





Genome Sequence of Australian Indigenous Wine Yeast Torulaspora delbrueckii COFT1 Using Nanopore Sequencing

Federico Tondini, a,b Vladimir Jiranek, a,b Paul R. Grbin, a,b Cristobal A. Onettoa

^aDepartment of Wine & Food Science, University of Adelaide, Adelaide, South Australia, Australia ^bAustralian Research Council Industrial Transformation Training Centre for Innovative Wine Production, Adelaide, South Australia, Australia

ABSTRACT Here, we report the first sequenced genome of an indigenous Australian wine isolate of *Torulaspora delbrueckii* using the Oxford Nanopore MinION and Illumina HiSeq sequencing platforms. The genome size is 9.4 Mb and contains 4,831 genes.

Torulaspora delbrueckii occurs saprophytically on wine grape surfaces worldwide (1). Under winemaking conditions, it displays a less vigorous fermentation phenotype than Saccharomyces cerevisiae, differing in flavor and aroma compound production (2). Its favorable oenological traits, such as low acetic acid production and osmotic tolerance, have led to its commercialization and adoption for use in the wine industry (3). Despite this, T. delbrueckii is still not well characterized at a molecular level, with no other genomes reported from wine isolates.

For a better understanding of *T. delbrueckii* oenological traits, RNA sequencing (RNA-seq) studies under wine-like conditions are required. However, in the absence of a closely related reference sequence, significant challenges remain when it comes to assembling short reads into full-length gene and transcript models. In order to facilitate future wine-related studies, the genome of a wine isolate of *T. delbrueckii* was sequenced and characterized.

T. delbrueckii strain COFT1 was isolated from a spontaneous wine fermentation at the Yalumba Wine Company (Angaston, South Australia, Australia). High-molecularweight genomic DNA was extracted according to the phenol-chloroform protocol (4), including a 2-h preincubation with Zymolase. DNA was prepared for sequencing with the MinION device using the SQK-LSK108 library prep kit (protocol GDE_9002_v108_ revT_18Oct2016) and R9.4 chemistry. A total of 138,992 reads were obtained, for a total of 1,214 Mbp (130 \times coverage) and an average length of 8,737 bp. Fast5 files were base called using Albacore version 2.0.2. Passed reads were trimmed for adapters using PoreChop version 0.2.3 and then assembled using SMARTdenovo version 1.0 (https:// github.com/ruanjue/smartdenovo). Contigs obtained from the assembly were polished using Racon version 0.5.0 (5) and Nanopolish version 0.8.5 (6). A final polish of the assembly was performed with Pilon version 1.22 using Illumina HiSeq RNA reads extracted from pure culture laboratory ferments with strain COFT1. The final assembly had no gaps, with a total length of 9,356,826 bp arranged in 9 chromosomes (1 mitochondrial chromosome) and an average GC content of 42%. The genome was first annotated with YGAP (7), and 5,231 genes were predicted. An improved annotation was performed with MAKER2 (8), providing STAR RNA alignment information (9) for the prediction of protein-coding genes. A total of 4,831 protein-coding genes were identified, compared to 4,714 and 4,972 protein-coding genes reported in previous genomes (10, 11). Functional annotation of the predicted protein sequences was performed using BLASTP (12) against the Swiss-Prot protein sequence database (E value = 1e-5) (13). BUSCO assessment (14) revealed a genome completeness of 98%.

Received 23 March 2018 **Accepted** 24 March 2018 **Published** 26 April 2018

Citation Tondini F, Jiranek V, Grbin PR, Onetto CA. 2018. Genome sequence of Australian indigenous wine yeast *Torulaspora delbrueckii* COFT1 using nanopore sequencing. Genome Announc 6:e00321-18. https://doi.org/10.1128/genomeA.00321-18.

Copyright © 2018 Tondini et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Vladimir Jiranek, vladimir.jiranek@adelaide.edu.au.

Tondini et al.

The functional annotation of the genome, as well as its completeness, allows comparison with the species *S. cerevisiae* and a better understanding of the oenological traits of this yeast. The genome sequence reported here will assist in delivering clearer transcriptional results in complex wine-like fermentations (e.g., mixed fermentations), providing useful insight into these processes for the wine industry.

Accession number(s). The final genome sequence has been deposited to NCBI GenBank database under accession numbers CP027647 to CP027655.

ACKNOWLEDGMENTS

We thank the Yalumba Wine Company for providing the wine samples and the Genomics Facility at SAHMRI (Adelaide, South Australia) for the use of the TapeStation 2200.

This work was supported by the Australian Research Council Industrial Transformation Training Centre for Innovative Wine Production (project IC130100005) and Wine Australia (project AGT1502).

REFERENCES

- 1. Renouf V, Claisse O, Lonvaud-Funel A. 2005. Understanding the microbial ecosystem on the grape berry surface through numeration and identification of yeast and bacteria. Aust J Grape Wine Res 11:316–327. https://doi.org/10.1111/j.1755-0238.2005.tb00031.x.
- Jolly NP, Varela C, Pretorius IS. 2014. Not your ordinary yeast: non-Saccharomyces yeasts in wine production uncovered. FEMS Yeast Res 14:215–237. https://doi.org/10.1111/1567-1364.12111.
- Ciani M, Morales P, Comitini F, Tronchoni J, Canonico L, Curiel JA, Oro L, Rodrigues AJ, Gonzalez R. 2016. Non-conventional yeast species for lowering ethanol content of wines. Front Microbiol 7:642. https://doi. org/10.3389/fmicb.2016.00642.
- Sambrook J, Russell DW. 2006. Purification of nucleic acids by extraction with phenol: chloroform. Cold Spring Harb Protoc 2006:pdb.prot4045. https://doi.org/10.1101/pdb.prot4045.
- Vaser R, Sović I, Nagarajan N, Šikić M. 2017. Fast and accurate *de novo* genome assembly from long uncorrected reads. Genome Res 27: 737–746. https://doi.org/10.1101/gr.214270.116.
- Loman NJ, Quick J, Simpson JT. 2015. A complete bacterial genome assembled *de novo* using only nanopore sequencing data. Nat Methods 12:733. https://doi.org/10.1038/nmeth.3444.
- Proux-Wéra E, Armisén D, Byrne KP, Wolfe KH. 2012. A pipeline for automated annotation of yeast genome sequences by a conservedsynteny approach. BMC Bioinformatics 13:237. https://doi.org/10.1186/ 1471-2105-13-237.
- 8. Holt C, Yandell M. 2011. MAKER2: an annotation pipeline and genome-

- database management tool for second-generation genome projects. BMC Bioinformatics 12:491. https://doi.org/10.1186/1471-2105-12-491.
- Dobin A, Gingeras TR. 2015. Mapping RNA-seq reads with STAR. Curr Protoc Bioinformatics 51:11–19. https://doi.org/10.1002/0471250953.bi11 14s51.
- Gomez-Angulo J, Vega-Alvarado L, Escalante-García Z, Grande R, Gschaedler-Mathis A, Amaya-Delgado L, Arrizon J, Sanchez-Flores A. 2015. Genome sequence of *Torulaspora delbrueckii* NRRL Y-50541, isolated from Mezcal fermentation. Genome Announc 3:e00438-15. https://doi.org/10.1128/genomeA.00438-15.
- Gordon JL, Armisén D, Proux-Wéra E, ÓhÉigeartaigh SS, Byrne KP, Wolfe KH. 2011. Evolutionary erosion of yeast sex chromosomes by matingtype switching accidents. Proc Natl Acad Sci U S A 108:20024. https:// doi.org/10.1073/pnas.1112808108.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403. https://doi.org/10.1016/S0022 -2836(05)80360-2.
- Bairoch A, Apweiler R. 2000. The Swiss-Prot protein sequence database and its supplement TrEMBL in 2000. Nucleic Acids Res 28:45–48. https:// doi.org/10.1093/nar/28.1.45.
- Simão FA, Waterhouse RM, Ioannidis P, Kriventseva EV, Zdobnov EM. 2015. BUSCO: assessing genome assembly and annotation completeness with single-copy orthologs. Bioinformatics 31:3210–3212. https://doi.org/10.1093/bioinformatics/btv351.