

Genetic Variations in the Regulator of G-Protein Signaling Genes Are Associated with Survival in Late-Stage Non-Small Cell Lung Cancer

Jingyao Dai¹, Jian Gu¹, Charles Lu², Jie Lin¹, David Stewart², David Chang¹, Jack A. Roth³, Xifeng Wu¹*

1 Department of Epidemiology, The University of Texas MD Anderson Cancer Center, Houston, Texas, United States of America, 2 Department of Thoracic/Head and Neck Medical Oncology, The University of Texas MD Anderson Cancer Center, Houston, Texas, United States of America, 3 Department of Thoracic and Cardiovascular Surgery, The University of Texas MD Anderson Cancer Center, Houston, Texas, United States of America

Abstract

The regulator of G-protein signaling (RGS) pathway plays an important role in signaling transduction, cellular activities, and carcinogenesis. We hypothesized that genetic variations in RGS gene family may be associated with the response of late-stage non-small cell lung cancer (NSCLC) patients to chemotherapy or chemoradiotherapy. We selected 95 tagging single nucleotide polymorphisms (SNPs) in 17 RGS genes and genotyped them in 598 late-stage NSCLC patients. Thirteen SNPs were significantly associated with overall survival. Among them, rs2749786 of *RGS12* was most significant. Stratified analysis by chemotherapy or chemoradiation further identified SNPs that were associated with overall survival in subgroups. Rs2816312 of *RGS1* and rs6689169 of *RGS7* were most significant in chemotherapy group and chemoradiotherapy group, respectively. A significant cumulative effect was observed when these SNPs were combined. Survival tree analyses identified potential interactions between rs944343, rs2816312, and rs1122794 in affecting survival time in patients treated with chemotherapy, while the genotype of rs6429264 affected survival in chemoradiation-treated patients. To our knowledge, this is the first study to reveal the importance of RGS gene family in the survival of late-stage NSCLC patients.

Citation: Dai J, Gu J, Lu C, Lin J, Stewart D, et al. (2011) Genetic Variations in the Regulator of G-Protein Signaling Genes Are Associated with Survival in Late-Stage Non-Small Cell Lung Cancer. PLoS ONE 6(6): e21120. doi:10.1371/journal.pone.0021120

Editor: Lin Zhang, University of Pennsylvania, United States of America

Received May 4, 2011; Accepted May 19, 2011; Published June 17, 2011

Copyright: © 2011 Dai et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by the National Institutes of Health grants R01 CA111646, P50 CA070907, and R01 CA055769. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. No additional external funding was received for this study.

1

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: xwu@mdanderson.org

Introduction

Non-small cell lung cancer (NSCLC) is the leading cause of cancer mortality worldwide [1]. Over 45% of NSCLC patients present with unresectable late-stage (stage IIIA/B or stage IV) disease in the United States [2]. A combined modality therapy is the current standard of care for patients with stage III NSCLC with good performance status (performance score 0 or 1). Numerous clinical trials have shown that concurrent chemoradiation offers a significant survival advantage over sequential chemoradiation [3]. Although concurrent chemoradiotherapy significantly improves the survival of patients with locally advanced disease, the majority of patients still die within 5 years because of locoregional or distant disease progression [4]. The stage IV patients are usually offered palliative chemotherapy and supportive care [5]. There is a wide variability in patients' response to chemoradiation and clinicopathological variables alone do not provide satisfactory guidance for the decision of treatment strategy. The application of pharmacogenomics may improve the prediction of response and help clinicians determine cancer treatments for individual NSCLC patient according to his unique genetic background. Therefore, in this study, we aimed to identify genetic predictors for clinical outcomes of late stage NSCLC patients.

G proteins (guanine nucleotide-binding proteins) are important cellular signal transduction molecules that are expressed in all

human cells [6,7]. They are activated by G protein-coupled receptors (GPCRs) and thereby may transduce extracellular signals into the interior of a cell [8]. GPCRs are a family of seventransmembrane domain receptors. When GPCRs traduce a signal inside the cell, the extracellular domain of GPCR first binds to the signal molecules, and then the intracellular domain of GPCR activates a heterotrimeric G-protein. The heterotrimeric G protein functions as "molecular switches" and can activate a cascade of signaling factors and downstream target activation [7]. This G protein-coupled biological process requires fine-tuning through accessory molecules such as the regulator of G-protein signaling (RGS) [9]. RGS proteins are a big family of over 30 intracellular proteins [10], which can negatively modulate GPCRs signaling pathways [11,12]. RGS are multi-functional, GTPase-accelerating proteins that promote GTP hydrolysis by the alpha subunit of heterotrimeric G proteins, thereby inactivating the G protein and rapidly switching off GPCR signaling pathways[11]. All RGS proteins contain a RGS domain (also referred as "RGS-box") ,which is required for their activities [13], and these RGS domains mediate the interaction with other signaling proteins, allowing RGS proteins to serve as signaling scaffolds [8]. Malfunctions of RGS proteins have been reported to be related to the pathogenesis of many common human diseases and drug addiction [14,15,16,17]. Multiple RGS proteins were found differentially expressed in a variety of solid and hematological malignancies[18,19,20,21,22, 23,24,25,26,27,28,29,30,31,32,33,34,35,36].

The single nucleotide polymorphisms (SNPs) of RGS have been associated with several human diseases, suggesting that genetic variation in the RGS pathway may play a significant role in these diseases' pathogenesis [37,38]. Recently, RGS SNPs have also been reported to play important roles in lung cancer. For instance, SNPs in RGS17 on chromosome 6q23-25 was associated with familial lung cancer susceptibility [39]. SNPs in RGS2 and RGS6 may modulate the risks of bladder and lung cancers [37,40]. Whether genetic variants in the RGS pathway could influence clinical outcomes in patients with NSCLC remains unknown. In this study, we tested the hypothesis that genetic variations of RGS are associated with the survival of late-stage NSCLC patients receiving chemotherapy or chemoradiation.

Results

We included 598 NSCLC patients in this study, with a mean age of 59.7 years (**Table 1**). Of the 598 patients, 456 were dead and 142 were alive. We found no significant difference in age (P=0.884), ethnicity (P=0.937), smoking status (P=0.860), and pack-years of smoking (P=0.926) between the two groups of patients (**Table 1**). However, we observed a significant difference

Table 1. Distribution of demographic and clinical variables by survival status.

Variable	Dead (n = 456)	Alive (n = 142)	P value*
	Number (%)	Number (%)	
Gender			
Men	262 (81.11)	61 (18.89)	
Women	194 (70.55)	81 (29.45)	0.002
Ethnicity			
Caucasian	358 (76.17)	112 (23.83)	
African American	72 (75.79)	23 (24.21)	
Other	26 (78.79)	7 (21.21)	0.937
Smoking status			
Never smoker	84 (75.00)	28 (25.00)	
Former smoker	184 (74.72)	59 (24.28)	
Current smoker	188 (77.37)	55 (22.63)	0.860
Clinical stage			
IIIA	57 (69.51)	25 (30.49)	
IIIB (Dry)	100 (70.42)	42 (29.58)	
IIIB (Wet)	25 (60.10)	14 (35.90)	
IV	274 (81.79)	61 (18.21)	0.004
Performance status			
0	96 (67.13)	47 (32.87)	
1	254 (76.05)	80 (23.95)	
2-4	66 (89.19)	8 (10.81)	<0.002
Therapy			
Chemotherapy	295 (64.69)	60 (16.90)	
Chemoradiotherapy	104 (76.47)	32 (23.53)	
Both	57 (53.27)	50 (46.73)	<0.001
Age, years (mean±SD)59.7±10.6	59.6±10.0	0.884
Pack-years (mean±SE)36.6±30.4	36.9±31.3	0.926

*Significant P values are in bold font.

in mortality status by gender (P=0.002), clinical stage (P=0.004), and performance status (P=0.002) (**Table 1**).

Associations between SNPs and overall survival in latestage NSCLC patients

A total of 13 SNPs in 6 genes were significantly associated with the risk of death at P<0.05 (**Table 2**). Among them, the variant alleles of four SNPs, rs7549021 and rs1056515 of RGS5, rs944343 of RGS3, and rs2749786 of RGS12, were associated with decreased risks of death, with adjusted HRs of 0.42 (95% CI, 0.22 to 0.83), 0.72 (95% CI, 0.54 to 0.97), 0.80 (95% CI, 0.67 to 0.95), and 0.58 (95% CI, 0.40 to 0.85), respectively. Other SNPs conferred increased risks of death. All SNPs in the RGS1 gene were in linkage disequilibrium (with $r^2>0.8$) with similar HRs in a dominant model.

The bootstrap re-sampling analysis was then performed for the 13 SNPs to internally validate the results. We found that only 5 SNPs, rs944343 (RGS3), rs6678136 (RGS4), rs7549021 (RGS5), rs3820487 (RGS5), and rs2749786 (RGS12), had bootstrap P values <0.05 at least 70 times out of 100 times (**Table 2**). The other SNPs had bootstrap P values <0.05 less than 50 times, indicating that those SNPs were likely false-positive results.

Associations between SNPs and risk of death stratified by treatment

We then performed a stratified analysis by treatment modality. chemotherapy or chemoradiation (**Tables 3 and 4**). Nine SNPs were associated with overall survival in patients who received chemotherapy only, 5 of which had bootstrap P values <0.05 at least 70 times out of 100 times (**Table 3**). Among these five SNPs, three (rs2816312 [RGS1], rs10218752 [RGS5], and rs1122794 [RGS11]) were associated with an increased risk of death, with HRs of 1.80 (95% CI, 1.32 to 2.45), 1.76 (95% CI, 1.06 to 2.90), and 1.37 (95% CI, 1.07 to 1.77), respectively. On the other hand, rs944343 (RGS3) and rs1051013 (RGS3) were associated with a decreased risk of death, with HRs of 0.73 (95% CI, 0.57 to 0.94) and 0.77 (95% CI, 0.60 to 0.98), respectively. In Kaplan-Meier analyses, four of the five significant SNPs (rs2816312 [RGSI], rs944343 [RGS3], rs1051013 [RGS3], and rs1122794 [RGS11]) were significantly associated with altered median-survival time (MST) (log-rank P value < 0.05) (**Table 3**).

There were eight SNPs significantly associated with survival status in patients who received chemoradiation, five of which had bootstrap P values <0.05 more than 70 times out of 100 times (**Table 4**). Among these five SNPs, three (rs2344673 [*RGS5*], rs12127281 [*RGS5*], and rs6429264 [*RGS7*]) were associated with an increased risk of death, with HRs of 1.86 (95% CI, 1.00 to 3.47), 1.83 (95% CI, 1.09 to 3.10), and 1.89 (95% CI, 1.06 to 3.38), respectively; while rs12757054 (*RGS7*) and rs6689169 (*RGS7*) were associated with a decreased risk of death, with HRs of 0.48 (95% CI, 0.26 to 0.86) and 0.47 (95% CI, 0.27 to 0.80), respectively. Two of these 5 SNPs on *RGS7* (rs6429264 and rs6689169) were associated with altered MST for NSCLC patients (log-rank P=0.0055 and 0.0441, respectively) (**Table 4**).

Cumulative effects of the unfavorable genotypes on survival

We further assessed the cumulative effects of the unfavorable genotypes in either treatment groups using the SNPs with bootstrap P values <0.05 at least 70 times out of 100 times in each group (**Table 5**). There were significant gene-dose effects in patients receiving both treatments (**Table 5**). In those patients receiving chemotherapy only and taking the low-risk reference

Table 2. Significant SNPs associated with overall survival.

Gene and SNPs	No. of Dead/Alive	HR* (95% CI)	Smallest P	MST	Log-rank <i>P</i> value	No. of times in boot- strap samples <i>P</i> <0.05
RGS3						
rs944343 ^{&}						99
GG	274/77	Reference		11.68		
CG	158/60	0.78 (0.63-0.97)	0.022435	14.21		
CC Trend	24/5	0.69 (0.43–1.10) 0.80 (0.67–0.95)	0.118902 P _{trend} = 0.011796	18.29	0.1278	
RGS4						
rs6678136 ^{&}						90
GG/GA	363/120	Reference		13.45		
AA	93/22	1.36 (1.07– 1.73)	0.012744	10.92	0.0364	
RGS5						
rs7549021 ^{&}						93
AA/AG	444/132	Reference		12.83		
GG	12/10	0.42 (0.22-0.83)	0.012568	24.51	0.0647	
rs3820487 ^{&}						95
CC/CA	431/141	Reference		13.45		
AA	25/1	1.81 (1.09–3.01)	0.022834	9.57	0.0001	
rs1056515						28
CC/CA	395/119	Reference		12.86		
AA	61/23	0.72 (0.54–0.97)	0.027911	13.62	0.1242	
RGS12		,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,				
rs2749786 ^{&}						100
AA/AG	425/125	Reference		12.70		
GG	31/17	0.58 (0.40–0.85)	0.005532	26.84	0.0015	
RGS22	31,17	0.50 (0.10 0.05)	0.003332	20.01	0.0015	
rs2453627						41
CC	133/52	Reference		15.43		71
CG/GG	323/90	1.26 (1.01–1.56)	0.039106	12.04	0.0247	
RGS1	323/90	1.20 (1.01-1.50)	0.039100	12.04	0.0247	
rs2760535 [#]						47
GG	240/111	Reference		12.50		47
	348/111		0.021054	13.59	0.1550	
GA/AA	108/31	1.31 (1.02–1.67)	0.031854	10.72	0.1558	11
rs1323291#	247/440	2.6		42.50		11
AA	347/110	Reference		13.59		
AC/CC	109/33	1.28 (1.00–1.65)	0.049051	10.72	0.2181	
rs16834456 [#]						47
CC	348/111	Reference		13.59		
CA/AA	108/31	1.31 (1.02–1.67)	0.031854	10.72	0.1558	
rs9427560 [#]						47
AA	348/111	Reference		13.59		
AG/GG	108/31	1.31 (1.02–1.67)	0.031854	10.72	0.1558	
rs2816310 [#]						11
CC	347/110	Reference		13.59		
CA/AA	109/33	1.28 (1.00–1.65)	0.049051	10.72	0.2181	
rs2816311 [#]						11
AA	347/110	Reference		13.59		
AG/GG	109/33	1.28 (1.00-1.65)	0.049051	10.72	0.2181	

Table 2. Cont.

#Linkage SNPs.

Abbreviations: SNPs, single nucleotide polymorphism; No., number; HR, hazard ratio; Cl, confidence interval; MST, median survival time.

&SNPs which had bootstrap P values < 0.05 at least 90% of times.

*HR adjusted by age, gender, ethnicity, smoking status and pack-years, performance status, clinical stage, and treatments.

doi:10.1371/journal.pone.0021120.t002

group as reference (0 unfavorable genotypes), the medium-risk (1 or 2 unfavorable genotypes) and the high-risk groups (3 or 4 unfavorable genotypes) had 1.69-fold (95% CI, 1.19 to 2.41; $P\!=\!0.004$) and 2.52-fold (95% CI, 1.71–3.71; $P\!<\!0.001$) increased risk of death, respectively (P for trend $<\!0.001$). The MST for the medium-risk and the high-risk groups were 11.22 months and 8.19 months, respectively, compared to 18.22 months for the low-risk

reference group (log-rank P value <0.0001). Similar dose-response trends were observed among patients receiving chemoradiation (**Table 5** and **Figure 1**).

Higher-order gene-gene interactions

The results of STREE program analysis for the interaction of the 10 bootstrap-validated significant SNPs (the SNPs which had

Table 3. Stratified analysis by treatment modality: Chemotherapy.

SNPs	Gene	No. of Dead/Alive	HR* (95%CI)	Smallest P	MST	Log-rank <i>P</i> value	No. of times in Boot- strap samples P<0.05
rs2816312 ^{&}	RGS1						100
AA		229/48	Reference		11.64		
AG/GG		66/12	1.80 (1.32–2.45)	0.000199	8.16	0.0134	
rs944343 ^{&}	RGS3						99
GG		178/30	Reference		9.44		
GC/CC		117/30	0.73 (0.57-0.94)	0.013565	12.76	0.0009	
rs1051013 ^{&}	RGS3						70
GG		186/33	Reference		9.57		
GA/AA		109/27	0.77 (0.60-0.98)	0.03534	12.70	0.0147	
rs10218752 ^{&}	RGS5						97
AA/AG		278/59	Reference		10.92		
GG		17/1	1.76 (1.06–2.90)	0.027803	9.01	0.0861	
rs1122794 ^{&}	RGS11						96
СС		193/48	Reference		11.55		
CA/AA		102/12	1.37 (1.07–1.77)	0.013747	9.47	0.0249	
rs12339493	RGS3						38
GG		239/43	Reference		10.53		
GA/AA		56/17	0.73 (0.53-0.99)	0.043020	12.40	0.0311	
rs7549021	RGS5						0
AA/AG		286/58	Reference		10.72		
GG		9/2	0.42(0.22-0.83)	0.013	9.57	0.6077	
rs2749786	RGS12						4
AA/AG		277/54	Reference		10.66		
GG		18/6	0.60 (0.37-1.00)	0.049813	11.71	0.015	
rs594149	RGS16						42
CC		205/35	Reference		10.59		
CG/GG		90/25	0.80 (0.61-1.04)	0.0984	11.71	0.0467	
P for trend				0.0446			
AA/AG		102/30	Reference		16.58		
GG		2/2	0.10 (0.01–0.78)	0.027524	24.51	0.0987	

Abbreviations: SNPs, single nucleotide polymorphism; No., number; HR, hazard ratio; CI, confidence interval; MST, median survival time.

doi:10.1371/journal.pone.0021120.t003



PLoS ONE | www.plosone.org

 $^{^{\&}amp;}$ SNPs which had bootstrap *P* values < 0.05 at least 70% of time.

^{*}HR adjusted by age, gender, ethnicity, smoking status and pack-years, performance status, clinical stage.

[#]MST, median survival time (months).

Table 4. Stratified analysis by treatment modality: Chemoradiation.

SNPs	Gene	No. of Dead/Alive	HR* (95%CI)	Smallest P	MST	Log-rank <i>P</i> value	No. of times in Boot- strap samples P<0.05
rs2344673 ^{&}	RGS5						88
GG		79/28	Reference		17.86		
GA/AA		25/4	1.86 (1.00–3.47)	0.049508	11.48	0.1024	
rs12127281 ^{&}	RGS5						86
GG		59/25	Reference		19.14		
GA/AA		45/7	1.83 (1.09–3.10)	0.023387	12.30	0.0602	
rs12757054 ^{&}	RGS7						73
GG		86/24	Reference		15.89		
GA/AA		18/8	0.48 (0.26-0.86)	0.013902	24.51	0.2311	
rs6429264 ^{&}	RGS7						90
GG		82/28	Reference		19.28		
GA/AA		22/4	1.89 (1.06–3.38)	0.031160	12.37	0.0055	
rs6689169 ^{&}	RGS7						96
AA		78/18	Reference		13.22		
AG/GG		26/14	0.47 (0.27-0.80)	0.005776	24.51	0.0441	
rs11586945	RGS5						3
GG		64/24	Reference		19.67		
GC/CC		40/8	1.63 (1.02–2.61)	0.040998	12.53	0.0623	
rs2999966	RGS5						0
CC/CA		96/27	Reference		16.05		
AA		8/5	0.38 (0.15-0.99)	0.047362	35.92	0.0642	
rs7549021	RGS5						0
AA/AG		102/30	Reference		16.58		
GG		2/2	0.10 (0.01-0.78)	0.027524	24.51	0.0987	

Abbreviations: SNPs, single nucleotide polymorphism; No., number; HR, hazard ratio; CI, confidence interval; MST, median survival time. &SNPs which had bootstrap P values <0.05 at least 70% of time.

doi:10.1371/journal.pone.0021120.t004

bootstrap P values < 0.05 at least 70% of time in **Tables 3 and 4**) in stratified analysis were presented in Figure 2. Survival tree analysis resulted in four terminal nodes in the chemotherapy group and two terminal nodes in the chemoradiation group (Figure 2A). In the chemotherapy group, the initial split was rs944343 (RGS3), and subsequent splits were rs2816312 (RGS1) and rs1122794 (RGS11). Different nodes had different percentages of death event. To assess the risk associated with each of the terminal nodes, node 1 in the chemotherapy branch was taken as the reference group, composed of individuals with the heterozygous and the homozygous variant genotypes of rs944343 (RGS3) and the homozygous wild-type genotype of rs1122794 (RGS11). Compared with the reference group, patients in the terminal nodes in the chemotherapy group had HRs ranging from 1.49 to 2.92. Patients in node 1 had the longest MST of 13.58 months. The highest at-risk group (node 3), which was composed of patients carrying the homozygous wild-type genotype of rs944343 (RGS3) and variant-containing genotypes of rs2816312 (RGSI), had a HR of 2.92 (95% CI, 1.92 to 4.44). The MSTs were shown to be significantly different in Kaplan-Meier survival analysis (log-rank P value <0.0001) (**Figure 2B** and **Table 6**). In the chemoradiation group, there was only one additional split. Compared to the patients with the homozygous wild-type genotype of rs6429264 (RGS7) (node 5), who had an MST of 19.28 months, the patients

carrying variant-containing genotypes of rs6429264 (*RGST*) exhibited a 1.89-fold increased risk of death (95% CI, 1.06 to 3.38), with an MST of 12.37 months (**Figure 2C** and **Table 6**).

Discussion

In this study, we found that genetic variations in RGS genes were associated with overall survival in late-stage NSCLC patients. Our findings also reinforced the importance of evaluating the cumulative and interaction effects of genetic variations when predicting clinical outcomes of patients with NSCLC.

NSCLC patients are mostly treated with the platinum-based chemotherapy, often in combination with radiation therapy. The platinum-based chemotherapy may be related to several cellular pathways, such as the DNA damage/repair, cell cycle control, and apoptosis pathways [41]. However, there has been no study reporting that RGS is involved in the platinum-based chemotherapy related pathways.

NSCLC cells can invade adjacent tissues and metastasize to nonadjacent organs and tissues, processes that may be attributed to altered cellular signaling pathways [42,43]. Oncogenic transformation is often the direct result of mutations of the signaling molecules, which constitute these pathways. In this study, 5 SNPs were associated with the overall risk of death with

^{*}HR adjusted by age, gender, ethnicity, smoking status and pack-years, performance status, clinical stage.

[#]MST, median survival time (months).

bootstrap P values <0.05 at least 90 times out of 100 times. Three of these 5 SNPs, rs6678136 (RGS4), rs3820487 (RGS5) and rs2749786 (RGS12) conferred significantly different MST in Kaplan-Meier curve (**Table 4**). Previous studies reported that RGS4 gene expression were associated with invasion of several cancer [36,44]. In addition, RGS4 protein acts as an inhibitor of

Table 5. The cumulative effects of unfavorable genotypes on survival.

Chemotherapy						Chemoradiation					
No. of unfavor- able genotypes	No. dead/ alive	HR* (95% CI)	P value	#LSM	Log-rank P value	No. of unfavor- able genotypes	No. dead/ alive	HR* (95% CI)	P value	MST#	Log-rank P value
0	42/18	Reference		18.22		0-1	15/12	Reference		39.80	
1–2	164/31	1.69(1.19–2.41)	0.004	11.22		2–3	67/16	3.48(1.70–7.11)	0.001	13.22	
3-4	89/11	2.52(1.71–3.71)	<0.001	8.19	< 0.0001	4–5	22/4	5.07(2.04-12.64)	<0.001	12.37	0.0119
			$P_{trend}{<}0.001$						$P_{trend}{<}0.001$		

*HR adjusted by age, gender, ethnicity, smoking status and pack-years, performance status, clinical stage #MST, median survival time (months).

doi:10.1371/journal.pone.0021120.t005

epithelial and endothelial cell tubulogenesis by regulating mitogenactivated protein kinases and vascular endothelial growth factor signaling, thereby inhibiting cell proliferation, migration, and invasion [45]. Xiao et a.l reported that multiple SNPs in combination in RGS5 may confer risk for hypertension in Chinese population [46]. RGS5 was reported to be a key modulator of tumor pericyte maturation and play a pivotal role in tumor neovascularization [9]. RGS5 knockout mice showed larger tumor burden and earlier death which may be caused by pericyte maturation and vascular normalization [33]. RGS5 has also been identified as a broadly expressed tumor antigen in multiple types of cancer [47]. RGS12 is a large RGS protein with multiple functional domains such as PDZ, PTB (phospho-tyrosine binding) and Rap binding domains [48]. PDZ domain of RGS12 interacts with a GPCR, CXCR2, and thereby contributes to the GAP action of RGS12 on CXCR2-mediated G-protein signals [49]. Therefore, it is biologically plausible that RGS4, RGS5, and RGS12 are associated with lung cancer survival. The functions of the significant SNPs on these genes are not clear since they are most likely tagging SNPs. Future studies are needed to find the causal SNPs. In stratified analyses, 5 SNPs in the chemotherapy group and

another 5 in the chemoradiation group were associated with the risk of death with bootstrap P values < 0.05 at least 70 times out of 100 times. The genotypes of four SNPs: rs2816312 (RGSI), rs944343 (RGS3), rs1051013 (RGS3), and rs1122794 (RGS11) were found to be significantly associated with MST in chemotherapy group. The most significant one was rs944343 (Log-rank P = 0.0009), which was a tagging SNP located at the 3' flanking region of RGS3. Increased RGS3 expression has been used as a diagnostic marker for soft tissue sarcoma and was associated with resistance to chemotherapy in breast cancer [50,51]. In addition, RGS3 has been reported to modulate glioma cell mobility [36]. The other host genes of SNPs in chemotherapy group, RGS1, and RGS11, have also been associated with the etiology and prognosis of cancer. Rangel et al. reported that RGS1 may be a prognostic marker in melanoma progression and its expression was associated with survival for melanoma patients [20]. Martinez-Cardus, et al. reported that RGS11 expression was significantly associated with the resistance to platinum therapy in colorectal cancer [52]. These studies support the role of RGS1, RGS3, and RGS11 in lung cancer prognosis. There were only two SNPs in chemoradiation group, rs6429264 and rs6689169, significantly associated with MST (log-rank P = 0.0055 and 0.0441, respectively). both of which are tagging SNP and located in RGS7. Several studies demonstrated that tumor necrosis factor-\alpha, a major inflammation cytokine that plays an important role in many human cancers, can rapidly activate the expression level of RGS7 [53,54]. The mechanisms by which these genotypes determine their phenotypes and affect the outcome of NSCLC are not clear. Further studies are warranted to identify the causal variants and the biologic mechanisms underlying our observed associations.

We also observed cumulative effects of RGS SNPs on the survival of late-stage NSCLC patients. In addition, we used survival tree analysis to identify interactions among these SNPs. These gene-gene interactions resulted in four terminal nodes with different risks of death in the chemotherapy-only group and two terminal nodes with different risks of death the chemoradiation

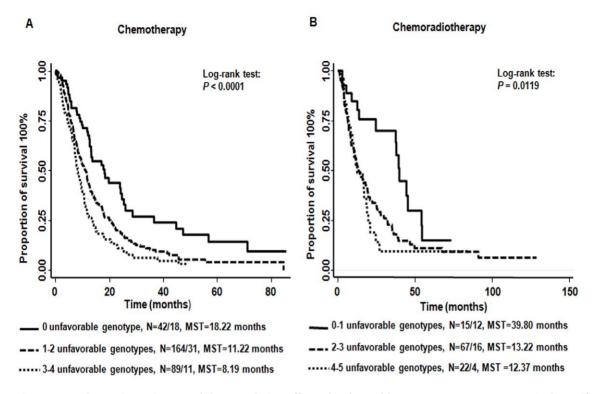


Figure 1. Kaplan-Meier estimates of the cumulative effect of unfavorable genotypes on NSCLC survival stratified by treatment. Solid line represents low-risk group, carrying 0 unfavorable genotypes in chemotherapy (**A**) and 0-1 unfavorable genotypes in chemoradiotherapy (**B**). Dashed line represents the medium-risk group, carrying 1-2 unfavorable genotypes in chemotherapy (**A**) and 2-3 unfavorable genotypes in chemoradiotherapy (**B**). Dotted line represents high risk group, carrying 3-4 unfavorable genotypes in chemotherapy (**A**) and 4–5 unfavorable genotypes in chemoradiotherapy (**B**). N: Number of Dead/Alive. MST: median survival time. doi:10.1371/journal.pone.0021120.g001

group. The cumulative-effects analysis and survival tree analysis may allow us to identify more powerful prognostic or predictive markers and signatures based on the combination of each patient's genetic variations. It should be noted that these types of analyses were exploratory, and the results need to be validated in independent studies.

There are a few strengths to our study. First, our current pathway-based approach is a logical extension of the candidate gene approach and avoids the requirement of much larger sample size by genome-wide association study. Second, we have collected a relative large population of NSCLC patients from the same institution. The uniform standard operation procedures in the cancer identification, pathological staging, and even strategy determination for cancer treatment made our findings more comprehensive and applicable to future clinical studies. Third, we have performed internal statistical validation by a bootstrap resampling procedure to minimize false discoveries. Fourth, we have performed exploratory gene-gene interaction analysis to establish a novel combination of SNPs to predict the outcome of NSCLC patients for their therapy, which could help clinicians in determining the optimal personalized treatment and the quality of care for survival.

To the best of our knowledge, this is the first study investigating the association of genetic variations in RGS family with survival for NSCLC. Our results have provided not only SNP-based analysis, but also a more comprehensive pathway-based approach in the clinical outcome prediction for NSCLC patients who underwent chemotherapy or chemoradiation. Future independent validation in larger population and detailed functional assays are necessary before these findings can be translated to the clinics.

Methods

Ethics Statement

All patients signed a written informed consent and this study has been reviewed and approved by the Institutional Review Board (IRB) of MD Anderson.

Study population and collection of epidemiologic and clinical data

A total of 598 patients with late-stage NSCLC, including stages IIIA, IIIB (Dry), IIIB (Wet), and IV, recruited between 1995 and 2007 from an epidemiological lung cancer study being conducted at The University of Texas MD Anderson Cancer Center. None of them had been previously treated by surgery chemotherapy, and/or radiotherapy before enrollment into the study. All participants had completed a risk factor questionnaire that collected data on demographic characteristics, tobacco use history, occupational and environmental exposures, prior medical history, and any history of cancer in first-degree relatives, and also had donated a 40-ml blood sample for genotyping. We extracted the clinical information from the patients' medical records of their co-morbid conditions, tumor size, clinical stage, pathologic stage, histological type, tumor grade, treatment type, tumor recurrence, survival, and tumor progression for all the analyses. The median follow-up time was 11.8 months.

SNP selection and genotyping

A comprehensive panel of cancer-related genes including RGS gene family was identified and classified in each specific pathway according to their major reported functions. In the gene list, seventeen genes in the RGS family (RGS 1–5, 7–14, 16, 18, 20, and

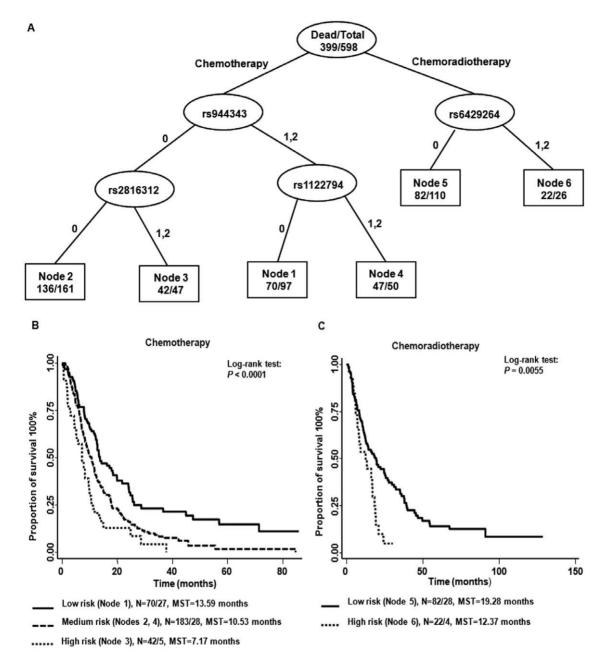


Figure 2. Survival tree analyses and Kaplan-Meier estimates of the significant RGS SNPs associated with survival. (A) Survival tree analyses; 0:homozygous wild type genotype; 1:heterozygous genotype; 2:homozygous variant genotype. (B) and (C) represent the survival curves of the risk group in chemotherapy group and chemoradiotherapy group; N: Number of Dead/Alive. MST: median survival time. doi:10.1371/journal.pone.0021120.g002

22) were selected for this panel. The detailed procedure for compiling the panel of genes and SNPs were reported previously [55]. Genomic DNA was extracted from the peripheral blood lymphocytes of the patients' blood samples, and all the genotyping work were performed according to the standard protocol provided by Illumina Inc. Then the results of genotyping were automatically generated by the Illumina's BeadStudio software. Finally, 95 SNPs in the RGS pathway were selected and successfully genotyped in these patients, as shown in **Table S1** in the Supporting Information.

Statistical analysis

STATA statistical software (StataCorp LP, College Station, TX) version 10.2 was used for the analysis of hazard ratios (HRs), P

values, median survival time (MST), P values for log-rank test and Kaplan-Meier survival estimate. χ^2 test (for categorical variables) and Student's t-test (for continuous variables) were used to assess differences in variables between dead and alive patients. For each SNP, the risk of death as a hazard ratio (HR) and 95% confidence interval (CI) were estimated with the Cox proportional hazards regression model. In addition, multivariate adjustment was used to control for potential confounding factors (age, gender, ethnicity, smoking status and pack-years, performance status, clinical stage, and treatment). For each SNP, the genetic distribution were assessed by three genetic models (dominant, recessive, and additive), and the model with the smallest P value was selected as the best-fitting model [56]. To validate the results, the bootstap

Table 6. Cox proportional hazards regression model in late-stage NSCLC patients based on the Survival tree analysis.

Risk Group	Dead (N%)	Alive (N%)	HR* (95%CI)	P-value	MST [#]	Log-rank P value
Chemotherapy						
Low (Node 1)	70 (72.16)	27 (27.84)	Reference		13.59	
Medium (Nodes 2, 4)	183 (86.73)	28 (13.27)	1.55 (1.16–2.09)	0.003	10.53	
High (Node3)	42 (89.36)	5 (10.64)	2.94 (1.93-4.47)	< 0.001	7.17	< 0.0001
Chemoradiation						
Low (Node 5)	82 (74.55)	28 (25.45)	Reference		19.28	
High (Node 6)	22 (84.62)	4 (15.38)	1.89 (1.06–3.38)	0.031	12.37	0.0055

*HR adjusted by age, gender, ethnicity, smoking status and pack-years, performance status, clinical stage. #MST, median survival time (months).

doi:10.1371/journal.pone.0021120.t006

resampling method was used. For each bootstrap sample drawn from the original data set, 100 bootstrap samples were generated. We obtained the *P* value for each SNP among the dominant, recessive, and additive models. The cumulative effects of different genotypes were calculated by summing up the individual effects of significant SNPs, that is, SNPs that showed significant association in single-SNP analysis and also had a bootstrap *P* value <0.05 at least 70 times. We used Cox proportional hazards regression model to estimate the HRs and 95% CIs. The Kaplan-Meier method and the log-rank test were used to estimate their effects on survival duration for these SNPs. Finally, the STREE program (http://masal.med.yale.edu/stree/) was used to perform survival tree analysis for the higher-order gene-gene interactions of the

SNPs. For these analyses, we only included SNPs that had been validated internally by bootstrapping. A two-sided P < 0.05 was considered statistically significant.

Supporting Information

Table S1 NPs in the RGS pathway. (DOC)

Author Contributions

Conceived and designed the experiments: XW. Performed the experiments: JD JG DC. Analyzed the data: JL. Contributed reagents/materials/analysis tools: CL DS JAR XW. Wrote the paper: JD XW.

References

- Parkin DM (2001) Global cancer statistics in the year 2000. Lancet Oncol 2: 533–543.
- Stinchcombe TE, Fried D, Morris DE, Socinski MA (2006) Combined modality therapy for stage III non-small cell lung cancer. Oncologist 11: 809–823.
- Fournel P, Robinet G, Thomas P, Souquet PJ, Lena H, et al. (2005) Randomized phase III trial of sequential chemoradiotherapy compared with concurrent chemoradiotherapy in locally advanced non-small-cell lung cancer: Groupe Lyon-Saint-Etienne d'Oncologie Thoracique-Groupe Francais de Pneumo-Cancerologie NPC 95-01 Study. J Clin Oncol 23: 5910–5917.
- Penland SK, Socinski MA (2004) Management of unresectable stage III nonsmall cell lung cancer: the role of combined chemoradiation. Semin Radiat Oncol 14: 326–334.
- van Meerbeeck JP, Koning CC, Tjan-Heijnen VC, Boekema AG, Kaandorp CJ, et al. (2005) [Guideline on 'non-small cell lung carcinoma; staging and treatment']. Ned Tijdschr Geneeskd 149: 72–77.
- Exton JH (1997) Cell signalling through guanine-nucleotide-binding regulatory proteins (G proteins) and phospholipases. Eur J Biochem 243: 10–20.
- Oldham WM, Hamm HE (2008) Heterotrimeric G protein activation by Gprotein-coupled receptors. Nat Rev Mol Cell Biol 9: 60–71.
- Hurst JH, Hooks SB (2009) Regulator of G-protein signaling (RGS) proteins in cancer biology. Biochem Pharmacol 78: 1289–1297.
- Manzur M, Hamzah J, Ganss R (2009) Modulation of g protein signaling normalizes tumor vessels. Cancer Res 69: 396–399.
- Berman DM, Gilman AG (1998) Mammalian RGS proteins: barbarians at the gate. J Biol Chem 273: 1269–1272.
 De Vries L, Zheng B, Fischer T, Elenko E, Farquhar MG (2000) The regulator
- of G protein signaling family. Annu Rev Pharmacol Toxicol 40: 235–271.

 12. Hollinger S, Hepler JR (2002) Cellular regulation of RGS proteins: modulators
- and integrators of G protein signaling. Pharmacol Rev 54: 527–559.
- 13. Burchett SA (2000) Regulators of G protein signaling: a bestiary of modular protein binding domains. J Neurochem 75: 1335–1351.
- Gu S, Cifelli C, Wang S, Heximer SP (2009) RGS proteins: identifying new GAPs in the understanding of blood pressure regulation and cardiovascular function. Clin Sci (Lond) 116: 391–399.
- Hendriks-Balk MC, Peters SL, Michel MC, Alewijnse AE (2008) Regulation of G protein-coupled receptor signalling: focus on the cardiovascular system and regulator of G protein signalling proteins. Eur J Pharmacol 585: 278–291.
- Riddle EL, Schwartzman RA, Bond M, Insel PA (2005) Multi-tasking RGS proteins in the heart: the next therapeutic target? Circ Res 96: 401–411.
- Traynor J Regulator of G protein-signaling proteins and addictive drugs. Ann N Y Acad Sci 1187: 341–352.

- Hurst JH, Mendpara N, Hooks SB (2009) Regulator of G-protein signalling expression and function in ovarian cancer cell lines. Cell Mol Biol Lett 14: 153–174.
- Grunebach F, Erndt S, Hantschel M, Heine A, Brossart P (2008) Generation of antigen-specific CTL responses using RGS1 mRNA transfected dendritic cells. Cancer Immunol Immunother 57: 1483–1491.
- Rangel J, Nosrati M, Leong SP, Haqq C, Miller JR, 3rd, et al. (2008) Novel role for RGS1 in melanoma progression. Am J Surg Pathol 32: 1207–1212.
- Furuya M, Nishiyama M, Kimura S, Suyama T, Naya Y, et al. (2004) Expression of regulator of G protein signalling protein 5 (RGS5) in the tumour vasculature of human renal cell carcinoma. J Pathol 203: 551–558.
- Rae FK, Stephenson SA, Nicol DL, Clements JA (2000) Novel association of a diverse range of genes with renal cell carcinoma as identified by differential display. Int J Cancer 88: 726–732.
- Han JI, Huang NN, Kim DU, Kehrl JH (2006) RGS1 and RGS13 mRNA silencing in a human B lymphoma line enhances responsiveness to chemoattractants and impairs desensitization. J Leukoc Biol 79: 1357–1368.
- Islam TC, Asplund AC, Lindvall JM, Nygren L, Liden J, et al. (2003) High level
 of cannabinoid receptor 1, absence of regulator of G protein signalling 13 and
 differential expression of Cyclin D1 in mantle cell lymphoma. Leukemia 17:
 1880–1890.
- Tsai CC, Huang KW, Chen HF, Zhan BW, Lai YH, et al. (2006) Gene expression analysis of human hepatocellular carcinoma by using full-length cDNA library. J Biomed Sci 13: 241–249.
- Chen X, Higgins J, Cheung ST, Li R, Mason V, et al. (2004) Novel endothelial cell markers in hepatocellular carcinoma. Mod Pathol 17: 1198–1210.
- Silva AP, Salim AC, Bulgarelli A, de Souza JE, Osorio E, et al. (2003) Identification of 9 novel transcripts and two RGSL genes within the hereditary prostate cancer region (HPC1) at 1q25. Gene 310: 49–57.
- Sood R, Bonner TI, Makalowska I, Stephan DA, Robbins CM, et al. (2001) Cloning and characterization of 13 novel transcripts and the human RGS8 gene from the 1q25 region encompassing the hereditary prostate cancer (HPC1) locus. Genomics 73: 211–222.
- Wiechec E, Overgaard J, Hansen LL (2008) A fragile site within the HPC1 region at 1q25.3 affecting RGS16, RGSL1, and RGSL2 in human breast carcinomas. Genes Chromosomes Cancer 47: 766–780.
- Smalley MJ, Iravani M, Leao M, Grigoriadis A, Kendrick H, et al. (2007) Regulator of G-protein signalling 2 mRNA is differentially expressed in mammary epithelial subpopulations and over-expressed in the majority of breast cancers. Breast Cancer Res 9: R85.



- 31. Nikolova DN, Zembutsu H, Sechanov T, Vidinov K, Kee LS, et al. (2008) Genome-wide gene expression profiles of thyroid carcinoma: Identification of molecular targets for treatment of thyroid carcinoma. Oncol Rep 20: 105-121.
- Tonjes A, Miedlich S, Holzapfel HP, Eszlinger M, Arkenau C, et al. (2004) Expression of regulators of g protein signaling mRNA is differentially regulated in hot and cold thyroid nodules. Thyroid 14: 896-901.
- 33. Hamzah J, Jugold M, Kiessling F, Rigby P, Manzur M, et al. (2008) Vascular normalization in Rgs5-deficient tumours promotes immune destruction. Nature
- 34. Schwable J, Choudhary C, Thiede C, Tickenbrock L, Sargin B, et al. (2005) RGS2 is an important target gene of Flt3-ITD mutations in AML and functions in myeloid differentiation and leukemic transformation. Blood 105: 2107-2114.
- Koga H, Imada K, Ueda M, Hishizawa M, Uchiyama T (2004) Identification of differentially expressed molecules in adult T-cell leukemia cells proliferating in vivo. Cancer Sci 95: 411-417.
- 36. Tatenhorst L, Senner V, Puttmann S, Paulus W (2004) Regulators of G-protein signaling 3 and 4 (RGS3, RGS4) are associated with glioma cell motility. J Neuropathol Exp Neurol 63: 210-222.
- 37. Gu J, Wu X, Dong Q, Romeo MJ, Lin X, et al. (2006) A nonsynonymous singlenucleotide polymorphism in the PDZ-Rho guanine nucleotide exchange factor (Ser1416Gly) modulates the risk of lung cancer in Mexican Americans. Cancer 106: 2716-2724.
- 38. Wang J, Lippman SM, Lee JJ, Yang H, Khuri FR, et al. (2010) Genetic variations in regulator of G-protein signaling genes as susceptibility loci for second primary tumor/recurrence in head and neck squamous cell carcinoma. Carcinogenesis 31: 1755-1761.
- 39. You M, Wang D, Liu P, Vikis H, James M, et al. (2009) Fine mapping of chromosome 6q23-25 region in familial lung cancer families reveals RGS17 as a likely candidate gene. Clin Cancer Res 15: 2666-2674.
- 40. Berman DM, Wang Y, Liu Z, Dong Q, Burke LA, et al. (2004) A functional polymorphism in RGS6 modulates the risk of bladder cancer. Cancer Res 64: 6820-6826
- 41. Hildebrandt MA, Gu J, Wu X (2009) Pharmacogenomics of platinum-based chemotherapy in NSCLC. Expert Opin Drug Metab Toxicol 5: 745-755.
- 42. Sliva D (2004) Signaling pathways responsible for cancer cell invasion as targets
- for cancer therapy. Curr Cancer Drug Targets 4: 327–336. Sarkar FH, Li Y (2004) Cell signaling pathways altered by natural chemopreventive agents. Mutat Res 555: 53-64.
- 44. Puiffe ML, Le Page C, Filali-Mouhim A, Zietarska M, Ouellet V, et al. (2007) Characterization of ovarian cancer ascites on cell invasion, proliferation,

- spheroid formation, and gene expression in an in vitro model of epithelial ovarian cancer. Neoplasia 9: 820-829.
- 45. Albig AR, Schiemann WP (2005) Identification and characterization of regulator of G protein signaling 4 (RGS4) as a novel inhibitor of tubulogenesis: RGS4 inhibits mitogen-activated protein kinases and vascular endothelial growth factor signaling. Mol Biol Cell 16: 609-625.
- 46. Xiao B, Zhang Y, Niu WQ, Gao PJ, Zhu DL (2009) Haplotype-based association of regulator of G-protein signaling 5 gene polymorphisms with essential hypertension and metabolic parameters in Chinese. Clin Chem Lab Med 47: 1483-1488.
- 47. Boss CN, Grunebach F, Brauer K, Hantschel M, Mirakaj V, et al. (2007) Identification and characterization of T-cell epitopes deduced from RGS5, a novel broadly expressed tumor antigen. Clin Cancer Res 13: 3347-3355.
- Siderovski DP, Willard FS (2005) The GAPs, GEFs, and GDIs of heterotrimeric G-protein alpha subunits. Int J Biol Sci 1: 51-66.
- 49. Ishii M, Kurachi Y (2003) Physiological actions of regulators of G-protein signaling (RGS) proteins. Life Sci 74: 163-171.
- Ooe A, Kato K, Noguchi S (2007) Possible involvement of CCT5, RGS3, and YKT6 genes up-regulated in p53-mutated tumors in resistance to docetaxel in human breast cancers. Breast Cancer Res Treat 101: 305-315
- 51. Takahashi H, Nemoto T, Yoshida T, Honda H, Hasegawa T (2006) Cancer diagnosis marker extraction for soft tissue sarcomas based on gene expression profiling data by using projective adaptive resonance theory (PART) filtering method. BMC Bioinformatics 7: 399.
- 52. Martinez-Cardus A. Martinez-Balibrea E. Bandres E. Malumbres R. Gines A. et al. (2009) Pharmacogenomic approach for the identification of novel determinants of acquired resistance to oxaliplatin in colorectal cancer. Mol Cancer Ther 8: 194-202.
- 53. Benzing T, Brandes R, Sellin L, Schermer B, Lecker S, et al. (1999) Upregulation of RGS7 may contribute to tumor necrosis factor-induced changes in central nervous function. Nat Med 5: 913-918.
- Balkwill F (2009) Tumour necrosis factor and cancer. Nat Rev Cancer 9: 361-371.
- 55. Wu X, Spitz MR, Lee JJ, Lippman SM, Ye Y, et al. (2009) Novel susceptibility loci for second primary tumors/recurrence in head and neck cancer patients: large-scale evaluation of genetic variants. Cancer Prev Res (Phila) 2: 617-624.
- Zheng SL, Sun J, Wiklund F, Smith S, Stattin P, et al. (2008) Cumulative association of five genetic variants with prostate cancer. N Engl J Med 358: 910-919.