

## RESEARCH ARTICLE

# Identifying the snake: First scoping review on practices of communities and healthcare providers confronted with snakebite across the world

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## Abstract

### Background

Snakebite envenoming is a major global health problem that kills or disables half a million people in the world's poorest countries. Biting snake identification is key to understanding snakebite eco-epidemiology and optimizing its clinical management. The role of snakebite victims and healthcare providers in biting snake identification has not been studied globally.

### Objective

This scoping review aims to identify and characterize the practices in biting snake identification across the globe.

### Methods

Epidemiological studies of snakebite in humans that provide information on biting snake identification were systematically searched in *Web of Science* and *Pubmed* from inception to 2<sup>nd</sup> February 2019. This search was further extended by snowball search, hand searching literature reviews, and using Google Scholar. Two independent reviewers screened publications and charted the data.

### Results

We analysed 150 publications reporting 33,827 snakebite cases across 35 countries. On average 70% of victims/bystanders spotted the snake responsible for the bite and 38% captured/killed it and brought it to the healthcare facility. This practice occurred in 30 countries with both fast-moving, active-foraging as well as more secretive snake species. Methods for identifying biting snakes included snake body examination, victim/bystander biting snake description, interpretation of clinical features, and laboratory tests. In nine publications, a

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picture of the biting snake was taken and examined by snake experts. Snakes were identified at the species/genus level in only 18,065/33,827 (53%) snakebite cases. 106 misidentifications led to inadequate victim management. The 8,885 biting snakes captured and identified were from 149 species including 71 (48%) non-venomous species.

## Conclusion

Snakebite victims and healthcare providers can play a central role in biting snake identification and novel approaches (e.g. photographing the snake, crowdsourcing) could help increase biting snake taxonomy collection to better understand snake ecology and snakebite epidemiology and ultimately improve snakebite management.

## Introduction

An estimated 5.4 million snake bites occur globally every year. About half of these cause snakebite envenoming (SBE), killing 81,000–138,000 people and disabling 400,000 more in the poorest regions [1, 2]. In May 2019, the World Health Organization (WHO) launched a road map to halve these deaths and disabilities by 2030, particularly focusing on the development of antivenoms and on their adequate distribution in the most affected countries [3, 4]. For this, understanding what type of snakes and associated snake bites occur where is crucial, yet this depends first on the taxonomic identification (identification hereafter; i.e. family, genus and species) of these snakes [5–7]. At the clinical level, the identity of the biting snake (BSN) can help healthcare providers anticipate victims' syndromes and support decision making when treating the patient (i.e. whether or not to administer antivenom or which type of antivenom) [8, 9]. This decision is important because not only are antivenoms effective against a limited number of venomous snakes but they have potentially lethal side effects such as fatal allergic reactions and should not be used to treat bites from non-venomous snakes. This is especially important in view of the scarcity and high costs of antivenom vials in many countries.

However, identifying the BSN is challenging due to the high diversity of snake species [10] in snakebite endemic countries (e.g. 310 snake species in India), and the limited herpetological knowledge of communities and healthcare providers confronted with snakebite. In rural areas, traditional healers are often the first to be consulted for snakebite at the community level [11] and they could play a role in BSN identification, yet this remains to be assessed. Different behaviours and practices exist across the world depending on the development of health systems, local culture and perception of snakes and snakebite, and snake diversity. There is no standardized protocol for identifying BSNs and recommended practices are often specific to certain regions. For instance, the WHO's Regional Office for South-East Asia (2016) [12] recommends experts identify the BSN based on a photo (i.e. taken with a mobile phone) or the animal's body, when killed by the victim or bystanders and brought to the health facility.

To our knowledge, no review has systematically explored the literature to identify and synthesize the nature and extent of information available on the practices and challenges in BSN identification across the world. The objective of this scoping review is to provide a global and comprehensive description of the diversity of behaviours and practices of communities and healthcare providers, and the implications of these, when confronted with snakebite and the need to identify BSNs. This review also assesses the capabilities of these communities and healthcare providers to identify the BSN, and the frequency and consequences of

misidentifications. Finally, it addresses aspects related to the BSNs, including their diversity and behaviour, and how this may influence their identification.

## Materials and methods

We followed the scoping review methodology proposed by Arksey & O'Malley (2005) [13], Levac et al. (2010) [14], and Tricco et al. (2018) [15] (S1 File).

### Eligibility criteria

Original epidemiological studies and clinical case report/series reporting snakebite cases in humans caused by wild snakes and including information on their taxonomic identification were considered. Studies from all geographical areas and publication dates were eligible.

### Search strategy

*Web of Science* and *Pubmed* were searched from inception to February 2, 2019 using the following key words: snakebite, snake bite, snake envenoming, snake envenomation, case, victim, event, patient, biting snake, culprit, offending snake, species, identif\*, misidentif\*, unidentif\*, identity, and mistaking (see S1 Table for full search strategy). In addition to the database searches, we used extensive secondary search techniques, such as snowball search, hand searching literature reviews, and performing key word searches in Google Scholar. This secondary search was conducted in English, French, and Spanish by three of the authors (IB, SBM, AMD). With *PubMed*, we accessed primarily publications in the field of medicine and life sciences, while with *Web of Science* and Google Scholar we covered most scientific fields.

### Publication selection

Searched publications were merged using citation software EndNote X7 and duplicates were removed. IB, an expert in the domain of snakebite, screened all the titles and abstracts and excluded those publications that did not match eligibility criteria. Eligible publications were read by IB and SBM, and their relevance was further assessed, particularly focusing on information pertaining to the BSN (i.e. number of snakes identified and methods used). A final set of publications was produced by discussion and consensus between IB and SBM. No quality assessment of the publications selected was made.

### Charting the data

IB and SBM charted the data from selected publications independently. Their results were compared and discrepancies resolved through discussion and consensus. The variables extracted were: publication identifiers (authors, journal, year of publication, language); study characteristics (study design, country, setting, sample size); number of BSNs identified; number of victims that saw the BSN; number of BSNs captured or killed; taxonomic granularity of BSN identification (family, genus, or species); the way the BSNs were identified. We cross-referenced all common and scientific names of snakes reported in the publications with the Reptile Database [10] and kept track of changes in taxonomy where relevant. One of us (AMD) searched the literature for information on the activity time (diurnal/nocturnal/both), foraging strategy (active/passive/both), and general habitat (terrestrial, aquatic, arboreal, fossorial) of each species. Some species lacked species-specific information and their behaviour was extrapolated from those of congeners. We also gathered data on the number of snake species per country from the Reptile Database [10] and coded these as medically-important venomous snakes (MIVS) or not following the WHO. We used the `pcor.test` function in package `ppcor`

(v. 1.1) in R (v. 3.5.1) to estimate partial correlation coefficients among the number of BSN species per country, the total number of snake species in that country, the total number of snake bites reported in publications from that country, and the MIVS status of the snake species.

## Results

### Selection and characteristics of publications

A total of 467 unique publications resulted from the initial search, and 150 of these were included in the review (Fig 1, S2 Table). These publications covered 35 countries on all continents (Table 1) and were mostly retrospective or prospective hospital-based epidemiological studies (76%), conducted in Asia (50%), published after 2000 (77%) in English (94%). In total,

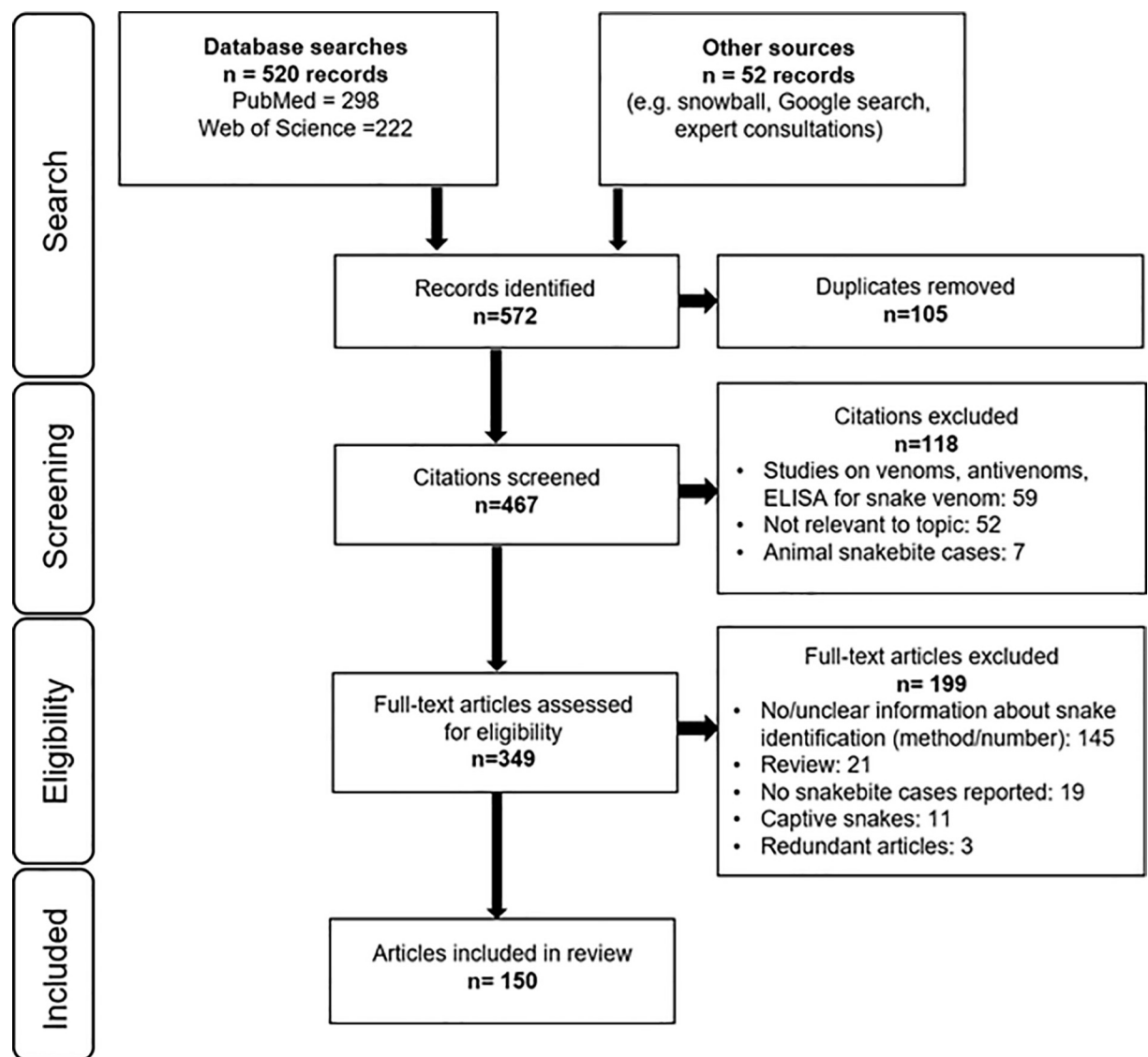


Fig 1. PRISMA flow diagram.

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**Table 1. General characteristics of included publications (number of publications = 150).**

Category		Count (%)
Type of study	Hospital-based retrospective study	58 (38.7)
	Hospital-based prospective study	56 (37.3)
	Case report or case-series	26 (17.3)
	Community-based survey	5 (3.3)
	Randomized controlled trial	2 (1.3)
	Mixed study design	3 (2.0)
Geographical Region <sup>a</sup>	Asia	74 (49.3)
	Central and South America	30 (20.0)
	Africa	19 (12.7)
	Australo-Papua	14 (9.3)
	North America	11 (7.3)
	Europe	2 (1.3)
Year of publication	1978–1979	1 (0.6)
	1980–1989	8 (5.3)
	1990–1999	26 (17.3)
	2000–2009	39 (26.0)
	2010–2019	76 (50.7)
Language	English	141 (94.0)
	French	4 (2.7)
	Spanish	3 (2.0)
	Portuguese	2 (1.3)

<sup>a</sup> Based on study site

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33,827 snakebite cases were reported in these studies and the median number of cases per study was 90–91 (range 1–3,411).

### Spotting the BSN

Out of the total, twenty-three publications (15%) across 13 countries mentioned the number of victims that saw the BSN. In total, 2,723 victims out of 3,865 (70.5%) have seen the BSN (range 51.9% to 93.2% across studies) (Table 2). Factors associated with the circumstances/context of the bite were often described for those cases where the snake was not spotted. This includes, for example, bites that occurred at night, assumed to be caused by nocturnal snakes (e.g. *Trimeresurus sp.*, *Bungarus sp.*) (e.g. Gabon, Hong Kong, Nepal, Saudi Arabia, Sri Lanka [16–20] or in habitats with tall grass and thick plantation vegetation (e.g. Papua New Guinea, Nepal, central hills of Sri Lanka [21–23]) or dense rain forest vegetation (e.g. Ecuador [24]).

### BSN captured/killed and brought to health facilities

A total of 114 (76%) publications in 30 countries described snakebite cases where victims or bystanders captured the BSN and brought it to the health facility (S2 Table). In 78 (68%) of these 114 publications and 24 countries, the BSN was killed. Globally, the BSN was captured/killed in 9,671/25,188 (38%) snakebite cases, but this practice varied among countries (Table 3). Cultural perceptions affected this behaviour. In Nepal, an Indian cobra (*Naja naja*) was not killed by the victim “in fear of revenge by its partner” [20]. In Sri Lanka “there may be a reluctance to capture or kill the animal because of fear or superstition” [44], and a snakebite victim “refrained from catching the snake due to religious ethics” [27]. In South Africa

Table 2. Number of snakebite cases where the BSN was spotted by the victims.

Country	References	Number of publications	Total snakebite cases	Number BSNs spotted	BSN spotted (%)
Gabon	[16]	1	27	14	51.9
Nepal	[20, 22, 25]	3	149	78	52.3
Sri Lanka	[26–28]	3	1036	567	54.7
Papua New Guinea	[21]	1	205	117	57.1
Australia	[29–31]	3	345	218	63.2
Zimbabwe	[32]	1	84	54	64.3
Cameroon	[33]	1	57	37	64.9
Singapore	[34]	1	52	35	67.3
South Africa	[35, 36]	2	496	380	76.6
India	[37–40]	4	240	184	76.7
Colombia	[41]	1	485	414	85.4
Croatia	[42]	1	542	488	90.0
Greece	[43]	1	147	137	93.2
<b>Total/Average</b>		<b>23</b>	<b>3865</b>	<b>2723</b>	<b>70.5</b>

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(northern Natal), killed BSNs were often not brought to hospitals because “most snakes are incinerated immediately after having been killed for purposes of protection from the bones of the snake, which are believed to be dangerous even after death” [45].

In four publications, victims were bitten while attempting to kill a snake. This was the case for 53% and 100% of patients bitten on the fingers in two Australian studies [29, 30] and caused the death of an 80-year-old traditional healer in Cameroon [53] and a worker in Hong Kong [17].

### BSN identification methods

From the methods section of selected publications, we extracted information on the way the BSNs were identified when victims reached a healthcare facility (S2 Table). One or a combination of the following methods were used:

i) examining the captured BSN brought to the healthcare facility (110 publications). The person who identified the BSN was not always reported. In 43 publications, and particularly in Brazil (n = 10), Australia (n = 7), Sri Lanka (n = 7), and India (n = 6), the identification was made by a snake expert. This occurred particularly for case report/case series to confirm unusual snakebite events [85, 87, 94] and for randomized controlled trials of antivenoms or epidemiological studies focusing on a particular snake species. The latter involves patients bitten by a specific snake species and therefore giving precise taxonomic attribution to the biting snake is key.

ii) verbal description of the BSN by victims/bystanders (52 publications). They described the colour (e.g. brown or black) or size (e.g. large) of the BSN. In some studies, photographs or preserved specimens in curated collections of local material were shown to victims or bystanders to assist in the identification. Several publications mentioned that a majority of patients were unable to recognize snake species based on photographs (e.g. [35, 45, 66, 110]). Victims claimed to have recognized the BSN in 16 publications.

iii) clinical features (26 publications). Distinctive clinical syndromes associated with bites by individual species have been defined and algorithms developed to infer the BSN species in India and Sri Lanka [44, 76, 111]. In Brazil, health care providers identified the BSN at the genus level (*Bothrops/Crotalus/Lachesis/Micrurus*) based on patients' signs and symptoms [102, 112, 113]. In these studies, most patients were effectively treated, but two deaths occurred

**Table 3. Geography and number of snakebite cases where the BSN was captured/killed by victims/bystanders.**

Geographical region	Country	References	Number of publications <sup>a</sup>	Total snakebite cases	Number BSNs captured	BSN captured %
Africa	Morocco	[46, 47]	2	905	14	1.5
	Tanzania	[48]	1	85	4	4.7
	Zimbabwe	[32]	1	250	22	8.8
	South Africa	[35, 45, 49, 50]	4	749	98	13.1
	Nigeria	[51, 52]	2	103	44	42.7
	Cameroon	[33, 53, 54]	3	118	60	50.8
Asia	Hong Kong	[17]	1	242	2	0.8
	Bangladesh	[55]	1	484	22	4.5
	Nepal	[20, 22, 25, 56]	4	898	74	8.2
	Taiwan	[57, 58]	2	71	8	11.3
	Sri Lanka	[23, 26, 28, 44, 59–67]	13	8 677	1 642	18.9
	Laos	[68, 69]	2	179	38	21.2
	Pakistan	[70]	1	90	21	23.3
	India	[37–40, 71–76]	10	775	189	24.4
	Saudi Arabia	[19, 77]	2	98	26	26.5
	Thailand	[78–82]	5	378	102	27.0
	Myanmar	[83, 84]	2	659	229	34.7
	Australo-Papua	Papua New Guinea	[21]	1	335	10
Australia		[29, 85–90]	7	667	181	27.1
Central and South America	Peru	[91]	1	170	10	5.9
	Ecuador	[92]	1	221	29	13.1
	Brazil	[93–106]	14	1 133	214	18.9
	Colombia	[41]	1	485	232	47.8
Europe	Croatia	[42]	1	542	49	9.0
	Greece	[43]	1	147	37	25.2
North America	USA	[107, 108]	2	176	27	15.3
	Puerto Rico	[109]	1	6	5	83.3
	<b>Total/Average</b>		<b>86</b>	<b>18 643</b>	<b>3 389</b>	<b>18.2</b>

<sup>a</sup> This includes only publications where number of BSN captured/killed was available and excludes studies whose inclusion criteria required BSN to be brought to the hospital (13 publications) and countries with only one study reporting only a single snakebite case (i.e. case report) (2 publications).

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possibly caused by the use of non-specific antivenom according to the authors [113]. The practice of bringing the BSN to the health facility is uncommon in northeastern Brazil compared to other Brazilian regions [104, 113]. Two studies focusing on krait bites also took into account bite circumstances (i.e. bite at night while victims were asleep).

iv) laboratory methods (23 publications). Immunoassays (EIA, ELISA) that detect venom antigens of some snake species in victims' blood were used in Australia, Bangladesh, Brazil, Ecuador, Myanmar, Sri Lanka, and Thailand mainly to retrospectively identify BSNs and for research purposes. A pilot study explored the use of molecular tools to identify snakes in Nepal [56].

v) examining a picture of the BSN (9 publications). This practice was reported in the US (n = 3), Morocco (n = 2), Malaysia (n = 1), Australia (n = 1), Colombia (n = 1) and Laos (n = 1). The picture of the BSN was taken by the victim/bystanders, some with a mobile phone, or by the medical staff when the dead snake was brought to the hospital. The picture was then sent to a herpetologist (Laos, USA) or a Poison Control Center (Colombia, Malaysia, Morocco, USA).



Table 4. BSN identifications done by victims.

Country	References	Total BSNs seen	Victims who claimed they can identify the BSN Number (%)	Snake names reported by victims
Ecuador	[24]	142	43 (30)	Moash/Macanche, Equis, Sobrecama, Lora, Coral <sup>a</sup>
Nepal	[22]	40	10 (25)	Cobra, krait, other venomous species not identified
Nepal	[25]	27	14 (52)	Cobra, krait, pit viper
Nepal	[115]	143 <sup>b</sup>	87 (61)	Cobra, water snake, common krait
Nigeria	[116]	142	110 (77.5)	Cobra, carpet viper, black mamba, python
Nigeria	[52]	72	54 (75)	Carpet viper
South Africa	[45]	162	62 (38)	NR (53/62 identifications presumably correct)
South Africa	[36]	202	14 (6.3)	NR
Sri Lanka	[28]	206	158 (76.8) <sup>c</sup>	NR (104/158 accurate identifications)

<sup>a</sup> Moash/Macanche (*Bothrops atrox*), Equis (*B. brazili*), Sobrecama (*B. asper*), Lora (*Bothrops bilineatus*), Coral (*Micrurus* species)

<sup>b</sup> Includes all snakebite cases, not only cases where the snake was spotted

<sup>c</sup> Identification based on dead snakes brought to health facility

NR: Not Reported

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BSNs were identified at the species or genus level in 18,065 out of 33,827 snakebite cases (53%) (S2 Table).

### Capability of victims and healthcare providers to identify snakes

**Victims/bystanders.** In nine studies, victims claimed they could identify the BSN and reported its common or local name (Table 4). In a Bangladeshi hospital, most snakebite victims were too distressed to describe the BSN and often misidentified it even when it was brought to the health facility [55]. Victims may not know the name of the snake [24] or the diversity of vernacular names given to a single snake species and/or taxonomic synonymies generate confusion for doctors (e.g. in the Brazilian Amazon [103, 112, 114]). In South Africa the word 'mamba' is often used by the local population to mean 'snake' of any species [50]. A survey of 150 inhabitants in southern Nepal showed that respondents were generally unable to identify different snake species [20]. In many parts of the world, authors considered snake identification by victims too unreliable to assist in routine snakebite management or in clinical snakebite research [24, 28, 66, 110].

**Healthcare providers.** The identity and credentials of the person doing the identification of BSN brought to health facility was often not reported. This limits our assessment of healthcare provider capability in identifying snakes. In three studies in India, Sri Lanka, and Thailand, experts systematically re-examined dead snakes initially identified by hospital personnel and misidentifications were reported in respectively 17/44, 51/860, and 27/1631 cases [44, 117, 118]. In the US, pictures of the BSN were sent to a Poison Control Center and identification by a snake expert was compared to that of healthcare providers. Healthcare accuracy for copperhead (*Agkistrodon contortrix*) and cottonmouth (*A. piscivorus*) identifications was respectively 68% and 74% [119]. Many authors highlighted the lack of training of healthcare providers in identifying biting species including in Brazil, India, Nepal, Singapore, South Africa, and Thailand [34, 45, 56, 97, 113, 117, 118, 120, 121].

### BSN misidentifications

The BSNs were misidentified in 106 snakebite cases in Australia (n = 3), Brazil (n = 10), Hong Kong (n = 1), India (n = 6), Malaysia (n = 1), Sri Lanka (n = 54), and Thailand (n = 31). The consequences for the victims are detailed in Table 5 according to three scenarios. Most



**Table 5. Cases of BSN misidentification published in the literature.**

Scenario 1: The BSN is venomous but misidentified as another venomous snake							
Country	References	Number of misidentified BSNs	Initial identification (incorrect)	Person who did the incorrect identification <sup>a</sup>	Final identification (correct)	Person who did the correct identification <sup>a</sup>	Consequences for the victim(s)
Australia	[30]	1	<i>Pseudonaja nuchalis</i>	Park ranger	<i>Notechis ater</i>	SVDK at hospital	Treated with Brown snake ( <i>Pseudonaja sp</i> ) antivenom and then Tiger snake ( <i>Notechis sp</i> ) antivenom. The 13-year old girl required prolonged ventilation and 53-day stay at hospital
Australia	[86]	1	<i>Pseudechis australis</i>	Other patients at hospital	<i>Pseudonaja nuchalis</i>	Flying Doctor at hospital	Treated with Black snake ( <i>Pseudechis sp</i> ) antivenom and then Brown snake ( <i>Pseudonaja sp</i> ) antivenom 7 hours after the bite. The patient died due to cerebral trauma (the patient had collapsed to the ground) aggravated by coagulation defect).
Brazil	[122]	1	Viperidae	NR	<i>Crotalus durissus terrificus</i>	Physician expert in toxicology	Treated with antithrotophic serum and then intravenous crotalid antivenom. The patient remained with a severe behavioral and cognitive impairment.
Hong Kong	[17]	1	<i>Ophiophagus hannah</i>	Physician at hospital & Hong Kong Government Agriculture and Fisheries Department	<i>Naja naja</i>	NR	Below elbow amputation and death. Antivenom not administered due to uncertainty in snake species.
India	[40]	1	<i>Echis sochureki</i>	A colleague	<i>Macrovipera lebetina</i>	Experienced herpetologist	Inappropriate treatment with serum institute of India polyvalent <sup>b</sup> that does not cover <i>M. Lebetina</i> , finger amputation
India	[117]	5	<i>Echis carinatus</i>	Hospital staff	<i>Hypnale hypnale</i>	Snake experts	Inappropriate treatment with Indian polyvalent antivenom that does not cover <i>H. Hypnale</i> (2/4 developed anaphylactoid antivenom reaction)
Sri Lanka	[59]	1	<i>Hypnale hypnale</i>	Native physician	<i>Daboia russelii</i>	NR, at hospital	Delayed treatment with indian polyvalent antivenom but patient died probably due to herniation of brain stem.
Sri Lanka	[44]	36	<i>Daboia russelii</i>	Hospital staff	<i>Hypnale hypnale</i>	Snake experts	Unnecessary use of antivenom
Thailand	[78]	8	<i>Bungarus fasciatus</i>	Medical staff	<i>Bungarus candidus</i>	Snake experts	Inappropriate treatment with <i>Bungarus fasciatus</i> antivenom. The patients died.
Thailand	[79]	1	<i>Naja kaouthia</i>	Assumption by the victim (in the dark)	<i>Bungarus fasciatus</i>	ELISA confirmation	Inappropriate treatment with monospecific cobra antivenom. The patient died.
Thailand	[118]	5	<i>Bungarus fasciatus</i>	Hospital staff	<i>Bungarus candidus</i>	Snake experts	Inappropriate treatment with <i>Bungarus fasciatus</i> antivenom
Scenario 2: The BSN is non or mildly venomous (opisthogyphous) but identified as a venomous snake							
Country	References	Number of misidentified BSNs	Initial identification (incorrect)	Person who did the incorrect identification <sup>a</sup>	Final identification (correct)	Person who did the correct identification <sup>a</sup>	Consequences for the victim(s)
Australia	[85]	1	Black snake	the family & SVDK	<i>Denisonia maculata</i>	Snake Experts	Inappropriate treatment with Australian Black Snake ( <i>Pseudechis sp</i> ) antivenom
Brazil	[123]	4	<i>Bothrops sp.</i>	Medical staff	<i>Tomodon dorsatus</i>	Laboratório de Herpetologia and the Laboratório Especial de Coleções Zoológicas of Butantan Institute.	Inappropriate treatment with Bothrops antivenom
Brazil	[97]	1	<i>Bothrops sp.</i>	Medical staff	<i>Philodryas patagonensis</i>	NR	Inappropriate treatment with Bothrops antivenom
Brazil	[94]	1	<i>Bothrops sp.</i>	Medical staff / clinical signs	<i>Clelia clelia plumbea</i>	Centra de Estudos e Pesquisas Biológicas at Universidade Católica de Goias	Inappropriate treatment with Bothrops antivenom
Brazil	[124]	1	<i>Bothrops sp.</i>	NR	<i>Drymarcon corais</i>	NR	Inappropriate treatment with Bothrops antivenom, nearly died of severe anaphylaxis
Brazil	[124]	1	<i>Bothrops sp.</i>	NR	<i>Sibynomorphus mikáinii</i>	NR	Inappropriate treatment with Bothrops antivenom, no side effect

(Continued)

Table 5. (Continued)

Country	References	Number of misidentified BSNs	Initial identification (incorrect)	Person who did the incorrect identification <sup>a</sup>	Final identification (correct)	Person who did the correct identification <sup>a</sup>	Consequences for the victim(s)
Malaysia	[125]	1	<i>Bungarus Sp.</i>	National Poison Centre / description of a photo of the snake	<i>Chrysopelea pelias</i>	Snake Experts	Inappropriate treatment with Indian polyvalent antivenom
Sri Lanka	[64]	1	<i>Echis carinatus</i>	Hospital staff	<i>Boiga trigonata</i>	Snake expert / Faculty of Medicine, Peradeniya	NR
Sri Lanka	[44]	2	<i>Hypnale hypnale</i>	Hospital staff	<i>Boiga ceylonensis/B. trigonata</i>	Snake experts	Unnecessary use of antivenom
Sri Lanka	[44]	3	<i>Daboia russelii</i>	Hospital staff	<i>Boiga ceylonensis/B. trigonata</i>	Snake experts	Unnecessary use of antivenom
Sri Lanka	[44]	1	<i>Echis carinatus</i>	Hospital staff	<i>Boiga ceylonensis/B. trigonata</i>	Snake experts	Unnecessary use of antivenom
Sri Lanka	[44]	2	<i>Echis carinatus</i>	Hospital staff	Pythons	Snake experts	Unnecessary use of antivenom
Sri Lanka	[44]	5	<i>Bungarus caeruleus</i>	Hospital staff	<i>Lycodon aulicus/L. striatus sithalayu</i>	Snake experts	Unnecessary use of antivenom
Sri Lanka	[44]	2	Cobras	Hospital staff	Rat snakes	Snake experts	Unnecessary use of antivenom
Thailand	[118]	2	<i>Calloselasma rhodostoma</i>	Hospital staff	<i>Oligodon dorsolateralis</i>	Snake experts	Unnecessary use of antivenom
Thailand	[118]	1	<i>Daboia russelii</i>	Hospital staff	<i>Oligodon cyclurus</i>	Snake experts	Unnecessary use of antivenom
Thailand	[118]	1	<i>Calloselasma rhodostoma</i>	Hospital staff	<i>Oligodon cyclurus</i>	Snake experts	Unnecessary use of antivenom
Thailand	[118]	7	<i>Daboia russelii</i>	Hospital staff	<i>Boiga multomaculata</i>	Snake experts	Unnecessary use of antivenom
Thailand	[118]	1	<i>Bungarus candidus</i>	Hospital staff	<i>Lycodon laoensis</i>	Snake experts	Unnecessary use of antivenom
Thailand	[118]	1	<i>Calloselasma rhodostoma</i>	Hospital staff	<i>Rhabdophis subminiatus</i>	Snake experts	Unnecessary use of antivenom
Thailand	[118]	4	<i>B. fasciatus/B. candidus</i>	Hospital staff	<i>Dryocalamus davisonii</i>	Snake experts	Unnecessary use of antivenom
<b>Scenario 3: The BSN is venomous but identified as non-venomous</b>							
Brazil	[98]	1	Nonpoisonous	Healthcare provider	<i>Bothrops jararacussu</i>	NR	Delayed treatment with anti-Bothrops serum, intracranial bleeding and gradual improvement
Sri Lanka	[60]	1	<i>Lycodon aulicus</i>	Doctor	<i>Bungarus caeruleus</i>	Identified by expert after patient death	Delayed administration of Indian polyvalent anti-venom, death 46 hours after the bite

<sup>a</sup> Identification done by examining the snake unless otherwise specified

<sup>b</sup> Indian polyvalent antivenom is raised against the venom of *B. caeruleus*, *Daboia russelii*, *Echis carinatus*, and *N. naja* (the so-called Big Four).

NR: Not Reported

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frequently, a venomous species was confused with another venomous species or a non or mildly venomous (opisthoglyphous) species was misidentified as a venomous snake.

### Diversity of captured/killed BSN

A total of 8,885 BSNs from 149 species in 12 families were captured and identified. Of these, 6,750 BSNs were identified to species, 2,082 to genus, and 53 as “non-venomous”. In total, 7,628 individual snakes of 69 species were MIVS, 1,205 individuals of 71 species were non-venomous, and 52 individuals of 9 species were potentially dangerous but understudied species (species that lack specific data on clinical symptoms or venom toxicity but which are closely related to dangerous snakes and possess fangs and venom glands (see [126]) (Table 6 and S3 Table). The 149 species included 40 species in the family Viperidae, 35 species in the family Elapidae, 49 species in the family Colubridae, 4 species in the family Lamprophiidae, and 21 species from 8 other families (Acrochordidae, Boidae, Cyliophoridae, Homalopsidae, Pareidae, Pythonidae, Typhlopidae, Xenopeltidae; S3 Table), as well as one amphisbaenid lizard (*Amphisbaena mertensii*) [101] and one serpentine amphibian (caecilian [118]) (not included in snake totals above). Of these 149 snake species, 32 were diurnal, 76 were nocturnal, and 41 were or could be active throughout the day, depending on the season. A total of 84 species were active foragers, 45 were ambush predators, 14 used both strategies, and 6 were unknown. General habitat use of 26 species was arboreal, 7 fossorial, 22 aquatic or semi-aquatic, and 94 were primarily terrestrial, with 4 of these partially aquatic, 5 partially arboreal, and 7 partially fossorial (S4 Table).

Within a country, the total number of snake bites across all publications was positively correlated with the number of species of BSNs reported (Panel A in S1 Fig; PPC = 0.55,  $p < 0.001$ ), especially among non-venomous snakes, whereas the number of snake species per country was more weakly correlated with the number of species of BSNs reported (Panel B in S1 Fig; PPC = 0.36,  $p = 0.04$ ).

A substantial number of snakes—1,859 (21%) individuals of 22 species—were reported in the literature using only the common name. Of those reported using the scientific name, 20 species (92 individuals) had been moved to another genus since the time of publication, 10 species (241 individuals) had experienced other taxonomic changes (splits or lumps) (e.g. *Echis ocellatus* in Cameroon has been renamed *E. romani* [127]), and the names of 3 species (6 individuals) contained minor misspellings as reported (S3 Table).

### Discussion

Based on a final selection of 150 publications from across the world, this first scoping review on BSN identification practices of communities and healthcare providers confronted with snakebite shows that: (I) BSN identification is important for snakebite epidemiology and clinical management yet there is a diversity of practices depending on cultural, ecological and healthcare contexts and there are no official standards (II) the majority of victims see the BSN but they are unreliable in their identification (III) the practice of capturing or killing the BSN occurs in all continents, especially in Asian countries with a diversity of snakes, and (IV) healthcare providers struggle to identify a BSN presented in the health facility and misidentifications occur.

Snakebite victims/bystanders in many snakebite endemic countries are aware of the importance of BSN identification and can play an important role. We found that on average 70% of victims/bystanders spotted the BSN and 38% managed to capture or kill it and brought it to the health facility for identification. Although this is a dangerous practice (i.e. due to the risk of secondary bites [17, 29, 30, 53, 128–133]), it occurs worldwide but particularly in Asia (e.g.

Table 6. Diversity and abundance of BSNs captured/killed and identified.

Geographical region	MIVS (n)	Non-venomous snakes (n)	Potentially dangerous snakes (n)
Africa	<i>Atractaspis bibronii</i> (22)	<i>Afrotyphlops schlegelii</i> (1)	<i>Causus defilippi</i> (8)
	<i>Bitis arietans</i> (8)	<i>Boaedon capensis</i> (2)	<i>Causus rhombeatus</i> (9)
	<i>Bitis atropos</i> (1)	<i>Crotaphopeltis hotamboeia</i> (3)	
	<i>Daboia mauritanica</i> (3)	<i>Dasyeltis scabra</i> (1)	
	<i>Dendroaspis polylepis</i> (12)	<i>Philothamnus</i> spp. (2)	
	<i>Echis romani</i> (103)		
	<i>Hemachatus haemachatus</i> (1)		
	<i>Naja annulifera</i> (3)		
	<i>Naja melanoleuca</i> (1)		
<i>Naja mossambica</i> (18)			
Asia	<i>Bitis arietans</i> (1)	<i>Acrochordus javanicus</i> (NR)	
	<i>Bungarus caeruleus</i> (623)	<i>Ahaetulla nasuta</i> (10)	
	<i>Bungarus candidus</i> (17)	<i>Ahaetulla prasina</i> (NR)	
	<i>Bungarus fasciatus</i> (6)	<i>Argyrogena fasciolata</i> (1)	
	<i>Bungarus lividus</i> (3)	<i>Atreitum schistosum</i> (1)	
	<i>Bungarus multicinctus</i> (6)	<i>Boiga ceylonensis</i> (10)	
	<i>Bungarus niger</i> (1)	<i>Boiga cyanea</i> (NR)	
	<i>Bungarus</i> spp. (20)	<i>Boiga cynodon</i> (NR)	
	<i>Calloselasma rhodostoma</i> (448)	<i>Boiga forstenii</i> (8)	
	<i>Cerastes cerastes</i> (7)	<i>Boiga multomaculata</i> (NR)	
	<i>Cerastes gasparetti</i> (7)	<i>Boiga</i> sp. (4)	
	<i>Cerastes vipera</i> (1)	<i>Boiga trigonata</i> (4)	
	<i>Daboia russelii</i> (985)	<i>Chrysopelea ornata</i> (1)	
	<i>Daboia siamensis</i> (383)	<i>Chrysopelea pelias</i> (1)	
	<i>Echis carinatus</i> (229)	<i>Coelognathus helena</i> (5)	
	<i>Echis coloratus</i> (5)	<i>Coelognathus radiatus</i> (4)	
	<i>Hydrophis cyanocinctus</i> (2)	<i>Cylindrophis maculatus</i> (1)	
	<i>Hydrophis platurus</i> (2)	<i>Cylindrophis ruffus</i> (NR)	
	<i>Hydrophis schistosus</i> (2)	<i>Enhydris enhydris</i> (NR)	
	<i>Hydrophis spiralis</i> (1)	<i>Enhydris jagorii</i> (NR)	
	<i>Hypnale hypnale</i> (502)	<i>Gonyosoma oxycephalum</i> (NR)	
	<i>Hypnale nepa</i> (5)	<i>Homolopsis buccata</i> (NR)	
	<i>Hypnale</i> spp. (1930)	<i>Hypsiglossus plumbea</i> (NR)	
	<i>Hypnale zara</i> (17)	<i>Indotyphlops braminus</i> (NR)	
	<i>Macrovipera lebetina</i> (1)	<i>Lycodon aulicus</i> (57)	
	<i>Naja atra</i> (1)	<i>Lycodon capucinus</i> (NR)	
	<i>Naja kaouthia</i> (108)	<i>Lycodon davisonii</i> (NR)	
	<i>Naja naja</i> (328)	<i>Lycodon laoensis</i> (NR)	
	<i>Naja siamensis</i> (118)	<i>Lycodon</i> spp. (3)	
	<i>Naja</i> spp. (8)	<i>Lycodon striatus</i> (15)	
	<i>Naja sumatrana</i> (1)	<i>Malpolon moileensis</i> (4)	
	<i>Ovophis monticola</i> (1)	« non-venomous snakes » (53)	
	<i>Rhabdophis subminiatus</i> (5)	<i>Oligodon arnensis</i> (7)	
	<i>Trimeresurus albolabris</i> (344)	<i>Oligodon cyclurus</i> (NR)	
	<i>Trimeresurus gramineus</i> (20)	<i>Oligodon</i> spp (7)	
	<i>Trimeresurus kanburiensis</i> (1)	<i>Oligodon taeniatus</i> (NR)	
	<i>Trimeresurus macrops</i> (23)	<i>Oligodon taeniolatus</i> (2)	
	<i>Trimeresurus malabaricus</i> (26)	<i>Pareas carinatus</i> (NR)	
	<i>Trimeresurus purpureomaculatus</i> (3)	<i>Psammodynastes pulverulentus</i> (2)	
	<i>Trimeresurus</i> spp. (36)	<i>Ptyas mucosa</i> (13)	
	<i>Trimeresurus trigonocephalus</i> (19)	<i>Ptyas</i> spp. (2)	
<i>Tropidolaemus wagleri</i> (2)	<i>Python molurus</i> (2)		
<i>Walterinnesia aegyptia</i> (1)	<i>Subsessor bocourti</i> (NR)		
	<i>Xenochrophis asperrimus</i> (2)		
	<i>Xenochrophis flavipunctatus</i> (1)		
	<i>Xenochrophis flavipunctatus</i> (NR)		
	<i>Xenochrophis piscator</i> (12)		
	<i>Xenochrophis</i> spp. (2)		
	<i>Xenopeltis unicolor</i> (NR)		

(Continued)

Table 6. (Continued)

Geographical region	MIVS (n)	Non-venomous snakes (n)	Potentially dangerous snakes (n)
Australo-papua	<i>Acanthophis sp.</i> (13)	<i>Antaresia childreni</i> (12)	<i>Boiga irregularis</i> (3)
	<i>Notechis scutatus</i> (NR)	<i>Dendrelaphis punctulatus</i> (2)	<i>Cryptophis pallidiceps</i> (4)
	<i>Oxyuranus microlepidotus</i> (1)	<i>Fordonia leucobalia</i> (1)	<i>Demansia olivacea</i> (5)
	<i>Oxyuranus scutellatus</i> (1)	<i>Liasis fuscus</i> (9)	<i>Demansia spp.</i> (17)
	<i>Pseudechis australis</i> (30)	<i>Liasis olivaceus</i> (4)	<i>Denisonia maculata</i> (1)
	<i>Pseudechis guttatus</i> (1)	<i>Morelia spilota</i> (8)	<i>Furina ornata</i> (3)
	<i>Pseudonaja nuchalis</i> (13)	<i>Pseudoferania polylepis</i> (1)	<i>Hemiaspis signata</i> (2)
	<i>Pseudonaja spp.</i> (20)	<i>Stegonotus australis</i> (10)	
		<i>Tropidonophis mairii</i> (2)	
Central and South America	<i>Bothrops alternatus</i> (6)	<i>Apostolepis assimilis</i> (1)	
	<i>Bothrops atrox</i> (45)	<i>Boa constrictor</i> (2)	
	<i>Bothrops bilineatus</i> (12)	<i>Clelia plumbea</i> (1)	
	<i>Bothrops erythromelas</i> (35)	<i>Dipsas mikanii</i> (6)	
	<i>Bothrops jararaca</i> (779)	<i>Drymarchon corais</i> (1)	
	<i>Bothrops jararacussu</i> (1)	<i>Erythrolamprus poecilogyrus</i> (2)	
	<i>Bothrops moojeni</i> (70)	<i>Erythrolamprus spp.</i> (1)	
	<i>Bothrops neuwiedi</i> (30)	<i>Eunectes murinus</i> (1)	
	<i>Bothrops sp.</i> (15)	<i>Oxyrhopus trigeminus</i> (3)	
	<i>Bothrops taeniatus</i> (2)	<i>Palusophis bifossatus</i> (3)	
	<i>Crotalus durissus</i> (14)	<i>Philodryas mattogrossensis</i> (1)	
	<i>Micrurus frontalis</i> (2)	<i>Philodryas olfersii</i> (4)	
	<i>Micrurus lemniscatus</i> (2)	<i>Philodryas patagoniensis</i> (300)	
	<i>Micrurus spp.</i> (2)	<i>Simophis rhinostoma</i> (2)	
		<i>Tomodon dorsatus</i> (86)	
	<i>Xenodon merremii</i> (3)		
Europe	<i>Vipera ammodytes</i> (83)		
	<i>Vipera berus</i> (3)		
North America	<i>Agkistrodon contortrix</i> (26)	<i>Borikenophis portoricensis</i> (5)	
	<i>Crotalus cerastes</i> (1)		

In this table, snake scientific names follow the taxonomy of the Reptile Database as of October, 2019 [10]

NR: Not reported

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Myanmar, India, and Sri Lanka). Communities in these snakebite endemic countries believe that bringing the BSN to the health facility for identification is essential for treating the victim [11, 134]. Myths and beliefs regarding snakes are present in many societies (e.g. [135, 136]) and could prevent victims/bystanders from killing the snake, yet the influence of these cultural aspects on BSN identification was rarely addressed in the selected publications and would deserve more research. Traditional healers may have a good knowledge on the type of snakes in their local environment, their distribution and behaviours. Yet, in this review, we did not find articles that report their role in BSN identification. Further studies could explore if their snakebite remedies are species-specific and assess their knowledge of local snake diversity. Overall, traditional healers can act as partners with healthcare providers promoting prompt referral to health facilities in case of snake bites inflicted by venomous snakes.

Photographing the BSN (e.g. with mobile phones) is an emerging practice in snakebite endemic countries worldwide. This was described for Australia, Colombia, Laos, Malaysia, Morocco, and the US, where snake experts are involved in snake photo identification (e.g. at Poison Control Center). This procedure is recommended by the WHO [12], the Snakebite Healing and Education Society in India [137] and the African Snakebite Institute [138] among others and it is certainly less dangerous than killing or capturing the snake. Even fast-moving, active-foraging snakes (e.g. *Demansia*, *Dendroaspis*, *Naja*) and secretive, fossorial species (e.g.

*Atractaspis*, *Micrurus*, *Xenopeltis*) were captured/killed in our review and could have been photographed instead. The advantages of using photos for BSN identification are numerous. Firstly, photos are less subject to interpretations and thus more reliable than victim descriptions. We showed that victims in general described the BSN colour or size, which is of limited value for the identification, and their identification of the BSN is often unreliable even with the assistance of preserved specimens or local snake photographs. Secondly, photos can be rapidly shared between victims, healthcare providers, and snake biologists, accelerating the identification process and improving its accuracy. Snake identification services based on snake photos have been developed in India [139], Sri Lanka [140], or Thailand [141] and are provided by poison control centres in the US [142] and Colombia [143]. This could be developed in other countries and, in low-resource settings, snake identification could be even crowdsourced involving online communities of snake experts (e.g. open biodiversity platforms like HerpMapper, iNaturalist, or snake identification Facebook groups) [144, 145]. Thirdly, photos can be securely and indefinitely stored building a digital dataset. Besides the ecological and epidemiological value of such a dataset, it could serve to train healthcare providers and machine learning algorithms in snake classification [146, 147].

BSN identification is particularly important in certain SBE endemic regions where monovalent or bi-/tri valent antivenoms are the only affordable treatment. This is the case, for example, in Myanmar, Taiwan, and Thailand in East and South East Asia, but also in many sub-Saharan Africa regions where *Echis* snake bites are prevalent [12]. Besides this, BSN identification is important for ecological and epidemiological purposes and subsequently for an optimal antivenom coverage [146] (e.g. Hump-nosed viper (*Hypnale hypnale*) causes frequent bites with morbidity and mortality in Sri Lanka and southwest India but currently lacks effective antivenom) [117, 148]. Healthcare providers should be encouraged to photograph and archive images of BSN brought to health facilities to help build national or regional BSN photo repositories for further epidemiological studies.

BSN identification could also complement syndromic approach to snakebite [36, 44, 76], particularly in those cases where symptoms caused by venoms of different snake species overlap (e.g. Russell's viper causes paralytic signs suggesting elapid neurotoxicity in Sri Lanka) [12].

Snake species diversity and the fact that bites from some “non-venomous” snakes can cause signs of envenoming (e.g. *Clelia plumbea* and *Philodryas olfersii* in Brazil) [64, 94, 124, 126] complicate snakebite clinical management and have led to snake misidentifications. Snake identification can be challenging for communities and healthcare providers (i.e. diversity of snakes, mimicry). In two villages in rural Tanzania, most of the respondents to a survey could not precisely differentiate between venomous and non-venomous snakes [149]. In a cross-sectional survey of 119 healthcare providers in Laos, 86 participants (72.3%) had inadequate knowledge of snake identification [150]. Although healthcare providers could be trained to recognize locally prevalent snake species (e.g. at the Damak Snakebite Treatment Center in Nepal [151]), reinforcing the collaboration between communities at risk, healthcare providers and snake experts could be a more effective approach to increase the number of BSNs correctly identified.

With this review, we retrospectively collated all the species names of the 8,885 BSN that were captured and identified and we have built a first extensive global list of snakes having bitten humans. This list includes 69 MIVS and 71 non-venomous snake species, as well as nine that are potentially dangerous but understudied species [126]. MIVS species are already listed by the WHO (2010) [152]. This review extends this list to non-venomous snakes, although there is significant bias in which non-venomous snakes are brought to health facilities and the number of non-venomous BSN species was strongly correlated with the number of snake

bites, suggesting that many more species of non-venomous BSNs exist and a limit has not been reached within epidemiological data. Nevertheless, this confirms that many snake bites globally are caused by non-venomous snakes, although victims may not always realize this immediately or at all.

### Limitations of this study

Many publications had to be excluded from the review because the way the BSN identification was done was unclear or not reported even though the BSN genus or species was mentioned. The list of BSN species we built is limited to the publications we retrieved and included in the review. It could have been extended, had more publications met the eligibility criteria. We recommend that future epidemiological studies on snakebite clearly describe the method(s) used to identify the BSN, including the credentials of the person who did the identification and their confidence in the identification. We used key words related to 'identification' to specifically gather snakebite publications describing snake identification, although we may have missed relevant publications that do not mention these keywords or that are published in other languages (e.g. Russian, Chinese). Information on snake identification was reported in an inconsistent and fragmented manner. We managed this problem by involving two authors in data extraction and comparing data collected until a consensus was reached. Some publications, particularly prospective studies, applied specific inclusion criteria (e.g. a specific snake species, dead snake brought to hospital). These were excluded from analyses where they were sources of bias (e.g. calculation of proportion of captured snakes). The selected publications are mainly hospital-based studies with very few community-based studies. We missed the behaviour of snakebite victims who did not seek treatment at hospitals because of asymptomatic bite or use of traditional healers. Snake taxonomy is constantly changing and an average of 30 new species per year have been described since the year 2000. Although we were always able to definitively decide which species/genera were meant, rare or newly-described species may be missed by all identification methodologies, and taxonomic instability further complicates an already-challenging situation [153, 154]. Finally, we cannot account for situations where snake misidentification was never discovered and incorrect names have been published, which seems likely in a subset of cases.

### Conclusion

This global scoping review showed that BSN identification in snakebite endemic countries includes a diversity of methods and practices: capturing/killing the BSN and examination of its body, description of the BSN by victim/bystanders, interpretation of clinical features, laboratory tests, and photographing the BSN. The capacity of snakebite victims, bystanders and healthcare providers to spot and identify the BSN is context-specific and depends on circumstances of the bite, the local snake diversity, and their own knowledge of local snakes. BSN misidentifications occur and lead to inappropriate management of the victims. The influence of cultural perceptions about snakes and role of traditional healers in snake identification are largely unexplored in the literature and urges for further research. Victims/bystanders managed to capture a diversity of BSNs, including fast-moving nervous snakes. This is dangerous and not recommended, and photographing the snake could be an alternative option [12]. We provided the first evidence-based list of venomous and non-venomous snake species involved in bites to humans. This list could be further extended by implementing snake identification as part of the clinical practice. Such a systematic collection of the taxonomy of BSNs at the global level is of considerable interest to better understand snake ecology and snakebite epidemiology and ultimately improve SBE management.



## Supporting information

**S1 Table. Search strategy syntax for each bibliographic database.**

(DOCX)

**S2 Table. Dataset—Scoping review identification.**

(XLSX)

**S3 Table. Taxonomic identification of captured biting snakes.**

(XLSX)

**S4 Table. Behaviour and ecology of captured biting snakes.**

(XLSX)

**S1 Fig. Correlation between number of snake bites and occurring snake species with number of biting snake species.** Correlations between A) the total number of snake bites across all publications and B) the total number of snake species occurring in a country with the number of species of BSNs reported. Each dot represents a country. Thailand is missing from the non-venomous panel in part A because quantitative data are not given for non-venomous BSNs in [31]. MIVS = medically-important venomous snakes.

(TIFF)

**S1 File. PRISMA-ScR checklist.**

(PDF)

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## References

1. Gutiérrez JM, Calvete JJ, Habib AG, Harrison RA, Williams DJ, Warrell DA. Snakebite envenoming. *Nat Rev Dis Primers*. 2017; 3:17063. <https://doi.org/10.1038/nrdp.2017.63> PMID: 28905944
2. WHO. Snakebite envenoming—Key Facts 2019 [cited 5 November 2019]. Available from: <https://www.who.int/news-room/fact-sheets/detail/snakebite-envenoming>
3. Williams DJ, Faiz MA, Abela-Ridder B, Ainsworth S, Bulfone TC, Nickerson AD, et al. Strategy for a globally coordinated response to a priority neglected tropical disease: Snakebite envenoming. *PLoS Negl Trop Dis*. 2019; 13(2):e0007059. <https://doi.org/10.1371/journal.pntd.0007059> PMID: 30789906
4. WHO. Snakebite envenoming—A strategy for prevention and control. 2019. Available from: <https://www.who.int/snakebites/resources/9789241515641/en/>
5. Isbister G. Snake antivenom research: the importance of case definition. *Emerg Med J*. 2005; 22(6):399–400. <https://doi.org/10.1136/emj.2004.022251> PMID: 15911943
6. Williams DJ, Habib AG, Warrell DA. Clinical studies of the effectiveness and safety of antivenoms. *Toxicon*. 2018; 150:1–10. Epub 2018/05/11. <https://doi.org/10.1016/j.toxicon.2018.05.001> PMID: 29746978.
7. Ralph R, Sharma SK, Faiz MA, Ribeiro I, Rijal S, Chappuis F, et al. The timing is right to end snakebite deaths in South Asia. *BMJ*. 2019; 364:k5317. <https://doi.org/10.1136/bmj.k5317> PMID: 30670457
8. Alirol E, Sharma SK, Bawaskar HS, Kuch U, Chappuis F. Snake bite in South Asia: a review. *PLoS Negl Trop Dis*. 2010; 4(1):e603. Epub 2010/02/04. <https://doi.org/10.1371/journal.pntd.0000603> PMID: 20126271; PubMed Central PMCID: PMC2811174.
9. Williams HF, Layfield HJ, Vallance T, Patel K, Bicknell AB, Trim SA, et al. The Urgent Need to Develop Novel Strategies for the Diagnosis and Treatment of Snakebites. *Toxins (Basel)*. 2019; 11(6):363.
10. Uetz. The Reptile Database 2019 [cited 5 November 2019]. Available from: <http://reptile-database.reptarium.cz>
11. Schioldann E, Mahmood MA, Kyaw MM, Halliday D, Thwin KT, Chit NN, et al. Why snakebite patients in Myanmar seek traditional healers despite availability of biomedical care at hospitals? Community perspectives on reasons. *PLoS Negl Trop Dis*. 2018; 12(2):e0006299. <https://doi.org/10.1371/journal.pntd.0006299> PMID: 29489824
12. WHO. Regional Office for South-East Asia, Guidelines for the management of snakebite 2nd edition 2016. Available from: <https://www.who.int/snakebites/resources/9789290225300/en/>
13. Arksey H, O'Malley L. Scoping studies: towards a methodological framework. *Int J Soc Res Methodol*. 2005; 8(1):19–32.
14. Levac D, Colquhoun H, O'Brien KK. Scoping studies: advancing the methodology. *Implementation Sci*. 2010; 5(1):69.
15. Tricco AC, Lillie E, Zarin W, O'Brien KK, Colquhoun H, Levac D, et al. PRISMA extension for scoping reviews (PRISMA-ScR): checklist and explanation. *Ann Intern Med*. 2018; 169(7):467–73. <https://doi.org/10.7326/M18-0850> PMID: 30178033
16. Tchoua R, Raouf A, Ogandaga A, Mouloungui C, Mbanga Loussou J, Kombila NND, et al. Analyse des envenimations par morsures de serpent au Gabon. *Bull Soc Pathol Exot*. 2002; 95(3):188–90. PMID: 12404868
17. Cockram CS, Chan JC, Chow KY. Bites by the white-lipped pit viper (*Trimeresurus albolabris*) and other species in Hong Kong. A survey of 4 years' experience at the Prince of Wales Hospital. *J Trop Med Hyg*. 1990; 93(2):79–86. Epub 1990/04/01. PMID: 2325197.
18. Ariaratnam CA, Sheriff MHR, David R, Theakston G, Warrell DA. Distinctive epidemiologic and clinical features of common krait (*Bungarus caeruleus*) bites in Sri Lanka. *Am J Trop Med Hyg*. 2008; 79(3):458–62. <https://doi.org/10.4269/ajtmh.2008.79.458> WOS:000259307800027. PMID: 18784244
19. Al Durihim H, Al Hussaini M, Bin Salih S, Hassan I, Harakati M, Al Hajjaj A. Snake bite envenomation: experience at King Abdulaziz Medical City, Riyadh. 2010. *East Mediterr Health J Apr*; 16(4):438–41
20. Pandey DP, Vohra R, Stalcup P, Shrestha BR. A season of snakebite envenomation: presentation patterns, timing of care, anti-venom use, and case fatality rates from a hospital of southcentral Nepal. *J Venom Res*. 2016; 7:1–9. MEDLINE:26998219. PMID: 26998219
21. Laloo D, Trevett A, Saweri A, Naraq S, Theakston R, Warrell D. The epidemiology of snake bite in Central Province and National Capital District, Papua New Guinea. *Trans R Soc Trop Med Hyg*. 1995; 89(2):178–82. [https://doi.org/10.1016/0035-9203\(95\)90485-9](https://doi.org/10.1016/0035-9203(95)90485-9) PMID: 7778143

22. Hansdak SG, Lallar KS, Pokharel P, Shyangwa P, Karki P, Koirala S. A clinico-epidemiological study of snake bite in Nepal. *Trop Doct.* 1998; 28(4):223–6. Epub 1998/11/06. <https://doi.org/10.1177/004947559802800412> PMID: 9803844.
23. Kularatne K, Budagoda S, Maduwage K, Naser K, Kumarasiri R, Kularatne S. Parallels between Russell's viper (*Daboia russelii*) and hump-nosed viper (*Hypnale* species) bites in the central hills of Sri Lanka amidst the heavy burden of unidentified snake bites. *Asian Pac J Trop Med.* 2011; 4(7):564–7. Epub 2011/08/02. [https://doi.org/10.1016/S1995-7645\(11\)60147-8](https://doi.org/10.1016/S1995-7645(11)60147-8) PMID: 21803310.
24. Praba-Egge AD, Cone SW, Araim O, Freire I, Paida G, Escalante J, et al. Snakebites in the rainforests of Ecuador. *World J Surg.* 2003; 27(2):234–40. Epub 2003/03/05. <https://doi.org/10.1007/s00268-002-6552-9> PMID: 12616443.
25. Heap BJ, Cowan GO. The epidemiology of snake bite presenting to British Military Hospital Dharan during 1989. *J R Army Med Corps.* 1991; 137(3):123–5. Epub 1991/10/01. <https://doi.org/10.1136/jramc-137-03-03> PMID: 1744818.
26. Rathnayaka RN, Ranathunga P, Kularatne S. Epidemiology and clinical features of Green pit viper (*Trimeresurus trigonocephalus*) envenoming in Sri Lanka. *Toxicon.* 2017; 137:99–105. <https://doi.org/10.1016/j.toxicon.2017.07.017> PMID: 28735968
27. Rathnayaka RN, Kularatne S, Ranathunga P. Coagulopathy and extensive local swelling following Green pit viper (*Trimeresurus trigonocephalus*) envenoming in Sri Lanka. *Toxicon.* 2017; 129:95–9. <https://doi.org/10.1016/j.toxicon.2017.02.011> PMID: 28216410
28. Kularatne AM, Silva A, Maduwage K, Ratnayake I, Walathara C, Ratnayake C, et al. Victims' response to snakebite and socio-epidemiological factors of 1018 snakebites in a tertiary care hospital in Sri Lanka. *Wilderness Environ Med.* 2014; 25(1):35–40. <https://doi.org/10.1016/j.wem.2013.10.009> PMID: 24412659
29. Jelinek GA, Hamilton T, Hirsch RL. Admissions for suspected snake bite to the Perth adult teaching hospitals, 1979 to 1988. *Med J Aust.* 1991; 155(11–12):761–4. Epub 1991/12/02. PMID: 1745167.
30. Jelinek GA, Breheny FX. Ten years of snake bites at Fremantle Hospital. *Med J Aust.* 1990; 153(11–12):658–61. Epub 1990/12/03. PMID: 2246987.
31. Munro J, Pearn JH. Snake bite in children: a five year population study from South-East Queensland. *J Paediatr Child Health.* 1978; 14(4):248–53.
32. Blaylock R. Snake bites at Triangle hospital: January 1975 to June 1981. *Cent Afr J Med.* 1982; 28(1):1–11. PMID: 7083313
33. Chippaux J-P, Amadi-Eddine S, Fagot P. Diagnostic et surveillance des hémorragies dues aux envenimations vipérines en savane africaine. *Bull Soc Pathol Exot.* 1999; 92(2):109–13. PMID: 10399601
34. Tan HH. Epidemiology of snakebites from a general hospital in Singapore: a 5-year retrospective review (2004–2008). *Ann Acad Med Singapore.* 2010; 39(8):640–7. Epub 2010/09/15. PMID: 20838707.
35. Blaylock R. Epidemiology of snakebite in Eshowe, KwaZulu-Natal, South Africa. *Toxicon.* 2004; 43(2):159–66. <https://doi.org/10.1016/j.toxicon.2003.11.019> PMID: 15019475
36. Pattinson JP, Kong VY, Bruce JL, Oosthuizen GV, Bekker W, Laing GL, et al. Defining the need for surgical intervention following a snakebite still relies heavily on clinical assessment: The experience in Pietermaritzburg, South Africa. *SAMJ, S. Afr. Med* 2017; 107(12):1082–5. <https://doi.org/10.7196/SAMJ.2017.v107i12.12628> WOS:000418339200021. PMID: 29262961
37. Bawaskar HS, Bawaskar PH. Profile of snakebite envenoming in western Maharashtra, India. *Trans R Soc Trop Med Hyg.* 2002; 96(1):79–84. Epub 2002/04/03. [https://doi.org/10.1016/S0035-9203\(02\)90250-6](https://doi.org/10.1016/S0035-9203(02)90250-6) PMID: 11926002.
38. Logaraj M, Thirumavalavan R, Gopalakrishnan S. Epidemiology of snakebite reported in a Medical College Hospital in Tamil Nadu, *Int J Health Allied Sci.* 2013; 2(1):53.
39. Matthai TP, Date A. Acute renal failure in children following snake bite. *Ann Trop Paediatr.* 1981; 1(2):73–6. Epub 1981/06/01. <https://doi.org/10.1080/02724936.1981.11748064> PMID: 6185055.
40. Sharma LR, Lal V, Simpson ID. Snakes of medical significance in India: the first reported case of envenoming by the Levantine viper (*Macrovipera lebetina*). *Wilderness Environ Med.* 2008; 19(3):195–8. Epub 2008/08/22. <https://doi.org/10.1580/07-WEME-CR-175.1> PMID: 18715121.
41. Manosalva-Sánchez C, Zuleta-Dueñas LP, Castañeda-Porras O. Estudio descriptivo del accidente ofídico, Casanare-Colombia, 2012–2014. *MedUNAB.* 2018; 20(3):338–48.
42. Lukšić B, Bradarić N, Prgomet S. Venomous snakebites in southern Croatia. *Coll Antropol* 2006; 30(1):191–7. PMID: 16617597
43. Frangides CY, Koulouras V, Kouni SN, Tzortzatos GV, Nikolaou A, Pneumaticsos J, et al. Snake venom poisoning in Greece. Experiences with 147 cases. *Eur J Intern Med.* 2006; 17(1):24–7. <https://doi.org/10.1016/j.ejim.2005.10.001> PMID: 16378881

44. Ariaratnam CA, Sheriff MHR, Arambepola C, Theakston RDG, Warrell DA. Syndromic Approach to Treatment of Snake Bite in Sri Lanka Based on Results of a Prospective National Hospital-Based Survey of Patients Envenomed by Identified Snakes. *Am J Trop Med Hyg.* 2009; 81(4):725–31. <https://doi.org/10.4269/ajtmh.2009.09-0225> WOS:000270474000035. PMID: 19815895
45. Coetzer PW, Tilbury CR. The epidemiology of snakebite in northern Natal. *S Afr Med J.* 1982; 62(7):206–12. Epub 1982/08/07. PMID: 7101072.
46. Elayoubi S. Envenimation par morsure de vipère en milieu pédiatrique 2015.
47. Chafiq F, El Hattimy F, Rhalem N, Chippaux J-P, Soulaymani A, Mokhtari A, et al. Snakebites notified to the poison control center of Morocco between 2009 and 2013. *J Venom Anim Toxins incl Trop Dis.* 2016; 22. <https://doi.org/10.1186/s40409-016-0065-8> WOS:000372121400001. PMID: 26985186
48. Yates VM, Lebas E, Orpiay R, Bale BJ. Management of snakebites by the staff of a rural clinic: the impact of providing free antivenom in a nurse-led clinic in Meserani, Tanzania. *Ann Trop Med Parasitol.* 2010; 104(5):439–48. Epub 2010/09/08. <https://doi.org/10.1179/136485910X12743554760306> PMID: 20819312.
49. Wagener M. Haemotoxic snakebite in rural KwaZulu-Natal, South Africa: A case presenting with haematemesis. *S Afr Med J.* 2016; 106(5):44–5. Epub 2016/05/04. <https://doi.org/10.7196/SAMJ.2016.v106i5.9124> PMID: 27138661.
50. McNally S, Reitz C. Victims of snakebite. A 5-year study at Shongwe Hospital, Kangwane, 1978–1982. *South Afr Med J.* 1987; 72(12):855–60.
51. Pugh RN, Theakston RD. A clinical study of viper bite poisoning. *Ann Trop Med Parasitol.* 1987; 81(2):135–49. Epub 1987/04/01. <https://doi.org/10.1080/00034983.1987.11812106> PMID: 3689023.
52. Michael GC, Thacher TD, Shehu MI. The effect of pre-hospital care for venomous snake bite on outcome in Nigeria. *Trans R Soc Trop Med Hyg.* 2011; 105(2):95–101. Epub 2010/11/03. <https://doi.org/10.1016/j.trstmh.2010.09.005> PMID: 21035155.
53. Tianyi FL, Agbor VN, Tochie JN, Kadia BM, Nkwescheu AS. Community-based audits of snake envenomations in a resource-challenged setting of Cameroon: case series. *BMC Res Notes.* 2018; 11(1):317. Epub 2018/05/20. <https://doi.org/10.1186/s13104-018-3409-3> PMID: 29776445; PubMed Central PMCID: PMC5960191.
54. Einterz EM, Bates ME. Snakebite in northern Cameroon: 134 victims of bites by the saw-scaled or carpet viper, *Echis ocellatus*. *Trans R Soc Trop Med Hyg.* 2003; 97(6):693–6. Epub 2005/08/25. [https://doi.org/10.1016/s0035-9203\(03\)80105-0](https://doi.org/10.1016/s0035-9203(03)80105-0) PMID: 16117965.
55. Harris JB, Faiz MA, Rahman MR, Jalil MM, Ahsan MF, Theakston RDG, et al. Snake bite in Chittagong Division, Bangladesh: a study of bitten patients who developed no signs of systemic envenoming. *Trans R Soc Trop Med Hyg.* 2010; 104(5):320–7. <https://doi.org/10.1016/j.trstmh.2009.12.006> PMID: 20096910
56. Sharma SK, Kuch U, Hoede P, Bruhse L, Pandey DP, Ghimire A, et al. Use of Molecular Diagnostic Tools for the Identification of Species Responsible for Snakebite in Nepal: A Pilot Study. *PLoS Negl Trop Dis.* 2016; 10(4). <https://doi.org/10.1371/journal.pntd.0004620> WOS:000375376700054. PMID: 27105074
57. Su H-Y, Huang S-W, Mao Y-C, Liu M-W, Lee K-H, Lai P-F, et al. Clinical and laboratory features distinguishing between *Deinagkistrodon acutus* and *Daboia siamensis* envenomation. *J Venom Anim Toxins incl Trop Dis.* 2018; 24. <https://doi.org/10.1186/s40409-018-0179-2> WOS:000454502200001. PMID: 30607144
58. Mao Y-C, Liu P-Y, Chiang L-C, Liao S-C, Su H-Y, Hsieh S-Y, et al. *Bungarus multicinctus multicinctus* Snakebite in Taiwan. *Am J Trop Med Hyg.* 2017; 96(6):1497–504. <https://doi.org/10.4269/ajtmh.17-0005> PMID: 28719273
59. Rathnayaka RN, Kularatne S, Kumarasinghe K, Ranaweera J, Ranathunga PN. Ischemic brain infarcts and intracranial haemorrhages following Russell's viper (*Daboia russelii*) bite in Sri Lanka. *Toxicon.* 2017; 125:70–3. <https://doi.org/10.1016/j.toxicon.2016.11.253> PMID: 27871786
60. Silva A, Gamlaksha D, Waidyaratne D. Medico-legal significance of the identification of offending snake in a fatal snake bite: a case report. *J Forensic Leg Med.* 2013; 20(8):965–7. Epub 2013/11/19. <https://doi.org/10.1016/j.jflm.2013.09.009> PMID: 24237800.
61. Senanayake MP, Ariaratnam CA, Abeywickrema S, Belligaswatte A. Two Sri Lankan cases of identified sea snake bites, without envenoming. *Toxicon.* 2005; 45(7):861–3. Epub 2005/05/21. <https://doi.org/10.1016/j.toxicon.2005.02.010> PMID: 15904681.
62. Seneviratne U, Dissanayake S. Neurological manifestations of snake bite in Sri Lanka. *J Postgrad Med.* 2002; 48(4):275–8; discussion 8–9. Epub 2003/02/07. PMID: 12571382.

63. Kularatne S, Budagoda B, Gawarammana I, Kularatne W. Epidemiology, clinical profile and management issues of cobra (*Naja naja*) bites in Sri Lanka: first authenticated case series. *Trans R Soc Trop Med Hyg.* 2009; 103(9):924–30. <https://doi.org/10.1016/j.trstmh.2009.04.002> PMID: 19439335
64. Kularatne SAM, Sivansuthan S, Medagedara SC, Maduwage K, de Silva A. Revisiting saw-scaled viper (*Echis carinatus*) bites in the Jaffna Peninsula of Sri Lanka: distribution, epidemiology and clinical manifestations. *Trans R Soc Trop Med Hyg.* 2011; 105(10):591–7. <https://doi.org/10.1016/j.trstmh.2011.07.010> WOS:000295771300009. PMID: 21868049
65. Shahmy S, Kularatne SAM, Rathnayake SS, Dawson AH. A prospective cohort study of the effectiveness of the primary hospital management of all snakebites in Kurunegala district of Sri Lanka. *PLoS Negl Trop Dis.* 2017; 11(8):e0005847. Epub 2017/08/23. <https://doi.org/10.1371/journal.pntd.0005847> PMID: 28827807; PubMed Central PMCID: PMC5578683.
66. Seneviratne SL, Opanayaka CJ, Ratnayake NS, Kumara KE, Sugathadasa AM, Weerasuriya N, et al. Use of antivenom serum in snake bite: a prospective study of hospital practice in the Gampaha district. *Ceylon Med J.* 2000; 45(2):65–8. Epub 2000/10/29. <https://doi.org/10.4038/cmj.v45i2.8003> PMID: 11051703.
67. Kularatne SA. Common krait (*Bungarus caeruleus*) bite in Anuradhapura, Sri Lanka: a prospective clinical study, 1996–98. *Postgrad Med J.* 2002; 78(919):276–80. Epub 2002/08/02. <https://doi.org/10.1136/pmj.78.919.276> PMID: 12151569; PubMed Central PMCID: PMC1742360.
68. Blessmann J, Khonesavanh C, Outhaithit P, Manichanh S, Somphanthabansouk K, Siboulapha P. Venomous snake bites in Lao PDR: a retrospective study of 21 snakebite victims in a provincial hospital. *Southeast Asian J Trop Med Public Health.* 2010; 41(1):195–202. Epub 2010/06/29. PMID: 20578499.
69. Vongphoumy I, Chanthilat P, Vilayvong P, Blessmann J. Prospective, consecutive case series of 158 snakebite patients treated at Savannakhet provincial hospital, Lao People's Democratic Republic with high incidence of anaphylactic shock to horse derived F(ab')<sub>2</sub> antivenom. *Toxicon.* 2016; 117:13–21. Epub 2016/03/21. <https://doi.org/10.1016/j.toxicon.2016.03.011> PMID: 26995210.
70. Asif N, Akhtar F, Kamal K. A study of ninety snake bite cases at Pakistan Air Force (PAF) Hospital, Shorkot, Pakistan. *Pak Armed Forces Med J.* 2015; 65(3):333–8.
71. Tripathy S, Routray PK, Mohapatra AK, Mohapatra M, Dash SC. Acute demyelinating encephalomyelitis after anti-venom therapy in Russell's viper bite. *J Med Toxicol.* 2010; 6(3):318–21. Epub 2010/03/20. <https://doi.org/10.1007/s13181-010-0015-8> PMID: 20237970; PubMed Central PMCID: PMC3550494.
72. Bawaskar HS, Bawaskar PH. Envenoming by the common krait (*Bungarus caeruleus*) and Asian cobra (*Naja naja*): clinical manifestations and their management in a rural setting. *Wilderness Environ Med.* 2004; 15(4):257–66. Epub 2005/01/08. [https://doi.org/10.1580/1080-6032\(2004\)015\[0257:ebtckbj\]2.0.co;2](https://doi.org/10.1580/1080-6032(2004)015[0257:ebtckbj]2.0.co;2) PMID: 15636376.
73. Kumar V, Sabitha P. Inadequacy of present polyspecific anti snakevenom—a study from central Kerala. *Indian J Pediatr.* 2011; 78(10):1225–8. Epub 2011/03/04. <https://doi.org/10.1007/s12098-011-0396-y> PMID: 21369925.
74. Farooqui JM, Mukherjee BB, Manjhi SNM, Farooqui AAJ, Datir S. Incidence of fatal snake bite in Loni, Maharashtra: An autopsy based retrospective study (2004–2014). *J Forensic Leg Med.* 2016; 39:61–4. <https://doi.org/10.1016/j.jflm.2016.01.013> WOS:000371791700010. PMID: 26854851
75. Vaiyapuri S, Vaiyapuri R, Ashokan R, Ramasamy K, Nattamaisundar K, Jeyaraj A, et al. Snakebite and its socio-economic impact on the rural population of Tamil Nadu, India. *PLoS One.* 2013; 8(11):e80090. <https://doi.org/10.1371/journal.pone.0080090> PMID: 24278244
76. Gopalakrishnan M, Vinod KV, Dutta TK, Shaha KK, Sridhar MG, Saurabh S. Exploring circulatory shock and mortality in viper envenomation: a prospective observational study from India. *QJM.* 2018; 111(11):799–806. Epub 2018/08/15. <https://doi.org/10.1093/qjmed/hcy175> PMID: 30107433.
77. Mahaba HM. Snakebite: Epidemiology, prevention, clinical presentation and management. *Ann Saudi Med.* 2000; 20(1):66–8. <https://doi.org/10.5144/0256-4947.2000.66> PMID: 17322751
78. Looareesuwan S, Viravan C, Warrell DA. Factors contributing to fatal snake bite in the rural tropics: analysis of 46 cases in Thailand. *Trans R Soc Trop Med Hyg.* 1988; 82(6):930–4. Epub 1988/01/01. [https://doi.org/10.1016/0035-9203\(88\)90046-6](https://doi.org/10.1016/0035-9203(88)90046-6) PMID: 3257001.
79. Viravan C, Veeravat U, Warrell M, Theakston R, Warrell D. ELISA confirmation of acute and past envenoming by the monocellate Thai cobra (*Naja kaouthia*). *Am J Trop Med Hyg.* 1986; 35(1):173–81. <https://doi.org/10.4269/ajtmh.1986.35.173> PMID: 3946735
80. Hutton RA, Looareesuwan S, Ho M, Silamut K, Chanthavanich P, Karbwang J, et al. Arboreal green pit vipers (genus *Trimeresurus*) of South-East Asia: bites by *T. albolabris* and *T. macrops* in Thailand and a review of the literature. *Trans R Soc Trop Med Hyg.* 1990; 84(6):866–74. Epub 1990/11/01. [https://doi.org/10.1016/0035-9203\(90\)90111-q](https://doi.org/10.1016/0035-9203(90)90111-q) PMID: 2096527.



81. Rojnuckarin P, Intragumtornchai T, Sattapiboon R, Muanpasitporn C, Pakmanee N, Khoo O, et al. The effects of green pit viper (*Trimeresurus albolabris* and *Trimeresurus macrops*) venom on the fibrinolytic system in human. *Toxicon*. 1999; 37(5):743–55. Epub 1999/04/29. [https://doi.org/10.1016/S0041-0101\(98\)00214-1](https://doi.org/10.1016/S0041-0101(98)00214-1) PMID: 10219986.
82. Buranasin P. Snakebites at Maharat Nakhon Ratchasima Regional Hospital. *Southeast Asian J Trop Med Public Health*. 1993; 24(1):186–92. Epub 1993/03/01. PMID: 8362295.
83. Aye K-P, Thanachartwet V, Soe C, Desakorn V, Chamnanchanunt S, Sahassananda D, et al. Predictive Factors for Death After Snake Envenomation in Myanmar. *Wilderness Environ Med*. 2018; 29(2):166–75. <https://doi.org/10.1016/j.wem.2018.01.001> WOS:000435429100004. PMID: 29572088
84. Warrell D. Bites by Russell's vipers (*Daboia russelii siamensis*) in Myanmar: effect of the snake's length and recent feeding on venom antigenaemia and severity of envenoming. *Trans R Soc Trop Med Hyg*. 1991; 85(6):804–8. [https://doi.org/10.1016/0035-9203\(91\)90464-a](https://doi.org/10.1016/0035-9203(91)90464-a) PMID: 1839340
85. Isbister GK, Gault A, Tasoulis T, O'Leary MA. A definite bite by the Ornamental Snake (*Denisonia maculata*) causing mild envenoming. *Clin Toxicol (Phila)*. 2016; 54(3):241–4. Epub 2016/02/09. <https://doi.org/10.3109/15563650.2015.1128545> PMID: 26852775.
86. Sprivilis P. Fatal intracranial haematomas in two patients with brown snake envenomation. *Med J Austr*. 1995; 162(10):557–.
87. Isbister GK, Dawson AH, Whyte IM. Two cases of bites by the black-bellied swamp snake (*Hemiaspis signata*). *Toxicon*. 2002; 40(3):317–9. Epub 2001/11/17. [https://doi.org/10.1016/S0041-0101\(01\)00221-5](https://doi.org/10.1016/S0041-0101(01)00221-5) PMID: 11711130.
88. Allen GE, Brown SG, Buckley NA, O'Leary MA, Page CB, Currie BJ, et al. Clinical effects and anti-venom dosing in brown snake (*Pseudonaja* spp.) envenoming—Australian snakebite project (ASP-14). *PLoS One*. 2012; 7(12):e53188. Epub 2013/01/10. <https://doi.org/10.1371/journal.pone.0053188> PMID: 23300888; PubMed Central PMCID: PMC3532501.
89. Razavi S, Weinstein SA, Bates DJ, Alfred S, White J. The Australian mulga snake (*Pseudechis australis*: Elapidae): report of a large case series of bites and review of current knowledge. *Toxicon*. 2014; 85:17–26. Epub 2014/04/15. <https://doi.org/10.1016/j.toxicon.2014.04.003> PMID: 24726467.
90. Currie BJ. Snakebite in tropical Australia: a prospective study in the "Top End" of the Northern Territory. *Med J Aust*. 2004; 181(11–12):693–7. Epub 2004/12/14. PMID: 15588215.
91. Villanueva M, Maguiña C, Cabada M, De Marini J, Alvarez H, Gotuzzo E. Ofidismo en la provincia de Chanchamayo, Junín: revisión de 170 casos consecutivos en el Hospital de Apoyo de La Merced. *Rev Med Hered*. 2004; 15(2):82–7.
92. Smalligan R, Cole J, Brito N, Laing GD, Mertz BL, Manock S, et al. Crotaline snake bite in the Ecuadorian Amazon: randomised double blind comparative trial of three South American polyspecific antivenoms. *BMJ*. 2004; 329(7475):1129. Epub 2004/11/13. <https://doi.org/10.1136/bmj.329.7475.1129> PMID: 15539665; PubMed Central PMCID: PMC527684.
93. Bucarechi F, De Capitani EM, Branco MM, Fernandes LC, Hyslop S. Coagulopathy as the main systemic manifestation after envenoming by a juvenile South American rattlesnake (*Crotalus durissus terrificus*): case report. *Clin Toxicol (Phila)*. 2013; 51(6):505–8. Epub 2013/05/30. <https://doi.org/10.3109/15563650.2013.802796> PMID: 23713821.
94. Pinto RNL, Dasilva NJ, Aird SD. Human envenomation by the South American opisthoglyph *Clelia clelia plumbea* (Wied). *Toxicon*. 1991; 29(12):1512–6. [https://doi.org/10.1016/0041-0101\(91\)90008-f](https://doi.org/10.1016/0041-0101(91)90008-f) WOS:A1991GY76200009. PMID: 1801328
95. Brandao EO, de Bastos HC, Nishioka Sde A, Silveira PV. Lance-headed viper (*Bothrops moojeni*) bite wounding the eye. *Rev Inst Med Trop Sao Paulo*. 1993; 35(4):381–3. Epub 1993/07/01. <https://doi.org/10.1590/S0036-46651993000400014> PMID: 8115801.
96. Correia JM, Santana Neto PdL, Sabino Pinho MS, da Silva JA, Porto Amorim ML, Costa Escobar JA. Poisoning due to *Philodryas oifersii* (Lichtenstein, 1823) attended at Restauracao Hospital in Recife, State of Pernambuco, Brazil: case report. *Rev Soc Bras Med Trop*. 2010; 43(3):336–8. <https://doi.org/10.1590/S0037-86822010000300025> WOS:000279011700025. PMID: 20563508
97. Nishioka SA, Silveira PV. *Philodryas patagoniensis* bite and local envenoming. *Rev Inst Med Trop Sao Paulo*. 1994; 36(3):279–81. Epub 1994/05/01. <https://doi.org/10.1590/S0036-46651994000300013> PMID: 7855493.
98. Silveira GG, Machado CR, Tuyama M, Lima MA. Intracranial Bleeding Following *Bothrops* sp. Snakebite. *Neurologist*. 2016; 21(1):11–2. Epub 2015/12/26. <https://doi.org/10.1097/NRL.000000000000067> PMID: 26703003.
99. Parda PPD, Souza SM, Monteiro M, Fan HW, Cardoso JLC, Franca FOS, et al. Clinical trial of two antivenoms for the treatment of *Bothrops* and *Lachesis* bites in the north eastern Amazon region of Brazil. *Trans R Soc Trop Med Hyg*. 2004; 98(1):28–42. [https://doi.org/10.1016/S0035-9203\(03\)00005-1](https://doi.org/10.1016/S0035-9203(03)00005-1) WOS:000187423600004. PMID: 14702836

100. Bucarechi F, Hyslop S, Vieira RJ, Toledo AS, Madureira PR, de Capitani EM. Bites by coral snakes (*Micrurus* spp.) in Campinas, State of Sao Paulo, Southeastern Brazil. *Rev Inst Med Trop Sao Paulo*. 2006; 48(3):141–5. Epub 2006/07/19. <https://doi.org/10.1590/s0036-46652006000300005> PMID: 16847503.
101. Silveria PV, Nishioka Sde A. Non-venomous snake bite and snake bite without envenoming in a Brazilian teaching hospital. Analysis of 91 cases. *Rev Inst Med Trop Sao Paulo*. 1992; 34(6):499–503. Epub 1992/11/01. <https://doi.org/10.1590/s0036-46651992000600002> PMID: 1342117.
102. Roriz K, Zaqueo KD, Setubal SS, Katsuragawa TH, Silva RRD, Fernandes CFC, et al. Epidemiological study of snakebite cases in Brazilian Western Amazonia. *Rev Soc Bras Med Trop*. 2018; 51(3):338–46. Epub 2018/07/05. <https://doi.org/10.1590/0037-8682-0489-2017> PMID: 29972565.
103. Waldez F, Vogt RC. Aspectos ecológicos e epidemiológicos de acidentes ofídicos em comunidades ribeirinhas do baixo rio Purus, Amazonas, Brasil. *Acta Amazonica*. 2009; 39(3):681–92. <https://doi.org/10.1590/s0044-59672009000300025> SCIELO:S0044-59672009000300025.
104. Oliveira FN, Brito MT, Oliveira de Moraes IC, Lia Fook SM, de Albuquerque HN. Accidents caused by Bothrops and Bothropoides in the State of Paraíba: epidemiological and clinical aspects. *Rev Soc Bras Med Trop*. 2010; 43(6):662–7. <https://doi.org/10.1590/s0037-86822010000600012> WOS:000285513300012. PMID: 21181019
105. Nishioka SD, Silveira PVP. A clinical and epidemiologic study of 292 cases of Lance-headed viper bite in a Brazilian teaching hospital. *Am J Trop Med Hyg*. 1992; 47(6):805–10. WOS:A1992KF92000012.
106. Nishioka Sde A, Silveira PV, Bauab FA. Bite marks are useful for the differential diagnosis of snakebite in Brazil. *Wilderness Environ Med*. 1995; 6(2):183–8. Epub 1995/05/01. [https://doi.org/10.1580/1080-6032\(1995\)006\[0183:bmauft\]2.3.co;2](https://doi.org/10.1580/1080-6032(1995)006[0183:bmauft]2.3.co;2) PMID: 11995906.
107. Bosak AR, Ruha AM, Graeme KA. A case of neurotoxicity following envenomation by the Sidewinder rattlesnake, *Crotalus cerastes*. *J Med Toxicol*. 2014; 10(2):229–31. Epub 2014/01/15. <https://doi.org/10.1007/s13181-013-0373-0> PMID: 24414250; PubMed Central PMCID: PMC4057548.
108. Thorson A, Lavonas EJ, Rouse AM, Kerns WP, 2nd. Copperhead envenomations in the Carolinas. *J Toxicol Clin Toxicol*. 2003; 41(1):29–35. Epub 2003/03/21. <https://doi.org/10.1081/clt-120018268> PMID: 12645965.
109. Garcia-Gubern C, Bello R, Rivera V, Rocafort A, Colon-Rolon L, Acosta-Tapia H. Is the Puerto Rican racer, *Alsophis portoricensis*, really harmless? A case report series. *Wilderness Environ Med*. 2010; 21(4):353–6. Epub 2010/12/21. <https://doi.org/10.1016/j.wem.2010.07.001> PMID: 21168790.
110. Heiner JD, Bebaria VS, Varney SM, Bothwell JD, Cronin AJ. Clinical effects and antivenom use for snake bite victims treated at three US hospitals in Afghanistan. *Wilderness Environ Med*. 2013; 24(4):412–6. <https://doi.org/10.1016/j.wem.2013.05.001> PMID: 23870762
111. Bawaskar HS, Bawaskar PH, Punde DP, Inamdar MK, Dongare RB, Bhoite RR. Profile of snakebite envenoming in rural Maharashtra, India. *J Assoc Physicians India*. 2008; 56:88–95. Epub 2008/05/14. PMID: 18472507.
112. Bernarde PS, Gomes JdO. Venomous snakes and ophidism in Cruzeiro do Sul, Alto Juruá, State of Acre, Brazil. *Acta Amazonica*. 2012; 42(1):65–72.
113. Leite Rde S, Targino IT, Lopes YA, Barros RM, Vieira AA. Epidemiology of snakebite accidents in the municipalities of the state of Paraíba, Brazil. *Cien Saude Colet*. 2013; 18(5):1463–71. Epub 2013/05/15. <https://doi.org/10.1590/s1413-81232013000500032> PMID: 23670475.
114. Pierini S, Warrell D, De Paulo A, Theakston R. High incidence of bites and stings by snakes and other animals among rubber tappers and Amazonian Indians of the Juruá Valley, Acre State, Brazil. *Toxicol*. 1996; 34(2):225–36. [https://doi.org/10.1016/0041-0101\(95\)00125-5](https://doi.org/10.1016/0041-0101(95)00125-5) PMID: 8711756
115. Sharma SK, Chappuis F, Jha N, Bovier PA, Loutan L, Koirala S. Impact of snake bites and determinants of fatal outcomes in southeastern Nepal. *Am J Trop Med Hyg*. 2004; 71(2):234–8. PMID: 15306717
116. Habila I. Factors associated with snake bite and health seeking behavior among residents of Kaltungo Iga, Gombe State, Nigeria. Doctoral dissertation, Ahmadu Bello University, Zaria, 2014.
117. Joseph JK, Simpson ID, Menon NCS, Jose MP, Kulkarni KJ, Raghavendra GB, et al. First authenticated cases of life-threatening envenoming by the hump-nosed pit viper (*Hypnale hypnale*) in India. *Trans R Soc Trop Med Hyg*. 2007; 101(1):85–90. <https://doi.org/10.1016/j.trstmh.2006.03.008> WOS:000242759400011. PMID: 16839578
118. Viravan C, Looareesuwan S, Kosakarn W, Wuthiekanun V, McCarthy CJ, Stimson AF, et al. A national hospital-based survey of snakes responsible for bites in Thailand. *Trans R Soc Trop Med Hyg*. 1992; 86(1):100–6. [https://doi.org/10.1016/0035-9203\(92\)90463-m](https://doi.org/10.1016/0035-9203(92)90463-m) WOS:A1992HH64200040. PMID: 1566285



119. Cox RD, Parker CS, Cox ECE, Marlin MB, Galli RL. Misidentification of copperhead and cottonmouth snakes following snakebites. *Clin Toxicol (Phila)*. 2018; 56(12):1195–9. <https://doi.org/10.1080/15563650.2018.1473583> MEDLINE:29792342. PMID: 29792342
120. Ogunbanjo GA. Management of snakebites at a rural South African hospital. *South African Fam Pract*. 2009; 51(3).
121. de Medeiros CR, Hess PL, Nicoletti AF, Sueiro LR, Duarte MR, de Almeida-Santos SM, et al. Bites by the colubrid snake *Philodryas patagoniensis*: a clinical and epidemiological study of 297 cases. *Toxicol*. 2010; 56(6):1018–24. Epub 2010/07/21. <https://doi.org/10.1016/j.toxicol.2010.07.006> PMID: 20643156.
122. Vale TC, Leite AF, Hora PR, Coury MI, Silva RC, Teixeira AL. Bilateral posterior circulation stroke secondary to a crotalid envenomation: case report. *Rev Soc Bras Med Trop*. 2013; 46(2):255–6. Epub 2013/06/07. <https://doi.org/10.1590/0037-8682-1667-2013> PMID: 23740059.
123. de Medeiros CR, de Souza SN, da Silva MC, de Souza Ventura J, de Oliveira Pirelli R, Puerto G. Bites by *Tomodon dorsatus* (serpentes, dipsadidae): Clinical and epidemiological study of 86 cases. *Toxicol*. 2019.
124. Silveira PVP, Nishioka SdA. Non-venomous snake bite and snake bite without envenoming in a Brazilian teaching hospital: analysis of 91 cases. *Rev Inst Med Trop Sao Paulo*. 1992; 34(6):499–503. <https://doi.org/10.1590/s0036-46651992000600002> PMID: 1342117
125. Ismail AK, Weinstein SA, Auliya M, Sabardin DM, Herbosa TJ, Saiboon IM, et al. A bite by the Twin-Barred Tree Snake, *Chrysopelea pelias* (Linnaeus, 1758). *Clin Toxicol*. 2010; 48(3):222–6. <https://doi.org/10.3109/15563650903550964> WOS:000276762200011. PMID: 20345298
126. Weinstein SA, Warrell DA, White J, Keyler DE. “Venomous Bites from Non-Venomous Snakes: A Critical Analysis of Risk and Management of “Colubrid Snake Bites: Elsevier; 2011.
127. Trape JF. Partition d’*Echis ocellatus* Stemmler, 1970 (Squamata: Viperidae), avec la description d’une espèce nouvelle. *Bull Soc Herp Fr*. 2018; 167:13–34.
128. Cockrell M, Swanson K, Sanders A, Prater S, von Wenckstern T, Mick J. Safe Handling of Snakes in an ED Setting. *J Emerg Nurs*. 2017; 43(1):21–3. Epub 2016/11/16. <https://doi.org/10.1016/j.jen.2016.07.009> PMID: 27842799.
129. Suchard JR, LoVecchio F. Envenomations by rattlesnakes thought to be dead. *N Engl J Med*. 1999; 340(24):1930–. <https://doi.org/10.1056/NEJM199906173402420> PMID: 10375322
130. Griffen D, Donovan JW. Significant envenomation from a preserved rattlesnake head (in a patient with a history of immediate hypersensitivity to antivenin). *Ann Emerg Med*. 1986; 15(8):955–8. [https://doi.org/10.1016/s0196-0644\(86\)80685-0](https://doi.org/10.1016/s0196-0644(86)80685-0) PMID: 3740586
131. Willhite LA, Willenbring BA, Orozco BS, Cole JB. Death after bite from severed snake head. *Clin Toxicol (Phila)*. 2018; 56(9):864–5. Epub 2018/02/20. <https://doi.org/10.1080/15563650.2018.1439951> PMID: 29457505.
132. Emswiler MP, Griffith 4th FP, Cumpston KL. Clinically significant envenomation from postmortem copperhead (*Agkistrodon contortrix*). *Wilderness Environ Med*. 2017; 28(1):43–5. <https://doi.org/10.1016/j.wem.2016.09.007> PMID: 27876196
133. Silva AMd Monteiro WM, Bernarde PS. Envenomation by a juvenile pit viper (*Bothrops atrox*) presumed to be dead. *Rev Soc Bras Med Trop*. 2019; 52.
134. Silva A, Marikar F, Murugananthan A, Agampodi S. Awareness and perceptions on prevention, first aid and treatment of snakebites among Sri Lankan farmers: a knowledge practice mismatch? *J Occup Med Toxicol*. 2014; 9(1):20.
135. Price L., Governance and Ecology: Managing the Menace of Venomous Snakes in Colonial India. *Cult Soc Hist*. 2017; 14(2):201–17.
136. Fita DS, Neto ECM, Schiavetti A. ‘Offensive’ snakes: cultural beliefs and practices related to snakebites in a Brazilian rural settlement. *J Ethnobiol Ethnomed*. 2010; 6(1):13.
137. Snakebite Healing and Education Society [cited 5 November 2019]. Available from: <http://www.she-india.org/>
138. African Snakebite Institute [cited 5 November 2019]. Available from: <https://www.africansnakebiteinstitute.com/snakebite/>
139. BIG 4 Mapping Project [cited 5 November 2019]. Available from: <http://snakebiteinitiative.in/snake/>
140. Maduwage K. Snake Identification Service of Sri Lanka 2017 [cited 5 November 2019]. Available from: <https://snakesidentification.org/>
141. Thailand snakes. 2019 [cited 5 November 2019]. Available from: <https://www.thailandsnakes.com/thailand-snake-id/>

142. Florida's. Florida's Poison Control Centers—Snakes 2019 [cited 5 November 2019]. Available from: <https://floridapoisontcontrol.org/poisoning-in-florida/snakes/>
143. Ciemto UdA-FdM-. 2019 [cited 5 November 2019]. Available from: <http://ciemto.medicinaudea.co/>
144. Geneviève LD, Ray N, Chappuis F, Alcoba G, Mondardini MR, Bolon I, et al. Participatory approaches and open data on venomous snakes: A neglected opportunity in the global snakebite crisis? *PLoS Negl Trop Dis*. 2018; 12(3):e0006162. <https://doi.org/10.1371/journal.pntd.0006162> PMID: 29518075
145. Ruiz de Castañeda R, Grey F, Williams D. Citizen science could map snakebite risk. *Nature*. 2019; 571(7766):478.
146. Ruiz de Castañeda R, Durso AM, Ray N, Fernández JL, Williams DJ, Alcoba G, et al. Snakebite and snake identification: empowering neglected communities and health-care providers with AI. *The Lancet Digital Health*. 2019; 1(5):e202–e3.
147. ITUNews. Artificial Intelligence for Health: ITU and WHO accept 8 new use cases 2018 [cited 5 November 2019]. Available from: <https://news.itu.int/artificial-intelligence-health-new-use-cases/>
148. Ariaratnam CA, Thuraisingam V, Kularatne SA, Sheriff MH, Theakston RD, de Silva A, et al. Frequent and potentially fatal envenoming by hump-nosed pit vipers (*Hypnale hypnale* and *H. nepa*) in Sri Lanka: lack of effective antivenom. *Trans R Soc Trop Med Hyg*. 2008; 102(11):1120–6. Epub 2008/05/06. <https://doi.org/10.1016/j.trstmh.2008.03.023> PMID: 18455743.
149. Kipanyula MJ, Kimaro WH. Snakes and snakebite envenoming in Northern Tanzania: a neglected tropical health problem. *J Venom Anim Toxins Incl. Trop Dis*. 2015; 21. <https://doi.org/10.1186/s40409-015-0033-8> WOS:000359978800001. PMID: 26309444
150. Inthanomchanh V, Reyer JA, Blessmen J, Phrasisombath K, Yamamoto E, Hamajima N. Assessment of knowledge about snakebite management amongst healthcare providers in the provincial and two district hospitals in Savannakhet Province, Lao PDR. *Nagoya J Med Sci*. 2017; 79(3):299–311. Epub 2017/09/08. <https://doi.org/10.18999/najims.79.3.299> PMID: 28878435; PubMed Central PMCID: PMC5577016.
151. Sharma SK. Venomous snakes of Nepal: a photographic guide: BP Koirala Institute of Health Sciences; 2013. Available from: [http://www.bik-f.de/files/publications/kuch\\_venomous\\_snakes\\_of\\_nepal\\_-\\_english\\_edition.pdf](http://www.bik-f.de/files/publications/kuch_venomous_snakes_of_nepal_-_english_edition.pdf)
152. WHO. Venomous snakes and antivenoms search interface 2010 [cited 5 November 2019]. Available from: <http://apps.who.int/bloodproducts/snakeantivenoms/database/snakeframeset.html>
153. Carrasco PA, Venegas PJ, Chaparro JC, Scrocchi GJ. Nomenclatural instability in the venomous snakes of the *Bothrops* complex: Implications in toxinology and public health. *Toxicon*. 2016; 119:122–8. <https://doi.org/10.1016/j.toxicon.2016.05.014> PMID: 27242040
154. Williams D, Wüster W, Fry BG. The good, the bad and the ugly: Australian snake taxonomists and a history of the taxonomy of Australia's venomous snakes. *Toxicon*. 2006; 48(7):919–30. <https://doi.org/10.1016/j.toxicon.2006.07.016> PMID: 16999982