



Draft Genome Sequence of *Diplodia seriata* F98.1, a Fungal Species Involved in Grapevine Trunk Diseases

Guillaume Robert-Siegwald,^a Julie Vallet,^b Eliane Abou-Mansour,^c Jiabao Xu,^d Patrice Rey,^e Christophe Bertsch,^f Cecilia Rego,^g Philippe Larignon,^h Florence Fontaine,^b Marc-Henri Lebrun^a

UMR 1290 BIOGER, INRA, AgroParisTech, Campus AgroParistech, Thiverval-Grignon, France^a; Université de Reims Champagne-Ardenne, URVVC EA 4707, Laboratoire Stress, Défenses et Reproduction des Plantes, Reims, France^b; Plant Biology Department, University of Fribourg, Fribourg, Switzerland^c; BGI-Shenzhen, Shenzhen, China^d; UMR1065 SAVE, Santé et Agroécologie du Vignoble, INRA, Bordeaux Sciences Agro, Villenave d'Ornon, France^e; Université Haute Alsace, Laboratoire Vigne Biotechnologie et Environnement EA 3991, Colmar, France^f; Institut Supérieur d'Agronomie, Tapada da Ajuda, Lisbon, Portugal^g; Institut Français de la Vigne et du Vin Pôle Rhône-Méditerranée, Rodilhan, France^h

ABSTRACT The ascomycete *Diplodia seriata* is a causal agent of grapevine trunk diseases. Here, we present the draft genome sequence of *D. seriata* isolate F98.1 (37.27 Mb, 512 contigs, 112 scaffolds, and 8,087 predicted protein-coding genes).

Diplodia seriata (1, 2) is one of the most common species in the family *Botryosphaeriaceae* that is associated with grapevine trunk diseases (3, 4). In grapevine, *D. seriata* is frequently isolated from necrotic tissues (black spots or sectors) localized in the wood of infected trunks or arms (3–5). This species is also frequently isolated from woody tissues of trees such as *Acer* sp., *Prunus* sp., and *Quercus* sp. (2). Pycnidia are produced on infected grapevine wood or pruning shoots, and liberate pycnidiospores dispersed by rainfall or sprinkler irrigation (6, 7). On grapevine surfaces, pycnidiospores germinate and infectious hyphae penetrate into the plant tissues through pruning wounds (4, 5). Inoculation of *D. seriata* on wounded stems from grafted grapevines of the highly susceptible cultivar tempranillo induces a local brown necrosis in all plants and foliar symptoms in 50% of plants (8). *D. seriata* is known to secrete several plant polymers degrading enzymes such as cellulases, xylanases, lipases, and laccases (9, 10). This fungal species also produces phytotoxic secondary metabolites such as (–)-mellein and its derivatives (11, 12).

D. seriata F98.1 was isolated in 1998 at Perpignan, France, from the trunk of a grapevine exhibiting foliar symptoms typical of Syrah decline (13). *D. seriata* F98.1 is pathogenic on grapevine (8, 13). Sequencing was performed by BGI-Tech (China) using Illumina HiSeq 2500 at a coverage of 270× (170-, 500-, and 6,000-bp libraries). After quality filtering, a total of 88,596,792 paired-end reads of 125 bp were obtained. Assembly with SOAPdenovo version 1.05 (14) led to 512 contigs and 112 scaffolds (37.27 Mb; G+C%: 56.8). This high-quality genome (scaffold N_{50} : 2.9 Mb; minimum scaffold length: 1,007 bp; gaps: 250 kb) has 13 scaffolds with a size greater than 1 Mb (90% of the total sequence), likely corresponding to chromosomes. Using GLEAN (15), 8,087 coding sequences (CDSs) were identified, 93% being supported by RNAseq (mycelium on potato dextrose broth or minimal medium for 4 days). Recently, the genome sequence of *D. seriata* DS831, isolated from an infected grapevine (United States, 2011), was released (16). The genome size of DS831 (37.13 Mb) is similar to the genome size of F98.1, but its assembly is five times more fragmented (1,391 contigs; 695 scaffolds), and it carries 9,398 CDSs. Bidirectional best BLAST hit (BDBH) analysis revealed that 82% of F98.1's and DS831's CDSs are similar. OrthoFinder (17) identified

Received 23 January 2017 Accepted 2 February 2017 Published 6 April 2017

Citation Robert-Siegwald G, Vallet J, Abou-Mansour E, Xu J, Rey P, Bertsch C, Rego C, Larignon P, Fontaine F, Lebrun M-H. 2017. Draft genome sequence of *Diplodia seriata* F98.1, a fungal species involved in grapevine trunk diseases. *Genome Announc* 5:e00061-17. <https://doi.org/10.1128/genomeA.00061-17>.

Copyright © 2017 Robert-Siegwald et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Marc-Henri Lebrun, marc-henri.lebrun@inra.fr.

F.F. and M.-H.L. contributed equally to this work.

5,935 orthologous single-copy gene families shared between the two genomes. According to BDBH analysis, 1,507 genes are specific to strain F98.1, and 2,763 are specific to strain DS831. Using BLASTN, we found that 2,686 (97.2%) of the genes thought to be specific to strain DS831 were present in the F98.1 genome sequence, and 1,440 (95.6%) of the genes thought to be specific to strain F98.1 are present in the DS831 genome sequence. The difference in CDS numbers between DS831 and F98.1 is likely a consequence of using different annotation software (GLEAN versus Augustus). Fusing the two annotations produces a set of 10,773 CDSs (8,087 from F98.1 and 2,686 from DS831).

Accession number(s). This whole-genome project has been deposited at NCBI GenBank under the accession number [MSZU00000000](https://www.ncbi.nlm.nih.gov/nuccore/MSZU00000000). The version described in this paper is the first version, MSZU01000000.

ACKNOWLEDGMENTS

This project was funded by grants V1301 and V1302 from CASDAR (Compte d’Affectation Spéciale au Développement Agricole et Rural, Ministère de l’Agriculture, France) and CNIV (Comité National Interprofessionnel des Vins, France).

REFERENCES

- Phillips AJL, Crous PW, Alves A. 2007. *Diplodia seriata*, the anamorph of “*Botryosphaeria*” *obtusa*. *Fungal Divers* 25:141–155.
- Phillips AJL, Alves A, Abdollahzadeh J, Slippers B, Wingfield MJ, Groenewald JZ, Crous PW. 2013. The *Botryosphaeriaceae*: genera and species known from culture. *Stud Mycol* 76:51–167. <https://doi.org/10.3114/sim0021>.
- Urbez-Torres JR. 2011. The status of *Botryosphaeriaceae* species infecting grapevines. *Phytopathologia Mediterr* 50:5–45.
- Bertsch C, Ramírez-Suero M, Magnin-Robert M, Larignon P, Chong J, Abou-Mansour E, Spagnolo A, Clément C, Fontaine F. 2013. Grapevine trunk diseases: complex and still poorly understood. *Plant Pathol* 62:243–265. <https://doi.org/10.1111/j.1365-3059.2012.02674.x>.
- Larignon P. 2011. Les maladies du bois de la vigne. Quelques éléments sur la biologie de deux champignons associés, *Phaeoacremonium aleophilum* & *Diplodia seriata*. *Phytoma* 646:41–44.
- Kuntzmann P, Villaume S, Bertsch C. 2009. Conidia dispersal of *Diplodia* species in a French vineyard. *Phytopathologia Mediterr* 48:150–154.
- Urbez-Torres JR, Peduto F, Gubler WD. 2010. First report of grapevine cankers caused by *Lasiodiplodia crassispora* and *Neofusicoccum mediterraneum* in California. *Plant Dis* 94:785–785. <https://doi.org/10.1094/PDIS-94-6-0785B>.
- Reis P, Magnin-Robert M, Nascimento T, Spagnolo A, Abou-Mansour E, Fioretti C, Clément C, Rego C, Fontaine F. 2016. Reproducing *Botryosphaeria* dieback foliar symptoms in a simple model system. *Plant Dis* 100:1071–1079. <https://doi.org/10.1094/PDIS-10-15-1194-RE>.
- Bénard-Gellon M, Farine S, Goddard ML, Schmitt M, Stempien E, Pensec F, Laloue H, Mazet-Kieffer F, Fontaine F, Larignon P, Chong J, Tarnus C, Bertsch C. 2015. Toxicity of extracellular proteins from *Diplodia seriata* and *Neofusicoccum parvum* involved in grapevine *Botryosphaeria* dieback. *Protoplasma* 252:679–687. <https://doi.org/10.1007/s00709-014-0716-y>.
- Esteves AC, Saraiva M, Correia A, Alves A. 2014. *Botryosphaeriales* fungi produce extracellular enzymes with biotechnological potential. *Can J Microbiol* 60:332–342. <https://doi.org/10.1139/cjm-2014-0134>.
- Djoukeng JD, Polli S, Larignon P, Abou-Mansour E. 2009. Identification of phytotoxins from *Botryosphaeria obtusa*, a pathogen of black dead arm disease of grapevine. *Eur J Plant Pathol* 124:303–308. <https://doi.org/10.1007/s10658-008-9419-6>.
- Andolfi A, Mugnai L, Luque J, Surico G, Cimmino A, Evidente A. 2011. Phytotoxins produced by fungi associated with grapevine trunk diseases. *Toxins (Basel)* 3:1569–1605. <https://doi.org/10.3390/toxins3121569>.
- Larignon P, Fulchic R, Cere L, Dubos B. 2001. Observation on black dead arm in French vineyards. *Phytopathologia Mediterr* 40:336–342.
- Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, Tang J, Wu G, Zhang H, Shi Y, Liu Y, Yu C, Wang B, Lu Y, Han C, Cheung DW, Yiu SM, Peng S, Xiaoqian Z, Liu G, Liao X, Li Y, Yang H, Wang J, Lam TW, Wang J. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. *Gigascience* 1:18. <https://doi.org/10.1186/2047-217X-1-18>.
- Elsik CG, Mackey AJ, Reese JT, Milshina NV, Roos DS, Weinstock GM. 2007. Creating a honey bee consensus gene set. *Genome Biol* 8:R13. <https://doi.org/10.1186/gb-2007-8-1-r13>.
- Morales-Cruz A, Amrine KC, Blanco-Ulate B, Lawrence DP, Travadon R, Rolshausen PE, Baumgartner K, Cantu D. 2015. Distinctive expansion of gene families associated with plant cell wall degradation, secondary metabolism, and nutrient uptake in the genomes of grapevine trunk pathogens. *BMC Genomics* 16:469. <https://doi.org/10.1186/s12864-015-1624-z>.
- Emms DM, Kelly S. 2015. OrthoFinder: solving fundamental biases in whole genome comparisons dramatically improves orthogroup inference accuracy. *Genome Biol* 16:157. <https://doi.org/10.1186/s13059-015-0721-2>.