



Case report

Roseomonas mucosa bacteremia in a neutropenic child: A case report and literature review

Keigo Kimura^a, Hideharu Hagiya^{b,*}, Isao Nishi^a, Hisao Yoshida^{b,c}, Kazunori Tomono^b

^a Laboratory for Clinical Investigation, Osaka University Hospital, Japan

^b Division of Infection Control and Prevention, Osaka University Hospital, Japan

^c Department of Pediatrics, Osaka University Hospital, Japan

ARTICLE INFO

Article history:

Received 15 August 2018

Received in revised form 9 November 2018

Accepted 9 November 2018

Keywords:

Bloodstream infection

Febrile neutropenia

Roseomonas mucosa

ABSTRACT

Roseomonas species is rarely found to be pathogenic to humans and there are few clinical cases that have been described in the literature. We report a case of *Roseomonas mucosa* bacteremia that involved a 9-year-old Japanese boy who was in a condition of febrile neutropenia caused by chemotherapy for cerebellar medulloblastoma. Conventional phenotyping failed to identify the organism; however, genetic analysis using 16S rDNA sequencing confirmed the pathogen to be *R. mucosa*. The patient recovered following treatment by meropenem without any complications. A literature review of pediatric cases of *Roseomonas* bacteremia identified 12 other documented cases, and these revealed that a common clinical situation for the infection is an immunocompromised state with malignancy and/or an indwelling intravenous catheter. Because of the low number of cases, the overall picture of *Roseomonas* bacteremia in children remains to be elucidated; however, the prognosis of the infection appears to be satisfactory. © 2018 Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Introduction

Members of the genus *Roseomonas*, which was first reported by Rihs et al. in 1993 [1], are slow-growing, aerobic, non-fermentative Gram-negative bacteria, which appear as pink-pigmented colonies. More than 20 *Roseomonas* species have been isolated from environmental samples, including water, soil, and plants [2–5]. These species are opportunistic pathogens with low pathogenicity to humans; however, the occurrence of human infections has increasingly been reported over the last two decades, predominantly in immunocompromised patients [6–8]. These organisms frequently cause central line-associated bloodstream infections, but potentially give rise to respiratory, skin and soft tissue, peritoneal, and urinary tract infections, as well as spondylitis and subretinal abscesses [9–11].

The major human pathogens of *Roseomonas* species are *Roseomonas gilardii* subsp. *gilardii*, *Roseomonas gilardii* subsp. *rosea*, and *Roseomonas mucosa* [2,8,12]. Of these, *R. mucosa* appears to be the most frequently identified in clinical samples [8] and has the ability to cause infections in immunocompetent patients [8,13] and even life-threatening diseases [14]. However, the clinical

picture of the infection caused by the pathogen remains to be fully elucidated because of the paucity of reported cases. Here, we report a case of *R. mucosa* bacteremia, with a literature review of pediatric cases of bacteremia caused by *Roseomonas* species.

Case

A 9-year-old Japanese boy (body weight, 27 kg) diagnosed with cerebellar medulloblastoma was admitted to our hospital. He had undergone cranial surgery for tumor resection and had subsequently undergone monthly cancer chemotherapy with a combination of cisplatin, cyclophosphamide, and vincristine. For the infusion of anticancer drugs, an indwelling central line catheter was placed. Three days subsequent to the third course, the patient developed febrile neutropenia, for which meropenem (2.7 g per day) was empirically initiated. His vital signs remained stable, but high fever accompanying diarrhea persisted. Whole-body tomography did not reveal any abnormal findings. Four days later, the patient's laboratory findings revealed Grade 4 neutropenia (40 cells/ μ L), a mild elevation in C-reactive protein levels (2.77 mg/dL), and decreased serum gamma globulin levels (immunoglobulin G, 493 mg/dL). Following the collection of blood through the central line for culture, teicoplanin was administered. In addition, his stool specimen was positive for *Clostridium difficile* toxins, and treatment with intravenous metronidazole was initiated.

* Corresponding author: Division of Infection Control and Prevention, Osaka University Hospital, 2-15 Yamadaoka, Suita, Osaka, 565-0871, Japan.

E-mail address: highgear@hp-infect.med.osaka-u.ac.jp (H. Hagiya).

Blood was collected into in BD BACTEC™ Peds Plus/F culture vials (Becton Dickinson, Sparks, MD, USA) and was incubated in a BD BACTEC™ FX blood culture system (Becton Dickinson). After 48 h at 35 °C, there was a positive signal from the automated blood culture apparatus; however, no organisms were detected under microscopic examination. The fluid was centrifuged to collect bacteria, and Gram-negative plump coccoid rods were successfully confirmed (Fig. 1A). These were subcultured on trypticase soy agar with 5% sheep blood (Becton Dickinson) and on chocolate agar (Kyokuto Pharmaceutical Industrial, Tokyo, Japan) in 5% CO₂ at 35 °C. Over subsequent days, colonies on the chocolate agar demonstrated a pink color (Fig. 1B). The organism, which was positive for catalase, oxidase, and urease in biochemical identification tests, could not be identified using Neg Combo Panel NNFC1J (Beckman Coulter, Brea, CA, USA).

Full-base 16S rDNA polymerase chain reaction analysis was conducted and 1,411 base pairs of the targeted gene were amplified using the universal primers 8UA (5'-AGA GTT TGA TCM TGG CTC AG-3') and 1485B (5'-TAC GGT TAC CTT GTT ACG AC-3'). The sequence data were analyzed using BLAST sequence homology search programs (GenBank, EzBioCloud, and leBIBI), and the pathogen was finally confirmed to correspond with the type strain of *R. mucosa* (ATCC BAA-692; accession number, AF538712) with an accuracy of 100%. The organism was also identified as *R. mucosa* by matrix-assisted laser desorption/ionization time-of-flight mass spectrometry using a MALDI Biotyper (Bruker Daltonics, Bremen, Germany), with a score of 2.378. The isolate showed resistance to piperacillin, aztreonam, and ceftazidime, and was susceptible to carbapenems, aminoglycosides, minocycline, fluoroquinolones, and trimethoprim/sulfamethoxazole (Table 1).

The patient gradually recovered from the neutropenia, and a subsequent blood culture examination was negative. He was treated with meropenem for 2 weeks with a satisfactory clinical course.

Discussion

Because of their low pathogenicity, *Roseomonas* species rarely cause infections in humans; thus, its clinical features are not yet fully understood. To elucidate the clinical characteristics of *Roseomonas* bacteremia in children, we reviewed previous literature (Table 2) [8,9,14–18]. A search of MEDLINE from its inception in 1996 revealed only 12 such cases. Of the 13 cases, including our patient, the majority of cases had any underlying diseases (11 cases, 84.6%); almost half of the cases had malignancies (6 cases, 46.2%) and neutropenia was observed in at least three cases (23.1%). Only one child was free from any underlying disease [8]. Since the characterization of *R. mucosa* in 2003 [12], this species

Table 1

Antimicrobial susceptibility testing of the pathogen after 48-hour incubation in ambient air at 35 °C.

	MIC (μg/mL)	Susceptibility
Piperacillin	>64	R
Ceftazidime	>16	R
Cefepime	16	I
Imipenem	≤1	S
Meropenem	≤1	S
Aztreonam	>16	R
Gentamicin	≤2	S
Tobramycin	≤2	S
Amikacin	≤8	S
Minocycline	≤2	S
Ciprofloxacin	0.5	S
Levofloxacin	≤0.5	S
Trimethoprim/sulphamethoxazole	≤2	S

MIC, minimum inhibitory concentration. Antimicrobial susceptibility was interpreted on the basis of Clinical & Laboratory Standards Institute Guideline (M100-S27, other non-*Enterobacteriaceae*).

has accounted for the majority of clinical cases (6/8 cases, 75%). According to a 16S rDNA-based study in Taiwan, *R. mucosa* was the most prevalent strain among various *Roseomonas* species [8]. Central lines appear to be common infectious sites for *Roseomonas* bacteremia in children, as has been described in adult cases [7–9,16]. A recent investigation found that opportunistic infections due to *R. mucosa* are associated with skin microbiota rather than the environment [2]. Considering these findings, *Roseomonas* bacteremia may occur as a result of the direct invasion of the pathogen through a catheter penetration site, particularly when the patient's neutrophil immunity has decreased. All of the pediatric patients were reported to be recovered from the infection, suggesting a preferable prognosis for cases of *Roseomonas* bacteremia.

The choice of treatment for *Roseomonas* species is difficult as there is no standard laboratory method at present, and drug susceptibility varies between species. Of note, the majority of previously reported isolates were not susceptible to third- or fourth-generation cephalosporins [12], as was the case in the patient described here. Among the *Roseomonas* species, *R. mucosa*, the most frequent isolate in clinical settings, exhibits the highest antimicrobial resistance [8,12]. The result of whole-genome sequencing for a clinical isolate of *R. mucosa* in a previous study indicated that the organism possesses innate antimicrobial-resistant characteristics [19]. However, *Roseomonas* species usually show *in vitro* susceptibility to other antimicrobial classes, such as carbapenems, aminoglycosides, and fluoroquinolones, and the clinical outcome with these antimicrobials has been reported to be satisfactory in adult cases [8].

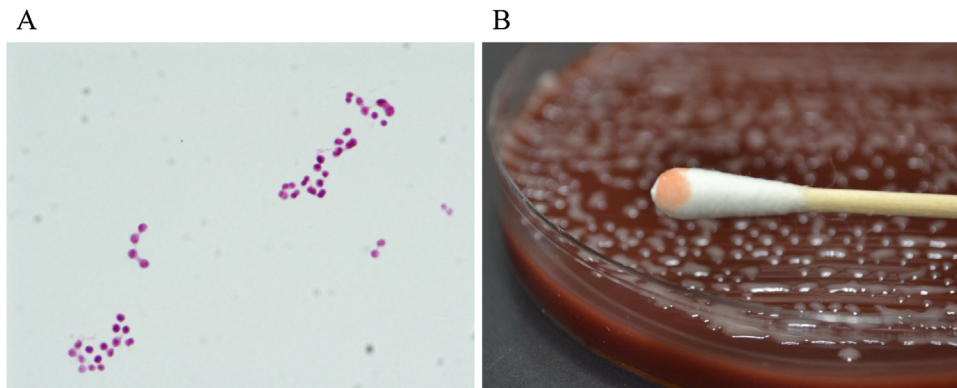


Fig. 1. Gram staining of blood culture fluid (A) and pink-pigmented colonies on a chocolate agar plate (B). A, stained following centrifugation. B, incubated in 5% CO₂ at 35 °C for 2 days.

Table 2
Summary of *Roseomonas* species bacteremia in pediatric cases.

No	Age	Sex	Underlying diseases	Neutropenia at the onset	Species	Infectious sites	Treatment	Prognosis	year	Ref
1	1m	n.	Premature	n.d.	<i>R. gilardii</i>	n.d.	n.d.	n.d.	1996	9
2	2m	n.	none	n.d.	<i>Roseomonas</i> (not identified)	n.d.	n.d.	Transient Colonization	1996	9
3	6y	F	Cystic fibrosis	n.d.	<i>Roseomonas</i> (not identified)	n.d.	n.d.	Transient Colonization	1996	9
4	15y	F	ALL	<100 / μ L	<i>Roseomonas</i> (not identified)	n.d.	GM, CAZ	"Response was positive"	1996	9
5	2y	M	ALL	No	<i>R. gilardii</i>	CVC	AMK, CAZ	Recover	2001	15
6	11m	M	ALL	No	<i>R. gilardii</i>	CVC	GM, CAZ	Recover	2006	16
7	18y	M	TPN dependence	n.d.	<i>R. mucosa</i>	CVC	CPFX	Recover	2010	14
8	8m	M	Tethered cord syndrome with dermal tract	No	<i>R. mucosa</i>	Soft tissue infection	CTX	Recover	2012	8
9	1y	F	none	No	<i>Roseomonas</i> genomospecies 5	Primary bacteremia	ABPC/SBT	Recover	2012	8
10	3y	F	Pompe disease	No	<i>R. mucosa</i>	Primary bacteremia	ABPC/SBT	Recover	2012	8
11	3y	n.	ALL	385 / μ L	<i>R. mucosa</i>	Probably CVC	IPM, AMK	Recover	2014	17
12	17y	M	AML	n.d.	<i>R. mucosa</i>	Probably CVC	carbapenem	Recover	2016	18
13	9y	M	Cerebellar medulloblastoma	40 / μ L	<i>R. mucosa</i>	Probably CVC	MEM	Recover	2017	Present case

ABPC/SBT, ampicillin/sulbactam; ALL, acute lymphoblastic leukemia; AMK, amikacin; AML, acute myeloid leukemia; CAZ, ceftazidime; CPFX, ciprofloxacin; CTX, cefotaxime; CVC, central venous catheter; GM, gentamicin; IPM, imipenem; MEM, meropenem; NB, neuroblastoma. n.d., not described; TPN, total parenteral nutrition.

Roseomonas organisms form characteristic pink-pigmented colonies on agar plates [11], which show positive reactions to catalase and urease, and thus identification at the genus level is not such difficult. However, commercial microbiologic kits using a phenotypic approach may result in misidentification, and accurate bacterial identification of the organisms at the species level requires genetic techniques. As in previous cases [14,17,20], we identified the *R. mucosa* strain using 16S rDNA sequencing.

To conclude, we described a case of *R. mucosa* bacteremia in a Japanese boy with febrile neutropenia following chemotherapy for cerebellar medulloblastoma. A literature review revealed that a common clinical feature of bacteremia caused by *Roseomonas* in children is a healthcare-associated catheter-related infection in an immunocompromised child with an underlying disease, particularly a malignancy. *Roseomonas* species are comparatively resistant to various antimicrobials; however, good outcomes can be expected, even in pediatric cases.

Declarations of interest

None.

Consent

Written informed consent was obtained from the parents for publication. A copy of the written consent is available for review by the Editor-in-Chief of this journal on request.

Contribution

Writing, H. Hagiya and K. Kimura. Supervision, I. Nishi, H. Yoshida, and K. Tomono.

Acknowledgments

This study was supported by the Center for Medical Research and Education, Graduate School of Medicine, Osaka University. We

would like to thank enago (www.enago.jp/) for English language editing.

References

- [1] Rihs J.D., Brenner DJ, Weaver RE, Steigerwalt AG, Hollis DG, Yu VL. *Roseomonas*, a new genus associated with bacteremia and other human infections. *J Clin Microbiol* 1993;31:3275–83.
- [2] Romano-Bertrand S, Bourdier A, Aujoulat F, et al. Skin microbiota is the main reservoir of *Roseomonas mucosa*, an emerging opportunistic pathogen so far assumed to be environmental. *Clin Microbiol Infect* 2016;22(737): e731–7.
- [3] Chung EJ, Yoon HS, Kim KH, Jeon CO, Chung YR. *Roseomonas oryzaicola* sp. Nov., isolated from the rhizosphere of rice (*Oryza sativa* L.). *Int J Syst Evol Microbiol* 2015;65:4839–44.
- [4] Gallego V, Sanchez-Porro C, Garcia MT, Ventosa A. *Roseomonas Aquatica* sp. Nov., isolated from drinking water. *Int J Syst Evol Microbiol* 2006;56:2291–5.
- [5] Zhang YQ, Yu LY, Wang D, et al. *Roseomonas Vinacea* Sp. Nov., a gram-negative coccobacillus isolated from a soil sample. *Int J Syst Evol Microbiol* 2008;58:2070–4.
- [6] Shokar NK, Shokar GS, Islam J, Cass AR. *Roseomonas gilardii* infection: case report and review. *J Clin Microbiol* 2002;40:4789–91.
- [7] De I, Rolston KV, Han XY. Clinical significance of *Roseomonas* species isolated from catheter and blood samples: analysis of 36 cases in patients with Cancer. *Clin Infect Dis* 2004;38:1579–84.
- [8] Wang CM, Lai CC, Tan CK, et al. Clinical characteristics of infections caused by *Roseomonas* species and antimicrobial susceptibilities of the isolates. *Diagn Microbiol Infect Dis* 2012;72:199–203.
- [9] Struthers M, Wong J, Janda JM. An initial appraisal of the clinical significance of *Roseomonas* species associated with human infections. *Clin Infect Dis* 1996;23:729–33.
- [10] Bhende M, Karpe A, Arunachalam S, Therese KL, Biswas J. Endogenous endophthalmitis due to *Roseomonas mucosa* presenting as a subretinal abscess. *J Ophthalmic Inflamm Infect* 2017;7:5.
- [11] Wallace PL, Hollis DG, Weaver RE, Moss CW. Biochemical and chemical characterization of pink-pigmented oxidative Bacteria. *J Clin Microbiol* 1990;28:689–93.
- [12] Han XY, Pham AS, Tarrand JJ, Rolston KV, Helsel LO, Levett PN. Bacteriologic characterization of 36 strains of *Roseomonas* species and proposal of *Roseomonas mucosa* sp. Nov. and *Roseomonas gilardii* Subsp. *Rosea* subsp. nov. *Am J Clin Pathol* 2003;120:256–64.
- [13] Kim KY, Hur J, Jo W, et al. Infectious spondylitis with bacteremia caused by *Roseomonas mucosa* in an immunocompetent patient. *Infect Chemother* 2015;47:194–6.
- [14] Bard JD, Deville JG, Summanen PH, Lewinski MA. *Roseomonas mucosa* isolated from bloodstream of pediatric patient. *J Clin Microbiol* 2010;48:3027–9.

- [15] Marin ME, Marco Del Pont J, Dibar E, et al. Catheter-related bacteremia caused by *Roseomonas gilardii* in an immunocompromised patient. *Int J Infect Dis* 2001;5:170–1.
- [16] McLean TW, Rouster-Stevens K, Woods CR, Shetty AK. Catheter-related bacteremia due to *Roseomonas* species in pediatric hematology/oncology patients. *Pediatr Blood Cancer* 2006;46:514–6.
- [17] Michon AL, Saumet L, Bourdier A, Haouy S, Sirvent N, Marchandin H. Bacteremia due to imipenem-resistant *Roseomonas mucosa* in a child with acute lymphoblastic leukemia. *J Pediatr Hematol Oncol* 2014;36:e165–168.
- [18] Kim YK, Moon JS, Song KE, Lee WK. Two cases of bacteremia due to *Roseomonas mucosa*. *Ann Lab Med* 2016;36:367–70.
- [19] Abu Choudhury M, Wailan AM, Sidjabat HE, et al. Draft genome sequence of *Roseomonas mucosa* strain Au37, isolated from a peripheral intravenous catheter. *Genome Announc* 2017;5:.
- [20] Fanella S, Schantz D, Karlowsky J, Rubinstein E. Septic arthritis due to *Roseomonas gilardii* in an immunocompetent adolescent. *J Med Microbiol* 2009;58:1514–6.