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Research

Gene expression profiling of *IncRNA-HOTAIR* and *IncRNA-MALAT1* in esophageal cancer: uncovering links to lifestyle factors and diagnostic significance

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Abstract

Background Esophageal cancer (EC) is the sixth most common cause of cancer-related deaths globally. Genetic and environmental factors could be affected in EC's onset and development.

The potential involvement of IncRNA-HOTAIR and IncRNA-MALAT1 in EC has garnered significant attention in recent studies. Our investigation aimed to examine IncRNA-HOTAIR and IncRNA-MALAT1 gene expression changes in EC patients. **Materials and methods** Our experimental study focused on 140 patients with malignant EC, comprising 70 paraffinembedded tumor tissues (FFPE) blocks and 70 FFPE blocks with marginal tissue samples. The relative gene expression levels of IncRNA-HOTAIR and IncRNA-MALAT1 were measured using Real-Time PCR. The data were analyzed using ANOVA and $2^{-\Delta\Delta CT}$ tests.

Results Our analysis revealed a significant increase in tumor expression compared to marginal tissues (P < 0.05). Besides, our research revealed a significant correlation between *IncRNA-HOTAIR* expression and hot drinks (P = 0.019), metastasis (P = 0.001), and the 5-year survival rate (P = 0.001). We found a significant correlation between *IncRNA-MALAT1* expression and alcohol abuse (P = 0.039), hot drinks (P = 0.001), and metastasis (P = 0.039).

Conclusion The findings indicate a potential carcinogenic effect of *IncRNA-HOTAIR* and *IncRNA-MALAT1* gene expression alterations in EC patients. Also, studying the IncRNA genes can help us identify biomarkers, emphasizing the significance of early diagnosis and treatment.

Keywords LncRNA · HOTAIR · MALAT1 · Esophageal cancer

1 Introduction

Esophageal cancer (EC) is the eighth most frequent cancer globally and causes the sixth-highest number of cancer-related fatalities. Histologically, esophageal squamous-cell carcinomas (ESCC) and esophageal adenocarcinomas (EAC) are the two main subtypes of EC [1]. The frequency of EAC is steadily rising in Western nations. ESCC continues to hold a strong position globally [2]. Surgery is an option for treating early-stage EC patients even though most people are still diagnosed with advanced malignancy. Therefore, it appears that more research on the prognosis or early diagnosis of esophageal cancer is required [3–5]. The migration and invasion of tumor cells originating from epithelial cells is known as the epithelial-mesenchymal transition (EMT), an essential biological event. The research's preliminary focus has been

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investigating EMT-related pathways. Dysphagia that manifests at an advanced stage is frequently used to diagnose EC, which is characterized by invasive tumor growth [6, 7]. Due to the invasive disease, individuals may potentially experience tumor recurrence or re-metastasis after therapy. Currently, less than 50% of EC patients survive overall; the percentage may even be as low as 15–25 percent. Depending on the type of cancer category, the inheritance of significant genetic variables influences the susceptibility to cancer to some extent. Genome-wide association studies (GWAS) have identified numerous genetic loci associated with cancer risk within non-coding genome regions. GWA analyses revealed that alterations in the protein's amino acid sequence connected to a minor proportion (about 3.3%) of the 301 SNPs related to cancer successfully uncovered. This highly controversial finding has sparked numerous studies to examine the noncoding regions and their role in the emergence of cancer. The investigations have determined long non-coding RNAs transcribed from non-coding regions associated with cancer risk, where single nucleotide polymorphisms increase the risk of developing cancer. Investigations have revealed that IncRNA plays an important function in regulating EMT. One specific type of IncRNA transcribed from the HOX locus at q13.13.12 is referred to as LncRNA-HOTAIR [2, 3, 7–9]. Several deconstructions have studied the overexpression of IncRNA-HOTAIR in ESCC, suggesting its association with the progressive TNM stage and weak biological differentiation [10]. Investigators uncovered that IncRNA-HOTAIR can control the polycomb repressive complex 2 (PRC2) by methylating the H3K27 of the WIF-1 promoter region. This inhibition leads to decreased WIF-1 expression and production, raising TCF/LEF and reduced beta-catenin levels, finally triggering the Wnt/beta-catenin signaling pathway [11]. This mechanism causes the target gene to become overexpressed, leading to raised invasion and distance of tumor cells, which LncRNA-HOTAIR may also support EMT. LncRNA-HOTAIR depletion leads to increased E-cadherin expression and reduced vitamin and MMP-9 levels in colorectal cancer cells. Thus, LncRNA-HOTAIR is offered as a conceivable new molecular controller for EMT [12]. Recent research on carcinogenesis has shown two forms of LncRNA that are essential players in the process: MALAT1 and NEAT2. Overexpression of IncRNA-MALAT1, which acts as a critical prognostic factor, has been noticed in some malignancies, including lung carcinoma and gallbladder cancer [13, 14].

Reduced hybridization in EAC, NSCL-1, and ESCC patients has disclosed that *IncRNA-MALAT1*, an 8.7 kb nucleus-specific IncRNA, is a predictive characteristic of metastasis [15]. The downregulation of *IncRNA-MALAT1* inhibits the apoptosis pathway, resulting in the upregulation of caspase-3 and -8 and the downregulation of *Bcl-2* and *Bcl-xl* [16]. This IncRNA upregulates in different solid tumors, such as breast, pancreatic, colon, prostate, and liver malignancies. Moreover, differentiable gene expression links it to tumor recurrence and metastasis [17]. The present study aimed to measure the differences in gene expression levels of *IncRNA-HOTAIR* and *IncRNA-MALAT1* in EC patients and emphasize the importance of understanding these IncRNAs in cancer.

2 Materials and methods

2.1 Tissue samples

This experimental study, which analyzed 140 individuals diagnosed with malignant EC at Tabriz International Hospital from 2012 to 2019, has constructed results that significantly improve our understanding of this disease. The investigation involved 70 patients in the two groups. The first group received tumor tissue samples, while the second group received marginal tissue samples. Pathologists and gastroenterologists diagnosed the paraffin samples from the pathology archive used for data collection. The international medical oncology community considers all patients included based on the clinical criteria for the diagnosis of EC. According to the International Union Against Cancer (UICC), patients considered using the tumor-node-metastasis staging system (TNM). Pathologists considered individuals with distinctly malignant EC stages I through IV suitable for inclusion in the study. Patients undergoing chemotherapy or having formerly surgery were excluded from the study. We received signed reported permission and questionnaires from each patient, and the Ahar Branch of Islamic Azad University validated our investigation.

2.2 Sample processing

The Nucleo Spin total RNA FFPE kit (Germany) was used to extract RNA from tumor and non-tumor tissues. RNA extraction was performed during DNase treatment. Before cDNA synthesis, the extracted RNA's quality and quantity were evaluated by UV spectrophotometry at 260/280 nm and 2% Agarose gel electrophoresis (using the NanoDrop TM ND-1,000, NanoDrop Technology, Wilmington, DE, USA). All extracted RNA samples were stored at -80 °C until analysis.



The RevertAid™ First Strand cDNA Synthesis Kit from Thermo Fisher, Inc., in the United States, was used for reverse transcription (RT).

2.3 REAL-TIME PCR

We achieved quantitative real-time polymerase chain reaction (qRT-PCR) employing specific primers to investigate the gene expression differences of IncRNA-HOTAIR and IncRNA-MALAT1 in cancerous versus non-cancerous tissues. For IncRNA-HOTAIR [18], forward: 5'-CAGTGGGGAACTCTGACTCG-3' and reverse: 5'-GTGCCTGGTGCTCTTACC-3'; for IncRNA-MALAT1; for GAPDH as the housekeeping gene, forward: 5'-GTAAGACCCCTGGACCACCA-3' and Reverse: 5'-CAAGGGGTCTACATG GCAACT-3' [19, 20]. To prepare the reaction mixture for qRT-PCR, use the following components: 1 μ l of each primer (at a concentration of 10 pmol), 16 μ l of SYBR Green Real-Time PCR Master Mix (produced by Takara, Japan), and 2 μ l of cDNA. The qRT-PCR examination was achieved employing the ABI7500 System from Applied Biosystems in California, USA. qRT-PCR was used to analyze the IncRNA-HOTAIR and IncRNA-MALAT1 genes. A total of 45 amplification cycles were completed. Each cycle contained an initial denaturation phase at 94 °C for 3 min, followed by three stages: denaturation at 94 °C for 30 s, annealing at 59 °C for 30 s, and extension at 72 °C for 40 s.

2.4 Statistical analysis

We employed the Pearson correlation test to evaluate the levels of *IncRNA-HOTAIR* and *IncRNA-MALAT1* gene expression changes in tumor and margin tissues. Furthermore, we used a chi-square test to investigate the relationship between different clinical characteristics and the expression levels of the IncRNAs *MALAT1* and *HOTAIR*. A two-tailed p-value of less than 0.05 was regarded as showing a significant difference in statistical significance.

3 Results

3.1 LncRNA-HOTAIR gene expression levels in EC patients

Our study, conceived to evaluate the *IncRNA-HOTAIR* gene expression levels in EC patients and identify its potential as a powerful prognostic marker, promises to pave the way for a more precise and effective approach to managing EC, instilling hope and optimism in the field. Our analysis, which included 70 confirmed cases of EC, was comprehensive and inclusive, with a balanced distribution of 33 (47.14%) male and 37 (52.85%) female patients, ensuring a robust and representative sample for our study. Each subject in our investigation, carefully selected to ensure the accuracy and reliability of our findings, underwent surgical intervention as part of their treatment. The cohort, with an age range of 33 to 70 and a mean age of 61.11 ± 3.25 , was meticulously chosen, instilling confidence in the robustness of our study. In the current investigation, the expression levels of the *IncRNA-HOTAIR* gene in tumor and margin tissues were assessed using real-time PCR. The expression of *IncRNA-HOTAIR* was significantly higher in tumor samples compared to non-cancerous tissues (P < 0.05) (Fig. 1).

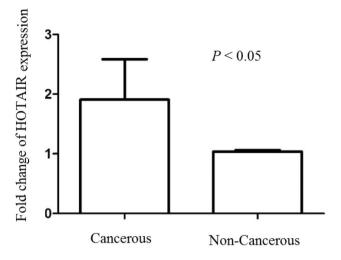
We categorized subjects into two groups based on the median expression levels of IncRNA-HOTAIR: high-expression (median amount \geq 1) and low-expression (median amount < 1). In addition to assessing expression levels, we investigated the relationship between various clinicopathological variables and IncRNA-HOTAIR expression in EC patients. We identified significant associations between IncRNA-HOTAIR expression and factors such as hot drink use, metastasis, and the 5-year survival rate (P = 0.019, P = 0.001, P = 0.001, respectively). None of the following factors showed significant relationships with IncRNA-HOTAIR expression: socioeconomic status (P = 0.739), smoking status (P = 0.309), alcohol abuse (P = 0.301), or phases (P = 0.907) (see Table 1). The discoveries mentioned provide critical insights into the potential role of IncRNA-HOTAIR as a prognostic marker and its association with specific clinical attributes in patients with EC.

3.2 LncRNA- MALAT1 gene expression levels in EC subjects

Compared to the non-cancerous margin samples, tumor samples exhibited a significant increase in the expression levels of the *IncRNA-MALAT1* gene (p < 0.001) (Fig. 2). We divided the participants into two groups based on *IncRNA-MALAT1* expression levels: a high-expression group (median amount \geq 1) and a low-expression group (median amount < 1). This classification facilitated a thorough comparison and analysis of gene expression levels among the



Fig. 1 The fold change of *LncRNA-HOTAIR* gene in cancerous versus noncancerous tissues (*P* < 0.05)



Esophageal Carcinoma

Table 1 The relationship between *IncRNA-HOTAIR* and *IncRNA-MALAT1* expression and patient's demographic and clinical features

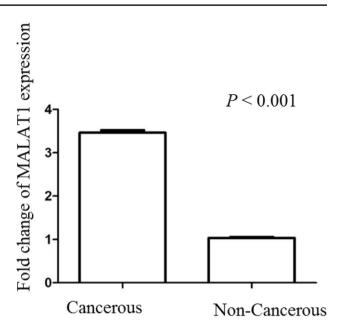
Variables	HOTAIR			MALAT1		
	Low	High	P-value	Low	High	P-value
Stage of disease			0.907			0.304
Stages I, II	23.0	14.0		20.0	17.0	
Stages III, IV	22.0	16.0		12.0	26.0	
Smoking status			0.309			0.642
High	19.0	9.0		23.0	5.0	
Low	27.0	20.0		32.0	15.0	
Alcohol abuse			0.301			0.039
High	12.0	9.0		6.0	15.0	
Low	36.0	18.0		32.0	22.0	
Hot drinks			0.019			0.001
Yes	19.0	27.0		12.0	34.0	
No	12.0	17.0		14.0	15.0	
Socioeconomic status			0.739			0.552
Good	41.0	16.0		32.0	25.0	
Poor	8.0	10.0		6.0	12.0	
Metastasis			0.001			0.001
Non-distant metastasis	12.0	18.0		16.0	14.0	
Distant metastasis	19.0	26.0		13.0	32.0	
5-year survival rate			0.001			0.193
Positive	14.0	20.0		16.0	18.0	
Negative	11.0	30.0		20.0	21.0	

research population. We found significant correlations between changes in *IncRNA-MALAT1* gene expression and factors like hot drink intake, metastasis, and alcohol addiction (P = 0.001).

However, the expression of the *IncRNA-MALAT1* gene and other tumor characteristics did not show a significant correlation (P > 0.05) (Table 1). The results indicate the potential role of *IncRNA-MALAT1* in tumor differentiation and its correlation with specific demographic factors. These results emphasize the potential role of *IncRNA-MALAT1* in tumor differentiation and its correlation with specific demographic factors. Only weak correlations existed between the *IncRNA-HOTAIR* expression changes and specific clinical traits.



Fig. 2 The fold change of *LncRNA-MALAT1* gene in cancerous versus non-cancerous tissues (*P* < 0.001)



Esophageal Carcinoma

4 Discussion

Epigenetic regulation by the IncRNA-HOTAIR is a crucial mechanism in cancer control. Research shows that chromatinmodifying complexes, including LSD1, PRC2, and COREST/REST, regulate the transcription of human HOX genes throughout the genome. LncRNA-HOTAIR serves as a guiding framework, steering these sets to their specific target genes. Unlike the LSD1/COREST combination, which removes methyl groups from H3K4, the PRC2 complex adds methyl groups to H3K27, deactivating IncRNA-HOTAIR target genes. Research suggests that IncRNA-HOTAIR functions as a negative regulator of osteogenic genes such as BMP2 and ALPL, indicating its role as an inhibitor of osteogenesis. Furthermore, IncRNA-HOTAIR is involved in cellular senescence through its interactions with E3 ubiquitin ligases. Additionally, it plays a crucial role in cell proliferation by modulating the expression of proteins and kinases associated with the cell cycle. Investigations have shown that LncRNA-HOTAIR is significantly expressed in BC; measuring its levels is crucial for predicting patient prognosis and evaluating the risk of tumor spread in early-stage BC. Different types of cancers are associated with the function of IncRNA-HOTAIR. It is overexpressed not only in primary pancreatic cancer but also in sarcoma, hepatocellular carcinoma, CRC, laryngeal squamous cell carcinoma, and nasopharyngeal cancer [21-26]. The study examined the expression of IncRNA-HOTAIR in EC patients, finding that tumor tissues exhibited significantly higher gene expression levels than marginal tissues. Studies have shown that IncRNA-HOTAIR is a competitive endogenous RNA in the IncRNA-mRNA network, which is critical for EC. LncRNA-HOTAIR levels have significantly increased in ESCC cells. This increase in IncRNA-HOTAIR led to the upregulation of sponging miR-1 and CCND1, which supports the proliferation of ESCCs. These findings highlight the significance of understanding the role of *IncRNA-HOTAIR* in EC and its potential as a therapeutic target for treatment [27]. Xu et al. indicate that IncRNA-HOTAIR may enhance the expression of snail2 by functioning as a sponge for miR-148a, thereby stimulating EMT. According to reports, homeobox C8 (HOXC8) is up-regulated in several cancer types and has a role in the creation of tumors [28]. Han et al. revealed that IncRNA-HOTAIR can attach to miR-204 as an endogenous RNA candidate to regulate HOXC8. The involvement of CC motif chemokine ligand 18 (CCL18) in tumor metastasis and progression is substantial [29]. Additionally, the determinations displayed that the miR-130a-5p and zinc finger E-box binding homeobox 1 axis, in which CCL18 promoted IncRNA-HOTAIR, accelerated the malignant growth of ESCC. Of particular note, tumors demonstrate a heightened hexokinase 2 (HK2) expression, a significant contributor to aerobic glycolysis. Ma et al. (2017) discovered that IncRNA-HOTAIR sponging of miR-125/miR-143 led to an increased expression of HK2, a crucial factor in the development of ESCC. Understanding the intricate mechanisms of IncRNA-HOTAIR influence on HK2 is critical in our battle against ESCC (29). LncRNA-MALAT1 plays



a significant role in controlling the phosphorylation and distribution of serine-arginine processing factors in nuclear speckles, thereby influencing intermittent processing. The upregulation of IncRNA-MALAT1 in many tumor tissues enhances the proliferation and migration [30]. Research indicates that the expression of IncRNA-MALAT1 increases significantly in certain types of prostate, bladder, and kidney cancers. It enhances cancer cells' growth, survival, and movement and is notably overexpressed in urothelial carcinoma. This IncRNA activates the Wnt signaling pathway, facilitating the transition from an epithelial to a mesenchymal state in laboratory conditions. In renal cell carcinoma, IncRNA-MALAT1 interacts with TFEB, a regulatory transcription factor involved in developmental pathways [31]. The complete TFEB coding sequence remains intact during the integration process. Research has uncovered a significant mechanism in cancer development. The fusion IncRNA-MALAT1 with malignant cells leads to increased cancer development and higher TFEB protein levels. Furthermore, IncRNA-MALAT1 directly enhances gene expression, a critical factor in cancer progression [32]. Two IncRNAs, MALAT1 and TUG1, promote PRC2 interaction with repressed and activated growth-regulating genes. TUG1 expression significantly increases in the later stages of the illness and is elevated in genitourinary cancers [33]. Overexpression of IncRNA-MALAT1 has been identified in multiple tumor types, including gastric, endometrial, colon, lung, bladder, breast, cervical, and colorectal cancers [34–39]. The results of this analysis were in line with the findings of earlier investigations. Wang et al. [40] investigated the impact of IncRNA-MALAT1 expression on patients with EC. The research indicated that IncRNA-MALAT1 plays a crucial role in cancer development and tumor growth, serving as both a molecular marker and a target for cancer therapy. McCabe et al. noted that IncRNA-MALAT1 plays a critical role in facilitating the EMT induced by TGF-B. Inhibiting IncRNA-MALAT1 may serve as a potential approach for monitoring bladder cancer progression [41]. Hu et al. [42] demonstrated that IncRNA-MALAT1 functions as an oncogene by modulating the ATM-CHK2 pathway, thereby regulating the development of ESCC tumors. Additionally, the expression of IncRNA-MALAT1 in tumor tissues may be critical for its regulation. During the progression of ESCC, gene amplification occurs within the tumor tissues. In individuals diagnosed with ESCC, the expression of IncRNA-MALAT1 is significantly higher in cancerous tissues than in the adjacent normal tissues. Suppressing the expression of IncRNA-MALAT1 in cancer cell lines leads to several significant effects: it reduces cell growth, increases programmed cell death, decreases cell movement and invasion, lowers the formation of cell clusters, and halts the cell cycle at the G2/M phase [43]. Research indicates that lowering IncRNA-MALAT1 levels halts the G2/M phase of the cell cycle by activating the checkpoint kinase 2/ataxia-telangiectasia mutated pathway through phosphorylation. Furthermore, it increases the number of cells that undergo apoptosis. LncRNA-MALAT1 is associated with the sensitivity of ESCC cells to chemotherapy and radiation. The suppression of IncRNA-MALAT1 leads to the enhancement of the sensitivity of ESCC cells to chemotherapy and radiation. Cyclin-dependent kinase subunit 1 (Cks1) expression was elevated in ESCCs. Upregulation of Cks1 was associated with increased radiation resistance [43]. Li et al. discovered that Cks1 and IncRNA-MALAT1 expression levels decreased in ESCC mice xenografts and cells after irradiation. The upregulation of IncRNA-MALAT1 also prevented irradiation-induced decreases in cell viability, increases in apoptosis, and reductions in Cks1 levels. The authors conclude that IncRNA-MALAT1 is a positive regulator of radioresistance in ESCC, potentially enhancing the effectiveness of radiotherapy in this condition [44]. The suppression of *IncRNA-MALAT1* has revealed a decrease in cell migration and invasion, a sensation related to increased miR-1-3p expression, and a decrease in the activity of CORO1C/TPM3. The findings exhibited that IncRNA-MALAT1 is directly associated with the seed sequence of miR-1-3p, resulting in the downregulation of miR-1-3p levels. This interchange improves the activity of the CORO1C/TPM3 signaling pathway and promotes the expression of IncRNA-MALAT1, suggesting an interdependent connection between IncRNA-MALAT1 and miR-1-3p in the context of EC invasion and metastasis. In addition, research has shown that miR-101 and miR-217 significantly affect the expression of IncRNA-MALAT1, which in turn is crucial for regulating cell invasion and metastasis. This influence is mediated through their effects on downstream genes, including MIA2, HNF4G, ROBO1, CCT4, and CTHRC1 [45]. Several significant obstacles are evident when examining the constraints related to research on IncRNA-MALAT1 and IncRNA-HOTAIR, particularly regarding their involvement in esophageal cancer and other areas. The challenges originate from the fundamental characteristics of IncRNAs, the techniques employed to study these molecules, and the real-world implications of the findings. One significant limitation of this research is its small sample size, which may affect the reliability of the findings. Engaging a larger number of medical centers and obtaining a broader spectrum of patient samples is essential to enhancing the diagnostic relevance of IncRNA-MALAT1 and IncRNA-HOTAIR as clinical biomarkers for EC. This approach will facilitate a more thorough assessment of treatment efficacy and progress. A notable limitation highlighted in the research on IncRNA-MALAT1 and IncRNA-HOTAIR is the relatively small number of studies conducted across different geographical locations. This scarcity may impact the ability to generalize the findings to a broader range of ethnic populations. The restricted geographical focus of this research may limit the extent to which the findings could be applied to other regions.



Findings indicate a need for more extensive and diverse study samples to verify the predictive potential of *IncRNA-MALAT1* and *IncRNA-HOTAIR* across various cancer types, including EC.

5 Conclusion

The research indicates a significant association between esophageal cancer and the overexpression of *IncRNA-HOTAIR* and *IncRNA-MALAT1*. These IncRNAs appear significantly elevated in tumor tissues and cancer cells, suggesting their potential role in the disease's progression. The findings suggest that their gene expression may serve as a potential genetic marker for esophageal cancer. Furthermore, future research should explore the impact of other cancer-associated IncRNAs on the development and progression of esophageal cancer. Additionally, further research is necessary to understand the methylation processes affecting the promoters of *IncRNA-HOTAIR* and *IncRNA-MALAT1*.

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Author contributions Saeid Ghorbian: Conceived and designed the experiments; Analyzed and interpreted the data. Vahid Ghorbani: Performed the experiments; Contributed reagents, materials, analysis tools or data; Wrote the paper.

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Data availability The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate The work was approved by the Ahar Branch Islamic Azad University (MSc.ID: 950187436), and informed written consent was filled out by all participants. All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional research committee.

Consent for publication Not applicable.

Competing interests The authors declare no competing interests.

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References

- 1. Uhlenhopp DJ, Then EO, Sunkara T, Gaduputi V. Epidemiology of esophageal cancer: update in global trends, etiology and risk factors. Clin J Gastroenterol. 2020;13(6):1010–21.
- 2. Amirmahani F, Vallian S, Asadi MH. The LncRNA MIAT is identified as a regulator of stemness-associated transcript in glioma. Mol Biol Rep. 2023;50(1):517–30.
- 3. Razavi M, Ghorbian S. Up-regulation of long non-coding RNA-PCAT-1 promotes invasion and metastasis in esophageal squamous cell carcinoma. EXCLI J. 2019;18:422.
- 4. Sadeghpour S, Ghorbian S. Evaluation of the potential clinical prognostic value of lncRNA-BANCR gene in esophageal squamous cell carcinoma. Mol Biol Rep. 2019;46(1):991–5.
- 5. Aalijahan H, Ghorbian S. Clinical application of long non-coding RNA-UCA1 as a candidate gene in progression of esophageal cancer. Pathol Oncol Res. 2020;26(3):1441–6.
- Ghorbian S, Ardekani AM. Non-invasive detection of esophageal cancer using genetic changes in circulating cell-free DNA. Avicenna J Med Biotechnol. 2012;4(1):3.
- 7. Ghasemzadeh S, Ghorbian S. Investigation of clinical significant utility of LncRNA-linc02389 in patients with esophageal squamous cell carcinoma. J Kermanshah Univ Med Sci.2023; 27(2):e136290. https://doi.org/10.5812/jkums-136290.
- 8. Oliayi AJ, Asadi MH, Amirmahani F. SNHG6 203 transcript could be applied as an auxiliary factor for more precise staging of breast cancer. J Kerman Univ Med Sci. 2019;26(4):253–9.



- Amirmahani F, Asadi MH, Jannat AF. LncRNA MIAT promotes the proliferation and invasion of colorectal cancer via suppressing apoptosis and senescence. Middle East J Cancer. 2023;14(2):219–29.
- 10. Zhang L, Wang Y, Gao J, Zhou X, Huang M, Wang X, He Z. Non-coding RNA: a promising diagnostic biomarker and therapeutic target for esophageal squamous cell carcinoma. Oncol Lett. 2024;27(6):1–5.
- 11. Li P, Ma X, Huang D. Role of the IncRNA/Wnt signaling pathway in digestive system cancer: a literature review. Eur J Med Res. 2024;29(1):447.
- 12. Cantile M, Belli V, Scognamiglio G, Martorana A, De Pietro G, Tracey M, Budillon A. The role of HOTAIR in the modulation of resistance to anticancer therapy. Front Mol Biosci. 2024;11:1414651.
- 13. Mishra S, Srivastava P, Pandey A, Agarwal A, Shukla S, Husain N. Panel of serum long non-coding RNAs as potential non-invasive biomarkers for gallbladder carcinoma. Non-coding RNA Res. 2024;9(2):583–93.
- 14. Li J, Dhilipkannah P, Holden VK, Sachdeva A, Todd NW, Jiang F. Dysregulation of IncRNA MALAT1 contributes to lung cancer in African Americans by modulating the tumor immune microenvironment. Cancers. 2024;16(10):1876.
- Zhang J, Wu L, Wang C, Xie X, Han Y. Research progress of long non-coding RNA in tumor drug resistance: a new paradigm. Drug Des Dev Ther. 2024;31:1385–98.
- 16. Guo F, Li Y, Liu Y, Wang J, Li Y, Li G. Inhibition of metastasis-associated lung adenocarcinoma transcript 1 in CaSki human cervical cancer cells suppresses cell proliferation and invasion. Acta Biochim Biophys Sin. 2010;42(3):224–9.
- 17. Hjazi A, Jasim SA, Altalbawy FM, Kaur H, Hamzah HF, Kaur I, Deorari M, Kumar A, Elawady A, Fenjan MN. Relationship between lncRNA MALAT1 and chemo-radiotherapy resistance of cancer cells: uncovered truths. Cell Biochem Biophys. 2024;28:1–5.
- 18. Lin K, Jiang H, Zhang LL, Jiang Y, Yang YX, Qiu GD, She YQ, Zheng JT, Chen C, Fang L, Zhang SY. Down-regulated LncRNA-HOTAIR suppressed colorectal cancer cell proliferation, invasion, and migration by mediating p21. Dig Dis Sci. 2018;63:2320–31.
- Gupta DG, Varma N, Kumar A, Naseem S, Sachdeva MU, Bose P, Binota J, Gupta M, Sonam P, Rana P, Malhotra P. Identification and validation of suitable housekeeping genes for gene expression studies in BCR-ABL1 positive B-lineage acute lymphoblastic leukemia. Mol Biol Rep. 2022;49(6):4841–8.
- 20. Gupta DG, Varma N, Abdulkadir SA, Singh P, Sachdeva MU, Naseem S, Siddiqui MR, Bose P, Binota J, Malhotra P, Khadwal A. Identification and validation of the optimal reference genes for standardizing the gene expression profiling diagnostic panel of Ph-like B-lineage acute lymphoblastic leukemia. Clin Exp Med. 2023;23(8):4539–51.
- 21. Abam F, Ghorbian S. The dual role of LncRNAs in hepatocellular carcinoma: friend and foe. Gastroenterol Endosc. 2024;2(4):186–95.
- 22. Moghimi A, Bani Hosseinian N, Mahdipour M, Ahmadpour E, Miranda-Bedate A, Ghorbian S. Deciphering the molecular complexity of hepatocellular carcinoma: unveiling novel biomarkers and therapeutic targets through advanced bioinformatics analysis. Cancer Rep. 2024;7(8):e2152.
- 23. Bagheri R, Ghorbian M, Ghorbian S. Tumor circulating biomarkers in colorectal cancer. Cancer Treat Res Commun. 2024;38:100787.
- 24. Kim K, Jutooru I, Chadalapaka G, Johnson G, Frank J, Burghardt R, Kim S, Safe S. HOTAIR is a negative prognostic factor and exhibits pro-oncogenic activity in pancreatic cancer. Oncogene. 2013;32(13):1616–25.
- 25. Saeedi N, Ghorbian S. Analysis of clinical important of LncRNA-HOTAIR gene variations and ovarian cancer susceptibility. Mol Biol Rep. 2020;47(10):7421–7.
- 26. Zhu C, Wang X, Wang Y, Wang K. Functions and underlying mechanisms of IncRNA HOTAIR in cancer chemotherapy resistance. Cell Death Discov. 2022;8(1):383.
- 27. Ren K, Li Y, Lu H, Li Z, Li Z, Wu K, Li Z, Han X. Long noncoding RNA HOTAIR controls cell cycle by functioning as a competing endogenous RNA in esophageal squamous cell carcinoma. Transl Oncol. 2016;9(6):489–97.
- 28. Xu F, Zhang J. Long non-coding RNA HOTAIR functions as miRNA sponge to promote the epithelial to mesenchymal transition in esophageal cancer. Biomed Pharmacother. 2017;90:888–96.
- 29. Han Y, Zhao G, Shi X, Wang Y, Wen X, Zhang L, Guo X. The emerging role of long non-coding RNAs in esophageal cancer: functions in tumorigenesis and clinical implications. Front Pharmacol. 2022;13:885075.
- 30. Miyashita A, Kobayashi M, Ishibashi S, Nagata T, Chandrasekhar A, Zochodne DW, Yokota T. The role of long noncoding RNA MALAT1 in diabetic polyneuropathy and the impact of its silencing in the dorsal root ganglion by a DNA/RNA heteroduplex oligonucleotide. Diabetes. 2022;71(6):1299–312.
- 31. Bhat R, Shanbhag P. Long non-coding RNAs in kidney injury: a comprehensive review. J Prev Diagn Manag Hum Dis. 2024;4(2):39–52.
- 32. Hirata H, Hinoda Y, Shahryari V, Deng G, Nakajima K, Tabatabai ZL, Ishii N, Dahiya R. Long noncoding RNA MALAT1 promotes aggressive renal cell carcinoma through Ezh2 and interacts with miR-205. Can Res. 2015;75(7):1322–31.
- 33. Bunch H. Gene regulation of mammalian long non-coding RNA. Mol Genet Genomics. 2018;293(1):1-5.
- 34. Zhao Z, Chen C, Liu Y, Wu C. 17β-Estradiol treatment inhibits breast cell proliferation, migration and invasion by decreasing MALAT-1 RNA level. Biochem Biophys Res Commun. 2014;445(2):388–93.
- 35. Iordanishvili S, Metreveli T, Lipartia E, Gachechiladze K, Khuntsaria I, Qobulashvili T, Jorbenadze M, Revazishvili T, Kldiashvili E, Kaufmann AM. The HPV-TP53-MALAT1 Axis: unravelling interactions in cervical cancer development. PLoS ONE. 2023;18(10):e0291725.
- 36. Kan JY, Wu DC, Yu FJ, Wu CY, Ho YW, Chiu YJ, Jian SF, Hung JY, Wang JY, Kuo PL. Chemokine (C-C motif) ligand 5 is involved in tumor-associated dendritic cell-mediated colon cancer progression through non-coding RNA MALAT-1. J Cell Physiol. 2015;230(8):1883–94.
- 37. Ji Q, Liu X, Fu X, Zhang L, Sui H, Zhou L, Sun J, Cai J, Qin J, Ren J, Li Q. Resveratrol inhibits invasion and metastasis of colorectal cancer cells via MALAT1 mediated Wnt/β-catenin signal pathway. PLoS ONE. 2013;8(11):e78700.
- 38. Wang X, Li M, Wang Z, Han S, Tang X, Ge Y, Zhou L, Zhou C, Yuan Q, Yang M. Silencing of long noncoding RNA MALAT1 by miR-101 and miR-217 inhibits proliferation, migration, and invasion of esophageal squamous cell carcinoma cells. J Biol Chem. 2015;290(7):3925–35.
- 39. Okugawa Y, Toiyama Y, Hur K, Toden S, Saigusa S, Tanaka K, Inoue Y, Mohri Y, Kusunoki M, Boland CR, Goel A. Metastasis-associated long non-coding RNA drives gastric cancer development and promotes peritoneal metastasis. Carcinogenesis. 2014;35(12):2731–9.
- 40. Wang C, Zhang Q, Hu Y, Zhu J, Yang J. Emerging role of long non-coding RNA MALAT1 in predicting clinical outcomes of patients with digestive system malignancies: a meta-analysis. Oncol Lett. 2019;17(2):2159–70.
- 41. McCabe EM, Rasmussen TP. IncRNA involvement in cancer stem cell function and epithelial-mesenchymal transitions. Semin Cancer Biol. 2021;75:38–48.



- 42. Hu L, Wu Y, Tan D, Meng H, Wang K, Bai Y, Yang K. Up-regulation of long noncoding RNA MALAT1 contributes to proliferation and metastasis in esophageal squamous cell carcinoma. J Exp Clin Cancer Res. 2015;34(1):7.
- 43. Syllaios A, Moris D, Karachaliou GS, Sakellariou S, Karavokyros I, Gazouli M, Schizas D. Pathways and role of MALAT1 in esophageal and gastric cancer. Oncol Lett. 2021;21(5):1–7.
- 44. Li Z, Zhou Y, Tu B, Bu Y, Liu A, Kong J. Long noncoding RNA MALAT 1 affects the efficacy of radiotherapy for esophageal squamous cell carcinoma by regulating Cks1 expression. J Oral Pathol Med. 2017;46(8):583–90.
- 45. Huang T, Wu Z, Zhu S. The roles and mechanisms of the IncRNA-miRNA axis in the progression of esophageal cancer: a narrative review. J Thorac Dis. 2022;14(11):4545.

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