





Complete Genome Resequencing of Thermus thermophilus Strain TMY by Hybrid Assembly of Long- and Short-Read **Sequencing Technologies**

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ABSTRACT Complete genome resequencing was conducted for Thermus thermophilus strain TMY by hybrid assembly of Oxford Nanopore Technologies long-read and MGI shortread data. Errors in the previously reported genome sequence determined by PacBio technology alone were corrected, allowing for high-quality comparative genomic analysis of closely related T. thermophilus genomes.

hermus thermophilus is an aerobic, thermophilic bacterium that grows optimally at around 70 to 75°C. Since the first isolation of this species from Mine Hot Spring in Japan in 1968 (1, 2), many T. thermophilus strains have been isolated from various thermal areas worldwide (3-11). Among them, strains HB8 (type strain) and HB27 have been rigorously studied biochemically (12, 13), structurally (14, 15), and genetically (16, 17).

So far, 13 complete genome sequences have been determined for *T. thermophilus* strains (https://www.ncbi.nlm.nih.gov/genome/browse#!/prokaryotes/461/). One of the strains, TMY, was isolated from the Otake geothermal power plant in Japan (6), and its genome sequence has been reported previously (18). However, because the sequencing was performed using PacBio technology alone, it contained numerous errors (frameshifted proteins), resulting in exclusion from the RefSeq database. For fine-scale comparative analysis of closely related T. thermophilus genomes, here, we reinvestigated the genome of TMY by combining Oxford Nanopore Technologies (ONT) long-read and MGI short-read sequencing technologies.

Freeze-dried TMY cells (JCM 10668) obtained from JCM (RIKEN, Japan) were inoculated into 5 ml of Thermus medium (ATCC 697), containing 0.4 mM MgCl₂ and 0.35 mM CaCl₂. After 24 h of cultivation at 70°C, genomic DNA was purified from pelleted cells using a blood and cell culture DNA midi kit (Qiagen). For long-read sequencing, unsheared genomic DNA $(1 \mu g)$ was pretreated using a short-read eliminator kit (Circulomics) to remove fragments of <10 Kbp, and a library was constructed using a ligation sequencing kit (ONT). Sequencing was performed using a GridlON X5 system on a FLO-MIN106 R9.41 flow cell (ONT). Base calling was conducted using Guppy v.4.0.11 to generate 375,245 reads (average, 4,578 bases; total, 1.72 Gb). For all software, default parameters were used. The raw sequencing data were filtered (Q < 10; length, <1,000 bases) using NanoFilt v.2.7.1 (19), yielding 213,707 reads (longest read, 163,227 bases; N_{so} , 9,231 bases; total, 1.15 Gb). For short-read sequencing, a library was constructed using an MGIEasy FS PCR-free DNA library prep set (MGI) with a \sim 400 to 500-bp insert. Paired-end sequencing was then performed on a DNBSEQ-400 instrument (MGI), yielding 8,454,552 paired-end reads (2 \times 150 bases). The raw sequencing data were filtered (Q < 30; length, <10 bases) using fastp v.0.20.1 (20), yielding 5,182,387 paired-end reads (average, 150 bases; total, 2.53 Gb). The trimmed long- and short-read data were assembled using Unicycler v.0.4.8 (21), and the assembly was polished using Pilon v.1.23 (22), generating a single circular chromosome and a single circular plasmid. Automatic annotation was conducted using DFAST v.1.4.0 (23), and the genomic features are summarized in Table 1.

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TABLE 1 Genome statistics and features of *Thermus thermophilus* strain TMY

Genetic element	Length (bp)	GC content (%)	No. of coding DNA sequences	No. of rRNAs	No. of tRNAs	GenBank accession no.
Chromosome	2,151,326	69.0	2,309	6	52	AP025158
Plasmid	19,144	67.4	26	0	1	AP025159

Comparative sequence analysis between the previously reported sequence (chromosome, GenBank accession number AP017920.1; plasmid pTMY, AP017921.1) (18) and the present result revealed 98.2% pairwise identity. The majority of differences were single-nucleotide deletions in the PacBio sequence, while the presence of two large insertions (~30 Kbp total) in our sequence was experimentally confirmed by Sanger sequencing.

Data availability. The complete genome sequence of *T. thermophilus* TMY is available from DDBJ/EMBL/GenBank under the accession numbers summarized in Table 1. The raw sequencing data were deposited in the SRA database under the accession numbers DRR313875 (Nanopore) and DRR313876 (DNBSEQ).

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REFERENCES

- Oshima T, Imahori K. 1971. Isolation of an extreme thermophile and thermostability of its transfer ribonucleic acid and ribosomes. J Gen Appl Microbiol 17:513–517. https://doi.org/10.2323/jgam.17.513.
- Oshima T, Imahori K. 1974. Description of *Thermus thermophilus* (Yoshida and Oshima) comb. nov., a nonsporulating thermophilic bacterium from a Japanese thermal spa. Int J Sys Evol Microbiol 24:102–112. https://doi.org/10 .1099/00207713-24-1-102.
- Manaia CM, Hoste B, Gutierrez MC, Gillis M, Ventosa A, Kersters K, Da Costa MS. 1995. Halotolerant *Thermus* strains from marine and terrestrial hot springs belong to *Thermus thermophilus* (ex Oshima and Imahori, 1974) nom. rev. emend. Syst Appl Microbiol 17:526–532. https://doi.org/ 10.1016/S0723-2020(11)80072-X.
- Beffa T, Blanc M, Lyon PF, Vogt G, Marchiani M, Fischer JL, Aragno M. 1996. Isolation of *Thermus* strains from hot composts (60 to 80 degrees C). Appl Environ Microbiol 62:1723–1727. https://doi.org/10.1128/aem.62.5.1723-1727.1996.
- Lyon PF, Beffa T, Blanc M, Auling G, Aragno M. 2000. Isolation and characterization of highly thermophilic xylanolytic *Thermus thermophilus* strains from hot composts. Can J Microbiol 46:1029–1035. https://doi.org/10.1139/w00-075.
- Fujino Y, Kawatsu R, Inagaki F, Umeda A, Yokoyama T, Okaue Y, Iwai S, Ogata S, Ohshima T, Doi K. 2008. *Thermus thermophilus* TMY isolated from silica scale taken from a geothermal power plant. J Appl Microbiol 104: 70–78. https://doi.org/10.1111/j.1365-2672.2007.03528.x.
- Miyazaki K. 2019. Complete genome sequencing of *Thermus thermophilus* strain HC11, isolated from Mine Geyser in Japan. Microbiol Resour Announc 8:e00873-19. https://doi.org/10.1128/MRA.00873-19.
- Miyazaki K, Tomariguchi N. 2019. Complete genome sequences of *Thermus ther-mophilus* strains AA2-20 and AA2-29, isolated from Arima Onsen in Japan. Microbiol Resour Announc 8:e00820-19. https://doi.org/10.1128/MRA.00820-19.
- Miyazaki K, Moriya T, Nemoto N, Oshima T, Yura K, Bessho Y. 2021. Complete genome sequence of *Thermus thermophilus* strain HB5018, isolated from Mine Hot Spring in Japan. Microbiol Resour Announc 10:e00039-21. https://doi.org/10.1128/MRA.00039-21.
- Miyazaki K, Moriya T, Tokito N, Oshima T, Yura K, Bessho Y. 2021. Complete genome sequences of *Thermus thermophilus* strains HB5002 and HB5008, isolated from Mine Hot Spring in Japan. Microbiol Resour Announc 10:e00272-21. https://doi.org/10.1128/MRA.00272-21.
- Miyazaki K, Tomariguchi N, Ueno Y. 2021. Complete genome sequences of four halophilic *Thermus thermophilus* strains isolated from Arima Hot Spring in Japan. Microbiol Resour Announc 10:e00874-21. https://doi.org/10.1128/ MRA.00874-21.
- Miyazaki K. 2005. A hyperthermophilic laccase from *Thermus thermophilus* HB27. Extremophiles 9:415–425. https://doi.org/10.1007/s00792-005-0458-z.

- Yamada T, Akutsu N, Miyazaki K, Kakinuma K, Yoshida M, Oshima T. 1990. Purification, catalytic properties, and thermal stability of threo-Ds-3-iso-propylmalate dehydrogenase coded by *leuB* gene from an extreme thermophile, *Thermus thermophilus* strain HB8. J Biochem 108:449–456. https://doi.org/10.1093/oxfordjournals.jbchem.a123220.
- Imada K, Sato M, Tanaka N, Katsube Y, Matsuura Y, Oshima T. 1991. Threedimensional structure of a highly thermostable enzyme, 3-isopropylmalate dehydrogenase of *Thermus thermophilus* at 2.2 Å resolution. J Mol Biol 222:725–738. https://doi.org/10.1016/0022-2836(91)90508-4.
- Yokoyama S, Hirota H, Kigawa T, Yabuki T, Shirouzu M, Terada T, Ito Y, Matsuo Y, Kuroda Y, Nishimura Y, Kyogoku Y, Miki K, Masui R, Kuramitsu S. 2000. Structural genomics projects in Japan. Nat Struct Biol 7:943–945. https://doi.org/10.1038/80712.
- Tamakoshi M, Yaoi T, Oshima T, Yamagishi A. 1999. An efficient gene replacement and deletion system for an extreme thermophile, *Thermus ther-mophilus*. FEMS Microbiol Lett 173:431–437. https://doi.org/10.1111/j.1574 -6968.1999.tb13535.x.
- Miyazaki K, Tomariguchi N. 2019. Occurrence of randomly recombined functional 16S rRNA genes in *Thermus thermophilus* suggests genetic interoperability and promiscuity of bacterial 16S rRNAs. Sci Rep 9:11233. https://doi.org/10.1038/s41598-019-47807-z.
- Fujino Y, Nagayoshi Y, Ohshima T, Ogata S, Doi K. 2017. Complete genome sequence of *Thermus thermophilus* TMY, isolated from a geothermal power plant. Genome Announc 5:e01596-16. https://doi.org/10.1128/genomeA.01596-16.
- De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. Bioinformatics 34:2666–2669. https://doi.org/10.1093/bioinformatics/bty149.
- Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. https://doi.org/10.1093/bioinformatics/ bty560.
- 21. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. PLoS Comput Biol 13:e1005595. https://doi.org/10.1371/journal.pcbi.1005595.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. https://doi.org/10.1371/journal.pone.0112963.
- Tanizawa Y, Fujisawa T, Kaminuma E, Nakamura Y, Arita M. 2016. DFAST and DAGA: Web-based integrated genome annotation tools and resources. Biosci Microbiota Food Health 35:173–184. https://doi.org/10.12938/bmfh.16-003.

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