

CASE REPORT

Storming the castle: A case report of multi-system dysregulation in a child with Castleman disease

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Abstract

Castleman disease is a non-clonal, lymphoproliferative disorder rarely seen in children. Presented is a 12-year-old male with progressive abdominal pain, vomiting, and fever. Diagnostic testing revealed multi-organ system involvement and the diagnosis was ultimately made with tissue biopsy. Marked disease regression occurred after high-dose steroids and continued interleukin-6 inhibition.

KEYWORDS

angiofollicular, Castleman, hyperplasia, interleukin-6, lymphoproliferative

1 | INTRODUCTION

Angiofollicular lymph node hyperplasia, known as Castleman disease (CD), is a group of histopathologically similar, non-clonal, lymphoproliferative disorders. CD is sub-typed as unicentric (UCD) or multicentric (MCD), depending on the number of lymph node regions involved and presence of systemic inflammatory symptoms. In MCD patients, testing for HHV-8 (a known association) needs to be performed and if negative, disease can be further classified as POEMs-associated MCD, TAFRO syndrome, or idiopathic MCD/not otherwise specified (iMCD/MCD-NOS).^{1,2} Given that CD largely

affects the adult population, little is known regarding the clinical course in children.^{2,3} With informed consent obtained and documented, we present a pediatric case of Castleman disease.

2 | CASE DESCRIPTION

A 12-year-old male presented with three weeks of persistent abdominal pain, emesis, fatigue, and fever. At arrival, he was tachycardic, tachypneic, febrile, dehydrated, pale, and ill-appearing. Physical examination showed obesity, anasarca, altered mental status, global

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abdominal tenderness, and distention, but clear lung fields and no cardiac murmur. Initial laboratory studies identified anemia (7.1 g/dL), thrombocytopenia ($20 \times 10^3/\mu\text{L}$), neutrophilia (83%) without leukocytosis ($11.3 \times 10^3/\mu\text{L}$), hypocalcemia (5.8 mg/dL), elevated serum creatinine (1.52 mg/dL), and elevated inflammatory markers (erythrocyte sedimentation rate (ESR) 75 mm/hr, c-reactive protein (CRP) 31 mg/L). Abdominal computed tomography (CT) showed a 3.5 cm left adrenal mass, hepatosplenomegaly, colonic wall thickening, solitary lung nodule, and small pericardial effusion.

Empiric antibiotics were initiated but testing for bacterial, viral, and fungal infections, including HHV-8, remained negative. Laboratory results showed severe hypothyroidism with undetectable free thyroxine and negative thyroid antibody panel, arguing against a diagnosis of central hypothyroidism. Parathyroid hormone was elevated (79 pg/dL, RR 10–65 pg/dL), precluding hypoparathyroidism as the cause for hypocalcemia. He required titration of levothyroxine and aggressive supplementation with calcium gluconate injections, calcium carbonate, and calcitriol for refractory hypocalcemia.

Delirium improved with control of fever and correction of hypocalcemia and hypothyroidism. Throughout his admission, he developed hypertension and a pericardial effusion. Multiple antihypertensive medications (enalapril, labetalol, nifedipine, furosemide, and isradipine) were required to control hypertension. A renal biopsy was performed to evaluate proteinuria and hematuria but showed no significant abnormalities. Additional laboratory testing identified hyperuricemia (15 mg/dL, RR 2–7 mg/dL) and elevated lactate dehydrogenase (1437 U/L, RR 550–900 U/L). Hyperuricemia resolved after rasburicase and allopurinol treatment. Abdominal magnetic resonance imaging (MRI) showed a cystic, hemorrhagic adrenal lesion (Figure 1).

In addition to persistent thrombocytopenia and microcytic, hypochromic anemia, he also had prolonged prothrombin time (17.1 seconds) with normal activated partial thromboplastin time (31 seconds) and elevations in fibrinogen (864 mg/dL, RR 170–410 mg/dL), d-dimer (14.3 $\mu\text{g}/\text{mL}$, RR $<0.5 \mu\text{g}/\text{mL}$), and immature platelet fraction (16%, RR 1.1–8.5%). Peripheral smear showed spherocytes, polychromasia, schistocytes, and normal neutrophils. Peripheral blood flow cytometry showed no signs

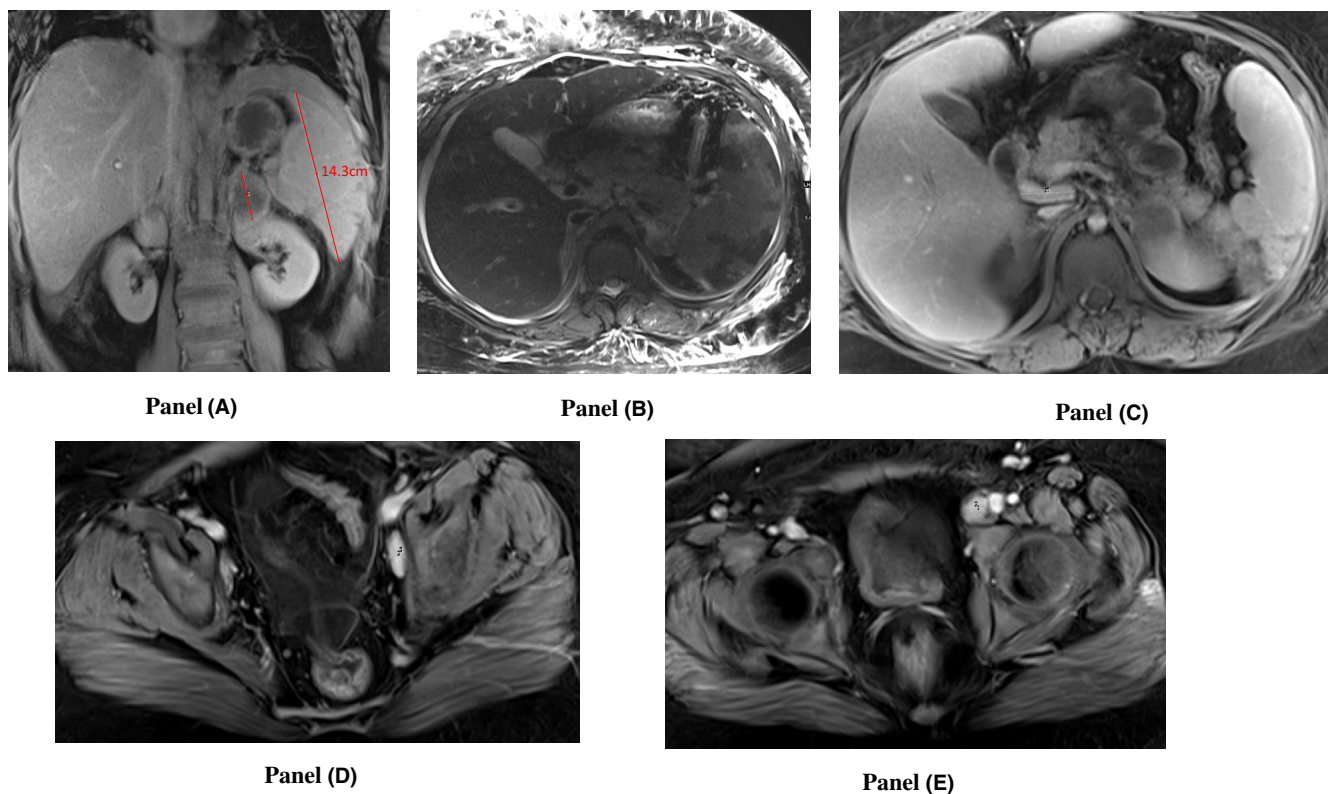


FIGURE 1 Initial abdominal and pelvic MRI. (A) Splenomegaly present with craniocaudal length of 14.3 cm and left adrenal gland with presence of a circumscribed lesion without additionally discerning features. (B) patchy confluent and geographic peripheral segmental T2 dark hypo-enhancing regions typical for splenic infarcts without discrete splenic mass. (C) Mildly enlarged lymph nodes present throughout abdomen most notably in the pelvis porta hepatis. (D) Left pelvic sidewall lymph node. (E) Left external iliac chain lymph node. Radiographic interpretation credit: Dr. Adam Bobby

of myeloid neoplasm or lymphoma. Bone marrow biopsy demonstrated a hypercellular marrow (80–90%) with mild erythroid and megakaryocytic hyperplasia. These findings were largely attributed to reactive and regenerative responses to the anemia and thrombocytopenia but with slight concern for hemolytic anemia. ADAMTS13 activity was decreased (52% then 29%, RR >68%) with a negative ADAMTS13 antibody inhibitory titer (<0.5), inconsistent with thrombotic thrombocytopenic purpura. Blood product transfusions were given as needed for support.

Investigation for autoimmune disease identified a low positive antinuclear antibody (1:40), mildly positive anti-cardiolipin immunoglobulin M antibody, and mild hypocomplementemia, with negative testing for anti-neutrophil cytoplasmic antibody (ANCA) and cryoglobulins. Testing was also negative for antibodies to myeloperoxidase, proteinase 3, crithidia, Sjögren's syndrome-related antigen A and B, Smith, and ribonuclear protein. Notable elevations of soluble interleukin-2 (IL-2) receptor assay/soluble CD25 (2,029 U/mL, RR 137–838 U/mL), soluble IL-2 receptor-alpha (6500 pg/mL, RR 622–1619 pg/mL), ferritin (560 ng/mL, RR 7–142 ng/mL), and vascular endothelial growth factor (98 pg/mL, RR 9–86 pg/mL) were discovered with a normal serum interleukin-6 (IL-6, RR <5 pg/mL). Given these elevations in conjunction with evidence of fever, splenomegaly, pancytopenia, and hypertriglyceridemia (204 mg/dL, RR 60–134mg/dL, although the patient was notably obese), investigation for hemophagocytic lymphohistiocytosis (HLH) was warranted. However, no hemophagocytosis was appreciated on the bone marrow biopsy and none of the genetic variants associated with HLH were present on the primary immunodeficiency panel. Furthermore, on immunophenotype lymphocyte subset panel, presence of natural killer cells CD56/CD16 cells was normal at 160, 4.2% (RR: 70–480, 3–22%), whereas in HLH, there is low or absent natural killer cell activity.⁴

An inguinal lymph node biopsy, obtained to evaluate for lymphoproliferative disease, showed atrophic germinal centers with increased numbers of follicles ranging from hyperplastic to regressed. The follicles were surrounded by prominent, thickened mantle zones containing concentrically arranged small lymphocytes, consistent with CD (Figure 2). Positron emission tomography-computed tomography (PET-CT) demonstrated generalized lymphadenopathy and bone marrow uptake without a discrete focal suspicious lesion, consistent with reactive change or mild marrow hyperplasia in the setting of CD (Figure 3). Diffuse, mild uptake was seen in a mildly enlarged spleen, thought to be reactive to non-specific cytokine release (Figure 3).

Treatment included initiation of prednisone (60mg/day) with a prolonged taper and tocilizumab infusions.

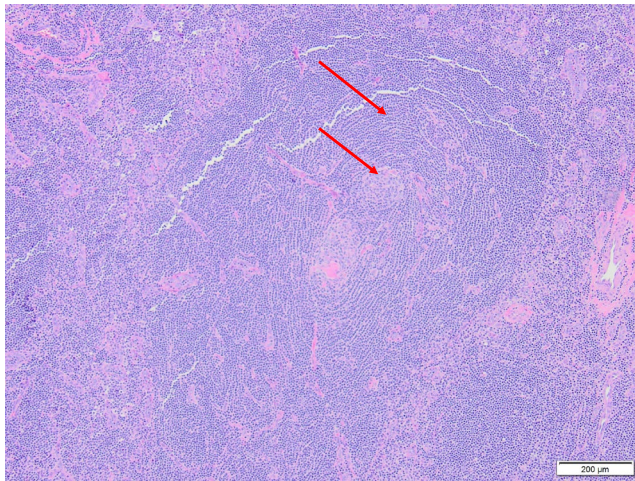
His clinical status and laboratory findings improved significantly and rapidly after initiation of treatment and PET-CT six months after initial presentation demonstrated marked improvement. The initially concerning left adrenal lesion appeared consistent with a resolving hemorrhage. Disease burden remains low on continued monthly tocilizumab infusions with stable cell lines and inflammatory markers showing consistent downtrend and evidence of stabilization. Of note, inflammatory markers did show slight increase (CRP, ESR) after trialing spacing of treatments to 5–6 weeks but returned to within normal limits (WNL) once treatments were resumed every 4 weeks. He has not had any recurrence of clinical symptoms since therapy was initiated. He remains on levothyroxine, labetalol, enalapril, and prophylactic dosages of acyclovir and trimethoprim-sulfamethoxazole while receiving IL-6 inhibition therapy.

3 | DISCUSSION

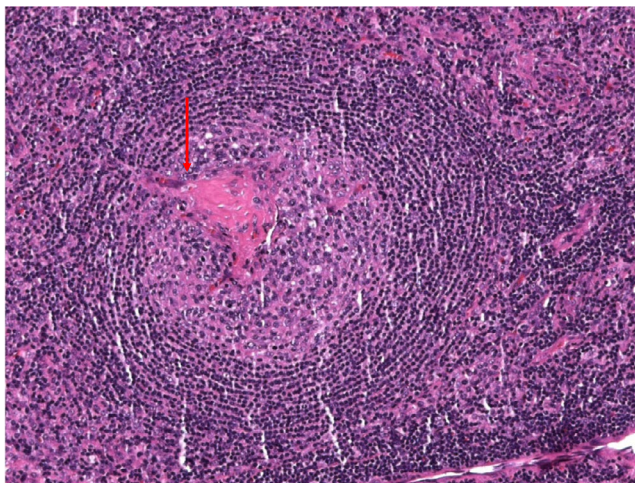
The diagnosis of CD is both clinical and pathologic, often requiring extensive evaluation.^{1–3} Per the international, evidence-based consensus diagnostic criteria for MCD, there must be multicentric lymphadenopathy with defined histopathology, two or more clinical or laboratory changes, and exclusion of MCD mimics, as delineated in Table 1.⁵ The patient presented had negative HHV-8 and HIV testing and meets both major diagnostic criteria and eight of the eleven minor diagnostic criteria for a diagnosis of MCD. Using the Japanese diagnostic criteria, this patient also meets criteria for iMCD-TAFRO syndrome given his lymph node pathology (Figure 2), negative HHV-8 testing, and clinical evidence of thrombocytopenia, anasarca, fever, reticulin fibrosis, organomegaly, absence of hypergammaglobulinemia, small volume lymphadenopathy, and hyperplasia of megakaryocytes in the bone marrow.² Serum transaminases and alkaline phosphatase values were also WNL, and immunoglobulins were as follows: IgA 142 (WNL), IgE 99 (WNL), IgG 783 (WNL), IgD 2.2 (WNL), IgM 29 (low), all consistent with this diagnosis.²

Patients with MCD typically have profound systemic inflammatory disease, due to IL-6 predominant cytokine storm and resultant multi-organ dysfunction.^{3,5,6} The underlying trigger for the cytokine storm remains unknown. One hypothesis of pathogenesis involves an auto-antibody mediated IL-6 pathway in which auto-antibodies trigger self-perpetuating, pro-inflammatory cytokine release and dysregulate signaling in the antigen-presenting cells.

The CDCN International Consensus Guidelines for treatment of iMCD were consulted to stratify our patient based on disease severity and choose the best therapeutic



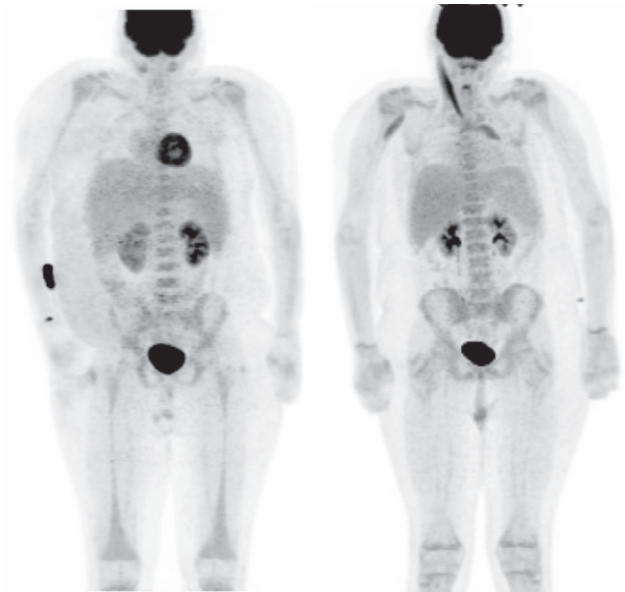
Panel (A)



Panel (B)

FIGURE 2 Histopathology of inguinal lymph node biopsy (H&E stain). (A) Small, atrophic germinal centers with increased numbers of follicles that vary in size from hyperplastic to regressed. The follicles are surrounded by prominent and widened mantle zones containing small lymphocytes arranged in a concentric fashion giving an “onion-skin appearance” of the mantle zone around the germinal centers. (B) A lymphoid follicle with a hyalinized atrophic germinal center, a feeder blood vessel, and an onion skin-shaped arrangement of mantle zone lymphocytes. These changes are characteristic of the hyaline vascular variant of Castleman disease. (Photographs courtesy of Samir Kahwash, M.D.)

option.⁶ Treatment of CD is aimed at cytokine blockade, including high-dose steroids and IL-6 inhibition.⁶⁻⁸ Tocilizumab, a recombinant humanized monoclonal antibody directed against the IL-6 receptor, and siltuximab, a monoclonal antibody that binds IL-6, are two of the treatments used most frequently⁹. Tocilizumab was used in this case due to equal effectiveness of both drugs and its availability on the formulary of our hospital. The IL-6 inhibitors can be efficacious even in the absence of elevated serum IL-6, as in this case. Additionally, current



Panel (A)

Panel (B)

FIGURE 3 Initial and follow-up nuclear medicine positron emission computed tomography. (A) Evidence of moderate/intense uptake involving left external iliac, left pelvic sidewall, subcarinal, and right hilar lymph nodes. Mild generalized uptake of the bone marrow without discrete focal suspicious lesion. Splenomegaly present with diffuse mild increased uptake and asymmetric right renal function noted. Resolved increased uptake in the right hilar, subcarinal lymph nodes with mild uptake in the nonenlarged left pelvic sidewall and left external iliac lymph nodes. Increased uptake within the thymus consistent with rebound thymic hyperplasia. (B) Left adrenal lesion smaller compared to the prior, consistent with resolving hematoma, and resolved previously identified asymmetry of renal uptake. Radiographic interpretation credit: Dr. Adam Bobbey and Dr. Ellen Chung

testing modalities may fail to recognize complexed IL-6 and provide falsely low IL-6 values on many commercial tests.⁹ As such, normal IL-6 levels should not preclude anti-IL-6 therapy.^{6,7} Further, novel therapies targeting upstream and downstream pathways such as Janus kinase/signal transducer activator of transcription 3 (JAK/STAT3), mitogen-activated protein kinase (MAPK), and phosphatidylinositol-3-kinase (PI3K)/mammalian target of rapamycin (mTOR) are currently under investigation and may have benefit.⁵

Treatment response ranges from complete remission to refractory, progressive disease.^{10,11} Due to potential for repeated, dangerous flares, therapy is indefinite and clinical, laboratory, and radiographic indices must be closely trended, particularly if weaning immunosuppression.^{5,10} The necessary duration of immunosuppressive or immunomodulatory therapy to prevent disease relapse has not yet been defined.^{8,10,11}

Of note, on the primary immunodeficiency panel obtained for this patient, there was a pathologic variant in

TABLE 1 Major and minor diagnostic criteria of Castleman disease**I. Major criteria (need both)**

- Histopathologic lymph node features consistent with the iMCD spectrum (need grade 2–3 for either regressive or plasmacytosis at minimum).
 - Regressed/atrophic/atretic germinal centers, often with expanded mantle zones composed of concentric rings of lymphocytes in an “onion-skinning” appearance
 - Follicular dendritic cell prominence
 - Vascularity, often with prominent endothelium in the interfollicular space and vessels penetrating into the germinal centers with a “lollipop” appearance
 - Sheet-like, polytypic plasmacytosis in the interfollicular space
 - Hyperplastic germinal centers

- Enlarged lymph nodes (≥ 1 cm in short-axis diameter) in ≥ 2 lymph node stations

II. Minor criteria (need at least 2 of 11 criteria with at least 1 laboratory criterion)

1. Laboratory

- Elevated inflammatory markers: c-reactive protein >10 mg/L or erythrocyte sedimentation rate >15 mm/hour
- Anemia: Hemoglobin <12.5 g/dL (males) or <11.5 g/dL (females)
- Abnormal platelet quantity: Platelet count <150 k/microL or >400 k/microL
- Hypoalbuminemia: Albumin <3.5 g/dL
- Renal dysfunction: estimated glomerular filtration rate <60 mL/min/1.73m² or proteinuria (total protein 150 mg/24hrs or 10 mg/100mL)
- Polyclonal hypergammaglobulinemia (IgG >1700 g/dL)

2. Clinical

- Constitutional symptoms: night sweats, fever, weight loss, or fatigue (≥ 2 CTCAE lymphoma score for B-symptoms)
- Hepatomegaly and/or splenomegaly
- Fluid accumulation: edema, anasarca, ascites, or pleural effusion
- Eruptive cherry hemangiomas or violaceous papules
- Lymphocytic interstitial pneumonitis

Note: CTCAE, Common terminology criteria for adverse events. Fajgenbaum DC et al.⁵

the MVK gene which has been associated with autosomal recessive mevalonate kinase deficiency, hyperimmunoglobulinemia D syndrome (HIDS), and periodic fever syndrome.^{12,13} While this patient's clinical presentation is not fully consistent with any of these diagnoses, the overlap between diagnostic clinical features is notable. The latter two syndromes are characterized by recurrent episodes of fever associated with lymphadenopathy, arthralgia,

gastrointestinal dismay, and skin rash.^{12,13} To our knowledge, no previous associations between CD and MVK gene alteration have been reported.

CD remains a diagnostic challenge given the rarity of the condition in the pediatric population and its non-specific clinical, laboratory, and radiographic features impacting multiple organ systems. The importance of awareness of the clinical presentation in a pediatric patient to prevent delayed diagnosis and provide optimal care cannot be understated.

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CONFLICT OF INTEREST

All authors have no conflicts of interest relevant to this article to disclose.

AUTHOR CONTRIBUTIONS

SD served as the lead and corresponding author, contributed to the conception and analysis of the case report, drafted the manuscript, gave final approval for publication, and agreed to be accountable for all aspects of the work. NK and CB both contributed to the conception and analysis of the case report, participated in drafting the manuscript, gave final approval for publication, and agreed to be accountable for all aspects of the work. SPA and SK contributed to the analysis of the case report, revised the manuscript, gave final approval for publication, and agreed to be accountable for all aspects of the work. MR served as the senior author on the project, contributed to the conception and analysis of the case report, revised the manuscript, gave final approval for publication, and agreed to be accountable for all aspects of the work.

ETHICAL APPROVAL

This case report conforms to recognized standards and was conducted in accordance with the Helinski declaration as revised in 2013. This work was deemed exempt from formal review by Nationwide Children's Hospital's Institutional Review Board. Patient anonymity is preserved throughout.

CONSENT

Patient consent has been signed and collected in accordance with the journal's patient consent policy and can be provided upon request.

DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were created or analyzed in this study.

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REFERENCES

1. Farruggia P, Trizzino A, Scibetta N, et al. Castleman's disease in childhood: report of three cases and review of the literature. *Italian Journal of Pediatrics*. 2011;37:50. doi:10.1186/1824-7288-37-50
2. Iwaki N, Fajgenbaum DC, Nabel CS, et al. Clinicopathologic analysis of TAFRO syndrome demonstrates a distinct subtype of HHV-8-negative multicentric Castleman disease. *Am J Hematol*. 2016;91(2):220-226. doi:10.1002/ajh.24242
3. Perez N, Bader-Meunier B, Roy CC, Dommergues JP. Paediatric Castleman disease: report of seven cases and review of the literature. *Eur J Pediatr*. 1999;158(8):631-637. doi:10.1007/s004310051166
4. Jordan MB, Allen CE, Greenberg J, et al. Challenges in the diagnosis of hemophagocytic lymphohistiocytosis: Recommendations from the North American Consortium for Histiocytosis (NACHO). *Pediatr Blood Cancer*. 2019;66(11):e27929. doi:10.1002/pbc.27929
5. Fajgenbaum DC, Uldrick TS, Bagg A, et al. International, evidence-based consensus diagnostic criteria for HHV-8-negative/idiopathic multicentric Castleman disease. *Blood*. 2017;129(12):1646-1657. doi:10.1182/blood-2016-10-746933
6. van Rhee F, Voorhees P, Dispenzieri A, et al. International, evidence-based consensus treatment guidelines for idiopathic multicentric Castleman disease. *Blood*. 2018;132(20):2115-2124. doi:10.1182/blood-2018-07-862334
7. Fajgenbaum DC. Novel insights and therapeutic approaches in idiopathic multicentric Castleman disease. *Blood*. 2018;132(22):2323-2330. doi:10.1182/blood-2018-05-848671
8. Nishimoto N, Kanakura Y, Aozasa K, et al. Humanized anti-interleukin-6 receptor antibody treatment of multicentric Castleman disease. *Blood*. 2005;106(8):2627-2632. doi:10.1182/blood-2004-12-4602
9. Chaturvedi S, Siegel D, Wagner CL, et al. Development and validation of panoptic Meso scale discovery assay to quantify total systemic interleukin-6. *Br J Clin Pharmacol*. 2015;80(4):687-697. doi:10.1111/bcp.12652
10. Yoshizaki K, Murayama S, Ito H, Koga T. The role of interleukin-6 in castleman disease. *Hematol Oncol Clin North Am*. 2018;32(1):23-36. doi:10.1016/j.hoc.2017.09.003
11. Cervantes CE, Correa R. Castleman disease: A rare condition with endocrine manifestations. *Cureus*. 2015;7(11):e380. doi:10.7759/cureus.380
12. Cuisset L, Drenth JPH, Simon A, et al. Molecular analysis of MVK mutations and enzymatic activity in hyper-IgD and periodic fever syndrome. *Eur J Hum Genet*. 2001;9(4):260-266. doi:10.1038/sj.ejhg.5200614
13. D'Osualdo A, Picco P, Caroli F, et al. MVK mutations and associated clinical features in Italian patients affected with auto-inflammatory disorders and recurrent fever. *Eur J Hum Genet*. 2005;13:314-320. doi:10.1038/sj.ejhg.5201323

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