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CLINICAL RESEARCH

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MONITOR

Elevated Plasma P-Selectin Autoantibodies in Primary Sjögren Syndrome Patients with Thrombocytopenia

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	kground:	thrombocytopenia is one of the hematological manif activated, P-selectin is expressed on the cell surface. toantibodies in the pathogenesis of thrombocytopen	•			
Material/N	Nethods:		e-linked immunosorbent assay (ELISA) in 38 pSS patients thrombocytopenia, 32 idiopathic thrombocytopenic pur-			
	Results:	The plasma P-selectin autoantibodies (A_{490}) in ITP pa were significantly higher than those in healthy contro- patients and pSS patients with thrombocytopenia. T tients with thrombocytopenia was significantly higher in P-selectin autoantibodies-positive patients, while a count was lower in P-selectin autoantibodies-positive	tients and pSS patients with/without thrombocytopenia ols, but there were no significant differences between ITP The positive rate of P-selectin autoantibodies in pSS pa- er than that in ITP patients. The platelet count was lower among pSS patients with thrombocytopenia, the platelet e patients than in P-selectin autoantibodies-negative pa- bocytopenia, the platelet count was lower in P-selectin au-			
Conclusions:		Elevated plasma P-selectin autoantibodies may play a role in the pathogenesis of thrombocytopenia in pSS patients.				
MeSH Keywords:		Autoantibodies, P-Selectin • Endothelial Cells • Purpura, Thrombocytopenic, Idiopathic • Sjögren's Syndrome, Primary • Thrombocytopenia				
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Background

Primary Sjögren's syndrome (pSS) is one of the most common chronic systemic autoimmune diseases. It primarily affects the salivary and lachrymal glands, and is characterized by the classical symptoms of oral and ocular dryness [1,2], along with widespread pain and intense fatigue. However, a significant proportion of patients develop extraglandular systemic manifestations involving several organs (e.g., central nervous system, lung, liver, and kidney). pSS has been estimated to affect 0.01–0.1% of the general population [3]. Secondary SS (sSS) is usually associated with other rheumatic conditions, such as rheumatoid arthritis (RA) and systemic lupus erythematosus (SLE), of which the most common is RA [4-6]. As a systemic autoimmune disease, B cells play an important role in the pathogenesis of pSS [7]. There are several autoantibodies present during pSS [8], and antinuclear antibodies (ANA) are the most common, detectable in up to 80% of pSS patients. However, the most specific autoantibodies in pSS are the intracellular antigens Ro52/TRIM21, Ro60/TROVE2, and La/SSB ribonucleoproteins [9]. These autoantibodies are detectable in patients several years before the first clinical manifestation of pSS, and may be used to predict the disease [10]. Rheumatoid factors (RF) are also frequently found in these patients, and are often associated with higher disease activity [8]. Hematological manifestations are also noted in pSS patients and include amenia, leukopenia, and thrombocytopenia [11]. Previous studies have reported that thrombocytopenia is present in 5-7% of pSS patients [12]; it is an isolated manifestation or a component of pancytopenia, but life-threatening, severe thrombocytopenia is very rare.

The cause of thrombocytopenia in pSS patients is still poorly understood. It has been suggested that thrombocytopenia in pSS involves a humoral autoimmune mechanism in which an antibody produced is able to bind to the cell membrane [13]. Idiopathic thrombocytopenic purpura (ITP) is also an autoimmune disorder characterized by thrombocytopenia [14]. It has been demonstrated that platelet autoantibodies are detectable in ITP patients. Platelet autoantibodies often recognize platelet glycoproteins (GP) such as GPIb, GP IX, GPIIb/IIIa, and P-selectin [15,16]. Platelet-derived microparticles could highlight platelet activation in pSS [17]. P-selectin is a member of the selectin family of cell-adhesion molecules, and may modulate the interaction of leukocytes and platelets with the endothelium [18]. P-selectin is stored in the granules of endothelial cells and platelets; it is expressed on their surface after activation, and shed into the plasma in the soluble form. P-selectin plays an important role in thrombosis and prothrombotic states [19-22]. P-selectin autoantibodies may affect the functions of platelets and endothelial cells. pSS is a chronic inflammatory systemic autoimmune disease with endothelial injury.

In the present study, enzyme-linked immunosorbent assay (ELISA) was used to detect the plasma P-selectin autoantibodies, and the role of P-selectin autoantibodies in the thrombocytopenia of pSS patients was evaluated. To the best of our knowledge, this is the first study to measure P-selectin autoantibodies in pSS patients.

Material and Methods

Patients and healthy controls

A retrospective study was conducted from July 1, 2012, to July 1, 2014. Seventy pSS patients were treated in our department and pSS was diagnosed according to the criteria of the American-European Consensus Group for pSS in 2002 [23]. Patients with secondary Sjögren's syndrome (sSS) with any other autoimmune diseases, such as RA, SLE, and other connective tissue diseases, were excluded from this study. Of the 70 pSS patients, 32 pSS patients had thrombocytopenia, and 38 pSS patients had no thrombocytopenia. In addition, ITP patients (n=32) were also enrolled as controls and ITP was diagnosed according to the criteria for ITP [24]. Moreover, 35 healthy subjects were recruited as healthy controls. This study was approved by the Ethics Committee of The First Affiliated Hospital of Soochow University, and all participants gave their written informed consent before study. The plasma samples were collected from all the subjects and stored at -80°C until use.

Detection of plasma P-selectin autoantibodies

We pre-coated 96-well Microtiter plates with goat anti-mouse polyclonal antibody at 5 µg/µL (100 µL/well) at 4°C overnight. These plates were washed 3 times with phosphate-buffered saline (PBS) containing 0.05% Tween 20 (PBS-T), and treated with blocking buffer (1% bovine serum albumin [BSA] in PBS-T) at 4°C overnight. After washing 3 times, P-selectin monoclonal antibody at 10 µg/ml was added to the plates followed by incubation overnight at 4°C. The plates were blocked on the second day, and then washed for use. We added 1×109/ml lysed platelets diluted at 1: 5 to the plates (100 µl/well) followed by incubation at 37°C for 2 h. After washing with PBS-T, the diluted plasma samples (1:5, 100 µl/well) were added, followed by incubation at 37°C for 1.5 h. After washing 3 times, 100 µl of rabbit anti-human IgG conjugated to horseradish peroxidase was added to each well followed by incubation at 37°C for 1 h. After washing 6 times, TMB was added and incubated for 10 min, and the reaction was stopped by addition of 3M sulfuric acid. Then, the optical density (OD) was measured at 490 nm with a microplate reader. All measurements were performed in duplicate. The mean OD +2 fold of standard deviation from the plasma samples of healthy controls was considered as the upper limit of normal. It was defined as positive Table 1. Demographic features of subjects enrolled in this study.

	Number	Gender, n (%)		Median age,	
		Male	Female	y (range)	
pSS	70	6 (8.6)	64 (91.4)	42 (20–60)	
Without thrombocytopenia	38	4 (10.5)	34 (89.5)	43 (20–53)	
With thrombocytopenia	32	2 (6.3)	30 (93.7)	42 (23–60)	
ITP	32	7 (21.9)	25 (78.1)	36 (19–54)	
Normal control	35	18 (51.4)	17 (48.6)	39 (18–59)	

if mean OD was higher than the upper limit of normal; otherwise, it was defined as negative.

Blood platelet counting

Blood platelet count was determined with an XE-5000 multiparameter automatic hematology analyzer (Sysmex, Kobe, Japan).

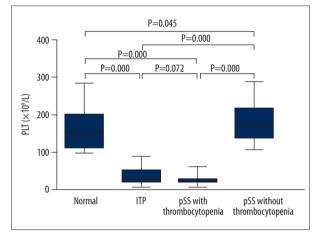
Statistical analysis

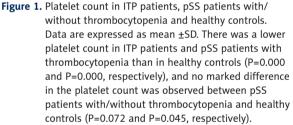
Statistical analysis was done with SPSS version 18.0 for Windows. Data are presented as means \pm standard deviation (SD). Data were compared with *t*-test between 2 groups. A value of *P*<0.05 was considered statistically significant.

Results

Patient characteristics

A total of 102 patients (13 males and 89 females) and 35 healthy controls (18 males and 17 females) were enrolled into the present study between 2012 and 2014. There were 70 pSS patients (6 males and 64 females) and 32 ITP patients (7 males and 25 females). Of the 70 pSS patients, 38 had no thrombocytopenia (4 males and 34 females) and the other 32 had concomitant thrombocytopenia (2 males and 30 females). The demographic characteristics of subjects included are summarized in Table 1. There were no significant differences in sex and age between pSS patients with thrombocytopenia and ITP patients. We analyzed the platelet count of patients and normal controls (Figure 1). The platelet count in ITP patients and pSS patients with thrombocytopenia (35.7±22.6×10⁹/L and 27.5±11.1×10⁹/L, respectively) was lower than that in healthy controls (157.3±52.5×10⁹/L) (P=0.000 and P=0.000, respectively), and there was no significant difference in the platelet count between pSS patients without thrombocytopenia (181.6±49.1×10⁹/L) and healthy controls. There was no significant difference in the platelet count between pSS patients with thrombocytopenia and ITP controls (P=0.072).





Plasma P-selectin autoantibodies

P-selectin autoantibodies were measured in pSS patients with/ without thrombocytopenia, ITP patients, and healthy controls (Figure 2). The plasma P-selectin autoantibodies in ITP patients, pSS patients with thrombocytopenia, and pSS patients without thrombocytopenia (0.796 ± 0.097 , 0.854 ± 0.142 , and 0.738 ± 0.090 , respectively) was significantly higher than that in healthy controls (0.684 ± 0.110) (P=0.000, P=0.000, and P=0.024, respectively), but there was no significant difference in the P-selectin autoantibodies between ITP patients and pSS patients with thrombocytopenia (P=0.062).

Proportion of P-selectin autoantibodies-positive patients

The proportion of P-selectin autoantibodies-positive patients was also calculated (Figure 3). Of all the patients,

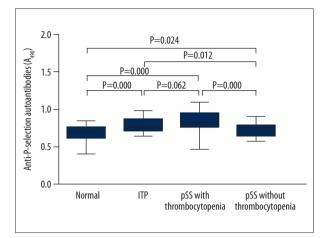


Figure 2. Plasma P-selectin autoantibodies (A490) in ITP patients, pSS patients with/without thrombocytopenia and healthy controls. Data are expressed as mean ±SD. The plasma P-selectin autoantibodies in ITP patients, pSS patients with thrombocytopenia and pSS patients without thrombocytopenia was significantly higher than in healthy controls (P=0.000, P=0.000 and P=0.024, respectively), but there was no significant difference in the P-selectin autoantibodies between ITP patients and pSS patients with thrombocytopenia (P=0.062).

20.6% (21/102) were positive for P-selectin autoantibodies. The proportion of P-selectin autoantibodies-positive patients was 15.63% (5/32), 40.63% (13/32), and 7.89% (3/38) in ITP patients, pSS patients with thrombocytopenia, and pSS patients without thrombocytopenia, respectively. The proportion of P-selectin autoantibodies-positive patients in ITP patients and pSS patients with thrombocytopenia was markedly higher than that in healthy controls (P=0.015 and P=0.000, respectively), but no significant difference was observed between healthy controls and pSS patients without thrombocytopenia (P=0.090), and the positive rate in pSS patients with thrombocytopenia was also comparable to that in ITP patients (P=0.05). The anti-P-selectin autoantibodies-positive rate in pSS patients without thrombocytopenia (P=0.001).

Relationship between P-selectin autoantibodies and platelet count

The relationship between P-selectin autoantibodies and platelet count was further evaluated in ITP patients and pSS patients with/without thrombocytopenia (Figure 4). There was a lower platelet count in P-selectin autoantibodies-positive patients ($44.4\pm10.7\times10^{9}/L$) than in P-selectin autoantibodies-negative patients ($98.6\pm80.9\times10^{9}/L$) (P=0.002) (Figure 4A). In ITP patients and pSS patients with thrombocytopenia, the platelet count in P-selectin autoantibodies-positive patients ($20.8\pm6.1\times10^{9}/L$) was lower than in anti-P-selectin autoantibodies-negative

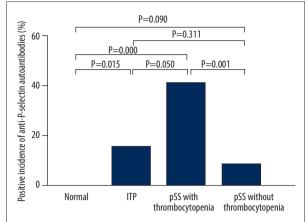


Figure 3. Proportion of P-selectin autoantibodies-positive patients. The proportion of P-selectin autoantibodiespositive patients in ITP patients, pSS patients with thrombocytopenia and pSS patients without thrombocytopenia was 15.63% (5/32), 40.63% (13/32) and 7.89% (3/38), respectively. There was significantly higher positive rate in ITP patients and pSS patients with thrombocytopenia than in healthy controls (P=0.015 and P=0.000, respectively), but no difference was observed between healthy controls and pSS patients without thrombocytopenia (P=0.090).

patients $(35.9\pm 19.5\times 10^9/L)$ (P=0.000) (Figure 4B). There was a lower platelet count in P-selectin autoantibodies-positive pSS patients $(53.1\pm 67.5\times 10^9/L)$ than in P-selectin autoantibodiesnegative pSS patients $(128.4\pm 83.1\times 10^9/L)$ (P=0.002) (Figure 4C), while the platelet count in P-selectin autoantibodies-positive patients $(22.3\pm 5.2\times 10^9/L)$ was markedly lower than in P-selectin autoantibodies-negative patients $(31.1\pm 12.7\times 10^9/L)$ (P=0.012) among pSS patients with thrombocytopenia (Figure 4D).

Discussion

The present study demonstrated that the plasma P-selectin autoantibodies significantly increased in pSS patients with thrombocytopenia as compared to ITP patients, pSS patients without thrombocytopenia, and healthy controls. Additionally, the proportion of P-selectin autoantibodies-positive patients was markedly higher in pSS patients with thrombocytopenia, and being positive for P-selectin autoantibodies was associated with a low platelet count, indicating that P-selectin autoantibodies may be involved in the destruction of platelets and the pathogenesis of thrombocytopenia in pSS.

pSS is a systemic autoimmune disease characterized by inflammation that can impair multiple organs, and the impairment of the hematopoietic system is common. Both T cells [25] and B cells [7] play important roles in the development of pSS. B-cell hyperactivity is a major characteristic of

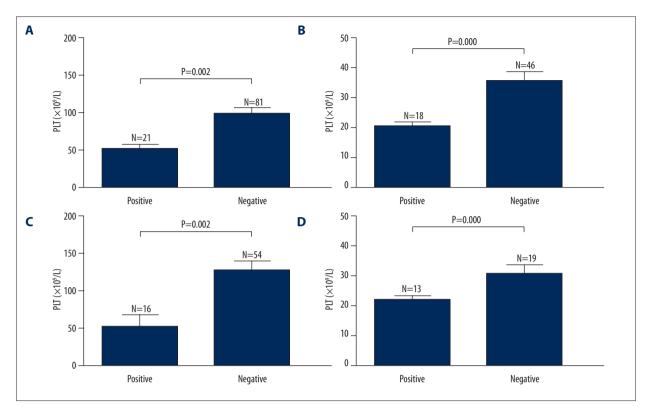


Figure 4. Platelet count in anti-P-selectin autoantibodies-positive/negative patients with ITP and pSS. (A) Platelet count in P-selectin autoantibodies-positive/negative patients with ITP and pSS. There was a lower platelet count in P-selectin autoantibodies-positive patients than in P-selectin autoantibodies-negative patients (P=0.002). (B) Platelet count in P-selectin autoantibodies-positive/negative ITP patients and pSS patients with thrombocytopenia. There was a lower platelet count in P-selectin autoantibodies-positive/negative patients than in P-selectin autoantibodies-negative patients (P=0.002). (B) Platelet count in P-selectin autoantibodies-positive/negative ITP patients and pSS patients with thrombocytopenia. There was a lower platelet count in P-selectin autoantibodies-negative patients (P=0.000) among ITP patients and pSS patients with thrombocytopenia. (C) Platelet count in P-selectin autoantibodies-positive/negative pSS patients. There was a lower platelet count in P-selectin autoantibodies-positive/negative pSS patients. There was a lower platelet count in P-selectin autoantibodies-positive/negative pSS patients (P=0.002). (D) Platelet count in P-selectin autoantibodies-positive/negative pSS patients with thrombocytopenia. There was a lower platelet count in P-selectin autoantibodies-positive/negative pSS patients with thrombocytopenia. There was a lower platelet count in P-selectin autoantibodies-positive/negative pSS patients with thrombocytopenia. There was a lower platelet count in P-selectin autoantibodies-positive/negative pSS patients with thrombocytopenia. There was a lower platelet count in P-selectin autoantibodies-positive/negative pSS patients with thrombocytopenia. There was a lower platelet count in P-selectin autoantibodies-positive patients than in anti-P-selectin autoantibodies-positive patients than in anti-P-selectin autoantibodies-negative patients (P=0.012).

pSS and is predominantly characterized by the production of a series of autoantibodies against various autoantigens, such as Ro52/TRIM21, Ro60/TROVE2, La/SSB, U1RNP, RF, cryoglobulins, centromere (ACA), mitochondria (AMA), smooth muscle, cyclic citrullinated peptides (anti-CCP), and others [26]. ANA is found in 59-85% of pSS patients [27], but is not specific for pSS. Ro/SSA antibody and La/SSB antibody are the major autoantibodies in pSS and can be used in the diagnosis of pSS [23]. Ro/SSA antibodies represent 2 distinct entities of autoantibodies: Ro52/TRIM21 antibodies and Ro60/TROVE2 antibodies. Ro52/TRIM21 antibodies are the most common specific autoantibody, and are detectable in 66.7% of pSS patients [28]. Ro60/TROVE2 antibodies and La/SSB antibodies are found in 52.1% and 49% of pSS patients, respectively [29]. RF is another family of autoantibodies and is present in 36–74% of pSS patients [6,30,31].

As a member of the selectin family, P-selectin is stored in the granules of platelets and the Weibel-Palade bodies of endothelial cells [32,33]. When the platelets and endothelial cells are activated, P-selectin is expressed on the cell surface and released into the plasma as a soluble form [34]. Increased soluble P-selectin is related to thrombotic diseases [20] and ITP is also an autoimmune disorder; several autoantibodies against GP such as GPIb, GP IX, GPIIb/IIIa, and P-selectin have been identified in ITP patients [16]. P-selectin autoantibodies may lead to platelet destruction and impair the platelet function. Although there is a role of P-selectin autoantibodies in the pathogenesis of ITP, the association between P-selectin autoantibodies antibodies and pSS with thrombocytopenia remains unknown.

Our results demonstrated that the plasma P-selectin autoantibodies were significantly higher in pSS patients with thrombocytopenia than in ITP patients. P-selectin autoantibodies-positive rate in pSS patients with thrombocytopenia was also markedly higher than in ITP patients. Moreover, there was a lower platelet count in P-selectin autoantibodies-positive patients. There were both endothelial injury and platelet

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tion. Therefore, the increase in P-selectin autoantibodies in pSS patients with thrombocytopenia was more obvious than in ITP patients, and the platelet count in P-selectin autoantibodies-positive pSS patients with thrombocytopenia was lower than in ITP patients. Our results suggest that the increased plasma P-selectin autoantibodies may contribute to the platelet destruction in pSS patients and finally result in thrombocytopenia. To the best of our knowledge, this is the first report on the role of P-selectin autoantibodies in the thrombocytopenia of pSS patients.

There were several limitations in this study. First, this was a retrospective study and patients were not followed up. Second,

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only patients older than 18 years were enrolled; therefore, the results might not be generalizable to children. Third, the sample size was small and patients were enrolled from a single center.

Conclusions

We for the first time report the role of P-selectin autoantibodies in the thrombocytopenia of pSS patients. P-selectin autoantibodies may lead to platelet destruction and endothelial injury and play a role in the pathogenesis of thrombocytopenia in pSS. Further prospective studies are still needed to understand the potential mechanism underlying the role of P-selectin autoantibodies in thrombocytopenia of pSS patients.

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