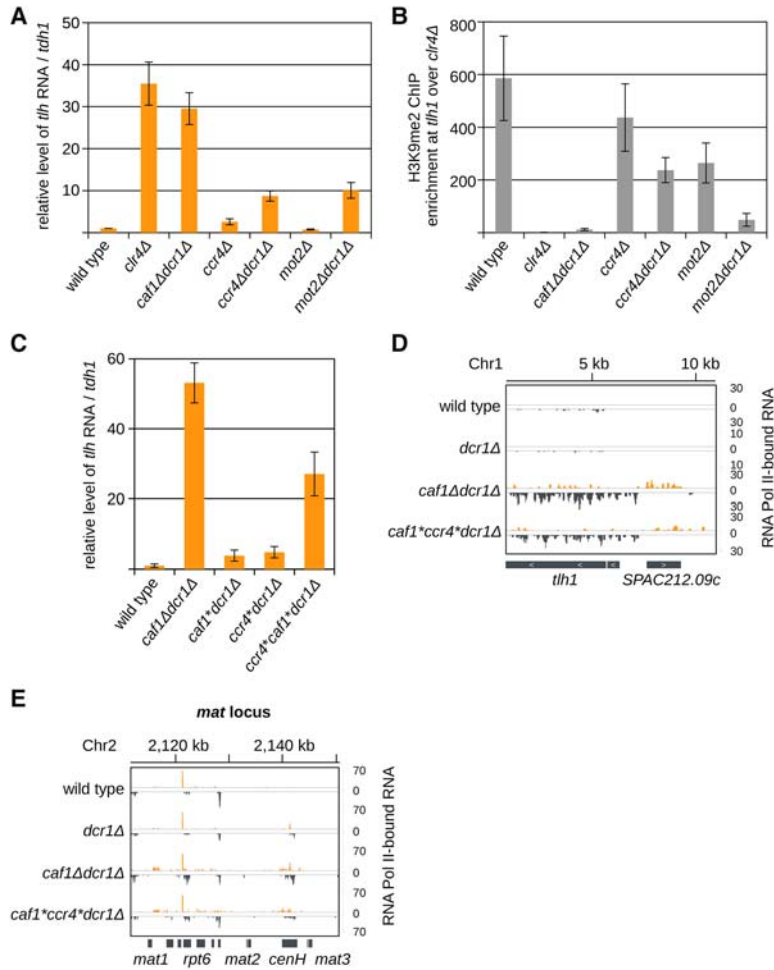


Figure 2. Caf1 and Ccr4 nuclease activity is required for heterochromatic silencing. (A) Quantification of subtelomeric *thl* transcripts in indicated strains by RT-qPCR. In *ccr4Δdcr1Δ* and *mot2Δdcr1Δ* cells, silencing of subtelomeric repeats is defective, but not as much as in *caf1Δdcr1Δ* or *clr4Δ* cells. Error bars, SE of three or more independent experiments. Reverse transcription was performed with specific primers; the wild type was set to one. (B) ChIP experiment showing that H3K9me2 is lost at subtelomeric *thl* repeats in *mot2Δdcr1Δ* cells but not in *ccr4Δdcr1Δ*. Error bars, SE of two (*mot2Δ* and *mot2Δdcr1Δ*) or more independent experiments. *clr4Δ* was set to one. (C) Quantification of subtelomeric *thl* transcripts by RT-qPCR in the wild type or *caf1Δdcr1Δ* controls or in *dcr1Δ* strains expressing a Caf1* (Caf1D53AD243AD174A) or/ and a Ccr4* (Ccr4H665A) activity mutant. Expression of Caf1* or Ccr4* silences *thl*, but when both deadenylases are mutated, silencing of *thl* is lost. Error bars, SE of three independent experiments. Reverse transcription was performed with specific primers; the wild type was set to one. (D,E) Sequencing reads of RNA Polymerase II (Pol II)-associated RNA in indicated cells are plotted over the subtelomeric *thl* region (D) and over the *mat* locus (E). In *caf1*ccr4*dcr1Δ* cells, transcription at *etl1* and *SPAC212.09c* and at the *mat* locus is increased compared with wild-type cells. Reads from + and - strands are depicted in orange and gray, respectively. Scale bars on the right denote RNA read numbers normalized to the total number of reads mapping to protein coding genes.

subtelomeric, centromeric, and *mat* locus in *caf1Δdcr1Δ* and *caf1Δago1Δ* cells (Fig. 1D,E; Supplemental Fig. S2A,B). These results show that H3K9me2 and heterochromatic silencing cannot be maintained in these mutants at all constitutive heterochromatic loci.

Caf1 and Ccr4 activity is required for heterochromatin assembly at the *thl* region

Next, we analyzed whether other subunits of the Ccr4-Not complex are required for subtelomeric silencing. In the double mutant of the

second deadenylase Ccr4 and RNAi (*ccr4Δdcr1Δ*), subtelomeric transcripts were also accumulating, but H3K9me2 was not completely lost at the subtelomeric *thl* region (Fig. 2A,B). *not2Δdcr1Δ*, *rcd1Δdcr1Δ* (*RQCD1*, *Caf40*), and *caf16Δdcr1Δ* cells showed no or little effect on *thl* RNA levels; however, in *mot2Δdcr1Δ* cells, *thl* transcripts were depressed (Fig. 2A; Supplemental Fig. S2C). H3K9me2 was also reduced at subtelomeric *thl* and centromeric *dg* repeats in *mot2Δdcr1Δ* cells (Fig. 2B; Supplemental Fig. 2D). Our data indicate that the Ccr4-Not complex subunits Caf1, Ccr4, and also the E3 ubiquitin ligase Mot2 are involved in silencing and heterochromatin formation at constitutive heterochromatic loci. At the most distal region (toward centromeres), subtelomeric H3K9me2 was also lost in *caf1Δ* (Supplemental Fig. S2E), *ccr4Δ*, and *mot2Δ* cells (Cotobal et al. 2015).

Of all subunits of the Ccr4-Not complex, deletion of the deadenylase Caf1 had the greatest loss of silencing of heterochromatic transcripts. We investigated if the nuclease activity of Caf1 was required for *thl* silencing and heterochromatin assembly. We genomically integrated Caf1* (Caf1D53AD243AD174A) and Ccr4* (H665A) activity mutants into *caf1Δdcr1Δ* and *dcr1Δ* cells (Chen et al. 2002; Jonstrup et al. 2007). Introduction of the Caf1* or Ccr4* activity mutants showed only a minor increase in *thl* RNA in *caf1*dcr1Δ* and *ccr4*dcr1Δ* cells (Fig. 2C), suggesting that both deadenylases, Caf1 and Ccr4, might act redundantly. Since Caf1 is required for Ccr4 recruitment to the Ccr4-Not complex, deletion of Caf1 eliminates the activity of both deadenylases (Basquin et al. 2012). Mutation of the active site of both Caf1 and Ccr4 resulted in a strong accumulation of subtelomeric transcripts and reduction in H3K9me2 in *caf1*ccr4*dcr1Δ* cells (Fig. 2C; Supplemental Fig. S3A). Sequencing of nascent RNA associated with RNA Polymerase II (Pol II) showed increased transcription in *caf1*ccr4*dcr1Δ* cells at subtelomeric, *mat*, and centromeric loci, similar to *caf1Δdcr1Δ* cells (Fig. 2D,E; Supplemental Fig. S3B). These results show that deadenylation by Caf1 and Ccr4 is required for transcriptional silencing at all heterochromatic loci. We note that *caf1*ccr4*dcr1Δ* cells have a slightly weaker defect compared with *caf1Δdcr1Δ* cells, suggesting that Caf1 might recruit an additional factor contributing to RNA degradation or heterochromatin formation.

This suggested that other nucleases and RNA processing factors could also be involved in degradation of heterochromatic RNAs. In addition to *caf1Δ*, deletion of the 5'-3' exonuclease

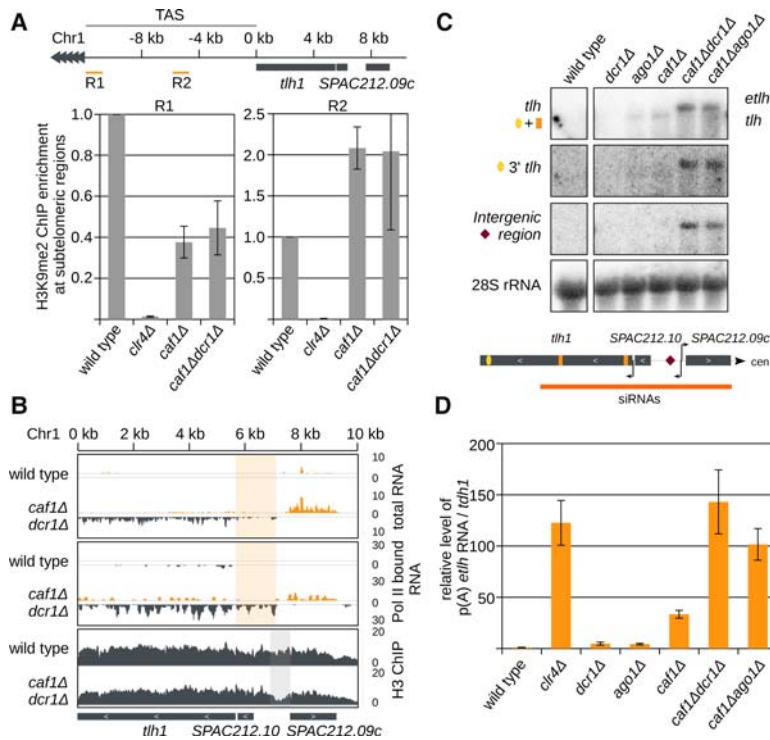


Figure 3. In *caf1Δdcr1Δ* cells, transcriptional silencing is lost. (A) ChIP experiment showing that in *caf1Δdcr1Δ* cells H3K9me2 is not lost at regions between telomeric repeats and the transcribed *tlh* gene. The *S. pombe* genome assembly is incomplete in the subtelomeric region, but additional insert clones from the telomere plasmid library with the sequence of the telomere-associated region (TAS) are available at www.pombase.com. Error bars, SE of three independent experiments. Region 1 (R1) is next to the telomere, and region 2 (R2) is ~6 kb away from telomeric repeats. *tlh* is ~10 kb upstream of the telomeric repeats. (B) Sequencing reads in indicated cells are plotted over the subtelomeric region. (Top) In *caf1Δdcr1Δ* cells, silencing in the subtelomeric region is lost. Reads between *SPAC212.10* and *SPAC212.09c* can be detected. Reads from + and – strands are depicted in orange and gray, respectively. Scale bars on the right denote total RNA read numbers normalized to all reads mapping to protein coding genes. Highlighted in orange are reads that map upstream of *tlh1*. (Middle) RNA Pol II-associated RNA. Pol II starts transcription in *caf1Δdcr1Δ* cells upstream of the annotated *tlh* gene; in wild-type cells, it starts at the annotated transcription start. Reads from + and – strands are depicted in orange and gray, respectively. Scale bars on the right denote RNA read numbers normalized to total number of reads mapping to protein coding genes. Highlighted in orange are reads that map upstream of *tlh1*. (Bottom) H3 ChIP sequencing, showing a nucleosome-free region (highlighted in gray) upstream of the Pol II-associated RNA *tlh* reads. Scale bars on the right denote reads per million. (C) Northern blot showing accumulation of two distinct *tlh* products. (*tlh*) three probes mapping to the annotated *tlh* sequence; (*3' tlh*) one probe mapping to the 3' end of *tlh*; (Intergenic region) one probe hybridizing between *SPAC212.10* and *SPAC212.09c*; and (28S rRNA) 28S rRNA probe as loading control. In *caf1Δdcr1Δ* and *caf1Δago1Δ* cells, a longer transcript (*ethl*) is accumulating. (D) Quantification of subtelomeric *ethl* transcripts in indicated strains by RT-qPCR reverse transcribed with oligo(dT). In *caf1Δ* cells, *ethl* silencing is strongly reduced. In *caf1Δdcr1Δ* and *caf1Δago1Δ* cells, silencing of the *ethl* transcript is lost. Error bars, SE of more than three independent experiments. Reverse transcription was performed with oligo(dT) primer, and qRT-PCR was performed with primers for *ethl*. The wild type was set to one.

Exo2 (Xrn1) showed strong accumulation of subtelomeric siRNAs (Supplemental Fig. S3C). Subtelomeric siRNAs were also increased in the deletion of the RNA processing factor *mlo3* (Zhang et al. 2011). In the deletion of the exosome subunit *rrp6*, the TRAMP component *cid14*, the PAF complex subunit *leo1* (Sadeghi et al. 2015), HP1 protein *swi6*, and in the *dis3-53* mutant, we observed only a smaller accumulation of subtelomeric siRNAs (Supplemental Fig. S3C,D). There was little or no effect on siRNA generation at centromeric repeats in these mutants (Supplemental Fig. S3C). Our results indicate that the Ccr4-Not/Xrn1 pathway eliminates heterochromatic transcripts, and this is required for heterochromatic silencing.

Heterochromatin is lost at transcribed regions

The Shelterin complex recruits CLRC and SHREC to the telomeres, from where they spread to maintain subtelomeric heterochromatin (Kano et al. 2005; Sugiyama et al. 2007; Wang et al. 2016). In *caf1Δdcr1Δ* cells, Taz1, a DNA binding protein of the Shelterin complex, is still recruited to the telomeric repeats (Supplemental Fig. S4A). This indicates that the Shelterin complex could still recruit CLRC to maintain heterochromatin. In *taz1Δ* cells, we observe a small increase in subtelomeric siRNAs, and consistent with previous studies, in *taz1Δdcr1Δ* cells when both CLRC recruitment pathways are eliminated, *tlh* transcripts accumulate (Supplemental Fig. S4B,C,E; Kano et al. 2005; Hansen et al. 2006). In *caf1Δtaz1Δ* cells, we did not observe an additional loss of H3K9me2 at *tlh* compared with *caf1Δ* (Supplemental Fig. S4D).

Consistent with Taz1 localization (Supplemental Fig. S4A), H3K9me2 was not lost at the telomere-associated sequence (TAS) between telomeric repeats and *tlh* in *caf1Δdcr1Δ* cells (Fig. 3A). This indicates that the Shelterin complex can still deposit H3K9me2 at the telomeric borders, which can spread until the *tlh* locus. Heterochromatin, however, cannot spread over the transcribed *tlh* gene in *caf1Δdcr1Δ* cells (Supplemental Fig. S2E). These data show that Caf1 and RNAi are not essential for the maintenance and spreading of H3K9me from telomeric repeats until the *tlh* region.

Absence of RNA degradation leads to changes in chromatin organization

Small RNA sequencing data in *caf1Δ* cells show that siRNAs are generated from *tlh*, *SPAC212.10*, *SPAC212.09c*, and their intergenic regions (Fig. 1A). Sequencing of RNA Pol II-associated nascent RNA and also total RNA showed increased transcriptional activity in the intergenic region in *caf1Δdcr1Δ* cells but not in wild-type cells (Fig. 3B). This is consistent with the H3 ChIP-seq and nucleosome positioning data that show a nucleosome-free region between *SPAC212.10* and *SPAC212.09c* in *caf1Δdcr1Δ* (Fig. 3B) and *clr4Δ* cells (Garcia et al. 2010). Northern blot analysis confirms that *tlh* RNA is longer in *caf1Δdcr1Δ* and *caf1Δago1Δ* cells than in *ago1Δ* or *caf1Δ* cells (Fig. 3C). *tlh* probes from the 3' end as well as from the 5' intergenic region hybridized to the longer transcript, indicating that in *caf1Δdcr1Δ* cells, transcripts start 2 kb upstream of the annotated *tlh* promoter and terminate at the *tlh* terminator (Fig. 3C). The extended *tlh* (*ethl*) transcript is polyadenylated and shows increased amounts in *caf1Δ*, but not in *ago1Δ* cells, and is completely de-

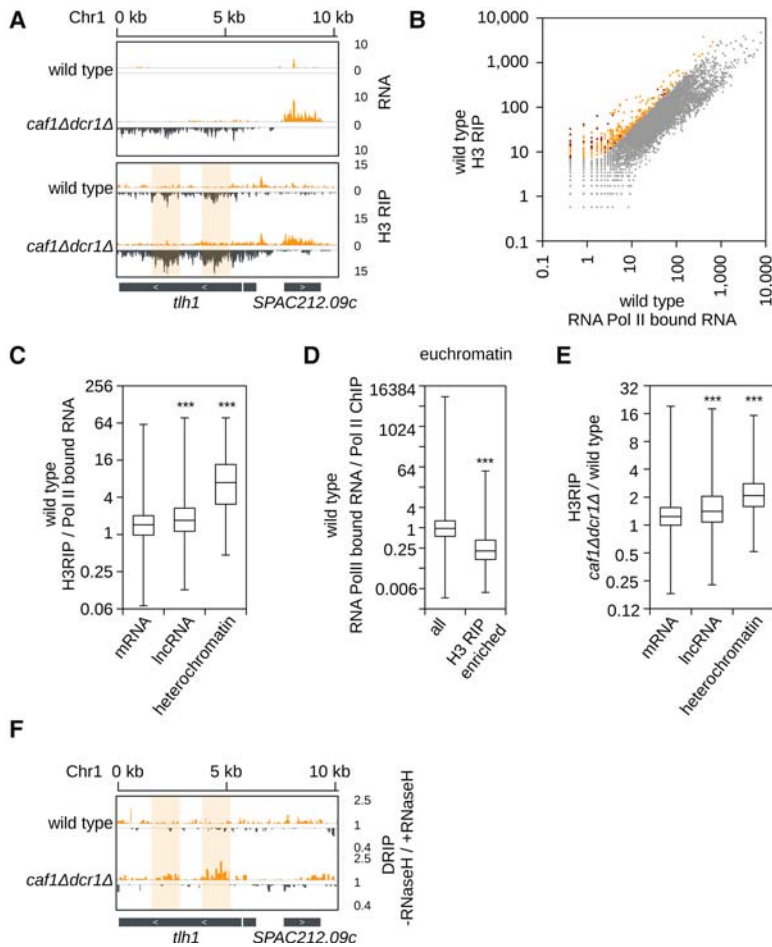


Figure 4. Heterochromatic transcripts accumulate on chromatin. (A) H3-RIP sequencing reads in indicated cells are plotted over the subtelomeric region. Total RNA sequencing reads on *top* as control. Reads from + and – strands are depicted in orange and gray, respectively. Scale bars on the *right* denote RNA read numbers per total number of reads mapping to protein coding genes. Compared with total RNA, *tlh* is enriched at the chromatin in wild-type and *caf1Δdcr1Δ* cells. Region where DNA:RNA hybrids are enriched is highlighted. Scale bars on the *right* denote RNA read numbers normalized to total number of reads mapping to protein coding genes. (B) Scatter plot showing H3-associated RNA relative to RNA Pol II-bound nascent RNA. A subset of genes is more enriched in H3 RIP than bound to RNA Pol II, indicating post-transcriptional retention on chromatin. Lowly expressed genes show stronger enrichment on chromatin. RNAs enriched on chromatin are shown in orange or red for heterochromatic RNA. (C) Box plot analysis of H3-associated RNA relative to nascent RNA bound to RNA Pol II in wild-type cells. (lncRNA) Euchromatic noncoding RNA with annotation SPNCRNA, $n = 1354$; (mRNA) all protein coding genes without genes in heterochromatic areas, $n = 5014$; (heterochromatic) genes which are located in constitutive heterochromatin areas, $n = 62$. Relative to their transcript levels, ncRNAs and especially heterochromatic RNA are more likely to be bound to chromatin than mRNA. (***) $P < 0.001$. (D) Box plot analysis of nascent RNA bound to RNA Pol II relative to RNA Pol II CHIP in wild-type cells. Genes that have RNA enriched on chromatin show reduced transcription compared with the quantity of RNA Pol II on the chromatin. (all) All mRNA and lncRNA, $n = 6345$; (chromatin enriched) euchromatic genes (mRNA and lncRNA) with greater than fourfold enrichment in H3-RIP over nascent RNA, $n = 258$. (***) $P < 0.001$. (E) Box plot analysis of H3-associated RNA in *caf1Δdcr1Δ* cells relative to wild-type cells. In *caf1Δdcr1Δ* cells, heterochromatic RNA and lncRNA are even more associated with chromatin than in wild-type cells. (mRNA) $n = 5058$; (lncRNA) $n = 1467$; (heterochromatin) $n = 90$. (***) $P < 0.001$. (F) DNA:RNA hybrid sequencing in wild-type and *caf1Δdcr1Δ* cells plotted over the subtelomeric region. DNA:RNA hybrids are formed with same sequences, which are also enriched in H3-RIP (highlighted). Scale bars on the *right* denote the ratio of “sample not treated” over “sample treated” with RNase H (–RNase H/+RNase H) before immunoprecipitation.

transcript and that, in the absence of RNA degradation, subtelomeric chromatin organization is changed.

Heterochromatic RNAs accumulate on chromatin and form DNA:RNA hybrids

Our data show that RNA degradation is required for heterochromatic silencing. This suggests that heterochromatic transcripts might accumulate on chromatin. We performed chromatin fractionation and also sequenced histone H3-bound RNA and observed that *tlh* RNA is enriched at chromatin in wild-type cells (Fig. 4A; Supplemental Fig. S5A,B). In *caf1Δ* and even more in *caf1Δdcr1Δ* cells, we observed increased levels of *tlh* transcripts in the chromatin fraction compared with wild-type cells (Fig. 4A; Supplemental Fig. S5A,B). Genome-wide comparison of histone H3-bound RNA and RNA Pol II-bound nascent RNA revealed that in wild-type cells, heterochromatic transcripts and lncRNA are more retained at chromatin than mRNAs relative to their transcription (Fig. 4B,C; Supplemental Figs. S5C, S6A). We found that transcripts from all constitutive heterochromatic loci accumulate on chromatin (Supplemental Fig. S6B). Our data suggest that heterochromatic transcripts and euchromatic lncRNA are processed less efficiently and are retained on chromatin post-transcriptionally.

In euchromatin we observed that genes with chromatin-retained RNAs were less efficiently transcribed by RNA Pol II (Fig. 4D). The same amount of RNA Pol II synthesized less nascent RNAs at these genes than at genes with no chromatin-bound RNAs (Fig. 4D; Supplemental Fig. S6C). These data show that in wild-type cells, chromatin-bound RNAs reduce transcription at euchromatic genes. In *caf1Δdcr1Δ* cells, heterochromatic transcripts from all loci and euchromatic lncRNA associate even more with chromatin than in wild-type cells, suggesting that the Ccr4-Not complex is degrading these transcripts on chromatin (Fig. 4E; Supplemental Fig. S6D,E).

Consistent with a previous observation (Nakama et al. 2012), we found low levels of DNA:RNA hybrids at subtelomeres in wild-type cells (Fig. 4F). In *caf1Δdcr1Δ* cells, heterochromatin is lost, and heterochromatic transcripts show a higher amount of DNA:RNA hybrids than in wild-type cells (Fig. 4F). Our data indicate that in wild-type cells, RNAi and Ccr4-Not degrade heterochromatic RNAs on chromatin

repressed in *caf1Δdcr1Δ* and *caf1Δago1Δ* cells (Fig. 3D). The *etlh* transcript was also accumulating in *chr4Δ* and at lower levels in *ccr4Δdcr1Δ* cells (Fig. 3D; Supplemental Fig. S4F). These data indicate that small RNAs in *caf1Δ* cells are generated from one longer

caf1Δdcr1Δ cells, heterochromatin is lost, and heterochromatic transcripts show a higher amount of DNA:RNA hybrids than in wild-type cells (Fig. 4F). Our data indicate that in wild-type cells, RNAi and Ccr4-Not degrade heterochromatic RNAs on chromatin

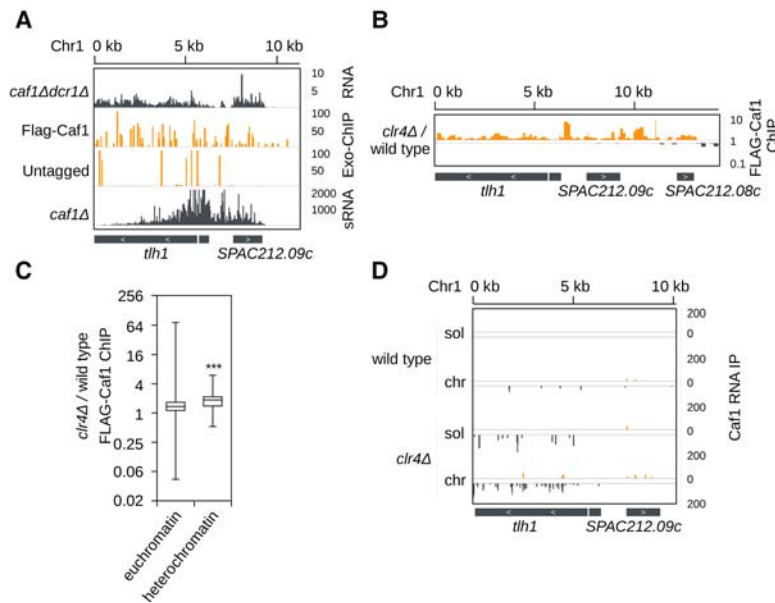


Figure 5. Heterochromatic transcripts are degraded on chromatin by the Ccr4-Not complex. (A) ChIP-exo (orange; middle) sequencing of endogenously tagged Flag-Caf1 in wild-type cells and untagged cells showing that Caf1 is enriched at subtelomeric *tlh* and *SPAC212.09c* genes. Caf1 localization overlaps with transcription (top; gray) and siRNA generation (bottom; gray). Scale bars on the right denote RNA read numbers normalized to total number of reads mapping to protein coding genes (RNA) or reads per million (Exo-ChIP and sRNA). (B) ChIP sequencing of endogenously tagged Flag-Caf1 in *clr4Δ* cells over wild-type cells. Caf1 is enriched at chromatin when *clr4* is deleted. Scale bars on the right denote fold change of Flag-Caf1 ChIP in *clr4Δ* over wild-type cells. (C) Box plots of Flag-Caf1 ChIP showing that in *clr4Δ* cells, Caf1 is more localized to heterochromatic loci than in wild-type cells. (***) $P < 0.001$. (D) Sequencing of Flag-Caf1-bound RNA from soluble (sol) and chromatin (chr) fractions. *tlh* RNA associates more with Caf1 from the chromatin than the soluble fraction in both wild-type and in *clr4Δ* cells. Reads from + and – strands are depicted in orange and gray, respectively. Scale bars on the right denote RNA read numbers normalized to total number of reads mapping to protein coding genes.

to prevent formation of DNA:RNA hybrids. These results also show that DNA:RNA hybrid formation is not heterochromatin dependent.

DNA:RNA hybrid formation was shown to induce H3S10 phosphorylation (H3S10P) (Castellano-Pozo et al. 2013), a chromatin mark that reduces HP1 binding to H3K9 methylated nucleosomes and interferes with heterochromatin formation and silencing (Fischle et al. 2005; Kloc et al. 2008). We determined H3S10P levels; however, we observed reduction of H3S10P in *caf1Δdcr1Δ* cells at *tlh* (Supplemental Fig. 6F). Consistent with our findings, an H3S10A mutant was previously shown to have reduced H3K9me2, indicating rather a codependence of these two histone marks (Kloc et al. 2008).

Caf1 eliminates heterochromatic RNA on chromatin

Our results show that heterochromatic transcripts are chromatin bound and suggest that Caf1 degrades them on chromatin. By using ChIP and ChIP-exo sequencing, we found that Caf1 is localized at the *tlh* and *SPAC212.09c* region in wild-type cells (Fig. 5A). This resembles the localization of Ccr4 and Not subunits that were found over open reading frames in *Saccharomyces cerevisiae* (Kruk et al. 2011; Venters et al. 2011). Caf1 localization on the chromatin was weak but detectable, indicating that Caf1 is not tightly bound to the chromatin. In *clr4Δ* cells, where heterochromatin is lost, even higher amounts of Caf1 were bound to the heterochromatic regions (Fig. 5B,C). In wild-type cells, Caf1 is bound to the chroma-

tin in the same region where siRNAs were generated in *caf1Δ* cells (Fig. 5A; Supplemental Fig. S7A); however in *clr4Δ* cells, Caf1 was enriched over the larger subtelomeric region (Fig. 5B; Supplemental Fig. S7B). Our data show that the Ccr4-Not complex is associated with chromatin; however, the complex is not recruited by heterochromatin. This suggests that RNAs, which are more bound to chromatin in *clr4Δ* cells, recruit Caf1 to chromatin.

We also performed Caf1 RNA-IP from chromatin and soluble fractions. In wild-type cells, chromatin-bound Caf1 associates with more *tlh* RNA than soluble Caf1 (Fig. 5D). This suggests that Caf1 degrades *tlh* RNA on chromatin. Other heterochromatic transcripts from subtelomeric and *mat* regions are degraded by Caf1 on chromatin as well (Supplemental Fig. S7C). At centromeric repeats we found that Caf1 does not degrade chromatin-associated RNAs in wild-type cells, which is consistent with RNAi being the primary degradation machinery at this locus (Supplemental Fig. S7C). In *clr4Δ* cells, chromatin-bound Caf1 associates with higher amounts of heterochromatic transcripts than soluble Caf1 (Fig. 5D; Supplemental Fig. S7C). This is consistent with increased Caf1 localization to heterochromatic DNA in *clr4Δ* cells and shows that Caf1 degrades heterochromatic RNA on chromatin

(Fig. 5B–D). We also found that Caf1 binds Caf1 mRNA and, in this way, might regulate its own activity (Supplemental Fig. S7D). Our data suggest that chromatin-bound RNAs are degraded by Ccr4-Not on chromatin, while mRNAs that are exported are degraded in cytosol.

Expression of heterochromatic *tlh* RNA leads to loss of silencing

Our data suggest that chromatin-bound RNAs negatively regulate heterochromatin assembly. To determine if chromatin-bound RNAs cause the loss of heterochromatin and to exclude secondary effects of mutant backgrounds, we generated wild-type strains expressing the heterochromatic *tlh* transcript under the thiamine-repressible *nmt1* promoter at a locus 5.5 kb upstream of *tlh* (Fig. 6A; Supplemental Fig. S8A). We split the *tlh* gene into two halves (*5'tlh* and *3'tlh*) and inserted both halves into the genome at the place of *SPNCRNA.70*. As a control, we generated the same construct expressing the euchromatic *LEU2* RNA.

We inserted an *ade6* reporter gene 5 kb upstream of the *tlh/LEU2*-expressing constructs (Fig. 6A). When grown on low adenine medium, cells that silence the *ade6* reporter gene are red, and cells that express *ade6* are white. When grown on *nmt1*-repressive low adenine medium (YE), the cells expressing the *LEU2* gene had mainly red colonies, indicating that the *ade6* reporter is silenced (Fig. 6B,C; Supplemental Fig. S8B). Expression of both *5'tlh* and *3'tlh* constructs showed a higher percentage of white colonies

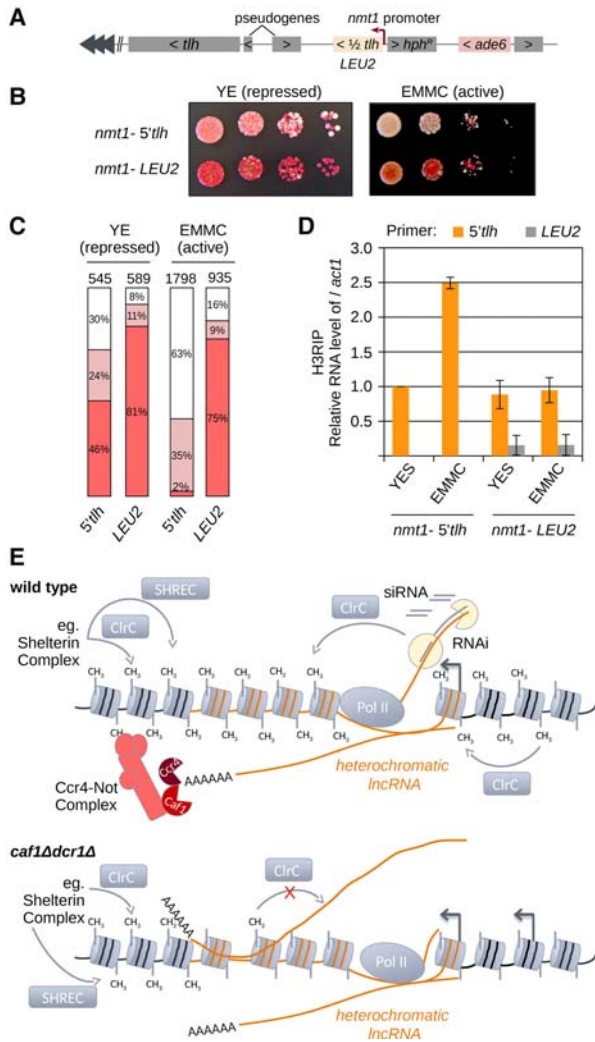


Figure 6. Accumulation of RNA on chromatin disrupts heterochromatin. (A) Scheme showing the constructs used. One half of *tlh* (5'*tlh*: first half; 3'*tlh*: second half) or *LEU2* were inserted ~5.5 kb upstream of *tlh* under the thiamine-inducible *nmt1* promoter. The *ade6* reporter gene was inserted 5 kb upstream of the inducible expression system. Cells that silence *ade6* are red, while *ade6*-expressing cells are white when grown on low adenine plates. Heterochromatin maintenance of a silenced, red colony was analyzed for each strain on YE (low adenine, thiamine +) or EMMC low ade (10 mg/L adenine, thiamine -) plates. (B) Growth assay with *ade6* reporter gene showing *nmt1-5'tlh* and *nmt1-LEU2* cells. *nmt1-5'tlh* cells show strong loss of heterochromatic silencing when grown under inducible conditions. Expression of a control *LEU2* gene did not lead to increased loss of heterochromatin when expression was induced. Cells were plated in 10-fold dilutions starting with 10^5 cells. (C) Quantification of different colored cells is shown for the indicated strains and expression, respectively. The total number of colonies counted is indicated above the bars. (D) Quantification of chromatin-associated RNA in indicated strains and conditions by H3 RIP RT-qPCR. In *nmt1-5'tlh* strains, 5'*tlh* RNA is enriched at chromatin in EMMC (induced) compared with YES medium (repressed). In *nmt1-LEU2* strains, neither *tlh* nor *LEU2* RNA is changed upon induction of *LEU2* expression. (E) RNAi or Shelterin complex recruit the CLRC methyltransferase and SHREC deacetylase complexes to establish heterochromatin in wild-type cells. CLRC and SHREC spread into repeats to establish heterochromatin. The Ccr4-Not complex eliminates heterochromatic RNA and promotes spreading over the transcribed region. In the absence of the Ccr4-Not complex and RNAi, heterochromatic transcripts accumulate on the chromatin. This leads to loss of heterochromatin. At the transcribed loci, heterochromatin is lost in *caf1Δdcr1Δ* cells, indicating that at transcribed regions, RNA degradation is required for heterochromatin assembly.

even under promoter-repressive conditions (YE) (Fig. 6B,C; Supplemental Fig. S8B).

When plated on EMMC low adenine media, the *nmt1* promoter is activated and RNA is transcribed at much higher levels with all constructs being expressed at similar levels (Supplemental Fig. S8C,D). Both 5'*tlh*- and 3'*tlh*-expressing cells had mainly white colonies on EMMC media, showing strong loss of heterochromatin (Fig. 6B,C; Supplemental Fig. S8E,F). On the contrary, the *LEU2*-expressing control cells were red, which indicates no loss of silencing on EMMC media (Fig. 6B,C). These data also show that the reporter gene is distant enough from the *tlh*/*LEU2*-expressing locus, and it is not affected by transcription from that locus.

In both *nmt1-5'tlh*- and *nmt1-3'tlh*-expressing cells, we found an increase in chromatin retention of *tlh* RNA after induction of *tlh* expression (Fig. 6D; Supplemental Fig. S8G). Although expressed at the same level, *LEU2* RNA shows very little enrichment on chromatin (Fig. 6D; Supplemental Fig. S8C). In the *nmt1-LEU2* strain, we also do not observe a change in chromatin retention of endogenous *tlh* RNA (Fig. 6D). Our data show that only chromatin-bound *tlh* RNAs lead to loss of heterochromatin, while released *LEU2* RNA did not have an effect on heterochromatic silencing. These results confirm that high amounts of chromatin-bound heterochromatic transcripts lead to loss of heterochromatin even in wild-type cells. This also excludes the possibility that chromatin association of heterochromatic RNAs is merely a consequence of increased transcription. Our data show that accumulation of RNA on chromatin leads to loss of heterochromatin and that degradation of chromatin-bound RNAs is essential for heterochromatin organization (Fig. 6E).

Discussion

Although RNA is required for heterochromatin formation, our data show that RNA needs to be degraded and chromatin-bound RNAs disrupt heterochromatin organization. We found that the Ccr4-Not complex eliminates heterochromatic transcripts on chromatin to maintain heterochromatin structure. The Ccr4-Not complex was initially described as a chromatin-associated complex involved in transcription (Miller and Reese 2012); however, its role on chromatin remained unclear. In fission yeast, the Ccr4-Not complex was recently shown also to be involved in deposition of H3K9 methylation at rDNA and meiotic genes (Cotobal et al. 2015; Sugiyama et al. 2016). Additionally, the Ccr4-Not complex was shown to play a role in silencing of subtelomeric transposons in *Caenorhabditis elegans* and *Drosophila melanogaster* (Fischer et al. 2013; Morgunova et al. 2015). Transposons are targeted by piRNAs in animal germline cells that establish heterochromatin in a similar way to siRNAs in fission yeast (Hirakata and Siomi 2016). The observations in *C. elegans* and *D. melanogaster* are analogous to our findings in *S. pombe* and imply a conserved role of the Ccr4-Not complex in degradation of chromatin-associated heterochromatic transcripts.

Our data suggest that degradation of heterochromatic transcripts by the Ccr4-Not complex and RNAi reduces the chance of DNA:RNA hybrid formation. It has been shown that chromatin retention of RNA and DNA:RNA hybrid formation establish RNAi-mediated heterochromatin (Sun et al. 2013; Skourti-Stathaki et al. 2014; Yu et al. 2014). Deletion of RNase H, however, has a defect in heterochromatin maintenance, indicating that an excess of DNA:RNA hybrids has a negative impact on heterochromatin assembly (Nakama et al. 2012). DNA:RNA hybrids were

also shown to inhibit nucleosome formation (Dunn and Griffith 1980), which would interfere with chromatin organization. These data show that an increase in DNA:RNA hybrids will interfere with chromatin organization and heterochromatin assembly.

We show that the deadenylation activity of the Ccr4-Not complex is important for heterochromatin maintenance, indicating that degradation of polyadenylated RNAs occurs on chromatin post-transcriptionally. We observed, however, a slightly weaker defect in the activity mutants than in the knock-out strains, suggesting that the activity mutant can recruit other RNA processing enzymes, like Exo2 (Xrn1) (Collart and Panasenko 2012; Miller and Reese 2012). Another possibility is that other functions of the Ccr4-Not complex are impaired in the deletion mutants. Recently, the Ccr4-Not complex was shown to act as a transcription elongation factor suggested to reactivate arrested RNA Pol II (Kruk et al. 2011; Dutta et al. 2015). Dicer was also implicated in release of stalled RNA Pol II, which reduces DNA:RNA hybrid formation (Zaratiegui et al. 2011; Castel et al. 2014). These functions might also contribute to heterochromatin formation.

Our data show that chromatin-bound *tlh* RNA will recruit the Ccr4-Not complex to chromatin. In *clr4Δ* cells, we observe higher amounts of the Ccr4-Not complex on chromatin than in wild-type cells. This suggests that higher amounts of chromatin-bound RNAs recruit more Ccr4-Not to chromatin in these mutants. A recent study suggests that in mammalian cells, some lncRNAs might be degraded cotranscriptionally on chromatin, while others are degraded in the nucleoplasm (Schlackow et al. 2017). It remains to be analyzed why specific RNAs are degraded on chromatin and others in the soluble fraction. The Ccr4-Not complex is found in the cytosol and the nucleus (Collart 2016), and our data suggest that Ccr4-Not localization might be a result of RNA localization.

We also found that many euchromatic lncRNAs are retained at the chromatin in wild-type cells and even more in the absence of the Ccr4-Not complex. Although many euchromatic RNAs are retained at the chromatin, they do not recruit RNAi (Marasovic et al. 2013). Our data show that RNA localization on chromatin is not sufficient to initiate RNAi-mediated heterochromatin formation. This indicates that mutations in the Paf1 complex, which promote establishment of ectopic RNAi-dependent heterochromatin, interfere with chromatin functions of the Paf1 complex rather than RNA release from chromatin (Sadeghi et al. 2015; Verrier et al. 2015).

Many lncRNAs have been implicated in chromatin-related processes from yeast to human cells and are overexpressed in numerous human diseases and cancer (Sánchez and Huarte 2013). Several human lncRNAs were also suggested to be bound at chromatin (Böhmdorfer and Wierzbicki 2015; Yamashita et al. 2016). This indicates that the degradation of chromatin-associated transcripts might be a conserved mechanism to maintain chromatin structure.

Methods

Strain construction

All *S. pombe* strains used in this study are listed in the Supplemental Table S1. Strains were generated as previously described (Marasovic et al. 2013). The point mutations for Caf1D53AD243AD174A were chosen as previously described (Jonstrup et al. 2007). D50A corresponds to D53A according to a new PomBase annotation. The Ccr4 activity mutant, Ccr4H665A, was taken corresponding to the homologous Ccr4H818A mutant in *S. cerevisiae* (Chen et al.

2002). The *tlh* overexpression strains contain one-half of *tlh1* (5'*tlh*: 1–2800 bp; 3'*tlh*: 2801–5664 bp) or *LEU2* (from *S. cerevisiae*; from start codon to stop codon without 3' UTR) under control of the *nmt1* promoter and *adh* terminator with a downstream *ade6* reporter. Cloning was done in a standard way with detailed description in the Supplemental Methods. Primers are listed in the Supplemental Table S2.

siRNA sequencing

Endogenously tagged 3×Flag-Ago1 was immunoprecipitated and Ago1-bound siRNA libraries were generated as previously described (Marasovic et al. 2013). A detailed description can be found in the Supplemental Methods.

Total RNA isolation, reverse transcription, and qRT-PCR

Total RNA isolation, reverse transcription, and quantitative real-time PCR (qRT-PCR) were performed according to standard procedures, described in detail in the Supplemental Methods. Primers are listed in the Supplemental Table S2.

Total RNA and poly(A) RNA sequencing

rRNA of 1 μg total RNA was degraded with Terminator nuclease (Epicentre) in buffer A for 2 h at 30°C. For poly(A) RNA sequencing, poly-adenylated RNA was extracted from total RNA with oligo d(T)₂₅ magnetic beads (NEB). The RNA library was obtained using the NEBNext Ultra Directional RNA library prep kit for Illumina (NEB).

Northern Blot

Ten micrograms of total RNA was used for Northern Blot, which was performed in a standard way as described in the Supplemental Methods.

Growth assay and *ade6* reporter spot assay

Tenfold serial dilutions of cultures with OD₆₀₀ between 0.7 and 1.5 were made so that the highest density spot contained 10⁵ cells. Cells were spotted on nonselective YES medium, not supplemented YE (low adenine) medium or EMMC low adenine (10 mg/L adenine, 226 mg/L leucine, 226 mg/uracil). The plates were incubated at 32°C for 2–3 d and imaged.

Chromatin immunoprecipitation

ChIP was performed as previously described (Marasovic et al. 2013) using 1.2 μg immobilized (Dynabeads Protein A, Thermo Scientific) antibody against dimethylated H3K9 (H3K9me2, Abcam AB1220), H3 (ab1791, Abcam), H3S10P (ab5176, Abcam), anti-Flag M2-magnetic beads (Sigma-Aldrich), or 5 μL agarose conjugated Pierce HA epitope tag antibody (no. 26181, Thermo Scientific). Oligonucleotides used for quantification are listed in the Supplemental Table S2. For sequencing, a ChIP-seq library was made using the NEBNext Ultra II DNA Library Prep Kit for Illumina (NEB).

DRIP sequencing

DNA–RNA hybrid immunoprecipitation (DRIP) was performed as previously described (Wahba et al. 2016). Lysis was performed like in ChIP from a 50-mL pellet without crosslinking. For immunoprecipitation, the S9.6 antibody (Kerafast) was used. Sample treated with RNase H was compared with sample not treated with RNase H. For a detailed protocol, see the Supplemental Methods.

RNA IP/Pol II-bound nascent RNA/H3-bound RNA

RNA IP was performed like ChIP but without RNase A digestion, using anti-RNA Pol II CTD repeat YSPTSPS antibody [8WG16] (ab817, Abcam), anti-H3 antibody (ab1791, Abcam), or anti-Flag M2-magnetic beads (Sigma-Aldrich). After phenol-chloroform-isoamyl alcohol extraction, DNA was digested with DNase I (Thermo Scientific) for 2 h at 37°C. RNA was recovered with a second phenol-chloroform-isoamyl alcohol purification and ethanol precipitation. Either RNA was taken for making a sequencing library using the NEBNext Ultra Directional RNA library prep kit for Illumina (NEB) or it was reverse transcribed into cDNA with specific primers for subsequent qRT-PCR.

ChIP-exo sequencing

ChIP-exo was performed similarly as previously described with minor modifications listed in the [Supplemental Methods](#) (Rhee and Pugh 2012; Serandour et al. 2013).

Chromatin fractionation

The frozen pellet of a 10-mL culture with an OD₆₀₀ of 1.0 was lysed with 0.25- to 0.5-mm glass beads (Roth) and the BioSpec FastPrep-24 bead beater (MP-Biomedicals), with eight cycles at 6.5 m/sec for 30 sec and 3 min on ice. The chromatin fraction was separated from the soluble fraction by centrifugation at 21,000g for 20 min. DNA sample was treated with Proteinase K and RNase A, and the RNA sample was treated with Proteinase K and DNase I. Reverse transcription was performed with specific primers for *tlh* and *tdh1*. For more details, see the [Supplemental Material](#).

3×Flag-Caf1 RNA IP with chromatin fractionation

Caf1-associated RNA in the chromatin and the nonchromatin fraction was isolated like the RNA IP described above. Just before sonication, the sample was centrifuged for 20 min at 21,000g at 4°C. The supernatant was taken as “soluble fraction”; the pellet was washed twice with lysis buffer and then resuspended in lysis buffer, which formed the “chromatin fraction.” IP was performed with anti-Flag M2-magnetic beads (Sigma-Aldrich).

Analysis of sequencing data

Single end sequencing of libraries was performed on an Illumina GA IIX sequencer at the LAFUGA core facility of the Gene Center, Munich. The Galaxy platform was used to demultiplex the obtained reads with Je-Demultiplex-Illu (Goecks et al. 2010).

Demultiplexed Illumina reads were mapped to the *S. pombe* genome, allowing a two-nucleotide mismatch to the genome using Novoalign (<http://www.novocraft.com>). The h90 *S. pombe* genome was assembled using the *mat* sequence from PomBase and imported in IGV. Small RNA reads mapping to multiple locations were randomly assigned. By using custom Perl scripts (see [Supplemental Table S4](#); [Scripts.tar.bz](#)), the data sets were normalized to the number of reads per million (rpm) sequences for small RNA-seq or rpm mapping to coding sequences for total RNA-seq, poly(A) RNA-seq, Caf1-RIP-seq, and H3 RIP-seq. ChIP data were either normalized by rpm if variation in read amounts was low; if centromeric heterochromatin was lost for example, ChIP-seq data were normalized to regions that were not changed in different mutants. Caf1 ChIP and DRIP reads were summed in a window of 100 nt and divided by a corresponding control to display the fold-change using the Integrative Genomics Viewer (IGV) (<http://www.broad.mit.edu/igv>). Sequencing data were done in two replicates or the data were confirmed by another method like qRT-PCR, with

the exception of small RNA sequencing, which was done only once. Sequenced strains are listed in the [Supplemental Table S3](#).

Statistical analysis

Two-sided *t*-test for two independent data sets with high variance was used to calculate the *P*-value. The *P*-value is displayed with asterisks: <0.05 (*), <0.01 (**), and <0.001 (***)

Data access

The sequencing data from this study have been submitted to the NCBI Gene Expression Omnibus (GEO; <http://www.ncbi.nlm.nih.gov/geo/>) under the accession number GSE94129.

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