

Defining protective epitopes for COVID-19 vaccination models

To the Editor,

Recent papers in the Journal provide tangible avenues for COVID-19 vaccine production as immunoreactive epitopes are brought to the forefront in these and many other emerging studies.^{1,2} The development of a consistent predictable animal model of COVID-19 infection is evidently also a welcome event for preliminary antiviral and vaccine assessments and surely brings us to another level of progression.³

The hamster model is not new to coronavirology but has the potential to provide a more stable and predictable model of infection in contrast to the murine models.⁴ Pulmonary infection, whether in the context of chemotherapy or vaccine trials, can be easily graded with a histopathological scoring method previously defined in another context and shown to be useful for small experimental animal groups.⁵ The latter has been applied to experimental endeavor with severe acute respiratory syndrome coronavirus (SARS-CoV).⁶

Initial enthusiasm to assess whole virus vaccines prepared in a variety of options have historically been followed by focused work on component vaccines. Regardless of the vaccine format, however, one major concern is that vaccination for some viruses and bacteria can be associated with adverse early recall responses after subsequent infections.^{7,8} Such a phenomenon was also postulated in early human vaccine trials after parenteral vaccination with *Mycoplasma pneumoniae* and respiratory syncytial virus.^{9,10} Hyperaccentuated immune responses after vaccination with SARS-CoV was previously recognized in murine models.^{6,11} Although antibody-dependent enhancement as an explanation of such post-vaccine pathology has been postulated by some for several vaccines, a confirmation of the latter and a workable solution have at times been elusive.¹²⁻¹⁴ Nevertheless, the critical lesson in vaccine assessment in animal models for COVID-19 is that the review of post-vaccine disease and prevention should therefore include an assessment of both the early and late lung in whichever model so adopted.^{6-8,11}

The current yet preliminary understanding of COVID-19 genome and structure offers several candidates for vaccination.^{1,2,15} In any such assessments, the examination of systemic humoral or cell-mediated responses to the vaccine are often sought, and thereafter, their association with vaccination outcomes is determined. One lesser sought method for looking at protective antibody at least at the entry-level is to examine the mucosal immune response postinfection that develops in lactating females.¹⁶ Immunoblotting for secretory Immunoglobulin A (IgA) (rather than IgA generally) with breast milk samples from those previously documented to have had COVID-19 infection has the

potential to identify immunogens as a surrogate to the finding of protective secretory IgA in the respiratory tract. This would not preclude other research that may focus on systemic protection rather than mucosal or on protection simultaneously from both aspects.

CONFLICT OF INTERESTS

The authors declare that there are no conflict of interests.

Nevio Cimolai MD, FRCP(C) 

Children's and Women's Health Centre of British Columbia, Pathology and Laboratory Medicine, The University of British Columbia, Vancouver, British Columbia, Canada

Correspondence

Nevio Cimolai, MD, FRCP(C), Faculty of Medicine, The University of British Columbia, Children's and Women's Health Centre of British Columbia, 4480 Oak Street, Vancouver, BC V6H3V4, Canada.

Email: ncimolai@mail.ubc.ca

ORCID

Nevio Cimolai  <http://orcid.org/0000-0003-2743-0556>

REFERENCES

- Bhattacharya M, Sharma AR, Patra P, et al. Development of epitope-based peptide vaccine against novel coronavirus 2019 (SARS-COV-2): immunoinformatics approach. *J Med Virol*. 2020. <https://doi.org/10.1002/jmv.25736>
- Baruah V, Bose S. Immunoinformatics-aided identification of T cell and B cell epitopes in the surface glycoprotein of 2019-nCoV. *J Med Virol*. 2020;92(5):495-500.
- Chan JF, Zhang AJ, Yuan S, et al. Simulation of the clinical and pathological manifestations of Coronavirus Disease 2019 (COVID-19) in golden Syrian hamster model: implications for disease pathogenesis and transmissibility. *Clin Infect Dis*. 2020. <https://doi.org/10.1093/cid/ciaa325>
- Roberts A, Vogel L, Guarner J, et al. Severe acute respiratory syndrome coronavirus infection of Golden Syrian hamsters. *J Virol*. 2005; 79(1):503-511.
- Cimolai N, Taylor GP, Mah D, Morrison BJ. Definition and application of a histopathological scoring scheme for an animal model of acute *Mycoplasma pneumoniae* pulmonary infection. *Microbiol Immunol*. 1992;36(5):465-478.
- Yasui F, Kai C, Kitabatake M, et al. Prior immunization with severe acute respiratory syndrome (SARS)-associated coronavirus (SARS-CoV) nucleocapsid protein causes severe pneumonia in mice infected with SARS-CoV. *J Immunol*. 2008;181(9):6337-6348.

7. Cimolai N, Mah DG, Taylor GP, Morrison BJ. Bases for the early immune response after rechallenge or component vaccination in an animal model of acute *Mycoplasma pneumoniae* pneumonitis. *Vaccine*. 1995;13(3):305-309.
8. Cimolai N, Cheong AC, Morrison BJ, Taylor GP. *Mycoplasma pneumoniae* re-infection and vaccination: protective oral vaccination and harmful immunoreactivity after re-infection and parenteral immunization. *Vaccine*. 1996;14(15):1479-1483.
9. Smith CB, Friedewald WT, Chanock RM. Inactivated *Mycoplasma pneumoniae* vaccine. *JAMA*. 1967;199(6):353-358.
10. Kim HW, Canchola JG, Brandt CD, et al. Respiratory syncytial virus disease in infants despite prior administration of antigenic inactivated vaccine. *Am J Epidemiol*. 1969;89(4):422-434.
11. Bolles M, Deming D, Long K, et al. A double-inactivated severe acute respiratory syndrome coronavirus vaccine provides incomplete protection in mice and induces increased eosinophilic proinflammatory pulmonary response upon challenge. *J Virol*. 2011;85(23):12201-12215.
12. Yip MS, Leung HL, Li PH, et al. Antibody-dependent enhancement of SARS coronavirus infection and its role in the pathogenesis of SARS. *Hong Kong Med J*. 2016;22(3 suppl 4):25-31.
13. Delgado MF, Coviello S, Monsalvo AC, et al. Lack of antibody affinity maturation due to poor Toll-like receptor stimulation leads to enhanced respiratory syncytial virus disease. *Nat Med*. 2009;15(1):34-41.
14. Kam YW, Kien F, Roberts A, et al. Antibodies against trimeric S glycoprotein protect hamsters against SARS-CoV challenge despite their capacity to mediate FcγRII-dependent entry into B cells *in vitro*. *Vaccine*. 2009;25(4):729-740.
15. Dong S, Sun J, Mao Z, Wang L, Lu YL, Li J. A guideline for homology modelling of the proteins from newly discovered betacoronavirus, 2019 novel coronavirus (2019-nCoV). *J Med Virol*. 2020. <https://doi.org/10.1002/jmv.25768>
16. Macadam S, Cimolai N. Anti-*Mycoplasma pneumoniae* secretory antibody in human breast milk. *Diagn Microbiol Infect Dis*. 2002;43(3):247-250.