

Biological Roles and Clinical Therapeutic Applications of Tumor-Associated Macrophages in Colorectal Liver Metastasis

Shenghao Li^{1,2,*}, Liyuan Hao^{1,2,*}, Xiaoyu Hu²

¹School of Clinical Medicine, Chengdu University of Traditional Chinese Medicine, Chengdu, Sichuan Province, People's Republic of China;

²Department of Infectious Diseases, Hospital of Chengdu University of Traditional Chinese Medicine, Chengdu, Sichuan Province, People's Republic of China

*These authors contributed equally to this work

Correspondence: Xiaoyu Hu, Hospital of Chengdu University of Traditional Chinese Medicine, Chengdu, Sichuan Province, People's Republic of China, Email xiaoyuhu202206@163.com

Abstract: Colorectal cancer (CRC) commonly metastasizes to the liver, and this poses a significant clinical challenge. Tumor-associated macrophages (TAMs), key players within the TME, play a significant role in promoting CRC metastasis by secreting various chemokines, growth factors, and cytokines. This review not only aims to enhance our knowledge of TAMs' functions in CRC progression and metastasis but also examines innovative therapeutic strategies to address the clinical problem of colorectal liver metastasis (CLM). By targeting TAMs, we may be able to develop more effective treatments and offer hope to patients suffering from this devastating disease.

Keywords: colorectal liver metastasis, macrophage, TAMs

Introduction

Colorectal cancer (CRC) ranks as the third most prevalent cancer worldwide and is the second leading cause of cancer-related mortality.¹ While early-stage CRC can often be effectively managed with radical interventions,² a significant portion of patients (25–50%) with early-stage disease eventually develop metastatic CRC,³ with the liver being the most common site for metastasis.⁴ The high incidence of liver metastasis in CRC can be attributed to the liver's anatomical position, as it receives blood from the gastrointestinal tract via the portal vein, facilitating the spread of CRC cells.⁵ Once CRC metastasizes, the prognosis worsens significantly, with a notable decline in the 5-year survival rate,⁶ highlighting the urgent need to understand the mechanisms underlying CRC metastasis for the development of effective clinical treatments.

In recent years, extensive research has been conducted on colorectal liver metastasis (CLM), which is influenced by both the complex interactions between cancer cells and their surrounding microenvironment and inherent changes within cancer cells. The “seed and soil” theory, proposed by Stephen Paget in 1889, emphasizes the role of the microenvironment in metastasis, suggesting that tumor cells (the “seed”) require a supportive environment (the “soil”) to establish and grow in metastatic sites.⁷ The interactions between tumor cells and the tumor microenvironment (TME) are thus critical for metastasis.^{8,9} The TME consists of various non-cancerous cells, including fibroblasts, and immune cells, along with non-cellular components such as the extracellular vesicles (EVs), cytokines and extracellular matrix (ECM).¹⁰ A growing body of evidence suggests that the TME plays a crucial role in the initiation, progression, and metastasis of CRC.^{11,12}

Among the immune cells in the TME, tumor-associated macrophages (TAMs) are particularly important.¹³ TAMs contribute to tumor growth, metastasis, immunosuppression, and angiogenesis through the secretion of a range of cytokines and chemokines.^{14–16} TAM subtypes carry out diverse functions and can dynamically alter their behavior in

response to various signals from cancer cells or the TME. Research indicates that TAMs are closely linked to the occurrence of CLM and influence 5-year survival rates. Patients with higher pre-invasive macrophage infiltration tend to have better 5-year survival rates following radical resection compared to those with lower infiltration. Moreover, TAMs infiltration at the invasive front is associated with better outcomes in CLM and overall survival.¹⁷ In patients with CLM, TAMs comprise at least two major cell subtypes, namely large TAMs (L-TAMs) and small TAMs (S-TAMs). Notably, a S-TAM has a more favorable prognosis compared to a L-TAM. The 3-year disease-free survival (DFS) rates for S-TAMs and L-TAMs are 60% and 8.5%, respectively.¹⁸ On the other hand, the accumulation and activation of Kupffer cells (KCs) in the peritumoral area of CLM patients correlate with poor prognosis.¹⁹ TAMs interact with other immune cells in the TME and with tumor cells, shaping a microenvironment conducive to CLM. Despite the progress in understanding TAMs and their role in CLM, there are still challenges and gaps in current treatments. For example, targeting TAMs for therapeutic intervention may face difficulties in achieving specific delivery and avoiding off-target effects. Additionally, the complex nature of the TME and the dynamic interactions between different cell types make it challenging to develop effective therapeutic strategies. However, the potential for therapeutic interventions targeting TAMs in CLM remains significant. This review provides an overview of TAMs, discussing their definition, origin, and polarization, with a particular emphasis on their role in promoting CLM. Additionally, it explores therapeutic strategies that target TAMs in CLM patients.

Definition and Origin of TAMs

As a type of innate immune cell, macrophages primarily originating from monocytes, and they play a vital part in defending the host against pathogens, regulating tissue homeostasis, and maintaining structural integrity. Blood monocytes, which derive from bone marrow hematopoietic stem cells, are the primary source of these cells.^{20–22} Many resident macrophages, such as Kupffer cells, alveolar macrophages, and brain macrophages, originate from yolk sac progenitors and undergo in situ proliferation or differentiation.²³ Macrophages that infiltrate tumor tissues or populate the solid tumor microenvironment are classified as TAMs.²⁴ As essential components of the tumor microenvironment, TAMs influence tumor progression by promoting growth, modulating immune responses, facilitating tumor angiogenesis, enhancing metastasis, and contributing to drug resistance.^{25,26} TAMs are drawn to and activated by diverse signals within the TME, where they perform multiple roles in cancer progression, such as initiating and promoting tumors, modulating the immune response, facilitating metastasis, and supporting angiogenesis.

Polarization and Influencing Factors of TAMs

Macrophages can polarize into two distinct states: the classically activated M1-type (pro-inflammatory) and the alternatively activated M2-type (anti-inflammatory) (Figure 1).¹³ M1 macrophages are known for promoting the Th1 immune response and their ability to engulf and destroy tumor cells.²⁷ These macrophages are typically induced by cytokines such as interferon-gamma (IFN- γ), lipopolysaccharide (LPS), or tumor necrosis factor-alpha (TNF- α). Within the TME, M1 macrophages release various inflammatory factors, including interleukins (IL-1, IL-6, IL-12, IL-23), C-X-C motif ligand 10 (CXCL-10), and TNF- α , which enhance the anti-tumor immune response and help recruit and activate other immune cells to target the tumor.²⁸ Conversely, M2 macrophages, which arise under the influence of cytokines like IL-4, IL-10, IL-13, or glucocorticoids, secrete anti-inflammatory growth factors such as vascular endothelial growth factor (VEGF), transforming growth factor-beta (TGF- β), and matrix metalloproteinases (MMP-2, MMP-9), along with cytokines like IL-4, IL-10, and IL-13. These factors play crucial roles in promoting epithelial-mesenchymal transition (EMT), angiogenesis, and immunosuppression, which can ultimately lead to tumor progression and poor treatment outcomes.²⁹ Additionally, due to the diverse cytokines they produce and their varied functions, M2 macrophages can be further subdivided into M2a, M2b, M2c, and M2d subtypes.³⁰ Interestingly, macrophage polarization between the M1 and M2 states is not fixed, and they can transition from one state to another, highlighting the potential of macrophages as therapeutic targets. While M1 and M2 are the most commonly studied macrophage phenotypes, macrophage polarization is not limited to these two states.

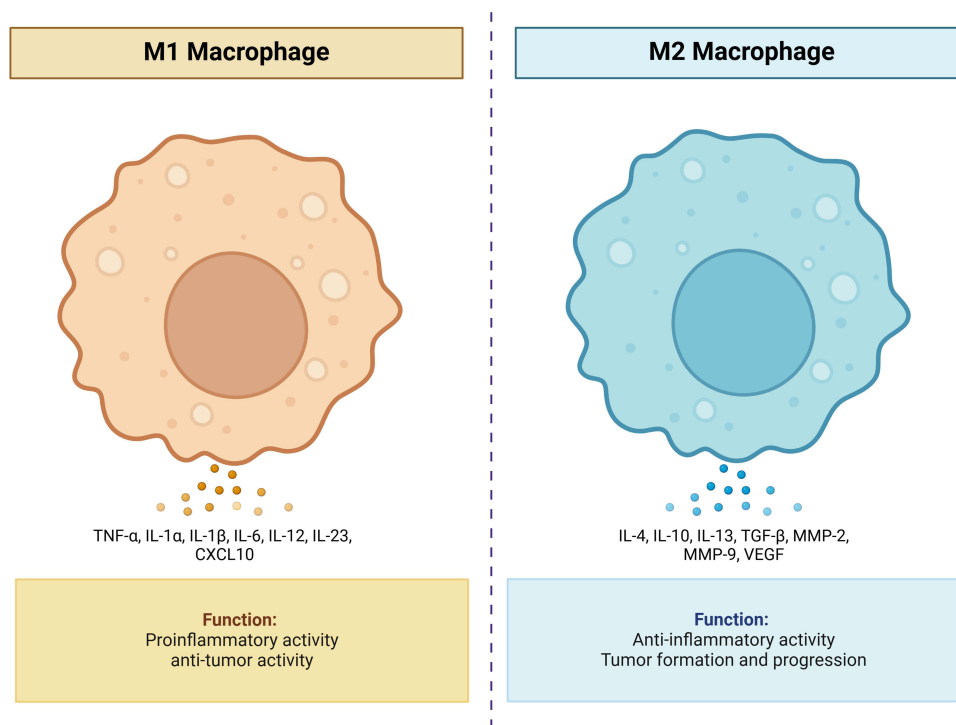


Figure 1 Functional Characteristics of M1 and M2 Macrophages in the Tumor Microenvironment. M1 macrophages produce pro-inflammatory and anti-tumor cytokines (TNF- α , IL-1 α , IL-1 β , IL-6, IL-12, IL-23, CXCL10). M2 macrophages secrete anti-inflammatory and pro-tumor factors (IL-4, IL-10, IL-13, TGF- β , MMP-2, MMP-9, VEGF), promoting tumor growth and progression. Created in BioRender. Hu, X. (2024) BioRender.com/k99w361.

The Role of TAMs Polarization in CLM

TAMs are integral to the development of CLM (Figure 2). Research indicates that different macrophage subtypes exhibit unique functions and distribution patterns within the liver metastasis microenvironment. Specifically,

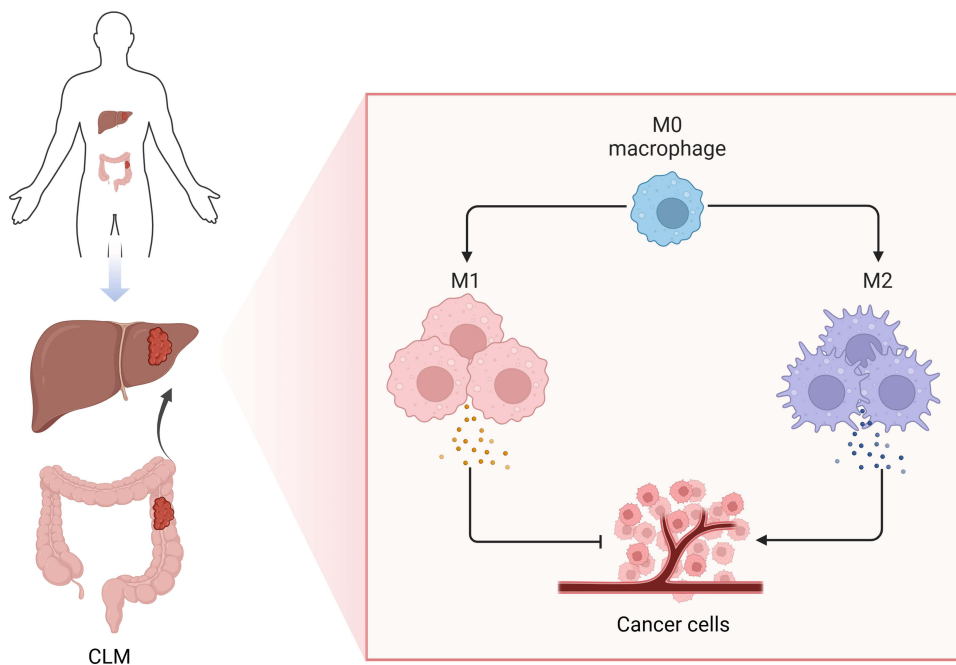


Figure 2 The role of TAMs in CLM. M0 macrophages can differentiate into M1 or M2 types. M1 macrophages are pro-inflammatory and anti-tumor, inhibiting cancer cell growth. M2 macrophages are anti-inflammatory and pro-tumor, promoting cancer growth and metastasis. The diagram illustrates colorectal cancer metastasis from the primary site to the liver and its interaction with macrophages. Created in BioRender. Hao, L. (2024) BioRender.com/b44g866.

CD68⁺CD163⁺CD206^{neg} M2-type macrophages predominate in CLM, particularly around the tumor periphery. Patients with higher levels of CD68⁺MRP8-14⁺CD86^{neg} M1-type macrophages in the tumor center had worse overall survival compared to those with lower M1 macrophage levels in this region.³¹ The expression patterns between the tumor invasive front (TF) area (<150 μm from the tumor center) and the peritumoral (PT) region (≥150 μm from the tumor center) differ between primary CRC and liver metastases. Notably, CD163 expression is more pronounced in the PT region than in the TF region. Additionally, CD163 expression is higher in liver metastases compared to primary sites, with CD68⁻CD163⁺ macrophages being the dominant type in liver metastases.³² Studies suggest that the impact of TAMs on liver metastasis in colorectal cancer is determined by the number and proportion of functional M1 and M2 subtypes, rather than the total TAMs count. The number of M2 macrophages and the M2/M1 ratio are more accurate predictors of CLM.³³ When connective tissue proliferates, there are more clusters of CD68-positive M1 macrophages and fewer CD206-positive M2 macrophages at the tumor-liver parenchyma interface. A better long-term prognosis is associated with fibrous tissue hyperplasia at the invasion front of metastatic tumors.³⁴ Consequently, further research into the roles and mechanisms of different TAM subtypes in CLM is essential for precise treatment strategies. These results indicate that in CLM, CD68⁺CD163⁺CD206^{neg} M2-type macrophages are dominant around the tumor. Moreover, a higher level of CD68⁺MRP8-14⁺CD86^{neg} M1-type macrophages in the tumor center is associated with a poorer overall survival rate. At the same time, it is pointed out that the number of M2 macrophages and the M2/M1 ratio are more accurate predictors for CLM. However, there is a potential conflict. Conclusions from different studies on which subtype of macrophages has what impact on prognosis under what circumstances may vary. For example, other studies may find different combinations of surface markers or in different tumor microenvironment regions, and the relationship between macrophage subtypes and prognosis is not so absolute. For critical evaluation, differences in research samples need to be considered, including sample size, individual differences of patients (such as age, gender, underlying diseases, etc.) and tumor staging and grading. Different sample characteristics may lead to different distributions and functional manifestations of macrophage subtypes, thus affecting the judgment of prognosis. In addition, the accuracy and specificity of research methods are also crucial. For example, surface markers used to identify macrophage subtypes may have cross-reactions or inaccurate situations, which may lead to misclassification of macrophage subtypes and further affect the conclusion on their relationship with prognosis.

TAM polarization is influenced by various cytokines, growth factors, chemokines, and other signals from tumor and stromal cells, playing a key role in CRC progression and metastasis. The CCL1-CCR8 axis, for example, is a positive regulator of CRC progression. Decreased CCL1 expression in TAMs at liver metastatic sites may contribute to the slower tumor progression in CRC, allowing time for radical resection of metastases.³⁵ Inhibiting TCF4 or CCL2 in tumor cells prevents CLM, while blocking the TCF4-CCL2-CCR2 axis significantly reduces CLM by hindering TAMs accumulation and M2 polarization in the tumor microenvironment.³⁶ The CCL2/CCR2 chemokine axis supports the recruitment of immunosuppressive TAMs to liver metastatic sites in CRC patients, thereby promoting tumor growth. Targeting CCR2 can reduce TAM accumulation in liver metastases and restore anti-tumor immunity.³⁷ CXCL16 suppresses CLM by promoting TNF-α secretion from TAMs, which induces tumor cell apoptosis.³⁸ High levels of CDC42 cargo from CRC cell-derived extracellular vesicles (CRC-EVs) are delivered to macrophages, activating NOD1 and subsequent phosphorylation of RIP2, leading to the release of downstream cytokines and chemokines, thus promoting CRC metastasis.³⁹ Human umbilical cord mesenchymal stem cells (hUCMSCs) secrete exosomes carrying miR-1827, which are delivered to CRC cells, inhibiting macrophage M2 polarization and preventing CLM.⁴⁰ Loss of Ndr2 inhibits CLM by activating the NF-κB pathway, which regulates macrophage polarization towards the M1 phenotype.⁴¹ Collagen triple helix repeat containing 1 (CTHRC1), an intrinsic marker of CRC metastasis, regulates macrophage polarization towards the M2 phenotype via the TGF-β signaling pathway, promoting CLM.⁴² Orosomucoid 1 (ORM1) mediates tumor immune tolerance by inducing macrophage M2 polarization, thereby advancing CRC progression and liver metastasis.⁴³ Osteoprotegerin (OPG) expression inhibits macrophage migration by blocking the RANKL-RANK pathway, and its downregulation in CRC cells promotes liver metastasis by activating TAMs.⁴⁴ STING activation promotes the nuclear translocation of TFEB by activating IRG1, inhibiting M2 macrophage polarization, and consequently suppressing CLM.⁴⁵ Additionally, KCs activation and their phagocytosis of tumor cells can reduce CRC cell metastasis to the liver.⁴⁶ KCs also induce liver metastasis via TGF-β1 signaling through the angiotensin II subtype receptor 1a (AT1a)

pathway.⁴⁷ In summary, TAMs play a pivotal role in CLM, with their polarization regulated by various cytokines, growth factors, chemokines, and other signals produced by tumor and stromal cells. Different signaling axes and factors distinctly influence TAMs accumulation and polarization, thereby affecting CLM progression. Furthermore, KCs activation and their tumor cell phagocytosis significantly contribute to suppressing CLM. These insights not only deepen our understanding of the mechanisms underlying CLM but also offer a theoretical basis and direction for developing novel targeted therapeutic strategies. The study lists the effects of various cytokines, growth factors, chemokines, etc. on the polarization of TAMs. For example, the CCL1-CCR8 axis, the TCF4-CCL2-CCR2 axis, CXCL16, and the CDC42 cargo carried by CRC-EVs all have regulatory effects on the polarization of TAMs and affect the progression of CLM. However, there are potential conflicts in the complex tumor microenvironment. Multiple factors interact, and different studies may have disagreements on the importance ranking or mechanism of action of certain factors. For instance, one study may emphasize the key role of a specific factor in the polarization of TAMs, while other studies may find that the role of this factor is masked or regulated by other factors. A critical evaluation shows that the tumor microenvironment is a highly complex system, and there are complex network relationships among various factors. When studying the effect of a single factor on the polarization of TAMs, it is difficult to completely exclude the interference of other factors. Therefore, it is necessary to comprehensively consider the synergistic or antagonistic effects of multiple factors. At the same time, the choice of experimental models will also affect the research results. *In vivo* models can better reflect the real physiological environment but are more difficult to control variables and *in vitro* models can better control the role of a single factor but may not be able to fully simulate the complex situation.

Metabolic Changes of TAMs

Macrophage metabolism plays a pivotal role in regulating their phenotypic polarization and function. The metabolic enzyme abhydrolase domain containing 5 (ABHD5) in macrophages has been shown to diminish the invasion and metastatic capabilities of CRC cells by inhibiting the production of matrix metalloproteinases (MMPs) through the NF- κ B signaling pathway. This finding suggests that targeting ABHD5 in TAMs could serve as a potential therapeutic strategy for CRC.⁴⁸ RNA sequencing of various macrophage populations revealed that the LXR/RXR pathway is highly enriched in large macrophages, contributing to the upregulation of cholesterol metabolism, clearance receptors, MERTK, and complement genes.⁴⁹ At the infiltrative edge of CLM, there is a significant presence of pro-inflammatory monocyte-derived macrophages (MoM ϕ) marked by the expression of SERPINB2, as well as more differentiated TAMs expressing glycoprotein nonmetastatic melanoma protein B (GPNMB).⁵⁰ SERPINB2⁺ MoM ϕ are indicative of early inflammatory responses, while GPNMB⁺ TAMs are involved in matrix degradation, angiogenesis, and lipid metabolism pathways.⁵⁰ A high infiltration of SERPINB2⁺ cells correlate with extended disease-free survival, whereas a greater density of GPNMB⁺ cells is associated with poorer survival outcomes.⁵⁰ Moreover, studies indicate that taurocholic acid (TCA) facilitates CLM, with notable increases in the proportions of myeloid-derived suppressor cells (MDSCs), neutrophils, and macrophages in the liver following TCA injection.¹² The effective establishment of secondary liver metastases may also be attributed to macrophage-induced immunosuppression within the tumor microenvironment. Liver metastases show a significant upregulation of “bile acid metabolism” pathways.⁵¹ These insights highlight the crucial influence of metabolic processes in the tumor microenvironment on CLM progression and suggest new potential targets for therapeutic intervention.

Role of miRNAs in TAM-Mediated CLM

Non-coding RNAs play a role in the progression of CLM by modulating TAMs. MicroRNAs (miRNAs), a broad category of small non-coding single-stranded RNAs, are recognized for their widespread presence and functional diversity, highlighting their significant biological roles. The decreased expression of miR-22 is associated with a supportive tumor microenvironment, which facilitates epithelial-to-mesenchymal transition and contributes to a cancer stem cell-like phenotype.⁵² Up-regulation of miR-216b inhibited the growth of CRC tumors and M2-type polarization of macrophages in mice. Overexpression of miR-216b also reduced liver and lung metastasis of mouse tumor cells.⁵³ Furthermore, CRC cell-derived exosomal miR-934 was found to induce M2 macrophage polarization, thereby enhancing CLM through a CXCL13/CXCR5/NF- κ B/p65 positive feedback loop.⁵⁴ In conclusion, non-coding

RNAs, especially miRNAs, are crucial in the progression of CLM by modulating TAMs. The dysregulation of miRNAs in the TME can influence critical processes, including epithelial-to-mesenchymal transition and macrophage polarization.

Cellular Interactions Between Macrophage and Components of TME

The complex interactions between macrophages, immune cells, and fibroblasts are vital in sustaining metastatic spread within the hepatic parenchyma. In patients with a high aspartate aminotransferase to platelet ratio index (APRI), there is an enrichment of inflammatory cancer-associated fibroblasts (CAFs) and SPP1-expressing macrophages. This enrichment is linked to the activation of malignant cells, the development of a fibrotic microenvironment, and a more suppressed T cell function.⁵⁵ Studies suggest that the increase in M2 macrophages and CAFs contributes to CLM progression.⁵⁶ Specifically, ANGPTL2 and SPP1 are notably enriched in CAFs and macrophages within CRC tissue, respectively, where ANGPTL2+ CAFs and SPP1+ macrophages facilitate CRC cell metastasis.⁵⁷ MYL9 in CAFs is implicated in modulating cytokine and chemokine secretion, which recruits M2 macrophages, inhibits dendritic cell activation, and fosters an immunosuppressive microenvironment.⁵⁸ SPP1⁺ macrophages are especially predominant in liver metastasis.⁵⁹ Complement 5a (C5a) promotes tumor metastasis through its regulation of inflammation in CLM, with C5a receptor deficiency in metastatic colon cancer leading to reduced infiltration of macrophages, neutrophils, and dendritic cells.⁶⁰ The STING signaling pathway further plays a role in inhibiting CLM by promoting NK cell activity via 4-1BBL/4-1BB co-stimulation.⁶¹ In summary, the interactions between macrophages, immune cells, and fibroblasts within the TME are pivotal in maintaining CLM, underscoring the potential of targeting these cellular interactions as a therapeutic approach for CLM management.

Crosstalk Between Macrophages and the Environmental Factors, Microbiota in CLM

KCs regulated by intestinal flora affect the occurrence and development of CLM. Liver metastases were observed in the Vanc group compared with the Coli group and the combined treatment group. *vulgatus* was more abundant in the Coli group before metastasis occurred. KCs might play a role in mediating the effects of gut microbiota on CLM. Specifically, *P. mirabilis* was found to enhance the migration of CT26 cells and promote CLM by decreasing the recruitment of KCs. In contrast, *B. vulgatus* appeared to inhibit CT26 cell migration and CLM by promoting the accumulation of KCs.⁶² In summary, the crosstalk between macrophages and the microbiota plays a crucial role in CLM. These insights suggest that targeting the gut-liver axis and modulating specific microbial populations could provide strategies for managing CLM. The study found that cathepsin K (CTSK) is an important mediator of intestinal flora imbalance and CRC metastasis. Experiments found that when the intestinal flora is imbalanced, overexpression of CTSK leads to larger tumors and more metastases. CTSK can bind to TLR4 to stimulate the polarization of tumor-associated macrophages M2. It can also stimulate M2 macrophages to secrete cytokines to promote the metastasis of CRC cells. Clinically, overexpression of CTSK is associated with high M2 macrophages, CRC metastasis, and poor prognosis.⁶³ It is found that tumor-infiltrating bacteria such as *Escherichia coli* exist in CLM. They increase lactate production. Lactate inhibits nuclear factor- κ B-gene binding (NF- κ B) signal transduction through the lactylation of retinoic acid-inducible gene 1 (RIG-I), mediates M2 macrophage polarization, and affects the immunosuppressive activity of regulatory T cells (Tregs) and the anti-tumor activity of CD8 T cells. The RIG-I lactylation inhibitor screened by small molecule compounds can inhibit M2 polarization and make CLM sensitive to 5-fluorouracil.⁶⁴ In clinical practice, PRM1201 can increase the number of short-chain fatty acid-producing bacteria and the production of short-chain fatty acids in the feces of patients with CRC. The anti-metastasis effect is positively correlated with the recovery of short-chain fatty acids. Its anti-cancer metastasis mechanism is related to inhibiting histone deacetylation and epithelial-mesenchymal transition, highlighting the dependence on the microbiota.⁶⁵ Many studies have shown that the environment has an important impact on the treatment of CLM and may become a future development direction. Plexin B2 is a key host regulatory factor for liver colonization in mouse models of colorectal cancer and others. It interacts with related substances on tumor cells to upregulate KLF4 and promote tumor cells to obtain epithelial characteristics. Blocking the plexin-B2-semaphorin axis can prevent liver metastasis.⁶⁶ Chronic low-dose cadmium exposure can promote the invasion and metastasis ability of colorectal cancer cells in vivo and in vitro. Cadmium activates epidermal growth factor receptor (EGFR) in a non-classical way and promotes a continuous EGFR signal to trigger the protein kinase B (Akt)/mammalian target of rapamycin (mTOR)

cascade reaction. Blocking EGFR can eliminate the promotion effect of cadmium on liver metastasis of CRC cells.⁶⁷ In mice fed an ethanol-containing diet, metastasis occurs earlier and more severely. Alcohol can increase the expression of liver cytokines and factors related to the colonization of colorectal cancer cells by 1.5–3.0 times. Moreover, alcoholic liver injury is related to changes in liver localization and increased circulating levels of CEA released by CRC cells.⁶⁸ Therefore, research on lifestyle, immunity, and personalized molecular biomarkers as well as molecular pathological epidemiology is a promising direction. The influence of external factors is related to molecular pathology and can evaluate the efficacy and side effects of the therapy.^{69,70}

Potential Mechanisms of TAMs to Promote CLM

CLM may have different growth patterns, angiogenesis characteristics, and immune microenvironments compared to the primary CRC. The potential of liver metastasis is related to the matrix composition of primary CRC. Differences in the expression patterns of type I and type IV collagen in CRC are associated with an increased risk of CLM.⁷¹ CLM patients had significantly higher levels of circulating type IV collagen.⁷² KCs may promote CLM by promoting ECM remodeling and promoting tumor cell invasion by CRC cells that evade initial phagocytosis.⁷³ Although numerous studies have explored the role and clinical relevance of TAMs in CRC, little is known about its role in CLM, which may be due to a different phenotype profile of macrophage expression in the liver.⁷⁴ Therefore, TAMs with different morphological and molecular fingerprints coexist in CLM and are associated with clinicopathological variables.²⁴ In patients with CLM, TAMs comprise at least two major cell subtypes, namely large TAMs (L-TAMs) and small TAMs (S-TAMs). The quantitative morphological characterization of TAMs can act as an easily quantifiable correlate of functional diversity with significant prognostic implications. TAMs can be used to reliably stratify patient outcomes and predict recurrence. Notably, a S-TAM has a more favorable prognosis compared to a L-TAM. The 3-year disease-free survival (DFS) rates for S-TAMs and L-TAMs are 60% and 8.5%, respectively.¹⁸

TAMs play crucial roles in various aspects of tumor progression, particularly in CLM. The mechanisms by which TAMs contribute to tumor liver metastasis are intricate, as they are implicated in almost every stage of metastasis.

TAMs Promote Angiogenesis of Tumor Cells

TAMs significantly influence tumor angiogenesis, a process essential for supplying nutrients and oxygen to support tumor growth, invasion, and migration.⁷⁵ Located within the tumor microenvironment (TME), TAMs are key players in promoting cancer cell survival and progression by producing various inflammatory mediators, growth factors, cytokines, and chemokines.⁷⁶ For example, SHP-2 deficiency in macrophages has been shown to enhance liver tumor metastasis by activating the Ang/Tie2-PI3K/Akt/mTOR pathway in Tie2-expressing monocyte/macrophages, which in turn promotes tumor microvascular formation.⁷⁷ Research indicates that macrophage infiltration is positively correlated with blood vessel density specifically in liver metastasis patients.⁷⁸ Notably, while TAMs generally exhibit an M1-like phenotype, metastasis-associated macrophages (MAMs) tend to be more M2-like and are particularly effective in promoting angiogenesis.⁷⁸ In both murine models and human studies, a subset of MAMs expressing VEGFR1 has been identified as key contributors to angiogenesis and metastasis.⁷⁸ KCs, the liver-resident macrophages, are instrumental in the formation of liver metastases.⁷⁹ The activation of TGF α -EGFR signaling in colon cancer cells leads to macrophage recruitment, resulting in an increase in both blood and lymphatic vessel surface area and enhanced lymphatic metastasis.⁸⁰ Additionally, the CX3CR1 receptor in macrophages has been linked to tumor metastasis and poor prognosis by promoting the survival of angiogenic macrophages.⁸¹ TAMs also release VEGF, a potent pro-angiogenic factor, contributing to blood vessel remodeling and the progression of CRC.⁸² VEGF-A secretion by CRC cells stimulates TAMs to produce CXCL1, which, in premetastatic liver tissue, recruits CXCR2-positive myeloid-derived suppressor cells (MDSCs) to establish a premetastatic niche, ultimately driving liver metastasis.⁸³ Furthermore, OPN-positive macrophages are known to produce VEGF, thereby enhancing angiogenesis and potentially facilitating cancer cell metastasis to the liver.⁸⁴ In the TME, tumor-derived VEGF enhances the Syk signaling pathway in macrophages, leading to the assembly of the CARD9-BCL10-MALT1 complex.⁸⁵ CARD9 plays a pivotal role in directing macrophage polarization towards the M2 phenotype through NF- κ B pathway activation.⁸⁵

TAMs Play a Critical Role in Facilitating Tumor Metastasis and Invasion

Metastasis requires tumor cells to spread from the primary tumor to different organs.⁸⁶ Metastasis involves the dissemination of tumor cells from the primary site to distant organs, and the ability of these cells to invade new tissues is a key indicator of malignancy.⁸⁷ A pivotal process in this context is the epithelial-mesenchymal transition (EMT), where epithelial cells gain mesenchymal traits, enhancing their migratory and invasive potential.⁸⁷ TAMs contribute to the metastatic process of colorectal cancer (CRC) by fostering a microenvironment conducive to EMT, primarily through the activation of the TGF- β signaling pathway. This allows tumor cells to acquire mesenchymal characteristics, enabling them to detach from the primary site and invade new tissues.⁸⁸

Further, TAMs are involved in promoting CRC cell migration, invasion, and metastasis by inducing the EMT process via the JAK2/STAT3/miR-506-3p/FoxQ1 axis, which also enhances the production of CCL2, facilitating macrophage recruitment to the tumor microenvironment.⁸⁹ The activation of NLRP3 signaling in TAMs further supports the migratory and invasive capabilities of CRC cells.⁹⁰ Moreover, cytokine TWEAK, secreted by Th17 cells, interacts with its receptor Fn14 on tumor cells, promoting EMT and, consequently, tumor migration and invasion.⁹¹ High TWEAK expression levels have been correlated with poor prognosis in CRC patients.⁹¹ Additionally, the interaction between CD163L1⁺ macrophages and Th17 cells recruits Th17 cells through the CCL4-CCR5 axis, further contributing to the invasive potential of the tumor.⁹¹ Interestingly, certain mechanisms within TAMs can also inhibit metastasis. For instance, Src homology 2 domain-containing tyrosine phosphatase 2 (Shp2) on TAMs promotes macrophage polarization towards the M1 phenotype, which is associated with anti-tumor effects, thus reducing metastasis.⁹² Similarly, the α 7 nicotinic acetylcholine receptor (α 7nAChR) in TAMs suppresses CRC metastasis through the JAK2/STAT3 signaling pathway⁹³.

Induction of Premetastatic Niche Formation

Formation of a premetastatic niche (PMN) is crucial in facilitating metastatic tumor growth in distant organs by creating an environment conducive to tumor cell colonization. The PMN is characterized by features such as increased vascular permeability, lymphangiogenesis, extracellular matrix (ECM) remodeling, and an immunosuppressive milieu.⁹⁴ For instance, exosomal miR-203a-3p contributes to M2 macrophage polarization by downregulating PTEN and activating the PI3K/AKT pathway.⁹⁵ These M2 macrophages secrete CXCL12, which enhances tumor metastasis via the CXCL12/CXCR4/NF- κ B pathway, thus establishing the PMN in CLM.⁹⁵ The tumor microenvironment (TME) in CLM is notably immunosuppressive, with myeloid cells, particularly myeloid-derived suppressor cells (MDSCs), being polarized to an M2 state and T cells becoming exhausted. Myeloid cells further interact with various TME components, influencing processes like angiogenesis, tumor cell invasiveness, and EMT, and thereby aiding in PMN formation and promoting CRC liver metastasis.⁹⁶ High levels of serum exosomal miR-203 are linked to distant metastasis and are an independent predictor of poor prognosis. This microRNA promotes distant metastasis by driving host M2 macrophages to form the PMN.⁹⁷ Similarly, miR-21-5p, abundant in CRC-derived small extracellular vesicles (sEVs), is pivotal in inducing a pro-inflammatory hepatic environment, facilitating CRC liver metastasis through the miR-21-Toll-like receptor 7 (TLR7)-IL6 axis.⁹⁸ Additionally, VEGFA from CRC cells triggers TAMs to produce CXCL1, which attracts CXCR2-positive MDSCs, whose accumulation in the liver furthers PMN formation and supports liver metastasis⁹⁹.

Promotion of Tumor Cell Intravasation and Extravasation

TAMs also play a vital role in the intravasation and extravasation of CRC cells, processes crucial for metastasis.¹⁰⁰ Activation of EGFR is necessary for tumor cell intravasation¹⁰¹, and TAM-derived EGF enhances CRC cell invasion and motility.¹⁰² In CRC, TAMs promote tumor progression by remodeling the ECM.¹⁰³ VEGF further facilitates tumor cell extravasation by inducing endothelial gaps, as demonstrated by experiments with VEGF-expressing CT26 cells in mice.¹⁰⁴ Thus, TAMs with pro-inflammatory and pro-angiogenic traits are key players in promoting the metastatic spread of CRC cells through intravasation and extravasation.

Targeting TAMs in the Treatment of Metastatic CRC

Inhibiting TAMs Survival

Targeting TAMs represents a promising therapeutic strategy in managing metastatic CRC. Inhibiting TAM survival can hinder tumor proliferation, invasion, migration, and angiogenesis while enhancing anti-tumor immunity. Overview of macrophage-targeting drugs and their functions (Table 1). For example, blocking CSF1R with the inhibitor PLX3397 depletes M2 TAMs, boosts CD8 T cell infiltration, and improves responses to chemotherapy and immunotherapy, effectively curbing tumor growth and metastasis.¹⁰⁵ The drug TMP195 reduces F4/80+ TAM populations, which significantly inhibits the growth and vascularization of CLM.¹⁰⁶ Additionally, the combination of Oxaliplatin with Trifluridine/Tipiracil (FTD/TPI), an antimetabolite for treating chemotherapy-refractory metastatic CRC, can eliminate M2 TAMs in CT26 tumor-bearing mice.¹⁰⁷ Moreover, inhibiting FGL1 secretion, which is targeted by the FDA-approved antiseptic benzethonium chloride, enhances the effectiveness of anti-PD-1 therapy in liver metastasis by suppressing TAM activity.¹⁰⁸ Thus, inhibiting TAM activation is a beneficial strategy in the treatment of CRC metastasis.

Reprogramming TAMs

Reprogramming TAMs from an M2 to an M1 phenotype offers potential therapeutic benefits for patients with metastatic CRC. In CRC mouse models, anti-MARCO monoclonal antibodies can shift TAMs towards an M1 phenotype, enhancing anti-tumor activity and potentially inhibiting CRC metastasis.¹⁰⁹ Tasquinimod induces a shift from M2-like to M1-like TAMs, thereby modifying the TME to enhance immune regulation, inhibit angiogenesis, and suppress metastasis.¹¹⁰ CCR5 antagonists have demonstrated the ability to repolarize TAMs to an M1 phenotype by affecting the STAT3/SOCS3 signaling pathway, showing promising anti-tumor effects in Phase I clinical trials for CLM.¹¹¹ M1-derived EVs containing oxaliplatin, retinoic acid, and *Libidibia ferrea* can induce TAM transition from M2 to M1 through the STAT3/NF- κ B/AKT signaling pathway, effectively inhibiting metastasis in CRC mouse models.¹¹² Additionally, dual blockade of CTLA-4 and PD-L1 synergistically enhances CRC growth and metastasis inhibition by increasing CD8⁺ and CD4⁺ T cell counts, promoting Th1 responses, and inducing M1 macrophage polarization.¹¹³ Nanodrugs like NCG inhibit MDSC differentiation, promote M1-like TAM polarization, disrupt the immunosuppressive barrier of tumor-associated fibroblasts, and increase effector T cell infiltration.¹¹⁴ Dahuang Zhechong Pill reduces CLM by decreasing hepatic macrophage infiltration and M2 polarization, mitigating CCL2-mediated M2 skewing, and improving the fibrotic microenvironment.¹¹⁵ Xiaoyaosan treatment significantly reduces the number of CD11b⁺F4/80⁺ TAMs and

Table 1 Overview of Macrophage-Targeting Drugs and Their Functions

Drug	Function	Reference
PLX3397	Depletion of M2 macrophages and increases CD8 ⁺ T cell infiltration	[105]
TMP195	Inhibits F4/80 ⁺ TAMs population	[106]
FTD/TPI	Depletes TAMs	[107]
Benzethonium chloride	Inhibits FGL1	[108]
Anti-MARCO	Increases M1 macrophages and decreases M2 macrophages	[109]
Tasquinimod	Leads to the transformation of M2 macrophages into M1 macrophages	[110]
CCR5 antagonist	Leads to the transformation of M2 macrophages into M1 macrophages	[111]
Oxaliplatin, retinoic acid, and <i>Libidibia ferrea</i>	Leads to the transformation of M2 macrophages into M1 macrophages	[112]
Anti CTLA-4 and anti-PD-L1	Induces polarization of macrophage M1	[113]
NCG(+) and anti-PD-L1	Induces polarization of macrophage M1	[114]
Dahuang Zhechong Pill	Decreases M2 polarization	[115]
Xiaoyaosan	Decreases CD11b ⁺ F4/80 ⁺ TAM	[116]
Bufalin	Decreases M2 polarization	[117]
Macelignan	Decreases M2 polarization	[118]
NT157	Inhibits expression of CCL2 to inhibit recruitment of TAMs	[119]
AMG 820	Blocks the CSF1-CSF1R signaling to inhibit recruitment of TAMs	[120]
Grapefruit-derived nanovectors deliver miR-18a	Induction of IL-12	[121]

CD11b⁺Gr1Ly6Chi MDSCs, thereby inhibiting the progression of CLM.¹¹⁶ Bufalin mitigates M2 polarization of KCs and downregulates IL-6 expression, inhibiting tumor metastasis.¹¹⁷ Macelignan reduces IL-1 β secretion by M2 macrophages, blocks NF- κ B p65 nuclear translocation, and prevents IL-1 β /NF- κ B-dependent CRC metastasis through the ROS-mediated PI3K/AKT signaling pathway.¹¹⁸ Therefore, influencing the polarization of TAMs is a promising strategy for the treatment of metastatic CRC.

Inhibiting TAM Recruitment

Inhibiting TAM recruitment involves targeting growth factors and chemokines that tumors use to attract TAMs, thereby promoting tumor growth.¹²² NT157 reduces CRC cell migration and invasion and decreases metastatic lesion formation in the liver by inhibiting oncogenic cytokines, chemokines, growth factors, and TGF- β , consequently lowering TAM levels in the TME.¹¹⁹ The anti-colony-stimulating factor 1 receptor (anti-CSF1R) monoclonal antibody AMG 820, used in combination with pembrolizumab, may inhibit TAM recruitment by reducing CSF-1R levels in metastatic CRC.¹²⁰ Moreover, miR-18a encapsulated in grapefruit-derived nanovectors (GNV) can inhibit liver metastasis by inducing M1 macrophages.¹²¹ Therefore, targeting TAMs recruitment signals is a significant therapeutic strategy for treating CRC metastasis.

The above research presents various treatment strategies targeting TAMs, including inhibiting TAMs' survival, reprogramming TAMs, and suppressing TAMs recruitment. It also lists the action mechanisms of related drugs and methods and their effects in animal models. However, it is noted that translating these strategies into clinical success poses challenges. There is a potential conflict in that treatment strategies effective in preclinical studies may not work well in clinical trials. Different clinical trials may yield different conclusions, for instance, some drugs may be effective in certain patient groups but not in others. A critical assessment reveals that there are significant differences between preclinical studies and clinical trials. Preclinical studies are typically conducted in animal models, which may not fully mimic the complexity and diversity of human diseases. In clinical trials, factors such as individual differences among patients, tumor heterogeneity, and the complexity of combination therapy regimens all affect the treatment outcome. Additionally, factors like drug dose, administration route, and treatment duration need further optimization. For example, a drug may have an inhibitory effect on TAMs at a specific dose, but an excessively high or low dose may lead to reduced efficacy or adverse reactions.

Discussion

Liver metastasis is a predominant site of spread in CRC and remains the leading cause of death for these patients. Traditional approaches—chemotherapy, surgical resection, and local therapies—are commonly used to manage or control these metastatic tumors. However, the challenge of recurrence persists, emphasizing the need for deeper insights into the mechanisms of liver metastasis and the development of novel therapeutic strategies.

The TME has become a focal point in cancer research. This environment is characterized by complex interactions among cancer cells, endothelial cells, fibroblasts, and immune cells, all of which contribute to tumor growth and metastasis.¹²³ TAMs within this milieu can adopt either M1 or M2 phenotypes, playing a significant role in metastasis through mechanisms such as promoting angiogenesis. This makes TAMs promising targets for therapeutic intervention.

TAMs demonstrate considerable heterogeneity in their role in metastasis. Their interactions with cancer cells and other immune components within the TME are dynamic and multifaceted, influencing therapeutic responses. To comprehensively understand the mechanisms driving liver metastasis, researchers must investigate gene mutations unique to the liver TME and consider how these factors contribute to adaptive phenotypes. Advances in single-cell RNA sequencing, spatial transcriptomics, and high-throughput multi-omics are proving valuable in uncovering the detailed molecular landscape of liver metastasis. For instance, multi-omics studies have revealed that memory CD8⁺ T cells, B cells, and CTSB⁺ macrophages are prevalent at liver metastatic sites, forming crucial immune memory niches that support antitumor responses.¹¹¹ Single-cell analysis shows that primary CRC tumors have fewer myeloid cells, mainly monocytes and M1-type TAMs, whereas liver metastases are marked by an immunosuppressive TME with MDSCs, M2-type TAMs, and exhausted T cells.⁸³ These advanced techniques offer new insights into CRC metastasis and open avenues for enhancing CRC prognosis.

Despite promising preclinical data, translating TAMs-targeting strategies into clinical success has been challenging. Multicenter clinical trials are vital for advancing the diagnosis and treatment of liver metastasis. Merely blocking TAMs infiltration or inhibiting their cancer-promoting functions may not suffice for effective therapy. Combining TAMs-targeting agents with chemotherapy, radiotherapy, immunotherapy, and other treatments might provide more effective solutions for CRC progression and metastasis. Nonetheless, these combined approaches have yet to achieve significant clinical results. Further research is needed to assess their efficacy both as standalone treatments and in combination therapies.

This review outlines the origins, polarization, and functions of TAMs, underscoring their potential as therapeutic targets for liver metastasis. Despite the existing challenges, the rapid progress in biotechnology holds promise for identifying TAMs as a critical focus for future therapies. Continued research and clinical trials are essential to fully exploit the therapeutic potential of targeting TAMs in metastatic CRC.

In conclusion, while TAMs represent a promising target for treating liver metastasis, further investigation is crucial to define their roles in disease progression and refine therapeutic strategies. Ongoing exploration of TAMs biology and their interactions within the TME may lead to innovative treatments that could significantly improve outcomes for patients with liver metastasis.

Acknowledgments

The authors acknowledge using Biorender (<https://app.biorender.com/>) to create the schemata (Figures 1 and 2).

Funding

The present study was financially supported by Science and Technology Program of Hebei (223777156D) and Clinical Medical School Graduate Research Innovation Practice Project (2023KCY06); National Natural Science Foundation of China (No. 81973840 and No. 81273748).

Disclosure

The authors declare that they have no competing interests in this work.

References

1. Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA*. 2021;71(3):209–249. doi:10.3322/caac.21660
2. Buccafusca G, Proserpio I, Tralongo AC, Rametta Giuliano S, Tralongo P. Early colorectal cancer: diagnosis, treatment and survivorship care. *Crit rev oncol/hematol*. 2019;136:20–30. doi:10.1016/j.critrevonc.2019.01.023
3. Ganesh K, Stadler ZK, Cercek A, et al. Immunotherapy in colorectal cancer: rationale, challenges and potential. *Nat Rev Gastroenterol Hepatol*. 2019;16(6):361–375. doi:10.1038/s41575-019-0126-x
4. Sathé A, Mason K, Grimes SM, et al. Colorectal cancer metastases in the liver establish immunosuppressive spatial networking between tumor-associated SPP1+ macrophages and fibroblasts. *Clin Cancer Res*. 2023;29(1):244–260. doi:10.1158/1078-0432.CCR-22-2041
5. Wang Y, Zhong X, He X, et al. Liver metastasis from colorectal cancer: pathogenetic development, immune landscape of the tumour microenvironment and therapeutic approaches. *J Exp Clin Cancer Res*. 2023;42(1):177.
6. Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer Statistics, 2021. *CA*. 2021;71(1):7–33. doi:10.3322/caac.21654
7. Paget S. The distribution of secondary growths in cancer of the breast. 1889. *Cancer Metastasis Rev*. 1989;8(2):98–101.
8. Quail DF, Joyce JA. Microenvironmental regulation of tumor progression and metastasis. *Nature Med*. 2013;19(11):1423–1437. doi:10.1038/nm.3394
9. McAllister SS, Weinberg RA. The tumour-induced systemic environment as a critical regulator of cancer progression and metastasis. *Nat Cell Biol*. 2014;16(8):717–727. doi:10.1038/ncb3015
10. Li C, Teixeira AF, Zhu HJ, Ten Dijke P. Cancer associated-fibroblast-derived exosomes in cancer progression. *Mol Cancer*. 2021;20(1):154. doi:10.1186/s12943-021-01463-y
11. Qi J, Sun H, Zhang Y, et al. Single-cell and spatial analysis reveal interaction of FAP(+) fibroblasts and SPP1(+) macrophages in colorectal cancer. *Nat Commun*. 2022;13(1):1742. doi:10.1038/s41467-022-29366-6
12. Zheng X, Ma Y, Bai Y, et al. Identification and validation of immunotherapy for four novel clusters of colorectal cancer based on the tumor microenvironment. *Front Immunol*. 2022;13:984480. doi:10.3389/fimmu.2022.984480
13. Chanmee T, Ontong P, Konno K, Itano N. Tumor-associated macrophages as major players in the tumor microenvironment. *Cancers*. 2014;6(3):1670–1690. doi:10.3390/cancers6031670
14. Ngambenjawang C, Gustafson HH, Pun SH. Progress in tumor-associated macrophage (TAM)-targeted therapeutics. *Adv Drug Delivery Rev*. 2017;114:206–221. doi:10.1016/j.addr.2017.04.010

15. Goswami KK, Ghosh T, Ghosh S, Sarkar M, Bose A, Baral R. Tumor promoting role of anti-tumor macrophages in tumor microenvironment. *Cell Immunol.* 2017;316:1–10. doi:10.1016/j.cellimm.2017.04.005
16. Malekghasemi S, Majidi J, Baghbanzadeh A, Abdolalizadeh J, Baradaran B, Aghebati-Maleki L. Tumor-associated macrophages: protumoral macrophages in inflammatory tumor microenvironment. *Adv Pharm Bull.* 2020;10(4):556–565. doi:10.34172/apb.2020.066
17. Zhou Q, Peng RQ, Wu XJ, et al. The density of macrophages in the invasive front is inversely correlated to liver metastasis in colon cancer. *J Transl Med.* 2010;8(1):13. doi:10.1186/1479-5876-8-13
18. Costa G, Sposito C, Soldani C, et al. Macrophage morphology and distribution are strong predictors of prognosis in resected colorectal liver metastases: results from an external retrospective observational study. *Int J Surg.* 2023;109(5):1311–1317. doi:10.1097/JS9.0000000000000374
19. Miyagawa S, Miwa S, Soeda J, Kobayashi A, Kawasaki S. Morphometric analysis of liver macrophages in patients with colorectal liver metastasis. *Clin Exp Metastasis.* 2002;19(2):119–125. doi:10.1023/A:1014571013978
20. Franklin RA, Liao W, Sarkar A, et al. The cellular and molecular origin of tumor-associated macrophages. *Science.* 2014;344(6186):921–925. doi:10.1126/science.1252510
21. Shand FHW, Ueha S, Otsuji M. Tracking of intertissue migration reveals the origins of tumor-infiltrating monocytes. *Proc Natl Acad Sci USA.* 2014;111(21):7771–7776. doi:10.1073/pnas.1402914111
22. Liu Y, Cao X. The origin and function of tumor-associated macrophages. *Cell Mol Immunol.* 2015;12(1):1–4. doi:10.1038/cmi.2014.83
23. Wynn TA, Chawla A, Pollard JW. Macrophage biology in development, homeostasis and disease. *Nature.* 2013;496(7446):445–455. doi:10.1038/nature12034
24. Boutilier AJ, Elswa SF. Macrophage polarization states in the tumor microenvironment. *Int J Mol Sci.* 2021;22(13):6995. doi:10.3390/ijms22136995
25. Chen D, Zhang X, Li Z, Zhu B. Metabolic regulatory crosstalk between tumor microenvironment and tumor-associated macrophages. *Theranostics.* 2021;11(3):1016–1030. doi:10.7150/thno.51777
26. Pan Y, Yu Y, Wang X, Zhang T. Tumor-associated macrophages in tumor immunity. *Front Immunol.* 2020;11:583084. doi:10.3389/fimmu.2020.583084
27. Sica A, Mantovani A. Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest.* 2012;122(3):787–795. doi:10.1172/JCI59643
28. Shueng PW, Yu LY, Chiu HC, et al. Early phago-/endosomal escape of platinum drugs via ROS-responsive micelles for dual cancer chemo/immunotherapy. *Biomaterials.* 2021;276:121012. doi:10.1016/j.biomaterials.2021.121012
29. Gao J, Liang Y, Wang L. Shaping polarization of tumor-associated macrophages in cancer immunotherapy. *Front Immunol.* 2022;13:888713. doi:10.3389/fimmu.2022.888713
30. Nikovics K, Morin H, Riccobono D, Bendahmane A, Favier AL. Hybridization-chain-reaction is a relevant method for in situ detection of M2d-like macrophages in a mini-pig model. *FASEB J.* 2020;34(12):15675–15686. doi:10.1096/fj.202001496R
31. Khanduri I, Maru DM, Parra ER. Exploratory study of macrophage polarization and spatial distribution in colorectal cancer liver metastasis: a pilot study. *Front Immunol.* 2023;14:1223864. doi:10.3389/fimmu.2023.1223864
32. He Y, Han Y, Fan AH, et al. Multi-perspective comparison of the immune microenvironment of primary colorectal cancer and liver metastases. *J Transl Med.* 2022;20(1):454. doi:10.1186/s12967-022-03667-2
33. Cui YL, Li HK, Zhou HY, Zhang T, Li Q. Correlations of tumor-associated macrophage subtypes with liver metastases of colorectal cancer. *Asian Pac J Cancer Prev.* 2013;14(2):1003–1007. doi:10.7314/APJCP.2013.14.2.1003
34. Takahashi Y, Matsuo K, Shiozawa T, Suzuki K, Shimizu H, Tanaka K. Prognostic implications of histologic growth patterns and tumor-infiltrating macrophages in colorectal liver metastases. *Langenbecks Arch Surg.* 2023;408(1):6. doi:10.1007/s00423-022-02741-z
35. Iwata M, Haraguchi R, Kitazawa R, et al. Reduced chemokine C-C motif ligand 1 expression may negatively regulate colorectal cancer progression at liver metastatic sites. *J Cell & Mol Med.* 2024;28(7):e18193. doi:10.1111/jcmm.18193
36. Tu W, Gong J, Zhou Z, Tian D, Wang Z. TCF4 enhances hepatic metastasis of colorectal cancer by regulating tumor-associated macrophage via CCL2/CCR2 signaling. *Cell Death Dis.* 2021;12(10):882. doi:10.1038/s41419-021-04166-w
37. Grossman JG, Nywening TM, Belt BA, et al. Recruitment of CCR2(+) tumor associated macrophage to sites of liver metastasis confers a poor prognosis in human colorectal cancer. *Oncotarget.* 2018;7(9):e1470729. doi:10.1080/2162402X.2018.1470729
38. Kee J-Y, Ito A, Hojo S. CXCL16 suppresses liver metastasis of colorectal cancer by promoting TNF- α -induced apoptosis by tumor-associated macrophages. *BMC Cancer.* 2014;14(1):949. doi:10.1186/1471-2407-14-949
39. Wei X, Ye J, Pei Y, et al. Extracellular vesicles from colorectal cancer cells promote metastasis via the NOD1 signalling pathway. *J Extracell Vesicles.* 2022;11(9):e12264. doi:10.1002/jev2.12264
40. Chen J, Li Z, Yue C, et al. Human umbilical cord mesenchymal stem cell-derived exosomes carrying miR-1827 downregulate SUCNR1 to inhibit macrophage M2 polarization and prevent colorectal liver metastasis. *Apoptosis.* 2023;28(3–4):549–565. doi:10.1007/s10495-022-01798-x
41. Li M, Lai X, Zhao Y, et al. Loss of NDRG2 in liver microenvironment inhibits cancer liver metastasis by regulating tumor associated macrophages polarization. *Cell Death Dis.* 2018;9(2):248. doi:10.1038/s41419-018-0284-8
42. Zhang XL, Hu LP, Yang Q, et al. CTHRC1 promotes liver metastasis by reshaping infiltrated macrophages through physical interactions with TGF- β receptors in colorectal cancer. *Oncogene.* 2021;40(23):3959–3973. doi:10.1038/s41388-021-01827-0
43. Yue L, Xu X, Dai S, et al. Orosomucoid 1 promotes colorectal cancer progression and liver metastasis by affecting PI3K/AKT pathway and inducing macrophage M2 polarization. *Sci Rep.* 2023;13(1):14092. doi:10.1038/s41598-023-40404-1
44. Hirata W, Itatani Y, Masui H. Downregulation of osteoprotegerin in colorectal cancer cells promotes liver metastasis via activating tumor-associated macrophage. *Sci Rep.* 2023;13(1):22217. doi:10.1038/s41598-023-49312-w
45. Liu Y, Sun Q, Zhang C. STING-IRG1 inhibits liver metastasis of colorectal cancer by regulating the polarization of tumor-associated macrophages. *iScience.* 2023;26(8):107376. doi:10.1016/j.isci.2023.107376
46. Matsumura H, Kondo T, Ogawa K, et al. Kupffer cells decrease metastasis of colon cancer cells to the liver in the early stage. *Int J Oncol.* 2014;45(6):2303–2310. doi:10.3892/ijo.2014.2662
47. Shimizu Y, Amano H, Ito Y, et al. Angiotensin II subtype 1a receptor signaling in resident hepatic macrophages induces liver metastasis formation. *Cancer Sci.* 2017;108(9):1757–1768. doi:10.1111/cas.13306
48. Shang S, Ji X, Zhang L, et al. Macrophage ABHD5 suppresses NF κ B-dependent matrix metalloproteinase expression and cancer metastasis. *Cancer Res.* 2019;79(21):5513–5526. doi:10.1158/0008-5472.CAN-19-1059

49. Donadon M, Torzilli G, Cortese N, et al. Macrophage morphology correlates with single-cell diversity and prognosis in colorectal liver metastasis. *J Exp Med*. 2020;217(11). doi:10.1084/jem.20191847
50. Cortese N, Carriero R, Barbagallo M, et al. High-Resolution Analysis of Mononuclear Phagocytes Reveals GPNMB as a Prognostic Marker in Human Colorectal Liver Metastasis. *Cancer Immunol Res*. 2023;11(4):405–420. doi:10.1158/2326-6066.CIR-22-0462
51. Wijler LA, Viergever BJ, Strating E, et al. Onward spread from liver metastases is a major cause of multi-organ metastasis in a mouse model of metastatic colon cancer. *Cancers*. 2024;16(5):1073. doi:10.3390/cancers16051073
52. Gerovska D, Garcia-Gallastegi P, Crende O, et al. Badiola I: geromirs are downregulated in the tumor microenvironment during colon cancer colonization of the liver in a murine metastasis model. *Int J Mol Sci*. 2021;22(9):4819. doi:10.3390/ijms22094819
53. Zou J, Wu B, Lin C, Ding Q, Li J. MiR-216b targets CPEB4 to suppress colorectal cancer progression through inhibiting IL-10-mediated M2 polarization of tumor-associated macrophages. *Am J Transl Res*. 2022;14(11):8129–8145.
54. Zhao S, Mi Y, Guan B. Tumor-derived exosomal miR-934 induces macrophage M2 polarization to promote liver metastasis of colorectal cancer. *J hematol oncol*. 2020;13(1):156. doi:10.1186/s13045-020-00991-2
55. Chen Q, Deng Y, Li Y, et al. Association of preoperative aspartate aminotransferase to platelet ratio index with outcomes and tumour microenvironment among colorectal cancer with liver metastases. *Cancer Lett*. 2024;588:216778. doi:10.1016/j.canlet.2024.216778
56. Feng Y, Qiao S, Chen J, et al. M2-type macrophages and cancer-associated fibroblasts combine to promote colorectal cancer liver metastases. *Oncotargets Ther*. 2024;17:243–260. doi:10.2147/OTT.S447502
57. Liu X, Qin J, Nie J, et al. ANGPTL2+cancer-associated fibroblasts and SPP1+macrophages are metastasis accelerators of colorectal cancer. *Front Immunol*. 2023;14:1185208. doi:10.3389/fimmu.2023.1185208
58. Deng S, Cheng D, Wang J, et al. MYL9 expressed in cancer-associated fibroblasts regulate the immune microenvironment of colorectal cancer and promotes tumor progression in an autocrine manner. *J Exp Clin Cancer Res*. 2023;42(1):294. doi:10.1186/s13046-023-02863-2
59. Liu Y, Zhang Q, Xing B, et al. Immune phenotypic linkage between colorectal cancer and liver metastasis. *Cancer Cell*. 2022;40(4):424–437. e425. doi:10.1016/j.ccell.2022.02.013
60. Piao C, Cai L, Qiu S, Jia L, Song W, Du J. Complement 5a enhances hepatic metastases of colon cancer via monocyte chemoattractant protein-1-mediated inflammatory cell infiltration. *J Biol Chem*. 2015;290(17):10667–10676. doi:10.1074/jbc.M114.612622
61. Sun Y, Hu H, Liu Z, et al. Macrophage STING signaling promotes NK cell to suppress colorectal cancer liver metastasis via 4-1BBL/4-1BB co-stimulation. *J Immun Cancer*. 2023;11(3):e006481. doi:10.1136/jitc-2022-006481
62. Yuan N, Li X, Wang M, et al. Gut microbiota alteration influences colorectal cancer metastasis to the liver by remodeling the liver immune microenvironment. *Gut Liver*. 2022;16(4):575–588. doi:10.5009/gnl210177
63. Li R, Zhou R, Wang H, et al. Gut microbiota-stimulated cathepsin K secretion mediates TLR4-dependent M2 macrophage polarization and promotes tumor metastasis in colorectal cancer. *Cell Death Differ*. 2019;26(11):2447–2463. doi:10.1038/s41418-019-0312-y
64. Gu J, Xu X, Li X, et al. Tumor-resident microbiota contributes to colorectal cancer liver metastasis by lactylation and immune modulation. *Oncogene*. 2024;43(31):2389–2404. doi:10.1038/s41388-024-03080-7
65. Jia R, Shao S, Zhang P, et al. PRM1201 effectively inhibits colorectal cancer metastasis via shaping gut microbiota and short-chain fatty acids. *Phytomedicine*. 2024;132:155795. doi:10.1016/j.phymed.2024.155795
66. Borrelli C, Roberts M, Eletto D, et al. In vivo interaction screening reveals liver-derived constraints to metastasis. *Nature*. 2024;632(8024):411–418. doi:10.1038/s41586-024-07715-3
67. Bautista-Puig N, Barreiro-Gen M, Statulevičiūtė G, et al. Corrigendum to “Unraveling public perceptions of the sustainable development goals for better policy implementation”. *Sci total environ*. 2024;912:169114. doi:10.1016/j.scitotenv.2023.169114
68. Mohr AM, Gould JJ, Kubik JL, et al. Enhanced colorectal cancer metastases in the alcohol-injured liver. *Clin Exp Metastasis*. 2017;34(2):171–184. doi:10.1007/s10585-017-9838-x
69. Ogino S, Nowak JA, Hamada T, Milner DA Jr, Nishihara R. Insights into pathogenic interactions among environment, host, and tumor at the crossroads of molecular pathology and epidemiology. *Annu Rev Pathol*. 2019;14(1):83–103. doi:10.1146/annurev-pathmechdis-012418-012818
70. Inamura K, Hamada T, Bullman S, Ugai T, Yachida S, Ogino S. Cancer as microenvironmental, systemic and environmental diseases: opportunity for transdisciplinary microbiomics science. *Gut*. 2022;71(10):2107–2122. doi:10.1136/gutjnl-2022-327209
71. Nyström H, Naredi P, Berglund A, Palmqvist R, Tavelin B, Sund M. Liver-metastatic potential of colorectal cancer is related to the stromal composition of the tumour. *Anticancer Res*. 2012;32(12):5183–5191.
72. Nyström H, Naredi P, Hafström L, Sund M. Type IV collagen as a tumour marker for colorectal liver metastases. *Eur J Surg Oncol*. 2011;37(7):611–617. doi:10.1016/j.ejso.2011.04.010
73. Tsilimigras DI, Brodt P, Clavien PA, et al. Liver metastases. *Nat Rev Dis Primers*. 2021;7(1):27. doi:10.1038/s41572-021-00261-6
74. Cortese N, Soldani C, Franceschini B, et al. Macrophages in Colorectal Cancer Liver Metastases. *Cancers*. 2019;11(5):633. doi:10.3390/cancers11050633
75. Liu ZL, Chen HH, Zheng LL, Sun LP, Shi L. Angiogenic signaling pathways and anti-angiogenic therapy for cancer. *Signal Transduct Target Ther*. 2023;8(1):198. doi:10.1038/s41392-023-01460-1
76. Dallavalasa S, Beeraka NM, Basavaraju CG, et al. The role of tumor associated macrophages (TAMs) in cancer progression, chemoresistance, angiogenesis and metastasis - current status. *Curr Med Chem*. 2021;28(39):8203–8236. doi:10.2174/0929867328666210720143721
77. Wu X, Guan S, Lu Y, et al. Macrophage-derived SHP-2 inhibits the metastasis of colorectal cancer via Tie2-PI3K signals. *Oncol Res*. 2023;31(2):125–139. doi:10.32604/or.2023.028657
78. Freire Valls A, Knipper K, Giannakouri E, et al. VEGFR1(+) metastasis-associated macrophages contribute to metastatic angiogenesis and influence colorectal cancer patient outcome. *Clin Cancer Res*. 2019;25(18):5674–5685. doi:10.1158/1078-0432.CCR-18-2123
79. Kruse J, von Bernstorff W, Evert K, et al. Macrophages promote tumour growth and liver metastasis in an orthotopic syngeneic mouse model of colon cancer. *Int J Colorectal Dis*. 2013;28(10):1337–1349. doi:10.1007/s00384-013-1703-z
80. Sasaki T, Nakamura T, Rebhun RB, et al. Modification of the primary tumor microenvironment by transforming growth factor alpha-epidermal growth factor receptor signaling promotes metastasis in an orthotopic colon cancer model. *Am J Pathol*. 2008;173(1):205–216. doi:10.2353/ajpath.2008.071147
81. Zheng J, Yang M, Shao J, Miao Y, Han J, Du J. Chemokine receptor CX3CR1 contributes to macrophage survival in tumor metastasis. *Mol Cancer*. 2013;12(1):141. doi:10.1186/1476-4598-12-141

82. Wang D, Wang X, Si M, et al. Exosome-encapsulated miRNAs contribute to CXCL12/CXCR4-induced liver metastasis of colorectal cancer by enhancing M2 polarization of macrophages. *Cancer Lett.* 2020;474:36–52. doi:10.1016/j.canlet.2020.01.005
83. Wang D, Sun H, Wei J, Cen B, DuBois RN. CXCL1 is critical for premetastatic niche formation and metastasis in colorectal cancer. *Cancer Res.* 2017;77(13):3655–3665. doi:10.1158/0008-5472.CAN-16-3199
84. Imano M, Okuno K, Itoh T, Ishimaru E, Satou T, Shiozaki H. Increased osteopontin-positive macrophage expression in colorectal cancer stroma with synchronous liver metastasis. *World J Surg.* 2010;34(8):1930–1936. doi:10.1007/s00268-010-0582-5
85. Yang M, Shao JH, Miao YJ, et al. Tumor cell-activated CARD9 signaling contributes to metastasis-associated macrophage polarization. *Cell Death Differ.* 2014;21(8):1290–1302. doi:10.1038/cdd.2014.45
86. Bravo-Cordero JJ, Hodgson L, Condeelis J. Directed cell invasion and migration during metastasis. *Curr Opin Cell Biol.* 2012;24(2):277–283. doi:10.1016/j.ceb.2011.12.004
87. Pastushenko I, Blanpain C. EMT transition states during tumor progression and metastasis. *Trends Cell Biol.* 2019;29(3):212–226. doi:10.1016/j.tcb.2018.12.001
88. Gazzillo A, Polidoro MA, Soldani C, Franceschini B, Lleo A, Donadon M. Relationship between epithelial-to-mesenchymal transition and tumor-associated macrophages in colorectal liver metastases. *Int J Mol Sci.* 2022;23(24):16197. doi:10.3390/ijms232416197
89. Wei C, Yang C, Wang S, et al. Crosstalk between cancer cells and tumor associated macrophages is required for mesenchymal circulating tumor cell-mediated colorectal cancer metastasis. *Mol Cancer.* 2019;18(1):64. doi:10.1186/s12943-019-0976-4
90. Deng Q, Geng Y, Zhao L, et al. NLRP3 inflammasomes in macrophages drive colorectal cancer metastasis to the liver. *Cancer Lett.* 2019;442:21–30. doi:10.1016/j.canlet.2018.10.030
91. Liu X, Wang X, Yang Q, et al. Th17 cells secrete TWEAK to trigger epithelial-mesenchymal transition and promote colorectal cancer liver metastasis. *Cancer Res.* 2024;84(8):1352–1371. doi:10.1158/0008-5472.CAN-23-2123
92. Wang S, Yao Y, Li H, Zheng G, Lu S, Chen W. Tumor-associated macrophages (TAMs) depend on Shp2 for their anti-tumor roles in colorectal cancer. *Am J Cancer Res.* 2019;9(9):1957–1969.
93. Fei R, Zhang Y, Wang S, Xiang T, Chen W. $\alpha 7$ nicotinic acetylcholine receptor in tumor-associated macrophages inhibits colorectal cancer metastasis through the JAK2/STAT3 signaling pathway. *Oncol Rep.* 2017;38(5):2619–2628. doi:10.3892/or.2017.5935
94. Patras L, Shaashua L, Matei I, Lyden D. Immune determinants of the pre-metastatic niche. *Cancer Cell.* 2023;41(3):546–572. doi:10.1016/j.ccell.2023.02.018
95. Pei W, Wei K, Wu Y, et al. Colorectal cancer tumor cell-derived exosomal miR-203a-3p promotes CRC metastasis by targeting PTEN-induced macrophage polarization. *Gene.* 2023;885:147692. doi:10.1016/j.gene.2023.147692
96. Geng Y, Feng J, Huang H, et al. Single-cell transcriptome analysis of tumor immune microenvironment characteristics in colorectal cancer liver metastasis. *Ann transl Med.* 2022;10(21):1170. doi:10.21037/atm-22-5270
97. Takano Y, Masuda T, Iinuma H, et al. Circulating exosomal microRNA-203 is associated with metastasis possibly via inducing tumor-associated macrophages in colorectal cancer. *Oncotarget.* 2017;8(45):78598–78613. doi:10.18632/oncotarget.20009
98. Shao Y, Chen T, Zheng X, et al. Colorectal cancer-derived small extracellular vesicles establish an inflammatory premetastatic niche in liver metastasis. *Carcinogenesis.* 2018;39(11):1368–1379. doi:10.1093/carcin/bgy115
99. Katoh H, Wang D, Daikoku T, Sun H, Dey SK, Dubois RN. CXCR2-expressing myeloid-derived suppressor cells are essential to promote colitis-associated tumorigenesis. *Cancer Cell.* 2013;24(5):631–644. doi:10.1016/j.ccr.2013.10.009
100. Salmaninejad A, Valilou SF, Soltani A, et al. Tumor-associated macrophages: role in cancer development and therapeutic implications. *Cell Oncol.* 2019;42(5):591–608. doi:10.1007/s13402-019-00453-z
101. Minder P, Zajac E, Quigley JP, Deryugina EI. EGFR regulates the development and microarchitecture of intratumoral angiogenic vasculature capable of sustaining cancer cell intravasation. *Neoplasia.* 2015;17(8):634–649. doi:10.1016/j.neo.2015.08.002
102. Cardoso AP, Pinto ML, Pinto AT, et al. Macrophages stimulate gastric and colorectal cancer invasion through EGFR Y(1086), c-Src, Erk1/2 and Akt phosphorylation and smallGTPase activity. *Oncogene.* 2014;33(16):2123–2133. doi:10.1038/onc.2013.154
103. Afik R, Zigmund E, Vugman M, et al. Tumor macrophages are pivotal constructors of tumor collagenous matrix. *J Exp Med.* 2016;213(11):2315–2331. doi:10.1084/jem.20151193
104. Weis S, Cui J, Barnes L, Cheresh D. Endothelial barrier disruption by VEGF-mediated Src activity potentiates tumor cell extravasation and metastasis. *J Cell Biol.* 2004;167(2):223–229. doi:10.1083/jcb.200408130
105. Zhu M, Bai L, Liu X, et al. Silence of a dependence receptor CSF1R in colorectal cancer cells activates tumor-associated macrophages. *J Immunol Cancer.* 2022;10(12):e005610. doi:10.1136/jitc-2022-005610
106. Qiao T, Yang W, He X, et al. Dynamic differentiation of F4/80+ tumor-associated macrophage and its role in tumor vascularization in a syngeneic mouse model of colorectal liver metastasis. *Cell Death Dis.* 2023;14(2):117. doi:10.1038/s41419-023-05626-1
107. Limagne E, Thibaudin M, Nuttin L, et al. Trifluridine/tipiracil plus oxaliplatin improves PD-1 blockade in colorectal cancer by inducing immunogenic cell death and depleting macrophages. *Cancer Immunol Res.* 2019;7(12):1958–1969. doi:10.1158/2326-6066.CIR-19-0228
108. Li JJ, Wang JH, Tian T, et al. The liver microenvironment orchestrates FGL1-mediated immune escape and progression of metastatic colorectal cancer. *Nat Commun.* 2023;14(1):6690. doi:10.1038/s41467-023-42332-0
109. Georgoudaki AM, Prokopec KE, Boura VF, et al. Reprogramming tumor-associated macrophages by antibody targeting inhibits cancer progression and metastasis. *Cell Rep.* 2016;15(9):2000–2011. doi:10.1016/j.celrep.2016.04.084
110. Shen L, Sundstedt A, Ciesielski M, et al. Tasquinimod modulates suppressive myeloid cells and enhances cancer immunotherapies in murine models. *Cancer Immunol Res.* 2015;3(2):136–148. doi:10.1158/2326-6066.CIR-14-0036
111. Halama N, Zoernig I, Berthel A, et al. Tumoral immune cell exploitation in colorectal cancer metastases can be targeted effectively by anti-CCR5 therapy in cancer patients. *Cancer Cell.* 2016;29(4):587–601. doi:10.1016/j.ccell.2016.03.005
112. de Carvalho TG, Lara P, Jorquera-Cordero C, et al. Inhibition of murine colorectal cancer metastasis by targeting M2-TAM through STAT3/NF-kB/AKT signaling using macrophage 1-derived extracellular vesicles loaded with oxaliplatin, retinoic acid, and Libidibia ferrea. *Biomed Pharmacother.* 2023;168:115663. doi:10.1016/j.biopha.2023.115663
113. Fiegle E, Doleschel D, Koletnik S, et al. Dual CTLA-4 and PD-L1 blockade inhibits tumor growth and liver metastasis in a highly aggressive orthotopic mouse model of colon cancer. *Neoplasia.* 2019;21(9):932–944. doi:10.1016/j.neo.2019.07.006

114. Huang S, Ding D, Lan T, et al. Multifunctional nanodrug performs sonodynamic therapy and inhibits TGF- β to boost immune response against colorectal cancer and liver metastasis. *Acta Biomater.* 2023;164:538–552. doi:10.1016/j.actbio.2023.04.001
115. Chen C, Yao X, Xu Y, et al. Dahuang Zhechong Pill suppresses colorectal cancer liver metastasis via ameliorating exosomal CCL2 primed pre-metastatic niche. *J Ethnopharmacol.* 2019;238:111878. doi:10.1016/j.jep.2019.111878
116. Zhao L, Zhu X, Ni Y, You J, Li A. Xiaoyaosan, a traditional Chinese medicine, inhibits the chronic restraint stress-induced liver metastasis of colon cancer in vivo. *Pharm Biol.* 2020;58(1):1085–1091. doi:10.1080/13880209.2020.1839513
117. Tang D, Wang H, Deng W, et al. Mechanism of bufalin inhibition of colon cancer liver metastasis by regulating M2-type polarization of Kupffer cells induced by highly metastatic colon cancer cells. *Apoptosis.* 2024;29(5–6):635–648. doi:10.1007/s10495-023-01930-5
118. Lv T, Lou Y, Yan Q, Nie L, Cheng Z, Zhou X. Phosphorylation: new star of pathogenesis and treatment in steatotic liver disease. *Lipids Health Dis.* 2024;23(1). doi:10.1186/s12944-024-02037-9
119. Sanchez-Lopez E, Flashner-Abramson E, Shalpour S, et al. Targeting colorectal cancer via its microenvironment by inhibiting IGF-1 receptor-insulin receptor substrate and STAT3 signaling. *Oncogene.* 2016;35(20):2634–2644. doi:10.1038/ncr.2015.326
120. Razak AR, Cleary JM, Moreno V, et al. Safety and efficacy of AMG 820, an anti-colony-stimulating factor 1 receptor antibody, in combination with pembrolizumab in adults with advanced solid tumors. *J Immun Cancer.* 2020;8(2):e001006. doi:10.1136/jitc-2020-001006
121. Teng Y, Mu J, Hu X, et al. Grapefruit-derived nanovectors deliver miR-18a for treatment of liver metastasis of colon cancer by induction of M1 macrophages. *Oncotarget.* 2016;7(18):25683–25697. doi:10.18632/oncotarget.8361
122. Gallo G, Vescio G, De Paola G, Sammarco G. Therapeutic targets and tumor microenvironment in colorectal cancer. *J Clin Med.* 2021;10(11):2295. doi:10.3390/jcm10112295
123. Yang Y, Guo J, Huang L. Tackling TAMs for cancer immunotherapy: it's nano time. *Trends Pharmacol Sci.* 2020;41(10):701–714. doi:10.1016/j.tips.2020.08.003

Publish your work in this journal

The Journal of Inflammation Research is an international, peer-reviewed open-access journal that welcomes laboratory and clinical findings on the molecular basis, cell biology and pharmacology of inflammation including original research, reviews, symposium reports, hypothesis formation and commentaries on: acute/chronic inflammation; mediators of inflammation; cellular processes; molecular mechanisms; pharmacology and novel anti-inflammatory drugs; clinical conditions involving inflammation. The manuscript management system is completely online and includes a very quick and fair peer-review system. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/journal-of-inflammation-research-journal>