

## Supplemental Material

### Molecular identification of bulbospinal ON neurons by GPER which drives pain and morphine tolerance

#### Authors

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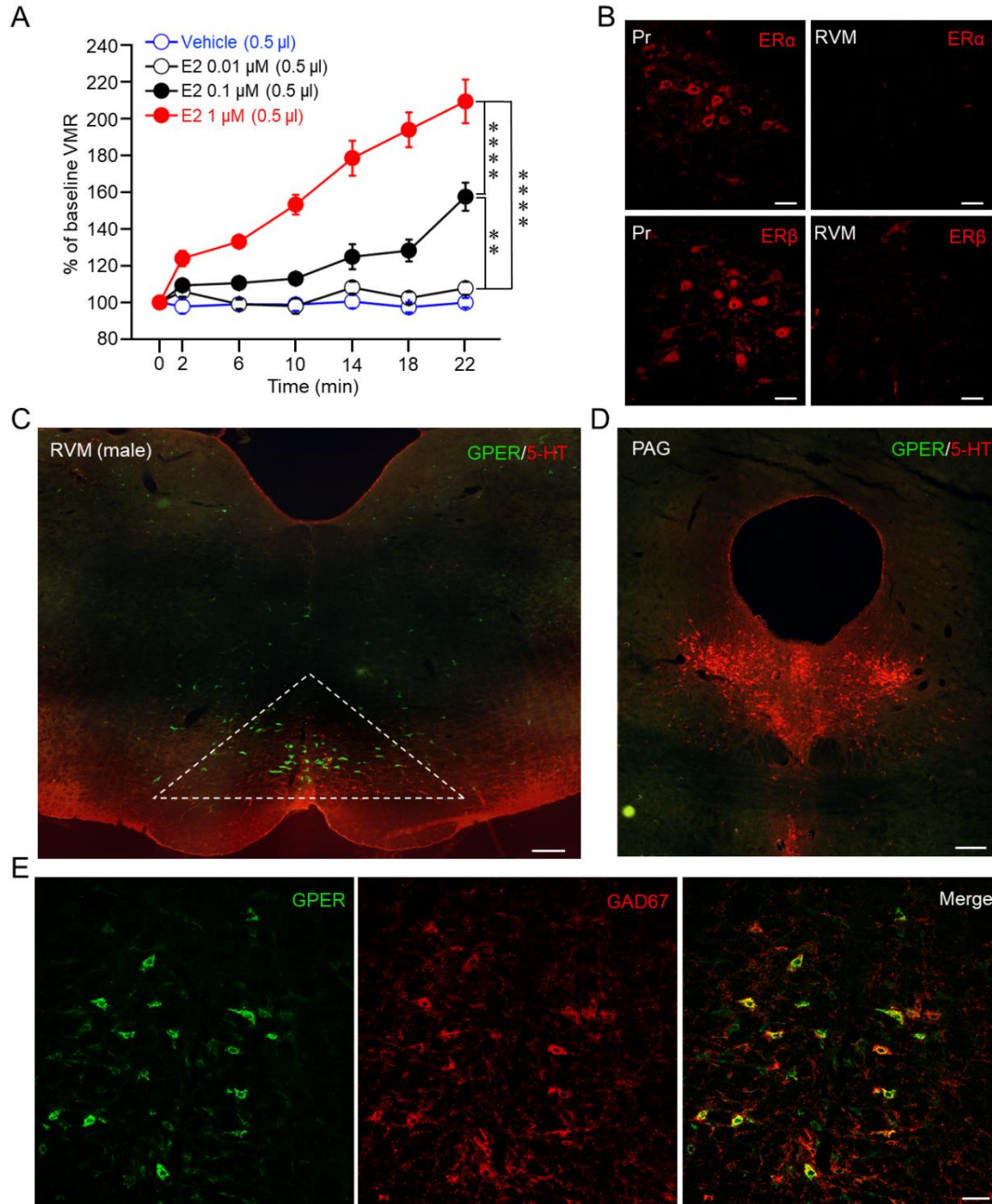
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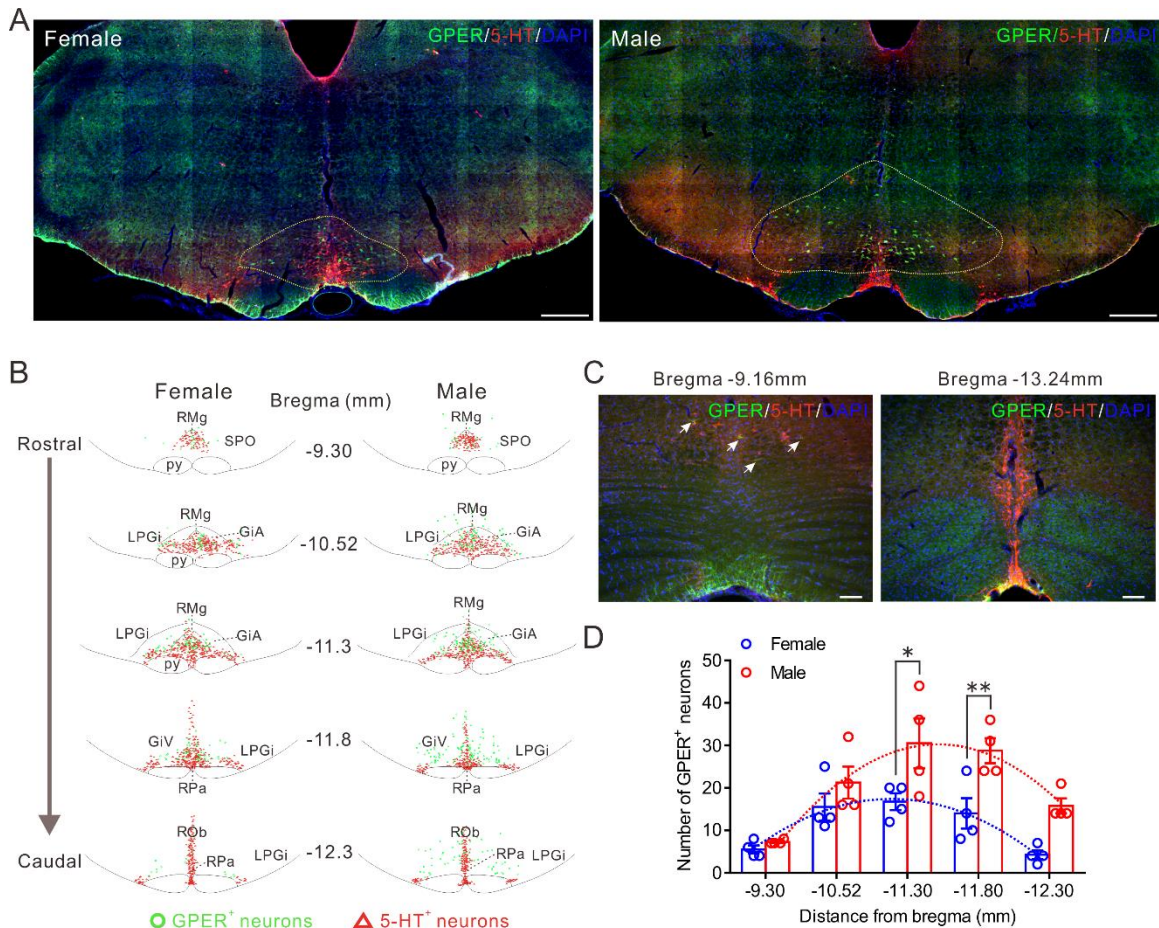
**Conflict of interest statement:** The authors have declared that no conflict of interest exists.

## Supplemental Figures

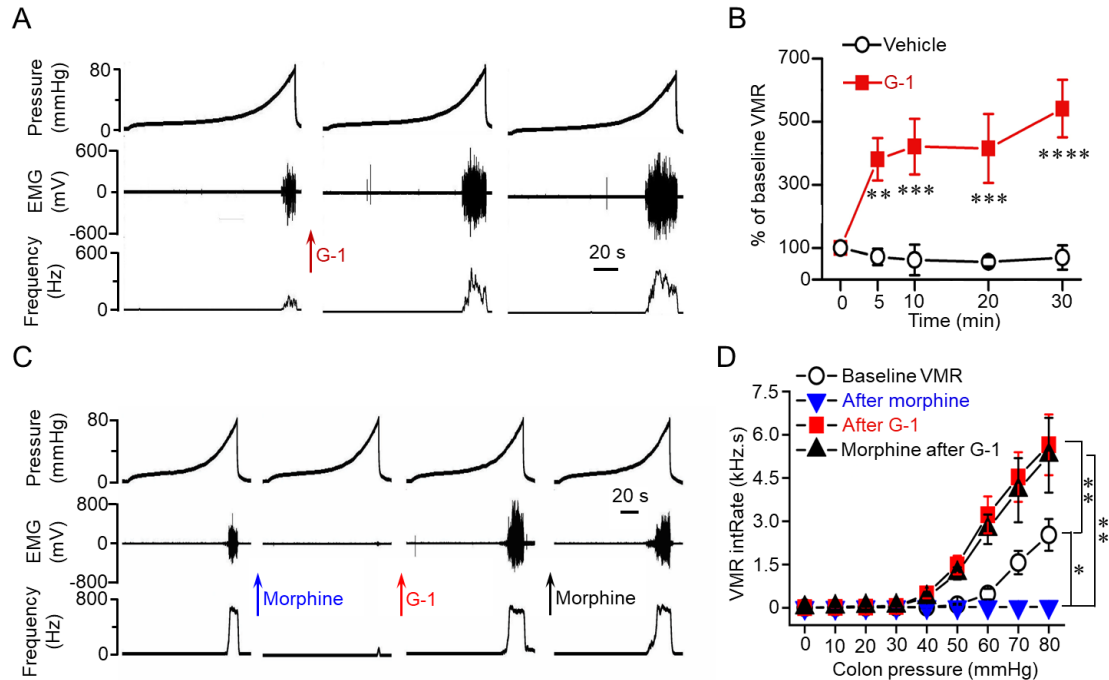


**Supplemental Figure 1. GPER is enriched in RVM.** (A) E2 applied to the RVM resulted the incremental increases in colorectal distension (CRD)-induced visceral motor responses (VMR) in a dose-dependent manner.  $n=6$  female rats for each group. (B) Immunofluorescent staining of the medulla oblongata revealed strong ER $\alpha$  and ER $\beta$  immunoreactivity in the prepositus nucleus (Pr) but not the RVM. Scale bars: 40  $\mu$ m. (C)

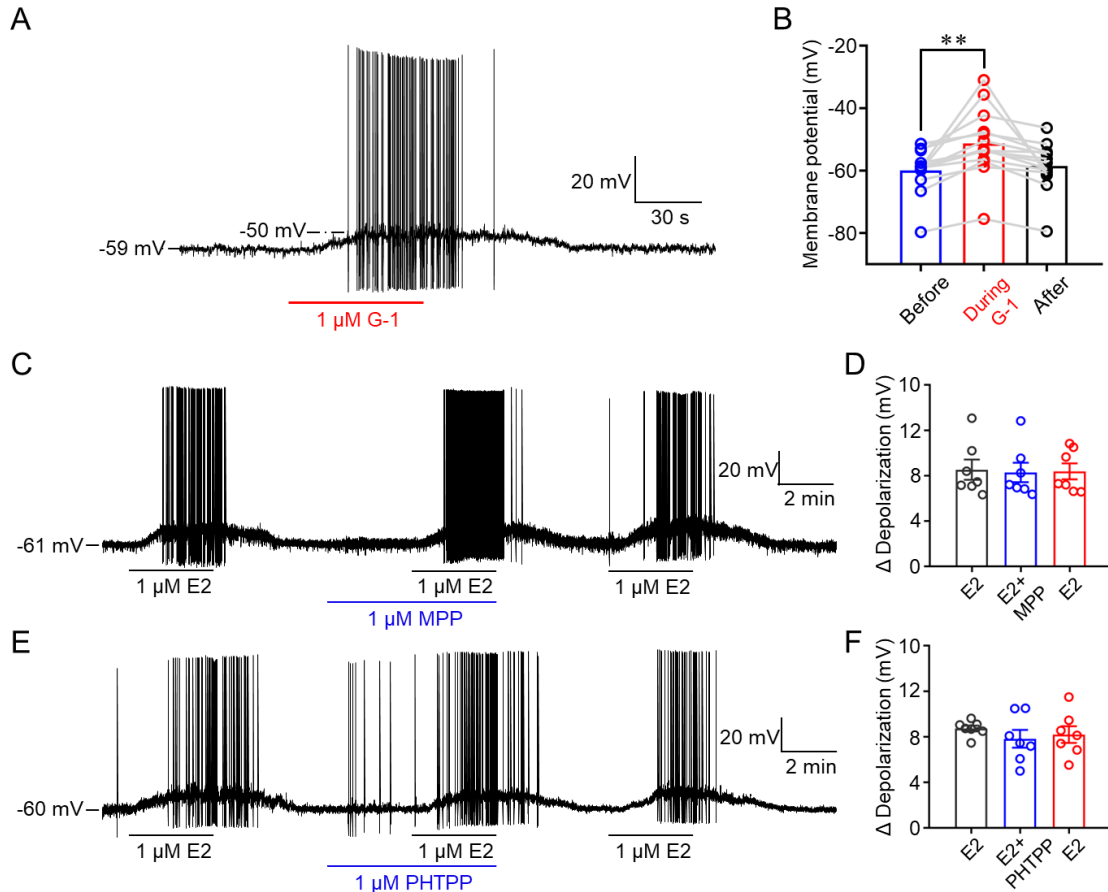
Distinctive GPER immunoreactivity in the RVM of male rats. Scale bar: 200  $\mu\text{m}$ . **(D)** GPER was hardly detectable in PAG and the adjacent serotonergic dorsal raphe nuclei. Scale bar: 200  $\mu\text{m}$ . **(E)** Co-labelling of GPER with GABA synthetase GAD67 in RVM. Scale bar: 50  $\mu\text{m}$ . Data are representative of at least 3 independent experiments. Data are presented as the mean  $\pm$  SEM.  $**P<0.01$  and  $****P<0.0001$ , by 1-way ANOVA with Tukey post hoc test **(A)**.



**Supplemental Figure 2. The rostral-caudal distribution of GPER<sup>+</sup> neurons in the ventral medulla of female and male rats.** (A) Representative double fluorescence IHC for GPER and 5-HT in a section of medulla from female (left) and male rats (right). Scale bars: 500  $\mu$ m. (B) A plot of the rostral-caudal distribution of GPER<sup>+</sup> and 5-HT<sup>+</sup> neurons in the medulla in female and male rats (pooled from 4 female and 4 male rats). (C) GPER and 5-HT immunofluorescence in the rostral (left) and caudal margin (right) of RVM from female rat. Arrow indicates a few GPER<sup>+</sup> neurons are scattered in the rostral margin of RVM. Scale bars: 100  $\mu$ m. (D) Statistical analysis of the rostral-caudal distribution of GPER<sup>+</sup> neurons in the RVM of female and male rats.  $n=4$  rats for each group. Data are representative of 4 independent experiments. Data are presented as the mean  $\pm$  SEM. \* $P<0.05$  and \*\* $P<0.01$ , by 2-way ANOVA followed by Bonferroni post hoc test (D).



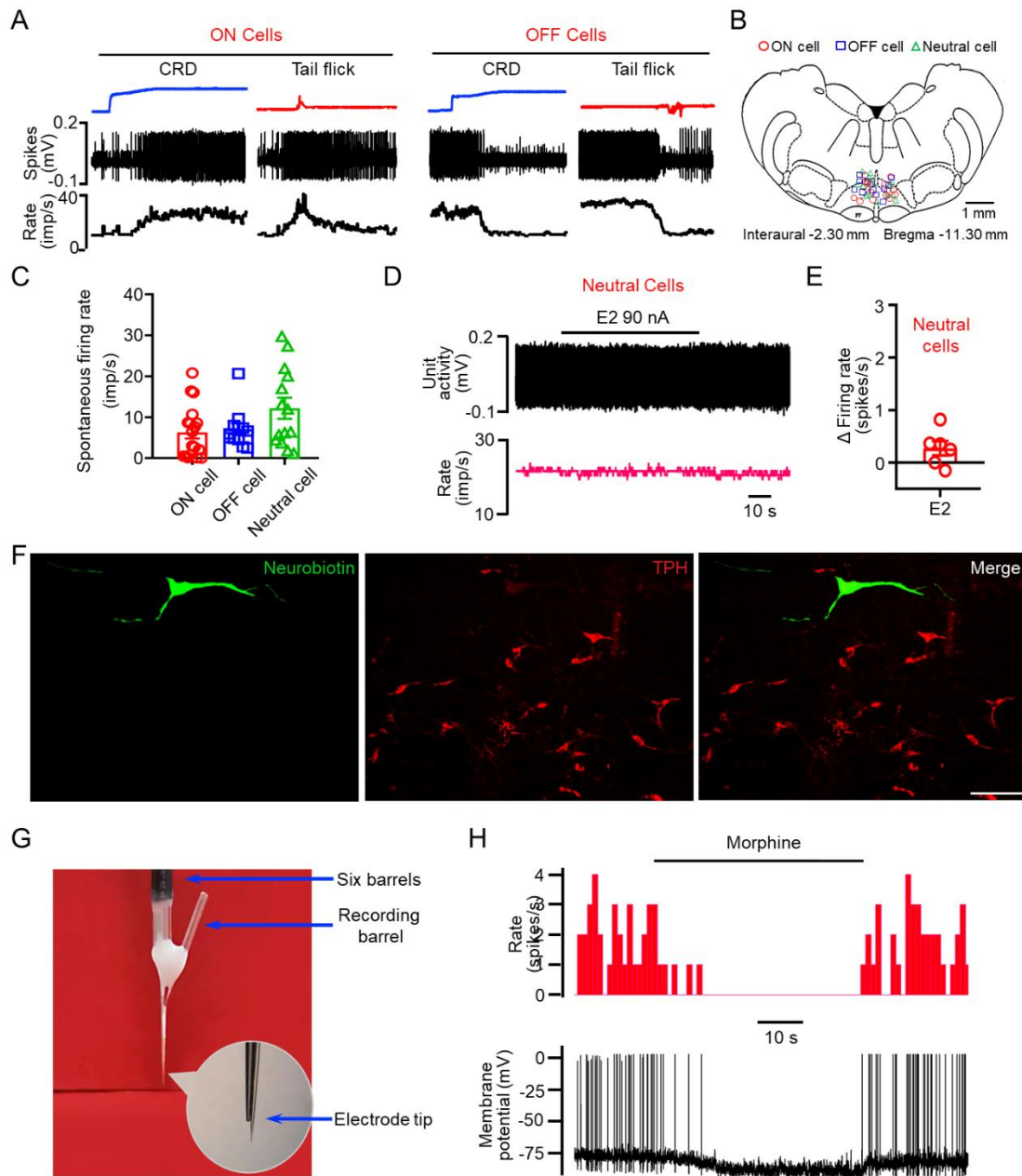
**Supplemental Figure 3. Activation of GPER results in colorectal hyperalgesia and attenuation of morphine analgesia.** (A and B) GPER agonist G-1 applied to RVM led to incremental increases in ramp CRD-induced VMR.  $n=6-7$  rats for each group. (C and D) G-1 applied to RVM negated the analgesic effect of morphine (also microinjected to RVM).  $n=5$  rats. Data are representative of at least 5 independent experiments. Data are presented as the mean  $\pm$  SEM. \* $P<0.05$ , \*\* $P<0.01$ , \*\*\* $P<0.001$  and \*\*\*\* $P<0.0001$ , by 2-way ANOVA followed by Bonferroni post hoc test (B) and paired 2-tailed Student's  $t$  test (D).



**Supplemental Figure 4. GPER mediates E2-induced depolarization of RVM neurons.**

(**A** and **B**) GPER selective agonist G-1 caused significant depolarization of RVM neurons in medulla slice in vitro.  $n=14$  cells from 3 rats. (**C** and **D**)  $ER\alpha$  antagonist MPP was without significant effect on E2-induced depolarization in RVM neurons.  $n=7$  cells from 2 rats. (**E** and **F**)  $ER\beta$  antagonist PHTPP did not affect E2-induced depolarization of RVM neurons.  $n=7$  cells from 2 rats. Data are representative of at least 2 independent experiments. Data are presented as the mean  $\pm$  SEM.  $**P<0.01$ , by paired 2-tailed Student's  $t$  test (**B**).

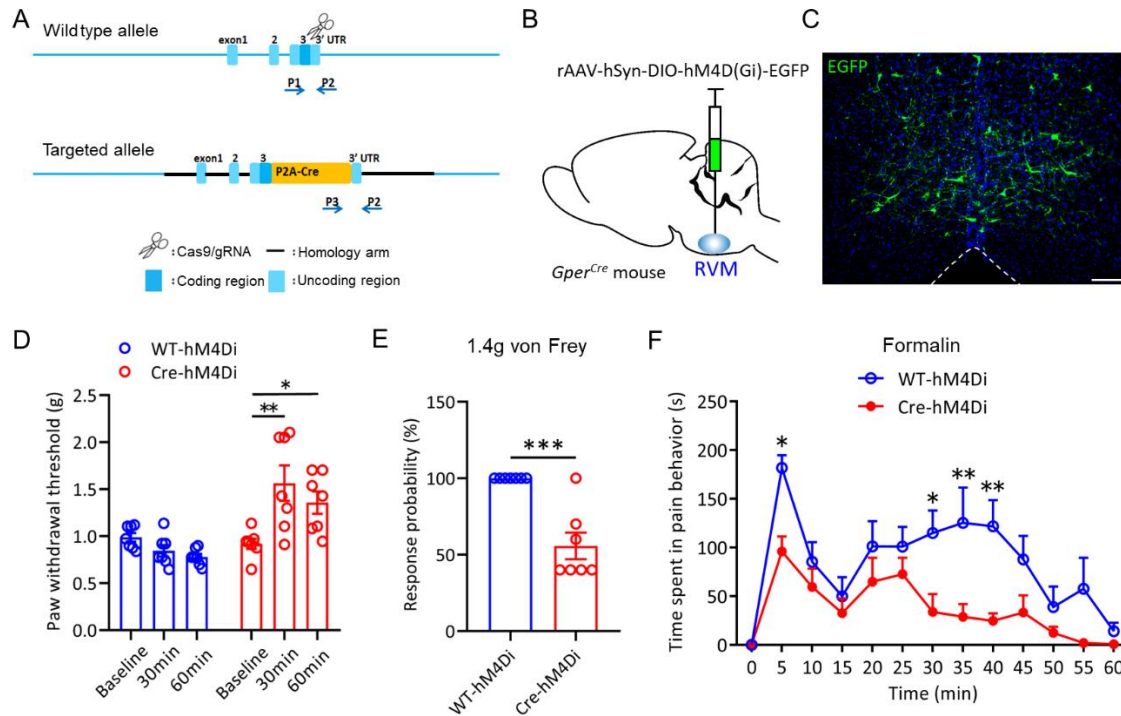




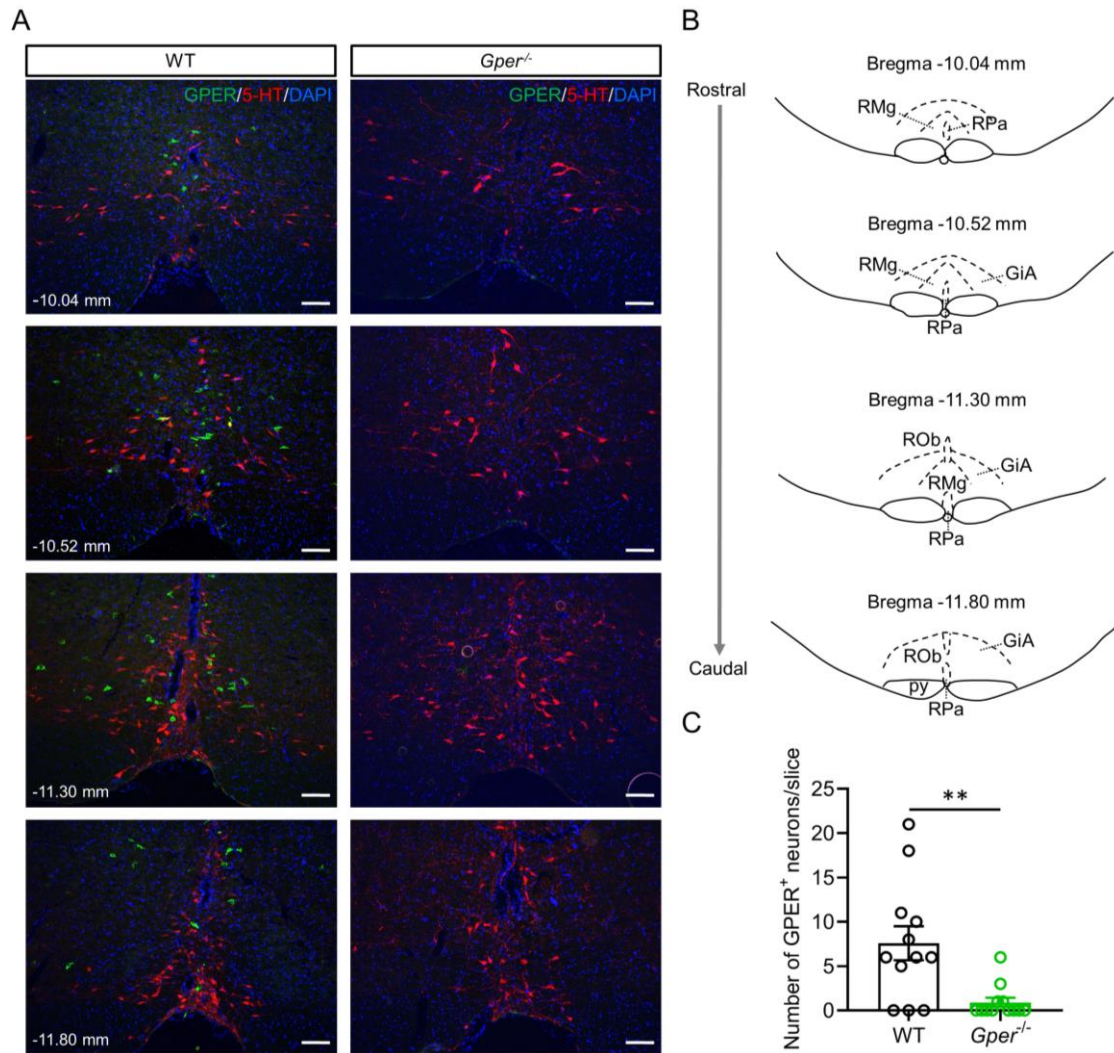
**Supplemental Figure 5. Electrophysiological recording and functional identification of ON and OFF neurons in the rat *in vivo*.** (A) Representative ON (left panels) or OFF cell (right panels) responses to noxious colorectal distention (CRD) and noxious thermal stimulation of the tail (Tail flick). (B and C) Anatomical position and baseline firing rate of RVM ON, OFF and Neutral cells recorded in our experiments (n=18 for ON cells, n=12 for OFF cells and n=14 for Neutral cells). (D and E) The firing rate of RVM neutral cells was not affected by iontophoretic application of E2 (n=6). (F) Representative IHC images illustrating neurobiotin-labeled RVM ON cell was not immunoreactive to TPH. Scale bar:

50  $\mu\text{m}$ . **(G)** Image of the multi-barrel piggyback electrodes used for intracellular recording and iontophoresis. **(H)** Iontophoretically applied morphine led to hyperpolarization and decreased firing rate of intracellularly-recorded RVM ON cells. Data are representative of at least 3 independent experiments. Data are presented as the mean  $\pm$  SEM.

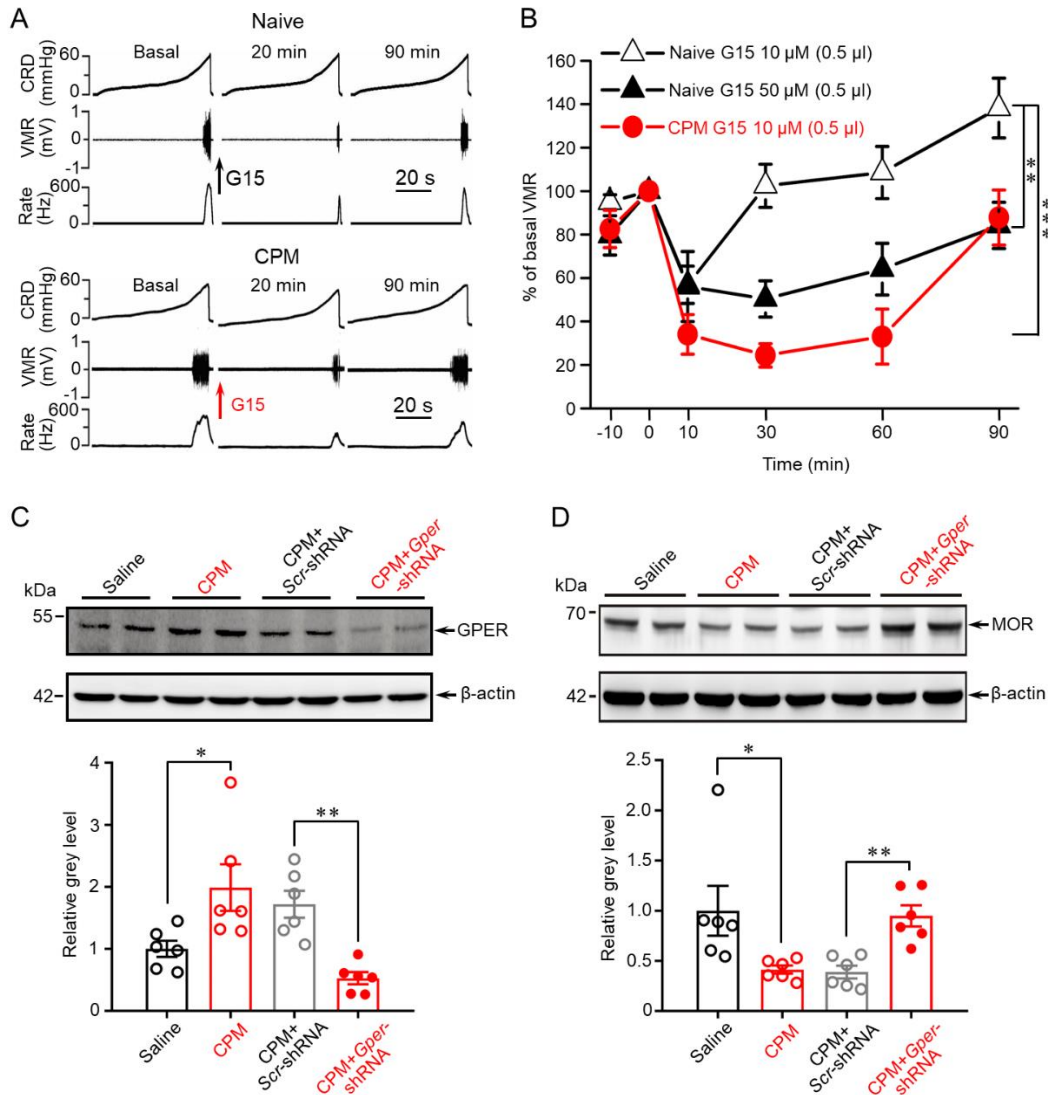




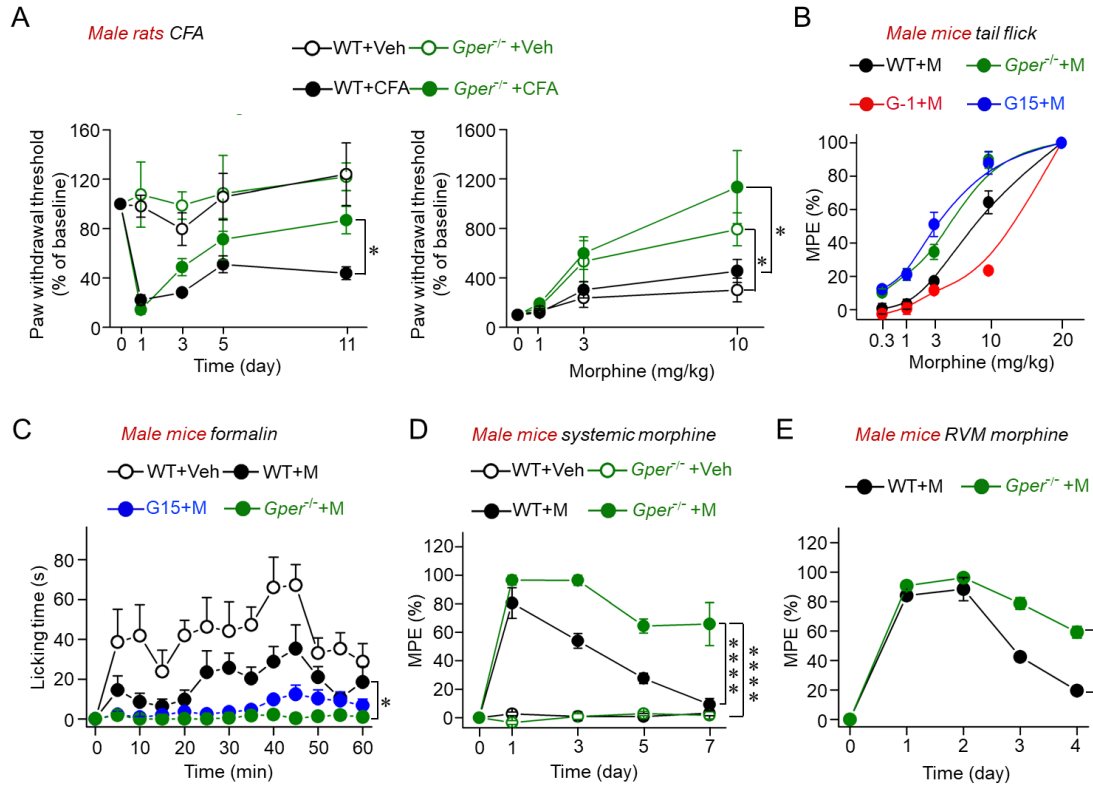
**Supplemental Figure 6. Chemogenetic silencing of RVM GPER<sup>+</sup> neurons attenuates acute mechanical and inflammatory pain.** (A) Schematic diagram of *Gper<sup>Cre</sup>* mice construction strategy. (B) Schematic showing the injection of rAAV-hSyn-DIO-hM4D(Gi)-EGFP into the RVM of *Gper<sup>Cre</sup>* mice. (C) Expression of rAAV-hSyn-DIO-hM4D(Gi)-EGFP in the RVM after viral infection. Scale bar: 100  $\mu$ m. (D) Paw withdrawal threshold was significantly increased following clozapine-N-oxide (CNO)-induced chemogenetic silencing of RVM GPER<sup>+</sup> neurons. n=7 mice for each group. (E) Frequency of withdrawal responses to repeated probing of hind paw with 1.4g von Frey filaments was decreased following CNO-induced chemogenetic silencing of RVM GPER<sup>+</sup> neurons. n=7 mice for each group. (F) Formalin-induced pain behavior was significantly decreased following CNO-induced chemogenetic silencing of RVM GPER<sup>+</sup> neurons. n=6 mice for each group. Data are representative of 3 independent experiments. Data are presented as the mean  $\pm$  SEM. \* $P$ <0.05, \*\* $P$ <0.01 and \*\*\* $P$ <0.001, by 1-way ANOVA with Tukey post hoc test (D), unpaired 2-tailed Student's  $t$  test (E), and 2-way ANOVA followed by Bonferroni post hoc test (F).



**Supplemental Figure 7. Validation of specificity for LS-A4272 by immunofluorescent staining in *Gper*<sup>-/-</sup> rats.** (A) Both GPER (green) and 5-HT (red) were abundantly expressed in RVM of WT female rats, and GPER immunoreactivity was exclusively confined to non-serotonergic neurons. The GPER immunoreactivity was markedly reduced in RVM of *Gper*<sup>-/-</sup> rats, in which only some non-specific green immunofluorescent responses to the GPER primary or secondary antibody could be seen in the middle edge of RVM slice. Scale bars: 100  $\mu$ m. From top to bottom are representative rostral to caudal RVM sections relative to the bregma, as illustrated in (B). (C) Statistical analysis of number of GPER<sup>+</sup> neurons in the RVM of WT and *Gper*<sup>-/-</sup> female rats. n=12 slices from 2 female rat for each group. Data are representative of 2 independent experiments. Data are presented as the mean  $\pm$  SEM. \*\* $P$ <0.01, by unpaired 2-tailed Student's  $t$  test (C).

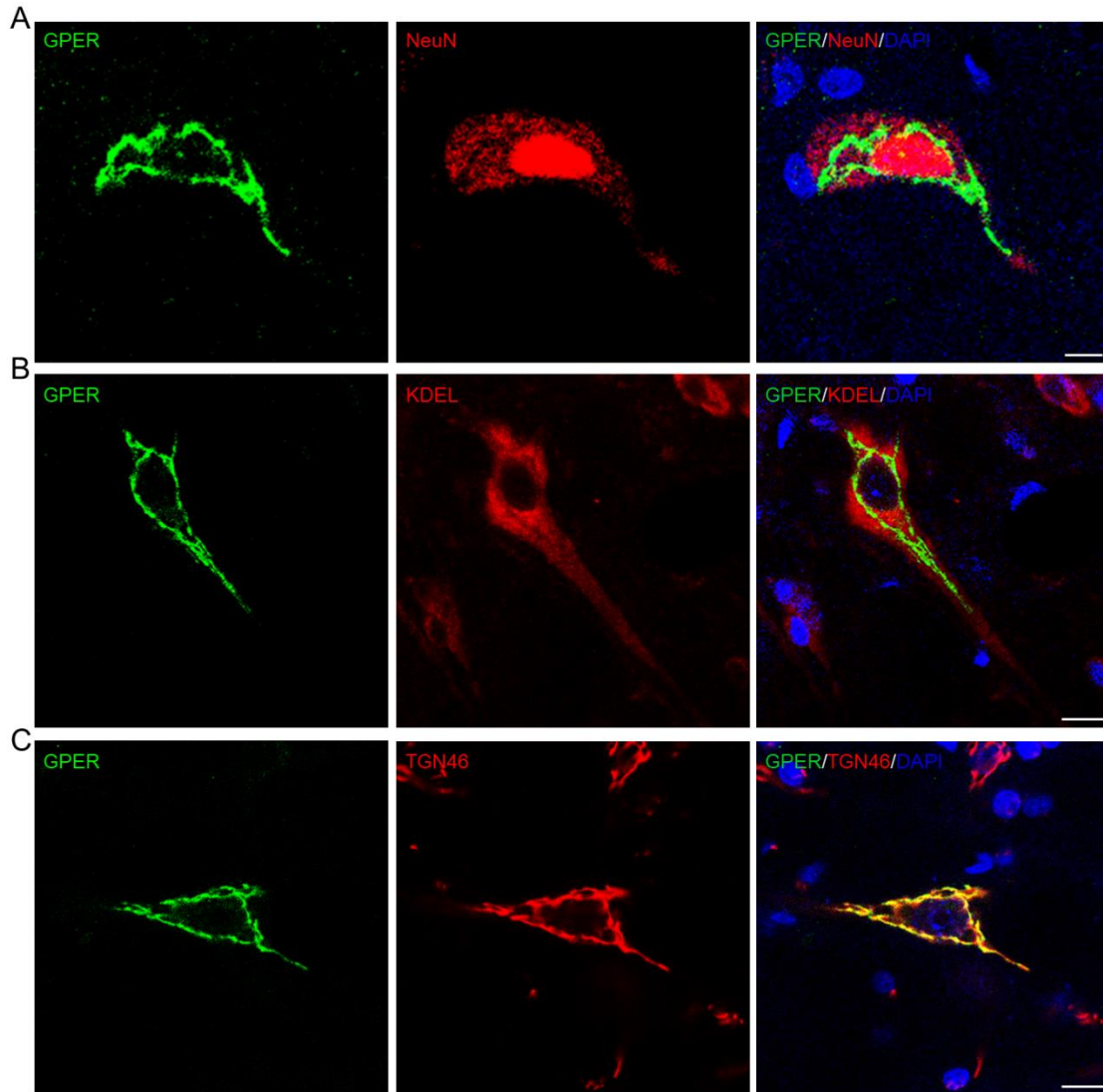


**Supplemental Figure 8. GPER signaling contributes to CPM-induced colorectal hypersensitivity.** (A and B) The effect of G15 microinjected to RVM on ramp CRD-induced visceral motor responses (VMR) in Naive or CPM-treated rats.  $n=6$  rats for each group. (C) GPER expression was increased in CPM model, while shRNA treatment knock down GPER in the RVM of CPM-treated female rats.  $n=6$  rats for each group. (D) MOR expression was decreased in CPM model, while *Gper* knockdown shRNA treatment reversed the down-regulation of MOR in RVM of CPM-treated female rats.  $n=6$  rats for each group. Data are representative of at least 2 independent experiments. Data are presented as the mean  $\pm$  SEM. \* $P$ <0.05, \*\* $P$ <0.01 and \*\*\* $P$ <0.001, by 1-way ANOVA with Tukey post hoc test (B-D).



**Supplemental Figure 9. GPER signaling modulates morphine analgesia and tolerance in male rats and mice.** (A) *Gper*<sup>-/-</sup> male rats displayed decreased hyperalgesia (left panel) and higher sensitivity to morphine analgesia compared with the WT male rats following intraplantar injection of CFA (right panel). n=5-6 rats. (B) In the tail flick test, either pharmacological blockade of GPER with G15 or genetic ablation of GPER (*Gper*<sup>-/-</sup>) led to a leftward shift of the dose-response curve of morphine analgesia in male rats. n=5-7 rats for each group. (C) In the formalin test, either pharmacological blockade of GPER with G15 or genetic ablation of GPER (*Gper*<sup>-/-</sup>) led to enhanced morphine analgesia in male mice. n=5-6 mice. (D) Development of systemic morphine tolerance was slowed in *Gper*<sup>-/-</sup> male mice compared with the wild type male mice. n=5 mice for each group. (E) Development of tolerance to locally-injected (RVM) morphine was slowed in *Gper*<sup>-/-</sup> male mice. n=6 mice for each group. Data are representative of at least 3 independent experiments. Data are presented as the mean  $\pm$  SEM. \* $P$ <0.05 and \*\*\*\* $P$ <0.0001, by 1-way ANOVA with Tukey post hoc test (A-D) and unpaired 2-tailed Student's  $t$  test (E).





**Supplemental Figure 10. GPER seems to be distinctively located on the golgi apparatus.** (A) Double staining for GPER and NeuN (a neuronal marker) showing intracellular localization of GPER immunoreactivity in RVM neurons. Scale bar: 5  $\mu$ m. (B) Double staining for GPER and KDEL (which stains endoplasmic reticulum) excluded localization of GPER in ER. Scale bar: 10  $\mu$ m. (C) Double staining for GPER and TGN46 (a marker of Golgi apparatus) showing localization of GPER in Golgi apparatus. Scale bar: 10  $\mu$ m. Data are representative of 3 independent experiments.