





## Genome Sequences of Five *Streptomyces* Strains Isolated at Microscale

Matthieu Nicault, a,b Abdoul-Razak Tidjani, a Eric Gelhaye, b Cyril Bontemps, a Pierre Leblonda

<sup>a</sup>Université de Lorraine, INRA, DynAMic, Nancy, France <sup>b</sup>Université de Lorraine, INRA, IAM, Nancy, France

**ABSTRACT** The genomes of five *Streptomyces* strains belonging to the same soil community were sequenced and assembled. The strains, which were isolated at microscale, belonged to different *Streptomyces* species. This sample provides access to understand the functioning of a *Streptomyces* community in an ecological context.

treptomyces organisms are soil-dwelling bacteria that play a central role in soil homeostasis (biogeochemical cycle and biotic interactions) (1). Their large linear chromosomes code for a wide variety of specialized metabolites (e.g., antibiotics and antifungals) and enzymes of high biotechnological and medical interest (2, 3). Here, we report the genome sequences of five Streptomyces strains isolated from four grains of soil. These grains had a size on the order of cubic millimeters and were collected at a distance of a few centimeters from each other in a French forest soil. Each grain was diluted in sterile water and serially diluted on Streptomyces isolation medium (4). The five selected strains had mycelial and sporulating phenotypes. After isolation and purification, spores were inoculated in Hickey-Tresner broth medium (30°C for 30 h), from which genomic DNA was extracted and purified (5). This procedure included a proteinase K treatment and a chloroform extraction step in order to remove the covalently bound terminal protein (6). An assembly was generated from Oxford Nanopore single reads using Canu (v1.7.1). Sequencing was carried out using the sequencing kits SQK-LSK109 and EXP-NBD104, the flow cell FLO-MIN106 (R9.4.1 revD) on the GridION system (v3.1.8), and the base-calling program Guppy (v2.0.8). When strains were multiplexed, Porechop (v0.2.4) was used for demultiplexing (and adaptor trimming). The coverage ranged from  $73\times$  to  $322\times$ . One large contig covering the whole genome of each strain was obtained (except for RPA4-5, for which two contigs were obtained). A circular extrachromosomal element of about 67 kb was predicted for strain RLA2-12. Illumina paired-end libraries (35 to 51 bp) were created from genomic DNA fragments using the NextSeq 500/550 high-output kit v2 (75 cycles) with the Westburg next-generation sequencing (NGS) DNA library preparation kit (Illumina TruSeq LT primers) (Table 1). Paired-end reads were generated using an Illumina NextSeq instrument (vNB552053). The minimum read size was set to 10 bp, and adaptor trimming was performed using Cutadapt (v1.15). The coverage of the paired-end reads varied between  $50\times$  and  $90\times$ . The final genome sequences were obtained by aligning the Illumina sequences to the Nanopore reference contigs using Pilon (v1.22) (7). Default parameters were used for all software. The total genome sizes, including extrachromosomal DNA when present, ranged from 9.04 Mb to 12.27 Mb, positioning the latter among the largest bacterial sequenced genomes (8) (Table 1). Although public annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline (9), we performed an annotation step using the RAST tool kit (RASTtk) (10, 11) and antiSMASH (12, 13) to predict the specialized metabolism repertoire. This showed that the sample had a large potential for synthesis, with 157 gene clusters. The strain RLA2-12 belonged

Citation Nicault M, Tidjani A-R, Gelhaye E, Bontemps C, Leblond P. 2020. Genome sequences of five *Streptomyces* strains isolated at microscale. Microbiol Resour Announc 9:e00428-20. https://doi.org/10.1128/MRA .00428-20.

Editor David A. Baltrus, University of Arizona
Copyright © 2020 Nicault et al. This is an
open-access article distributed under the terms
of the Creative Commons Attribution 4.0
International license.

Address correspondence to Pierre Leblond, pierre.leblond@univ-lorraine.fr.

Received 28 April 2020 Accepted 18 May 2020 Published 4 June 2020 Nicault et al. 

♣ Microbiology

TABLE 1 Genome features, sequencing statistics, and accession numbers for the five Streptomyces isolates

|  | Data for Illumina sequencing <sup>a</sup> |                   | Data for Oxford Nanopore sequencing <sup>b</sup> |                   |                        |                     |                    | GC             |                       |
|--|---|-------------------|--|-------------------|------------------------|---------------------|--------------------|----------------|-----------------------|
| Strain<br>(replicon)                               | No. of reads (coverage [×])               | SRA accession no. | No. of reads (coverage [×])                      | SRA accession no. | Replicon size (bp)     | Genome<br>size (bp) | Total no. of CDSsc | content<br>(%) | GenBank accession no. |
| RLA2-12<br>(pRLA2-12.1 <sup>c</sup> ) <sup>d</sup> | 21,079,784 (83)                           | SRX8039999        | 269,118 (175)                                    | SRX8039129        | 10,825,588<br>(67,358) | 10,892,946          | 10,116             | 70.3           | JABAQG000000000       |
| RLB1-33  | 14,156,938 (50)                           | SRX8040000        | 477,335 (222)                                    | SRX8039130        | 12,127,650             | 12,127,650          | 11,381             | 70.0           | CP050974              |
| RPA4-2   | 19,808,024 (86)                           | SRX8040001        | 231,331 (165)                                    | SRX8039131        | 9,856,149              | 9,856,149           | 9,287              | 70.9           | CP050975              |
| RPA4-5   | 19,657,766 (93)                           | SRX8040002        | 97,134 (73)                                      | SRX8039132        | 9,047,156              | 9,047,156           | 9,260              | 70.9           | CP050976              |
| S1D4-11  | 15,509,220 (53)                           | SRX8040003        | 180,498 (107)                                    | SRX8039133        | 12,276,515             | 12,276,515          | 12,065             | 69.9           | CP051010              |

<sup>&</sup>lt;sup>a</sup> Illumina libraries were generated using the universal TruSeq LT adapter 5'-AATGATACGGCGACCACCGAGATCTACACTCTTTCCCTACACGACGCTCTTCCGATCT-3' and the index adapter 5'-GATCGGAAGGCACACGTCTGAACTCCAGTCAC(index)ATCTCGTTATGCCGTCTTCTGCTTG-3'.

to a previously isolated cluster of strains (identical 16S rRNA gene sequences) (8, 14) that was closely related to *Streptomyces olivochromogenes* DSM 40451 (99.9% identity in 16S rRNA gene sequences) (15). The strains RLB1-33, RPA4-2, and S1D4-11 and *Streptomyces mirabilis* CSSP107 exhibited identities of 99.9% to 99.2% in their 16S rRNA genes, indicating that they might belong to the same species or constitute sister species. While these strains belong to the large *Streptomyces* clade II described by McDonald and Currie (16), strain RPA4-5 was closely related to *Streptomyces platensis* ATCC 23948 (identical 16S rRNA genes), which belongs to the third, unstructured "other" species group. These results indicate that closely related strains and distant *Streptomyces* species coexist in the same microhabitat.

**Data availability.** Genome sequences and sequence reads (Illumina and Nanopore technologies) are available at GenBank and at the NCBI Sequence Read Archive (SRA) under the accession numbers listed in Table 1.

## **ACKNOWLEDGMENTS**

This work was funded by the French National Research Agency (ANR LABEX ARBRE grant ANR-11-LABX-0002-01), by the French National Institute for Agricultural Research (INRA), and by Région Lorraine (now called Région Grand Est).

## **REFERENCES**

- Bontemps C, Toussaint M, Revol PV, Hotel L, Jeanbille M, Uroz S, Turpault MP, Blaudez D, Leblond P. 2013. Taxonomic and functional diversity of Streptomyces in a forest soil. FEMS Microbiol Lett 342:157–167. https:// doi.org/10.1111/1574-6968.12126.
- Thibessard A, Leblond P. 2014. Subtelomere plasticity in the bacterium Streptomyces, p 243–258. In Louis EJ, Becker MM (ed), Subtelomeres. Springer, Berlin, Germany.
- Barka EA, Vatsa P, Sanchez L, Gaveau-Vaillant N, Jacquard C, Meier-Kolthoff JP, Klenk H-P, Clément C, Ouhdouch Y, van Wezel GP. 2015. Taxonomy, physiology, and natural products of *Actinobacteria*. Microbiol Mol Biol Rev 80:1–43. https://doi.org/10.1128/MMBR.00019-15.
- D'Costa VM, McGrann KM, Hughes DW, Wright GD. 2006. Sampling the antibiotic resistome. Science 311:374–377. https://doi.org/10.1126/ science.1120800.
- Kieser T, Bibb M, Buttner M, Chater K, Hopwood D. 2000. Practical Streptomyces genetics. John Innes Foundation, Norwich, United Kingdom.
- Lin YS, Kieser HM, Hopwood DA, Chen CW. 1993. The chromosomal DNA of Streptomyces lividans 66 is linear. Mol Microbiol 10:923–933. https:// doi.org/10.1111/j.1365-2958.1993.tb00964.x.
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. https://doi.org/10.1371/journal .pone.0112963.
- 8. Tidjani AR, Lorenzi JN, Toussaint M, van Dijk E, Naquin D, Lespinet O, Bontemps C, Leblond P. 2019. Massive gene flux drives genome diversity

- between sympatric *Streptomyces* conspecifics. mBio 10:e01533-19. https://doi.org/10.1128/mBio.01533-19.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/nar/gkw569.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. https:// doi.org/10.1038/srep08365.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Bruccoleri R, Lee SY, Fischbach MA, Muller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0: a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res 43:W237–W243. https://doi.org/10.1093/nar/gkv437.
- Medema MH, Blin K, Cimermancic P, de Jager V, Zakrzewski P, Fischbach MA, Weber T, Takano E, Breitling R. 2011. antiSMASH: rapid identification, annotation and analysis of secondary metabolite biosynthesis gene

Volume 9 Issue 23 e00428-20 mra.asm.org **2** 

<sup>&</sup>lt;sup>b</sup> Nanopore adaptor sequences are not available and were automatically trimmed during base calling.

<sup>&</sup>lt;sup>c</sup> As determined through RAST automatic annotation. CDSs, coding sequences.

<sup>&</sup>lt;sup>d</sup>C indicates the circular configuration of the replicon, as predicted by the assembling.



- clusters in bacterial and fungal genome sequences. Nucleic Acids Res 39:W339 –W346. https://doi.org/10.1093/nar/gkr466.
- 14. Tidjani AR, Lorenzi JN, Toussaint M, van Dijk E, Naquin D, Lespinet O, Bontemps C, Leblond P. 2019. Genome sequences of 11 conspecific *Streptomyces* sp. Microbiol Resour Announc 8:e00863-19. https://doi.org/10.1128/MRA.00863-19.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.
- McDonald BR, Currie CR. 2017. Lateral gene transfer dynamics in the ancient bacterial genus *Streptomyces*. mBio 8:e00644-17. https://doi.org/ 10.1128/mBio.00644-17.

Volume 9 lssue 23 e00428-20 mra.asm.org **3**