Litton, Felix Oberender, Forbes McGain, Gavin Salt, Glenn Eastwood, Gopal Taori, Hannah Thompson, Hayden White, Hergen Buscher, Ian Seppelt, Ifrah Khan, Janelle Young, Jayshree Lavana, Jeremy Cohen, Jessica Lugsdin, Jill Garlick, Jim Buttery, John Botha, John Santamaria, Jonathan Barrett, Kasha Singh, Kevin Laupland, Khaled El-Khawas, Kristine Estensen, Kush Deshpande, Kyle White, Leigh Fitzpatrick, Lewis Campbell, Mahesh Ramanan, Manoj Saxena, Marie Draper, Marion Kainer, Mark Kol, Mark Page, Mark Plummer, Martin Sterba, Matthew Anstey, Matthew Brain, Matthew Maiden, Myrene Kilminster, Naomi Hammond, Neeraj Bhadange, Nicole Humphreys, Paras Jain, Paul Azzi, Paul Secombe, Paula Lister, Peter Chan, Peter McCanny, Phillip Britton, Pierre Janin, Rashmi Runiyar, Ravi Krishnamurthy, Ravikiran Sonawane, Ravindranath Tiruvoipati, Rebecca Jessup, Richard Totaro, Rinaldo Bellomo, Ritesh Sanghavi, Samantha Bates, Sandra Peake, Shailesh Bihari, Shane George, Sharon Waterson, Simon Erickson, Steve Webb, Subhash Arora, Subodh Ganu, Thomas Rozen, Toni McKenna, Umesh Kadam, Vineet Nayyar, Wei Han Choy, and Wisam Albassam.

References

- Majumder MS, Mandl KD. Early in the epidemic: impact of preprints on global discourse about COVID-19 transmissibility. *Lancet Glob Health* 2020;8:e627–e630.
- Horby P, Lim WS, Emberson JR, Mafham M, Bell JL, Linsell L, et al.; RECOVERY Collaborative Group. Dexamethasone in hospitalized patients with COVID-19: preliminary report. N Engl J Med [online ahead of print] 17 Jul 2020; DOI: 10.1056/NEJMoa2021436.
- Australian and New Zealand Intensive Care Society. COVID-19 guidelines: version 1. Melbourne, Australia: Australian and New Zealand Intensive Care Society; 2020 [updated 2020 Mar 16; accessed 2020 Nov 12]. Available from: https://www.anzics.com.au/wp-content/uploads/2020/ 03/ANZICS-COVID-19-Guidelines-Version-1.pdf.
- Horby P, Lim WS, Emberson JR, Mafham M, Bell JL, Linsell L, et al.; RECOVERY Collaborative Group. Effect of dexamethasone in hospitalized patients with COVID-19: preliminary report [preprint]. medRxiv; 2020 [accessed 2020 Sep 27]. Available from: https:// doi.org/10.1101/2020.06.22.20137273.
- Venkatesh B, Finfer S, Cohen J, Rajbhandari D, Arabi Y, Bellomo R, et al.; ADRENAL Trial Investigators; Australian–New Zealand Intensive Care Society Clinical Trials Group. Adjunctive glucocorticoid therapy in patients with septic shock. N Engl J Med 2018;378:797–808.
- Bauchner H, Fontanarosa PB. Randomized clinical trials and COVID-19: managing expectations. JAMA 2020;323:2262–2263.
- Sterne JAC, Murthy S, Diaz JV, Slutsky AS, Villar J, Angus DC, et al.; WHO Rapid Evidence Appraisal for COVID-19 Therapies (REACT) Working Group. Association between administration of systemic corticosteroids and mortality among critically ill patients with COVID-19: a meta-analysis. *JAMA* 2020;324:1330–1341.

Copyright © 2021 by the American Thoracic Society

Check for updates

High Prevalence of Pulmonary Sequelae at 3 Months after Hospital Discharge in Mechanically Ventilated Survivors of COVID-19

To the Editor:

Severe coronavirus disease (COVID-19) is characterized by acute hypoxemic respiratory failure, usually with extensive consolidations

and areas with ground glass on chest computed tomographic (CT) scans (1). Whether long-term respiratory sequelae persist in survivors of severe COVID-19 remains to be established. This report describes our findings of respiratory outcomes in mechanically ventilated survivors of COVID-19 at 3 months after hospital discharge.

Methods

We recorded clinical and follow-up data of all patients with COVID-19 treated at our ICU in the Maastricht Intensive Care COVID cohort (registered in the Netherlands Trial Register [NL8613]) (2). The institutional review board of Maastricht University Medical Center+ approved the study, and informed consent was obtained (METC2020–2287). During admission, ventilator strategies included lung-protective ventilation ($V_T \le 6$ ml/kg) and positive end-expiratory pressure titration using electrical impedance tomography. Prone positioning was considered when the Pa_{O_2}/F_{IO_2} ratio was less than 112.5 mm Hg (15 kPa) and maintained for at least 12 hours.

At 3 months after hospital discharge, survivors were screened at a multidisciplinary post-ICU outpatient clinic for respiratory outcomes with pulmonary function testing (PFT), including spirometry, lung volumes, and diffusing capacity for carbon monoxide adjusted for Hb, chest high-resolution CT (HRCT) imaging, and 6-minute-walk test (6-MWT). Two experienced radiologists systematically scored chest HRCT scans for the presence of pulmonary abnormalities, including ground-glass opacifications, reticulation, consolidations, bronchiectasis, atelectasis, presence of new emphysema, cystic changes, air trapping, extent of lobe involvement, and total lung involvement. The extent of lobe involvement was visually scored on a 0-5 scale, as follows: 0 = no involvement, 1 = 1-5%, 2 = 6-25%, 3 = 26-50%, 4 = 51-75%, and 5 = >75% involvement (3). The CT Severity Score (CTSS) was calculated by adding the lobar scores. HRCT scans were compared with scans performed at presentation (n = 33) at the emergency department or during admission (n = 5), depending on availability. All data are presented as median (interquartile range [IQR]). Correlations between CTSS, PFT results, and 6-MWT were assessed using Spearman's rank correlation.

Results

During the first European pandemic wave between March and May 2020, the Maastricht Intensive Care COVID cohort included 94 patients. Fifty-two (55%) patients were alive 3 months after hospital discharge, and 48 of them (92%) participated in the follow-up clinic. The four missing patients attended follow-up elsewhere. Follow-up (IQR) occurred at a median of 120 (103–135) days after intubation and 90 (80–99) days after hospital discharge. Baseline characteristics are detailed in Table 1.

We found diminished TLC and diffusion capacity in 23 and 36 participants, respectively, but no airway obstruction on PFT (Table 1), whereas five participants had no abnormalities. The median 6-MWT result was 482 m (82% of predicted distance). Two participants were on home supplemental oxygen, and four participants experienced a significant saturation drop during this test (>4% drop).

Only two participants had no signs of COVID-19-related abnormalities at follow-up HRCT scan. HRCT scans showed ground-glass opacities in 89% (n = 41) of cases. Signs of reticulation, including course fibrous bands either with or without obvious parenchymal distortion, bronchiectasis, and bronchiolectasis, were seen in 67% (n = 31) of cases and were

³This article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License 4.0 (http://creativecommons.org/licenses/by-nc-nd/4.0/). For commercial usage and reprints, please contact Diane Gern (dgern@thoracic.org).

Originally Published in Press as DOI: 10.1164/rccm.202010-3823LE on December 16, 2020

Table 1. Baseline Characteristics, PFT, and HRCT Imaging Results at 3-Month Follow-Up

Baseline Characteristics on Admission (N = 48)

Baseline Characteristics on Admission (N = 48	5)		
Age, yr Sex, M BMI, kg/m ² Origin of admission Emergency department Hospital ward Transfer from another ICU Pre-ICU length of stay, d APACHE II score Preexisting lung disease Asthma COPD Smoking status Current smoker Former smoker Charlson Comorbidity Index 0 1-2 3-4 5 or more Leukocyte count, 10 ⁻⁹ /L C-reactive protein, mg/L D-dimer, $\mu g/L$ Pac_/Flo_ratio, mm Hg Prone positioned during ICU admission Pisap, cm H ₂ O PEEP, cm H ₂ O VT/kg bodyweight, ml/kg Dynamic compliance, ml/cm H ₂ O Received steroids during admission* IMV duration, d ICU length of stay, d Hospital length of stay, d Hospital discharge location Home Nursing home Rehabilitation center Rehabilitation center IECMO during admission	$\begin{array}{c} 33 \ (6\\ 27.68 \ (2\\ 27.68 \ (2\\ 27 \ (5\\ 10 \ (2\\ 3.00 \ (1\\ 15.0 \ (1\\ 15.0 \ (1\\ 3.6\\ 0 \ (0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0$	5.18-30.47) 2.9) 6.2) 0.8) 0.0-5.00) 3.0-17.3) .2 .2) .2) .2) .2) .2) .30 .2) .2 .2) .2 .2) .2 .2) .2 .2 .2) .2 .2) .2 .2 .2) .2 .2 .2) .2 .2 .2) .2	
PFT (N = 43) FEV_1, L $FEV_1/VC, \%$ FVC, L RV, L ILC, L D_{LCO}, L^{\ddagger} 6-MWT, m [§] MRC Dyspnea score Grade 0–1 (none/mild) Grade 2–3 (moderate) Grade 4–5 (severe)	Absolute Value 2.9 (2.6–3.5) 79.9 (76.1–86.6) 3.6 (3.1–4.2) 2.0 (1.6–2.2) 5.6 (4.6–6.7) 5.4 (4.6–6.3) 480.0 (386.0–536.0) 27 (62.8) 14 (32.5) 2 (4.7)	Percentage of Predicted 95.0 (77.0-104.5) — 87.0 (70.0-106.0) 88.0 (70.0-103.0) 84.0 (71.5-102.5) 61.0 (50.0-69.0) 81.5 (69.5-99.5)	Below LLN 11 (25.6) 0 (0.0) 16 (37.2) 9 (20.9) 23 (53.5) 36 (87.8) —
HRCT Imaging Results (N = 46)			
Fibrosis Ground glass Atelectasis Dominant pattern Reticular Ground glass No abnormalities Decreased attenuation Due to small-airways disease Due to new emphysema CTSS	42 (91.3) 41 (89.1) 15 (32.6) 31 (67.4) 13 (28.3) 25 (54.3) 25 (54.3) 21 (45.7) 12 (25.0) 11.0 (5.0–15.0)		

Definition of abbreviations: 6-MWT = 6-minute-walk test; APACHE = Acute Physiology And Chronic Health Evaluation; BMI = body mass index; COPD = chronic obstructive pulmonary disease; CTSS = Computed Tomographic Severity Score; D_{LCO_c} = diffusing capacity for carbon monoxide adjusted for Hb; ECMO = extracorporeal membrane oxygenation; HRCT = high-resolution chest computed tomographic; IMV = invasive mechanical ventilation; LLN = lower limit of normal; MRC Dyspnea = Medical Research Council Dyspnea questionnaire; PEEP = positive end-expiratory pressure; PFT = pulmonary function testing; Pinsp = inspiratory pressure in bilevel pressure-controlled ventilation; RV = residual volume.

Data are presented as median (interquartile range) or n (%) unless indicated otherwise. For laboratory results and ventilator settings, the worst value for the first 24 hours of admission was recorded.

*Defined as receiving steroid treatment for at least 2 days or more.

[†]Three patients were still admitted to a rehabilitation center at the moment of follow-up.

 $^{\ddagger}_{2}D_{LCO_{c}}$ failed in two patients.

[§]Two patients were on supplemental oxygen while performing the 6-MWT.

assumed to represent fibrosis. One-quarter of the survivors showed new emphysematous destruction or cavitation that was not present at baseline scan or showed obvious deterioration of preexistent emphysema (Figure 1). Some air trapping was common, but it was not a dominant feature. Traction bronchiectasis was rare and not a dominant feature at follow-up.

Total severity scores for the 3-month follow-up scans ranged from 0 to 25, with a median score of 11. Participants with limited residual changes mainly showed subpleural parenchymal bands or small plate atelectasis. No predilection for a certain part of the lungs was noted. Residual lesions were predominantly located in the areas that showed crazy paving (ground glass with reticulation) at presentation, whereas areas with consolidations observed during admission appeared to be spared at 3 months. Diffusion capacity was significantly correlated with both TLC (ρ = 0.56; *P* < 0.001) and 6-MWT (ρ = 0.53, *P* < 0.001) but not with CTSS.

Discussion

We assessed respiratory sequelae of invasively mechanically ventilated patients with COVID-19 detailing both pulmonary function and HRCT scan results at 3 months after hospital discharge. Key findings were high prevalence of diminished diffusion capacity and TLC and fibrotic changes on HRCT images. These findings add to previous studies reporting a lower prevalence of pulmonary dysfunction in patients with COVID-19. However, these reports were based on less severely ill patients with COVID-19 who were not supported by invasive mechanical ventilation (4) or who were ventilated for a shorter duration (5). As such, our data represent the more severe spectrum of COVID-19 disease.

Based on our HRCT findings, fibrosis (evident from reticular pattern found in the majority of participants) and ground-glass opacifications were the dominant pathophysiological hallmarks observed. Notably, ground-glass opacifications were still present at 3 months after hospital discharge, mainly with a subpleural distribution similar to a nonspecific interstitial pneumonia pattern or a diffuse distribution with lower density than in areas of ground glass seen at baseline imaging. Whether these are signs of fibrosis or of ongoing inflammation is speculative, but this may become clear from follow-up imaging studies. Finally, we noted new emphysematous abnormalities both in areas showing a so-called vacuole sign at baseline imaging as well as in areas outside the areas with infiltration. The former might be explained by direct parenchymal destruction caused by infection; the latter finding could be a manifestation of ventilator-induced injury.

Long-term pulmonary effects have been documented in patients with acute respiratory distress syndrome (ARDS) and, to a lesser extent, in respiratory syndromes caused by coronaviruses such as severe acute respiratory syndrome (SARS) and Middle East Respiratory Syndrome (MERS) (6–8). Comparisons with our findings in COVID-19 should be interpreted with caution, as populations and follow-up strategies varied highly. In general, fibrotic changes on chest imaging were described in survivors of ARDS, SARS, and MERS (6, 7, 9). Fibrosis in our cohort was diffusely distributed in the lungs, which contrasted with the usually described localization in the anterior parts of the lungs in severe ARDS (9). To our knowledge, the new areas of emphysema we observed seem specific for COVID-19 compared with SARS or MERS (10).

In addition, the sparse data available suggest similar predominant reduction in diffusion capacity in survivors of COVID-19 compared with survivors of ARDS, SARS, and MERS (6–8). Whether this reduction in diffusion capacity is aggravated by the vascular and thrombotic complications witnessed in patients with COVID-19 remains to be investigated (11). Furthermore, as reported for ARDS, physical capacity was clearly diminished in our cohort, which correlated as expected with reduced lung function (8).

Strengths of our follow-up cohort include the inclusion of only mechanically ventilated patients and the high follow-up response rate. An important limitation is the lack of a direct comparison between survivors of COVID-19 and survivors of non-COVID-19 ARDS, which remains speculative at this point. Other limitations include the single-center character and the lack of baseline information on pulmonary function before the infection. Moreover, at the time of study initiation, the high incidence of pulmonary embolism in COVID-19 was not (yet) acknowledged and therefore not systematically screened for.

It is likely that the observed HRCT scan and PFT abnormalities will—at least partially—resolve over time, as was shown in

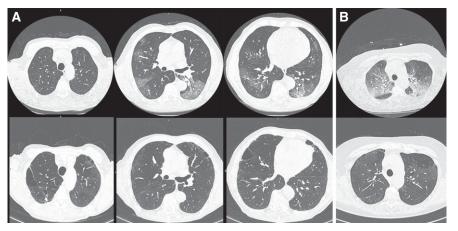


Figure 1. Representative high-resolution chest computed tomographic (HRCT) images of two of the survivors. (A) HRCT imaging performed at admission (upper row) and at 3-month follow-up (lower row). Chest computed tomographic (CT) imaging at admission shows typical bilateral subpleural ground-glass opacities. No signs of previous emphysema were detected. However, follow-up HRCT imaging shows obvious emphysematous destruction. (B) CT image at presentation at emergency department with evident ground areas with reticulation (crazy paving). Follow-up reveals diffuse areas of persistent ground glass without reticulation, as well as areas with low density in previously normal areas, possibly due to hypoperfusion.

long-term evaluations of survivors of SARS, MERS, and ARDS (6, 8, 12). Nevertheless, the long-term detrimental impact of these pulmonary sequalae on patient health and quality of life in survivors of ARDS is well established (13). Whether respiratory effects of COVID-19 hold similar implications has yet to be investigated. As such, our findings support long-term respiratory follow-up of mechanically ventilated patients with COVID-19.

Conclusions

The majority of invasively mechanically ventilated survivors of COVID-19 still had abnormal pulmonary function tests and residual changes on HRCT scans at 3 months after hospital discharge. Diminished diffusion capacity, diminished TLC, and fibrosis on HRCT were the dominant features. Our findings warrant intensive respiratory follow-up of mechanically ventilated patients with COVID-19.

Author disclosures are available with the text of this letter at www.atsjournals.org.

Rob J. J. van Gassel, M.D. Julia L. M. Bels, M.D. Anne Raafs, M.D. Bas C. T. van Bussel, M.D., Ph.D. Marcel C. G. van de Poll, M.D., Ph.D. Sami O. Simons, M.D., Ph.D. Lieke W. L. van der Meer, M.D. Hester A. Gietema, M.D., Ph.D. *Maastricht University Medical Centre Maastricht, the Netherlands*

Rein Posthuma, M.D. Maastricht University Medical Centre Maastricht, the Netherlands and

Centre of Expertise for Chronic Organ Failure (CIRO) Horn, the Netherlands

Susanne van Santen, M.D., Ph.D.* Maastricht University Medical Centre Maastricht, the Netherlands

ORCID IDs: 0000-0002-0780-2052 (R.J.J.v.G.); 0000-0001-9228-8045 (A.R.); 0000-0003-1621-7848 (B.C.T.v.B.); 0000-0002-3302-4063 (M.C.G.v.d.P.); 0000-0002-4296-5076 (S.O.S.); 0000-0001-9080-5977 (L.W.L.v.d.M.); 0000-0001-7036-3307 (H.A.G.); 0000-0003-0015-0116 (R.P.).

*Corresponding author (e-mail: susanne.van.santen@mumc.nl).

References

- Ferrando C, Suarez-Sipmann F, Mellado-Artigas R, Hernandez M, Gea A, Arruti E, et al.; COVID-19 Spanish ICU Network. Clinical features, ventilatory management, and outcome of ARDS caused by COVID-19 are similar to other causes of ARDS. *Intensive Care Med* 2020;46: 2200–2211. [Published erratum appears in *Intensive Care Med* 1–3.]
- Tas J, van Gassel RJJ, Heines SJH, Mulder MMG, Heijnen NFL, Acampo-de Jong MJ, *et al.* Serial measurements in COVID-19induced acute respiratory disease to unravel heterogeneity of the disease course: design of the Maastricht Intensive Care COVID cohort (MaastrlCCht). *BMJ Open* 2020;10:e040175.
- Gietema HA, Müller NL, Fauerbach PV, Sharma S, Edwards LD, Camp PG, et al.; Evaluation of COPD Longitudinally to Identify Predictive Surrogate Endpoints (ECLIPSE) investigators. Quantifying the extent of emphysema: factors associated with radiologists' estimations and quantitative indices of emphysema severity using the ECLIPSE cohort. Acad Radiol 2011;18:661–671.

- Frija-Masson J, Debray MP, Gilbert M, Lescure FX, Travert F, Borie R, et al. Functional characteristics of patients with SARS-CoV-2 pneumonia at 30 days post-infection. *Eur Respir J* 2020;56:2001754.
- Ramani C, Davis EM, Kim JS, Provencio JJ, Enfield KB, Kadl A. Post-ICU COVID-19 outcomes: a case series. *Chest* [online ahead of print] 21 Aug 2020; DOI: 10.1016/j.chest.2020.08.2056.
- Hui DS, Wong KT, Ko FW, Tam LS, Chan DP, Woo J, et al. The 1-year impact of severe acute respiratory syndrome on pulmonary function, exercise capacity, and quality of life in a cohort of survivors. Chest 2005;128:2247–2261.
- Park WB, Jun KI, Kim G, Choi JP, Rhee JY, Cheon S, *et al.* Correlation between pneumonia severity and pulmonary complications in Middle East respiratory syndrome. *J Korean Med Sci* 2018;33:e169.
- Herridge MS, Cheung AM, Tansey CM, Matte-Martyn A, Diaz-Granados N, Al-Saidi F, *et al.*; Canadian Critical Care Trials Group. One-year outcomes in survivors of the acute respiratory distress syndrome. *N Engl J Med* 2003;348:683–693.
- Desai SR, Wells AU, Rubens MB, Evans TW, Hansell DM. Acute respiratory distress syndrome: CT abnormalities at long-term followup. *Radiology* 1999;210:29–35.
- Antonio GE, Wong KT, Chu WC, Hui DS, Cheng FW, Yuen EH, et al. Imaging in severe acute respiratory syndrome (SARS). *Clin Radiol* 2003;58:825–832.
- 11. Patel BV, Arachchillage DJ, Ridge CA, Bianchi P, Doyle JF, Garfield B, et al. Pulmonary angiopathy in severe COVID-19: physiologic, imaging, and hematologic observations. *Am J Respir Crit Care Med* 2020;202:690–699.
- 12. Wilcox ME, Patsios D, Murphy G, Kudlow P, Paul N, Tansey CM, *et al.* Radiologic outcomes at 5 years after severe ARDS. *Chest* 2013;143: 920–926.
- Heyland DK, Groll D, Caeser M. Survivors of acute respiratory distress syndrome: relationship between pulmonary dysfunction and long-term health-related quality of life. *Crit Care Med* 2005;33:1549–1556.

Copyright © 2021 by the American Thoracic Society

Check for updates

SARS-CoV-2 Detected on Environmental Fomites for Both Asymptomatic and Symptomatic Patients with COVID-19

To the Editor:

Severe acute respiratory syndrome coronavirus (SARS-CoV) and Middle East respiratory syndrome coronavirus (MERS-CoV), relatives of SARS-CoV-2, are implicated in nosocomial outbreaks

Supported by the Ministry of Science and Technology (2020YFC0846300), Shenzhen Science and Technology Research and Development Project (202002073000001), Ministry of Science and Technology (2020YFC0846300), National Natural Science Foundation of China (81900071) and China Postdoctoral Science Foundation (2019M660836). The funding institutions had no roles in study design, data collection, data analysis, interpretation, or writing of the report in this study.

Author Contributions: M.Y., L.L., T.H., and Y.L. contributed to the study design. M.Y., T.H., S.L., Y.Y., Y.J., X.L., and J.Y. contributed to the collection of clinical data. M.Z. contributed to the viral RNA detection. M.Y. and Y.Y. contributed to the data analysis. M.Y., L.L., Y.Y., and Y.L. contributed to the manuscript preparation. All the authors have read, edited, and approved the manuscript.

Originally Published in Press as DOI: 10.1164/rccm.202006-2136LE on December 16, 2020

^aThis article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial No Derivatives License 4.0 (https://creativecommons.org/licenses/by-nc-nd/4.0/). For commercial usage and reprints, please contact Diane Gern (dgern@thoracic.org).