

This article is licensed under a Creative Commons Attribution-NonCommercial NoDerivatives 4.0 International License.

Clinical Characteristics and Molecular Patterns of *RET*-Rearranged Lung Cancer in Chinese Patients

Kai Zhang,* Huajun Chen,† Ye Wang,‡ Lin Yang,§ Chengzhi Zhou,¶ Weiqiang Yin,# Guangsuo Wang,§ Xinru Mao,** Jianxing Xiang,** Bing Li,** Tengfei Zhang,** and Shihong Fei*

*Cancer Center, Union Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan, P.R. China

†Guangdong Lung Cancer Institute, Guangdong General Hospital and Guangdong Academy of Medical Sciences, Guangzhou, Guangdong, P.R. China

‡Department of Respiratory Medicine, West China Hospital, Chengdu, Sichuan, P.R. China

§Department of Thoracic Surgery, Shenzhen People's Hospital, Second Clinical Medical College of Jinan University, Shenzhen, Guangdong, P.R. China

¶State Key Laboratory of Respiratory Diseases, The First Affiliated Hospital of Guangzhou Medical University, Guangzhou, Guangdong, P.R. China

#Department of Thoracic Surgery, The First Affiliated Hospital of Guangzhou Medical University, Guangzhou, Guangdong, P.R. China

**Burning Rock Biotech, Guangzhou, Guangdong, P.R. China

RET rearrangement has been proven as an oncogenic driver in patients with lung cancer. However, the prevalence, clinical characteristics, molecular features, and therapeutic options in *RET*-rearranged patients remain unclear, especially in Chinese lung cancer patients. We retrospectively collected 6,125 Chinese lung cancer patients who have been profiled using next-generation sequencing (NGS). The clinical demographics and molecular features of *RET* rearrangement-positive patients were analyzed. *RET* rearrangements were identified in 84 patients with a proportion of 1.4% in our cohort. The median age at diagnosis was 58 years, and it mainly occurred in females with adenocarcinoma histology. *KIF5B-RET* was the most frequent fusion type and accounted for 53.8% (57/106) of all *RET* fusions identified, with K15-R12 as the most frequent variant (71.9%). Among 47 *RET*⁺ patients profiled with larger panels, 72.3% (34/47) harbored concurrent alterations. *TP53* ranked as the most common concurrent alteration, and concomitant *EGFR* oncogenic alterations were identified in seven patients. Moreover, an adenocarcinoma patient harboring concurrent *RET* fusion and *EGFR* L858R responded to combinatorial treatment of cabozantinib and osimertinib, with a progression-free survival of 5 months. Our study improved knowledge of clinical characteristics and molecular features of *RET*-rearranged lung cancers in China. It might be helpful to guide clinicians for more effective personalized diagnostic and therapeutic approaches.

Key words: *RET* rearrangement; Lung cancer; Adenocarcinoma; Clinical characteristics; Concurrent gene alteration

INTRODUCTION

The proto-oncogene gene *RET* encodes a receptor tyrosine kinase that can activate downstream pathways such as MAPK/ERK, PI3K/AKT, and JAK/STAT¹. *RET* plays a critical role in cell proliferation, migration, and differentiation^{2–5}. *RET* rearrangement was first identified in NIH-3T3 cells transfected with lymphoma DNA in 1985⁶, and chromosomal rearrangement could lead to constitutive activation of *RET* kinase and downstream signaling events, which cause tumorigenesis.

Fusion of *RET* more frequently occurs in radiation-induced papillary thyroid cancer^{7,8}. The prevalence of *RET* rearrangement is 0.7%–2% in lung cancer and 1%–2% in non-small cell lung cancer (NSCLC)⁹. To date, several fusion partners have been identified, such as *KIF5B*, *CCDC6*, *NCOA4*, and *TRIM33* in lung cancer, with *KIF5B-RET* fusion accounting for the major proportion¹⁰. The coiled-coil domain of *RET* partner gene *KIF5B* can activate *RET* tyrosine kinase domain by ligand-independent homodimerization and autophosphorylation, leading to

the constitutive activation of downstream pathways and tumorigenesis¹¹.

Investigation of the prevalence, clinical demographics, and molecular pattern of oncogenic rearrangements may provide comprehensive genomic profiling as well as aiding in the selection of patients for optimal therapies. However, results from studies of *RET* rearrangements in lung cancer are still inconclusive, especially in Chinese patients. Moreover, previous reports about the prevalence and clinical characteristics are conflicting. Lin et al. reported that *RET* rearrangements were more prone to occur in younger age, never-smokers, females with adenocarcinoma in lung cancer¹². Some studies revealed that there was no statistically significant difference in gender and smoking status, or even drew an opposite conclusion in terms of gender^{13,14}. Therefore, we retrospectively analyzed 6,125 Chinese lung cancer patients for *RET* rearrangement using next-generation sequencing and identified 84 *RET* fusion-positive patients. This study demonstrated clinical demographics and molecular features of *RET* rearrangement in Chinese lung cancer patients.

MATERIALS AND METHODS

Patient Information

A total of 6,125 samples (either tissue or plasma) from lung cancer patients were consecutively collected from September 2015 to July 2017 in this retrospective *RET* rearrangement study. There was no preselection with smoking, gender, clinical stage, or age of patients. Eligible patients were histologically diagnosed as lung cancer according to the latest World Health Organization Criteria. Cancer stage was evaluated based on the 7th TNM classification. *RET* rearrangements were identified using next-generation sequencing. Our profiling panels (Burning Rock Biotech, Guangzhou, P.R. China), consisting of 8, 56, 168, or 295 cancer-related genes, were designed and validated for identification of base substitutions, insertions, deletions, copy number variations, and gene fusion. This entire study was approved by the institutional review board of Union Hospital, Tongji Medical College.

Preparation of Tissue DNA and Plasma Cell-Free DNA

Tissue DNA and plasma cell-free DNA were extracted using QIAamp DNA formalin-fixed paraffin-embedded (FFPE) tissue kit (Qiagen, Valencia, CA, USA) and QIAamp Circulating Nucleic Acid kit (Qiagen), respectively, following the manufacturer's instructions.

NGS Library Preparation and Sequencing

DNA shearing was performed using Covaris M220 (Covaris Inc., Woburn, MA, USA) and followed by end repair, phosphorylation, and adaptor ligation. Then

200- to 400-bp fragments were selected by bead (Agen-court AMPure XP Kit; Beckman Coulter, Brea, CA, USA) and hybridized with capture probes baits (SureSelectXT Custom 1kb-499kb; Agilent, Santa Clara, CA, USA). Hybrid selection was performed using magnetic beads (Dynabeads™ MyOne™ Streptavidin T1; Thermo Fisher Scientific, Waltham, MA, USA) and followed by PCR amplification. Bioanalyzer (LabChip GX Touch 24 Nucleic Acid Analyzer; Perkin-Elmer, Waltham, MA, USA) was used to evaluate DNA quality and size by high-sensitivity DNA assay. Indexed samples were sequenced on NextSeq 500 (Illumina, Inc., San Diego, CA, USA).

NGS Data Analysis

All the reads were mapped to the human genome (hg19) with Burrows–Wheeler Aligner (BWA)¹⁵. Local alignment optimization, mark duplication, and variant calling were performed using Genome Analysis ToolKit (GATK) 3.2¹⁶, picards, and VarScan¹⁷. Gene rearrangements were called with FACTERA¹⁸, and CNVs were analyzed based on sequencing depth. Variants were filtered using the VarScan ffilter pipeline, and loci with depth less than 100 were filtered out. At least two and five supporting reads were needed for INDELs, while eight supporting reads were needed to call SNVs, in both plasma and tissue samples. According to the ExAC, 1000 Genomes, dbSNP, ESP6500SI-V2 database, variants with population frequency over 0.1% were grouped as SNP and excluded from further analysis. Remaining variants were annotated with ANNOVAR¹⁹ and SnpEff v3.6²⁰.

RESULTS

Patient Characteristics

To interrogate *RET* fusion patterns, we retrospectively screened 6,125 NSCLC patients from September 2015 to July 2017 and identified 84 patients harboring *RET* fusion, giving an overall frequency of 1.4%. Thirty-six (42.9%) of them were males and 47 (56.0%) were females. Gender information of the remaining patient (1.2%) was not recorded. Compared to patients in the original cohort ($n=6,125$; 3,411 males, 2,578 females, and 136 unknown), the prevalence of females was significantly higher in the *RET*⁺ cohort ($n=84$, $p=0.0228$, Pearson's chi-squared test). Median age of patients harboring *RET* fusion was 58 years, ranging from 35 to 81 years. Compared to patient age in the 6,125 original cohort (median age=61 years), no preference pattern in terms of age was found in the *RET*⁺ cohort. As to histological subtype, a majority of them (62/84, 73.8%) were diagnosed as lung adenocarcinoma. One patient was diagnosed with lung squamous cell carcinoma, and three patients had a mixture of adenocarcinoma and squamous cell carcinoma. Histological information of the other

Table 1. Summary of Baseline Characteristics of Patients Harboring *RET* Rearrangement (*N* = 84)

| Patient Characteristics | <i>n</i> (%) |
|-------------------------|--------------|
| Gender | |
| Male | 36 (42.9%) |
| Female | 47 (56.0%) |
| Unknown | 1 (1.2%) |
| Age (years) | |
| Median | 58 |
| Range | 35–81 |
| Histological types | |
| LUAD | 62 (72.8%) |
| LUSC | 1 (1.2%) |
| Mixed LUAD and LUSC | 3 (3.6%) |
| Other lung cancers | 18 (21.4%) |

LUAD, lung adenocarcinoma; LUSC, lung squamous cell carcinoma.

18 lung cancer patients was not recorded. The detailed patient characteristics are summarized in Table 1.

Overview of RET Fusion Patterns

To interrogate *RET* fusion patterns, we performed ultra-deep targeted sequencing on plasma using panels

covering critical exons and introns of lung cancer-related genes. Among them, 32 patients used a panel consisting of 168 lung cancer-related genes, spanning 170 kb of human genome; 10 patients used a 56-gene panel, and the 37 patients used a panel consisting of 7 well-known lung cancer driver genes plus *KRAS*, a well-established prognostic factor. Five patients used a panel consisting of 295 cancer-related genes.

In this cohort, a total of 106 *RET* rearrangements in 84 patients were identified with *RET* rearrangement. Structures of all the *RET* rearrangements with detailed demonstration of breakpoints are presented in Figure 1. The most commonly seen partner was *KIF5B*, and 57 *KIF5B-RET* fusion events were identified with a frequency of 53.8% (57/106) in all *RET* rearrangements and in 67.9% (57/84) patients (Fig. 2). The most frequent variant of *KIF5B-RET* was K15-R12, occurring in 71.9% (41/57) of *KIF5B-RET*⁺ patients (Fig. 1). This result was in agreement with previous literature, which reported the occurring frequency of about 75%^{11,21–23}.

The second- and third-ranked fusion partners were *CCDC6* and *NCOA4*, occurring in 17.0% (18/106) and 2.8% (3/106) of all *RET* fusions, and in 21.4% (18/84) and 3.6% (3/84) of patients, respectively. Several rare and novel *RET* fusion partners were identified in our study,



Figure 1. Structure and breakpoints of 106 *RET* fusions detected in 84 lung cancer patients by next-generation sequencing.

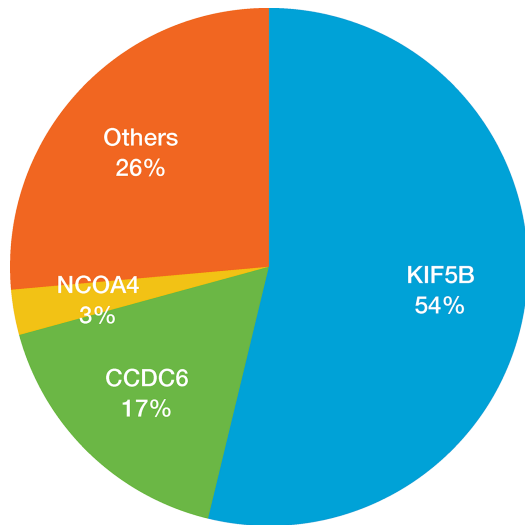


Figure 2. Distribution of various *RET* rearrangement partners identified in the 84 lung cancer patients. Different colors and sizes indicate the occurrence frequency of each *RET* fusion partner in the overall *RET* fusion identified ($n = 106$).

including *TSSK4*, *SORBS1*, *SIRT1*, *PTPRK*, *ADD3-AS1*, *PRKG1*, *IL2RA*, *CCNYL2*, *CCDC186*, and *ANKS1B*. Table 2 lists detailed breakpoints and patient histology of novel *RET* fusions identified in this study, which have not been reported before to the best of our knowledge.

In addition, we observed that multiple *RET* fusions can be detected in individual patients, and 21 patients harbored more than one *RET* fusion. Most studies revealed that for an individual patient harboring multiple fusions, the hotspot partner commonly serves as driver mutation²⁴.

Concurrent Genomic Alterations in *RET* Fusion-Positive Patients

Since the discovery of *RET* fusion, there has been no definitive conclusion about the mutual exclusivity between *RET* fusion and other genomic alterations^{14,25,26}. We interrogated the mutation spectrum of lung cancer patients with positive *RET* fusions. Patients tested with the eight-gene panel were excluded from this analysis due to the rare chance of harboring dual drivers. Among 47 patients tested with larger panels, 34 of them (72.3%) harbored concurrent mutations, with *TP53* being the most commonly seen concurrent mutation, occurring in 42.5% (20/47) of patients. It was followed by *EGFR* and *MYC*, occurring in 14.9% (7/47) and 10.6% (5/47) of patients, respectively (Fig. 3). The underlying mechanisms of *RET* co-occurrence with other mutations and influence on clinical outcomes are needed to be addressed in further studies.

It has been regarded that actionable driver mutations were commonly mutually exclusive^{27–29}. However, the coalteration of *EGFR* and other driver mutations such as

Table 2. Novel *RET* Fusion Partners Identified in Chinese Lung Cancer Patients

| Fusion | Breakpoints | Histology |
|---------------------|---------------------|----------------|
| <i>TSSK4-RET</i> | Intron1_Intron11 | Adenocarcinoma |
| <i>SORBS1-RET</i> | Intron8_Intron11 | Adenocarcinoma |
| <i>SIRT1-RET</i> | Intron8_Intron11 | Adenocarcinoma |
| <i>PTPRK-RET</i> | Intron3_Intron11 | Adenocarcinoma |
| <i>ADD3-AS1-RET</i> | Intron1_Intron11 | Adenocarcinoma |
| <i>PRKG1-RET</i> | Intron7_Intron11 | Adenocarcinoma |
| <i>IL2RA-RET</i> | Intergenic_Intron11 | Adenocarcinoma |
| <i>CCNYL2-RET</i> | Intron6_Intron15 | Not available |
| <i>CCDC186-RET</i> | Intron10_Intron11 | Adenocarcinoma |
| <i>ANKS1B-RET</i> | Intron1_Intron11 | Adenocarcinoma |

ALK in a subset of lung cancers has been reported and challenged previous dogma^{30–32}. In this cohort, the co-occurrence of *RET* fusion with *EGFR* oncogenic genetic alterations was observed in seven patients, consisting of five exon 19 deletions, two L858R mutations, and two T790M mutations. All seven patients had received previous treatment before the positive detection of *RET* and *EGFR* alterations. One case received previous chemotherapy, and the other six cases received previous EGFR-TKI treatment (including the two T790M⁺ patient), indicating that *RET* fusion maybe one of the mechanisms that contributes to resistance of EGFR TKI.

Interestingly, we found that no *KIF5B-RET* was identified in the seven patients who harbored *EGFR* mutations. The mutual exclusivity of *KIF5B-RET* and *EGFR* alterations suggested that *KIF5B-RET* was a strong driver mutation. For the seven patients harboring concurrent *EGFR* and non-*KIF5B-RET* fusion, the fusion types of *RET* included *CCNYL2-RET* ($n = 1$), *PCMI-RET* ($n = 1$), *CCDC6-RET* ($n = 3$), and *NCOA4-RET* ($n = 2$). Among them, sequencing samples of six patients were plasma. We observed that overall allelic fraction (AF) ratio of first-generation EGFR-TKI sensitizing mutations was higher than non-*KIF5B-RET* in each of the six patients (AF ratio of *EGFR/RET* = 2.4) (Fig. 4), indicating that non-*KIF5B-RET* fusion might function as a potential acquired resistance mechanism to EGFR tyrosine kinase inhibitors. The *RET* rearrangement may exist as a minor clone with *EGFR*-sensitive alterations and expanded while the *EGFR*-sensitive alterations were inhibited by EGFR-TKI.

Clinical Outcomes of an *EGFR* and *RET* Fusion Concurrent Patient Treated With Cabozantinib

Several tyrosine kinase inhibitors (TKIs), such as vandetanib, cabozantinib, and sunitinib, have been proven with anti-*RET* activity. Cabozantinib inhibits a broad range of tyrosine kinases and displayed a 28% ORR, and median PFS and OS of 5.5 months and 9.9 months, respectively, for patients with advanced *RET*-rearranged NSCLC³³.

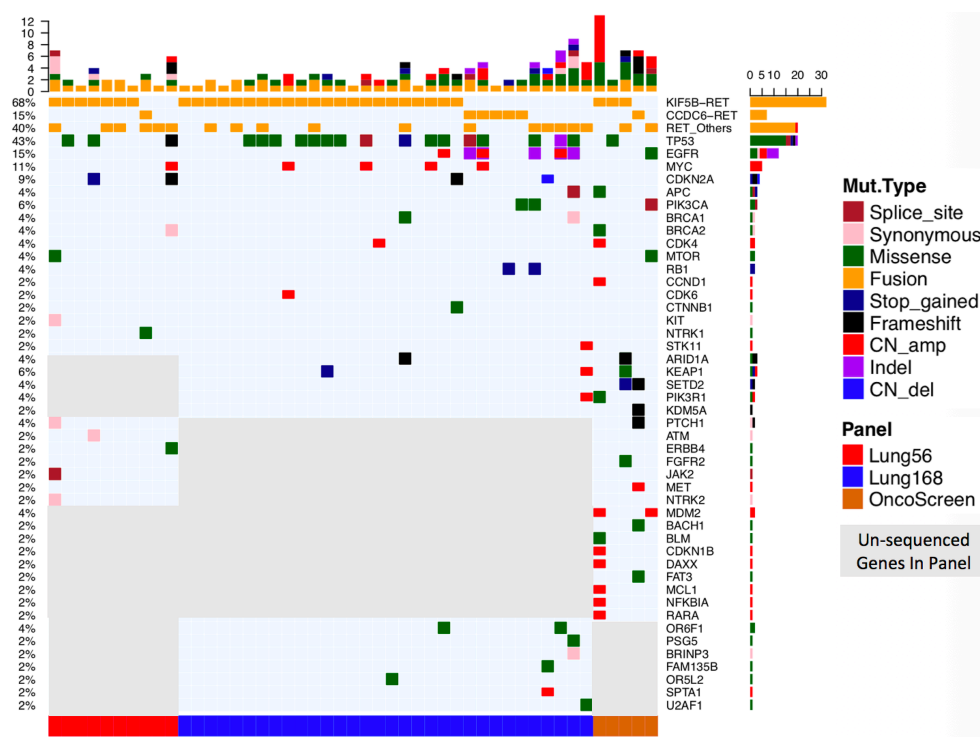


Figure 3. Concurrent genetic alternation analysis demonstrated by oncoPrint. The top bar indicates the number of mutation in each patient. The right-side bar demonstrates the number of patient harboring a specific mutation. Different colors mean different mutation type categories.

In our cohort, clinical outcomes of cabozantinib were available for six *RET*-rearranged patients. The median treatment period of cabozantinib for these patients was 5 months. Among them, one patient was identified harboring concurrent *RET* fusion and *EGFR* mutation. It was a 65-year-old male patient diagnosed with stage IV lung adenocarcinoma with bone metastasis. He was treated with gefitinib and osimertinib as the first- and second-line therapy, and achieved stable disease after the two lines of treatment, with progression-free survival of 8 months and 4 months, respectively. After development of resistance to osimertinib, NGS revealed that this patient had concurrent *CCNYL2-RET* fusion and *EGFR* L858R. Combination of osimertinib and cabozantinib was used to treat this patient after the positive detection of the two concurrent alterations. He achieved stable disease (SD) with a tumor shrinkage of 13% 1 month after treatment initiation. Finally, he experienced disease progression after a PFS of 5 months.

DISCUSSION

In this study, we retrospectively analyzed molecular profiling data of 6,125 Chinese lung cancer patients and identified 84 patients harboring *RET* rearrangements, accounting for 1.4% of this cohort. This ratio was in agreement with a previous study about *RET* fusion²⁴.

We investigated the distinct clinical characteristics of *RET* fusion patients. Furthermore, we analyzed the fusion partners, demonstrated their molecular pattern, and investigated the mutual exclusivity of *RET* fusion with other concomitant gene alterations.

Several studies have investigated the correlation of *RET* fusion and clinical demographics in lung cancer, and most of them revealed that *RET* rearrangements were more prone to occur in lung adenocarcinomas.³⁴ In our study, we observed similar results that most of *RET*⁺ patients were lung adenocarcinomas. However, there were discrepancies among previous studies about other factors such as gender and age. Michels et al. reported that rearrangements of *RET* occurred with a high proportion of men (59% vs. 41%) and median age of 62 years in a European cohort¹⁴. Another study carried out in Japanese patients revealed that *RET* fusion was not associated with gender ($p=0.524$) but significantly correlated with younger age (57.5 years)¹³. A meta-analysis reported that *RET* fusions were identified at significantly high frequency in younger (<60 years) females¹². In our Chinese cohort, we observed that *RET* rearrangement had a tendency to occur in females with a median age of 58 years at diagnosis. One explanation is that this discrepancy may have resulted from differences in ethnicity, lifestyles, environmental factors, or molecular heterogeneity. Therefore, studies are needed

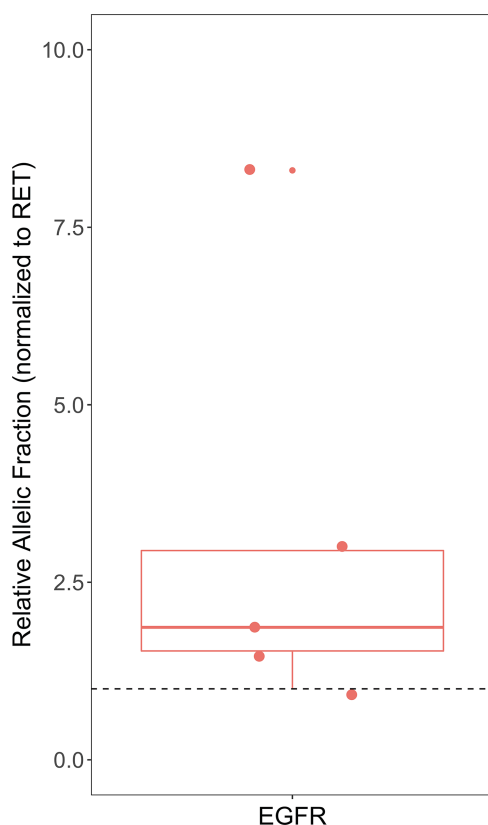


Figure 4. Relative allelic fraction of *EGFR* in seven *RET*-rearranged patients. The y-axis indicates *EGFR* relative allelic fraction that was normalized to *RET* fusion in each individual patient.

to further investigate the underlying relationship of *RET* fusion patients and these factors.

KIF5B was the most frequently appeared partner of *RET* fusion and accounted for 53.8% of all the *RET* fusions identified in our study. For *KIF5B-RET*, the most common variant was K15-R12, occurring in 71.9% of all *KIF5B-RET* fusions. The occurring frequencies in this study was similar with previous literature^{11,21–23}. However, no consistent conclusion was obtained in terms of clinical outcomes to *RET* inhibitors across different *RET* fusion types¹⁰. Further studies need to address this issue to provide more guidance for patients at high risks so that optimal treatment strategy may be implemented.

The mutual exclusivity of *RET* fusion and other molecular alterations has been poorly investigated, and no definitive conclusion has been obtained to date. Some studies reported that *RET* fusion was exclusive with other gene alterations. However, Song et al. reported several concomitant genomic alterations, including *EGFR*, *MAP2K1*, *CTNNB1*, and *AKT1*, occurred in 4 of 11 *RET*-rearranged NSCLC patients, with a frequency of 36.4%.²⁵ Similar co-occurrence frequency of genetic alterations was also found in another study in 10 of

22 *RET*⁺ patients (45.5%), consisting of eight *TP53*, one *MET* amplification, one *CTNNB1*, and one *EGFR* rare mutation L833F¹⁴. In our study, we also identified several *RET* fusion co-occurring mutations, and *EGFR* oncogenic driver mutation was identified in seven *RET*⁺ patients. It was reported that patients harboring both *RET* fusion and *EGFR* mutation were resistant to *EGFR* TKIs^{26,35,36}, which suggested that *RET* fusion maybe one of the mechanisms that contributed to resistance of *EGFR* TKI. This study provided the basis for the hypothesis that an actionable driver mutation could function as an acquired resistant mechanism for another actionable driver alteration.

To date, chemotherapy is still the standard first-line treatment for *RET*-rearranged patients. Several *RET* inhibitors have been developed, but the overall outcomes to *RET* inhibitors were inferior to targeted therapies in other lung cancer oncogenic mutations like *ALK* and *ROS1*. Cabozantinib displayed an overall response rate (ORR) of 28% and median PFS of 5.5 months in a phase II clinical trial of *RET*-rearranged lung cancers ($n=26$)³³. The ORR to vandetanib was reported as 18% ($n=19$) and to lenvatinib was 16% ($n=25$) in Korean patients. Several reasons are attributed to less sensitivity of *RET* inhibitors. One reason was that the *RET* inhibitors could lead to toxicities due to the activity to VEGFR kinase; thus, the clinical uses of these inhibitors were often reduced to 70% at a suboptimal dose³³. Therefore, more potent and selective *RET* inhibitors that do not target VEGFR are needed to increase the sensitivity and reduce the off-target toxicity. Moreover, combinatorial treatment strategy is another approach to be taken into account.

There were still some limitations in this study. First, we only analyzed the prevalence of *RET* fusion in terms of gender, age, and histology, and other clinical characteristics like smoking status, tumor size, and metastatic status were not included due to incompleteness of clinical records. Second, owing to the lack of response and survival information, we did not perform survival analysis to interrogate the clinical outcomes across different *RET* variants. Last, but not the least, further validations are needed to support the hypothesis that non-*KIF5B-RET* may serve as an acquired resistance mechanism for another driver mutation.

The development of next-generation sequencing greatly improved the molecular diagnosis of cancer. The knowledge of specific clinical features associated with *RET* fusion can guide patients at high risk for precise diagnosis and treatment strategy. We further propose that studies to be carried out include more clinical feature analysis in *RET* fusion-positive patients and prognostic prediction evaluation among different *RET* fusions variants.

ACKNOWLEDGMENTS: The authors would like to thank all the patients and their families. This work was supported by the National Natural Science Foundation of China (Grant

Nos. 81773056 and 81372260). K.Z., H.C., Y.W., L.Y., C.Z., W.Y., G.W., and S.F. contributed to the collection of clinical samples, pathological diagnosis, and experimental design. X.M. and J.X. performed data analysis. B.L. contributed to bioinformatics analyses and figure generation. T.Z. conducted manuscript writing. The authors declare no conflicts of interest.

REFERENCES

- Phay JE, Shah MH. Targeting RET receptor tyrosine kinase activation in cancer. *Clin Cancer Res*. 2010; 16(24):5936–41.
- Alberti L, Carniti C, Miranda C, Roccato E, Pierotti MA. RET and NTRK1 proto-oncogenes in human diseases. *J Cell Physiol*. 2003;195(2):168–86.
- Faivre S, Djelloul S, Raymond E. New paradigms in anti-cancer therapy: Targeting multiple signaling pathways with kinase inhibitors. *Semin Oncol*. 2006;33(4):407–20.
- Qian Y, Chai S, Liang Z, Wang Y, Zhou Y, Xu X, Zhang C, Zhang M, Si J, Huang F, Huang Z, Hong W, Wang K. KIF5B-RET fusion kinase promotes cell growth by multilevel activation of STAT3 in lung cancer. *Mol Cancer* 2014;13:176.
- Gainor JF, Shaw AT. Novel targets in non-small cell lung cancer: ROS1 and RET fusions. *Oncologist* 2013;18(7): 865–75.
- Takahashi M, Ritz J, Cooper GM. Activation of a novel human transforming gene, *ret*, by DNA rearrangement. *Cell* 1985;42(2):581–8.
- Bounacer A, Wicker R, Schlumberger M, Sarasin A, Suarez HG. Oncogenic rearrangements of the *ret* proto-oncogene in thyroid tumors induced after exposure to ionizing radiation. *Biochimie* 1997;79(9–10):619–23.
- Hamatani K, Eguchi H, Ito R, Mukai M, Takahashi K, Taga M, Imai K, Cologne J, Soda M, Arihiro K, Fujihara M, Abe K, Hayashi T, Nakashima M, Sekine I, Yasui W, Hayashi Y, Nakachi K. RET/PTC rearrangements preferentially occurred in papillary thyroid cancer among atomic bomb survivors exposed to high radiation dose. *Cancer Res*. 2008;68(17):7176–82.
- Lipson D, Capelletti M, Yelensky R, Otto G, Parker A, Jarosz M, Curran JA, Balasubramanian S, Bloom T, Brennan KW, Donahue A, Downing SR, Frampton GM, Garcia L, Juhn F, Mitchell KC, White E, White J, Zvirko Z, Peretz T, Nechushtan H, Soussan-Gutman L, Kim J, Sasaki H, Kim HR, Park SI, Ercan D, Sheehan CE, Ross JS, Cronin MT, Janne PA, Stephens PJ. Identification of new ALK and RET gene fusions from colorectal and lung cancer biopsies. *Nat Med*. 2012;18(3):382–4.
- Gautschi O, Milia J, Filleron T, Wolf J, Carbone DP, Owen D, Camidge R, Narayanan V, Doebele RC, Besse B, Remon-Masip J, Janne PA, Awad MM, Peled N, Byoung CC, Karp DD, Van Den Heuvel M, Wakelee HA, Neal JW, Mok TSK, Yang JCH, Ou SI, Pall G, Froesch P, Zalcman G, Gandara DR, Riess JW, Velcheti V, Zeidler K, Diebold J, Fruh M, Michels S, Monnet I, Popat S, Rosell R, Karachaliou N, Rothschild SI, Shih JY, Warth A, Muley T, Cabillie F, Mazieres J, Drilon A. Targeting RET in patients with RET-rearranged lung cancers: Results from the Global, Multicenter RET Registry. *J Clin Oncol*. 2017;35(13):1403–10.
- Ju YS, Lee WC, Shin JY, Lee S, Bleazard T, Won JK, Kim YT, Kim JI, Kang JH, Seo JS. A transforming KIF5B and RET gene fusion in lung adenocarcinoma revealed from whole-genome and transcriptome sequencing. *Genome Res*. 2012;22(3):436–45.
- Lin C, Wang S, Xie W, Chang J, Gan Y. The RET fusion gene and its correlation with demographic and clinicopathological features of non-small cell lung cancer: A meta-analysis. *Cancer Biol Ther*. 2015;16(7):1019–28.
- Tsuta K, Kohno T, Yoshida A, Shimada Y, Asamura H, Furuta K, Kushima R. RET-rearranged non-small-cell lung carcinoma: A clinicopathological and molecular analysis. *Br J Cancer* 2014;110(6):1571–8.
- Michels S, Scheel AH, Scheffler M, Schultheis AM, Gautschi O, Aebbersold F, Diebold J, Pall G, Rothschild S, Bubendorf L, Hartmann W, Heukamp L, Schildhaus HU, Fassunke J, Ihle MA, Kunstlinger H, Heydt C, Fischer R, Nogova L, Mattonet C, Hein R, Adams A, Gerigk U, Schulte W, Luders H, Grohe C, Graeven U, Muller-Naendrup C, Draube A, Kambartel KO, Kruger S, Schulze-Olden S, Serke M, Engel-Riedel W, Kaminsky B, Randerath W, Merkelbach-Bruse S, Buttner R, Wolf J. Clinicopathological characteristics of RET rearranged lung cancer in European patients. *J Thorac Oncol*. 2016;11(1):122–7.
- Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 2009; 25(14):1754–60.
- McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernytsky A, Garimella K, Altshuler D, Gabriel S, Daly M, DePristo MA. The Genome Analysis Toolkit: A MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res*. 2010;20(9):1297–303.
- Koboldt DC, Zhang Q, Larson DE, Shen D, McLellan MD, Lin L, Miller CA, Mardis ER, Ding L, Wilson RK. VarScan 2: Somatic mutation and copy number alteration discovery in cancer by exome sequencing. *Genome Res*. 2012;22(3):568–76.
- Newman AM, Bratman SV, Stehr H, Lee LJ, Liu CL, Diehn M, Alizadeh AA. FACTERA: A practical method for the discovery of genomic rearrangements at breakpoint resolution. *Bioinformatics* 2014;30(23):3390–3.
- Wang K, Li M, Hakonarson H. ANNOVAR: Functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res*. 2010;38(16):e164.
- Cingolani P, Platts A, Wang le L, Coon M, Nguyen T, Wang L, Land SJ, Lu X, Ruden DM. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of *Drosophila melanogaster* strain w1118; iso-2; iso-3. *Fly (Austin)* 2012; 6(2):80–92.
- Yokota K, Sasaki H, Okuda K, Shimizu S, Shitara M, Hikosaka Y, Moriyama S, Yano M, Fujii Y. KIF5B/RET fusion gene in surgically-treated adenocarcinoma of the lung. *Oncol Rep*. 2012;28(4):1187–92.
- Cai W, Su C, Li X, Fan L, Zheng L, Fei K, Zhou C. KIF5B-RET fusions in Chinese patients with non-small cell lung cancer. *Cancer* 2013;119(8):1486–94.
- Suehara Y, Arcila M, Wang L, Hasanovic A, Ang D, Ito T, Kimura Y, Drilon A, Guha U, Rusch V, Kris MG, Zakowski MF, Rizvi N, Khanin R, Ladanyi M. Identification of KIF5B-RET and GOPC-ROS1 fusions in lung adenocarcinomas through a comprehensive mRNA-based screen for tyrosine kinase fusions. *Clin Cancer Res*. 2012;18(24):6599–608.
- Ferrara R, Auger N, Auclin E, Besse B. Clinical and translational implications of RET rearrangements in non-small cell lung cancer. *J Thorac Oncol*. 2018;13(1):27–45.
- Song Z, Yu X, Zhang Y. Clinicopathologic characteristics, genetic variability and therapeutic options of RET

- rearrangements patients in lung adenocarcinoma. *Lung Cancer* 2016;101:16–21.
26. Klemptner SJ, Bazhenova LA, Braiteh FS, Nikolinakos PG, Gowen K, Cervantes CM, Chmielecki J, Greenbowe JR, Ross JS, Stephens PJ, Miller VA, Ali SM, Ou SH. Emergence of RET rearrangement co-existing with activated EGFR mutation in EGFR-mutated NSCLC patients who had progressed on first- or second-generation EGFR TKI. *Lung Cancer* 2015;89(3):357–9.
 27. Shaw AT, Yeap BY, Mino-Kenudson M, Digumarthy SR, Costa DB, Heist RS, Solomon B, Stubbs H, Admane S, McDermott U, Settleman J, Kobayashi S, Mark EJ, Rodig SJ, Chirieac LR, Kwak EL, Lynch TJ, Iafrate AJ. Clinical features and outcome of patients with non-small-cell lung cancer who harbor EML4-ALK. *J Clin Oncol*. 2009;27(26):4247–53.
 28. Soda M, Choi YL, Enomoto M, Takada S, Yamashita Y, Ishikawa S, Fujiwara S, Watanabe H, Kurashina K, Hatanaka H, Bando M, Ohno S, Ishikawa Y, Aburatani H, Niki T, Sohara Y, Sugiyama Y, Mano H. Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature* 2007;448(7153):561–6.
 29. Horn L, Pao W. EML4-ALK: Honing in on a new target in non-small-cell lung cancer. *J Clin Oncol*. 2009;27(26):4232–5.
 30. Yang JJ, Zhang XC, Su J, Xu CR, Zhou Q, Tian HX, Xie Z, Chen HJ, Huang YS, Jiang BY, Wang Z, Wang BC, Yang XN, Zhong WZ, Nie Q, Liao RQ, Mok TS, Wu YL. Lung cancers with concomitant EGFR mutations and ALK rearrangements: Diverse responses to EGFR-TKI and crizotinib in relation to diverse receptors phosphorylation. *Clin Cancer Res*. 2014;20(5):1383–92.
 31. Ulivi P, Chiadini E, Dazzi C, Dubini A, Costantini M, Medri L, Puccetti M, Capelli L, Calistri D, Verlicchi A, Gamboni A, Papi M, Mariotti M, De Luigi N, Scarpi E, Bravaccini S, Turolla GM, Amadori D, Crino L, Delmonte A. Nonsquamous, non-small-cell lung cancer patients who carry a double mutation of EGFR, EML4-ALK or KRAS: Frequency, clinical-pathological characteristics, and response to therapy. *Clin Lung Cancer* 2016;17(5):384–90.
 32. Won JK, Keam B, Koh J, Cho HJ, Jeon YK, Kim TM, Lee SH, Lee DS, Kim DW, Chung DH. Concomitant ALK translocation and EGFR mutation in lung cancer: A comparison of direct sequencing and sensitive assays and the impact on responsiveness to tyrosine kinase inhibitor. *Ann Oncol*. 2015;26(2):348–54.
 33. Drilon A, Rekhtman N, Arcila M, Wang L, Ni A, Albano M, Van Voorthuysen M, Somwar R, Smith RS, Montecalvo J, Plodkowski A, Ginsberg MS, Riely GJ, Rudin CM, Ladanyi M, Kris MG. Cabozantinib in patients with advanced RET-rearranged non-small-cell lung cancer: An open-label, single-centre, phase 2, single-arm trial. *Lancet Oncol*. 2016;17(12):1653–60.
 34. Platt A, Morten J, Ji Q, Elvin P, Womack C, Su X, Donald E, Gray N, Read J, Bigley G, Blockley L, Cresswell C, Dale A, Davies A, Zhang T, Fan S, Fu H, Gladwin A, Harrod G, Stevens J, Williams V, Ye Q, Zheng L, de Boer R, Herbst RS, Lee JS, Vasselli J. A retrospective analysis of RET translocation, gene copy number gain and expression in NSCLC patients treated with vandetanib in four randomized Phase III studies. *BMC Cancer* 2015;15:171.
 35. Hu W, Liu Y, Chen J. Concurrent gene alterations with EGFR mutation and treatment efficacy of EGFR-TKIs in Chinese patients with non-small cell lung cancer. *Oncotarget* 2017;8(15):25046–54.
 36. Schrock AB, Zhu VW, Hsieh WS, Madison R, Creelan B, Silberberg J, Costin D, Bharne A, Bonta I, Bosemani T, Nikolinakos P, Ross JS, Miller VA, Ali SM, Klemptner SJ, Ignatius Ou SH. Receptor tyrosine kinase fusions and BRAF kinase fusions are rare but actionable resistance mechanisms to EGFR tyrosine kinase inhibitors. *J Thorac Oncol*. 2018;13(9):1313–23.