www.nature.com/tp

CD4⁺ but not CD8⁺ T cells revert the impaired emotional behavior of immunocompromised *RAG-1*-deficient mice

L Rattazzi¹, G Piras¹, M Ono², R Deacon³, CM Pariante⁴ and F D'Acquisto¹

An imbalanced immune system has long been known to influence a variety of mood disorders including anxiety, obsessivecompulsive disorders and depression. In this study, we sought to model the impact of an immunocompromised state on these emotional behaviors using $RAG-1^{-/-}$ mice, which lack T and B cells. We also investigated the relative contribution of CD4⁺ or CD8⁺ T cells to these manifestations using $RAG-1^{-/-}$ /OT-II and $RAG-1^{-/-}$ /OT-I transgenic mice, respectively. Our results show that $RAG-1^{-/-}$ mice present a significant increase in digging and marble-burying activities compared with wildtype mice. Surprisingly, these anxiety-like behaviors were significantly reverted in $RAG-1^{-/-}$ /OT-II but not $RAG-1^{-/-}$ /OT-I transgenic mice. Immunodepletion experiments with anti-CD4 or anti-CD8 in C57/BL6 mice or repopulation studies in $RAG-1^{-/-}$ mice did not reproduce these findings. Microarray analysis of the brain of $RAG-1^{-/-}$ and $RAG-1^{-/-}$ /OT-II mice revealed a significantly different gene fingerprint, with the latter being more similar to wild-type mice than the former. Further analysis revealed nine main signaling pathways as being significantly modulated in $RAG-1^{-/-}$ compared with wild-type mice. Taken together, these results suggest that life-long rather than transient immunodeficient conditions influence the emotional behaviors in mice. Most interestingly, these effects seem to correlate with a specific absence of CD4⁺ rather than CD8⁺ T cells. Validation of these findings in man might provide new clues on the mechanism by which early life immune modulation might impact mod response in adults and provide a further link between immune and emotional well-being.

Translational Psychiatry (2013) 3, e280; doi:10.1038/tp.2013.54; published online 9 July 2013

Introduction

A correlation between mental diseases and immune dysfunction has been reported and debated in the literature since the late 1980s.¹ Indeed, direct and indirect evidences in both human and animal experimental systems indicate that changes in the immune repertoire significantly influence cognitive functions^{2,3} and neurodegeneration.^{4–6} More recent studies also suggest that a healthy immune system is a prerequisite for a balanced and functional emotional response.^{7–10}

The link between emotion and immunity has been documented in a variety of studies addressing psychosocial changes in patients treated with immunosuppressive drugs. Cyclosporine, a drug widely used in organ transplantation, has been shown to induce a range of neuropsychological problems ranging from depression to anxiety.^{11–15} Similarly, other studies described psychological side effects like anxiety, depression and obsessive-compulsive disorders in patients treated with a variety of structurally unrelated immunosuppressant including methotrexate,¹⁶ azathioprine,¹⁷ and chemotherapy.¹⁸

The recombination-activating gene *RAG-1* encodes proteins necessary for immunoglobulin and T-cell receptor gene recombination. *RAG-1*-deficient mice have small lymphoid organs that do not contain mature B and T lymphocytes.¹⁹ Seminal work on $RAG-1^{-/-}$ mice by Cushman and co-workers²⁰ reported an increased locomotor activity, reduced levels of fearfulness and no differences in spatial learning and memory. However, given the shared expression of the *RAG-1* protein by lymphocytes and central nervous system tissues, the authors concluded their seminal paper stating: 'Whether these changes are due to the loss of *RAG-1* gene expression in the brain, the result of the absence of the *RAG-1* gene in the immune system or some combination of both effects remains to be determined in future research'.

To address this question, McGowan *et al.*²¹ used a very elegant approach to assess the function of *RAG-1* in the central nervous system and dissect it from the lack of T and B cells in the periphery.²¹ The authors compared the behavior of *RAG-1* to *RAG-2*-deficient mice and found an impaired social recognition memory in the first but not the latter. Because both lines are immunodeficient and *RAG-2* is not expressed in the brain, the authors claimed a specific function of *RAG-1* in controlling memory formation.

In this study, we expanded on these notions and explored first if the immunodeficient state of $RAG-1^{-/-}$ mice had any

E-mail: F.Dacquisto@qmul.ac.uk

Keywords: CD4 T cells; CD8 T cells; immunodeficient mice; OT-I and OT-II transgenic mice; RAG knockout mice Received 3 Decemeber 2012; revised 8 April 2013; accepted 23 May 2013 npg

¹Centre for Biochemical Pharmacology, The William Harvey Research Institute, Barts and the London School of Medicine and Dentistry, Queen Mary University of London, London, UK; ²Institute of Child Health, University College London, London, UK; ³Department of Experimental Psychology, University of Oxford, Oxford, UK and ⁴Department of Psychological Medicine, Institute of Psychiatry, King's College London, London, UK

Correspondence: Dr F D'Acquisto, Centre for Biochemical Pharmacology, The William Harvey Research Institute, Barts and the London School of Medicine, Queen Mary University of London, Charterhouse Square, London EC1M 6BQ, UK.

influence on their emotional behavior and second the specific contribution of CD4⁺ or CD8⁺ T cells to these changes using *RAG-1^{-/-}/*OT-II and the *RAG-1^{-/-}/*OT-I transgenic lines, respectively. Second, we explored possible changes in mood-modifying circulating factors or gross brain structural differences in these mice. The results obtained suggest that CD4⁺ but not CD8⁺ T cells are capable of partially reverting anxiety-like behavior and lack of self-care characteristic of immunodeficient mice. Interestingly, these behavioral changes were mirrored in specific brain gene fingerprint. These unexpected new findings might provide a mechanistic explanation for the increased emotional distress observed in patients suffering from a wide variety of immune disorders.

Materials and methods

Mice. We used 5-week-old male mice for all the experiments. Mice were housed in groups of six per cage under specific pathogen-free conditions and with free access to food and water. Mice were housed for at least 7 days before testing. Wild-type (WT) C57BL/6 mice purchased from Charles River (Manston, UK) or RAG-1+/+ littermate were used as control. B6.SJL-Ptprc^a Pepc^b/BoyJ-Tg(TcraTcrb)1100Mjb/J-B6.129S7-*Rag1^{im1Mom}* (RAG^{-/-}/ OT-I) and B6.SJL-Ptprc^a Pepc^b/BoyJ-Tg(TcraTcrb)425Cbn/ J-B6.129S7-Rag1^{tm1Mom} (RAG^{-/-}/OT-II) mice were kindly provided by Professor Hans Stauss (University College London, London, UK) and bred in our animal facility. Apart from the nest construction test, all experiments were performed during the light phase of the light-dark cycle, and no more than two tests per day were performed. All tests were conducted in a blinded manner and according to the UK Animals (Scientific Procedures) Act, 1986.

Flow cytometric analysis. Thymocytes and lymphocytes were stained in 100 μ l of fluorescence-activated cell sorting buffer (phosphate-buffered saline containing 5% fetal calf serum and 0.02% of NaN₂). The antibodies used were anti-CD3 phycoerythrin (clone 145-2C11), anti-CD4 fluorescein isothiocyanate (clone GK 1.5), anti-CD8 Cy5 (clone 53-6.7) (all from eBioscience, San Diego, CA, USA). Cells were labeled with the appropriate concentration of conjugated antibodies for 1 h at 4 °C as described previously.²² After labeling, cells were washed and analyzed. In all experiments, stained cells were acquired with FACScalibur flow cytometer and analyzed using the FlowJoTM software (Tree Star, Ashland, OR, USA, Oregon Corporation).

In vivo **T-cell depletion.** Male C57/BL6 mice (6 weeks old) received an intraperitoneal injection of anti-CD4 (250 μ g; clone GK1.5; BioLegend) or anti-CD8 (250 μ g; clone 53-6.7; BioLegend, San Diego, CA, USA) or immunoglobulin G control. T-cell immunodepletion was verified by staining peripheral blood mononuclear cells at different time points (Day 2, Day 5 and Day 7) after the treatment. Briefly, blood samples were collected by intracardiac puncture in syringes containing sodium citrate 3.2% (wv⁻¹). The cells were centrifuged to pellet at 300 *g* and resuspended in fluorescence-activated cell sorting buffer containing 1:500 Fc blocking (anti-mouse CD16/32) and stained with anti-CD4

or anti-CD8. Red blood cells were lysed with RBC Lysis Buffer according to the manufacturer's instruction (eBioscience).

RAG1^{-/-} repopulation studies. Purified CD4 or CD8 T cells were obtained from male C57/BL6 mice (6 weeks old) by negative selection following the manufacturer's instructions (Dynabeads[®] Untouched[™] Mouse CD8 Cells and Dynabeads[®] Untouched[™] Mouse CD4 Cells; Invitrogen, Invitrogen Life Technologies Ltd, Paisley, UK). Purity was tested by fluorescence-activated cell sorter and was >98%. Cells were resuspended in phosphate-buffered saline (2 × 10⁶/300 l) and transferred into male *RAG1*^{-/-} mice (6 weeks old) by intraperitoneal injection.

Digging and marble-burying tests. Marble-burying and digging tests were carried out as described previously²³ with some modifications. Briefly, mice were individually placed in a clear plastic box ($14 \text{ cm} \times 10 \text{ cm} \times 11 \text{ cm}$) filled with approximately 5-cm-deep wood chip bedding lightly pressed to give a flat surface. The same bedding substrate was used for all the mice and flattened after each test. Fifteen glass marbles were placed on the surface in five rows of three marbles each. The latency to start digging, the number of digging bouts and the number of buried marbles (to 2/3 their depth) were recorded during the 15-min test. Two trials were performed, the second trial taking place 24 h after the first trial.

Open field activity test. The open filed test was performed as described previously with some modifications.²⁴ The open field consisted of a white PVC arena ($50 \text{ cm} \times 30 \text{ cm}$) divided into $10 \text{ cm} \times 10 \text{ cm}$ squares (n = 15). Mice were brought into the experimental room 15 min before testing. Each mouse was placed in one of the corner squares facing the wall, observed and recorded for 5 min. The total number of squares crossed, latency to the first rear and the total number of rears were recorded. After each test, the arena was cleaned with water to attenuate and homogenize olfactory traces. Two trials were performed, the second trial taking place 24 h after completion of the first trial.

Nest construction test. The nest construction test was performed as described previously.²⁵ Mice were transferred into individual cages 1 h before the dark phase (1700 hours) and individually housed overnight. The results were assessed the next morning. Food, water and wood chip bedding were provided. No other environmental enrichment was added. One 2–3 g, $5 \text{ cm} \times 5 \text{ cm}$ pressed cotton square (nestlet; Ancare, Ancare Bellmore, NY, USA) was placed in each cage. The weight of nesting material shredded was calculated by weighing the nestlet before and after the overnight test. The quality of the nest was evaluated on a five-point scale as detailed in Deacon.²⁵

Plasma corticosterone and cytokine measurement. Blood was collected by intracardiac puncture performed under anesthesia. Serum was obtained from the clotted blood by centrifugation (8000 r.p.m., 5 min) and stored at -80 °C before the assay. Corticosterone concentrations were measured in diluted (1:32) plasma by EIA assay following the manufacturer's instructions (Enzo Life Sciences, Exeter, UK). Cytokine levels in the same samples were measured (dil. 1:100) using mouse T-helper type 1 (Th1)/Th2/Th17/Th22 16plex Kit Flowcytomix and according to the manufacturer's instructions (eBioscience).

Histology. Brains were collected either before or at the end of experiments and fixed in 4% paraformaldehyde for 72 h. Thereafter, tissues were sectioned on a sagittal or coronal plane and embedded in paraffin by our in-house histology facility. Sections (5 μ m) were deparaffinized and stained with hematoxylin and eosin. In all cases, a minimum of three sections per animal was evaluated. Phase-contrast digital images were taken using the Image Image-Pro (Media Cybernetics, Rockville, MD, USA) analysis software package.

Microarray analysis. Total RNA was extracted from brains of WT (n=3), $RAG^{-/-}$ (n=2) and OT-II/ $RAG^{-/-}$ (n=2) mice using RNeasy[®] Microarray Tissue Mini Kit (Qiagen[®], West Sussex, UK). Total RNA was hybridized to Affymetrix Mouse Gene 1.0 ST array chips at UCL Genomics (London, UK) with standard Affymetrix protocols, using GeneChip Fluidics Station 450, and scanned using the Affymetrix GeneChip Scanner (Affvmetrix, Santa Clara, CA, USA), Data were normalized by rma of the Bioconductor package, affy. Differentially expressed genes were identified by the Bioconductor package, limma, considering the false discovery rate (adjusted *P*-value < 0.05). The gene and sample scoring system was made by canonical correspondence analysis. Canonical correspondence analysis is a variant of correspondence analysis, where the main data are linearly regressed onto explanatory variables (environmental variables), and subsequently the regressed data are analyzed by correspondence analysis. In this study, we regressed the whole data set onto an explanatory variable, which was defined as the difference between 'average' WT and 'average' $RAG-1^{-/-}$. Detailed methodology is described elsewhere.²⁶ Signaling pathway impact analysis was performed using the Bioconductor package, SPIA, by comparing WT and $RAG-1^{-/-}$.

Real-time polymerase chain reaction. Total RNA was extracted from whole brains (n = 6 for each mouse line) with RNeasy[®] Microarray Tissue Mini Kit (Qiagen[®]) according to the manufacturer's protocol and reverse transcribed using 2 µg oligo(dT)15 primer, 10 U AMV reverse transcriptase, 40 U RNase inhibitor (all from Promega Corporation, Madison, WI, USA) and 1.25 mm each dNTP (Bioline, London, UK) for 45 min at 42 °C. Real-time polymerase chain reaction was carried out by using ABsoluteTM QPCR ROX Mix (Thermo Scientific, Epsom, UK) and fluorescent QuantiTect primers. Cycling conditions were set according to the manufacturer's instructions. Sequence-specific fluorescent signal was detected by 7900HT Fast Real-Time PCR System (Applied Biosystems, Warrington, Cheshire, UK). mRNA data were normalized relative to glyceraldehyde 3-phosphate dehydrogenase and then used to calculate expression levels. We used the comparative Ct method to measure the gene transcription in samples. The results are expressed as relative units based on calculation of $2^{-\Delta\Delta Ct}$, which gives the relative amount of gene normalized to endogenous control (glyceraldehyde 3-phosphate dehydrogenase) and to the sample with the lowest expression set as 1.

Data analysis. Initially, we determined if the data distribution was parametric. Pairwise comparisons were made by *t*test and the results expressed as mean \pm s.e.m. For nonparametric data, the Mann–Whitney *U*-test was applied, and results were expressed as medians (interquartile range).

Results

Immune repertoire of RAG-1^{-/-}, RAG-1^{-/-}/OT-I and RAG-1-/-/OT-II. Crossing OT-I and OT-II TCR transgenic mice onto $RAG-1^{-/-}$ background generates mice with a single population of mature CD8⁺ or CD4⁺ T cells. Figure 1 shows a typical immature and mature T-cell profiles of , RAG-1^{-/-}/OT-I and RAG-1^{-/-}/OT-II compared RAG-1-/with WΤ C57BL/6 control mice. Control mice show a typical profile with a 1:2 ratio of CD8⁺ and CD4⁺ single-positive T cells in the thymus as well as in the periphery (Figure 1, first top and bottom panels, respectively). As expected, the majority of $RAG-1^{-/-}$ thymocytes are CD4 and CD8 double-negative cells and have no mature CD4 or CD8 single-positive \tilde{T} cells in the periphery (Figure 1, second top and bottom panels, respectively).¹⁹ The presence of OT-I and OT-II TCR transgene overcomes the block at the stage of double-negative 3 (DN3) of RAG-1^{-/-} thymocytes and allows the generation of a peripheral T-cell immune repertoire constituted by 72% of CD8⁺ in RAG-1^{-/-}/OT-I and 65% of CD4⁺ T cells RAG-1^{-/-}/OT-II mice (Figure 1, third top and bottom panels).27

Increased digging and marble-burying behavior of *RAG-1*^{-/-} mice. We first investigated differences in anxiety- and obsessive-compulsive-like behavior in WT and *RAG-1*^{-/-} mice using the digging and marble-burying tests. As shown in Figure 2a, *RAG-1*^{-/-} mice presented a significant increase (two- to threefold) in the number of digging bouts and buried marble compared with WT mice. The latency to dig was higher in control mice compared with *RAG-1*^{-/-} mice, although the difference was not significant.

To further corroborate the anxiety-like behavior and simultaneously rule out possible intrinsic impairment in locomotor activity, we used the open field activity test. Here, we considered a number of parameters including exploration (number of rears and latency) and locomotor activity (number of squares crossed). Although $RAG-1^{-/-}$ mice showed a slight reduction compared with the mice in all the parameters observed, these differences were not statistically significant (Figure 2b).

CD4⁺ but not CD8⁺ T cells revert the increased digging and marble-burying behavior of $RAG-1^{-/-}$ mice. We next tested the hypothesis that the presence of CD4⁺ or CD8⁺ T cells might influence the heightened digging and marble-burying behavior of the $RAG-1^{-/-}$ mice. $RAG-1^{-/-}$

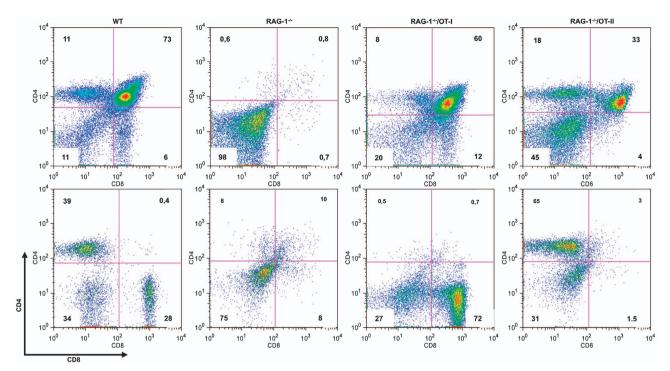


Figure 1 Immune repertoire of $RAG-1^{-/-}$, $RAG-1^{-/-}/OT-I$ and $RAG-1^{-/-}/OT-I$ mice. Thymocytes and lymphocytes from male wild-type (WT), $RAG-1^{-/-}$, $RAG-1^{-/-}/OT-I$ and $RAG-1^{-/-}/OT-I$

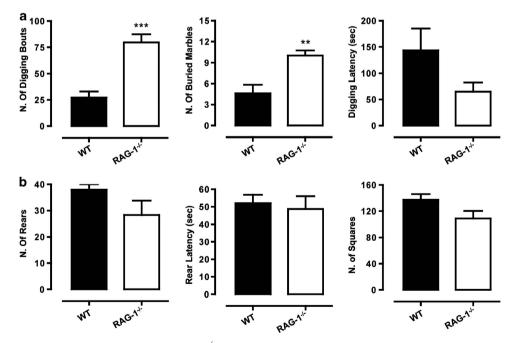


Figure 2 Increased digging and marble-burying behavior of $RAG-1^{-/-}$ mice (upper panel). The bar graphs in **a** shows the total number of digging bouts, buried marbles and the latency to dig (expressed in seconds) measured during the 15min marble-burying test. The bar graphs in **b** show the total number of rears, the latency to rear (expressed in seconds) and the total number of squares crossed assessed during the 5-min open field test. Values are expressed as mean ± s.e.m. of six mice and are representative of n = 3-4 separate experiments. **P < 0.01 and ***P < 0.001 indicate significant values compared with wild-type (WT) C57BL/6 control mice (Mann–Whitney *U*-test).

OT-I showed no difference in either number of bouts, buried marbles or latency compared with $RAG-1^{-/-}$ mice (Figure 3a). In contrast, $RAG-1^{-/-}$ /OT-II behaved differently

from $RAG-1^{-/-}$ mice, showing a significant reduction in the number of bouts (about 25%) and buried marbles (about 20%), and almost doubled latency time (P<0.05; Figure 3b).

PS

T cells and anxiety-related disorders L Rattazzi *et al*

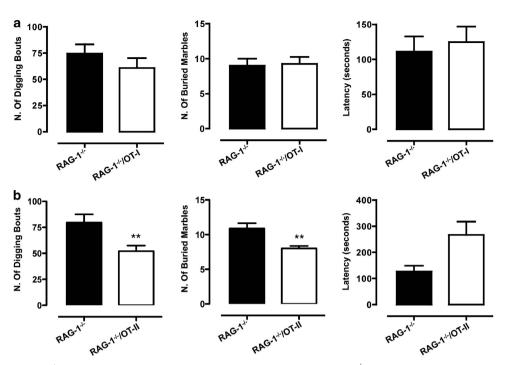


Figure 3 CD4⁺ but not CD8⁺ T cells revert the increased digging and marble-burying behavior of $RAG-1^{-/-}$ mice. The bar graphs show the total number of digging bouts, buried marbles and the latency to dig (expressed in seconds) in $RAG-1^{-/-}$ /OT-I (a) or $RAG-1^{-/-}$ /OT-II (b) compared with $RAG-1^{-/-}$ during the 15-min marble-burying test. Values are expressed as mean \pm s.e.m. of six mice and are representative of n=3-4 separate experiments. **P<0.01 indicates significant values compared with $RAG-1^{-/-}$ mice (Mann–Whitney U-test).

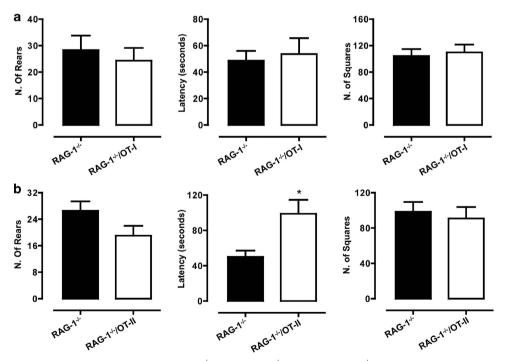


Figure 4 No differences in the open field activity between $RAG-1^{-/-}/OT-I$, $RAG-1^{-/-}/OT-I$ and $RAG-1^{-/-}$. The bar graphs show the total number of rears, the latency to rear (expressed in seconds) and squares crossed in $RAG-1^{-/-}/OT-I$ (a) or $RAG-1^{-/-}/OT-I$ (b) compared with $RAG-1^{-/-}$ during the 5-min open field test. Values are expressed as mean ± s.e.m. of six mice and are representative of n = 3-4 separate experiments. *P < 0.05 indicates significant values compared with $RAG-1^{-/-}$ mice (Mann–Whitney *U*-test).

With the open field test, we observed no difference in the behavior of $RAG-1^{-/-}/OT-I$ and $RAG-1^{-/-}$ (Figure 4a). Similarly, $RAG-1^{-/-}/OT-II$ showed no significant difference in either

the number of rears or of squares crossed as compared with $RAG-1^{-/-}$, except for an increase in the latency, which this time reached a statistical significance (Figure 4b). When we

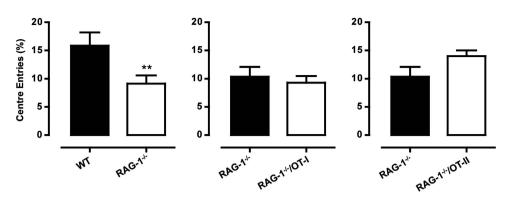


Figure 5 CD4⁺ but not CD8⁺ T cells might revert the decreased number of center entries showed by $RAG-1^{-/-}$ mice. The bar graphs show the comparison of the entries into the center between $RAG-1^{-/-}$ and wild-type mice (left panel), $RAG-1^{-/-}$ and $RAG^{-/-}$ /OT-I (middle panel) and $RAG^{-/-}$ and $RAG^{-/-}$ /OT-I (right panel) during the 5-min open field test. Values are expressed as mean ± s.e.m. of six mice and are representative of n = 3-4 separate experiments. **P < 0.01 indicates significant values compared with wild-type C57BL/6 control mice (Mann–Whitney *U*-test).

compared the number of center entries (considered an anxiety-like behavior parameter) in the three lines, we observed a significant reduction in $RAG-1^{-/-}$ mice compared with WT mice. However, this difference was not significantly reverted in either $RAG-1^{-/-}/OT-I$ or $RAG-1^{-/-}/OT-II$ (Figure 5).

Transient depletion of CD4⁺ or CD8⁺ T cells does not affect the emotional behavior of C57/BL6 mice. We next wondered if we could reproduce the results obtained with $RAG-1^{-/-}$ /OT-I and $RAG-1^{-/-}$ /OT-II using anti-CD4- or anti-CD8-depleting antibodies in C57/BL6 mice. As shown in Figure 6, neither anti-CD4 nor anti-CD8 antibodies significantly modified the digging, or marble-burying activities or the latency to digging (top, middle and bottom panels, respectively) compared with control IgG-treated mice at the indicated times.

To further confirm these results, we sought to investigate if the repopulation of $RAG-1^{-/-}$ with T cells would rescue the increased anxiety observed in these mice. Consistent with previous results, reconstitution of $RAG-1^{-/-}$ mice with purified CD4 or CD8 T cells did not affect the increase in the number of digging bouts compared with vehicle phosphatebuffered saline-injected mice (Figure 7).

CD4⁺ **T** cells revert the impaired nest construction of **RAG-1**^{-/-} mice. To explore the impact of T cells on other emotional behavior, we tested the three transgenic lines for their nesting activity, a standard test for measuring activities of daily living. Figure 8a shows representative pictures of the results obtained, whereas in Figure 8b, we report the quantitative results. Measurement of nestlet shredding showed a decreased ability of $RAG-1^{-/-}$ mice to perform this task compared with WT mice (Figure 8b). Similar to previous analysis, this impaired behavior was significantly reverted in $RAG-1^{-/-}$ /OT-II but not $RAG-1^{-/-}$ /OT-I mice (Figure 8b). Comparable results were obtained scoring nest quality (Supplementary Figure 1).

No differences in systemic or gross brain structure between $RAG-1^{-\prime-}$, $RAG-1^{-\prime-}$ /OT-I and $RAG-1^{-\prime-}$ /OT-II mice. We next investigated whether the behavioral changes

Translational Psychiatry

observed were due to changes in known behavioral modulating factors such as corticosterone. As shown in Figure 9a, there were no significant differences in the levels of circulating corticosterone between WT and $RAG-1^{-/-}$ mice or between $RAG-1^{-/-}$ and $RAG-1^{-/-}/OT-I$ and $RAG-1^{-/-}/OT-I$ and $RAG-1^{-/-}/OT-I$ mice.

Cytokines can induce behavioral changes (also known as sickness behavior²⁸), a consequence of their modulatory effects on brain function. When we scanned the same samples for classical Th1, Th2, Th17 and ThGM-CSF cytokines, only interleukin (IL)-17, IL-18 and interferon- γ could be detected. However, none of these mediators was differentially modulated in *RAG-1^{-/-}* and *RAG-1^{-/-}*/OT-I and *RAG-1^{-/-}*/OT-II mice compared with WT mice (Figure 9b), excluding the possibility that cytokines released by T cells or a latent state of infection as being responsible for the behavioral changes observed.

Similarly, analysis of general brain morphology and architecture of WT, $RAG-1^{-/-}$, $RAG-1^{-/-}/OT-I$ and $RAG-1^{-/-}/OT-I$ oT-II mice showed comparable hematoxylin–eosin (Figures 10a–d, respectively) or luxol fast blue staining (data not shown) ruling out any contribution of infiltrated immune cells or general neuronal defect in the altered emotional behavior of tested mice.

Brain gene fingerprint of *RAG-1*^{-/-} **and** *RAG-1*^{-/-} **/OT-II.** To unveil cellular and molecular mechanisms potentially responsible for the observed changes in emotional behavior, we took an unbiased approach and compared whole brain gene fingerprint of WT, *RAG-1*^{-/-}, *RAG-1*^{-/-}/OT-II mice. The flowchart in Figure 11a summarizes the results of this analysis. From the 34760 probes present in the chip, 6635 were significantly modulated (*P*<0.05). This corresponded to 782 differentially expressed genes with a fold change (FC) value < -1 or >1, and 111 of them were annotated genes (genes with Entrez ID.)

Hierarchical clustering and heatmap analysis of these selected 111 genes and brain samples (Figure 11b and Table 1) showed that $RAG-1^{-/-}$ mice showed a distinct pattern of gene expression compared with $RAG-1^{-/-}$ /OT-II and WT mice. This result also suggested that $RAG-1^{-/-}$ /OT-II was similar to WT at the transcriptomic level. To further

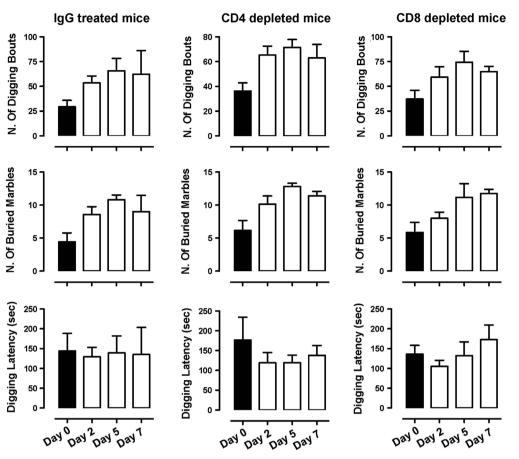


Figure 6 Depletion of CD4 or CD8 T cells does not induce anxiety-like behavior in C57/BL6 mice. C57/BL6 mice received an intraperitoneal injection of anti-CD4 (250 µg), anti-CD8- (250 µg) depleting antibodies or IgG control, and then tested in the digging and marble-burying test. The bar graphs show the total number of digging bouts, buried marbles and the latency to dig (expressed in seconds) in mice treated as indicated and assessed before the treatment (day 0) or after 2, 5 and 7 days (day 2, day 5 and day 7, respectively). Values are expressed as mean ± s.e.m. of 6–8 mice.

determine whether and how much $RAG-1^{-/-}/OT$ -II was more similar to WT than $RAG-1^{-/-}$, we analyzed the similarities between the samples based on the gene expression pattern, focusing on the difference between WT and $RAG-1^{-/-}$, which is our major interest in this study (see Materials and methods). The similarity analysis using these 111 genes showed that $RAG-1^{-/-}/OT$ -II was far more similar to WT than $RAG-1^{-/-}$ (Figure 11c). Importantly, the result of this similarity analysis was very similar using all the differentially expressed genes (data not shown), indicating that the result is robust and not dependent on the selection of genes by FC and annotation. These results were compatible with those of the behavioral analysis.

Pathway analysis provided further clues on the main differences between WT and $RAG-1^{-/-}$. Using a third-generation pathway analysis approach,²⁹ nine pathways were identified as been significantly modulated in $RAG-1^{-/-}$ compared with WT: two being activated (Parkinson's disease and RNA transport) and seven inhibited (Huntington's diseases, Alzheimer's disease, extracellular matrix-receptor interaction, olfactory transduction, focal adhesion, calcium signaling and small-cell lung cancer; summarized in Table 2 and reported singularly in Supplementary Figures 2–10).

Discussion

The idea that a balanced mental state is a directly associated with general well-being can be traced back to the time of Decimus lunius luvenalis. He was the first to state that a healthy mind is found in a healthy body (*'mens sana in corpore sano'*). However, evidences gathered in our modern time suggests also the reverse, that is, that a *corpore sano*, and in particular a healthy immune system, might contribute to our mental well-being. In this study, we sought to provide direct experimental evidence of this hypothesis investigating first the emotional behavior of mice genetically void of T and B cells (the *RAG-1^{-/-}* mice) and secondly assessing the specific contribution of CD4⁺ or CD8⁺ T cells.

Our results show a significant increase in anxiety-like behavior in RAG-1^{-/-} mice as evaluated by the number of center entries in the open field test as well as the increased digging and marble-burying activities. Most interestingly, we also observed that CD4⁺ but not CD8⁺ T cells are able to revert significantly the exaggerated emotional response of RAG-1^{-/-} mice. These results were not due to T-cell activation upon exposure of the mice to the behavioral paradigms (Supplementary Figure 11). The notion that CD4⁺ T cells has a preferential role as 'mood stabilizer'

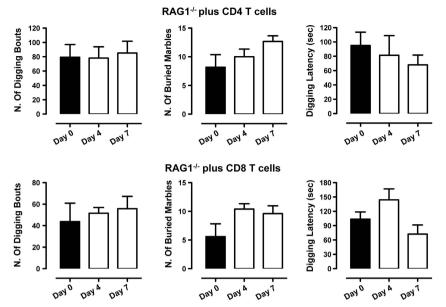


Figure 7 Repopulation of $RAG1^{-/-}$ mice with CD4 or CD8 T cells does not affect their anxiety-like behavior. $RAG1^{-/-}$ mice received an intraperitoneal injection of purified CD4 (2 × 10⁶) or CD8 (2 × 10⁶) T cells and then tested in the digging and marble-burying test. The bar graphs show the total number of digging bouts, buried marbles and the latency to dig (expressed in seconds) in mice assessed before the cell transfer (day 0) or after 4 and 7 days (day 4 and day 7, respectively). Values are expressed as mean ± s.e.m. of 6–8 mice.

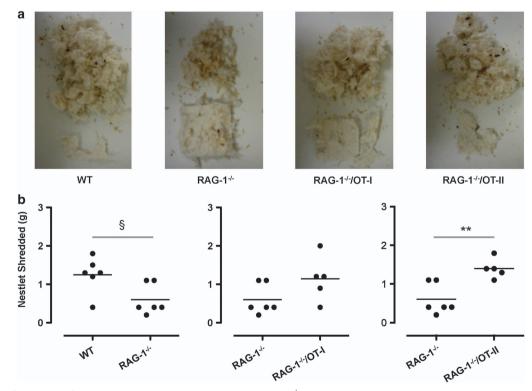


Figure 8 CD4⁺ but not CD8⁺ T cells revert the impaired nest construction of $RAG-1^{-/-}$ mice. (a) Representative pictures of the nestlet shredding activity of wild-type (WT), $RAG-1^{-/-}$, $RAG-1^{-/-}$ /OT-I and $RAG-1^{-/-}$ /OT-I during an overnight test. (b) Quantitative analysis of nestlet shredding activity expressed as grams of nestlet shredded after an overnight test. Values are expressed as mean ± s.e.m. of six mice and are representative of n=3-4 separate experiments. [§]P<0.05 and ^{**}P<0.01 indicate significant values compared with WT C57BL/6 control and $RAG^{-/-}$ mice, respectively (Mann–Whitney *U*-test).

compared with CD8 $^+$ T cells has long been suggested. Early studies on adult patients suffering from anxiety and obsessive-compulsive disorders have demonstrated immunological

alterations including a significant increase of CD8⁺ and decrease of CD4⁺ lymphocytes compared with the healthy control group.³⁰ The same abnormality has been observed in

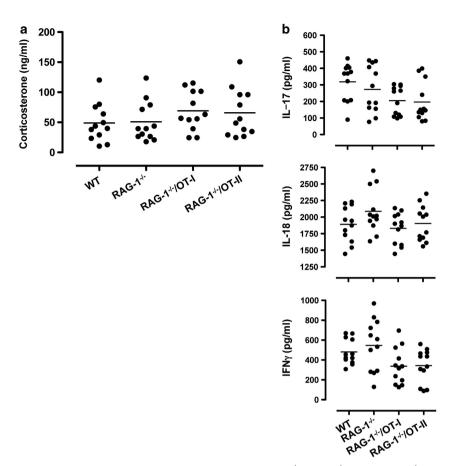


Figure 9 No differences in corticosterone or inflammatory cytokine serum levels between $RAG-1^{-/-}$, $RAG-1^{-/-}$ /OT-I and $RAG-1^{-/-}$ /OT-II mice. Levels of corticosterone (a) or interleukin (IL)-17, IL-18 and interferon (IFN)- γ (b) in the serum of wild-type (WT), $RAG-1^{-/-}$, $RAG-1^{-/-}$ /OT-I and $RAG-1^{-/-}$ /OT-II. Values are expressed as ng ml⁻¹ or as pg ml⁻¹ and are cumulative of n = 2-3 experiments.

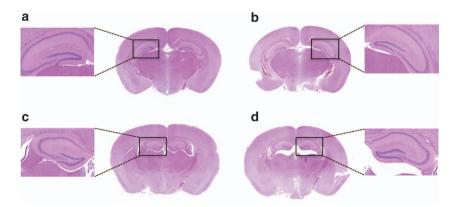


Figure 10 No differences in gross brain structure between $RAG-1^{-/-}$, $RAG-1^{-/-}/OT-I$ and $RAG-1^{-/-}/OT-II$ mice. The pictures show the coronal hematoxylin and eosin-stained sections of brain from wild-type (**a**), $RAG-1^{-/-}$ (**b**), $RAG-1^{-/-}/OT-I$ (**c**) and $RAG-1^{-/-}/OT-II$ (**d**). The higher magnification represents the hippocampal area. The figures are representative of n = 3-4 mice.

patients with autism, a disorder characterized also by obsessive-compulsive symptoms and anxiety disorders.³¹ A recent study has also demonstrated in a quantitative manner an inverse relationship between CD4 count and hospital-associated anxiety and depression.³² Finally, in one of his recent review AH Miller³³ provided a comprehensive account of the multiple links between CD4⁺ T cells and depression,

highlighting the importance of trafficking of T cells to the brain following stress as a way to reduce stress-induced anxiety-like behavior.³³ However, a direct functional association between T cells and altered emotional behavior is still missing.

One of the major drawbacks in the life of patients suffering from anxiety-like behavior and/or obsessive-compulsive disorders is their inability to perform normal daily activities. npg

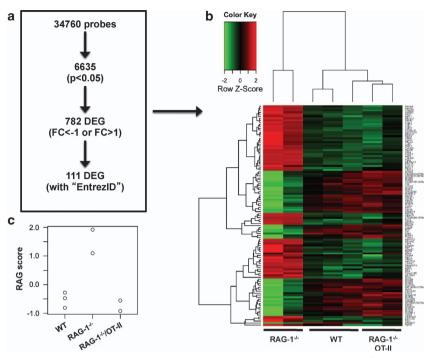


Figure 11 Heatmap and Canonical Correspondence Analysis on Microarray data of brain from wild-type (WT), $RAG-1^{-/-}$ and $RAG-1^{-/-}$ OT-II mice. (a) Genes were filtered by a moderated t-statistics and fold change (FC). The heatmap analysis used annotated genes only (genes with EntrezID). (b) Hierarchical clustering and heatmap analysis of the filtered genes. $RAG-1^{-/-}$ samples showed a distinct cluster. (c) Similarity analysis for the features of $RAG-1^{-/-}$ in comparison with WT. Note that $RAG-1^{-/-}$ /OT-II mice had low scores and were equivalent to WT mice.

Insightful epidemiological studies suggest that this 'inability to cope' might be due to an emotional rather than a cognitive impairment.³⁴ Our results are in line with this hypothesis and show a significant reduction in self-caring of $RAG-1^{-/-}$ mice compared with WT mice, as evaluated by the nesting test. Although these results suggest a link between self-neglect, anxiety-like behavior and immune suppression, more studies are needed to corroborate this hypothesis. These could include burrowing and hoarding, two other methods to test and quantify experimentally daily living activities.³⁵

Nestlet shredding together with marble burying is also a reliable experimental model for obsessive-compulsive disorders and anxiety.³⁶ We could not observe a significant difference in nestlet shredding after 60 min (data not shown), whereas we found a dramatic reduction in nest construction after an overnight test. We are tempted to explain these findings with the fact that, contrary to innate cells, adaptive immune cells are more involved in chronic disease and hence they usually exert their functions over a long period of time. In a similar way, one might speculate that immunosuppressed patients might present more difficulties in coping with long-term illness while being able to face problems as they came in.

Textbook immunology reports CD4⁺ T cells as 'helper' cells because of their ability to modulate other cell functions, and these 'helping' properties may go beyond antigenpresenting cell-mediated immunity. Groundbreaking studies by Kipnis and Schwartz^{37–39} has put CD4⁺ T cells at the center stage of neuroimmunology.² Repopulation of *scid* mice with T cells from WT donors has been shown to improve significantly the impaired cognitive functions of these mice.⁴⁰ Most interestingly, circulating and patrolling CD4⁺ T cells have been reported to convey constantly protective signaling to the brain, thus contributing to what is now known as 'protective autoimmunity'.^{41–44} In light of this concept, antigen-specific T cells, like myelin basic protein T-cell clones,⁴⁵ circulate through the brain and sustain key neuronal processes and function such as neurogenesis, cognition and memory.³⁹ Considering that the OT-II TCR transgene recognizes a non-endogenous antigenic peptide like OVA_{322–339}, it is tempting to speculate that, at difference from cognition and memory, emotional behavior might require a less-stringent condition, that is, the simple presence of circulating CD4⁺ T cells. Indeed, their circulation through the brain or meningeal spaces, as suggested by Derecki *et al.*,⁴⁶ might be enough to restore the emotional impairment that flares in immunocompromised conditions.⁴⁶

Previous studies assessing the role of T cells on brain functions have used immunodepleting antibodies or cellular replacement in immunodeficient host like the scid mice.40 When we adopted similar approaches in our settings, we could not find any significant differences. Neither the depletion of T cells in C57/BL6 mice using anti-CD4 or anti-CD8 antibodies nor the repopulation of $RAG-1^{-/-}$ with purified CD4 or CD8 T cells had any effect on emotional behavior. The reason behind this apparent discrepancy might lay in the nature of the immunodeficient state that has been studied. In the *RAG-1^{-/-}* mice, we have investigated how the absence of immune cells from prenatal development to adulthood influences neuronal networks and emotional behavior, whereas the immunodepletion or repopulation experiments refer to transient conditions (up to 2 weeks in our tests). This is quite an important aspect when one considers that

Table 1	Top-ranked genes	(n = 111) for the features	s of <i>RAG-1^{-/-}</i> I	brain in comparison with wild-type
---------	------------------	----------------------------	-----------------------------------	------------------------------------

Gene symbol	Probe ID	EntrezID	RAG score (gene score)	FC (RAG-WT)	<i>FC</i> (RAG <i>-OTII)</i>	P <i>-value adj.</i> RAG <i>vs WT</i>	P <i>-value adj.</i> RAG <i>vs OTI</i>
Mageb16	1 054 5096	71967	- 2.040851531	- 1.382	-2.412	0.080489	0.022479
Vmn2r42	1 055 9883	22310	- 1.93209053	- 2.152	- 1.793	0.021505	0.020203
Mir300	10398412	723 833	- 1.88308172	- 1.677	- 2.085	0.017506	0.016937
9530091C08Rik	10586718	320 440	- 1.794710154	- 1.667	- 2.619	0.104888	0.032618
Phxr4	10 583 203	18689	- 1.697861707	- 1.052	- 2.198	0.236372	0.040404
Mir539	10 398 418	723917	- 1.68592755	- 1.867	- 1.625	0.032518	0.037203
A130014H13Rik	10 402 560	319630	- 1.666278107	- 1.767	- 2.305	0.029595	0.018413
Mir380	10 398 388	723 859	- 1.579352318	- 1.198	- 1.896	0.039846	0.018076
Mir487b	10 398 416	723 940	- 1.515646393	- 1.591	- 1.606	0.029403	0.025115
Gm9911	10578017	100101427	- 1.495726048	- 1.097	- 1.857	0.171881	0.045496
Tbrg3	10 430 929	21 378	- 1.480314578	- 1.246	- 1.955	0.059237	0.02181
Air323	10 398 390	723 839	- 1.459997834	- 1.189	- 1.849	0.084182	0.027581
Air680-2	10 602 221	751 551	- 1.436650249	- 1.111	-2.109	0.136616	0.028476
Air665	10 398 338	751 555	- 1.426506241	- 1.668	- 1.866	0.0336	0.023984
Mir376b	10 398 408	723 934	- 1.390698725	- 1.236	- 1.411	0.034845	0.023984
<rtap2-4< td=""><td>10390913</td><td>71 453</td><td>- 1.383924657</td><td>- 1.713</td><td>- 2.176</td><td>0.049788</td><td>0.026248</td></rtap2-4<>	10390913	71 453	- 1.383924657	- 1.713	- 2.176	0.049788	0.026248
-830001A07Rik	10 586 722	320 055	- 1.360180695	- 1.16	- 1.984	0.170697	0.043923
Mir382	10 398 420	723912	- 1.358674308	- 1.517	- 1.887	0.029403	0.01938
Nir154	10 398 428	387 172	- 1.302334846	- 1.148	- 1.479	0.022707	0.016937
Gm10048	10 529 953	625 026	- 1.295861539	- 1.173	- 1.861	0.06009	0.02181
Air329	10 398 392	723 842	- 1.295645605	- 1.373	- 1.719	0.043559	0.024869
Ephb1	10 596 115	270 190	- 1.28329288	- 1.144	- 1.534	0.044139	0.022479
D230041D01Rik	10 548 727	1 000 38615	- 1.249725567	- 1.103	- 1.502	0.050443	0.023984
Nir679	10 398 396	751 539	- 1.239666734	- 1.275	- 1.743	0.092767	0.038914
Nir344	10 564 235	723 931	- 1.227043517	- 1.01	- 1.991	0.109837	0.023058
1931406H21Rik	10413216	77 592	- 1.20459512	- 1.236	- 1.209	0.030727	0.027375
Mir9-1	10 493 191	387 133	- 1.188744017	- 1.132	- 1.618	0.04865	0.02181
Nir341	10398350	723 846	- 1.185006997	- 1.323	-2.14	0.109543	0.032187
Snord95	10 375 501	1 002 16540	- 1.140100065	- 1.165	- 1.229	0.049459	0.037703
Tra2a	10 544 638	101 214	- 1.085592515	- 1.154	- 1.518	0.05765	0.027615
Gm5887	10 542 834	545 893	- 1.068780678	- 1.213	- 1.068	0.023897	0.024869
Snord37	10 365 003	100217454	- 1.052485206	- 1.061	- 0.627	0.017506	0.02181
Nir543	10 398 400	723 881	- 1.046459099	- 1.036	- 1.118	0.021685	0.017462
Myh9	10 430 245	17886	- 0.951892542	- 1.048	- 0.884	0.021822	0.021872
l11ra1	10 504 106	16157	- 0.935378718	- 1.033	- 1.167	0.02919	0.021613
/wf	10541910	22371	-0.913653403	- 1.116	- 1.213	0.024473	0.019653
Malat1	10465244	72289	- 0.881007189	- 1.116	- 1.756	0.136615	0.04157
Celsr3	10 589 130	107 934	-0.876228519	- 1.171	- 1.127	0.02257	0.019787
Dock6	10591614	319899	-0.837490045	- 1.295	- 1.147	0.014287	0.016937
Dock6	10 591 630	319899	-0.808120768	- 1.142	- 0.959	0.021097	0.018349
Dock6	10591612	319 899	- 0.805918085	- 1.083	- 1.032	0.021505	0.017355
Trank1	10589761	320 429	- 0.803294001	- 1.008	- 1.337	0.062753	0.029244
Pkd1	10 442 495	18763	-0.798266404	- 1.173	- 1.073	0.020247	0.016937
Snhg11	10478073	319317	- 0.792852186	- 1.038	- 0.89	0.03061	0.03547
-n1	10 355 403	14268	- 0.75577708	- 1.188	- 1.149	0.018698	0.016937
Leng8	10549615	232 798	- 0.755496969	- 1.053	- 1.088	0.029574	0.024595
All2	10 432 298	381 022	- 0.679798747	- 1.088	- 0.886	0.017506	0.016937
Erdr1	10 608 711	170 942	- 0.516986696	- 1.069	- 0.546	0.019742	0.035677
Nkks	10 488 048	59 030	0.457287879	1.043	0.341	0.022707	0.18131
Dynlt1c	10 548 785	100 040 563	0.459997269	1.004	0.654	0.017506	0.01714
Rpl10a	10 355 173	19896	0.485867583	1.117	0.793	0.021854	0.029315
Vsa2	10 411 363	59 050	0.552510543	1.192	0.814	0.018273	0.020521
Atp6v1f	10 536 895	66 144	0.573664963	1.027	1.039	0.021685	0.018076
Rpsa	10 385 034	16785	0.573730493	1.054	0.675	0.017506	0.017355
Pirg1	10 492 757	53317	0.578165545	1.006	0.899	0.017506	0.016937
ligi Hsd17b10	10 602 592	15 108	0.594214684	1.012	0.98	0.021685	0.018382
Snrpd2	10 367 073	107 686	0.601279671	1.012	0.881	0.021505	0.019125
.y6c2	10 429 573	100 041 546	0.616556062	1.066	0.885	0.024294	0.027767
Bloc1s1	10 373 594	14 533	0.639650101	1.016	1.1	0.021993	0.017462
Rpp38	10 479 749	227 522	0.640345029	1.128	0.834	0.017506	0.018347
Vdr61	10 593 740	66317	0.668426554	1.012	1.07	0.022823	0.018547
2d53	10 501 063		0.668970684	1.139		0.022823	0.040581
7053 Tm4sf1	10 498 273	12508 17112	0.675247588	1.016	0.627 0.827	0.021505	
							0.016937
Rpl29	10 505 090	19944	0.686092788	1.1	1.086	0.030792	0.026878
dh5	10 496 475	11532	0.693727155	1.039	1.025	0.017802	0.016937
Ikain4	10 490 551	58237	0.698189047	1.015	1.213	0.021505	0.016937
Rpl11	10454097	67 025	0.70163422	1.163	1.003	0.039409	0.046635
ipl11	10517457	67 025	0.706880397	1.009	0.903	0.021685	0.020078
Idufa2	10 458 386	17991	0.706987075	1.075	1.157	0.017506	0.016937
Trim13	10415784	66 597	0.708945695	1.285	0.768	0.021505	0.034966
Rpl10a	10 443 360	19896	0.711172011	1.402	1.194	0.021505	0.018349
Tspan6	10 606 609	56 496	0.71734053	1.097	1.001	0.017506	0.016937

npg 11

Table 1 C	ontinued
-----------	----------

Gene symbol	Probe ID	EntrezID	RAG score (gene score)	<i>FC</i> (RAG-WT)	<i>FC</i> (RAG- <i>OTII)</i>	P <i>-value adj.</i> RAG <i>vs WT</i>	P <i>-value adj.</i> RAG <i>vs OTII</i>
Slc39a12	10 469 389	277 468	0.724078447	1.08	1.054	0.018698	0.016937
Txndc9	10396064	98 258	0.737153463	1.094	1.077	0.021505	0.017462
Kcnj13	10356403	100 040 591	0.746292444	1.237	1.062	0.017506	0.016937
S100a10	10 493 995	20194	0.749982194	1.295	1.167	0.017506	0.016937
Trappc1	10 377 508	245 828	0.753853858	1.109	1.257	0.022707	0.017462
Ly6c1	10 429 568	17 067	0.755009369	1.168	1.228	0.032378	0.025501
Slc19a3	10356145	80721	0.76123195	1.042	0.765	0.02554	0.037851
Bloc1s1	10457924	14 533	0.763087509	1.124	1.198	0.021505	0.016937
Tmem100	10 380 285	67 888	0.763927693	1.001	0.715	0.021589	0.026199
P2ry13	10498367	74 191	0.766062492	1.278	0.969	0.017506	0.017355
Hddc2	10 362 394	69 692	0.799560244	1.01	1.071	0.021505	0.016937
Sncg	10418921	20618	0.812109509	1.236	1.33	0.011381	0.012815
Churc1	10 396 694	211 151	0.830334656	1.017	1.336	0.037868	0.021562
Scrg1	10571865	20284	0.838580022	1.22	1.228	0.025882	0.022109
Rpl29	10 490 824	19944	0.8434953	1.052	1.155	0.039564	0.028208
Hsd11b1	10361234	15 483	0.850759562	1.097	1.145	0.017506	0.016937
Snrpd2	10 498 595	107 686	0.851183191	1.075	1.138	0.026256	0.021562
Pgcp	10 423 556	54 381	0.868297012	1.256	1.28	0.017506	0.016937
Tpmt	10409021	22 0 17	0.88208296	1.027	1.412	0.021822	0.016937
Ċd63	10367436	12512	0.888313271	1.17	1.461	0.022707	0.016937
5730469M10Rik	10419082	70 564	0.895272141	1.268	1.91	0.03237	0.017355
Ly86	10 404 606	17 084	0.897001413	1.274	1.271	0.021685	0.018349
Åkr1b10	10537157	67 861	0.902194098	1.175	1.305	0.021505	0.016937
Lgals1	10425161	16852	0.912182029	1.268	1.398	0.021505	0.016937
N6amt2	10 420 385	68 043	0.91317463	1.241	1.256	0.021685	0.018347
Arhgdib	10 548 892	11 857	0.929045264	1.003	1.328	0.058924	0.027787
Ly6a	10 429 564	110454	0.94058187	1.438	1.499	0.025555	0.021402
Fcer1g	10360070	14 127	0.963847748	1.2	1.306	0.055767	0.039132
Akr1c18	10 407 435	105 349	0.964325333	1.047	0.762	0.024294	0.036537
Serpinb9	10 404 429	20723	0.966649984	1.023	0.968	0.017506	0.016937
Rfc4	10 438 690	106 344	0.99077814	1.162	1.23	0.020247	0.016937
Serpinb1a	10 408 557	66 222	1.057850202	1.247	1.558	0.017506	0.013705
Gstk1	10537712	76 263	1.150030127	1.592	1.697	0.021505	0.016937
Rpl11	1 050 2745	67 025	1.167088017	1.307	1.558	0.037379	0.023823
Stxbp4	1 038 9795	20913	1.173500167	1.086	1.948	0.091593	0.023603
Rpl11	1 045 1301	67 025	1.243624445	1.26	1.699	0.053335	0.025053
Ppia	1 054 5337	268 373	1.52055618	1.462	1.698	0.034726	0.023458
Mela	1 058 2545	17 276	3.221879948	4.675	2.776	0.002256	0.016937
Rpl36	10378783	54217	3.792674002	2.358	3.245	0.109543	0.045167

Abbreviations: FC, fold change; WT, wild type.

The table shows the identity and statistical values of the top-ranked genes that were used for heatmap analysis (Figure 9b) and calculating the similarity score (*RAG* score) for the *RAG-1^{-/-}* phenotype compared with the wild-type one (Figure 9c). Genes are ordered according to their relative contributions to the *RAG* score (i.e. the association with *RAG-1^{-/-}* or WT; positive values indicate association with *RAG-1^{-/-}*, whereas negative ones indicate association with WT). See Figure 9a for how genes were selected.

Probe ID is affymetrix ID.

FC is logged value.

neuropsychiatric disorders have often been linked to problems occurring at the developmental stage.^{47–49} Inflammation and infection at pre- and perinatal stages have been proved to be as powerful as maternal stress and trauma in causing long-term consequences on neuronal development and mental health.^{50–52} These clinical evidences hold true in experimental settings. Both peri- and prenatal administration of immunomodulatory agents such as TLR3 ligands and viral mimic polyinosine-polycytidylic, or TLR4 ligand and bacterial surrogate lipopolysaccharide, induced the development of schizophrenia- and autism-related behavioral changes including decreased exploratory activity and social interaction as adults.^{53–58}

Previous studies have shown that, like in pregnancy,⁵⁹ CD4⁺ T cells with a skewed Th2 phenotype contribute to a controlled and trophic microenvironment in the brain upregulating neurotrophic factors such as glial-derived neurotrophic factor, brain-derived neurotrophic factor and insulin growth

factor 1, or suppressing inflammatory mediators like tumor necrosis factor- α and IL-6.^{46,60} Conversely, direct and indirect evidence have linked Th1 and Th17 cells to emotional disorders. T cells from individuals with generalized anxiety disorders show an enhanced capability to differentiate in Th17 cells.61,62 Experimental evidences also suggest that Th17 cells preferentially accumulate in the brain of mice subjected to chronic restrain stress, whereas mice deficient in the RORyT (transcription factor necessary for Th17 differentiation) exhibited resistance to learned helplessness.63 T-bet (transcription factor necessary for Th1 differentiation) knockout mice show significantly reduced depressive-like behaviors provoked by repeated restraint stress.⁶⁴ This is in line with clinical studies showing, for instance, the contribution of Th1 cytokines to the pathogenesis of neuropsychiatric manifestations of systemic lupus erythematosus.65 In light of these findings, we decided to investigate whether the absence or presence of circulating CD4⁺ T cells would impact neuronal

12

Name	<u>D</u> I	pSize	NDE	pNDE	tA	pPERT	рG	pGFdr	pGFWER	Status	KEGGLINK
Parkinson's disease	5012	117	62	4.21E – 19	4.097	0.2	3.78E – 18	4.73E – 16	4.73E 16	Activated	http://www.genome.jp/dbget.bin/show_pattwary? mmu05012 + 66725 + 22202 + 22195 + 57320 + 68943 + 13063 + 104130 + 17991 + 17992 + 17993 + 2268487 + 226646 + 527197 + 5230075 + 54405 + 565166 + 660108 + 66218 + 668495 + 67130 + 67130 + 67784 + 67273 + 68198 + 68349 + 69875 + 72900 + 66925 + 66944 + 66168 + 66694 + 66108 + 66594 + 66594 + 66594 + 66594 + 66594 + 66594 + 66594 + 66594 + 66173 + 11328 + 113858 + 12868 + 12868 + 12868 + 20463 + 66142 + 75483 + 11949 + 11950 + 11957 + 228033 + 228000 + 67726 + 677942 + 71679 + 11739 + 11740 + 22334 + 22335 + 140499 + 67728 + 34406 + 67748 + 67728 + 77678 + 11739 + 11740 + 22334 + 22335 + 140499 + 67728 + 24406 + 67728 + 75483 + 11740 + 22334 + 22335 + 140499 + 67728 + 75483 + 74406 + 67728 + 7768
Huntington's disease	5016	172	77	9.54E – 18	- 0.372	0.797	3.07E – 16	1.92E – 14	3.84E – 14	Inhibited	T13047 T17047 T17047 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T17097 T170
Alzheimer's disease	5010	164	70	5.83E – 15	- 1.6765	0.558	1.12E – 13	4.66E – 12	1.40E – 11	Inhibited	21/80+12/20/+381917+68922+12913 http://www.genome.jp/dbget-in/show_pathway?mmu05010+66340+11487+130634+ 104130+17991+17992+17993+2256846+227197+230075+407785+ 54405+595136+66046+66108+66218+66495+67130+67184+67264+67273+ 68198+68349+69875+72900+66922+66945+66152+66576+66594+67733+ 67003+67530+11949+11956+11957+228033+28090+67126+67342+71679+ 110223+12858+12865+12865+12865+12865+12869+12869+12869+20463+66142+75483+ 20192+16438+16439+18798+14810+14811+14812+12289+15299+15108+
ECM-receptor interaction	4512	84	28 0	0.000175059	- 13.7865	5.00E - 06	1.91E – 08	5.98E – 07	2.39E – 06	Inhibited	14/102-118/125+7/8943+11816+19009+169/1+14433+2/34664 http://www.genome.jp/dbget-bin/show_pathway?mmu04512+16772+16774+16775+ 16776+16779-256519+22928+12814+12825+12826+12827+12830+12833+ 16835+16842+12843+245026 #1877+22371+20750+14268+21825+15529+
RNA transport	3013	161	56	2.67E – 08	0.7715	0.271	1.43E – 07	3.57E – 06	1.79E - 05	Activated	209/11 + 103304 1 0400 + 109039 + 10003 http://www.genome.jpd/0961451000 53975 + 69731 + 237221 + 20901 + 66069 + 60365 + 192170 + 56215 + 56009 + 237082 + 71805 + 114671 + 408191 + 72134 + 69912 + 54563 + 77595 + 107939 + 227720 + 20610 + 22218 + 13827 + 97112 + 66235 + 68969 + 53356 + 54709 + 56347 + 68135 + 78655 + 70047 + 10514 + 117109 + 208366 + 227522 + 54364 + 66161 + 60533 + 69961 + 386612 + 66231 + 26905 + 67204 + 218693 + 13682 +
Small-cell lung cancer	5222	86	22 0	0.026137813 - 2	- 20.07421825	5.00E - 06	2.20E - 06	4.59E - 05	0.000275271	Inhibited	218268+230861+21/869+10806/+1366/+209394+69482 http://www.genome.jp/dbget-bin/show_pathwa??mmu05222+16400+56469+218772+ 17187+19211+30955+13063+12826+12827+12820+12820+16772+16774+
Olfactory transduction	4740	989	40	-	- 23.1386	5.00E - 06	6.60E - 05	0.001179114	0.008253795	Inhibited	10/13-110/04 10/19 + ZE051 2-23454 11/37 1-225 + L5207 -12571 1534 454 11/57 41/52
Focal adhesion	4510	199	41 0	0.095748092 - 2	- 20.21166302	0.001	0.000981781	0.01486948	0.122722602	Inhibited	33.3529 + 19092 http://www.jenome.jp/dbget-bin/show_pathway?mmu04510 + 16400 + 192176 + 286940 + 67268 + 667938 + 19211 + 30955 + 21894 + 70549 + 12814 + 12825 + 12826 + 12827 + 12830 + 12833 + 12835 + 12842 + 12843 + 14268 + 16772 + 16775 + 16775 + 16776 + 16779 + 19699 + 20750 + 21825 + 22371 + 226519 + 23928 + 245026 + 81877 + 12389 +
Calcium signal- ing pathway	4020	179	47 0	0.000966948	- 5.8395	0.109	0.001070603	0.01486948	0.133825321 Inhibited		$\begin{array}{l} 12300 + 11797 + 12445 + 16001 + 18596 + 107746 + 109905 + 57257 \\ http://www.genome.jp/dbget-bin/show_pathway?mmu04020 + 320404 + 432530 + 18125 \\ + 22334 + 22334 + 108091 + 105641 + 19059 + 12494 + 13869 + 18679 + 14810 + 14811 \\ + 18438 + 228139 + 18803 + 18802 + 16438 + 16439 + 102093 + 18679 + 18682 + 68961 \\ + 20190 + 20191 + 20192 + 18798 + 12288 + 12289 + 12286 + 12287 + 12290 + 108071 \\ + 11550 + 12669 + 21338 + 21390 - 243764 + 26361 + 21924 + 11739 + 11740 + 11515 \\ + 12291 + 239566 + 58226 + 11941 \\ \end{array}$
Abbreviations: Bonferroni's m perturbation m Microarray datt package, <i>SPIA</i> The analysis w	ECM, e: ethod; p easured a of <i>RAC</i> Signific 'as perfo	xtracellt NDE, <i>P</i> lin eact 3 - $t^{-/-}$ cantly r med by	ular ma -value n pathw and wi nodulat / SPIA	Abbreviations: ECM, extracellular matrix; pG, global <i>P</i> -values, obtained Babnferroni's method; pNDE, <i>P</i> -value by the number of differentially expr perturbation measured in each pathway). Microarray data of RAG - $t^{-/-}$ and wild-type brains were analyzed by a package, $SPIA$. Significantly modulated pathways were selected by a Q The analysis was performed by SPIA (Bioconductor package).	<i>P</i> -values, obtain of differentially <i>lere</i> analyzed ere selected t package).	ained by com r expressed ç by a modere yy a global pr	ibining the pPt genes (classic: ate t-statistics a athway signific	ERT and pND ⁱ al test for the ε and fold chanç cance <i>P</i> -value	E using Fisher enrichment of (ge, and subser	's method; genes in a quently an ing false d	Abbreviations: ECM, extracellular matrix; pG, global <i>P</i> -values, obtained by combining the pPERT and pNDE using Fisher's method; pGFdr, global <i>P</i> -values after fdf correction; pGFWER, global <i>P</i> -value adjusted by the Bonferroni's method; pNDE, <i>P</i> -value by the number of differentially expressed genes (classical test for the enrichment of genes in a certain pathway); pPERT, <i>P</i> -value by perturbation (calculated based on the amount of perturbation measured in each pathway). Microarray data of <i>RAG-1^{-/-}</i> and wild-type brains were analyzed by a moderate t-statistics and fold change, and subsequently analyzed for the pathway enrichment with a bootstrap technique using the Bioconductor package, <i>SPIA</i> . Significantly modulated pathways were selected by a global pathway significance <i>P</i> -value with considering false discovery rate (<i>P</i> <0.05), which combines the enrichment and perturbation <i>P</i> -values. The analysis was performed by SPIA (Bioconductor package).

Table 2 Signaling pathway impact analysis of $RAG-t^{-/-}$ brain

Translational Psychiatry

13

Table 3 miRNA modulated in *RAG-1*^{-/-} brain and their relative targets.

miRNA	Cited in	Known targets
Mir539	Bao B <i>et al. J Nutr</i> 2010; Haga CL and Phinney DG. <i>J Biol Chem</i> 2012	Holocarboxylase synthetase Twist-related protein 1, polycomb complex protein BMI-1
Mir380	Hu K et al. BMC Neurosci 2012; Matsumoto S et al. Biochem Biophys Res Commun 2012	Unknown
Mir487b	Xi S et al. J Clin Invest 2013	Polycomb protein SUZ12, polycomb complex protein BMI-1, protein Wnt-5a, Myc proto-oncogene protein, GTPase KRas
Mir323	Qiu S <i>et al. J Transl Med</i> 2013; Fenoglio C <i>et al.</i> Int J Mol Sci 2012	Unknown
Mir680-2	None	Unknown
Mir665	Si H et al. J Cancer Res Clin Oncol 2013	Unknown
Mir376b	Korkmaz G et al. Autophagy 2012	Cysteine protease ATG4C and beclin-1
Mir382	Kriegel AJ et al. Physiol Genom 2012	Kallikrein 5
	Haga CL and Phinney DG. J Biol Chem 2012	Twist-related protein 1, polycomb complex protein BMI-1
Mir154	Milosevic J et al. Am J Respir Cell Mol Biol 2012	WNT/B-catenin pathway
Mir329	Khudayberdiev S et al. Commun Integr Biol 2009;	Unknown
	Qiu S et al. J Transl Med 2013	
Mir679	None	Unknown
Mir344	Qin L et al. BMC Genom 2010	WNT/β-catenin pathway
Mir9-2	Rodriguez-Otero P et al. Br J Haematol 2011	Fibroblast growth factor receptor 1 and cyclin-dependent kinase 6
Mir341	None	Unknown
Mir543	Haga CL and Phinney DG. J Biol Chem 2012	Twist-related protein 1, polycomb complex protein BMI-1
	,	

Abbreviations: miR, micro-RNA.

The table shows a list of miRNAs modulated in RAG-1^{-/-} compared to RAG-1^{-/-}/OT-II and wild type brains and the relative studies describing their molecular target(s).

gene development. Our microarray analysis provided us with a number of interesting findings.

A large number of micro-RNA(s) was found among the genes that were mostly downregulated. These gene expression modulators have been recently highlighted for their role in mental heath and are becoming increasingly popular in this field.⁶⁶ Literature search for known targets of the micro-RNA we have identified provided us with few indications (Table 3). Further studies are needed to investigate the functions of these micro-RNA and their role in emotional behavior. Several quantitative polymerase chain reaction validated genes (data not shown) in our screening are known to control a variety of neuropsychological conditions. Synuclein- γ has been associated with Parkinson's disease⁶⁷ and reported to be implicated in both cognitive and emotional functions.⁶⁸ Von Willebrand factor has been shown to be significantly increased in schizophrenia,69-71 whereas changes in polycystic kidney disease 1 and tetratricopeptide repeat and ankyrin repeat containing 1) expression have been associated with bipolar disorders.⁷²⁻⁷⁴ S100a10, a recently suggested potential biomarker for suicide risk in mental disorders,⁷⁵ was upregulated in $RAG-1^{-/-}$ and downregulated in $RAG-1^{-/-}/OT-II$ to WT level. Other interesting differentially expressed genes included ephrin type-B receptor 1 (Ephb1), whose genetic deletion in mice causes neuronal loss in the substantia nigra and spontaneous locomotor hyperactivity,⁷⁶ myeloid/lymphoid or mixed-lineage leukemia 2 (MII2), whose activity is required for memory formation,77 and Churchill domain containing protein 1 (Churc1), a neuronal development gene implicated with the occurrence of autism.78

Some of the implications of these changes in gene expression have been investigated with SPIA pathway analysis software. This showed inhibition of signaling pathways that control neurodegenerative disorders, such as Alzheimer's and Huntington's disease, in WT compared with $RAG-1^{-/-}$ mice further supporting the emerging view of these diseases as endowed with an autoimmune-related diseases component.^{79–81} Further differences were observed in a wide range of neuronal, sensory and basic cellular pathways that will be explored in future studies to detail the complex crosstalk between the neuronal and immune systems.

In conclusion, the results of this study shed new light on the complex crosstalk between the immune system and our emotional well-being, although future investigations are needed to corroborate our hypothesis. In fact, it would be interesting to explore the possible contribution of B cells to the emotional behavior of the $RAG-1^{-/-}$ mice as well as to confirm these results in mice expressing TCR transgene with different strength of signaling. Equally important, one might speculate the existence of CD4⁺ T-cell-specific factors that control emotional behaviors and their exploitation for the treatment of wide variety of mental disorders.

Beyond these experimental questions, the most important challenge for the future is to understand how T cells influence behavior and *vice versa*. The answer might lay in shared signaling pathway like *RAG* or the immune synapse: a signaling complex that has been named after the neuromuscular synapse and that allows the exchanges of information between antigen-presenting cells and T cells.^{82–84} Along these lines, recent studies have shown the existence of a subset of memory T cell in mice that produces acetylcholine in response to noradrenaline providing another way by which the immune system communicate with the nervous system.^{85,86} Taken together, these findings might help the design of new therapies for mental health by restoring an impaired or absent

1.4

immune system as observed in several autoimmune diseases.

Conflict of interest

The authors declare no conflict of interest.

Acknowledgements. We thank Dr Mangesh Thorat (Cancer Research UK Center for Epidemiology, Mathematics and Statistics) for helping us with brain section picture acquisition and Dr Dianne Cooper for carefully reading the manuscript. MO is a Human Frontier Science Program Long-term Fellow.

- Irwin M, Patterson T, Smith TL, Caldwell C, Brown SA, Gillin JC et al. Reduction of immune function in life stress and depression. *Biol Psychiatry* 1990; 27: 22–30.
- Ziv Y, Schwartz M. Immune-based regulation of adult neurogenesis: implications for learning and memory. Brain Behav Immun 2008; 22: 167–176.
- Yirmiya R, Goshen I. İmmune modulation of learning, memory, neural plasticity and neurogenesis. Brain Behav Immun 2011: 25: 181–213.
- Aktas O, Kieseier B, Hartung HP. Neuroprotection, regeneration and immunomodulation: broadening the therapeutic repertoire in multiple sclerosis. *Trends Neurosci* 2010; 33: 140–152.
- 5. German DC, Eagar T, Sonsalla PK. Parkinson's disease: a role for the immune system. *Curr Mol Pharmacol* 2011 (e-pub ahead of print).
- Lucin KM, Wyss-Coray T. Immune activation in brain aging and neurodegeneration: too much or too little? *Neuron* 2009; 64: 110–122.
- Littrell J. The mind-body connection: not just a theory anymore. Soc Work Health Care 2008; 46: 17–37.
- Costa-Pinto FA, Palermo-Neto J. Neuroimmune interactions in stress. *Neuroimmunomodulation* 2010; 17: 196–199.
- Rook GA, Lowry CA, Raison CL. Lymphocytes in neuroprotection, cognition and emotion: is intolerance really the answer? *Brain Behav Immun* 2011; 25: 591–601.
- 10. Barak Y. The immune system and happiness. Autoimmun Rev 2006; 5: 523-527.
- Sato Y, Takayanagi Y, Onaka T, Kobayashi E. Impact of cyclosporine upon emotional and social behavior in mice. *Transplantation* 2007; 83: 1365–1370.
- von Horsten S, Exton MS, Schult M, Nagel E, Stalp M, Schweitzer G et al. Behaviorally conditioned effects of cyclosporine A on the immune system of rats: specific alterations of blood leukocyte numbers and decrease of granulocyte function. J Neuroimmunol 1998; 85: 193–201.
- von Horsten S, Exton MS, Voge J, Schult M, Nagel E, Schmidt RE et al. Cyclosporine A affects open field behavior in DA rats. Pharmacol Biochem Behav 1998; 60: 71–76.
- Bechstein WO. Neurotoxicity of calcineurin inhibitors: impact and clinical management. Transpl Int 2000; 13: 313–326.
- 15. Wijdicks EF. Neurotoxicity of immunosuppressive drugs. Liver Transpl 2001; 7: 937-942.
- van der Meer A, Wulffraat NM, Prakken BJ, Gijsbers B, Rademaker CM, Sinnema G. Psychological side effects of MTX treatment in juvenile idiopathic arthritis: a pilot study. *Clin Exp Rheumatol* 2007; 25: 480–485.
- van der Hoeven J, Duyx J, de Langen JJ, van Royen A. Probable psychiatric side effects of azathioprine. *Psychosom Med* 2005; 67: 508.
- Pandey M, Sarita GP, Devi N, Thomas BC, Hussain BM, Krishnan R. Distress, anxiety, and depression in cancer patients undergoing chemotherapy. World J Surg Oncol 2006; 4: 68.
- Mombaerts P, Iacomini J, Johnson RS, Herrup K, Tonegawa S, Papaioannou VE. RAG-1-deficient mice have no mature B and T lymphocytes. *Cell* 1992; 68: 869–877.
- Cushman J, Lo J, Huang Z, Wasserfall C, Petitto JM. Neurobehavioral changes resulting from recombinase activation gene 1 deletion. *Clin Diagn Lab Immunol* 2003; 10: 13–18.
- McGowan PO, Hope TA, Meck WH, Kelsoe G, Williams CL. Impaired social recognition memory in recombination activating gene 1-deficient mice. *Brain Res* 2011; 1383: 187–195.
- Paschalidis N, Huggins A, Rowbotham NJ, Furmanski AL, Crompton T, Flower RJ et al. Role of endogenous annexin-A1 in the regulation of thymocyte positive and negative selection. *Cell Cycle* 2010; 9: 784–793.
- Deacon RM. Digging and marble burying in mice: simple methods for *in vivo* identification of biological impacts. *Nat Protoc* 2006; 1: 122–124.
- Deacon RM. Housing, husbandry and handling of rodents for behavioral experiments. Nat Protoc 2006; 1: 936–946.
- 25. Deacon RM. Assessing nest building in mice. Nat Protoc 2006; 1: 1117–1119.
- Ono M, Tanaka RJ, Kano M, Sugiman T. Visualising the cross-level relationships between pathological and physiological processes and gene expression: analyses of haematological diseases. *PLoS One* 2013; 8: e53544.
- Barnden MJ, Allison J, Heath WR, Carbone FR. Defective TCR expression in transgenic mice constructed using cDNA-based alpha- and beta-chain genes under the control of heterologous regulatory elements. *Immunol Cell Biol* 1998; 76: 34–40.
- Sharpley CF, Agnew LL. Cytokines and depression: findings, issues, and treatment implications. *Rev Neurosci* 2011; 22: 295–302.

- Tarca AL, Draghici S, Khatri P, Hassan SS, Mittal P, Kim JS et al. A novel signaling pathway impact analysis. *Bioinformatics* 2009; 25: 75–82.
- Marazziti D, Presta S, Pfanner C, Gemignani A, Rossi A, Sbrana S et al. Immunological alterations in adult obsessive-compulsive disorder. *Biol Psychiatry* 1999; 46: 810–814.
- Yonk LJ, Warren RP, Burger RA, Cole P, Odell JD, Warren WL et al. CD4 + helper T cell depression in autism. *Immunol Lett* 1990; 25: 341–345.
- Zena-Castillo D, Mezones-Holguin E, Valdiviezo-Garcia G, La-Chira-Alban A, Rodriguez-Morales AJ, Dickson-Gonzalez S. Impact of hospital-associated anxiety and depression on the CD4 counts of naive HIV/AIDS patients from locations in Northern Peru. Int J Infect Dis 2009; 13: e75–e76.
- 33. Miller AH. Depression and immunity: a role for T cells? Brain Behav Immun 2010; 24: 1-8.
- Huppert JD, Simpson HB, Nissenson KJ, Liebowitz MR, Foa EB. Quality of life and functional impairment in obsessive-compulsive disorder: a comparison of patients with and without comorbidity, patients in remission, and healthy controls. *Depress Anxiety* 2009; 26: 39–45.
- Deacon R. Assessing burrowing, nest construction, and hoarding in mice. J Vis Exp 2012; 59: e2607.
- Li X, Morrow D, Witkin JM. Decreases in nestlet shredding of mice by serotonin uptake inhibitors: comparison with marble burying. *Life Sci* 2006; 78: 1933–1939.
- Schwartz M. The emergence of a new science of the mind: immunology benefits the mind. Mol Psychiatry 2010; 15: 337–338.
- Kipnis J, Cohen H, Cardon M, Ziv Y, Schwartz M. T cell deficiency leads to cognitive dysfunction: implications for therapeutic vaccination for schizophrenia and other psychiatric conditions. *Proc Natl Acad Sci USA* 2004; **101**: 8180–8185.
- Kipnis J, Gadani S, Derecki NC. Pro-cognitive properties of T cells. Nat Rev Immunol 2012; 12: 663–669.
- Brynskikh A, Warren T, Zhu J, Kipnis J. Adaptive immunity affects learning behavior in mice. Brain Behav Immun 2008; 22: 861–869.
- Kipnis J, Mizrahi T, Hauben E, Shaked I, Shevach E, Schwartz M. Neuroprotective autoimmunity: naturally occurring CD4 + CD25 + regulatory T cells suppress the ability to withstand injury to the central nervous system. *Proc Natl Acad Sci USA* 2002; 99: 15620–15625.
- Schwartz M, Kipnis J. Protective autoimmunity and neuroprotection in inflammatory and noninflammatory neurodegenerative diseases. J Neurol Sci 2005; 233: 163–166.
- Schwartz M, Ziv Y. Immunity to self and self-maintenance: a unified theory of brain pathologies. Trends Immunol 2008; 29: 211–219.
- Schwartz M, Shechter R. Protective autoimmunity functions by intracranial immunosurveillance to support the mind: The missing link between health and disease. *Mol Psychiatry* 2010; 15: 342–354.
- Barouch R, Schwartz M. Autoreactive T cells induce neurotrophin production by immune and neural cells in injured rat optic nerve: implications for protective autoimmunity. *FASEB J* 2002; 16: 1304–1306.
- Derecki NC, Cardani AN, Yang CH, Quinnies KM, Crihfield A, Lynch KR et al. Regulation of learning and memory by meningeal immunity: a key role for IL-4. J Exp Med 2010; 207: 1067–1080.
- Aas M, Navari S, Gibbs A, Mondelli V, Fisher HL, Morgan C *et al.* Is there a link between childhood trauma, cognition, and amygdala and hippocampus volume in first-episode psychosis? *Schizophr Res* 2012; **137**: 73–79.
- Aas M, Dazzan P, Fisher HL, Morgan C, Morgan K, Reichenberg A et al. Childhood trauma and cognitive function in first-episode affective and non-affective psychosis. Schizophr Res 2011; 129: 12–19.
- Pawlby S, Hay D, Sharp D, Waters CS, Pariante CM. Antenatal depression and offspring psychopathology: the influence of childhood maltreatment. *Br J Psychiatry* 2011; 199: 106–112.
- Monk C, Fitelson EM, Werner E. Mood disorders and their pharmacological treatment during pregnancy: is the future child affected? *Pediatr Res* 2011; 69(Part 2): 3R–10R.
- Harvey L, Boksa P. Prenatal and postnatal animal models of immune activation: relevance to a range of neurodevelopmental disorders. *Dev Neurobiol* 2012; 72: 1335–1348.
- Schlotz W, Phillips DI. Fetal origins of mental health: evidence and mechanisms. Brain Behav Immun 2009; 23: 905–916.
- Ellis S, Mouihate A, Pittman QJ. Neonatal programming of the rat neuroimmune response: stimulus specific changes elicited by bacterial and viral mimetics. *J Physiol* 2006; 571 (Pt 3): 695–701.
- Hsiao EY, McBride SW, Chow J, Mazmanian SK, Patterson PH. Modeling an autism risk factor in mice leads to permanent immune dysregulation. *Proc Natl Acad Sci USA* 2012; 109: 12776–12781.
- Hsiao EY, Patterson PH. Placental regulation of maternal-fetal interactions and brain development. *Dev Neurobiol* 2012; 72: 1317–1326.
- Hodyl NA, Walker FR, Krivanek KM, Clifton VL, Hodgson DM. Prenatal endotoxin exposure alters behavioural pain responses to lipopolysaccharide in adult offspring. *Physiol Behav* 2010; **100**: 143–147.
- Hodyl NA, Krivanek KM, Lawrence E, Clifton VL, Hodgson DM. Prenatal exposure to a proinflammatory stimulus causes delays in the development of the innate immune response to LPS in the offspring. J Neuroimmunol 2007; 190: 61–71.
- Mouihate A, Galic MA, Ellis SL, Spencer SJ, Tsutsui S, Pittman QJ. Early life activation of toll-like receptor 4 reprograms neural anti-inflammatory pathways. *J Neurosci* 2010; 30: 7975–7983.

- Saito S, Nakashima A, Shima T, Ito M. Th1/Th2/Th17 and regulatory T-cell paradigm in pregnancy. Am J Reprod Immunol 2010; 63: 601–610.
- Gadani SP, Cronk JC, Norris GT, Kipnis J. IL-4 in the brain: a cytokine to remember. J Immunol 2012; 189: 4213–4219.
- Ferreira TB, Kasahara TM, Barros PO, Vieira MM, Bittencourt VC, Hygino J et al. Dopamine up-regulates Th17 phenotype from individuals with generalized anxiety disorder. J Neuroimmunol 2011; 238: 58–66.
- Vieira MM, Ferreira TB, Pacheco PA, Barros PO, Almeida CR, Araujo-Lima CF et al. Enhanced Th17 phenotype in individuals with generalized anxiety disorder. J Neuroimmunol 2010; 229: 212–218.
- Beurel E, Harrington LE, Jope RS. Inflammatory T helper 17 cells promote depression-like behavior in mice. *Biol Psychiatry* 2012; 73: 622–630.
- Kim SJ, Lee H, Joung HY, Lee G, Lee HJ, Shin MK et al. T-bet deficient mice exhibit resistance to stress-induced development of depression-like behaviors. J Neuroimmunol 2011; 240–241: 45–51.
- Postal M, Pelicari KO, Sinicato NA, Marini R, Costallat LT, Appenzeller S. Th1/Th2 cytokine profile in childhood-onset systemic lupus erythematosus. *Cytokine* 2013; 61: 785–791.
- O'Connor RM, Dinan TG, Cryan JF. Little things on which happiness depends: microRNAs as novel therapeutic targets for the treatment of anxiety and depression. *Mol Psychiatry* 2012; 17: 359–376.
- Nishioka K, Wider C, Vilarino-Guell C, Soto-Ortolaza AI, Lincoln SJ, Kachergus JM *et al.* Association of alpha-, beta-, and gamma-Synuclein with diffuse lewy body disease. *Arch Neurol* 2010; 67: 970–975.
- Kokhan VS, Bolkunov AV, Ustiugov AA, Van'kin GI, Shelkovnikova TA, Redkozubova OM et al. Targeted inactivation of gamma-synuclein gene affects anxiety and exploratory behaviour of mice. *Zh Vyssh Nerv Deiat Im I P Pavlova* 2011; 61: 85–93.
- Brietzke E, Teixeira AL. Similar immune profile in bipolar disorder and schizophrenia: selective increase in soluble tumor necrosis factor receptor I and von Willebrand factor. *Bipolar Disord* 2010; **12**: 453–454.
- Ho-Yen DO, Kay DS, Henderson R, Todd AS, Naylor GJ. Factor VIII-related antigen and procoagulant activity in manic-depressive patients. J Affect Disord 1983; 5: 209–213.
- Hope S, Melle I, Aukrust P, Steen NE, Birkenaes AB, Lorentzen S et al. Similar immune profile in bipolar disorder and schizophrenia: selective increase in soluble tumor necrosis factor receptor I and von Willebrand factor. *Bipolar Disord* 2009; **11**: 726–734.
- Kimmel RJ, Kovacs I, Vrabel C, Wood B, Schalling M, Kelsoe JR. Cosegregation of bipolar disorder and autosomal-dominant medullary cystic kidney disease in a large family. *Am J Psychiatry* 2005; **162**: 1972–1974.
- Wylie K, de Silva D, Jerram T, Mindham RH. Simultaneous kidney disease and manic-depressive psychosis. Br J Psychiatry 1993; 162: 275–276.

- Chen DT, Jiang X, Akula N, Shugart YY, Wendland JR, Steele CJ et al. Genome-wide association study meta-analysis of European and Asian-ancestry samples identifies three novel loci associated with bipolar disorder. *Mol Psychiatry* 2013; 18: 195–205.
- Zhang L, Su TP, Choi K, Maree W, Li CT, Chung MY et al. P11 (S100A10) as a potential biomarker of psychiatric patients at risk of suicide. J Psychiatr Res 2011; 45: 435–441.
- Richards AB, Scheel TA, Wang K, Henkemeyer M, Kromer LF. EphB1 null mice exhibit neuronal loss in substantia nigra pars reticulata and spontaneous locomotor hyperactivity. *Eur J Neurosci* 2007; 25: 2619–2628.
- Kerimoglu C, Agis-Balboa RC, Kranz A, Stilling R, Bahari-Javan S, Benito-Garagorri E *et al.* Histone-methyltransferase MLL2 (KMT2B) is required for memory formation in mice. *J Neurosci* 2013; 33: 3452–3464.
- Griswold AJ, Ma D, Sacharow SJ, Robinson JL, Jaworski JM, Wright HH et al. A de novo 1.5 Mb microdeletion on chromosome 14q23.2–23.3 in a patient with autism and spherocytosis. Autism Res 2011; 4: 221–227.
- Villoslada P, Moreno B, Melero I, Pablos JL, Martino G, Uccelli A et al. Immunotherapy for neurological diseases. *Clin Immunol* 2008; 128: 294–305.
- Wisniewski T, Konietzko U. Amyloid-beta immunisation for Alzheimer's disease. Lancet Neurol 2008: 7: 805–811.
- 81. Cardoso F. Huntington disease and other choreas. *Neurol Clin* 2009; **27**: 719–736; vi.
- Monks CR, Freiberg BA, Kupfer H, Sciaky N, Kupfer A. Three-dimensional segregation of supramolecular activation clusters in T cells. *Nature* 1998; 395: 82–86.
- Reichardt P, Dornbach B, Gunzer M. APC, T cells, and the immune synapse. *Curr Top Microbiol Immunol* 2010; 340: 229–249.
- Valitutti S, Dupre L. Plasticity of immunological synapses. Curr Top Microbiol Immunol 2010; 340: 209–228.
- Rosas-Ballina M, Olofsson PS, Ochani M, Valdes-Ferrer SI, Levine YA, Reardon C *et al.* Acetylcholine-synthesizing T cells relay neural signals in a vagus nerve circuit. *Science* 2011; **334**: 98–101.
- 86. Woodman I. Neuroimmunology: nervous ChAT. Nat Rev Immunol 2011; 11: 720.

Translational Psychiatry is an open-access journal published by Nature Publishing Group. This work is licensed under a Creative Commons Attribution 3.0 Unported License. To view a copy of this license, visit http://creativecommons.org/ licenses/by/3.0/

Supplementary Information accompanies the paper on the Translational Psychiatry website (http://www.nature.com/tp)