ORIGINAL ARTICLE

Examination of the efficacy of small genetic panels in genomic conservation of companion animal populations

Aaron J. Sams¹ | Brett Ford¹ | Adam Gardner¹ | Adam R. Boyko^{1,2}

¹Embark Veterinary, Incorporated, Boston, MA. USA

²Department of Biomedical Sciences, College of Veterinary Medicine, Cornell University, Ithaca, NY, USA

Correspondence Aaron J. Sams, Embark Veterinary, Incorporated, Boston, MA 02111, USA.

Email: asams@embarkvet.com

Funding information Embark

Abstract

In many ways, dogs are an ideal model for the study of genetic erosion and population recovery, problems of major concern in the field of conservation genetics. Genetic diversity in many dog breeds has been declining systematically since the beginning of the 1800s, when modern breeding practices came into fashion. As such, inbreeding in domestic dog breeds is substantial and widespread and has led to an increase in recessive deleterious mutations of high effect as well as general inbreeding depression. Pedigrees can in theory be used to guide breeding decisions, though are often incomplete and do not reflect the full history of inbreeding. Small microsatellite panels are also used in some cases to choose mating pairs to produce litters with low levels of inbreeding. However, the long-term impact of such practices has not been thoroughly evaluated. Here, we use forward simulation on a model of the dog genome to examine the impact of using limited marker panels to guide pairwise mating decisions on genome-wide population-level genetic diversity. Our results suggest that in unmanaged populations, where breeding decisions are made at the pairwise-rather than population-level, such panels can lead to accelerated loss of genetic diversity at genome regions unlinked to panel markers, compared to random mating. These results demonstrate the importance of genome-wide genetic panels for managing and conserving genetic diversity in dogs and other companion animals.

KEYWORDS

companion animal, conservation genetics, dogs, microsatellites, simulation

1 | INTRODUCTION

Loss of genetic diversity in small populations is a major concern of the global conservation community because it can reduce fitness and adaptability and can ultimately lead to breed, population, or species extinction (O'Grady et al., 2006). For example, Target 13 of the Convention on Biological Diversity's Strategic Plan (https:// www.cbd.int/sp/) is aimed at "minimizing genetic erosion" of both socio-economically and culturally valuable species and "safeguarding their genetic diversity."

It has been repeatedly demonstrated that the optimal way to reduce genetic erosion in management programs is to minimize the mean kinship within a population (Ballou & Lacy, 1995; Fernández, Toro, & Caballero, 2001; Sonesson & Meuwissen, 2001). Kinship can be estimated using pedigrees; however, pedigrees are frequently inaccurate, incomplete, or missing (Cassell, Adamec, &

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2020 The Authors. Evolutionary Applications published by John Wiley & Sons Ltd

ILEY Evolutionary Applications

Pearson, 2003). In lieu of deep, high-quality pedigrees, molecular data are used. While there has been a push in recent years to move toward monitoring and managing populations of conservation concern with genomic data in the form of whole-genome genotyping or whole-genome sequencing (Flanagan, Forester, Latch, Aitken, & Hoban, 2017; Ivy, Putnam, Navarro, Gurr, & Ryder, 2016; Leroy et al., 2017; Shafer et al., 2015), many populations are still monitored with small sets of neutral markers, typically tens to hundreds of microsatellites (Abdul-Muneer, 2014; Attard et al., 2016; Kaczmarczyk, 2016; Kirk & Freeland, 2011; Song et al., 2018; Pedersen, Pooch, & Liu, 2016; Toro, Fernández, & Caballero, 2009).

While many researchers have cautioned against using small marker panels alone to guide captive breeding (see for example Toro, Silió, Rodrigáñez, & Rodriguez, 1998; Wang & Hill, 2000)) at least some previous research has suggested that small microsatellite panels can be used to maintain genetic diversity in captive breeding programs (Kaczmarczyk, 2016). Importantly though, as Nicholas, Mellersh, and Lewis (2018) recently pointed out, such small microsatellite panels do not effectively survey genome-wide genetic diversity. Rather, they survey genetic diversity at and near (depending on the extent of linkage disequilibrium) the assayed microsatellites. Direct comparison of microsatellite panel and genome-wide (e.g. single nucleotide polymorphisms [SNPs]) panel estimates of genetic diversity is rare, but the argument of Nicholas and colleagues is supported by a recent direct comparison in Arabidopsis halleri, which found that microsatellite-based estimates of genetic diversity and population differentiation differ substantially from unbiased estimates from SNPs (Fischer et al., 2017).

In many ways, dogs are an ideal model for the study of genetic erosion and population recovery. Genetic diversity in many common domestic dog breeds has been declining systematically since the beginning of the 1800s, when modern breeding practices came into fashion (Jansson & Laikre, 2018). As such, inbreeding in domestic dog breeds is substantial and widespread (Freedman et al., 2014; Kettunen, Daverdin, Helfjord, & Berg, 2017; Pedersen et al., 2016; Sams & Boyko, 2018) and has led to an increase in recessive deleterious mutations of high effect (Jagannathan et al., 2019; Marsden et al., 2016) as well as general inbreeding depression (Chu et al., 2019).

Dog breeders and breed clubs are increasingly aware of the serious consequences of diversity loss, and with robust panels of both microsatellite markers and genome-wide SNP arrays widely available commercially, there is great potential for breeders to use genetic testing in ways that ultimately improve (or worsen) genetic diversity. However, a key challenge, at least in the United States, is a lack of population-level management. Rather, individual dog breeders or groups of breeders typically manage small subsets of a breed, often relying on pedigrees or commercially available molecular tests to minimize known genetic health risks and sometimes overall inbreeding in individual litters of dogs. For breeds still managed in such a way, it is critical to long-term breed health and survival to understand the long-term impacts on genome-wide genetic diversity of chosen mating strategies and the molecular tools used to guide those strategies.

As a first step in understanding the impact of such population management, we conducted individual-based forward-time population genetic simulations of linked genetic diversity on a model of the dog genome using SLiM 3 (Haller & Messer, 2019). We apply a range of human-directed mate choice models to ask how well different mate choice schemes applied to a restricted panel of 33 "microsatellite" locations in our model dog genome (referred to throughout as MS33) affect genetic diversity genome-wide. More specifically, we evaluate several combinations of metrics calculated on this set of 33 multi-allelic markers to guide diversity-based mate choice including heterozygosity, internal relatedness (IR; Amos et al., 2001), and average genetic relatedness (AGR; Wang, 2002). Importantly, we do not model the generally recommended strategy of selecting parents to minimize population-level kinship. This optimal strategy is ideal for populations that are small and managed by humans. Dog breeds, however, at least in the United States, are not managed as a whole by any single entity. Rather, mating decisions are most often made by individual breeders and dog owners. Successful strategies for preserving genetic diversity in individual dog breeds will need to take this into account. Therefore, as a first step, we simulate individual mating decisions aimed at optimizing genetic measurements for single offspring. Using these methods, we ask how mate choice impacts genome-wide genetic variation as reflected by heterozygosity and allelic richness (average number of alleles per locus) compared to random mate choice, as well as mate choice guided by two-generation pedigree awareness, genome-wide heterozygosity, and relatedness calculated from genome-wide identity by descent (see Section 2).

2 | MATERIALS AND METHODS

2.1 | Genome model

We implemented all simulations in SLiM (v3.3) (Haller & Messer, 2019), and all simulations were run in parallel using Amazon Web Services EC2, SQS, and auto-scaling services. The genome model in our simulations is a rough approximation of the canine genome. For computational efficiency, we model genetic variation as nonrecombining 0.5 megabase (Mb) multi-allelic haplotype blocks. These haplotype blocks are represented by mutation type 1 (m1) in the simulation template (Appendix S1). Chromosomes are created by dividing the genome into 38 sets of 120 haplotype blocks, approximating a genome size of 2.28 gigabases. The average recombination rate in dogs is approximately 1 cM/Mb (probability of crossover per base pair of 10⁻⁸) and is likely more uniform than in humans (Auton et al., 2013; Axelsson, Webster, Ratnakumar, Ponting, & Lindblad-Toh, 2012). Therefore, we model the recombination rate between chromosomes as 0.5 and within

chromosomes as 0.005 (500,000 bp \times 10⁻⁸). Additionally, we modeled 33 microsatellite loci (m2 in the simulation template) spaced across the first 25 chromosomes as such: 3 on chromosome 1, 2 each on chrs 2 – 7, 1 each on chrs 8 – 25, and with no microsatellites on chrs 26–38. This distribution of markers across chromosomes is similar to the 33 STR panel used by Pedersen et al. (2016). Given a number of unique haplotype blocks and microsatellite alleles at the start of the simulation burn-in, we evenly distributed those alleles across individuals in the founding population (see Figure B1, Appendix S2 for a graphical example of this model).

2.2 | Demographic model

We created a relatively simple demographic model in which a single ancestral population evolves for a burn-in and drift period (200 generations) to allow founding genetic diversity to recombine sufficiently and experience sufficient genetic drift. This is followed by a short immediate bottleneck of five generations. Finally, the population expands and goes through 40 mate choice generations. Population genetic data are collected at the beginning of the first generation of mate choice, once every five generations, and again at the very end of the simulations (see Figure B2, Appendix S2 for a graphical example of this model).

2.3 | Life cycle model

Generations in the initial burn-in/drift period and bottleneck follow the standard Wright-Fisher model in SLiM. However, during mate choice, we induce a slightly different model. Creation of offspring in the mate choice schema includes (see Figure B3, Appendix S2 for a graphical example of this model):

- Random sampling of a parent from the full population (sample individual that has not been mated > MAXIMUM_NUMBER_ OF_MATINGS times).
- 2. Choose second parent:
 - a. Subsampling a fraction of the remaining population randomly (according to MATING_POOL_SIZE) from which to choose potential mates. This step is intended to model the fact that within an effective population of purebred dogs, individual dogs only have access to a limited number of other dogs as mates. This sampling is repeated if no eligible (has been mated < MAXIMUM_NUMBER_OF_MATINGS times) dogs are sampled.
 - A mate for the first individual is chosen according to a specific mate choice model (see Mate Choice Models) from this pool.
- In the event of a layered mate choice model in which two statistics are used to choose mates, a PROPORTION_OF_MATES_ FOR_LAYERED_MATE_CHOICE parameter is used to subsample the potential mates based on each statistic used.

Mate choice models that we implemented in this study primarily differ in how a second parent is selected. First parents are selected randomly from the entire population as described above.

2.4.1 | Random

Second parent is randomly selected from the sampled mating pool.

2.4.2 | Pedigree

Second parent is the individual with the lowest relatedness as calculated from three-generation pedigrees, randomly sampled in ties. In other words, using the keepPedigrees = True option in SLiM 3, pedigree relatedness between individuals in the current generation can be calculated from pedigrees. This option maintains pedigrees including all current individual's parents and grandparents.

2.4.3 | Heterozygosity models [Microsatellite (MS33-HET) and Genome-wide (GW-HET)]

For these models, we calculate the expected heterozygosity for offspring between the first parent and all individuals in the mating pool as the average pairwise observed homozygosity across all four pairwise combinations of parental genomes (assuming no recombination). We choose as the best mate the individual that would produce offspring with the highest heterozygosity and select randomly among ties.

2.4.4 | Internal relatedness (MS33-IR)

For this model, as above, we calculate mean internal relatedness (Amos et al., 2001) among four possible gametic pairs at each microsatellite locus and then average across all loci and choose the individual that produces the lowest IR value, selecting randomly from ties.

2.4.5 | Average genetic relatedness (MS33-AGR)

Here, we apply a method designed to calculate relatedness between individuals based on small panels of SNPs or microsatellites (Wang, 2002). We based the code in our SLiM template on the implementation of this calculation found in the R package Demerelate (https://github.com/cran/Demerelate) and ran several tests to ensure that our SLiM implementation and the R version produced identical results. We chose as the second parent

WILEY

the individual that produced the lowest AGR value, selecting randomly from ties.

2.4.6 | Whole-genome relatedness (GW-REL)

We calculate whole-genome relatedness as in (Hedrick & Lacy, 2015) and as above, calculated relatedness across all four possible gametic pairs between two individuals. We chose as the second parent the individual least related to the first randomly selected parent and chose among ties randomly.

2.4.7 | Layered mate choice (MS33_IR_AGR)

We additionally investigated a single microsatellite mate choice model combining the IR and AGR statistics. First, a fraction of individuals in the mating pool are chosen based on the IR statistic, and then, an individual is chosen from that sample based on AGR (see Section 2.3 above).

2.5 | Population genetic statistics

2.5.1 | Observed heterozygosity

We calculated per individual as the fraction of all genotypes in an individual that are heterozygous.

2.5.2 | Allelic richness

We calculated allelic richness as the total number of unique alleles at each position. In some outputs (results not presented here but data available), we also calculated richness for alleles >=0.05 frequency (raw data available but not presented here).

2.5.3 | Coefficient of inbreeding

We calculated the coefficient of inbreeding (COI) for individual dogs as the fraction of all mutations of type "m1" that are identical within an individual divided by the total genomic length. Because each mutation of this type represents 0.5 Mb, this method is identical to that presented in (Sams & Boyko, 2018).

2.6 | Statistical analyses

For nearly all statistical comparisons, including calculations of 95% confidence intervals used in Figure 1, we have utilized estimation graphics and statistics (Ho, Tumkaya, Aryal, Choi, & Claridge-Chang, 2019) as implemented in the python package dabest (v0.2.5

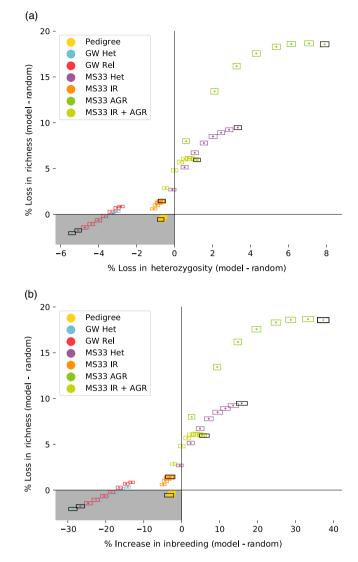


FIGURE 1 Small marker panel mate selection performs worse and genome-wide marker panel mate selection improves over time relative to random mating. (a) Percent loss of Heterozygosity versus percent loss in allelic richness, (b) percent increase in coefficient of inbreeding versus percent loss in allelic richness. Both panels illustrate data from parameter set 1 and represent change relative to the random mate selection model. Dots represent the mean difference between each mate choice model and random mating using a randomization method. Each model includes eight time points, one for every five generations of mate selection, with the final generation indicated with a black box. Height and width of boxes represent 95% confidence intervals for richness and heterozygosity, respectively (see Section 2). Gray box represents the quadrant in which both loss of heterozygosity and richness is slowed relative to random mating. GW models (including Pedigree) improve over the course of simulations, while MS models lose more diversity over time than random mating

 https://acclab.github.io/DABEST-python-docs/index.html). See python script used to generate results for specific uses (https:// github.com/ProjectAussie/embark-research/tree/master/sams_ etal_2020_evolutionary_applications).

3 | RESULTS

3.1 | Genetic variation in simulated populations compared to present-day dogs

For each of eight mate choice models (seven described in Section 2 and an additional compound mate choice model of MS33-IR and MS33-AGR), we generated 100 replicate simulations for each of 21 different demographic parameter sets. Three baseline parameter sets vary in the effective population size used during the burn-in period. Additional simulations are variants of these three baseline parameters, varying one other parameter (Table C1, Appendix S3).

The simulated populations show realistic genetic variation compared to present-day domestic dog breeds. For example, across all models, prior to mate selection the mean coefficient of inbreeding (COI) across all 800 replicates (100 replicates × 8 model types) for parameter set 1 (PS-1) is approximately 0.17 - 0.21 and across all simulations is ~0.12 - 0.38, well within the range of several common dog breeds today (Sams & Boyko, 2018). Similarly, mean internal relatedness (IR) varies in the range of -0.013 - 0.034 across PS-1 and -0.015 - 0.1 across all parameter sets (see Tables C2 and C3, Appendix S3 for COI and IR means by model and parameter set). Pedersen et al. (2016) observed a mean IR value of 0.007 in 102 Bulldogs.

3.2 | MS33-based mate choice models lead to more diversity loss than random mate choice or short pedigrees over time

Performance of all models was measured as the loss of genetic diversity relative to the loss of genetic diversity observed across the random mating model. Of the MS33-based models tested, internal relatedness-based mate choice (MS33-IR) performed best overall after 40 generations of mate choice. This model limited loss of heterozygosity and increase in coefficient of inbreeding comparably or slightly better than random mating across all parameter sets tested (Figure 1; Tables C4 and C5, Appendix S3). However, this model performed worse than random mating at preserving allelic richness across all parameter sets but one (PS-19) at which it was indistinguishable from random mating (Table C6, Appendix S3).

With a few exceptions, all other MS33-based models performed worse than random mating using both heterozygosity, coefficient of inbreeding, and allelic richness as a metric of diversity loss. In these cases, diversity loss is as great or in some cases substantially greater than random mate choice and recent pedigree-based mate choice. Importantly, even in cases where MS models preserve heterozygosity and limit increases in coefficient of inbreeding more than random mating, the trajectory of diversity loss over time in these simulations suggests that given enough time these models would also perform worse overall than random mating (Figure 1; Appendix S2, Figure S1).

Among the MS33-based models, MS33-IR mate choice lost the least amount of genetic diversity, and MS33-AGR mate choice lost

the greatest amount of genetic diversity. In all cases, the accelerated loss of genetic diversity compared to random mating is due to preserving diversity at a small number of loci at the expense of the remainder of the genome. In other words, by avoiding inbreeding with individuals more closely related at a small number of loci scattered throughout the genome, the effective population size at unlinked loci, which are evolving under drift, is further reduced.

As direct evidence of this reduction of effective size at unlinked loci, we observe that genetic diversity at MS loci is preserved well, but is lost more than random mating away from these MS positions (Figure 2a). This pattern is consistent regardless of the MS-based mate selection model we examine and is not observed in genome-wide models described below (Figure 2b). Although the rate of decay of diversity preservation near MS loci should correlate with local variation in recombination rate, the consistent difference between MS and GW models suggests that on average, using sparse microsatellites to manage genetic diversity will be ineffective.

3.3 | Number of repeated matings can moderate "popular parent" effects

We examined whether adjusting the total number of individuals that any single individual can access as potential mates within the population, termed here "mating pool size," as well as the maximum number of times any single individual can contribute to the next generation, termed here "number of repeated matings," has a substantial impact on the magnitude of the loss of diversity in MS-based mate selection models.

We reduced the maximum number of matings per individual per generation from 200 (the population size) to 50 (PS10-12) and 5 (PS13-15). Reducing from 200 to 50 had no significant impact on diversity metrics (Figure 3a; Tables C7–C9, , Appendix S3). However, reducing from 200 to 5 led to a substantial reduction in loss of heterozygosity and allelic richness and lower increase in COI (Figure 3b; Tables C7–C9, Appendix S3).

Additionally, we found that halving mating pool size from 50 to 25 (0.25 to 0.125 of the total population; PS16-18) and from 50 to 10 (PS-19-21), unexpectedly, led to a notable reduction in the amount of genome-wide diversity loss (heterozygosity, richness, or coefficient of inbreeding) in all MS-based mate choice schema (Figure 3c,d; Tables C7-C9, Appendix S3). Upon further inspection, we realized this result is consistent with the mating pool size acting to moderate the number of times the same individual is chosen as a parent in a single generation and that the two parameters discussed here are not as independent as we originally envisioned when designing our simulations. This may be because the "mating pool" is randomly chosen each time a first parent is sampled, such that if the same individual is chosen twice, it will have different mating pools each time. In other words, reducing the number of individuals available as mates for a given individual reduces the number of times that individuals with low mean genetic similarity to the rest of the population will be overrepresented in the next

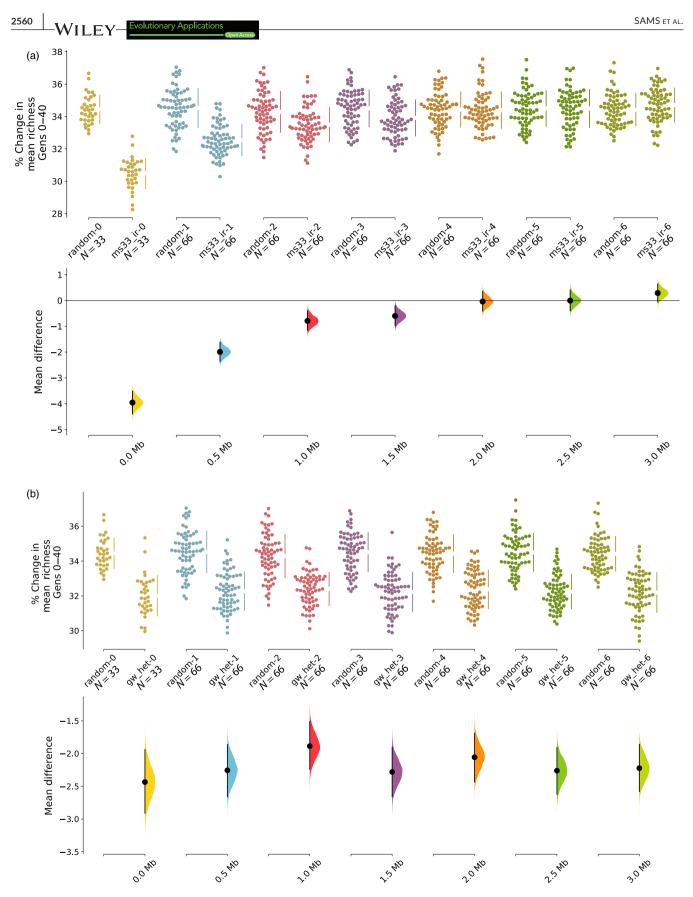


FIGURE 2 Local preservation of allelic richness degrades with distance to MS markers in MS but not GW mate choice models. Difference in percent allelic richness lost during 40 generations of mate choice between random and other models. The upper portion of each panel shows raw data from 100 replicate simulations, while the lower portion of each panel illustrates resampled mean differences between each model and the random mate choice model. The lower portion of each panel also indicates distance in megabases (Mb) from simulated microsatellite markers. (a) In MS models such as MS33-IR, richness is better preserved near markers used during mate choice but degrades with distance. (b) In GW models like GW-HET, richness is consistently preserved across all markers genome wide

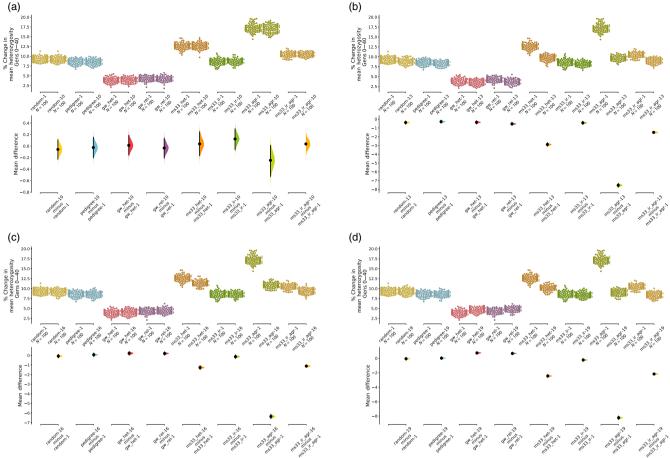


FIGURE 3 Limiting the number of matings per individual improves preservation of heterozygosity in MS models. Change in loss of heterozygosity over 40 mate choice generations. Lower values correspond to better preservation of heterozygosity. The upper portion of each panel shows raw data from 100 replicate simulations, while the lower portion of each panel illustrates resampled mean differences between two parameter sets for each mate choice model. Note the difference in scale of the mean difference statistic in each panel. (a) PS1 compared to PS10 (maximum_number_of_matings 200 to 50). (b) PS1 compared to PS13 (maximum_number_of_matings 200 to 5). (c) PS1 compared to PS16 (mating_pool_size 50 to 25). D. PS1 compared to PS19 (mating_pool_size 50 to 10)

generation. However, while limiting the number of possible mates for each individual decreased the loss of heterozygosity for MSbased mate choice models, it has no impact on the random and pedigree-based mate choice models and increases the loss of heterozygosity slightly in the two genome-wide genetic models (Figure 3c,d), supporting the idea above that higher variance in heterozygosity when marker sample size is low drives the unexpected relationship between preservation of heterozygosity and an individual's mating pool size.

3.4 | Genome-wide metrics improve diversity preservation

In addition to the MS-based mate selection models, we also included several models meant to capture the viability of using genome-wide metrics in general to preserve genetic diversity. These include tracking two-generation pedigrees (pedigree)—to avoid matings between very close relatives, genome-wide heterozygosity (GW-HET)—to select mates which maximize heterozygosity in the offspring, and genome-wide relatedness (GW-REL)—which prefers the most distantly related individuals as mates. We find that these three models all lead to greater preservation of genetic diversity than random mating. Perhaps more importantly, these models reduce the rate of genetic drift over time (random model), compared to the MS models which accelerate the rate of genetic drift (Figure 1).

4 | DISCUSSION

In this study, we used forward population genetic simulations of a model of the canine genome to investigate the efficacy of using a small genetic marker panel (e.g. a microsatellite panel) to guide mating aimed at preserving existing genetic variation in a population, when mating decisions are made at the pairwise, rather than population, level. We ran these simulations across a range of mate choice models and demographic parameterizations using a genomic model that included both genome-wide genetic markers and a set of 33 ILEY— Evolutionary Applications

markers distributed across the genome (herein referred to as the GW and MS33 marker sets, see Section 2).

Most previous work on conservation management with molecular data has focused on cases where a population can be managed by selecting the entire configuration of parents for the next generation with marker-assisted selection (MAS). For example, Fernández, Toro, and Caballero (2004) demonstrated that in a single population, where parental contributions are chosen to maximize either heterozygosity or allelic richness at a set of multi-allelic markers, management programs are optimal at maintaining each of those statistics, but that heterozygosity can be better at maintaining allelic richness than vice versa. However, to our knowledge, no prior work has addressed whether similar strategies that optimize at the level of individual mating pairs, rather than the entire population of parents, can similarly act to preserve diversity. Consistent with this prior work, our results suggest that selecting optimal mates for individuals from an entire population using heterozygosity and other kinship metrics can act to preserve genetic diversity at markers used to calculate the test statistic (for example see Figure 2).

Importantly, however, we also found that given enough generations using small panels of markers in such a mating scheme does not preserve diversity genome-wide. In fact, mate choice models using the MS33 marker set over time led to greater loss of genetic variation compared to random mating in the form of reduced heterozygosity and allelic richness measured using the GW marker set. López-Cortegano et al. (2019) simulated management of subdivided populations and found that using a restricted number of markers was less effective than whole-genome data but still more effective than random mating. However, the density of markers in their simulations is greater than typical microsatellite panels and they acknowledge that less dense panels would likely be less effective. Nonetheless, our results are partially consistent with this result, in that the MS33-IR model does preserve genome-wide genetic diversity better than random mating (but not allelic richness) during the course of our simulations.

Our results suggest that reducing the number of times that any given individual can contribute offspring to the next generation, either explicitly in the form of the "maximum number of matings" parameter or implicitly by reducing the "mating pool size" parameter, can act to moderate the severity of diversity loss compared to the random mating model. This finding is generally consistent with theory and prior simulation work which has demonstrated that optimal management schemes to preserve genetic diversity include limiting variance in family size, in other words, ensuring that no single individual contributes disproportionately to the next generation (Toro et al., 1998).

The better performance of the MS33-IR model compared to other microsatellite-based statistics may be explained by this statistic being less susceptible to such "popular parent" effects. As formulated, when considering between two potential mates, internal relatedness may favor a mate that is more genetically similar if the alleles carried by that individual are on average more common. We suspect that within each generation, this leads to fewer instances of the same outlier individual being repeatedly chosen as the optimal mate for other individuals. Comparing between baseline parameter sets and parameter sets where the maximum number of matings is most restricted (e.g. PS1 vs PS13), we see that this reduced allow-ance of multiple matings has a much weaker impact in MS33-IR mate selection compared to other MS-based mate selection schemes.

Toro, Silió, Rodrigañez, Rodriguez, and Fernández (1999) demonstrated that irrespective of variance in family size, MAS should lead to better preservation of diversity than using no genetic information at all. In contrast, our results suggest that in an unsupervised pairwise parental selection scheme, limited marker panels lead to substantially more diversity loss than using no genetic information at all. A general consensus in MAS of parental populations is that pedigrees should be the primary source of kinship calculations and that small microsatellite panels are generally only useful to supplement pedigrees (Toro et al., 2009). Our results from pairwise parental selection are consistent with this, as we have shown that using only shallow pedigrees to minimize loss of genetic diversity is preferable to using a small panel of genetic markers alone.

Genetic drift comes from two primary sources in a diploid population: variation in genetic contribution between individuals in a population and variation in genetic diversity at a given locus within an individual (Wang & Hill, 2000). Here, we have shown (Figure 2) that the added loss of genetic variation in our simulations relative to random mating is due to accelerated loss of diversity throughout the majority of the genome that is untagged by MS33 markers. While we did not specifically explore the causes of this difference, we suspect that even in our simulations which reduce the contributions of any given individual to the next generation, that groups of individuals which happen to be most distantly related to all other individuals in a given generation across the MS33 marker set are disproportionately chosen as mates for the next generation. This variance in contribution of families (as measured by the MS33 set) across generations will act to consistently reduce effective size at markers unlinked to the MS33 marker set.

Finally, for comparison we simulated several GW mate choice models and found, by and large, that using genome-wide genetic data to monitor genetic diversity and make mate selection decisions is far superior to small marker panels and, typically, random mating. This result has important implications for the preservation of domestic dog breeds. Most academic effort in the field of genetic diversity management over the past few decades has primarily been focused on optimal management for small populations of conservation concern where mating in the entire population can be controlled (Ballou & Lacy, 1995; Fernández, Toro, & Caballero, 2001, 2004; Kettunen et al., 2017; López-Cortegano et al., 2019; Sonesson & Meuwissen, 2001). Similarly, in livestock, conservation of genomic diversity in combination with genomic selection can occur at the level of entire herds or regional populations, although this approach also suffers from geographic partitioning and localization of conservation efforts (Bosse et al., 2015; Bruford et al., 2015; Herrero-Medrano et al., 2014; Ramljak et al., 2018; Zhao et al., 2019). In contrast, dog breeds, as well as breeds in other companion species

olutionary Application

such as cats and horses, have populations which are typically maintained by networks of individual breeders. Therefore, it is very important to understand the long-term impact of different types of breeding practices in these systems.

Here, we have shown that using small panels of molecular markers is no substitute for quality pedigree information or more importantly whole-genome characterization of genetic diversity using dense genetic markers or whole-genome sequence data. For context, assuming each microsatellite in our study can act to maintain diversity at up to 4 Mb each (basing this number on the distance over which diversity preservation decays in Figure 2), only ~5% of the genome (33 × 4 Mb/2.28 Gb) is covered by our simulated microsatellite panel. Our results suggest that optimal management of unsupervised companion animal populations should (a) include strictly limiting individual and family contributions to the next generation and (b) the selection of mating pairs to minimize inbreeding in offspring using deep pedigree information or, more optimally, using dense genotype data to maximize heterozygosity/minimize inbreeding in offspring, as pedigrees are often incomplete and do not incorporate variance in inheritance of IBD segments among related individuals (Cassell et al., 2003; Hill & Weir, 2011; Keller, Visscher, & Goddard, 2011).

We note that we did not directly compare our results to whole-population management schemes, and as such, management strategies are not currently feasible for most companion animals. We suspect that such schemes will be generally superior to the unsupervised mating methods examined here, as they are better able to optimize contributions from individuals and choose the optimal (or near optimal) configuration of pairwise matings to preserve existing genetic diversity.

Most companion animal species remain relatively unmanaged with respect to genetic diversity at the breed level. As such, genetic diversity has rapidly decayed in many breeds over the past century (Jansson & Laikre, 2018). While we have not focused on optimizing the use of whole-genome molecular data to preserve genetic diversity in this study, future species-specific analyses should aim to develop specific recommendations to individual breeders. For example, more realistic (non-Wright-Fisher) models would better reflect the breeding practices used in companion animal breeding. Further, in our whole-genome mate choice methods we have focused primarily on maximizing heterozygosity, but preservation of allelic diversity is also an important metric to optimize, as the number of unique alleles creates a limit on the maximum heterozygosity attainable (Fernández et al., 2004). Finally, we have not considered here the needs of diversity management schemes to also consider balancing the goals of preserving genetic diversity with simultaneously eliminating deleterious variation from a population. In particular, due to the lack of past management, many companion animal breeds carry high effect deleterious mutations, and care must be taken to purge such variation without reducing linked neutral variation (Fernández et al., 2004; Hedrick & Garcia-Dorado, 2016).

Eventually, companion animal breeding may benefit from largescale participation in databases and services aimed at tracking breed-wide whole-genome genetic diversity, including awareness of adaptive and deleterious variation, to limit variance in family contributions, maximize the inclusion of genetic variation in subsequent generations, and purge deleterious variation over time. Experimentation and optimization of such a system applied to breeds in a large and diverse species such as domestic dogs would provide critical case studies to the conservation genetics community (Shafer et al., 2015), help breeders and breed organizations understand the limits of truly closed breeding, and better conserve some of the world's most precious animal resources.

ACKNOWLEDGEMENTS

We thank the customers of Embark, who's participation and curiosity made this work possible. We also thank our colleagues at Embark (in particular Alison Ruhe and Tiago Antao for several helpful discussions), the Doberman Diversity Project, the Embark scientific advisory board, Cornell University, and the Kevin M. McGovern Family Center for Venture Development in the Life Sciences, for their guidance and encouragement. This study was funded by Embark.

DATA AVAILABILITY STATEMENT

Data for this study are available at Dryad: https://doi.org/10.5061/ dryad.k0p2ngf5h. Code to replicate analyses in this study using this simulation data is available on Github: https://github.com/Proje ctAussie/embark-research/tree/master/sams_etal_2020_evolu tionary_applications.

ORCID

Aaron J. Sams 🕩 https://orcid.org/0000-0002-3800-4459

REFERENCES

- Abdul-Muneer, P. M. (2014). Application of microsatellite markers in conservation genetics and fisheries management: Recent advances in population structure analysis and conservation strategies. Genetics Research International, 2014(6), 1–11. https://doi. org/10.1155/2014/691759
- Amos, W., Wilmer, J. W., Fullard, K., Burg, T. M., Croxall, J. P., Bloch, D., & Coulson, T. (2001). The influence of parental relatedness on reproductive success. *Proceedings of the Royal Society B: Biological Sciences*, 268(1480), 2021–2027. https://doi.org/10.1098/rspb.2001.1751
- Attard, C., Möller, L. M., Sasaki, M., Hammer, M. P., Bice, C. M., Brauer, C. J., ... Beheregaray, L. B. (2016). A novel holistic framework for genetic-based captive-breeding and reintroduction programs. *Conservation Biology*, 30(5), 1060–1069. https://doi.org/10.1111/cobi.12699
- Auton, A., Rui Li, Y., Kidd, J., Oliveira, K., Nadel, J., Holloway, J. K., ... Boyko, A. R. (2013). Genetic recombination is targeted towards gene promoter regions in dogs. *PLoS Genetics*, 9(12), e1003984. https:// doi.org/10.1371/journal.pgen.1003984
- Axelsson, E., Webster, M. T., Ratnakumar, A., Ponting, C. P., & Lindblad-Toh, K. (2012). Death of PRDM9 coincides with stabilization of the recombination landscape in the dog genome. *Genome Research*, 22(1), 51–63. https://doi.org/10.1101/gr.124123.111
- Ballou, J. D., & Lacy, R. C. (1995). Identifying genetically important individuals for management of genetic variation in pedigreed populations (pp. 76–111) In J. D. Ballou, M. Gilpin, & T. J. Foose (Eds.). New York, NY: Columbia University Press.
- Bosse, M., Megens, H.-J., Madsen, O., Crooijmans, R. P. M. A., Ryder, O. A., Austerlitz, F., ... de Cara, M. A. R. (2015). Using genome-wide measures of coancestry to maintain diversity and fitness in endangered

and domestic pig populations. *Genome Research*, *25*(7), 970–981. https://doi.org/10.1101/gr.187039.114

- Bruford, M. W., Ginja, C., Hoffmann, I., Joost, S., Orozco-terWengel, P., Alberto, F. J., ... Zhan, X. (2015). Prospects and challenges for the conservation of farm animal genomic resources, 2015-2025. *Frontiers in Genetics*, 6(109), 337-411. https://doi.org/10.3389/ fgene.2015.00314
- Cassell, B. G., Adamec, V., & Pearson, R. E. (2003). Effect of incomplete pedigrees on estimates of inbreeding and inbreeding depression for days to first service and summit milk yield in Holsteins and Jerseys. *Journal of Dairy Science*, 86(9), 2967–2976. https://doi.org/10.3168/ jds.s0022-0302(03)73894-6
- Chu, E. T., Simpson, M. J., Diehl, K., Page, R. L., Sams, A. J., & Boyko, A. R. (2019). Inbreeding depression causes reduced fecundity in Golden Retrievers. *Mammalian Genome*, 30(5-6), 166–172. https:// doi.org/10.1007/s00335-019-09805-4
- Fernández, J., Toro, M. A., & Caballero, A. (2001). Practical implementation of optimal management strategies in conservation programmes: A mate selection method. *Animal Biodiversity and Conservation*, 24(2), 17–24.
- Fernández, J., Toro, M. A., & Caballero, A. (2004). Managing individuals' contributions to maximize the allelic diversity maintained in small, conserved populations. *Conservation Biology*, 18(5), 1358–1367. https://doi.org/10.1111/j.1523-1739.2004.00341.x
- Fischer, M. C., Rellstab, C., Leuzinger, M., Roumet, M., Gugerli, F., Shimizu, K. K., ... Widmer, A. (2017). Estimating genomic diversity and population differentiation – An empirical comparison of microsatellite and SNP variation in *Arabidopsis halleri*. *BMC Genomics*, 18(1), 1–15. https://doi.org/10.1186/s12864-016-3459-7
- Flanagan, S. P., Forester, B. R., Latch, E. K., Aitken, S. N., & Hoban, S. (2017). Guidelines for planning genomic assessment and monitoring of locally adaptive variation to inform species conservation. *Evolutionary Applications*, 11(7), 1035–1052. https://doi.org/10.1111/eva.12569
- Freedman, A. H., Gronau, I., Schweizer, R. M., Ortega-Del Vecchyo, D., Han, E., Silva, P. M., ... Novembre, J. (2014). Genome sequencing highlights the dynamic early history of dogs. *PLoS Genetics*, 10(1), e1004016. https://doi.org/10.1371/journal.pgen.1004016
- Haller, B. C., & Messer, P. W. (2019). SLiM 3: Forward genetic simulations beyond the wright-fisher model. *Molecular Biology and Evolution*, 36(3), 632–637. https://doi.org/10.1093/molbev/ msy228
- Hedrick, P. W., & Garcia-Dorado, A. (2016). Understanding inbreeding depression, purging, and genetic rescue. *Trends in Ecology & Evolution*, 31(12), 940–952. https://doi.org/10.1016/j.tree.2016.09.005
- Hedrick, P. W., & Lacy, R. C. (2015). Measuring relatedness between inbred individuals. *The Journal of Heredity*, 106(1), 20–25. https://doi. org/10.1093/jhered/esu072
- Herrero-Medrano, J., Megens, H.-J., Groenen, M. A., Bosse, M., Perez-Enciso, M., & Crooijmans, R. P. (2014). Whole-genome sequence analysis reveals differences in population management and selection of European low-input pig breeds. *BMC Genomics*, 15(1), 601–612. https://doi.org/10.1186/1471-2164-15-601
- Hill, W. G., & Weir, B. S. (2011). Variation in actual relationship as a consequence of Mendelian sampling and linkage. *Genetics Research*, 93(1), 47–64. https://doi.org/10.1017/s0016672310000480
- Ho, J., Tumkaya, T., Aryal, S., Choi, H., & Claridge-Chang, A. (2019).
 Moving beyond P values: Data analysis with estimation graphics.
 Nature Methods, 16(7), 565–566. https://doi.org/10.1038/s4159
 2-019-0470-3
- Ivy, J. A., Putnam, A. S., Navarro, A. Y., Gurr, J., & Ryder, O. A. (2016). Applying SNP-derived molecular coancestry estimates to captive breeding programs. *Journal of Heredity*, 107(5), 403–412. https://doi. org/10.1093/jhered/esw029
- Jagannathan, V., Drögemüller, C., Leeb, T., Aguirre, G., André, C., Bannasch, D., ... Wade, C. (2019). A comprehensive biomedical

variant catalogue based on whole genome sequences of 582 dogs and eight wolves. *Animal Genetics*, 50(6), 695–704. https://doi. org/10.1111/age.12834

- Jansson, M., & Laikre, L. (2018). Pedigree data indicate rapid inbreeding and loss of genetic diversity within populations of native, traditional dog breeds of conservation concern. *PLoS One*, 13(9), e0202849–e202917. https://doi.org/10.1371/journal.pone.0202849
- Kaczmarczyk, D. (2016). Selection of optimal spawning pairs to maintain genetic variation among captive populations of Acipenseridae based on the polymorphism of microsatellite loci. Archives of Polish Fisheries, 24(2), 77–84. https://doi.org/10.1515/aopf-2016-0009
- Keller, M. C., Visscher, P. M., & Goddard, M. E. (2011). Quantification of inbreeding due to distant ancestors and its detection using dense single nucleotide polymorphism data. *Genetics*, 189(1), 237-249. https://doi.org/10.1534/genetics.111.130922
- Kettunen, A., Daverdin, M., Helfjord, T., & Berg, P. (2017). Cross-breeding is inevitable to conserve the highly inbred population of puffin hunter: The Norwegian Lundehund. *PLoS One*, 12(1), e0170039–e170116. https://doi.org/10.1371/journal.pone.0170039
- Kirk, H., & Freeland, J. R. (2011). Applications and implications of neutral versus non-neutral markers in molecular ecology. *International Journal of Molecular Sciences*, 12(6), 3966–3988. https://doi. org/10.3390/ijms12063966
- Leroy, G., Carroll, E. L., Bruford, M. W., DeWoody, J. A., Strand, A., Waits, L., & Wang, J. (2017). Next-generation metrics for monitoring genetic erosion within populations of conservation concern. *Evolutionary Applications*, 11(7), 1066–1083. https://doi.org/10.1111/eva.12564
- Liu, D., Zhou, Y. U., Yang, K., Zhang, X., Chen, Y., Li, C., ... Song, Z. (2018). Low genetic diversity in broodstocks of endangered Chinese sucker, *Myxocyprinus asiaticus*: Implications for artificial propagation and conservation. *ZooKeys*, 792(2), 117–132. https://doi.org/10.3897/ zookeys.792.23785
- López-Cortegano, E., Pouso, R., Labrador, A., Pérez-Figueroa, A., Fernández, J., & Caballero, A. (2019). Optimal management of genetic diversity in subdivided populations. *Frontiers in Genetics*, 10. https://doi.org/10.3389/fgene.2019.00843
- Marsden, C. D., Ortega-Del Vecchyo, D., O'Brien, D. P., Taylor, J. F., Ramirez, O., Vilà, C., ... Lohmueller, K. E. (2016). Bottlenecks and selective sweeps during domestication have increased deleterious genetic variation in dogs. Proceedings of the National Academy of Sciences of the United States of America, 113(1), 152–157. https://doi. org/10.1073/pnas.1512501113
- Nicholas, F. W., Mellersh, C., & Lewis, T. (2018). Letter to the editor regarding an autosomal recessive mutation in SCL24A4 causing enamel hypoplasia in Samoyed and its relationship to breed-wide genetic diversity. *Canine Genetics and Epidemiology*, 5(1), 1–4. https://doi. org/10.1186/s40575-018-0059-7
- O'Grady, J. J., Brook, B. W., Reed, D. H., Ballou, J. D., Tonkyn, D. W., & Frankham, R. (2006). Realistic levels of inbreeding depression strongly affect extinction risk in wild populations. *Biol. Conserv.*, 133, 42–51.
- Pedersen, N. C., Pooch, A. S., & Liu, H. (2016). A genetic assessment of the English bulldog. *Canine Genetics and Epidemiology*, 3(1), 1–16. https://doi.org/10.1186/s40575-016-0036-y
- Ramljak, J., Bunevski, G., Bytyqi, H., Marković, B., Brka, M., Ivanković, A., ... Medugorac, I. (2018). Conservation of a domestic metapopulation structured into related and partly admixed strains. *Molecular Ecology*, 27(7), 1633–1650. https://doi.org/10.1111/mec.14555
- Sams, A. J., & Boyko, A. R. (2018). Fine-scale resolution of runs of homozygosity reveal patterns of inbreeding and substantial overlap with recessive disease genotypes in domestic dogs. G3: Genes|Genomes|Genetics, 9(1), 117–123. https://doi.org/10.1534/g3.118.200836
- Shafer, A. B. A., Wolf, J. B. W., Alves, P. C., Bergström, L., Bruford, M. W., Brännström, I., ... Zieliński, P. (2015). Genomics and the challenging translation into conservation practice. *Trends in Ecology & Evolution*, 30(2), 78–87. https://doi.org/10.1016/j.tree.2014.11.009

- Sonesson, A. K., & Meuwissen, T. H. (2001). Minimization of rate of inbreeding for small populations with overlapping generations. *Genetics Research*, 77(3), 285–292. https://doi.org/10.1017/s0016 672301005079
- Toro, M. A., Fernández, J., & Caballero, A. (2009). Molecular characterization of breeds and its use in conservation. *Livestock Science*, 120, 174–195. https://doi.org/10.1016/j.livsci.2008.07.003
- Toro, M., Silió, L., Rodrigáñez, J., & Rodriguez, C. (1998). The use of molecular markers in conservation programmes of live animals. *Genetics Selection Evolution*, 30(6), 585–617. https://doi. org/10.1186/1297-9686-30-6-585
- Toro, M., Silió, L., Rodrigañez, J., Rodriguez, C., & Fernández, J. (1999). Optimal use of genetic markers in conservation programmes. *Genetics Selection Evolution*, 31(3), 255. https://doi. org/10.1186/1297-9686-31-3-255
- Wang, J. (2002). An estimator for pairwise relatedness using molecular markers. Genetics, 160(3), 1203–1215.
- Wang, J., & Hill, W. G. (2000). Marker-assisted selection to increase effective population size by reducing Mendelian segregation variance. *Genetics*, 154(1), 475–489.

Zhao, Q.-B., Sun, H., Zhang, Z., Xu, Z., Olasege, B. S., Ma, P.-P., ... Pan, Y.-C. (2019). Exploring the structure of haplotype blocks and genetic diversity in chinese indigenous pig populations for conservation purpose. *Evolutionary Bioinformatics Online*, 15, 1176934318825082. https://doi.org/10.1177/1176934318825082

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Sams AJ, Ford B, Gardner A, Boyko AR. Examination of the efficacy of small genetic panels in genomic conservation of companion animal populations. *Evol Appl*. 2020;13:2555–2565. https://doi.org/10.1111/eva.13038