


Roles of *BMI1* in the Initiation, Progression, and Treatment of Hepatocellular Carcinoma

Ru Wang, BD¹ , Hengwei Fan, MD² , Ming Sun, MD¹,
Zhongwei Lv, PhD¹, and Wanwan Yi, MD¹

Technology in Cancer Research & Treatment
Volume 21: 1–9
© The Author(s) 2022
Article reuse guidelines:
sagepub.com/journals-permissions
DOI: 10.1177/15330338211070689
journals.sagepub.com/home/tct


Abstract

Liver cancer has high rates of morbidity and mortality, and its treatment is a global health challenge. Hepatocellular carcinoma (HCC) accounts for 90% of all primary liver cancer cases. B-lymphoma Mo-MLV insertion region 1 (*BMI1*) has been identified as a proto-oncogene, which contributes to the initiation and progression of many malignant tumors. *BMI1* expression is upregulated in HCC, and it influences the occurrence and development of HCC by various mechanisms, such as the INK4a/ARF locus, NF- κ B signaling pathway, and PTEN/PI3K/AKT signaling pathway. In addition, the expression of *BMI1* is related to prognosis and recurrence of HCC. Hence, there is clear evidence that *BMI1* is a novel and valid therapeutic target for HCC. Accordingly, the development of therapeutic strategies targeting *BMI1* has been a focus of recent research, providing new directions for HCC treatment. This review summarizes the role of *BMI1* in the occurrence and treatment of HCC, which will provide a basis for using *BMI1* as a potential target for the development of therapeutic strategies for HCC.

Keywords

hepatocellular carcinoma, *BMI1*, miR-218, miR-203, INK4a/ARF locus

Abbreviations

AFP, Alpha-fetoprotein; *BMI1*, B-lymphoma Mo-MLV insertion region 1; CDK, cyclin-dependent kinase; CDDP, cis-dichlorodiamminoplatinum (II); CpsI, carbamoyl phosphate synthase I; CRISPR-Cas9, clustered regularly interspaced short palindromic repeats and CRISPR associated protein-9; CXCR4, CXC chemokine receptor 4; ECM, the extracellular matrix; EMT, epithelial-to-mesenchymal transition; GEMS, Gene Expression Modulation by Small Molecules; HCC, Hepatocellular carcinoma; HPCs, hepatic progenitor cells; JXY, Jiedu Xiaozheng Yin; MMP, matrix metalloproteinase; NPSC, Nanoplatin and siRNA co-loaded CaP nanoparticles; PcG, polycomb group; RKIP, Raf kinase inhibitor protein; siRNAs, small interfering RNAs; THA, 1,6,7-trihydroxyxanthone; VEGF, vascular endothelial growth factor.

Received: August 24, 2021; Revised: November 16, 2021; Accepted: December 15, 2021.

Introduction

Liver cancer poses a global health challenge, with 905 677 new cases detected in 2020 and an estimated incidence of >1 million cases by 2025.¹ Liver cancer is the sixth most common cancer worldwide and the fourth leading cause of cancer deaths globally. Hepatocellular carcinoma (HCC) accounts for more than 90% of all primary liver cancer cases and is the most common type of liver cancer. Current therapeutic methods for liver cancer mainly include surgical resection, liver transplantation, local ablation, external radiation, trans-arterial therapies, chemotherapy, targeted therapy, and immunotherapy.² However, the efficacies of these strategies remain insufficient, and the mortality rate of liver cancer is still high. Therefore, to improve survival

¹ Shanghai Tenth People's Hospital, Tongji University School of Medicine, Shanghai, China

² The Eastern Hepatobiliary Surgery Hospital, Navy Medical University (Second Military Medical University), Shanghai, China

Authors Ru Wang, Hengwei Fan, and Ming Sun contributed equally to this work.

Corresponding Authors:

Zhongwei Lv and Wanwan Yi, Department of Nuclear Medicine, Shanghai Tenth People's Hospital, Tongji University School of Medicine, Yanchang Middle Road 301, Shanghai 20072, China.

Emails: Lvzwjs2020@163.com; 1911608@tongji.edu.cn



and the quality of life, new and effective treatment methods for HCC are required.

Similar to the processes involved in other cancers, the occurrence and invasion of HCC involve complicated processes, involving multiple factors and processes. A series of physiological processes involving the polycomb group (*PcG*) gene family, including cellular differentiation, stem cell self-renewal, and correct gene silencing, are critical for normal tissue functioning. B-lymphoma Mo-MLV insertion region 1 (*BMI1*) encodes a key PcG protein that ubiquitylates histone H2A by forming a stable heterodimer with RING1B.³ *BMI1* has been identified as a proto-oncogene. It encodes a protein with 324 amino acids, which is principally localized in the nucleus and contributes to the generation of mouse pre-B cell lymphomas.⁴ *BMI1* has been found in various normal cells and is highly conserved evolutionarily; moreover, it contains a type of zinc finger motif.⁵ Accumulating evidence indicates that *BMI1* is expressed at significantly high levels and is closely related to

the occurrence, development, and outcomes of various human malignancies.^{6–8} In endometrial adenocarcinoma, the overexpression of *BMI1* is a poor prognostic factor correlated with the invasion of the myometrium and lymph nodes.⁹ In addition, in gastric cancer, the *BMI1* gene regulates tumor growth, metastasis, and chemoresistance by downregulating Raf kinase inhibitor protein (RKIP) expression.¹⁰ Therefore, a clear understanding of the role of *BMI1* in HCC is essential for the development of new therapeutic strategies. This review describes the roles of *BMI1* in the occurrence, progression, and prognosis of HCC as well as the implications of recent research for treatment strategies for HCC.

BMI1 Expression Is Upregulated in HCC

Neo et al identified 218 differentially expressed genes between HCC and normal liver tissues; these genes included *BMI1*.¹¹ *BMI1* expression is markedly higher in HCC tissues than in normal and adjacent non-cancerous liver tissues,^{12–14} consistent with the results for other cancers.^{8,15,16} However, changes in the expression of *BMI1* in HCC at different stages are still controversial. Sasaki et al established that the percentage of *BMI1*-positive cells is higher in poorly differentiated HCC ($91.22 \pm 5.64\%$) than in moderately differentiated HCC ($56.94 \pm 28.55\%$) and well-differentiated HCC ($8.48 \pm 11.97\%$).¹⁷ In contrast, Effendi et al concluded that *BMI1* expression level is the highest in well-differentiated HCC (including early HCC samples), followed by the expression in moderately differentiated and poorly differentiated HCC.¹⁸ These conflicting results could be because of differences in analysis methods, evaluation criteria, and population differences. Both were semi-quantitative analyses; however, Sasaki et al evaluated the proportion of *BMI1*-positive cells in 27 HCC tissue samples (among >100 cells) under a microscope,¹⁷ while Effendi et al evaluated tissues resected from patients with HCC by immunohistochemical staining based on a 3-point scale (0 for no staining, 1+ for focal and weak distribution, and 2+ for diffuse and clear distribution).¹⁸ To further explore the relationship between the expression of *BMI1* and the degree of HCC differentiation, Effendi et al determined *BMI1* mRNA expression levels in HCC samples with different levels of differentiation and obtained similar results. Both studies proved that *BMI1* is highly expressed in the nuclei in HCC. Nevertheless, correlations between high *BMI1* expression levels and clinical characteristics of patients with HCC should be investigated further. Based on data for 62 patients with HCC, Li et al determined that high *BMI1* expression is not statistically correlated with age, gender, satellite foci, tumor location and number, and the level of alpha-fetoprotein (AFP) but was highly correlated with neoplasm size, distant metastasis, vascular infiltration, and AJCC TNM stage.¹⁹ Sasaki et al also detected correlations between high *BMI1* expression levels and venous invasion and cancer cell proliferation, thus revealing that *BMI1* accelerates disease progression and is a poor prognostic factor in HCC.¹⁷ Surprisingly, Wang et al obtained contradictory results. They found that the expression

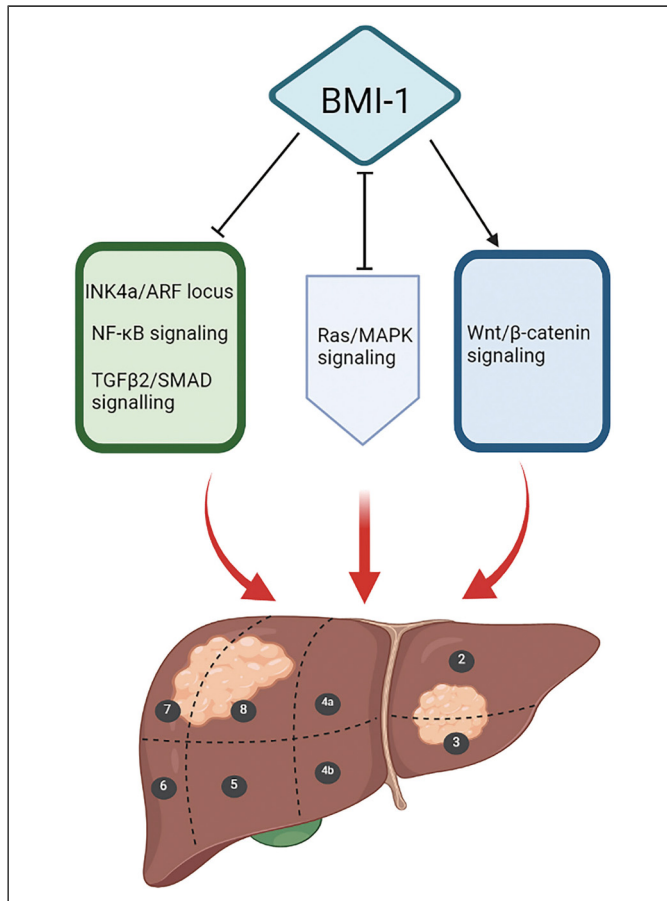


Figure 1. Mechanism underlying the initiation of hepatocellular carcinoma (HCC) by *BMI1*. Previous studies have shown that *BMI1* promotes the occurrence of HCC by blocking the *INK4a/ARF* locus, NF-κB signaling pathway, and TGFβ2/SMAD signaling axis and by stimulating the Wnt/β-catenin signaling axis. Besides, *BMI1* and Ras/MAPK signaling work together to activate the initiation of HCC. Black and red arrows represent promotion, T-shaped symbols represent inhibition, and H-shaped symbols represent interactive effects.

of *BMI1* was not related to the basic clinical characteristics of patients with HCC or to metastasis and postoperative recurrence, although they reported that high *BMI1* levels promoted the development and progression of HCC.²⁰ These inconsistent findings may be related to the differences in disease stage and indicate that more comprehensive and objective analyses are urgently required.

Mechanism by Which *BMI1* Contributes to the Initiation of HCC

Excessive proliferation of cells plays a pivotal role in the early formation of cancers.²¹ Several studies have demonstrated that *BMI1* not only participates in the self-renewal of normal hematopoietic stem cells but also contributes greatly to the continuous proliferation of acute myelocytic leukemia stem cells without senescence and apoptosis.^{22,23} It has been reported

that *BMI1* is required for hepatic stem cell expansion.²⁴ Interestingly, *BMI1* promotes the proliferation, colony formation, and cell cycle progression of hepatic progenitor cells in vitro, and high *BMI1* expression in hepatic progenitor cells results in the occurrence of poorly differentiated HCC in nude mice.²⁵

The initiation of HCC by *BMI1* is mediated by many mechanisms (Figure 1). Chiba et al revealed that *BMI1* determines the self-renewal capability of side population cells purified from HCC tissues and demonstrated the high proliferative potential and anti-apoptotic properties of these cells,²⁶ which directly contribute to its tumorigenic potential.²⁷ Although *BMI1* can promote the self-renewal of hepatic stem cells, it has no influence on the overall cell cycle, and there is no evidence that *BMI1* promotes the growth or self-renewal of differentiated cells.²⁸ However, Ma et al reported that the downregulation of *BMI1* expression could induce CD133⁺ Huh7 cell cycle arrest in the G0/G1 and S phases by blocking the NF- κ B signaling pathway, thereby promoting cell apoptosis.²⁹ It has previously been reported that *BMI1* regulates the cell cycle, apoptosis, and senescence by repressing *INK4a/ARF*, encoding p16^{ink4a}, which inhibits the activity of cyclin-dependent kinase (CDK), as well as p19^{ARF}, a well-known tumor suppressor (Figure 2).^{30–32} Direct binding of p16^{INK4a} to CDK4 and CDK6 maintains Rb in a hypophosphorylated state. Hypophosphorylated Rb represses E2F-dependent transcription, causing cell cycle arrest and senescence.³³ p19^{ARF} suppresses MDM2, which mediates the ubiquitin-dependent degradation of p53, and subsequently activates p53 target genes involved in cell cycle arrest and apoptosis.^{34,35} The increase in cell self-renewal ability induced by *BMI1* in neural stem cells and glioma cells is not completely *INK4a/ARF*-dependent.^{36,37} In a study by Chiba et al the expression levels of *INK4a* and *ARF* were increased in PLC/PRF/5 cells upon *BMI1* knockdown, with no remarkable changes in Huh7 cells compared with levels in the control group.²⁷ Intriguingly, *BMI1* increased the expression of p16^{INK4a} and inhibited the expression of p19^{ARF} in mouse hepatocytes.³⁸ Hence, Chiba et al further explored the role of the *INK4a/ARF* tumor suppressor gene locus in hepatic stem cell expansion and hepatocarcinogenesis. They ascertained that transfection with *BMI1* alone could enhance the renewal ability of Dlk⁺ cells, without *INK4a/ARF* expression, demonstrating that the *INK4a/ARF* locus does not directly contribute to early hepatoma formation.³⁹ Additionally, they identified five candidate genes downstream of *BMI1* in hepatic stem/progenitor cells, representing a major step forward. Among the five candidate genes, *Prom1* (CD133) and *EpCAM* are highly expressed in hepatic stem cells, whereas carbamoyl phosphate synthase I (*Cps1*), *Mat1a*, and *Gjb2* show low expression in hepatocyte-differentiated cells.³⁹ Knockout of the *Mat1a* gene in mice enhanced the sensitivity of hepatocytes to oxidative stress and accelerated the formation of HCC, providing powerful evidence that *Mat1a* is a hepatoma marker.⁴⁰ Furthermore, *Cps1* may be a key component in the progression of hepatocytes to malignant HCC cells by

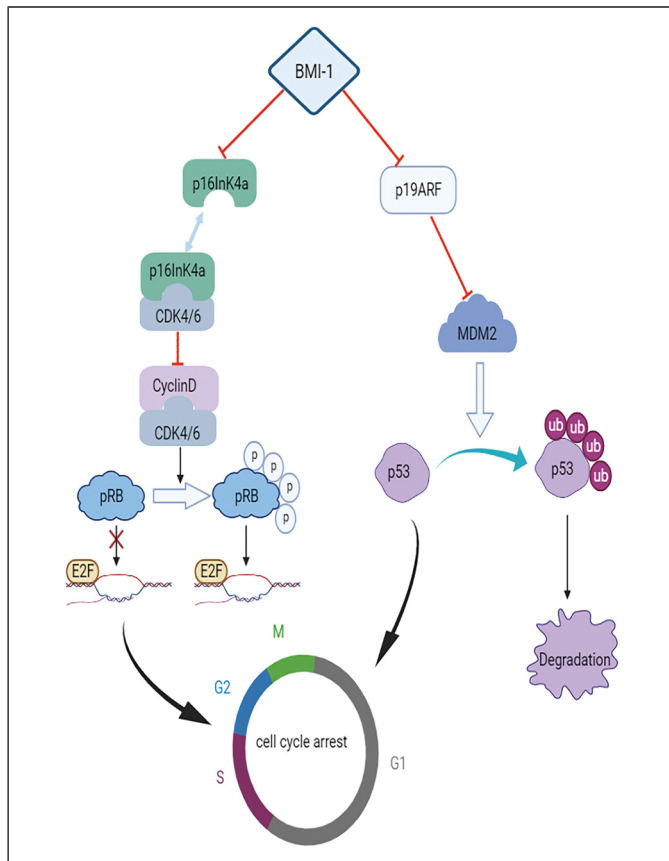


Figure 2. Mechanism by which *INK4a/ARF* mediates the effects of *BMI1*. The *BMI1* gene downregulates the expression of p16^{INK4a} and p19^{ARF}. Direct binding of p16^{INK4a} to CDK4 and CDK6 blocks the binding of CDK4/6 to cyclin D, resulting in the dephosphorylated state of Rb. Dephosphorylated Rb does not dissociate the repressive complexes of E2F, whose protein products dictate regular and correct DNA transcription, and thus prevents the release of E2F, causing cell cycle arrest and senescence. p19^{ARF} suppresses MDM2, which mediates the ubiquitination and degradation of p53, and can initiate related processes to promote abnormal cell cycle arrest and apoptosis during uncontrolled proliferation or irreparable damage.

metabolic reprogramming.⁴¹ Nevertheless, further research is necessary to determine the relationships between these five candidate downstream targets and carcinogenesis. Chiba et al ultimately determined that *BMII* promotes hepatic stem cell expansion and tumorigenicity in both *INK4a/ARF*-dependent and *INK4a/ARF*-independent manners in mice.³⁹ This conclusion is supported by the results of Fu et al who found that *BMII* knockdown induces G1-phase arrest and activates the p14^{ARF} and p16^{INK4a} signaling pathways in HepG2 cells,⁴² as well as the results of Xu et al who established that the silencing of *BMII* expression does not lead to the upregulation of p14^{ARF} and p16^{INK4a} in Huh7 and Hep3B cells.³⁸

Furthermore, some additional *INK4a/ARF*-independent mechanisms have been reported. It has been proposed that *BMII* functions synergistically with activated Ras/MAPK signaling and inhibits the TGF β 2/SMAD signaling axis to induce the occurrence of HCC.^{38,43} *BMII* stimulates the Wnt/ β -catenin signaling axis to increase the expression levels of cyclin D1, c-myc, and E-cadherin, thereby activating HCC cell proliferation.⁴⁴ These findings unequivocally demonstrate that *BMII* mediates the occurrence of HCC by complex and diverse mechanisms. In the future, these mechanisms should be explored further to provide more ideas for the treatment of HCC.

Relationship Between *BMII* and the Progression of HCC

Metastasis and invasion are crucial hallmarks of cancer progression. *BMII* is important for the invasion and metastasis of various cancers,^{45,46} including HCC.¹⁹ Cancer invasion and metastasis are complex and multi-factorial processes that involve tumor angiogenesis, extracellular matrix degradation, and the epithelial-to-mesenchymal transition (EMT).⁴⁷ Li et al have revealed that *BMII* can inhibit the PTEN/PI3K/AKT signaling pathway in HCC, ultimately stimulating metastasis by increasing the expression and activity of vascular endothelial growth factor (VEGF), matrix metalloproteinase (MMP)-9, and MMP-2.¹⁹ PTEN that encodes the protein phosphatase phosphatase and tensin homolog is mutated or lost in various human cancers, including HCC.⁴⁸ PTEN acts as a negative regulator of the PI3K/AKT signaling pathway in cells.⁴⁹ Tumor growth and migration are inseparable from the nutritional support provided by the blood; VEGF has important roles in tumor angiogenesis and metastasis. Matrix metalloproteinases are the common extracellular matrix-degrading enzymes. The expression levels of MMP-2 and MMP-9 are markedly upregulated in later TNM stages of HCC, and there is a significant correlation between MMP-9 (but not MMP-2) and PTEN in HCC.⁵⁰ MMP-9 is upregulated via the PI3K/AKT/NF- κ B signaling pathway, which promotes HCC metastasis and invasion.⁵¹

BMII overexpression accelerates the progression of HCC by inducing EMT. Vimentin is beneficial for EMT, whereas E-cadherin inhibits this process.⁵² Zhang et al demonstrated

that high *BMII* expression decreased E-cadherin levels and increased vimentin levels in CD133⁺ HepG2 cells, which are regarded as representative HCC stem cells.⁵³ These results are consistent with those of studies on breast and colon cancer.^{6,45} The NF- κ B signaling pathway has been identified as a key regulator of EMT in a variety of cancers.^{54,55} Intriguingly, Ma et al obtained similar results and found that *BMII* knockdown inhibits the metastasis of CD133⁺ Huh7 cells by inhibiting the NF- κ B pathway.²⁹ In addition, Twist2, a primary helix-loop-helix transcription factor, regulates the progression of EMT in many cancers.^{56–58} A previous study has indicated that Twist2-expressing HepG2 cells exhibit an increase in the expression of the stem cell marker *BMII*.⁵⁹ Therefore, further research is necessary to identify the links between the *BMII* proto-oncogene and EMT in HCC.

Recently, new mechanisms by which *BMII* affects the invasion and metastasis of cancers have been reported. *BMII* negatively regulates RKIP expression via miR-27a and miR-155, thus promoting tumor metastasis and chemoresistance.¹⁰ These findings provide new directions for research on the mechanisms mediating the effect of *BMII* on HCC progression.

High *BMII* Expression Effectively Indicates Prognosis in HCC

Consistent with the association between *BMII* and prognosis in other cancers,^{9,60} high *BMII* levels are related to relatively poor outcomes in HCC, indicating that *BMII* is a reliable prognostic factor.¹⁹ It is striking that *BMII* knockdown can enhance the sensitivity of HCC cells to sorafenib and 5-fluorouracil, which are promising and frequently used drugs for HCC.^{61,62} Wang et al found that the 5-year survival rate of patients with HCC from China with low *BMII* expression is 46%, compared with only 15% for the patients with high *BMII* expression after routine surgery without prior radiotherapy or chemotherapy.²⁰ There was an approximately three-fold gap in the survival rate between the two groups, which was quite substantial. Of note, there was no significant difference in the survival rate of patients with high *BMII* levels and those with low *BMII* levels in 2 years, as determined by a Kaplan–Meier curve analysis.²⁰ These findings suggest that *BMII* is an important biomarker for predicting the long-term survival of patients with HCC. However, Yonemitsu et al concluded that the expression level of *BMII* had no apparent effect on the survival of patients with HCC after resection in Japan, although high *BMII* protein levels were correlated with the recurrence rate after hepatectomy.⁶³ These inconsistent results imply that the prognostic value of *BMII* expression in HCC patients differs among regions/territories. A meta-analysis suggested that high *BMII* expression predicts an adverse overall survival rate for cancer patients in Asian populations and as a favorable predictor in Caucasian populations.⁶⁴ Another meta-analysis also suggested that high *BMII* expression is associated with lower overall survival rates in patients with colorectal cancer in Asia and is associated with higher overall survival rates in European populations.⁶⁵

Overview of HCC Therapy Targeting *BMII*

The link between *BMII* and the initiation and progression of HCC is well-established. Therefore, several researchers have investigated the potential of *BMII* as a target for the treatment of HCC (Table 1).

Previous studies have identified *BMII* inhibitors, including PTC-209, PTC-028, PTC-596, and QW24. PTC-209 was the first reported *BMII* inhibitor. It is a low-molecular-weight compound identified by high-throughput screening using the GEMS (Gene Expression Modulation by Small Molecules) platform.⁶⁶ Kreso et al demonstrated that PTC-209 decreases *BMII* expression at the protein level and causes the loss of tumor-initiating cells in primary colorectal cancer as well as irreversible intra-tumor injury.⁶⁶ PTC-028 decreases *BMII* function via hyperphosphorylation and induces apoptosis in epithelial ovarian cancer cells.⁶⁷ Likewise, PTC-596, identified as a novel *BMII* inhibitor, can promote the apoptosis of acute myeloid leukemia progenitor cells.⁶⁸ Wang et al demonstrated that QW24, a small-molecule inhibitor against *BMII*, could reduce the stability of the *BMII* protein and downregulate *BMII* expression via the autophagy-lysosome pathway, without affecting *BMII* mRNA levels in colorectal cancer cells.⁶⁹ Bartucci et al identified *BMII* inhibitors, named RU-A1 compounds, by medicinal chemistry and chemical engineering, which can inhibit HCC cell growth, chemosensitivity, and tumor-initiating capacity, with more effective tumor-suppressive effects than those of PTC-209.⁷⁰ These *BMII* inhibitors may be effective therapeutic agents for cancer in the future. However, *BMII* inhibitors have not been used in HCC clinical treatment to date and the efficacy of these inhibitors against HCC is not clear.

MicroRNAs are a group of endogenous noncoding single-stranded RNAs that negatively regulate the expression of target genes.⁷¹ MiR-218 has suppressive effects in many cancers.^{72–74} Furthermore, accumulating evidence indicates that miR-218 negatively regulates *BMII* expression in HCC^{14,75,76} and is an independent prognostic factor for

HCC.⁷⁷ Hence, reducing the expression of *BMII* by increasing the expression of miR-218 may be an effective treatment strategy for HCC. Fu et al found that 1,6,7-trihydroxyxanthone (THA), an active ingredient derived from *Goodyera oblongifolia*, could increase miR-218 expression and decrease *BMII* expression, thereby strongly inhibiting cancer cell growth and inducing abnormal cell apoptosis in HCC.⁷⁶ Similarly, some studies have demonstrated that miR-203 has suppressive effects in numerous tumors.⁷⁸ A study has shown that miR-203 levels are lower in HCC tissue samples than in normal tissues and the overexpression of miR-203 could suppress the proliferation and invasion of HCC cells.⁷⁹ Interestingly, miR-203 can enhance the radiosensitivity of HCC cells by directly targeting *BMII*.⁸⁰ Therefore, miR-203 may be a valid therapeutic target for HCC, as it inhibits *BMII* expression. Recently, some progress has been made in the elucidation of miR-203-based regulation. For example, luteolin, a natural flavonoid, possesses anti-breast cancer properties, as it upregulates miR-203 expression.⁸¹ Strikingly, sevoflurane, an inhalational anesthetic, suppresses cell proliferation in breast cancer and migration in colorectal cancer by upregulating miR-203 expression.^{82,83} In addition, *BMII* is a well-known target gene of the miR-200 family, which plays an essential role in cancer progression and metastasis.^{84,85} A recent study has proven that miR-200c suppresses the initiation of HCC, at least in part by inhibiting *BMII* expression.⁸⁶ Interestingly, the short-chain fatty acid sodium butyrate (NaB) produced by bacterial fermentation of dietary fiber in the colon limits colorectal cancer liver metastasis by downregulating *BMII* expression by enhanced miR-200c expression.⁸⁷ These results provide potential therapeutic agents for HCC.

Many studies have inspired new ideas for the development of HCC treatments targeting *BMII*. Jiedu Xiaozheng Yin (JXY), a Chinese herbal decoction, inhibits HCC cell growth by suppressing *BMII* expression.⁸⁸ Kaneta et al identified that wallichoside, a methanol-soluble extract from *Beaumontia*

Table 1. The Overview of Potential Therapeutic Strategies Targeting *BMII* in HCC Treatment.

Therapeutic strategies targeting <i>BMII</i>	Category	Reference
Compounds inhibiting <i>BMII</i> expression	PTC-209	66
	PTC-028	67
	PTC-596	68
	QW24	69
	RU-A1	70
Increasing the expression of MicroRNAs downregulating <i>BMII</i> level	1,6,7-trihydroxyxanthone(THA) upregulating miR-218	76
	Luteolin and sevoflurane upregulating miR-203	81 to 83
Natural extracts suppressing <i>BMII</i> expression	Short-chain fatty acid sodium butyrate (NaB) upregulating miR-200c	87
	Jiedu Xiaozheng Yin (JXY)	88
	Wallichoside	89
Combinations of chemotherapeutical agent and <i>BMII</i> - <i>siRNA</i>	Nanoplatin and siRNA co-loaded CaP nanoparticles (NPSC) delivering cisplatin and <i>BMII</i> - <i>siRNA</i>	92
	Nanocapsules delivering <i>BMII</i> - <i>siRNA</i> and cisplatin cationic	93
Genome editing	Clustered regularly interspaced short palindromic repeats and CRISPR associated protein-9(CRISPR-Cas9)	97 to 99

murtonii and *Eugenia operculate*, suppressed *BMII* promoter activity and inhibited the growth of Huh7 human hepatocellular carcinoma cells.⁸⁹ Small interfering RNAs (siRNAs) can silence specific cancer genes; thus, siRNA therapeutics are very promising and are a focus of cancer research.⁹⁰ Cisplatin (cis-dichlorodiamminoplatinum [II]; CDDP) is an anticancer agent used widely in HCC treatment.⁹¹ Li et al designed Nanoplatin and siRNA co-loaded CaP nanoparticles to deliver the chemotherapeutic agent CDDP and *BMII* siRNA to effectively and safely treat HCC; the upregulation of *BMII* expression under CDDP-exposure contributed to drug resistance in HCC cell lines.⁹² Yang et al found that *BMII* siRNA and cisplatin cationic nanocapsules delivered to HCC-bearing mice had higher anti-tumor activity than that of mice treated with nanocapsules alone.⁹³

Genome editing is an emerging area of research because of its ability to precisely alter genomes of target cells and regulate functional genes using targeted nucleases. Clustered regularly interspaced short palindromic repeats and CRISPR associated protein-9 (CRISPR-Cas9) acts as “molecular scissors” and is quicker, more precise, and more effective than alternative genome-editing strategies.⁹⁴ CRISPR-Cas9 has also shown great potential in the treatment of cancer, including HCC.^{95,96} Wang et al reported that the knockout of CXC chemokine receptor 4 (CXCR4), associated with a poor prognosis in HCC, by CRISPR/Cas9 not only suppresses proliferation and invasion but also increases sensitivity to the anticancer drug cisplatin in HCC.⁹⁷ Another study has shown that the CRISPR/Cas9-mediated knockout of nuclear receptor binding SET domain-containing protein 1 inhibits the ability of HCC cells to proliferate and migrate.⁹⁸ In addition, Zhao et al applied the CRISPR/Cas9 technique to knock out *BMII* in epithelial ovarian cancer cells and found that the knockout of *BMII* could promote cell apoptosis and reduce cell growth and metastasis.⁹⁹ These results suggest that CRISPR/Cas9-mediated *BMII* knockout is feasible for the treatment of HCC, and relevant research is urgently needed.

Conclusion

BMII is a key gene in tumorigenesis and contributes to the development of many cancers, making it a well-established therapeutic target. In this review, we introduced the characteristics of *BMII* expression in the occurrence and development of HCC and briefly summarized the mechanisms by which *BMII* influences disease progression. Accumulating evidence shows that in some cancers, suppressing the expression and activity of *BMII* by direct inhibition or by suppressing upstream factors is a highly efficient treatment strategy. In addition, considering the biological availability of *BMII* inhibitors in clinical application, innovative drug combinations and agents have also shown promising results. Although such inhibitors are not widely used in clinical practice, there is sufficient evidence that *BMII* is an effective therapeutic target for HCC. Therefore, further studies to elucidate the basic mechanism of action of *BMII* in HCC and its clinical correlations are needed.

Declaration of Conflicting Interests


The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This study was sponsored by the National Natural Science Foundation of China (grant number: 82071964), Shanghai Leading Talent Program funded by Shanghai Human Resources and Social Security Bureau (grant number: 03.05.19005), Shanghai Shengkang Three-year Action Project (grant number: SHDC2020CR2054B), Shanghai Tenth People’s Hospital (grant number: 2021SYPDR065 and 04.01.20065), and Shanghai Municipal Health Commission (grant number: 202040043; GWV-10.1-XK09 and 20214Y0159).

ORCID iDs

Ru Wang  <https://orcid.org/0000-0002-7939-7359>

Hengwei Fan  <https://orcid.org/0000-0002-4761-8217>

References

- Llovet JM, Kelley RK, Villanueva A, et al. Hepatocellular carcinoma. *Nat Rev Dis Primers*. 2021;7(1):6. doi:10.1038/s41572-020-00240-3
- European Association for the Study of the Liver. EASL Clinical practice guidelines: management of hepatocellular carcinoma. *J Hepatol*. 2018;69(1):182-236. doi:10.1016/j.jhep.2018.03.019
- Simon JA, Kingston RE. Mechanisms of polycomb gene silencing: knowns and unknowns. *Nat Rev Mol Cell Biol*. 2009;10(10):697-708. doi:10.1038/nrm2763
- van Lohuizen M, Verbeek S, Scheijen B, et al. Identification of cooperating oncogenes in E mu-myc transgenic mice by provirus tagging. *Cell*. 1991;65(5):737-752. doi:10.1016/0092-8674(91)90382-9
- Haupt Y, Alexander WS, Barri G, et al. Novel zinc finger gene implicated as myc collaborator by retrovirally accelerated lymphomagenesis in E mu-myc transgenic mice. *Cell*. 1991;65(5):753-763. doi:10.1016/0092-8674(91)90383-a
- Guo BH, Feng Y, Zhang R, et al. Bmi-1 promotes invasion and metastasis, and its elevated expression is correlated with an advanced stage of breast cancer. *Mol Cancer*. 2011;10(1):10. doi:10.1186/1476-4598-10-10
- Li DW, Tang HM, Fan JW, et al. Expression level of Bmi-1 oncoprotein is associated with progression and prognosis in colon cancer. *J Cancer Res Clin Oncol*. 2010;136(7):997-1006. doi:10.1007/s00432-009-0745-7
- Kang MK, Kim RH, Kim SJ, et al. Elevated Bmi-1 expression is associated with dysplastic cell transformation during oral carcinogenesis and is required for cancer cell replication and survival. *Br J Cancer*. 2007;96(1):126-133. doi:10.1038/sj.bjc.6603529
- Yu J, Chen L, Bao Z, et al. BMI-1 promotes invasion and metastasis in endometrial adenocarcinoma and is a poor prognostic factor. *Oncol Rep*. 2020;43(5):1630-1640. doi:10.3892/or.2020.7539

10. Li Y, Tian Z, Tan Y, et al. Bmi-1-induced miR-27a and miR-155 promote tumor metastasis and chemoresistance by targeting RKIP in gastric cancer. *Mol Cancer*. 2020;19(1):109. doi:10.1186/s12943-020-01229-y
11. Neo SY, Leow CK, Vega VB, et al. Identification of discriminators of hepatoma by gene expression profiling using a minimal dataset approach. *Hepatology*. 2004;39(4):944-953. doi:10.1002/hep.20105
12. Yu J, Hu M, Chen W, et al. Expression of B cell-specific moloney murine leukemia virus integration site 1 (BMI-1) and WW domain-containing oxidoreductase (WWOX) in liver cancer tissue and normal liver tissue. *Med Sci Monit*. 2018;9(24):6673-6679. doi:10.12659/msm.909675
13. Ma S, Chan KW, Hu L, et al. Identification and characterization of tumorigenic liver cancer stem/progenitor cells. *Gastroenterology*. 2007;132(7):2542-2556. doi:10.1053/j.gastro.2007.04.025
14. Tu K, Li C, Zheng X, et al. Prognostic significance of miR-218 in human hepatocellular carcinoma and its role in cell growth. *Oncol Rep*. 2014;32(4):1571-1577. doi:10.3892/or.2014.3386
15. Breuer RH, Snijders PJ, Smit EF, et al. Increased expression of the EZH2 polycomb group gene in BMI-1-positive neoplastic cells during bronchial carcinogenesis. *Neoplasia*. 2004;6(6):736-743. doi:10.1593/neo.04160
16. Tateishi K, Ohta M, Kanai F, et al. Dysregulated expression of stem cell factor Bmi1 in precancerous lesions of the gastrointestinal tract. *Clin Cancer Res*. 2006;12(23):6960-6966. doi:10.1158/1078-0432.Ccr-06-0449
17. Sasaki M, Ikeda H, Itatsu K, et al. The overexpression of polycomb group proteins Bmi1 and EZH2 is associated with the progression and aggressive biological behavior of hepatocellular carcinoma. *Lab Invest*. 2008;88(8):873-882. doi:10.1038/labinvest.2008.52
18. Effendi K, Mori T, Komuta M, et al. Bmi-1 gene is upregulated in early-stage hepatocellular carcinoma and correlates with ATP-binding cassette transporter B1 expression. *Cancer Sci*. 2010;101(3):666-672. doi:10.1111/j.1349-7006.2009.01431.x
19. Li X, Yang Z, Song W, et al. Overexpression of Bmi-1 contributes to the invasion and metastasis of hepatocellular carcinoma by increasing the expression of matrix metalloproteinase (MMP)-2, MMP-9 and vascular endothelial growth factor via the PTEN/PI3 K/Akt pathway. *Int J Oncol*. 2013;43(3):793-802. doi:10.3892/ijo.2013.1992
20. Wang H, Pan K, Zhang HK, et al. Increased polycomb-group oncogene Bmi-1 expression correlates with poor prognosis in hepatocellular carcinoma. *J Cancer Res Clin Oncol*. 2008;134(5):535-541. doi:10.1007/s00432-007-0316-8
21. Wicha MS, Liu S, Dontu G. Cancer stem cells: an old idea--a paradigm shift. *Cancer Res*. 2006;66(4):1883-1890; discussion 1895-6. doi:10.1158/0008-5472.Can-05-3153
22. Park IK, Qian D, Kiel M, et al. Bmi-1 is required for maintenance of adult self-renewing haematopoietic stem cells. *Nature*. 2003;423(6937):302-305. doi:10.1038/nature01587
23. Lessard J, Sauvageau G. Bmi-1 determines the proliferative capacity of normal and leukaemic stem cells. *Nature*. 2003;423(6937):255-260. doi:10.1038/nature01572
24. Fan L, Xu C, Wang C, et al. Bmi1 is required for hepatic progenitor cell expansion and liver tumor development. *PLoS One*. 2012;7(9):e46472. doi:10.1371/journal.pone.0046472
25. Zhang R, Wu WR, Shi XD, et al. Dysregulation of Bmi1 promotes malignant transformation of hepatic progenitor cells. *Oncogenesis*. 2016;5(2):e203. doi:10.1038/oncsis.2016.6
26. Chiba T, Kita K, Zheng YW, et al. Side population purified from hepatocellular carcinoma cells harbors cancer stem cell-like properties. *Hepatology*. 2006;44(1):240-251. doi:10.1002/hep.21227
27. Chiba T, Miyagi S, Saraya A, et al. The polycomb gene product BMI1 contributes to the maintenance of tumor-initiating side population cells in hepatocellular carcinoma. *Cancer Res*. 2008;68(19):7742-7749. doi:10.1158/0008-5472.Can-07-5882
28. Chiba T, Zheng YW, Kita K, et al. Enhanced self-renewal capability in hepatic stem/progenitor cells drives cancer initiation. *Gastroenterology*. 2007;133(3):937-950. doi:10.1053/j.gastro.2007.06.016
29. Ma DQ, Zhang YH, Ding DP, et al. Effect of Bmi-1-mediated NF- κ B signaling pathway on the stem-like properties of CD133⁺ human liver cancer cells. *Cancer Biomark*. 2018;22(3):575-585. doi:10.3233/cbm-181329
30. Sparmann A, van Lohuizen M. Polycomb silencers control cell fate, development and cancer. *Nat Rev Cancer*. 2006;6(11):846-856. doi:10.1038/nrc1991
31. Park IK, Morrison SJ, Clarke MF. Bmi1, stem cells, and senescence regulation. *J Clin Invest*. 2004;113(2):175-179. doi:10.1172/jci20800
32. Jacobs JJ, Kieboom K, Marino S, et al. The oncogene and polycomb-group gene bmi-1 regulates cell proliferation and senescence through the ink4a locus. *Nature*. 1999;397(6715):164-168. doi:10.1038/16476
33. Kent LN, Leone G. The broken cycle: e2F dysfunction in cancer. *Nat Rev Cancer*. 2019;19(6):326-338. doi:10.1038/s41568-019-0143-7
34. Gil J, Peters G. Regulation of the INK4b-ARF-INK4a tumour suppressor locus: all for one or one for all. *Nat Rev Mol Cell Biol*. 2006;7(9):667-677. doi:10.1038/nrm1987
35. Kanapathipillai M. Treating p53 mutant aggregation-associated cancer. *Cancers (Basel)*. 2018;10(6):154. doi:10.3390/cancers10060154
36. Fasano CA, Dimos JT, Ivanova NB, et al. shRNA knockdown of Bmi-1 reveals a critical role for p21-Rb pathway in NSC self-renewal during development. *Cell Stem Cell*. 2007;1(1):87-99. doi:10.1016/j.stem.2007.04.001
37. Bruggeman SW, Hulsman D, Tanger E, et al. Bmi1 controls tumor development in an Ink4a/Arf-independent manner in a mouse model for glioma. *Cancer Cell*. 2007;12(4):328-341. doi:10.1016/j.ccr.2007.08.032
38. Xu CR, Lee S, Ho C, et al. Bmi1 functions as an oncogene independent of Ink4A/Arf repression in hepatic carcinogenesis. *Mol Cancer Res*. 2009;7(12):1937-1945. doi:10.1158/1541-7786.Mcr-09-0333
39. Chiba T, Seki A, Aoki R, et al. Bmi1 promotes hepatic stem cell expansion and tumorigenicity in both Ink4a/Arf-dependent and -independent manners in mice. *Hepatology*. 2010;52(3):1111-1123. doi:10.1002/hep.23793

40. Lu SC, Alvarez L, Huang ZZ, et al. Methionine adenosyltransferase 1A knockout mice are predisposed to liver injury and exhibit increased expression of genes involved in proliferation. *Proc Natl Acad Sci U S A*. 2001;98(10):5560-5565. doi:10.1073/pnas.091016398
41. Cancer Genome Atlas Research Network. Comprehensive and integrative genomic characterization of hepatocellular carcinoma. *Cell*. 2017;169(7):1327-41.e23. doi:10.1016/j.cell.2017.05.046
42. Fu WM, Zhu X, Wang WM, et al. Hotair mediates hepatocarcinogenesis through suppressing miRNA-218 expression and activating P14 and P16 signaling. *J Hepatol*. 2015;63(4):886-895. doi:10.1016/j.jhep.2015.05.016
43. Li B, Chen Y, Wang F, et al. Bmi1 drives hepatocarcinogenesis by repressing the TGFβ2/SMAD signalling axis. *Oncogene*. 2020;39(5):1063-1079. doi:10.1038/s41388-019-1043-8
44. Chen MH, Fu LS, Zhang F, et al. LncAY controls BMI1 expression and activates BMI1/Wnt/β-catenin signaling axis in hepatocellular carcinoma. *Life Sci*. 2021;9 (280) :119748. doi:10.1016/j.lfs.2021.119748
45. Zhang Z, Bu X, Chen H, et al. Bmi-1 promotes the invasion and migration of colon cancer stem cells through the downregulation of E-cadherin. *Int J Mol Med*. 2016;38(4):1199-1207. doi:10.3892/ijmm.2016.2730
46. Gao FL, Li WS, Liu CL, et al. Silencing Bmi-1 enhances the senescence and decreases the metastasis of human gastric cancer cells. *World J Gastroenterol*. 2013;19(46):8764-8769. doi:10.3748/wjg.v19.i46.8764
47. Su Z, Yang Z, Xu Y, et al. Apoptosis, autophagy, necroptosis, and cancer metastasis. *Mol Cancer*. 2015;2 (14) :48. doi:10.1186/s12943-015-0321-5
48. Chalhoub N, Baker SJ. PTEN And the PI3-kinase pathway in cancer. *Annu Rev Pathol*. 2009;7 (4) :127-150. doi:10.1146/annurev.pathol.4.110807.092311
49. Stambolic V, Suzuki A, de la Pompa JL, et al. Negative regulation of PKB/Akt-dependent cell survival by the tumor suppressor PTEN. *Cell*. 1998;95(1):29-39. doi:10.1016/s0092-8674(00)81780-8
50. Chen JS, Wang Q, Fu XH, et al. Involvement of PI3 K/PTEN/AKT/mTOR pathway in invasion and metastasis in hepatocellular carcinoma: association with MMP-9. *Hepatol Res*. 2009;39(2):177-186. doi:10.1111/j.1872-034X.2008.00449.x
51. Cheng JC, Chou CH, Kuo ML, et al. Radiation-enhanced hepatocellular carcinoma cell invasion with MMP-9 expression through PI3 K/Akt/NF-kappaB signal transduction pathway. *Oncogene*. 2006;25(53):7009-7018. doi:10.1038/sj.onc.1209706
52. Onder TT, Gupta PB, Mani SA, et al. Loss of E-cadherin promotes metastasis via multiple downstream transcriptional pathways. *Cancer Res*. 2008;68(10):3645-3654. doi:10.1158/0008-5472.Can-07-2938
53. Zhang Z, Wang Q, Bu X, et al. Overexpression of Bmi-1 promotes epithelial-mesenchymal transition in CD133 + Hep G2 cells. *Mol Med Rep*. 2017;16(5):6156-6161. doi:10.3892/mmr.2017.7347
54. Lin C, Zong J, Lin W, et al. EBV-miR-BART8-3p induces epithelial-mesenchymal transition and promotes metastasis of nasopharyngeal carcinoma cells through activating NF-κB and Erk1/2 pathways. *J Exp Clin Cancer Res*. 2018;37(1):283. doi:10.1186/s13046-018-0953-6
55. Hseu YC, Lin YC, Rajendran P, et al. Antrodia salmonea suppresses invasion and metastasis in triple-negative breast cancer cells by reversing EMT through the NF-κB and Wnt/β-catenin signaling pathway. *Food Chem Toxicol*. 2019;2 (124) :219-230. doi:10.1016/j.fct.2018.12.009
56. Fang X, Cai Y, Liu J, et al. Twist2 contributes to breast cancer progression by promoting an epithelial-mesenchymal transition and cancer stem-like cell self-renewal. *Oncogene*. 2011;30(47):4707-4720. doi:10.1038/onc.2011.181
57. Li Y, Wang W, Wang W, et al. Correlation of TWIST2 up-regulation and epithelial-mesenchymal transition during tumorigenesis and progression of cervical carcinoma. *Gynecol Oncol*. 2012;124(1):112-118. doi:10.1016/j.ygyno.2011.09.003
58. Yang J, Zhang X, Zhang Y, et al. HIF-2α promotes epithelial-mesenchymal transition through regulating Twist2 binding to the promoter of E-cadherin in pancreatic cancer. *J Exp Clin Cancer Res*. 2016;2(35):26. doi:10.1186/s13046-016-0298-y
59. Liu AY, Cai Y, Mao Y, et al. Twist2 promotes self-renewal of liver cancer stem-like cells by regulating CD24. *Carcinogenesis*. 2014;35(3):537-545. doi:10.1093/carcin/bgt364
60. Peng HX, Liu XD, Luo ZY, et al. Upregulation of the proto-oncogene Bmi-1 predicts a poor prognosis in pediatric acute lymphoblastic leukemia. *BMC Cancer*. 2017;17(1):76. doi:10.1186/s12885-017-3049-3
61. Ruan ZP, Xu R, Lv Y, et al. Bmi1 knockdown inhibits hepatocarcinogenesis. *Int J Oncol*. 2013;42(1):261-268. doi:10.3892/ijo.2012.1693
62. Zhang R, Xu LB, Yue XJ, et al. Bmi1 gene silencing inhibits the proliferation and invasiveness of human hepatocellular carcinoma cells and increases their sensitivity to 5-fluorouracil. *Oncol Rep*. 2013;29(3):967-974. doi:10.3892/or.2012.2189
63. Yonemitsu Y, Imazeki F, Chiba T, et al. Distinct expression of polycomb group proteins EZH2 and BMI1 in hepatocellular carcinoma. *Hum Pathol*. 2009;40(9):1304-1311. doi:10.1016/j.humpath.2009.01.017
64. Shao Y, Geng Y, Gu W, et al. Prognostic role of high Bmi-1 expression in asian and caucasian patients with solid tumors: a meta-analysis. *Biomed Pharmacother*. 2014;68(8):969-977. doi:10.1016/j.biopha.2014.10.017
65. Pourjafar M, Samadi P, Karami M, et al. Assessment of clinicopathological and prognostic relevance of BMI-1 in patients with colorectal cancer: a meta-analysis. *Biotechnol Appl Biochem*. 2021;68(6):1313-1322. doi:10.1002/bab.2053
66. Kreso A, van Galen P, Pedley NM, et al. Self-renewal as a therapeutic target in human colorectal cancer. *Nat Med*. 2014;20(1):29-36. doi:10.1038/nm.3418
67. Dey A, Xiong X, Crim A, et al. Evaluating the mechanism and therapeutic potential of PTC-028, a novel inhibitor of BMI-1 function in ovarian cancer. *Mol Cancer Ther*. 2018;17(1):39-49. doi:10.1158/1535-7163.Mct-17-0574
68. Nishida Y, Maeda A, Kim MJ, et al. The novel BMI-1 inhibitor PTC596 downregulates MCL-1 and induces p53-independent mitochondrial apoptosis in acute myeloid leukemia progenitor cells. *Blood Cancer J*. 2017;7(2):e527. doi:10.1038/bcj.2017.8

69. Wang J, Xing Y, Wang Y, et al. A novel BMI-1 inhibitor QW24 for the treatment of stem-like colorectal cancer. *J Exp Clin Cancer Res.* 2019;38(1):422. doi:10.1186/s13046-019-1392-8
70. Bartucci M, Hussein MS, Huselid E, et al. Synthesis and characterization of novel BMI1 inhibitors targeting cellular self-renewal in hepatocellular carcinoma. *Target Oncol.* 2017;12(4):449-462. doi:10.1007/s11523-017-0501-x
71. Ambros V. The functions of animal microRNAs. *Nature.* 2004;431(7006):350-355. doi:10.1038/nature02871
72. Yang Y, Ding L, Hu Q, et al. MicroRNA-218 functions as a tumor suppressor in lung cancer by targeting IL-6/STAT3 and negatively correlates with poor prognosis. *Mol Cancer.* 2017;16(1):141. doi:10.1186/s12943-017-0710-z
73. Liu Z, Mao L, Wang L, et al. Mir-218 functions as a tumor suppressor gene in cervical cancer. *Mol Med Rep.* 2020;21(1):209-219. doi:10.3892/mmr.2019.10809
74. Wang T, Xu L, Jia R, et al. MiR-218 suppresses the metastasis and EMT of HCC cells via targeting SERBP1. *Acta Biochim Biophys Sin (Shanghai).* 2017;49(5):383-391. doi:10.1093/abbs/gmx017
75. Wu J, Jiang ZM, Xie Y, et al. miR-218 suppresses the growth of hepatocellular carcinoma by inhibiting the expression of proto-oncogene Bmi-1. *J Buon.* 2018;23(3):604-610.
76. Fu WM, Tang LP, Zhu X, et al. MiR-218-targeting-Bmi-1 mediates the suppressive effect of 1,6,7-trihydroxyxanthone on liver cancer cells. *Apoptosis.* 2015;20(1):75-82. doi:10.1007/s10495-014-1047-3
77. Tu K, Li C, Zheng X, et al. Prognostic significance of miR-218 in human hepatocellular carcinoma and its role in cell growth. *Oncol Rep.* 2014;32(4):1571-1577. doi:10.3892/or.2014.3386
78. Du SL, Xu LY, Gao P, et al. MiR-203 regulates DJ-1 expression and affects proliferation, apoptosis and DDP resistance of pancreatic cancer cells. *Eur Rev Med Pharmacol Sci.* 2019;23(20):8833-8840. doi:10.26355/eurrev_201910_19278
79. Yang F, Lv LZ, Cai QC, et al. Potential roles of EZH2, Bmi-1 and miR-203 in cell proliferation and invasion in hepatocellular carcinoma cell line Hep3B. *World J Gastroenterol.* 2015;21(47):13268-13276. doi:10.3748/wjg.v21.i47.13268
80. Shao Y, Zhang D, Li X, et al. MicroRNA-203 increases cell radiosensitivity via directly targeting Bmi-1 in hepatocellular carcinoma. *Mol Pharm.* 2018;15(8):3205-3215. doi:10.1021/acs.molpharmaceut.8b00302
81. Gao G, Ge R, Li Y, et al. Luteolin exhibits anti-breast cancer property through up-regulating miR-203. *Artif Cells Nanomed Biotechnol.* 2019;47(1):3265-3271. doi:10.1080/21691401.2019.1646749
82. Liu J, Yang L, Guo X, et al. Sevoflurane suppresses proliferation by upregulating microRNA-203 in breast cancer cells. *Mol Med Rep.* 2018;18(1):455-460. doi:10.3892/mmr.2018.8949
83. Fan L, Wu Y, Wang J, et al. Sevoflurane inhibits the migration and invasion of colorectal cancer cells through regulating ERK/MMP-9 pathway by up-regulating miR-203. *Eur J Pharmacol.* 2019;5(850):43-52. doi:10.1016/j.ejphar.2019.01.025
84. Liu Y, Sánchez-Tilló E, Lu X, et al. The ZEB1 transcription factor acts in a negative feedback loop with miR200 downstream of Ras and Rb1 to regulate Bmi1 expression. *J Biol Chem.* 2014;289(7):4116-4125. doi:10.1074/jbc.M113.533505
85. Xu Z, Tao J, Chen P, et al. Sodium butyrate inhibits colorectal cancer cell migration by downregulating Bmi-1 through enhanced miR-200c expression. *Mol Nutr Food Res.* 2018;62(6):e1700844. doi:10.1002/mnfr.201700844
86. Xu L, Lin J, Deng W, et al. EZH2 Facilitates BMI1-dependent hepatocarcinogenesis through epigenetically silencing microRNA-200c. *Oncogenesis.* 2020;9(11):101. doi:10.1038/s41389-020-00284-w
87. Xu Z, Zhou Z, Zhang J, et al. Targeting BMI-1-mediated epithelial-mesenchymal transition to inhibit colorectal cancer liver metastasis. *Acta Pharm Sin B.* 2021;11(5):1274-1285. doi:10.1016/j.apsb.2020.11.018
88. Chen XZ, Cao ZY, Li JN, et al. Ethyl acetate extract from jiedu Xiaozheng Yin inhibits the proliferation of human hepatocellular carcinoma cells by suppressing polycomb gene product Bmi1 and Wnt/ β -catenin signaling. *Oncol Rep.* 2014;32(6):2710-2718. doi:10.3892/or.2014.3541
89. Kaneta Y, Arai MA, Ishikawa N, et al. Identification of BMI1 promoter inhibitors from *Beaumontia murtonii* and *Eugenia operculata*. *J Nat Prod.* 2017;80(6):1853-1859. doi:10.1021/acs.jnatprod.7b00138
90. Subhan MA, Torchilin VP. Efficient nanocarriers of siRNA therapeutics for cancer treatment. *Transl Res.* 2019;12(214):62-91. doi:10.1016/j.trsl.2019.07.006
91. Lee JE, Bae SH, Choi JY, et al. Epirubicin, cisplatin, 5-FU combination chemotherapy in sorafenib-refractory metastatic hepatocellular carcinoma. *World J Gastroenterol.* 2014;20(1):235-241. doi:10.3748/wjg.v20.i1.235
92. Li M, Zhao P, Fan T, et al. Biocompatible co-loading vehicles for delivering both nanoplatin cores and siRNA to treat hepatocellular carcinoma. *Int J Pharm.* 2019;12 (572) :118769. doi:10.1016/j.ijpharm.2019.118769
93. Yang T, Chen Y, Zhao P, et al. Enhancing the therapeutic effect via elimination of hepatocellular carcinoma stem cells using Bmi1 siRNA delivered by cationic cisplatin nanocapsules. *Nanomedicine.* 2018;14(7):2009-2021. doi:10.1016/j.nano.2018.05.012
94. Lino CA, Harper JC, Carney JP, et al. Delivering CRISPR: a review of the challenges and approaches. *Drug Deliv.* 2018;25(1):1234-1257. doi:10.1080/10717544.2018.1474964
95. Daisy PS, Shreyas KS, Anitha TS. Will CRISPR-Cas9 have cards to play against cancer? An update on its applications. *Mol Biotechnol.* 2021;63(2):93-108. doi:10.1007/s12033-020-00289-1
96. Wu X, Ma W, Mei C, et al. Description of CRISPR/Cas9 development and its prospect in hepatocellular carcinoma treatment. *J Exp Clin Cancer Res.* 2020;39(1):97. doi:10.1186/s13046-020-01603-0
97. Wang X, Zhang W, Ding Y, et al. CRISPR/Cas9-mediated genome engineering of CXCR4 decreases the malignancy of hepatocellular carcinoma cells in vitro and in vivo. *Oncol Rep.* 2017;37(6):3565-3571. doi:10.3892/or.2017.5601
98. Zhang S, Zhang F, Chen Q, et al. CRISPR/Cas9-mediated knock-out of NSD1 suppresses the hepatocellular carcinoma development via the NSD1/H3/Wnt10b signaling pathway. *J Exp Clin Cancer Res.* 2019;38(1):467. doi:10.1186/s13046-019-1462-y
99. Zhao Q, Qian Q, Cao D, et al. Role of BMI1 in epithelial ovarian cancer: investigated via the CRISPR/Cas9 system and RNA sequencing. *J Ovarian Res.* 2018;11(1):31. doi:10.1186/s13048-018-0406-z