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Data Availability Statement: The data used in this study is a property of the Guarda Nacional Republicana, a governmental police force from Portugal and, by law, confidential. The authors obtained specific approval in order to use the data. Data request may be sent to the Divisão de Medicina Veterinária (cari.dsad.dmv@gnr.pt). Other researchers, who meet the criteria for access to confidential data, can access data in the same manner as the authors. The authors had no special access privileges. RESEARCH ARTICLE

The intra-articular administration of triamcinolone hexacetonide in the treatment of osteoarthritis. Its effects in a naturally occurring canine osteoarthritis model

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Abstract

Objective

To evaluate the effect of an intra-articular (IA) administration of triamcinolone hexacetonide, compared with saline.

Patients and methods

Forty (N = 40) hip joints were randomly assigned to a treatment group (THG, n = 20, receiving IA triamcinolone hexacetonide) and a control group (CG, n = 20, receiving IA saline). On treatment day (T0), and at 8, 15, 30, 90 and 180 days post-treatment, weight distribution, joint range of motion, thigh girth, digital thermography, radiographic signs, synovial fluid interleukin-1 and C-reactive protein levels were evaluated. Data from four Clinical Metrology Instruments was also gathered. Results were compared Repeated Measures ANOVA, with a Huynh-Feldt correction, Paired Samples T-Test or Wilcoxon Signed Ranks Test. A Kaplan-Meier test was performed to compare both groups, with p<0.05.

Results

Joints were graded as mild (65%), moderate (20%) and severe (15%). Patients of both sexes, with a mean age of 6.5 \pm 2.4 years and bodyweight of 26.7 \pm 5.2kg, were included. No differences were found between groups at T0. Comparing THG to CG, weight distribution showed significant improvements in THG from 8 (p = 0.05) up to 90 days (p = 0.01). THG showed lower values during thermographic evaluation in the Lt view (p<0.01). Pain and function scores also improved from 30 to 180 days. Increasing body weight, age, and presence of caudolateral curvilinear osteophyte corresponded to worse response to treatment.

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Results of the Kaplan Meier test showed significant differences between groups, with THG performing better considering several evaluations and scores.

Conclusion

THG recorded significant improvements in weight-bearing and in with the considered CMIs, particularly pain scores. Lower thermographic values were registered in THG up to the last evaluation day. Age, sex, and radiographic findings did significantly influenced response to treatment.

Introduction

Osteoarthritis (OA) is a disease transversal to all mammals and a source of chronic pain. For that reason, it represents a considerable burden to societies, representing a large investment in healthcare, while reducing productivity and quality of life [1-3]. Since OA is symptomatic only in the affected joint while, at the same time, lacking obvious extra-articular manifestations, it is well suited to administer local therapy by intra-articular (IA) injection [4, 5]. The changes that occur in slowly progressive spontaneous dog OA closely match those of human OA, while maintaining the same life stages that go by at a faster progression rate, and sharing many of the same environmental conditions. For those reasons, the naturally occurring canine model is considered the closest to a gold standard [6-11].

The medical approach to OA aims at slowing disease progression while relieving symptoms, particularly pain [9, 12]. IA corticosteroids have been used for several decades to palliate pain and inflammation associated with OA and of joint's surrounding tissues [13, 14]. Its use should be especially considered in patients with moderate to severe pain, nonresponding to oral analgesic/non-steroidal anti-inflammatory drugs. A human systematic review has deemed triam-cinolone more effective than betamethasone and methylprednisolone [15]. Triamcinolone hexacetonide (TH), in particular, is described as able to provide pain relief and improved mobility for prolonged periods [16, 17]. In a canine model of OA, animals treated with IA TH showed a significant reduction of osteophyte size compared with a control group. At the histological level, TH significantly reduced the severity of OA structural changes of cartilage and had no deleterious effects on normal cartilage [18]. By effect is obtained through a dose-dependent reduction in the cartilage proteolytic enzyme stromelysin, interleukin 1 β , and the oncogenes c-fos and c-myc, which are involved in the metalloproteinases synthesis [19]. In human patients, the mean duration of effect of TH in patients with OA is around seven months [20].

Since pain and functional limitations are the most relevant clinical signs of OA, clinical trials and studies need to assess these parameters in order to evaluate patients and assessment of response to treatment [21–23]. Clinical metrology instruments (CMI) represent a patient-centred approach, and the most commonly used are the Canine Brief Pain Inventory (CBPI, divided in a pain severity score—PSS, and a pain interference score—PIS) and the Liverpool Osteoarthritis in Dogs (LOAD). Further validated CMIs include the Hudson Visual Analogue Scale (HVAS), and the Canine Orthopaedic Index (COI, divided into four scores: stiffness, gait, function and quality of life—QOL). As a whole, CMIs complement the evaluation of the multi-dimensional experience that is OA related pain [9, 23–31]. Typically, OA pain is localized and related to movement or weight-bearing of the affected joints, and affected patients commonly bear less weight on a painful limb. Evaluating weight distribution through stance analysis is a sensitive evaluation canine lameness [22, 23, 32]. Additional functional evaluations aim at assessing activity levels and mobility impairments [26]. Pedometry is a simple and inexpensive method to assess mobility levels, which can measure ambulatory activity with an acceptable level of accuracy [33]. Additional clinical measurements include the examination of muscle masses and evaluation of the joint range of motion, which are consistently reduced and restricted in OA patients [34–37].

Imaging plays a key role in the assessment of patients with joint disease and, in cases of hip OA, the ventrodorsal (VD) hip extended view is the most common pelvic radiographic projection. The ventrodorsal flexed view, also called frog-legged view (FL), useful for further evaluation of the presence of the circumferential femoral head osteophyte (CFHO) and caudolateral curvilinear osteophyte (CCO), radiographic findings related with the development of clinical signs [38–43]. By correlating changes in temperature patterns with various disease, degenerative or injury processes, digital thermography can provide a reproducible diagnostic tool [44–46]. This diagnosis modality can differentiate normal from osteoarthritis subjects [47, 48].

Since OA is a low-grade inflammatory disease, the analysis of synovial fluid (SF) can add additional information regarding the disease's characterization [1]. Interleukin 1 (IL-1) is the most important proinflammatory catabolic cytokine in OA, with a highly potent capability of inducting cartilage degradation and relation with lameness duration [49-51]. C-reactive protein (CRP) can be produced at the level of the inflamed tissues, and its shifts occur from a very early stage [52, 53]. It has been highly associated with knee OA in humans [54].

The goal of this study is to compare the effect of triamcinolone hexacetonide to a control group in the management of OA in a naturally occurring canine model, using several outcome assessment modalities. We hypothesize the intra-articular administration of triamcinolone hexacetonide will be able to reduce the clinical signs of OA, compared to a control group.

Materials and methods

The study protocol was approved by the ethical review committee of the Universidade de Évora (ORBEA, approval n° GD/32055/2018/P1, September 25th, 2018), and complies with the ARRIVE guidelines. Written, informed consent was obtained from the Institution responsible for the animals. Twenty dogs were selected based on medical history records, physical, orthopaedic, neurological and radiographic examinations compatible with hip OA, and the sample comprised forty (N = 40) joints of twenty active police working dogs with bilateral hip OA. To be included in the study, patients should be over two years, have a bodyweight over 20kg and should not have received any medication or nutritional supplement for over six weeks before enrolment in the study. Patients with any other documented or suspected orthopaedic or neurological disease, or additional concomitant disease (ruled out through physical examination, complete blood count, and serum chemistry profile), were excluded.

In a double-blinded study, dogs with affect joints were randomly assigned to a control group (CG, n = 20 joints) or a treatment group (THG, n = 20 joints). Evaluations were conducted on days 0 (treatment day), 8, 15, 30, 90 and 180. Days were counted from treatment day (day 0). An outline of all procedures on each evaluation day is presented in <u>Table 1</u>. The same researcher performed all evaluations.

On treatment day, patients in CG received an IA administration of 2ml of 0.9%NaCl. On the same day, patients in THG received an IA administration of 20mg in a volume of 1 ml of triamcinolone hexacetonide (Bluxam, Riemser Pharma). IA administrations and radiographic examination were conducted under light sedation, induced with a combination of medetomidine (0.01mg/kg) and buthorphanol (0.1mg/kg), given intravenously. After all procedures were conducted, sedation was reversed with atipamezole (100–150µg/kg), administered intramuscularly. Both a VD extended legs and FL views were obtained. On the VD view, the

Procedure				Day		
	0	8	15	30	90	180
Treatment	X					
Digital Thermography	X	X	Х	X	X	X
Digital radiography	X			X	X	X
Stance analysis	X	X	X	X	X	X
Pedometer	X	X	Х	X	X	X
Goniometry	X	X	Х	X	X	X
Thigh girth measurement	X	X	X	X	X	X
Clinical Metrology Instruments	X	X	Х	X	X	X
Synovial fluid CRP	X	X		X	X	X
Synovial fluid IL-1	X	X		X	X	X

Table 1. Procedures conducted in each day.	Days are counted from treatment day
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CMI-Clinical Metrology Instruments; CRP-C-Reactive Protein; IL-1-Interleukin 1.

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presence of the following findings was assessed; an irregular wear on the femoral head, making it misshapen and with a loss of its rounded appearance; a flattened or shallow acetabulum, with irregular outline; CCO; new bone formation on the acetabulum and femoral head and neck; a worn away angle formed at the cranial effective acetabular rim; subchondral bone sclerosis along the cranial acetabular edge; and CFHO [43, 55, 56]. For the IA administration, patients were positioned in lateral recumbency, with the joint of interest uppermost. A window of 4x4cm in the area surrounding the greater trochanter was clipped and aseptically prepared. The limb was then placed in a neutral, parallel to the table position and a 21-gauge with 2.5" length needle was introduced just dorsal to the greater trochanter, perpendicular to the long axis of the limb until the joint was reached [57]. Confirmation of correct needle placement was obtained through the collection of SF. As much SF as possible was aspirated and kept for the posterior determination of IL-1 β and CRP concentrations, and the treatment or saline was administered.

Evaluation of weight distribution was carried out with a weight distribution platform (Companion Stance Analyzer; LiteCure LLC[®], Newark, Delaware, United States), following manufacturer's guidelines. The equipment was placed in the centre of a room, at least 1 meter from the walls. It was also calibrated at the beginning of each day and zeroed before each data collection. For the evaluation, patients were led to stand on the platform, with one foot on each quadrant, with their heads facing forward. The left-right symmetry index was calculated with the following formula: symmetry index = $[(WB_R-WB_L)/((WB_R+WB_L)x0.5)]x100$ [28, 58], where WB_R is the value of weight-bearing for the right pelvic limb, and WB_L is the value of weight-bearing for the left pelvic limb. Negative values were made positive. Additionally, we considered a deviation from the normal 20% weight-bearing for a pelvic limb [59], calculated by subtracting WB to 20.

Pedometers were worn around the patient's neck, attached to an adjustable lightweight collar, to measure ambulatory activity and mobility levels [60]. They were worn for a week before the first evaluation time, to order to establish a baseline value, and before each evaluation time. Mean daily counts were considered, calculated by dividing the register number of steps by the number of considered days. In a quiet room, with as much time as needed to answer all items, trainers completed a copy of HVAS, CBPI, COI and LOAD, in sequence by the same handler at all evaluation days.



Fig 1. A lateral view of a dog with moderate osteoarthritis, with the greater trochanter in the centre, at a distance of 60 cm. The range of temperature was set at 15–40°C and emissivity at 0.98. Thermographic images were analyzed with a Rainbow HC colour pallet. A temperature box is placed on the anatomical area of the hip joint.

For the digital thermography evaluation, animals were kept for 30 minutes in a room with controlled temperature, at 21°C. During this period, they were allowed to walk around the room calmly. A dorsoventral image was obtained with patients in a symmetrical upright standing, including the area from the last lumbar to the first coccygeal vertebrae, at a distance of 60 cm [61]. A lateral view was also obtained, with the greater trochanter in the centre, at the same distance. All images were taken with a FLIR ThermaCAM E25^(R) model (FLIR Systems, Wilsonville, Oregon, United States). The posterior analysis was conducted with free software (Tools, FLIR Systems, Inc), using a rainbow colour pallet. Temperature boxes were placed on the anatomical area of the hip joint, to determine mean and maximal temperatures (Fig 1).

Both thigh girth and joint range of motion were determined with the patient in lateral recumbency. A Gullick II measuring tape was used to evaluate thigh girth, at a distance of 70% thigh length, measured from the tip of the greater trochanter, with an extended leg [62]. Hip joint ROM was then determined with a goniometer at extension and flexion with a flexed stifle [63].

Determination of IL-1 β and CRP concentrations were made using Fuji Dri-Chem Slides VC-CRP PS (FUJIFILM Europe GmbH), read with a DRIChem NX500i (FUJIFILM Europe GmbH), and a DuoSet Ancillary Canine IL-1 β Reagent kit (R&D Systems, United Kingdom), read with a FLUOstar OPTIMA (BMG Labtech).

After treatment, animals were rested for three consecutive days and resumed their regular activity over five days. On days 1 and 3 after the procedure, the veterinarian examined all patients in order to determine existing signs of exacerbated pain, persistent stiffness of gait and changes in posture. If no complaints were registered, the animal could resume its normal activity [64, 65].

Normality was assessed with a Shapiro-Wilk test. Different group's results were compared in each evaluation day, and each measured parameter was compared with the result observed at treatment day. To assess the effect of different parameters on the patients' clinical evolution, results were compared by sex, age and different cut off values for body weight with Repeated Measures ANOVA, with a Huynh-Feldt correction, Paired Samples T-Test, or Wilcoxon Signed Ranks Test. A Kaplan-Meier test was performed to evaluate the time to return to base-line values of symmetry index and CMI scores, compared with the Breslow test. All results were analyzed with IBM SPSS Statistics version 20, and a significance level of p<0.05 was set.

Results

The sample included 40 hip joints (n = 20 left and n = 20 right) of active police working dogs with a mean age of 6.5 ± 2.4 years and bodyweight of 26.7 ± 5.2 kg. Both sexes (male n = 28, female n = 12) and four breeds were represented: German Shepherd Dogs (n = 8), Belgian Malinois Shepherd Dogs (n = 6), and Dutch Shepherd Dog (n = 6). At T0, 26 joints were classified as mild (65%), 8 as moderate (20%) and 6 as severe (15%), according to the Orthopedic Foundation for Animals hip grading scheme. No differences were found between groups at the initial evaluation. After the initial rest period, all animals resumed normal activity, with similar workload and movement compared to that before treatment.

Clinical and laboratorial findings

Values recorded for different assessments at the initial evaluation, and its variations throughout the study, for THG and CG, are presented in Table 2. Comparing results between groups with repeated measures ANOVA with a Huynh-Feldt correction, significant differences between groups were found concerning body weight (F(2.8,140) = 4.2, p<0.01), deviation (F (4.8,109) = 2.8, p = 0.02), symmetry index (F(2.8,77.8) = 7.5, p<0.01), mean temperature on a DV view (F(3.9,93.9) = 6.6, p<0.01), maximal temperature on a DV view (F(3.6,86.2) = 6.9, p<0.01), mean temperature on a Lt view (F(4.5,113.2) = 26.7, p<0.01), maximal temperature on a Lt view (F(4.1,101.6) = 96.2, p<0.01), joint flexion (F(4.9,146.7) = 19.5, p<0.01), IL-1 synovial concentration (F(1.9,58.3) = 4.9, p = 0.02). Significant differences were observed between groups regarding CMI scores, specifically PSS (F(5,120) = 2.4, p<0.05), PIS (F(5,120) = 2.6, p = 0.03) and function (F(2.8,69.2) = 2.4, p = 0.04). Evolution of the symmetry index in CG and THG is presented in Fig 2. Results of the Kaplan Meier test are presented in Table 3. Kaplan Meier curves for symmetry index and function score are presented in Figs 3 and 4, respectively. A dorsoventral digital thermography view is presented in Fig 5.

Radiographic findings

Frequency of different radiographic findings observed in CG and THG, at the initial evaluation, are presented in Table 4. In THG, an increase in the frequency of flattened or shallow acetabulum, with irregular outline was observed at 90 and 180 day (p<0.05). Increased new bone formation on the acetabulum and femoral head and neck was also observed at 90 day (p<0.05), as the frequency of CCO at 180 day (p<0.05).

In CG, an increase in the frequency of flattened or shallow acetabulum, with irregular outline, an increase was observed at 90 (p<0.01) and 180 day (p<0.01), compared with the initial evaluation day. In the THG, patients without CCO had higher LOAD (p<0.05), PSS (p = 0.02), PIS (p = 0.01), stiffness (p = 0.01), function (p<0.05), gait (p<0.01), QOL (p<0.01) and COI scores (p<0.01), and lower HVAS scores (p<0.01). At 15 day, they had lower mean and maximal thermographic evaluations on a DV (p = 0.01 for both) and mean on a Lt view (p = 0.04), and higher LOAD (p<0.01), stiffness (p = 0.04), function (p<0.01), gait (p<0.01), QOL (p<0.01) and COI scores (p<0.01). Again at the 30 and 90 day evaluations, those without CCO at the initial evaluation had higher HVAS (p<0.05 and p<0.01, respectively), and lower PSS (p<0.01 for both), PIS (p<0.01 and p = 0.02, respectively), LOAD (p<0.01 for both), stiffness (p = 0.04 and p = 0.02, respectively), function (p<0.01 and p = 0.04, respectively), gait

	fodality		Treatme	ant day					8 days			_			15 days				_	-	
		Ũ	U	HL	(g			ЭHG	ď		CG			THG		Р			
		mean	SD	mean	SD	mean	SD	p 1	mean	SD I		mea	n SD	d	mean	SD	р				
Goniometry	Flexion (°. mean±SD)	55.0	4.4	57.0	4.1	55.3	3.7	<0.01*	55.3	4.7 0.	.09 1.	0 57.2	2 5.2	0.14	58.0	5.3	0.53	1.0			
	Extension (°. mean±SD)	151.2	3.9	148.0	8.8	149.9	4.6	0.95 1	49.4	4.5 0.	41	151.1	3.5	0.07	151.3	5.1	< 0.05*	1.0			
	Thigh girth (cm. mean ±SD)	31.2	2.6	30.4	3.4	31.1	3.3	0.94	30.8	3.1 1.	0.	49 31.1	1 2.9	0.86	32.6	3.5	< 0.01*	0.58			
	Pedometer (daily steps ±SD)	1445.7	755.7	1539.6	397.3	829.5	931.3	0.58 5	89.8 15	54.6 0.	.85	606.0	309.5	0.15	1215.1	793.8	0.03*	1.0			
CMI	HVAS (0-10)	6.8	1.2	5.7	1.9	6.7	1.5	0.48	6.4	1.7 0.	.19 0.	16 6.8	3 1.2	0.6	6.3	1.6	0.72	0.16			
	CBPI-PSS (0-10)	3.1	1.9	4.2	2.8	3.4	2.3	0.69	2.9	1.8 0.	03* <0.	05* 3.7	7 2.8	0.2	3.2	1.9	0.04^{*}	0.04*			
	CBPI-PIS (0-10)	3.2	2.2	4.8	3.3	3.4	2.1	0.01*	3.3	2.1 0.	.04* 0.	02* 3.6	5 2.1	0.01*	3.5	2.1	0.03*	0.01*			
	COI-Stiffness (0-16)	3.4	3.4	6.8	4.2	4.1	3.3	0.31	5.5	3.6 0.	38 0.	51 4.1	1 3.2	0.31	5.1	4.0	0.46	10.19			
	COI-Function (0-16)	3.6	4.1	6.3	5.7	4.1	4.0	0.64	4.1	4.0 0.	21 <0.	05* 4.4	1 5.5	0.72	3.8	3.9	0.75	0.03*			
	COIGait (0-20)	4.7	5.2	10.5	5.9	5.4	6.1	0.29	7.3	4.7 0.	.18 0.	21 5.6	3 4.3	0.02*	6.8	5.6	< 0.05*	0.19			
	COI-QOL (0-12)	4.5	2.6	6.2	3.9	4.6	2.7	0.62	5.5	3.0 0.	28 0.	85 4.7	7 2.9	0.06	4.4	3.6	0.56	0.09			
	COI—Overall score (0-64)	16.4	14.7	29.8	19.1	18.2	13.8	0.22	23.8	14.6 0.	.19 0.	48 18.¢	5 13.8	0.14	21.0	16.7	0.83	0.19			
	LOAD (0-52)	13.6	10.5	23.2	14.1	14.4	12.7	1.0	18.3	12.0 0.	98 0.	33 14.3	3 10.7	0.83	18.0	12.3	0.88	0.14			
Digital	DV (°. mean±SD)	24.7	1.9	25.3	0.6	25.2	1.3	0.01*	25.3	0.6 0.	08 <0.	01* 24.4	1 1.6	0.61	23.4	2.5	0.03*	1.0			
Thermography	DV max (°. mean±SD)	26.3	1.9	26.4	1.6	25.8	1.0	0.06	24.9	1.6 0.	24 <0.	01* 26.5	7 1.6	0.97	24.6	2.5	0.03*	1.0			
	Lt (°. mean±SD)	28.7	2.7	26.5	1.9	31.6	2.1	<0.01*	30.5	2.8 <0.	01* <0.	01* 29.5	7 2.9	< 0.01*	28.5	4.0	0.02*	<0.01*			
	Lt max (°. mean±SD)	31.9	3.1	30.4	4.1	34.9	1.0	<0.01*	34.6	1.1 <0.	01* <0.	01* 34.9	9 0.8	< 0.01*	34.0	1.9	< 0.01*	<0.01*			
Synovial fluid	IL-1 (pg/mL. mean±SD)	170.9	120.4	208.5	95.2	72.3	42.4	<0.01*	98.4	80.8 0.	04* 0.	04* -	·	•	•						
	CRP (mg/mL. mean±SD)	0.4	1.0	2.5	3.5	0.3	1.2	< 0.01*	0.0	0.0 0.0	.18 1.	- 0	•	•	•						
Weight-bearing	Symmetry Index (mean ±SD)	24.7	20.3	53.9	50.4	18.7	17.1	0.06	19.2	18.1 <0.	.05* <0.	05* 23.5	9 16.3	0.18	18.9	10.9	0.01*	0.03*			
	Deviation (mean±SD)	2.8	3.6	4.7	4.4	2.78	1.987	0.3	2.1	1.9 0.	.08 0.	43 2.5	34 2.13	27 0.47	2.2	1.6	< 0.05*	0.02*			
4	Aodality			ē	0 days						90 d	lays					18	0 days			
			CG			THG		Р		CG		TH	(5)	Ч		CG		TI	ÐF		2
		mean	SD	Ч	mean	SD	Р	_	nean	SD I	me	an SD	đ		mean	SD	Р	mean	A	- -	
Goniometry	Flexion (°. mean±SD)	53.6	2.9	0.11	51.8	3.9	<0.01*	<0.01*	52.7	2.9 0.	.02* 52.	0 3.5	3 <0.0	1* <0.01*	51.6	2.2	0.00*	49.3	4.3 <0	.01* <0	.01*
	Extension (°. mean±SD)	150.8	3.4	0.06	152.3	3.7	0.09	1.0 1	50.8	2.9 0.	07 151.	9 3.() <0.0	1* 0.23	151.3	2.9	0.17	150.2	3.7 0	.26 1	0.
	Thigh girth (cm. mean ±SD)	30.6	2.7	0.39	29.5	3.1	0.25	0.44	31.6	2.7 0.	54 31.	5 3.5	0.0	9 0.43	31.5	2.2	0.2	30.2	3.7 0	96.	.32
	Pedometer (daily steps ±SD)	594.5	663.4	0.48	747.9	548.2	0.65	0.15 4	151.9 4	63.0 0.	4 410.	8 497.4	4 0.1:	5 0.08	434.9	455.8	0.2	376.0 26	3.3 <0	.01* 0	.63
CMI	HVAS (0-10)	6.4	1.4	0.14	6.3	1.9	0.64	0.17	6.6	1.7 0.	22 6.	5 1.5	3 0.8	4 0.21	6.5	1.4	0.04^{*}	6.4	1.6 0	.13 0	.12
	CBPI-PSS (0-10)	3.7	2.6	0.03*	3.8	2.6	0.02*	0.01*	4.1	2.9 0.	.02* 3.	2 2.1	0.5	7 0.32	3.6	3.1	0.02*	3.6	2.5 0	98 0	.23
	CBPI-PIS (0-10)	3.8	2.6	0.01^{*}	5.7	5.3	0.04*	<0.05*	3.9	2.8 0.	.01* 3.	.1 2.4	4 0.0:	7 0.33	3.5	2.4	0.01*	4.0	3.0 0	.63 0	.22
	COI-Stiffness (0-16)	4.6	4.1	0.87	5.3	4.0	0.78	0.48	4.6	3.9 0.	33 4.	9 3.6	5 0.5(0.39	4.0	5.7	0.82	4.8	4.4 0	.10	.48
	COI-Function (0-16)	5.7	5.3	0.2	5.7	5.3	0.79	< 0.05*	5.0	5.2 0.	21 4.	9 4.4	4 0.7:	5 < 0.01*	4.0	5.4	1.0	3.3	3.8 0	.39 0	.01*
	COI-Gait (0-20)	6.9	5.1	0.19	7.8	6.8	0.69	0.19	5.7	5.5 0.	.11 6.	7 4.6	5 0.15	8 0.16	4.4	5.4	0.87	6.5	5.8 0	.01* 0	.46
	COI-QOL (0-12)	5.3	3.3	0.39	5.0	3.7	<0.01*	0.57	5.1	2.8 0.	.02* 4.	3 2.7	7 0.45	9 0.59	4.7	2.6	0.09	4.4	3.2 0	.03* 0	.25
	COI—Overall score (0-64)	22.4	19.1	0.04^{*}	22.9	19.7	0.75	0.26	20.1	15.7 0.	29 20.	3 14.5	0.5	3 0.14	15.7	14.9	0.1	20.9 1	8.9 0	0 60.	21
	LOAD (0-52)	16.4	13.1	0.22	18.1	13.5	0.47	0.88	13.1	12.4 0.	72 15.	.1 9.4	4 0.0:	7 0.17	13.1	12.4	0.88	16.0 1	2.0 0	.03* 0	.07
Digital	DV (°. mean±SD)	25.3	1.5	0.36	24.4	0.8	0.36	1.0	26.1	1.2 0.	.04* 26.	3 1.6	5 0.02	2* 0.68	25.6	1.4	0.89	25.1	0.9 0	.57 1	0.
Thermography	DV max (°. mean±SD)	25.2	2.1	0.88	25.9	0.7	0.31	1.0	27.4	1.4 0.	.14* 27.	.6 1.1	<0.0.	1* 0.02*	26.9	1.4	0.74	26.5	0.9 0	.78 1	0.
	Lt (°. mean±SD)	29.8	2.2	$< 0.01^{*}$	29.5	2.3	<0.01*	<0.01*	28.4	1.8 <0.	01* 29.	0 2.6	S <0.0.	1* <0.01*	27.3	1.8	0.21	28.7	2.3 <0	.01* <0	.01*
	Lt max (°. mean±SD)	33.9	1.2	$< 0.01^{*}$	33.7	1.6	<0.01*	<0.01*	30.5	1.9 <0.	.01* 31.	4 2.6	5 <0.0	1* <0.01*	29.7	1.9	0.13	31.2	2.3 <0	.01* <0	.01*
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Synovial fluid	IL-1 (pg/mL. mean±SD)	122.9	108.9	0.05	186.6	104.5	0.24	1.0	159.6	59.1	0.13	169.1	55.3	0.36	1.0	184.2	68.5	0.25	145.1	33.5	0.07	1.0
	CRP (mg/mL. mean±SD)	0.48	6.0	0.18	0.1	0.2	0.43	1.0	0.4	0.8	0.36	0.0	0.0	0.42	1.0	0.0	0.0	0.5	0.0	0.0	0.18	1.0
Weight-bearing	Symmetry Index (mean ±SD)	18.9	12.2	0.04^{*}	13.3	8.6	<0.01*	<0.01*	27.4	12.1	0.29	14.0	13.5	0.02*	0.02	27.0	27.9	0.51	20.7	22.7	0.01*	0.22
	Deviation (mean±SD)	2.5	1.917	0.2	1.8	2.3	0.02^{*}	$< 0.01^{*}$	2.72	2.27	0.29	2.3	2.1	0.04^{*}	$< 0.01^{*}$	2.61	2.973	0.55	2.0	2.8	0.04^{*}	0.75
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CBPI-Canine Brief Pain Inventory; CRP-C-reactive protein; COI-Canine Orthopedic Index; DV-dorsoventral view; HVAS-Hudson Visual Analogue Scale; IL-1 -Interleukin 1; LOAD-Liverpool Osteoarthritis in Dogs, LT-lateral view; PIS-Pain Interference Score; PSS-Pain Severity Score; QOL-Quality of Life.

* indicates significance when comparing the value registered by a group at an evaluation day with T0 (p), and comparing both groups at each follow-up day (P).

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Simmetry Index by evaluation moment



(p<0.01 and p = 0.04, respectively), QOL (p<0.01 for both) and COI scores (p<0.01 and p = 0.02, respectively). At the final evaluation, they had only higher HVAS score (p = 0.03). In CG, animals without CCO at the initial evaluation did now show significant differences with those that did. However, at the 30-day evaluation, they had higher mean thermographic evaluation on a Lt view (p = 0.04), better joint flexion (p = 0.03), lower IL-1 and higher CRP concentration levels (p = 0.04 for both). On the 90 day evaluation, animals without CCO had lower maximal thermographic evaluation on a Lt view (p<0.05) and lower CRP values at 180 days (p = 0.02).

			Trea	atment	
Variable	Breslow test	C	G	T	HG
		mean±SD	95% CI	mean±SD	95% CI
Symmetry Index	0.003*	47.0±11.8	23.8±70.2	96.0±12.8	70.9±121.1
Deviation	0.022*	44.8±12.1	21.1±68.5	81.8±14.7	52.9±110.6
HVAS	0.269	48.7±12.4	25.4±73.9	66.1±14.2	38.3±93.9
PSS	0.065	63.2±17.2	29.6±96.8	90.2±17.6	55.7±124.7
PIS	0.000*	8.4±0.4	7.7±9.0	118.6±16.3	86.7±150.5
LOAD	0.000*	40.7±10.6	19.9±61.4	124.3±15.9	93.1±155.5
Stiffness	0.004*	64.7±16.9	31.4±97.9	130.8±11.6	108.1±153.5
Function	0.046*	65.4±13.4	39.2±91.6	81.9±143.2	
Gait	0.001*	52.7±14.6	23.9±81.4	117.0±15.1	87.5±146.5
QOL	0.044*	60.9±15.0	31.4±90.4	119.3±17.5	85.0±153.6
COI	0.146	52.7±13.4	26.5±78.9	85.6±15.9	54.4±116.9

Table 3. Time to return to baseline values for weight-bearing distributions (symmetry index and deviation) and CMIs, calculated with Kaplan-Meier estimators and compared with the Breslow test.

COI—Canine Orthopedic Index; HVAS—Hudson Visual Analogue Scale; LOAD—Liverpool Osteoarthritis in Dogs; PIS—Pain Interference Score; PSS—Pain Severity Score; QOL—Quality of Life. * indicates significance.

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Fig 4. Kaplan-Meier curve demonstrating a significant difference between the control group (CG) and triamcinolone hexacetonide group (THG) in time for function score to return to baseline values ($p = 0.046^{\circ}$).

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Fig 5. A dorsoventral view of a dog with moderate osteoarthritis, including the area from the last lumbar vertebra to the first coccygeal vertebra at a minimum, at a distance of 60 cm. Arrow indicates cranial direction. The range of temperature was set at 15–40°C and emissivity at 0.98. Thermographic images were analyzed with a Rainbow HC colour pallet.

In the THG, patients without CFHO at the initial evaluation had lower pedometer counts (p<0.01) and lower HVAS (p = 0.04) and higher PIS scores (p = 0.03) on that day. At 8 day, they had higher body weight (p = 0.02) and higher deviation (p = 0.01) and symmetry index (p<0.05). At 15 day, they had higher body weight (p = 0.04), lower pedometer count (p<0.01) and higher LOAD score (p<0.05). Again at 30 day, these patients showed higher deviation (p = 0.04) and symmetry index (p<0.01). At 90 day, they had higher deviation (p = 0.03), symmetry index (p<0.01) and higher synovial IL-1 concentration (p = 0.01). At the final

Table 4. Frequency of radiographic findings in the Control (CG) and Treatment Groups (THG) in a ventrodorsa
and frog-leg views, at the initial evaluation.

Radiographic finding		TH	IG			С	G	
	Pı	esent	At	osent	Pr	resent	At	osent
Irregular wear on the femoral head, making it misshapen and with a loss of its rounded appearance	20	100%	0	0%	17	85%	3	15%
Flattened or shallow acetabulum, with irregular outline	16	80%	4	20%	11	55%	9	45%
Caudolateral curvilinear osteophyte (CCO)	8	40%	12	60%	5	25%	15	75%
New bone formation on the acetabulum and on femoral head and neck	17	85%	3	15%	20	100%	0	0%
The angle formed at the cranial effective acetabular rim is worn away		90%	2	10%	18	90%	2	10%
Subchondral bone sclerosis along the cranial acetabular edge	20	100%	0	0%	19	95%	1	5%
Circumferential femoral head osteophyte (CFHO)	8	40%	12	60%	3	15%	17	85%

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evaluation day, they had lower pedometer counts (p = 0.02). In CG, joints without CFHO at the initial evaluation had higher joint extension (p<0.01) and HVAS (p = 0.02), lower PSS (p = 0.01) and PIS scores (p = 0.03) at the 8 day evaluation. At 15 day, they had higher mean thermographic values on a Lt view (p = 0.02), lower PSS (p = 0.02) and PIS scores (p<0.05). This higher mean thermographic values on a Lt view was again observed at 30 day (p = 0.01) and higher HVAS scores (p = 0.02) at 90 day. At the final evaluation, they had higher maximal thermographic values on a Lt view (p = 0.04), and lower PSS (p = 0.05) and PIS scores (p<0.03).

Comparisons by sex

In the THG, females had lower symmetry index (p < 0.01) and lower synovial CRP concentration (p < 0.01). At the 8 day evaluation day, females were lighter than males (p = 0.04) and had higher synovial IL-1 levels (p = 0.04). Additional differences were observed at 15 days, with female dogs having lower mean and maximal temperatures on a DV (p<0.01 for both) and Lt views (p = 0.04 and p < 0.01, respectively). At 30 days, females had higher pedometer counts (p<0.01) and, at 90 days, lower mean and maximal temperatures on a DV view (p<0.05 and)p<0.01, respectively). At the final evaluation day, no differences were observed between sexes. Female dogs of CG had significantly lower body weight throughout the study (p = 0.01). In the first evaluation, they also showed higher values in all thermographic evaluations (p < 0.01) and lower PIS scores (p = 0.04). Again at 8 days, higher thermographic evaluations were recorded (p < 0.01), except maximal value on a Lt view, as higher joint extension values (p < 0.01). A higher joint extension was again observed in female dogs at 15 days (p = 0.04), with lower PIS scores (p = 0.03). At the 30 days evaluation females again showed higher thermographic maximal values on an LT view max (p < 0.01). Female dogs at 90 days had lower thigh girth (p = 0.03) and lower PSS and PIS scores (p = 0.01). At the final evaluation day, they had higher extension values (p = 0.02), higher HVAS (p = 0.02), and lower PSS (p < 0.01) e PIS (p < 0.01), stiffness (p = 0.02), function (p = 0.02), gait (p<0.01), QOL (p = 0.02) and COI (p = 0.01) scores.

Comparisons by bodyweight

Comparing animals in THG with a weight below the mean value of the sample, had higher deviation (p = 0.03), symmetry index (p = 0.04), mean and maximal values on thermography DV view (p < 0.01 and p = 0.02, respectively), lower thigh girth (p < 0.01) and lower synovial IL-1 levels (p = 0.02). At 8 days, they had lower thigh girth (p < 0.01) and lower PSS (p = 0.01), PIS (p<0.01) and stiffness scores (p = 0.01). After 15 days, lighter patients had higher symmetry index (p = 0.03) and lower thigh girth (p < 0.01). At 30 days, these patients had higher pedometer counts (p = 0.01) and lower thigh girth (p < 0.01). At the 90-day evaluation, they had higher mean and maximal values on thermography Lt view (p = 0.02 and p < 0.01, respectively), lower thigh girth (p < 0.01) and higher joint flexion (p = 0.03). 30 At the final evaluation day, lighter animals had lower mean and maximal values on thermography a DV (p < 0.01 for both) but higher on a Lt view (p < 0.01 for both), lower thigh girth (p < 0.01) and higher joint flexion (p = 0.03). In CG, lighter patients registered had lower PIS scores (p = 0.04) at the initial evaluation. These patients, at the 8 day evaluation, had higher thermographic mean and maximal values on a DV (p = 0.03 and p = 0.02, respectively), lower thigh girth (p = 0.01), and higher stiffness (p = 0.03), function (p < 0.01), gait (p = 0.03) and COI scores (p < 0.01). Significant differences were again observed at 15 days, with lighter patients showing lower thigh girth (p = 0.04) and HVAS (p < 0.05), and higher stiffness, function, gait QOL e COI scores (p < 0.01). They also had lower CRP concentrations at 30 days (p = 0.04) and higher HVAS

scores (p = 0.02). At 90 days, they had lower thigh girth (p<0.01) and IL-1 levels (p = 0.02) at 90 days. At the final day of evaluation, lighter animals showed higher mean thermographic values on a DV view (p<0.01), and higher joint flexion (p = 0.02) and extension (p<0.01).

Comparisons by age

Considering patients above or below the mean age in the THG, at the initial evaluation younger patients had higher mean and maximal temperature on a Lt view (p < 0.01 for both), and lower PSS (p = 0.01), stiffness (p = 0.04), function (p = 0.02), gait (p = 0.02), QOL (p = 0.04), and COI scores (0.01). After treatment, at 8 days, they had lower deviation (p = 0.03) and symmetry index (p = 0.04), higher mean temperature on a Lt view (p < 0.01), lower synovial IL-1 concentration (p = 0.04), and lower PIS (p < 0.02) and function scores (p = 0.03). At 15 days, they had higher mean and maximal temperature on a DV view (p<0.01 for both) and Lt view (p<0.01 for both), and lower PIS (p<0.01), stiffness (p = 0.03), gait (p<0.01) and COI score (p = 0.04). After 30 days, younger animals had higher mean and maximal temperature on a Lt view (p = 0.01 for both), higher HVAS (p<0.01), and lower PSS (p<0.01), PIS (p<0.01), LOAD (p < 0.01), stiffness (p < 0.01) and QOL scores (p < 0.01). Differences regarding CMI scores were observed again at 90 days, with the same patients having higher HVAS (p<0.01), and lower PSS (p = 0.01), PIS (p < 0.01) and QOL scores (p = 0.01). At the final evaluation, patients below the mean age value had higher pedometer counts (p < 0.01), lower deviation (p = 0.02) and SI (p < 0.01), higher HVAS (p < 0.01), and lower PSS (p < 0.01), PIS (p < 0.01), LOAD (p < 0.01), stiffness (p < 0.01), function (p = 0.04), gait (p < 0.01), QOL (p < 0.01) and COI scores (p < 0.01). In the CG at the initial evaluation, younger patients had higher maximal values on the thermographic Lt view (p = 0.04), lower LOAD (p = 0.02), stiffness (p < 0.01), function (p < 0.01), gait (p < 0.01) and COI (p < 0.01) scores. After 8 days, they showed lower SI (p<0.01), higher maximal values on the thermographic Lt view (p = 0.02) and lower LOAD (p = 0.04), stiffness (p<0.01), function (p<0.01), gait (p<0.01), QOL (p<0.01) and COI (p<0.01) scores. Again at 15 days, younger patients presented lower LOAD (p<0.01), stiffness (p<0.01), function (p<0.01), gait (p<0.01), QOL (p<0.01) and COI (p<0.01) scores. At the 30 day evaluation, they again presented improved evaluations in several parameters, with lower mean and maximal values on the thermographic DV (p < 0.01 and p = 0.02, respectively) Lt view (p = 0.02, for the mean value), higher joint flexion (p = 0.01) and lower LOAD (p<0.01), stiffness (p<0.01), function (p<0.01), gait (p<0.01), QOL (p<0.01) and COI (p<0.01) scores. Better CMI scores was again observed at 90 days, specifically lower LOAD (p = 0.04), stiffness (p<0.01), function (p<0.01), gait (p<0.01), QOL (p<0.01) and COI (p < 0.01) scores. At the final evaluation, patients below the sample mean age had lower deviation and SI (p = 0.03 and p < 0.01, respectively), and stiffness (p < 0.01), function (p < 0.01), gait (p<0.01), QOL (p<0.01) and COI (p<0.01) scores.

Discussion

OA is a leading cause of disability around the world, impacting the physical and mental wellbeing of populations, posing a substantial toll on healthcare and financial resources [66]. To our knowledge, this is the first study to describe the effect of a single injection of triamcinolone hexacetonide on several clinical, imaging and laboratorial signs in a naturally occurring canine osteoarthritis model, with a long follow up period.

There are some reports evaluating the effect of IA TH in humans. A 2-year follow-up study showed that TH has long-term safety, with no deleterious effects being observed deriving from IA administration [67–69]. Also, patients treated had significant increases in ROM and improvements in pain [67]. These improvements are noticeable with the results of the Kaplan

Maier test for symmetry index, with results in SG taking significantly longer to return to baseline values. It was also observable with different scores, as function or stiffness. Comparing TH to a saline injection, TH (40mg) had higher effectiveness than the placebo group in the four weeks in terms of pain in movement, pain scale, and ultra-sound measurement of synovial hypertrophy [70]. Treatment with 20mg or 40mg of TH produced equal relapse after six months in patients with chronic polyarthritis and when treating medium-sized joints. With that in mind, since no difference in outcome was found between the compared doses, the authors advised that lower dose should be preferred, reducing pharmaceutical costs and metabolic side effects [71, 72]. As a whole, these reports present the overall safety and effectiveness of IA TH in the management of OA, measured with multiple validated CMI and other clinical evaluations. With this animal model, a single IA TH administration was able to significantly reduce weight-bearing changes in affected joints up to the 90-day evaluation compared with a control group. Besides changes in different CMI scores, particularly pain scores calculated with the CBPI, were observed. This is of particular interest, since pain is a hallmark of OA, and its characterization produces valuable data that may translate to humans [21, 73, 74]. Individual CMI scores in THG improved for a majority of animals were observed with several of the considered questionnaires, in many cases up to the last evaluation. A significant difference was also observed with the Kaplan Meier test for the majority of the considered scores. In contrast, patients in the CG had worse scores (meaning lower HVAS scores and higher values in the remaining considered CMIs scores) throughout the follow-up period, particularly with as time progressed. Our results are in line with previously described effects for TH, but since in dogs OA progresses faster while maintaining the same stages [9], it is possible that in humans results may be observed for a more extended period. Additionally, since patients who composed this sample are active working dogs, their musculoskeletal structures are under increased stress and effort [75], leading to an earlier decline in initially observed improvements. It would also be of interest to have intermediary follow-ups between the 90 and 180-day evaluations, in order to further precise the duration of treatment efficacy. Some patients in the CG also showed some improvements, particularly at the 8 and 15-day evaluations. It may be due to the natural evolution of osteoarthritis, with the possibility for spontaneous improvements in some stages of the disease. An additional possibility is related to the removal of cytokine loaded synovial fluid at the time of the evaluation and the posterior administration of saline, which can lead to an effect similar to a joint lavage. In fact, placebo saline injections have shown an effect in functional improvements that can last up to a 6-month follow-up [76].

IL-1 is commonly pointed out as the most important proinflammatory cytokine responsible for the catabolic events in OA [1, 49, 50]. Corticosteroids are, of the medications available for the treatment of OA, the ones with most potent anti-inflammatory activity, specifically through the downregulation of the synthesis of inflammatory mediators such as IL-1 β , TNF- α and COX-2 in the synovial fluid [64, 77–79]. In this study, significant differences between CG and THG were only observed at the 8-day evaluation and, even though IL-1 levels were lower in both groups compared to the initial evaluation, levels in CG were lower. This shows that TH can reduce IL-1 levels, which can be partly responsible for its ability to improve OA clinical signs, but the removal of synovial fluid and the administration of saline is also able to do so. Despite this effect, IL-1 levels cannot be the sole responsible for OA clinical signs, since, despite lower IL-1 levels of CG, this group still had worse clinical signs.

Radiographic evaluation is a staple of OA monitoring. Previous OA animal studies have demonstrated a decrease in disease progression or a protective role of corticosteroids injections, based on histological and biochemical findings [18, 80–84]. A recent systematic review of canine models of OA induction concludes that the reports regarding its IA use appear to be unanimously positive, with lower doses with sustained joint concentrations having a protective

effect [85]. In CG, the natural progression of the disease was observed as expected, and radiographic signs progressed throughout the follow-up period. In the THG, radiographic signs also progressed, particularly in the more advanced follow-up days. Although radiographic signs progressed, they seemed to be less severe in THG. We only characterized radiographic sings as present or absent, so this eventual protective role of IA TH may not be entirely recorded. The evaluation of CCO and CFHO is of particular clinical interest, as they represent early radiographic signs that predict the development of the clinical [42, 43, 86, 87]. Our results support this finding since animals with CCO or CFHO in both views showed worse clinical signs at the initial evaluations. Also, animals with these radiographic findings showed a worse response to treatment, despite THG having a positive evolution compared with CG. Digital thermography can assess inflammatory pain in osteoarthritic patients [47, 48]. Our findings showed mixed results regarding the thermographic evaluation. While, in some evaluation days, animals with higher temperatures recorded in different thermography evaluations corresponded to those patients with worse clinical signs, in other days, it did not. This may indicate that other characteristics may play an important role, e.g. the amount of muscle masses surrounding the joint, in this case, represented by thigh girth, or variations in body weight.

Documented risk factors for OA include higher bodyweight and increasing age [2]. To evaluate the effect of these factors, we considered results with a cut-off for weight and compared younger to older patients. Considering bodyweight at different cut-off points, heavier patients in both groups generally showed worse clinical signs, particularly with different CMI scores. This effect of body weight may also be responsible for the fact that females also had better CMI scores than males since they were also lighter. Similarly, patients with age above the mean sample age showed worse evaluations during the follow-up period and worse response to treatment in THG. This may reflect that older patients may have a more degenerate joint with more advanced OA-induced changes, and therefore worse clinical signs, with a reduced ability to show improvements in response to therapy.

Side effects of IA procedures are mainly related to discomfort from the procedure itself, localized pain post-injection and flushing. IA corticosteroids can also cause synovitis, in a reactive reaction called a steroid flare, with a described prevalence of 2–6% [4, 78, 88, 89]. More rare reported side effects include crystal-induced synovitis, calcification and steroid arthropathy [90]. We observed increased lameness in four patients, which spontaneously resolved within 48 hours. No additional medication was administered to the animals during the followup period. When compared to other therapeutic options, IA TH may be a more cost-effective option due to its lower cost [91].

Conclusions

To our knowledge, this is the first study to describe the effect of a single injection of triamcinolone hexacetonide in a naturally occurring canine model, with a long follow up period. THG recorded significant improvements in weight-bearing up to the 90-day follow-up. Improvements were also observed with the considered CMIs, particularly pain scores. Lower thermographic values were registered in THG up to the last evaluation day. Age, sex, and radiographic findings did significantly influenced response to treatment.

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References

- 1. Loeser RF, Goldring SR, Scanzello CR, Goldring MB. Osteoarthritis: A disease of the joint as an organ. Arthritis Rheum. 2012; 64: 1697–1707. https://doi.org/10.1002/art.34453 PMID: 22392533
- Anderson KL, O'Neill DG, Brodbelt DC, Church DB, Meeson RL, Sargan D, et al. Prevalence, duration and risk factors for appendicular osteoarthritis in a UK dog population under primary veterinary care. Sci Rep. 2018; 8: 5641. https://doi.org/10.1038/s41598-018-23940-z PMID: 29618832
- Cuervo B, Chicharro D, Del Romero A, Damia E, Carrillo J, Sopena J, et al. Objective and subjective evaluation of plasma rich in growth factors therapy for the treatment of osteoarthritis in dogs. Osteoarthr Cartil. 2019; 27: S482. https://doi.org/10.1016/j.joca.2019.02.532
- Edwards SHR. Intra-articular drug delivery: The challenge to extend drug residence time within the joint. Vet J. 2011; 190: 15–21. https://doi.org/10.1016/j.tvjl.2010.09.019 PMID: 20947396
- Larsen C, Østergaard J, Larsen SW, Jensen H, Jacobsen S, Lindegaard C, et al. Intra-articular depot formulation principles: Role in the management of postoperative pain and arthritic disorders. J Pharm Sci. 2008; 97: 4622–4654. https://doi.org/10.1002/jps.21346 PMID: 18306275
- Gregory MH, Capito N, Kuroki K, Stoker AM, Cook JL, Sherman SL. A Review of Translational Animal Models for Knee Osteoarthritis. Arthritis. 2012; 2012: 1–14. <u>https://doi.org/10.1155/2012/764621</u> PMID: 23326663
- McCoy AM. Animal Models of Osteoarthritis: Comparisons and Key Considerations. Vet Pathol. 2015; 52: 803–818. https://doi.org/10.1177/0300985815588611 PMID: 26063173
- Kol A, Arzi B, Athanasiou KA, Farmer DL, Nolta JA, Rebhun RB, et al. Companion animals: Translational scientist's new best friends. Sci Transl Med. 2015; 7: 308ps21–308ps21. <u>https://doi.org/10.1126/ scitranslmed.aaa9116</u> PMID: 26446953
- Meeson RL, Todhunter RJ, Blunn G, Nuki G, Pitsillides AA. Spontaneous dog osteoarthritis—a One Medicine vision. Nat Rev Rheumatol. 2019. <u>https://doi.org/10.1038/s41584-019-0202-1</u> PMID: 30953036
- Pascual-Garrido C, Guilak F, Rai MF, Harris MD, Lopez MJ, Todhunter RJ, et al. Canine hip dysplasia: A natural animal model for human developmental dysplasia of the hip. J Orthop Res. 2018; 36: 1807– 1817. https://doi.org/10.1002/jor.23828 PMID: 29227567
- Liu W, Burton-Wurster N, Glant TT, Tashman S, Sumner DR, Kamath R V., et al. Spontaneous and experimental osteoarthritis in dog: Similarities and differences in proteoglycan levels. J Orthop Res. 2003; 21: 730–737. https://doi.org/10.1016/S0736-0266(03)00002-0 PMID: 12798075
- Minnema L, Wheeler J, Enomoto M, Pitake S, Mishra SK, Lascelles BDX. Correlation of Artemin and GFRα3 With Osteoarthritis Pain: Early Evidence From Naturally Occurring Osteoarthritis-Associated Chronic Pain in Dogs. Front Neurosci. 2020; 14. https://doi.org/10.3389/fnins.2020.00077 PMID: 32116521

- Céleste C, Ionescu M, Poole AR, Laverty S. Repeated intraarticular injections of triamcinolone acetonide alter cartilage matrix metabolism measured by biomarkers in synovial fluid. J Orthop Res. 2005; 23: 602–610. https://doi.org/10.1016/j.orthres.2004.10.003 PMID: 15885481
- Garg N, Perry L, Deodhar A. Intra-articular and soft tissue injections, a systematic review of relative efficacy of various corticosteroids. Clin Rheumatol. 2014; 33: 1695–1706. <u>https://doi.org/10.1007/s10067-014-2572-8 PMID: 24651914</u>
- Cheng OT, Souzdalnitski D, Vrooman B, Cheng J. Evidence-Based Knee Injections for the Management of Arthritis. Pain Med. 2012; 13: 740–753. https://doi.org/10.1111/j.1526-4637.2012.01394.x PMID: 22621287
- Zhang W, Moskowitz RW, Nuki G, Abramson S, Altman RD, Arden N, et al. OARSI recommendations for the management of hip and knee osteoarthritis, Part II: OARSI evidence-based, expert consensus guidelines. Osteoarthr Cartil. 2008; 16: 137–162. <u>https://doi.org/10.1016/j.joca.2007.12.013</u> PMID: 18279766
- Park KD, Kim TK, Bae BW, Ahn J, Lee WY, Park Y. Ultrasound guided intra-articular ketorolac versus corticosteroid injection in osteoarthritis of the hip: a retrospective comparative study. Skeletal Radiol. 2015; 44: 1333–1340. https://doi.org/10.1007/s00256-015-2174-9 PMID: 26031217
- Pelletier J-P, Martel-Pelletier J. Protective effects of corticosteroids on cartilage lesions and osteophyte formation in the pond-nuki dog model of osteoarthritis. Arthritis Rheum. 1989; 32: 181–193. https://doi. org/10.1002/anr.1780320211 PMID: 2920053
- Pelletier J, DiBattista J, Raynauld J, Wilhelm S, Martel-Pelletier J. The in vivo effects of intraarticular corticosteroid injections on cartilage lesions, stromelysin, interleukin-1, and oncogene protein synthesis in experimental osteoarthritis. Lab Invest. 1995; 72: 578–86. Available: http://www.ncbi.nlm.nih.gov/ pubmed/7745952 PMID: 7745952
- Rocha RH, Natour J, dos Santos RM, Furtado RNV. Time Effect of Intra-articular Injection With Triamcinolone Hexacetonide and Its Correlations. Am J Phys Med Rehabil. 2019; 98: 872–878. <u>https://doi.org/10.1097/PHM.00000000001217 PMID: 31584880</u>
- Strasser T, Peham C, Bockstahler BA, Turmezei TD, Treece GM, Gee AH, et al. Identification of quantitative trait loci for osteoarthritis of hip joints in dogs. Am J Vet Res. 2016; 52: 369–77. https://doi.org/10. 2460/ajvr.69.10.1294 PMID: 18828685
- Piel MJ, Kroin JS, Van Wijnen AJ, Kc R, Im HJ. Pain assessment in animal models of osteoarthritis. Gene. 2014; 537: 184–188. https://doi.org/10.1016/j.gene.2013.11.091 PMID: 24333346
- 23. Reid J, Nolan AM, Scott EM. Measuring pain in dogs and cats using structured behavioural observation. Vet J. 2018; 236: 72–79. https://doi.org/10.1016/j.tvjl.2018.04.013 PMID: 29871754
- Stadig S, Lascelles BDX, Nyman G, Bergh A. Evaluation and comparison of pain questionnaires for clinical screening of osteoarthritis in cats. Vet Rec. 2019; 185: 757–757. <u>https://doi.org/10.1136/vr.105115</u> PMID: 31619513
- Gruen ME, Griffith EH, Thomson AE, Simpson W, Lascelles BDX. Criterion Validation Testing of Clinical Metrology Instruments for Measuring Degenerative Joint Disease Associated Mobility Impairment in Cats. Thamm D, editor. PLoS One. 2015; 10: e0131839. https://doi.org/10.1371/journal.pone.0131839 PMID: 26162101
- Lascelles BDX, Brown DC, Maixner W, Mogil JS. Spontaneous painful disease in companion animals can facilitate the development of chronic pain therapies for humans. Osteoarthr Cartil. 2018; 26: 175– 183. https://doi.org/10.1016/j.joca.2017.11.011 PMID: 29180098
- Hercock CA, Pinchbeck G, Giejda A, Clegg PD, Innes JF. Validation of a client-based clinical metrology instrument for the evaluation of canine elbow osteoarthritis. J Small Anim Pract. 2009; 50: 266–271. https://doi.org/10.1111/j.1748-5827.2009.00765.x PMID: 19527419
- Walton MB, Cowderoy E, Lascelles D, Innes JF. Evaluation of Construct and Criterion Validity for the 'Liverpool Osteoarthritis in Dogs' (LOAD) Clinical Metrology Instrument and Comparison to Two Other Instruments. Wade C, editor. PLoS One. 2013; 8: e58125. <u>https://doi.org/10.1371/journal.pone.</u> 0058125 PMID: 23505459
- 29. Walton B, Cox T, Innes J. 'How do I know my animal got better?'-measuring outcomes in small animal orthopaedics. In Pract. 2018; 40: 42–50. https://doi.org/10.1136/inp.k647
- Brown DC. The Canine Orthopedic Index. Step 2: Psychometric Testing. Vet Surg. 2014; 43: 241–246. https://doi.org/10.1111/j.1532-950X.2014.12141.x PMID: 24512284
- Hudson JT, Slater MR, Taylor L, Scott HM, Kerwin SC. Assessing repeatability and validity of a visual analogue scale questionnaire for use in assessing pain and lameness in dogs. Am J Vet Res. 2004; 65: 1634–1643. https://doi.org/10.2460/ajvr.2004.65.1634 PMID: 15631027
- 32. Clough W, Canapp S. Assessing Clinical Relevance of Weight Distribution as Measured on a Stance Analyzer through Comparison with Lameness Determined on a Pressure Sensitive Walkway and

Clinical Diagnosis. Vet Comp Orthop Traumatol. 2018; 31: A1–A25. https://doi.org/10.1055/s-0038-1667359 PMID: 30060271

- Tudor-Locke C, Williams JE, Reis JP, Pluto D. Utility of Pedometers for Assessing Physical Activity. Sport Med. 2002; 32: 795–808. <u>https://doi.org/10.2165/00007256-200232120-00004</u> PMID: 12238942
- Wilson L, Smith B. Canine lameness. 2nd ed. In: McGowan CM, Goff L, editors. Animal Physiotherapy: Assessment, Treatment and Rehabilitation of Animals. 2nd ed. Wiley Blackwell; 2016. pp. 112– 126.
- Lotsikas P, Lotsikas F, D. H, Dyce J, Ridge P. Disorders of the Pelvic Limb: Diagnosis and Treatment. 2nd ed. In: Zink C, J. van D, editors. Canine Sports Medicine and Rehabilitation. 2nd ed. Wiley Blackwell; 2016. pp. 353–388.
- Hyytiäinen HK, Mölsä SH, Junnila JT, Laitinen-Vapaavuori OM, Hielm-Björkman AK. Ranking of physiotherapeutic evaluation methods as outcome measures of stifle functionality in dogs. Acta Vet Scand. 2013; 55: 29. https://doi.org/10.1186/1751-0147-55-29 PMID: 23566355
- Henderson AL, Hecht S, Millis DL. Lumbar paraspinal muscle transverse area and symmetry in dogs with and without degenerative lumbosacral stenosis. J Small Anim Pract. 2015; 56: 618–622. <u>https://</u> doi.org/10.1111/jsap.12385 PMID: 26310387
- Turmezei TD, Treece GM, Gee AH, Houlden R, Poole KES. A new quantitative 3D approach to imaging of structural joint disease. Sci Rep. 2018; 8: 1–13. https://doi.org/10.1038/s41598-017-17765-5 PMID: 29311619
- Lafeber FPJG, van Spil WE. Osteoarthritis year 2013 in review: Biomarkers; reflecting before moving forward, one step at a time. Osteoarthr Cartil. 2013; 21: 1452–1464. <u>https://doi.org/10.1016/j.joca.2013</u>. 08.012 PMID: 23954702
- 40. Gordon WJ, Conzemius MG, Riedesel E, Besancon MF, Evans R, Wilke V, et al. The relationship between limb function and radiographic osteoarthrosis in dogs with stifle osteoarthrosis. Vet Surg. 2003; 32: 451–454. https://doi.org/10.1053/jvet.2003.50051 PMID: 14569573
- Bauer DC, Hunter DJ, Abramson SB, Attur M, Corr M, Felson D, et al. Classification of osteoarthritis biomarkers: a proposed approach. Osteoarthr Cartil. 2006; 14: 723–727. <u>https://doi.org/10.1016/j.joca.</u> 2006.04.001 PMID: 16733093
- 42. Mayhew PD, McKelvie PJ, Biery DN, Shofer FS, Smith GK. Evaluation of a radiographic caudolateral curvilinear osteophyte on the femoral neck and its relationship to degenerative joint disease and distraction index in dogs. J Am Vet Med Assoc. 2002; 220: 472–6. Available: <u>http://www.ncbi.nlm.nih.gov/pubmed/11860241</u>
- Puckler K, Tellhelm B, Kirberger R. The hip joint and pelvis. In: Kirberger R, McEvoy F, editors. BSAVA Manual of Canine and Feline Musculoskeletal Imaging. Wiley; 2016. pp. 212–231.
- Ring EFJ. The historical development of thermal imaging in medicine. Rheumatology. 2004; 43: 800– 802. https://doi.org/10.1093/rheumatology/keg009 PMID: 15163833
- Uematsu S, Edwin DH, Jankel WR, Kozikowski J, Trattner M. Quantification of thermal asymmetry. J Neurosurg. 1988; 69: 552–555. https://doi.org/10.3171/jns.1988.69.4.0552 PMID: 3418388
- Jin C. Automated Analysis Method for Screening Knee Osteoarthritis using Medical Infrared Thermography. J Med Biol Eng. 2013; 33: 471. https://doi.org/10.5405/jmbe.1054
- 47. Borojevic N, Darko K, Grazio S, Grubisic F, Antonini S, Nola IA, et al. thermography of rheumatoid arthritis and osteoarthritis. Period Biol. 2011; 113: 445–448.
- **48.** Fokam D, Lehmann C. Clinical assessment of arthritic knee pain by infrared thermography. J Basic Clin Physiol Pharmacol. 2019; 30. https://doi.org/10.1515/jbcpp-2017-0218 PMID: 30375348
- McIlwraith C. Traumatic Arthritis and Posttraumatic Osteoarthritis in the Horse. 2nd ed. In: McIlwraith C, editor. Joint Disease in the Horse. 2nd ed. Elsevier; 2016. pp. 33–56.
- Fujita Y, Hara Y, Nezu Y, Schulz KS, Tagawa M. Proinflammatory cytokine activities, matrix metalloproteinase-3 activity, and sulfated glycosaminoglycan content in synovial fluid of dogs with naturally acquired cranial cruciate ligament rupture. Vet Surg. 2006; 35: 369–376. https://doi.org/10.1111/j.1532-950X.2006.00159.x PMID: 16756618
- Vincent TL. IL-1 in osteoarthritis: time for a critical review of the literature. F1000Research. 2019; 8: 934. https://doi.org/10.12688/f1000research.18831.1 PMID: 31249675
- Bennett D, Eckersall PD, Waterston M, Marchetti V, Rota A, Mcculloch E, et al. The effect of robenacoxib on the concentration of C-reactive protein in synovial fluid from dogs with osteoarthritis. BMC Vet Res. 2013; 9. https://doi.org/10.1186/1746-6148-9-42 PMID: 23452411
- Eckersall PD, Conner JG. Bovine and canine acute phase proteins. Vet Res Commun. 1988; 12: 169– 178. https://doi.org/10.1007/BF00362798 PMID: 2460991

- Sowers M, Jannausch M, Stein E, Jamadar D, Hochberg M, Lachance L. C-reactive protein as a biomarker of emergent osteoarthritis. Osteoarthr Cartil. 2002; 10: 595–601. <u>https://doi.org/10.1053/joca.</u> 2002.0800 PMID: 12479380
- Smith G, Karbe G, Agnello K, McDonald-Lynch M. Pathogenesis, Diagnosis, and Control of Canine Hip Dysplasia. 1st ed. In: Tobias K, Johnston S, editors. Veterinary Surgery: Small Animal. 1st ed. Saunders; 2011. pp. 824–848.
- Fortrie RR, Verhoeven G, Broeckx B, Duchateau L, Janssens L, Samoy Y, et al. Intra- and Interobserver Agreement on Radiographic Phenotype in the Diagnosis of Canine Hip Dysplasia. Vet Surg. 2015; 44: 467–473. https://doi.org/10.1111/j.1532-950X.2014.12309.x PMID: 25414132
- Van Vynckt D, Samoy Y, Mosselmans L, Verhoeven G, Verschooten F, Van Ryssen B. The use of intraarticular anesthesia as a diagnostic tool in canine lameness. Vlaams Diergeneeskd Tijdschr. 2012; 81: 290–297.
- Volstad N, Sandberg G, Robb S, Budsberg S. The evaluation of limb symmetry indices using ground reaction forces collected with one or two force plates in healthy dogs. Vet Comp Orthop Traumatol. 2017; 30: 54–58. https://doi.org/10.3415/VCOT-16-04-0054 PMID: 27849103
- Clough W, Canapp S, Taboada L, Dycus D, Leasure C. Sensitivity and Specificity of a Weight Distribution Platform for the Detection of Objective Lameness and Orthopaedic Disease. Vet Comp Orthop Traumatol. 2018; 31: 391–395. https://doi.org/10.1055/s-0038-1667063 PMID: 30300913
- Chan CB, Spierenburg M, Ihle SL, Tudor-Locke C. Use of pedometers to measure physical activity in dogs. J Am Vet Med Assoc. 2005; 226: 2010–5. Available: http://www.ncbi.nlm.nih.gov/pubmed/ 15989183
- Vainionpää M, Raekallio M, Tuhkalainen E, Hänninen H, Alhopuro N, Savolainen M, et al. Comparison of three thermal cameras with canine hip area thermographic images. J Vet Med Sci. 2012; 74: 1539– 44. Available: http://www.ncbi.nlm.nih.gov/pubmed/22785576
- McCarthy DA, Millis DL, Levine D, Weigel JP. Variables Affecting Thigh Girth Measurement and Observer Reliability in Dogs. Front Vet Sci. 2018; 5. https://doi.org/10.3389/fvets.2018.00203 PMID: 30214905
- 63. Levine, D., Millis DL. Canine Rehabilitation and Physical Therapy. 2014.
- Caron JP. Intra-Articular Injections for Joint Disease in Horses. Vet Clin North Am Equine Pract. 2005; 21: 559–573. https://doi.org/10.1016/j.cveq.2005.07.003 PMID: 16297721
- Chakravarty K, Pharoah PDP, Scott DGI. A randomized controlled study of post-injection rest following intra-articular steroid therapy for knee synovitis. Rheumatology. 1994; 33: 464–468. https://doi.org/10. 1093/rheumatology/33.5.464 PMID: 8173852
- Vina ER, Kwoh CK. Epidemiology of osteoarthritis. Curr Opin Rheumatol. 2018; 30: 160–167. https:// doi.org/10.1097/BOR.0000000000479 PMID: 29227353
- Raynauld J-P, Buckland-Wright C, Ward R, Choquette D, Haraoui B, Martel-Pelletier J, et al. Safety and efficacy of long-term intraarticular steroid injections in osteoarthritis of the knee: A randomized, doubleblind, placebo-controlled trial. Arthritis Rheum. 2003; 48: 370–377. <u>https://doi.org/10.1002/art.10777</u> PMID: 12571845
- Spolidoro Paschoal N d. O, Natour J, Machado FS, de Oliveira HA V., Furtado. Effectiveness of Triamcinolone Hexacetonide Intraarticular Injection in Interphalangeal Joints: A 12-week Randomized Controlled Trial in Patients with Hand Osteoarthritis. J Rheumatol. 2015; 42: 1869–1877. https://doi.org/10. 3899/jrheum.140736 PMID: 26233501
- Meenagh GK. A randomized controlled trial of intra-articular corticosteroid injection of the carpometacarpal joint of the thumb in osteoarthritis. Ann Rheum Dis. 2004; 63: 1260–1263. https://doi.org/10. 1136/ard.2003.015438 PMID: 15361383
- Mendes JG, Natour J, Nunes-Tamashiro JC, Toffolo SR, Rosenfeld A, Furtado RNV. Comparison between intra-articular Botulinum toxin type A, corticosteroid, and saline in knee osteoarthritis: a randomized controlled trial. Clin Rehabil. 2019; 33: 1015–1026. <u>https://doi.org/10.1177/</u> 0269215519827996 PMID: 30782000
- Weitoft T, Öberg K. Dosing of intra-articular triamcinolone hexacetonide for knee synovitis in chronic polyarthritis: a randomized controlled study. Scand J Rheumatol. 2019; 48: 279–283. <u>https://doi.org/10. 1080/03009742.2019.1571222</u> PMID: 30843453
- 72. Cushman DM, Ofek E, Syed RH, Clements N, Gardner JE, Sams JM, et al. Comparison of Varying Corticosteroid Type, Dose, and Volume for the Treatment of Pain in Small- and Intermediate-Size Joint Injections: A Narrative Review. PM&R. 2019; 11: 758–770. <u>https://doi.org/10.1016/j.pmrj.2018.09.040</u> PMID: 31166662
- 73. Wiegant K, Intema F, van Roermund PM, Barten-van Rijbroek AD, Doornebal A, Hazewinkel HAW, et al. Evidence of Cartilage Repair by Joint Distraction in a Canine Model of Osteoarthritis. Arthritis Rheumatol. 2015; 67: 465–474. https://doi.org/10.1002/art.38906 PMID: 25303046

- 74. Robertson-Plouch C, Stille JR, Liu P, Smith C, Brown D, Warner M, et al. A randomized clinical efficacy study targeting mPGES1 or EP4 in dogs with spontaneous osteoarthritis. Sci Transl Med. 2019; 11: eaaw9993. https://doi.org/10.1126/scitranslmed.aaw9993 PMID: 31666405
- Alves JC, Santos A, Fernandes Â. Evaluation of the effect of mesotherapy in the management of back pain in police working dogs. Vet Anaesth Analg. 2018; 45: 123–128. https://doi.org/10.1016/j.vaa.2017. 07.006 PMID: 29222031
- 76. Previtali D, Merli G, Di Laura Frattura G, Candrian C, Zaffagnini S, Filardo G. The Long-Lasting Effects of "Placebo Injections" in Knee Osteoarthritis: A Meta-Analysis. Cartilage. 2020; 194760352090659. https://doi.org/10.1177/1947603520906597 PMID: 32186401
- 77. Sellam J, Berenbaum F. The role of synovitis in pathophysiology and clinical symptoms of osteoarthritis. Nat Rev Rheumatol. 2010; 6: 625–635. https://doi.org/10.1038/nrrheum.2010.159 PMID: 20924410
- Lavelle W, Lavelle ED, Lavelle L. Intra-Articular Injections. Anesthesiol Clin. 2007; 25: 853–862. https:// doi.org/10.1016/j.anclin.2007.07.002 PMID: 18054149
- 79. Vaishya R, Pandit R, Agarwal AK, Vijay V. Intra-articular hyaluronic acid is superior to steroids in knee osteoarthritis: A comparative, randomized study. J Clin Orthop Trauma. 2017; 8: 85–88. https://doi.org/ 10.1016/j.jcot.2016.09.008 PMID: 28360505
- Kumar A, Bendele AM, Blanks RC, Bodick N. Sustained efficacy of a single intra-articular dose of FX006 in a rat model of repeated localized knee arthritis. Osteoarthr Cartil. 2015; 23: 151–160. https:// doi.org/10.1016/j.joca.2014.09.019 PMID: 25266960
- Frisbie DD, Kawcak CE, Trotter GW, Powers BE, Walton RM, McIlwraith CW. Effects of triamcinolone acetonide on an in vivo equine osteochondral fragment exercise model. Equine Vet J. 1997; 29: 349– 359. https://doi.org/10.1111/j.2042-3306.1997.tb03138.x PMID: 9306060
- Augustine AJ, Oleksyszyn J. Glucocorticosteroids inhibit degradation in bovine cartilage explants stimulated with concomitant plasminogen and interleukin-1<alpha>. Inflamm Res. 1997; 46: 60–64. https://doi.org/10.1007/s000110050073 PMID: 9085145
- Pelletier JP, Martel-Pelletier J. In vivo protective effects of prophylactic treatment with tiaprofenic acid or intraarticular corticosteroids on osteoarthritic lesions in the experimental dog model. J Rheumatol Suppl. 1991; 27: 127–30. Available: http://www.ncbi.nlm.nih.gov/pubmed/2027112 PMID: 2027112
- Sieker JT, Ayturk UM, Proffen BL, Weissenberger MH, Kiapour AM, Murray MM. Immediate Administration of Intraarticular Triamcinolone Acetonide After Joint Injury Modulates Molecular Outcomes Associated With Early Synovitis. Arthritis Rheumatol. 2016; 68: 1637–1647. <u>https://doi.org/10.1002/art.39631</u> PMID: 26866935
- **85.** Vandeweerd J-M, Zhao Y, Nisolle J-F, Zhang W, Zhihong L, Clegg P, et al. Effect of corticosteroids on articular cartilage: have animal studies said everything? Fundam Clin Pharmacol. 2015; 29: 427–438. https://doi.org/10.1111/fcp.12137 PMID: 26211421
- 86. Powers MY, Biery DN, Lawler DE, Evans RH, Shofer FS, Mayhew P, et al. use of the caudolateral curvilinear osteophyte as an early marker for future development of osteoarthritis associated with hip dysplasia in dogs. J Am Vet Med Assoc. 2004; 225: 233–7. Available: <u>http://www.ncbi.nlm.nih.gov/pubmed/</u> 15323379
- Tôrres RCS, Ferreira PM, Araújo RB, Martins AS. Presença de "Linha Morgan" como indicador de displasia coxofemoral em cães da raça Pastor-Alemão. Arq Bras Med Veterinária e Zootec. 1999; 51: 157–158. https://doi.org/10.1590/S0102-09351999000200006
- Waddell DD. Viscosupplementation with Hyaluronans for Osteoarthritis of the Knee Clinical Efficacy and Economic Implications. Drugs Aging. 2007; 24: 629–642. https://doi.org/10.2165/00002512-200724080-00002 PMID: 17702533
- Popma JW, Snel FW, Haagsma CJ, Brummelhuis-Visser P, Oldenhof HGJ, van der Palen J, et al. Comparison of 2 Dosages of Intraarticular Triamcinolone for the Treatment of Knee Arthritis: Results of a 12-week Randomized Controlled Clinical Trial. J Rheumatol. 2015; 42: 1865–1868. <u>https://doi.org/10.3899/jrheum.141630 PMID: 26233499</u>
- Bellamy N, Campbell J, Robinson V, Gee T, Bourne R, Wells G. Intraarticular corticosteroid for treatment of osteoarthritis of the knee. In: Bellamy N, editor. The Cochrane Database of Systematic Reviews. Chichester, UK: John Wiley & Sons, Ltd; 2005. https://doi.org/10.1002/14651858.CD005328 PMID: 15846755
- Tammachote N, Kanitnate S, Yakumpor T, Panichkul P. Intra-Articular, Single-Shot Hylan G-F 20 Hyaluronic Acid Injection Compared with Corticosteroid in Knee Osteoarthritis. J Bone Jt Surg. 2016; 98: 885–892. https://doi.org/10.2106/JBJS.15.00544 PMID: 27252432