REVIEW

Open Access

From micro to macro, nanotechnology demystifies acute pancreatitis: a new generation of treatment options emerges



Wei Du¹, Xinyue Wang¹, Yuyan Zhou¹, Wencheng Wu^{2*}, Haojie Huang^{1*} and Zhendong Jin^{1*}

Abstract

Acute pancreatitis (AP) is a disease characterized by an acute inflammatory response in the pancreas. This is caused by the abnormal activation of pancreatic enzymes by a variety of etiologic factors, which results in a localized inflammatory response. The symptoms of this disease include abdominal pain, nausea and vomiting and fever. These symptoms are induced by a hyperinflammatory response and oxidative stress. In recent years, research has focused on developing anti-inflammatory and antioxidative therapies for the treatment of acute pancreatitis (AP). However, there are still limitations to this approach, including poor drug stability, low bioavailability and a short half-life. The advent of nanotechnology has opened up a novel avenue for the management of acute pancreatitis (AP). Nanomaterials can serve as an efficacious vehicle for conventional pharmaceuticals, enhancing their targeting ability, improving bioavailability and prolonging their half-life. Moreover, they can also exert a direct therapeutic effect. This review begins by introducing the general situation of acute pancreatitis (AP). It then discusses the pathogenesis of acute pancreatitis (AP) and the current status of treatment. Finally, it considers the literature related to the treatment of acute pancreatitis (AP) by nanomaterials. The objective of this study is to provide a comprehensive review of the existing literature on the use of nanomaterials in the treatment of acute pancreatitis (AP). In particular, the changes in inflammatory markers and therapeutic outcomes following the administration of nanomaterials are examined. This is done with the intention of offering insights that can inform subsequent research and facilitate the clinical application of nanomaterials in the management of acute pancreatitis (AP).

*Correspondence: Wencheng Wu wuwencheng@uestc.edu.cn Haojie Huang huanghaojie@smmu.edu.cn Zhendong Jin zhendong Jin@163.com

Full list of author information is available at the end of the article



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by-nc-nd/4.0/.



Introduction

Acute pancreatitis (AP) is defined as a localized inflammatory response to the activation of pancreatic enzymes by a variety of etiologic factors and their action on pancreatic tissue, with or without functional changes in other organs [1–3]. The global incidence of AP ranges from 13 to 45 cases per 100,000 people, with a predominance of adults, and shows an increasing trend year by year [4–7]. Common causes are cholelithiasis, alcohol and hyperlipidemia [8, 9]. Additionally, less frequent causes encompass medication side effects, endoscopic retrograde cholangiopancreatography (ERCP), genetic predispositions and traumatic incidents [10]. The primary clinical manifestations are abdominal discomfort, nausea and vomiting and fever. In severe cases, hypotension and shock may also occur [3, 11], pathological alterations encompass interstitial inflammation as well as necrosis of the pancreatic tissue at the periphery [1]. The clinical severity of AP can be categorized into mild, moderate or severe. Mild AP is characterized by a self-limiting nature and typically not require specific intervention [8, 12], it is estimated that approximately 20% of patients with mild AP will progress to moderate or severe AP, with the development of localized complications such as pancreatic pseudocysts and encapsulated necrosis. In such patients, persistent organ failure and mortality rates of 10–20% have been observed [13, 14]. The current range of treatment options for AP is inadequate to meet the clinical needs of patients, underscoring the urgent need for the development of novel therapeutic modalities.

As we all know, nanotechnology is a scientific and technological field that employs the manipulation of individual atoms and molecules to fabricate and alter materials. It is concerned with the properties and applications of materials with structural dimensions in the range of 1–100 nm [15]. In recent years, nanotechnology has continued to evolve, extending from the domain of physical chemistry to the realm of biomedicine and achieving a significant degree of implementation. It has become a pivotal tool in the prevention, diagnosis and treatment of a multitude of illnesses [16-21], and plays a role in cell tracking and visualizing specific disease regions [22]. Nanomaterials can be altered in size, structure and other physical properties through processes including such as physical, chemical and biosynthesis methods [21, 22], giving it larger specific surface area, the ability to load drugs and a surface that can be used for other structural modifications [19, 23, 24]. The advent of rapidly evolving nanomaterials has opened up new avenues for therapeutic strategies in the treatment of AP. These strategies primarily revolve around the use of nanocarriers and nanodrugs, which have the potential to enhance drug safety, facilitate tissue penetration and enable targeted delivery [25-29]. At present, an increasing number of researchers have engaged in the treatment research of AP with nanomaterials. This review begins by examining the pathogenesis and current status of AP treatment. It subsequently searches and summarizes relevant literature on nanomaterials for treating AP. It then analyzes changes in inflammatory indexes and therapeutic effects after the application of nanomaterials. Finally, it analyzes the opportunities and challenges of nanomaterials for treating AP in the future.

Current research on the pathogenesis and treatment of AP

Mechanisms of AP

The etiology of AP is intricate, as is the pathogenesis. The specific mechanism remains unclear, but there is now a consensus among researchers and scholars that trypsin plays a role in digesting pancreatic tissue [30, 31]. Abnormal activation of trypsinogen is recognized as a significant pathogenetic mechanism in the development of AP. In normal physiological conditions, the rough endoplasmic reticulum of acinar cells synthesizes zymogen granules and lysosomes, which are then processed by the Golgi apparatus and stored in separate secretory vesicles. The apical region of the follicular cell is capable of cytosolizing zymogen granules, thereby maintaining a low concentration of these granules within the cell.

Additionally, the presence of intracellular trypsin inhibitors prevents the premature activation of intracellular trypsinogen, which, if activated, could potentially lead to the development of AP [32, 33]. The synthesis of intracellular lysosomal and digestive enzymes is increased when cells are stimulated by substances such as alcohol and bile acids. Conversely, these toxic stimuli inhibit zymogen granule cytosolization, which results in an increase in intracellular zymogen granule and lysosome content [33–35]. The codomainization of zymogen granules and lysosomes occurs, and histone B within lysosomes activates trypsinogen under acidic conditions, resulting in its conversion to trypsin. Subsequently, lysosomal membrane rupture releases histone B and trypsin into the cytoplasm, where histone B then promotes RIP3. The formation of the RIP1 necroptosis complex through the mixed-series protein kinase structural domain (MLKL) signaling pathway results in the phosphorylation and oligomerization of MLKL protein molecules, leading to the rupture of the follicular cell membrane and necrosis, which ultimately causes AP [36–40]. In recent years, the calcium overload theory and the mitochondrial dysfunction theory, with pancreatic vesicles as the primary focus of investigation, have garnered significant attention within the scientific community.

Pathologic calcium signaling in pancreatic alveolar cells is mainly characterized by persistent, non-oscillatory, peak-to-plateau calcium signaling throughout the cell, also known as alveolar cell "calcium overload" [41, 42]. In typical physiological circumstances, cholecystokinin prompts the release of calcium from the endoplasmic reticulum through the inositol triphosphate (InsP3R) signaling pathway. The influx of calcium subsequently stimulates the generation of adenosine triphosphate (ATP) by mitochondria and the activation of apical secretory zymogen granules within follicular cells [43, 44]. Intracellular calcium overload resulting from alcohol, bile acids, and cholecystokinin stimuli represents a pivotal mechanism in the pathogenesis of AP. Calcium endocytosis reduces Ca²⁺ levels within the endoplasmic reticulum, thereby activating ORAI1 to facilitate Ca²⁺ influx from the extracellular milieu. Sustained calcium endocytosis culminates in intracellular calcium overload, which in turn induces mitochondrial membrane damage and mitochondrial dysfunction [45-47]. Normal mitochondrial membrane potential is essential for the maintenance of mitochondrial oxidative phosphorylation and ATP synthesis, in which the mitochondrial membrane permeability transition pore (MPTP) plays a pivotal role. The persistent influx of calcium ions into the cell results in an intracellular calcium overload, which in turn causes damage to the mitochondrial membrane and the opening of the MPTP. This alteration in the mitochondrial membrane potential subsequently leads to a reduction in



Fig. 1 Explanation picture of the AP Mechanism using the calcium overload theory and the mitochondrial disorder theory

ATP production. The reduction in ATP levels is insufficient to support the ATP-dependent transport of Ca²⁺ by the Sarcoendoplasmic Reticulum Calcium ATPase (SER-CAs) and the Plasma membrane Ca^{2+} -ATPase (PMCAs). The transfer of calcium from the cell to the extracellular space results in a reduction in intracellular calcium transport, leading to intracellular persistent calcium overload. This ultimately causes the massive activation of digestive zymogens, which in turn leads to pancreatic auto-digestion [48-51]. Concurrently, mitochondrial dysfunction results in impaired cellular autophagy and the formation of autophagic vesicles with a distinct vacuolar structure. These vesicles produce reactive oxygen species and cytokines, exacerbating additional cellular damage (Fig. 1) [52]. Damaged cells produce tissue factor, heat shock proteins and other harmful factors, which activate NF-κB, MAPK, signal transducers and phosphatidylinositol-3-kinase inflammatory signaling pathways. This results in an elevated release of TNF- α , IL-6 and other similar inflammatory and chemokine factors, which then trigger the inflammation cascade that leads to the progression of a local inflammatory response into a systemic inflammatory response [53, 54]. Furthermore, the presence of mitochondria-associated endoplasmic reticulum membranes (MAM) induces endoplasmic reticulum stress, lysosomal damage, and impaired protein folding and post-translational modification within the endoplasmic reticulum. This, in turn, impairs cellular digestion and defense mechanisms, leading to cytoplasmic protein degradation and cellular necrosis [31, 55–58].

Novel therapeutics and challenges

In recent years, researchers have identified a number of drugs that can be used in the treatment of AP to address the pathogenic mechanisms involved. For example, Auxora is a novel selective inhibitor of the Ca²⁺ channel ORAI1, which prevents Ca²⁺ from entering the alveolar cells [59]. In a rat AP model, intravenous infusion of Auxora resulted in a notable reduction in the severity of AP-associated features, including vacuolization of the adenohypophysis, activation of necrotic cell signaling, pancreatic edema and adenohypophysis death, decreased myeloperoxidase activity and inflammatory cytokine expression in pancreas tissues. FMLF peptide was observed to induce an oxidative burst in human neutrophils and stimulate cytokine production in human PBMCs and rodent PAMs, suggesting that ORAI1/ STIM1 channels play a role in the inflammation [60]. The

drug has successfully completed Phase II clinical trials in the United States. Compared to standard care, patients diagnosed with moderate AP who received Auxora exhibited a notable improvement in their symptoms, a reduction in the incidence of persistent systemic inflammatory response syndrome (SIRS), a lower rate of hospitalizations and a minimal prevalence of minor chromaturia adverse effects. These findings suggest that Auxora has the potential to provide personalized treatment pathways for patients presenting with AP-SIRS [61]. Moreover, additional researchers have substantiated that trehalose can be employed in the management of AP. Trehalose is a disaccharide that has been demonstrated to stimulate autophagic flux and autophagic vesicle clearance [62], which can inhibit the development of pancreatic necrosis, aberrant trypsinogen activation and inflammatory signaling pathways by enhancing the autophagic elimination of damaged mitochondria [31]. Aringenin is a naturally occurring dihydroflavonoid, derived predominantly from citrus fruits such as grapefruit and lemon. Some researchers have identified that naringenin can be modulated by PI3K/Akt [63], TLR4/NF-κB [64], MAPK/ NF-KB [65] and other pathways to exert anti-inflammatory effects, reduce blood amylase (AMS) and lipase (LPS) levels and diminish the expression of inflammatory cytokines and the number of white blood cells (WBCs). This has the potential to effectively mitigate the damage of pancreatic tissue [66, 67]. Furthermore, emodin, ascorbic acid, α -tocopherol, β -carotene, baicalein, and other substances have been demonstrated to be effective in the alleviation of clinical symptoms and the mitigation of the condition and complications associated with AP [68-73].

While these drugs can be utilized for the management of AP, they are constrained by several factors, including poor chemical stability, low bioavailability, a short half-life and limited targeting capabilities [74–77]. Consequently, enhancing the stability, bioavailability and reducing the adverse effects of AP drugs have emerged as a prominent research area in recent years.

Advantage analysis of nanomaterials for AP treatment

Conventional drugs used to treat AP have been demonstrated to possess a limited capacity to selectively target the pancreas and effectively penetrate the bloodpancreas barrier (BPB) [78, 79], and most of these trypsin inhibitors are peptides with short half-lives [80]. It is frequently unfeasible to attain an optimal drug concentration within the pancreas, and the suboptimal tissue microenvironment of AP patients can influence both drug release and pharmacological activity [27], limiting the effectiveness of AP treatment [26, 27]. Nanomaterials offer several advantages over conventional drug formulations for the treatment of AP. Nanomaterials possess a structural size of 1–100 nm, which is characterized by a minor particle size, enormous specific surface area and good solubility. When vascular permeability is increased at the inflammatory sites of AP patients, nanomaterials with a small particle size can penetrate through the endothelial gap and selectively target pancreatic inflammatory tissues by traversing the blood pancreatic barrier, the cellular biofilm barrier and other body barriers [81, 82]. The large specific surface area endows it with the capacity to serve as an efficacious drug carrier. The utilization of nanoscale carriers has been demonstrated to augment the hydrodynamic radius of the drug, diminish its glomerular filtration rate and extend the drug's half-life [83, 84]. Nanomaterials can also be designed to possess the requisite functional characteristics to serve as nanomedicines for the treatment of AP. They can be modified in a variety of ways to enhance their targeting properties [85]. For example, nanomedicines can leverage alterations in the inflammatory microenvironment (e.g., pH, ROS and trypsin) or leaky vessel extravasation and inflammatory cell-mediated chelation (ELVIS effect)to achieve targeted release [86], thereby facilitating drug uptake, reducing the impact of the first-pass effect of the drug, improving bioavailability while decreasing toxicity and enhancing biosafety [85, 87–89]. The use of nanomaterials has been demonstrated to prolong the release of drugs by wrapping biofilms or fibrous proteins around the surface of the drug. This results in longer-acting blood levels and a reduction in the number of required administrations [20, 85, 90, 91].

Application of nanomaterials in the treatment of AP

Nanocarriers

Liposomal nanocarriers

Typical liposomes possess a bilayer membrane structure analogous to that observed in cell membranes. They are sealed spheres comprising a hydrophilic polar head and a hydrophobic nonpolar tail, which can be modified to facilitate the delivery of lipid-soluble, water-soluble and amphoteric drugs. They are extensively utilized as nanomedicine carriers [92, 93]. Kaempferol (KA) is a natural antioxidant in Traditional Chinese Medicine (TCM). A substantial body of evidence indicates that KA inhibits mitochondrial damage and exerts anti-inflammatory effects through the LPS-TLR4-NF-κB and IRE1-JNK-CHOP pathways. Consequently, it has been widely recognized as a potential treatment for inflammation [94–96]. However, KA is poorly soluble in water and fat solubility and is easily oxidized and less stable, which limits its therapeutic efficacy [97]. Some researchers have designed a novel thicketals (TK)-modified based on DSPE-PEG2000 liposomal nanocarriers (DTM@KA NPs), which enhances the bioavailability of KA while



Fig. 2 DTP@KA NPs improved mitochondrial fission by activating TOM20-STAT6 signaling pathway. **A**. Protein expressions of STAT6, TOM20 and DRP1 in pancreas lysate. **B**. Quantitative analysis of STAT6, TOM20 and DRP1 protein expressions. **C**. Distributions and expressions of STAT6 and TOM20 in pancreas tissues. Scar bar = 20 μ m. **D**. Co-localization and expressions of TOM20 and STAT6 with IF staining in pancreas tissues. Scar bar = 50 μ m. **E**. IHC staining of DRP1 and Bax in pancreas tissues. Scar bar = 20 μ m. Data represent the mean ± SEM of at least three independent experiments; *n* = 5–8/group. Significance: *p* < 0.05 and *p* < 0.01 vs. the control group; *p* < 0.05, *p* < 0.01 vs. the SAP model group. Reprinted from [98] with permission. Copyright © The Author(s) 2024

simultaneously reducing the incidence of toxic side effects. Results from the AP mouse model demonstrated that DTM@KA NPs exhibited favorable biological safety with minimal side effects in vitro and in vivo. Subsequent studies revealed that DTM@KA NPs facilitated STAT6mediated mitochondrial precursor protein translocation by interacting with TOM20, and further promoted DRP1-dependent fission and Pink1/Parkin-regulated mitochondrial autophagy. Additionally, they enhanced lysosomal degradation, which facilitated the removal of damaged mitochondria from the PAC, thereby reducing inflammation and apoptosis (Fig. 2) [98].

Polymer nanocarriers

Polymeric nanomaterials encompass a range of synthetic and natural polymers. Synthetic polymers include PLGA, PEI, PEG and others. Natural polymers, such as chitosan (CTS) and sericin protein (SF), offer distinct advantages, including good biocompatibility, biodegradability, low cost and low toxicity [99, 100]. Festerone (FST) is a naturally occurring flavonol that has been the subject of several studies, which have identified a number of potential beneficial effects, including anti-inflammatory, antioxidant, anti-angiogenic and hypolipidemic properties [101–103], nevertheless, the material displays low water solubility and high permeability defects [104]. In accordance with the aforementioned methodology, a research team prepared FST-loaded lipid polymer hybrid nanoparticles by conjoined ultrasonication and double emulsion (w/o/w) techniques. These nanoparticles exhibited a high level of loading and mucosal adhesion strength, enabling sustained in vitro release of FST [105]. Oral pretreatment with LPHNP containing FST provided protection for rats from L-arginine-induced SAP and multiple organ damage in the in vivo study. Additionally, this treatment resulted in a reduction in amylase and lipase levels compared with FST, LPHNP alone and the untreated group. Further biochemical analysis demonstrated a reduction in pro-inflammatory factors, including NLRP3, IL-1 β , TNF- α and IL-6, were diminished, and a notable decrease in the observed inflammation in pancreatic tissue [105].

Biomolecule nanocarriers

In comparison to lipid and polymer carriers, biomolecule carriers demonstrate notable sensitivity to elevated amylase, protease and lipase levels in pancreatitis lesions. These compounds demonstrate greater specificity and offer distinct advantages in terms of half-life, stability, safety and manufacturing feasibility [106]. A research team has successfully developed a primary three-dimensional (3D) structure of a bilirubin-like DNA double helix by co-precipitation using silk fibroin as a carrier. It was collapsed into a nanoparticle (BRSNP), which could achieve targeted pancreatic tissue. In the presence of excess pancreatic enzymes (trypsin) at the AP site, the BRSNP was observed to release bilirubin, which demonstrated the ability to directly inhibit the production of cellular mitochondrial ROS. Additionally, bilirubin was observed to decrease malondialdehyde (MDA) levels and increase SOD levels in rats. The inhibitory effect of bilirubin on cellular mitochondrial ROS production, reduction in MDA levels and increase in SOD levels in rats has been demonstrated. Additionally, it activates the NRF2 pathway, increases HO-1, and thus inhibits the pro-inflammatory NF-кВ signaling pathway. BRSNPs can target pancreatic tissues by reducing oxidative stress and pro-inflammatory cytokine expression, and impairing macrophage and neutrophil recruitment. The efficacy of BRSNPs in treating AP has been substantiated by evidence indicating that they can mitigate oxidative stress, diminish the expression of pro-inflammatory cytokines, and impede macrophage and neutrophil recruitment [20].

Inorganic nanocarriers

In comparison to lipid and polymer carriers, inorganic nanoparticles typically exhibit smaller particle sizes, narrower size distributions and surface chemistry well suited for ligand coupling [107]. 1,2-Bis (2-aminophenoxy) ethane-N, N, N, N, N'-tetra acetic acid (BAPTA-AM) is a hydrophobic membrane-permeable calcium chelator that can directly bind to intracellular Ca²⁺, thereby eliminating cytoplasmic calcium overload at the root and blocking abnormal calcium-mediated sustained activation of trypsinogen at the source is interrupted by this mechanism, preventing the cascade of adenohypophysis cell death. However, the compound displays poor water solubility and a short half-life limit its clinical transfer [108]. A research team devised and synthesized an organosilicon precursor comprising a trypsin-cleavable arginine-based amide bond. It was incorporated into the framework of mesoporous silica to create trypsinresponsive organo-bridged MSNs (Arg-MSNs) for the effective encapsulation of a membrane-permeable calcium chelator, BAPTA-AM, to produce Arg-MSNs@BA. This approach was devised with the objective of facilitating the controlled release of drugs and rapidly eliminating intracellular Ca²⁺ overload during the initial stages of AP [109–111]. Arg-MSNs produced via a sol-gel process, exhibiting a small size, uniform size distribution, positive surface charge and exemplary storage stability properties, were then subjected to a series of modifications to create SL@M@Arg-MSNs@BA, which demonstrated immune evasion, inflammatory endothelial adhesion, transendothelial migration and precise targeting of injured cells [112, 113]. Upon arrival at the designated site, the core Arg-MSNs@BA in the AP mouse model responds to premature and massive activation of trypsin by rapid and direct biodegradation of its backbone, resulting in flexible payload release. The released BA will effectively chelate the overloaded Ca2+, thereby restoring the cellular redox state by blocking the I κ B α /NF- κ B/TNF- α / IL-6 and CAMK-II/p-RIP3/p pathways. The MLKL/ caspase-8,9 signaling pathways reduce the secretion of pro-inflammatory cytokines, block the inflammatory cascade and inhibit cell necrosis (Fig. 3). Additionally, they significantly reduce the adiposity of adipose tissue and lipase and amylase levels and restore pancreatic function [114]. Furthermore, the Arg-MSN backbone structure inhibits trypsin self-digestion while preventing tissue damage. This ultimately results in the inhibition and modulation of downstream local and systemic inflammatory responses, which disrupts the destructive cycle amplified by inflammation. This protective mechanism safeguards peripancreatic tissues and effectively reduces the risk of distal tissue damage [53].

Cell membrane nanocarriers

Cell membrane nanocarrier is a nanoparticle that has been coated with a partial or complete cell membrane. The surface of the nanocarrier mimics the properties of the source cells, carrying biological activity and homing effects. This enables the nanocarrier to replicate natural



Fig. 3 SL@M@Arg-MSNs@BA reduces recruitment of mononuclear macrophages and neutrophils, inhibits the release of neutrophil extracellular traps, and promotes M2-macrophage polarization. Western blot analysis of (A) p-lkBa, (B) NF- κ B, (C) TNF- α , and (D) IL-6 expression and their quantification of protein expression in the corresponding pancreas. n = 3. Data are shown as mean ± SEM. Reprinted from [114] with permission. Copyright © 2024 American Chemical Society

cellular interactions, effectively reducing the cytotoxicity and immunogenicity of nanoparticles. Additionally, the use of cell membrane nanocarriers improves the histocompatibility and bio targeting of nanoparticles. This makes them an effective drug carrier [115–117]. The aforementioned cells include red blood cells, macrophages, neutrophils, cancer cells, stem cells and others [118–120]. Pancreatitis is a severe inflammatory disease marked by the infiltration of the lesion by a substantial number of inflammatory cells, including neutrophils and macrophages [121]. In light of the aforementioned considerations, acid-responsive hollow mesoporous Prussian blue nanoparticles (HMPB-NPs) were developed for the co-delivery of the membrane-permeable calcium chelator, BAPTA-AM (BA) and the trypsin activity inhibitor, gabexate mesylate (GA). These nanoparticles were encapsulated with neutrophil membranes and partially surface-modified with N, N-dimethyl-1,3-propanediamine. In an AP mouse model, the agent demonstrated effective recruitment, trans endothelial migration and precise targeting of vesicular cells at the inflamed endothelium. Additionally, it inhibited endoplasmic reticulum stress (IRE1/XBP1 and ATF4/CHOP). Axis restoration was observed in impaired autophagy (Beclin-1/P62/LC3 axes), with a concomitant effective restoration of the cellular redox state and reduction in the proportion of apoptotic cells. This resulted in the hindrance of alveolar cell autodigestion and restoration of pancreatic function [53].

In addition to cell membranes as nanocarriers, the "free-rider" strategy of drug-loaded inflammatory cells has become a hot research topic in recent years, mainly involving macrophages and neutrophils [122, 123]. The capacity of surviving inflammatory cells to respond to environmental changes is superior, enabling them to deliver therapeutic payloads with greater precision and dynamism. Furthermore, the heightened sensitivity of these cells to inflammatory signals increases the potential for targeted delivery of therapies to cell types. In addition, the large intracellular volume of live inflammatory cells allows them to accommodate more nanoparticles, resulting in enhanced loading capacity and drug-to-cell ratios. In addition, numerous drugs can be simultaneously introduced into inflammatory cells, promoting a combined therapeutic approach [124]. An investigator has developed a β-glucan-functionalized zinc-doxorubicin nanoparticle system (\beta Glus-ZnD-NPs). In mice with pancreatic ductal adenocarcinoma (PDAC), oral administration of βGlus-ZnD NPs resulted in positive targeting and transgression of microfold (M) cells, overcoming the intestinal epithelial barrier and undergoing phagocytosis by endogenous macrophages to produce \beta Glus-ZnD@ Mφ. As a hitchhiking cell carrier, βGlus-ZnD@Mφ traverses the intestinal lymphatic system and gets into the circulation, and eventually accumulates in tumor tissues due to the tumor homing and "stealth" characteristics given by endogenous M ϕ (Fig. 4). Concurrently, the M ϕ of the hitchhiking ßGlus-ZnD NPs are activated to produce MMP, which disrupt the proliferative matrix barrier of junctional fibers, promote the degradation of stromal fibrosis and differentiate towards an M-like phenotype. This process also promotes T-cell infiltration and ultimately remodels the TME from immune-suppressing to immune-primary, which induces apoptosis of the tumor cells and enhances the efficacy of PDAC therapy [125]. The "free rider" strategy of inflammatory cells carrying drugs provides a new approach for the treatment of AP. Figure 5 illustrates the preparation process of the different nanocarriers described above or the mechanism of action of the therapeutic APs. Table 1 shows the research progress of nanocarriers loaded with different drugs for the treatment of AP.

Liposomal, polymer, biomolecule and cell membrane nanocarriers are organic nanomaterials formed by covalent or non-covalent assembly of organic molecules [126]. They can be easily functionalized to precisely control their chemical composition, shape, size and surface properties, and have good biocompatibility, making them highly adaptable to various biomedical applications [127]. However, their mechanical strength and stability are low, and such limitations may be imposed by the requirement for high structural integrity and thermal stability in specific applications [128]. In addition, batch-to-batch variations in synthesis can affect reproducibility and scalability [129]. In comparison to organic nanomaterials, inorganic nanomaterials demonstrate superior chemical and structure stability, in addition to distinctive physicochemical attributes, including electrical, magnetic and catalytic properties [130]. However, the toxicity and low biocompatibility can trigger negative immune responses that prompt immune response and permanent toxicity, giving great challenges for clinical translation [131]. In addition, inorganic nanomaterials typically have limited biodegradability, leading to accumulation and presenting large venture over long periods of time [132].

Nanomedicine

Nanodrug particle

In addition to the loading or encapsulation of therapeutic AP drugs with various nanocarriers, drugs can be congregated into nanodrug particles through the influence of external forces. For example, selenium has been demonstrated to possess anti-inflammatory and antioxidant properties [133]. Selenium is directly converted into nano-selenium, which is not only a simple and efficient process that reduces drug toxicity, but also improves the shortcomings of poor drug solubility and bioavailability. Furthermore, it regulates the sustained release of the drug and guarantees its therapeutic efficacy [134, 135]. Similarly, a research team applied yttrium oxide nanoparticles (NY) to a mouse AP model and found that NY could potentially play a role in restoring mitochondrial and ER homeostasis through NRF2/NFkB pathway. This resulted in the effective reduction of plasma amylase and lipase levels, as well as a notable attenuation of mitochondrial stress and inflammatory markers. Additionally, inflammatory cell recruitment around damaged pancreatic follicular cells was inhibited, which contributed to a favorable therapeutic AP effect [136]. In addition to inorganic substances, biological macromolecules can also be directly assembled into drug nanoparticles. Tetrahedral framework nucleic acid nanoparticles (TFNAs) have been designed through the self-assembly of four single-stranded DNA (ssDNA) molecules, resulting in a three-dimensional cage-like structure [137–139]. A number of studies have demonstrated that TFNAs possess robust anti-inflammatory and anti-apoptotic properties [140-142]. A research team was able to achieve a notable reduction in amylase, lipase, aspartate aminotransferase (AST), alanine aminotransferase (ALT), creatinine (CREA) and blood urea nitrogen (BUN) levels in mice with AP through the administration of TFNAs. This demonstrates that TFNAs can effectively inhibit inflammation and prevent pathological cell death [143].



Fig. 4 In vivo biodistribution of ZnD/ β Glus-ZnD NPs. **a**) Ex vivo IVIS images of fluorescence signals from DOX that accumulated in isolated major organs from test mice that had been treated using various modalities and their corresponding average radiances. **b**) Time-dependent fluorescence signals of DOX that accumulated in isolated major organs from test mice that had been orally treated with β Glus-ZnD NPs and their corresponding average radiances. **c**) Time course of DOX concentrations in plasma following treatment with i.v. DOX or oral β Glus-ZnD NPs. **d**) Time-dependent accumulation of DOX concentration in tumorous pancreatic tissues in test mice that had been treated with i.v. DOX or oral β Glus-ZnD NPs. **d**) Results of DOX accumulation in major organ tissues at 1 h (6 h) following i.v. DOX (Oral β Glus-ZnD NPs) administration. n.s.: not significant (*P* > 0.05). *: statistically significant (*P* < 0.05). Reprinted from [125] with permission. Copyright © 2023 Wiley-VCH GmbH



Fig. 5 The preparation process of the different nanocarriers described above or the mechanism of action of the therapeutic APs. (A) Schematic of study on DTM@KA NPs preparation and protective function as well as possible mechanism about mitochondrial function and oxidative stress regulated by TOM20-STAT6-DRP1-mitophagy signaling in experimental SAP. Reprinted from [98] with permission. Copyright © The Author(s) 2024. (B) Graph of bilirubin loaded silk fibroin nanoparticles (BRSNPs) for the experimental acute pancreatitis (AP) application. Reprinted from [20] with permission. Copyright © 2020 Elsevier B.V. (C) Schematic Representation Depicting the Fabrication Process of Biomimetic HMPB NPs and Their Therapeutic Mechanism in a Mouse Model of AP Induced by Sodium Taurocholate Retrograde Infusion. Reprinted from [114] with permission. Copyright © 2024 American Chemical Society. (D) Treatment mechanism diagram of the formulations 2 N@M@HMPB@BA#Ga. Reprinted from [53] with permission. Copyright © 2024 American Chemical Society

Tab	le 1	The researc	h progress of	f nanocarriers loa	aded wit	ch different c	lrugs for	the treatment of A	٩P

Nano carrier	Drug	Characteristics	Ref.
Liposome	Kaempferol (KA)	Liposomes exerts profound impacts on damaging intracellular redox homeostasis by reducing GSH depletion and activating Nrf2, which synergizes with KA to reinforce the inhibition of inadequate fission, excessive mitochondrial fusion and impaired mitophagy.	[98]
Polymer	Fisetin (FST)	Polymer owns a high level of load capacity and mucosal adhesion strength, and it can achieve sustained in vitro release of FST.	[105]
Silk fibrin	Bilirubin	BRSNPs can be selectively delivered to inflammatory lesions in the pancreas and release bilirubin in an enzyme-responsive manner, reducing oxidative stress, decreasing the expression of pro-inflammatory cytokines, and impairing the recruitment of macrophages and neutrophils.	[20]
Mesoporous silica nanoparticles(MSN)	BAPTA-AM	MSN has a high loading content and surface modifications confer MSN inflammatory recruitment and precise PAC-targeting ability; after skeletal biodegradation, BAPTA-AM is released on-demand in injured PACs, resulting in elimination of intracellular calcium overload.	[114]
Macrophage membrane	Emodin(ED)	Macrophage membranes not only provide immune evasion, but also show a tendency to target inflam- matory tissues with slow-release properties.	[121]
Neutrophil membrane	BAPTA-AM And Ga	The formulation exhibited efficient recruitment at the inflammatory endothelium, trans-endothelial migration, and precise acinar cell targeting, resulting in rapid pancreatic localization and higher accumulation.	[53]

Nanoenzyme

Natural enzymes are distinguished by their catalytic activity and substrate diversity. However, they remain susceptible to limitations, including high cost, poor thermal stability and a low recovery rate [144]. In comparison to natural enzymes, nanoenzymes exhibit tunable catalytic activity, high stability and biocompatibility, rendering them more suitable for bioassay and therapy [145]. Metal-organic frameworks (MOFs) represent a relatively novel class of porous solid materials. The diversity of metal nodes, connecting columns, and a vast range of coordination interactions in all potential orientations render MOFs the optimal alternative to natural enzymes. These have garnered significant research interest in the domain of catalysis due to the clarity of coordination networks, mesoporous structures and tunable porosities [146, 147]. A team of researchers has designed an optimized copper-based metal-organic framework nanoenzyme (Cu MOF) with special antioxidant activity. The activity of Cu MOF is at its greatest at pH 7.4 and 35 °C, and it displays considerable potential under physiological conditions. The majority of catalytic reactions associated with other nanoenzymes occur in acidic environments (pH < 5.0), which can lead to the denaturation of natural enzymes [148]. The newly developed compound displays remarkable excellent catalase-like activity and hydroxyl-removal ability. In addition, it scavenges ROS directly through its intrinsic enzyme. Furthermore, the Cu MOF displays antioxidant activity in AP, and also activates PINK1/PARK2-mediated mitochondrial autophagy (Fig. 6). This indicates that Cu MOF plays a role in mitochondrial autophagy-mediated inflammation and necrotic apoptosis in AP, in order to maintain mitochondrial homeostasis and attenuate inflammation [149]. Additionally, research indicates that the utilization of copper nanoparticles may potentially elevate the likelihood of developing neurological disorders [150].

Nevertheless, there is currently no consensus among medical professionals and toxicologists regarding the establishment of standardized methodologies for toxicological testing and analysis [132]. It is imperative to achieve a balance between efficacy and safety.

Nanomembrane

In complex pancreatic resections, the leakage of tissue fluid from medical trauma, accidents, and surgical treatments can result in significant complications, including acute necrotizing pancreatitis, hemorrhage, infection and mortality [151], the conventional techniques for wound closure, including suturing, ligating and cauterizing, are susceptible to stress concentrations that can result in secondary tissue trauma, capillary fragility and failure to seal pancreatic leaks. The use of traditional anti-leakage materials, such as fibrin glue and polyglycolic acid, has not demonstrated efficacy in enhancing tissue regeneration, wound healing, flexibility and adhesion at the pancreatic resection margin [152]. In response to these issues, a research team employed electrostatic spinning technology to develop a novel bilayer (AG-TH/PG-MC) multifunctional nanofiber membrane, wherein the inner layer consisted of a combination of sodium alginate (A), gelatin (G) and thrombin (TH), while the outer layer comprised a combination of polycaprolactone (P), gelatin (G) and mitomycin-c (MC). The elevated gelatin and alginate concentration within the internal layer diminishes fiber diameter and water contact angle. Furthermore, cross-linking enhances the membrane's hydrophilic properties, rendering it highly biodegradable and firmly adhering to tissue surfaces [153, 154]. The outer layer of MC has been demonstrated to possess antibiotic properties that reduce tissue adhesion by inhibiting fibrosis and vascular distribution. This effect can be observed within a few weeks. Furthermore, MC has been shown to inhibit fibroblast proliferation by downregulating $TGF\beta$



Fig. 6 Cu MOF regulated PINK/PARK-mediated mitophagy to control ROS-induced inflammation and necroptosis. **a**) Schematic diagram of the mitophagy, anti-inflammatory, and antioxidant mechanisms of Cu MOF. **b**) Heatmap of the differentially expressed genes. **c**) Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway terms. **d**) Significantly enriched Gene Ontology (GO) terms. **e**, **i**, **j**) Immunoblot analysis and quantification of P62 and LC3 in pancreatic tissues (n=3). **f**, **k**) Immunoblot analysis and quantification of PINK1 and PARK2 in pancreatic tissues (n=3). **g**, **l**) TUNEL positivity was revealed by immunohistochemical staining of pancreatic tissues and quantificative analysis (n=3). Scale bar: 100 µm. **h**, **m**, **n**) Immunoblot analysis and quantificate tissues (n=3). The data are represented as mean values, and error bars indicate SEM (mean ±SEM). n represents the number of samples for each group (p<0.05; p<0.01; p<0.001; p<0.0001). Reprinted from [149] with permission. Copyright © 2024 Wiley-VCH GmbH

receptor expression and fibroblast apoptosis [155, 156]. The results of external biocompatibility and hemostasis tests demonstrated that the bilayer exhibited enhanced cell proliferation and effective hemostatic properties. Furthermore, in vivo studies and computerized molecular simulations revealed that the bilayer effectively covered the wound site, prevented suture breakage and leakage, inhibited bleeding and reduced postoperative tissue adhesion (Fig. 7). These findings suggest that the bilayer has a promising therapeutic potential in preventing pancreatic leakage in the postoperative period [157]. Figure 8 illustrates the preparation process of the various nanomedicines mentioned above or the mechanism of action of therapeutic APs. Table 2 shows different types of nanomaterials used for the treatment of AP.

Summary and outlook

AP is a severe, life-threatening inflammatory disease with an unclear pathogenesis. It is primarily characterized by inflammatory infiltration and oxidative stress. Previous reports on the treatment of AP have focused on the suppression of the inflammatory response and the prevention of oxidation of pancreatic tissue. The traditional therapeutic AP drugs have several limitations, including poor drug stability, low bioavailability and a short halflife. To date, there has been limited utilization of these techniques in clinical settings. The ongoing advancement of nanotechnology has resulted in the development of a plethora of novel multifunctional nanomaterials, which have been employed in a multitude of clinical studies pertaining to AP. These developments have yielded novel insights and approaches for the treatment of AP. Conventionally synthesized nanomaterials have several disadvantages, including the need for expensive equipment, the generation of toxic chemicals and the strict control of reaction conditions. In contrast, the production of bio-nanomaterials based on derivatives from natural organisms, microorganisms, microalgae, enzymes and plant extracts is biologically safer and represents a safe, efficient, environmental-friendly and favorable approach of obtaining nanomaterials. This approach has attracted considerable attention and interest within the scientific community. It is important to acknowledge that nanomaterials employed in AP therapy also possess intrinsic limitations. Nanomaterials for the treatment of pancreatitis have many challenges and opportunities on the way. First, despite the development of numerous nanomaterials for the treatment of AP, the majority have only been confirmed in rudimentary models, lack pathological and statistical data and have not been subjected to extensive, multicenter investigations. Second, although the high potential of nanomaterials, their long-term safety and toxicity in vivo need to be thoroughly investigated. Nanomaterials may elicit an immune response or accumulate in the body with potential toxic effects on healthy tissues. The future may see the advent of therapeutic systems based on combinations of multiple nanomaterials, with a particular focus on the in vivo distribution, metabolic pathways and potential toxic effects of novel nanomaterials. This will ensure safety in clinical applications and improve therapeutic efficacy while reducing the risk of toxicity by decreasing the number of single nanomaterials used. Third, it is necessary to establish a unified scientific evaluation system, improve multiple animal models, evaluation indices and test technology, and vigorously develop nanotoxicology to promote the clinical translation and safe application of nanomaterials.

With the continuous development of technology, artificial intelligence (AI), big data analytics, 3D printing technology, single-cell sequencing, spatial transcriptomics and other fields continue to emerge, nanotechnology can be combined with these fields to promote technological innovation. AI technologies, particularly machine learning and deep learning algorithms, have the capacity to process and analyze vast quantities of biomedical data in order to identify pivotal parameters in drug delivery. AI is capable of constructing predictive models based on known drug-target interaction data, with the objective of predicting new drug targets and assisting in the optimization of the design of nanocarriers. This is done with the intention of more efficiently incorporating specific biomarkers, with the ultimate goal of improving drug delivery and targeting in diseased tissues. The application of big data analytics can facilitate the identification of the behavioral patterns exhibited by nanomaterials in diverse biological environments. Furthermore, it can assist in the screening of materials with optimal biodegradability properties and in the development of kinetic simulations, which can be employed to predict biodegradability and to optimize drug release under a range of conditions. By means of big data analysis of patient-specific biological information, in conjunction with the application of AI, it is possible to develop personalized drug delivery solutions that are tailored to the individual patient, thereby advancing the concept of precision medicine.

It is similarly anticipated that nanomaterials will eventually receive policy support from governments, including the signing of bilateral or multilateral cooperation agreements and the establishment of cross-border cooperation mechanisms. Additionally, the formation of an international nanotechnology standards committee to develop unified nanotechnology standards and norms is envisioned, as is the establishment of an international forum on the ethics and regulation of nanotechnology for regular discussion and updating of nanotechnology ethical and regulatory policies. Furthermore, the creation of an international fund for nanotechnology education is proposed to support cross-border nanotechnology



Fig. 7 (A) Samples extract preparation procedure and cell culture (B) Fluorescence microscope images of the L929 Fibroblast cell proliferation after 1, 3 and 7 days of culture. (C) F-actin are analysis of the L929 fibroblast cells after 7 days. (D) MTT cytotoxicity assay after 1, 3 and 7 days of culture. (E) Digital images of hemolytic activity of the composite membranes with PBS as a negative control and Triton x-100 as a positive control. (F) Digital images of the blood clotting formation on the samples. (G) Comparative blood hemolysis ratio analysis. (H) Blood clotting index (BCI) of the samples after 35 min. (I) Comparative bleeding time and (J) Blood loss analysis. Reprinted from [157] with permission. Copyright © 2024 Elsevier Ltd



Fig. 8 (A)Schematic diagram of TFNAs in treating AP in mice. Reprinted from [143]with permission. Copyright © 2022 American Chemical Society. (B) Schematic diagram of Cu MOF nanozyme synthesis and its application in treating AP. Reprinted from [149] with permission. Copyright © 2024 Wiley-VCH GmbH. (C) Schematic illustration of the preparation of electrospun AG-TH/PG-MC dual-layered membrane and application in the rat pancreas leakage model and its multifunctional ability. Reprinted from [157] with permission. Copyright © 2024 Elsevier Ltd

Phosphatidylinositol 3-kinase/protein serine-threonine

Types of Nanomaterials	Animal Model	Effects and Cellular Mechanisms DTM@KA NPs facilitated STAT6-regulated mitochondrial precursor proteins transport via interacting with TOM20 to further promote Drp1-dependent fission and Pink1/Parkin-regulated mitophagy with enhanced lysosomal degradation for removing damaged mitochondria in PAC.		
Liposomes (DTM@KA NPs)	NAT; mice			
Polymer (FST-LPHNPs)	L-arginine; mice	A reduction in NF- κ B activity, amylase and lipase levels, and pro-inflammatory factors (NLRP3, IL-1 β , TNF- α , and IL-6).	[105]	
Silk fibrin (BRSNPs) L-arginine; mice		Inhibition of cellular mitochondrial ROS production decreased malondialdehyde (MDA) levels and increased SOD levels in rats, as well as activation of the Nrf2 pathway and an increase in HO-1, which in turn inhibited the pro-inflammatory NF-κB signaling pathway.		
Mesoporous silica nanopar- ticles (SL@M@ArgMSNs@BA) socholate; mice		Inhibit cell necrosis by impeding the IкВа/NF-кВ/TNF-а/IL-6 and CAMK-II/p-RIP3/p-MLKL/cas- pase-8,9 signaling pathways.	[114]	
Macrophage membrane (MVs-UIO-ED)	Cerulein; mice	Inflammation-targeting ability, with both alpha-amylase and lipase levels showing the most significant reductions.	[120]	
Neutrophil membrane (2 N@M@HMPB@BA#Ga)	Sodium Tau- rocholate; mice	The formulation inhibited endoplasmic reticulum stress (IRE1/XBP1 and ATF4/CHOP axis) and restored impaired autophagy (Beclin-1/p62/LC3 axis), preserving dying acinar cells and restoring the cellular "health status".	[53]	
Nano yttria (NY)	Cerulein; mice	NY can attenuate experimental AP by restoration of mitochondrial and ER homeostasis through Nrf2/NFkB pathway modulation, reducing of endoplasmic reticulum (ER) stress markers (BIP, IRE1 and Ero1-Lq), and molecular chaperones (Hsp27 and Hsp70)	[136]	
Tetrahedral framework nucleic acids (TFNAs)	Sodium Tau- rocholate; mice	Decreased serum amylase, lipase, ALT, AST, CREA, and BUN levels, as well as decreased expression of the proteins Bax and caspase-3 associated with cell death and apoptosis.	[143]	
Nanozymes(Cu MOF)	Arginine; mice	Effectively scavenges reactive oxygen species and attenuates apoptosis and necrosis of aden hypophysial cells by activating PINK1/PARK2-mediated mitochondrial autophagy pathway.	[149]	
Nanomembrane (AG-TH/PG-MC)	Rat pancre- atic leakage	The high gelatin and alginate content of the inner layer reduces fiber diameter and water con- tact angle, making it highly biodegradable and firmly adhering to tissue surfaces. The MC in the outer layer has antibiotic properties that reduce tissue adhesion by inhibiting fibrosis and inhibit- ing vascular distribution and can inhibit fibroblast formation by down-regulating the expression of TGFβ receptor and fibroblast apoptosis.	[157]	

PI3K/AKT

TLR4

AMS

WBCs

LPS

kinase

Amylase

Lipase

Toll like receptor 4

White blood cells

Table 2 Different types of nanomaterials used for the treatment of AP

education and academic exchanges. It is recommended that the nanomaterials industry actively share their scientific research results, strengthen international exchanges and cooperation, and improve the international status and influence of the industry.

and influence	e of the industry.	BPB ROS	Blood-pancreas barrier Reactive oxygen species
Abbreviations		KA	Kaempterol Traditional Chinasa Madisina
	Acute papereatitis		Inactional Chinese Medicine
EDCD	Endoscopia retrogrado cholangionangroatography	IREI	inositoi-requiring enzyme i
DID1	Pacaptar interacting protain 1	JINK	C-Jun N-terminal kinase
	Receptor-interacting protein 1	CHOP	C/EBP nomologous protein
	Nived lineage kinase domain like	IK	I hioketals
IVIERE	Mixed III ledge Killase dollari I-like	DSPE-PEG2000	Hydrophilic 1,2-distearoyl-sn-glycero-3-
INSPOK ATD	Adenacina triphosphate receptor		phosphoethanolamine-N-[methoxy(polyethylene
AIP	Adenosine inprosphale		glycol)-2000
MPTP	Memorane permeability transition pore	STAT6	Signal transducer and activator of transcription 6
SERCAS	Sarcoendoplasmic Reticulum Calcium Al Pase	TOM20	Translocase of outer mitochondrial membrane 20
PMCAs	Plasma membrane Ca ²¹ -Al Pase	DRP1	Dynamin related protein 1
NF-KB	Nuclear factor-ĸB	PAC	Pancreatic acinar cell
MAPK	Mitogen-activated protein kinase	PLGA	Poly-lactic-co-glycolic acid
TNF-α	Tumor necrosis factor-alpha	PEI	Polyethyleneimine
IL-1	Interleukin-1	PEG	Polyethylene glycol
IL-6	Interleukin-6	CTS	Chitosan
MAM	Mitochondria-associated endoplasmic reticulum	SF	Sericin protein
	membranes	FST	Festerone
FMLF	N-Formylmethionine-leucyl-phenylalanine	LPHNP	Lipid polymeric Hybrid nanoparticle
PBMCs	Peripheral blood mononuclear cells	SAP	Severe acute pancreatitis
PASCs	Pancreatic stellate cells	NLRP3	NOD-like receptor protein 3
ORAI1	Calcium release-activated calcium modulator 1	BRSNP	Bilirubin encapsulated silk fibrin nanoparticles
STIM1	Stromal interaction molecule 1	MDA	Malondialdehvde
SIRS	Systemic inflammatory response syndrome	SOD	Superoxide dismutase

NRF2	Nuclear factor erythroid 2-related factor 2
HO-I	Heme oxygenase-1
BAPIA-AM	I,2-Bis(2-aminophenoxy) ethane-N, N, N, N, N'-tetraacetic acid
CAMK-II	Calmodulin-dependent protein kinase II
HMPB-NPs	Hollow mesoporous Prussian blue nanoparticles
GA	Gabexate mesylate
XBP1	X box binding protein 1
ATF4	Activating transcription factor 4
P62	Protein sequestosome 1
LC3	Light chain 3
βGlus-ZnD-NPs	β -glucan-functionalized zinc-doxorubicin nanoparticle system
PDAC	Pancreatic ductal adenocarcinoma
Μ	Microfold
NY	Nanoparticles
ER	Endoplasmic reticulum
TFNAs	Tetrahedral framework nucleic acid nanoparticles
ssDNA	Single-stranded DNA
AST	Aspartate aminotransferase
ALT	Alanine aminotransferase
CREA	Creatinine
BUN	Blood urea nitrogen
MOFs	Metal-organic frameworks
PINK1	PTEN-induced kinase 1
PARK2	Parkinson Disease Protein 2
AG-TH/PG-MC	Alginate gelatin- thrombin/ polycaprolactone gelatin- mitomycin-c
MC	Mitomycin-c
TGFβ	Transforming Growth Factor beta
Al	Artificial intelligence
MMP	Matrix metalloproteinases

Acknowledgements

Wei Du, Xinyue Wang and Yuyan Zhou contributed equally to this work. We thank the researchers who gave us permission to cite literature in the article.

Author contributions

W D: Writing-original draft and pictures. XY W and YY Z: Supervision; Validation. WC W, HJ H and ZD J: Writing – review & editing. All authors reviewed the manuscript.

Funding

This work was financially supported by the National Natural Science Foundation of China(Grants number 8217032680 and 82020108005).

Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

All authors in the paper agree to be published.

Competing interests

The authors declare no competing interests.

Author details

¹Department of Gastroenterology, Shanghai Institute of Pancreatic Diseases, National Key Laboratory of Immunity and Inflammation, Changhai Hospital, Naval Medical University, Shanghai 200433, China ²Central Laboratory, Department of Medical Ultrasound, Sichuan Provincial People's Hospital, Sichuan Academy of Medical Sciences, University of Electronic Science and Technology of China, Chengdu, Sichuan 610072, China

Received: 16 October 2024 / Accepted: 10 January 2025 Published online: 29 January 2025

References

- Mayerle J, Sendler M, Hegyi E, Beyer G, Lerch MM, Sahin-Toth M, Genetics. Cell Biology, and pathophysiology of pancreatitis. Gastroenterology. 2019;156(7):1951–68. e1.
- Tenner S, Vege SS, Sheth SG, Sauer B, Yang A, Conwell DL, et al. American College of Gastroenterology Guidelines: management of Acute Pancreatitis. Am J Gastroenterol. 2024;119(3):419–37.
- Boxhoorn L, Voermans RP, Bouwense SA, Bruno MJ, Verdonk RC, Boermeester MA, et al. Acute pancreatitis. Lancet. 2020;396(10252):726–34.
- Petrov MS, Yadav D. Global epidemiology and holistic prevention of pancreatitis. Nat Rev Gastroenterol Hepatol. 2019;16(3):175–84.
- Yadav D, Lowenfels AB. The epidemiology of pancreatitis and pancreatic cancer. Gastroenterology. 2013;144(6):1252–61.
- Iannuzzi JP, King JA, Leong JH, Quan J, Windsor JW, Tanyingoh D, et al. Global incidence of Acute Pancreatitis is increasing over time: a systematic review and Meta-analysis. Gastroenterology. 2022;162(1):122–34.
- Lee PJ, Papachristou GI. New insights into acute pancreatitis. Nat Rev Gastroenterol Hepatol. 2019;16(8):479–96.
- Mederos MA, Reber HA, Girgis MD. Acute Pancreatitis: a review. JAMA. 2021;325(4):382–90.
- Faghih M, Singh VK. Do elevated triglycerides truly trigger Acute Pancreatitis? Dig Dis Sci. 2019;64(3):616–8.
- Jia W, Xu L, Xu W, Yang M, Zhang Y. Application of nanotechnology in the diagnosis and treatment of acute pancreatitis. Nanoscale Adv. 2022;4(8):1949–61.
- 11. Lankisch PG, Apte M, Banks PA. Acute pancreatitis. Lancet. 2015;386(9988):85–96.
- Greenberg JA, Hsu J, Bawazeer M, Marshall J, Friedrich JO, Nathens A, et al. Clinical practice guideline: management of acute pancreatitis. Can J Surg. 2016;59(2):128–40.
- Schepers NJ, Bakker OJ, Besselink MG, Ahmed Ali U, Bollen TL, Gooszen HG, et al. Impact of characteristics of organ failure and infected necrosis on mortality in necrotising pancreatitis. Gut. 2019;68(6):1044–51.
- van Dijk SM, Hallensleben NDL, van Santvoort HC, Fockens P, van Goor H, Bruno MJ, et al. Acute pancreatitis: recent advances through randomised trials. Gut. 2017;66(11):2024–32.
- Bayda S, Adeel M, Tuccinardi T, Cordani M, Rizzolio F. The history of Nanoscience and Nanotechnology: from chemical-physical applications to Nanomedicine. Molecules. 2019;25(1).
- Xie M, Liu X, Wang S. Degradation of methylene blue through Fenton-like reaction catalyzed by MoS(2)-doped sodium alginate/Fe hydrogel. Colloids Surf B Biointerfaces. 2022;214:112443.
- Patil-Sen Y. Advances in nano-biomaterials and their applications in biomedicine. Emerg Top Life Sci. 2021;5(1):169–76.
- Pelaz B, Alexiou C, Alvarez-Puebla RA, Alves F, Andrews AM, Ashraf S, et al. Diverse applications of Nanomedicine. ACS Nano. 2017;11(3):2313–81.
- Kim BY, Rutka JT, Chan WC, Nanomedicine. N Engl J Med. 2010;363(25):2434–43.
- 20. Yao Q, Jiang X, Zhai YY, Luo LZ, Xu HL, Xiao J, et al. Protective effects and mechanisms of bilirubin nanomedicine against acute pancreatitis. J Control Release. 2020;322:312–25.
- Ahmad A, Rashid S, Chaudhary AA, Alawam AS, Alghonaim MI, Raza SS, et al. Nanomedicine as potential cancer therapy via targeting dysregulated transcription factors. Semin Cancer Biol. 2023;89:38–60.
- Mujahid MH, Upadhyay TK, Khan F, Pandey P, Park MN, Sharangi AB, et al. Metallic and metal oxide-derived nanohybrid as a tool for biomedical applications. Biomed Pharmacother. 2022;155:113791.
- Lammers T, Aime S, Hennink WE, Storm G, Kiessling F. Theranostic nanomedicine. Acc Chem Res. 2011;44(10):1029–38.
- 24. Ravi Kumar MN. Nano and microparticles as controlled drug delivery devices. J Pharm Pharm Sci. 2000;3(2):234–58.
- Hou X, Zaks T, Langer R, Dong Y. Lipid nanoparticles for mRNA delivery. Nat Rev Mater. 2021;6(12):1078–94.
- 26. Zhang Q, Li S, Yu Y, Zhu Y, Tong R. A Mini-review of Diagnostic and Therapeutic Nano-Tools for Pancreatitis. Int J Nanomed. 2022;17:4367–81.
- 27. Qiang H, Li J, Wang S, Feng T, Cai H, Liu Z, et al. Distribution of systemically administered nanoparticles during acute pancreatitis: effects of particle size and disease severity. Pharmazie. 2021;76(5):180–8.
- Xu X, An H, Zhang D, Tao H, Dou Y, Li X, et al. A self-illuminating nanoparticle for inflammation imaging and cancer therapy. Sci Adv. 2019;5(1):eaat2953.

- Gowd V, Ahmad A, Tarique M, Suhail M, Zughaibi TA, Tabrez S, et al. Advancement of cancer immunotherapy using nanoparticles-based nanomedicine. Semin Cancer Biol. 2022;86(Pt 2):624–44.
- Hashimoto D, Ohmuraya M, Hirota M, Yamamoto A, Suyama K, Ida S, et al. Involvement of autophagy in trypsinogen activation within the pancreatic acinar cells. J Cell Biol. 2008;181(7):1065–72.
- Biczo G, Vegh ET, Shalbueva N, Mareninova OA, Elperin J, Lotshaw E, et al. Mitochondrial dysfunction, through impaired autophagy, leads to endoplasmic reticulum stress, deregulated lipid metabolism, and pancreatitis in animal models. Gastroenterology. 2018;154(3):689–703.
- Gorelick FS, Thrower E. The acinar cell and early pancreatitis responses. Clin Gastroenterol Hepatol. 2009;7(11 Suppl):S10–4.
- Witt H, Apte MV, Keim V, Wilson JS. Chronic pancreatitis: challenges and advances in pathogenesis, genetics, diagnosis, and therapy. Gastroenterology. 2007;132(4):1557–73.
- Jin S, Orabi AI, Le T, Javed TA, Sah S, Eisses JF, et al. Exposure to Radiocontrast agents induces pancreatic inflammation by activation of Nuclear Factor-kappaB, Calcium Signaling, and Calcineurin. Gastroenterology. 2015;149(3):753– 64. e11.
- Wang Y, Sternfeld L, Yang F, Rodriguez JA, Ross C, Hayden MR, et al. Enhanced susceptibility to pancreatitis in severe hypertriglyceridaemic lipoprotein lipase-deficient mice and agonist-like function of pancreatic lipase in pancreatic cells. Gut. 2009;58(3):422–30.
- Zierke L, John D, Gischke M, Tran QT, Sendler M, Weiss FU, et al. Initiation of acute pancreatitis in mice is independent of fusion between lysosomes and zymogen granules. Cell Mol Life Sci. 2024;81(1):207.
- Ren J, Jia X, Zhao Y, Shi W, Lu J, Zhang Y, et al. The RIP3-RIP1-NF-kappaB signaling axis is dispensable for necroptotic cells to elicit cross-priming of CD8(+) T cells. Cell Mol Immunol. 2017;14(7):639–42.
- Liang QQ, Shi ZJ, Yuan T, Chen SY, Li YP, Zhang HR, et al. Celastrol inhibits necroptosis by attenuating the RIPK1/RIPK3/MLKL pathway and confers protection against acute pancreatitis in mice. Int Immunopharmacol. 2023;117:109974.
- Boonchan M, Arimochi H, Otsuka K, Kobayashi T, Uehara H, Jaroonwitchawan T, et al. Necroptosis protects against exacerbation of acute pancreatitis. Cell Death Dis. 2021;12(6):601.
- Duan PY, Ma Y, Li XN, Qu FZ, Ji L, Guo XY, et al. Inhibition of RIPK1-dependent regulated acinar cell necrosis provides protection against acute pancreatitis via the RIPK1/NF-kappaB/AQP8 pathway. Exp Mol Med. 2019;51(8):1–17.
- Wen L, Javed TA, Yimlamai D, Mukherjee A, Xiao X, Husain SZ. Transient high pressure in pancreatic ducts promotes inflammation and alters tight junctions via Calcineurin Signaling in mice. Gastroenterology. 2018;155(4):1250– 63. e5.
- Husain SZ, Orabi AI, Muili KA, Luo Y, Sarwar S, Mahmood SM, et al. Ryanodine receptors contribute to bile acid-induced pathological calcium signaling and pancreatitis in mice. Am J Physiol Gastrointest Liver Physiol. 2012;302(12):G1423–33.
- Chvanov M, Voronina S, Jefferson M, Mayer U, Sutton R, Criddle DN, et al. Deletion of the WD40 domain of ATG16L1 exacerbates acute pancreatitis, abolishes LAP-like non-canonical autophagy and slows trypsin degradation. Autophagy. 2025;21(1):210–22.
- Huang W, Cane MC, Mukherjee R, Szatmary P, Zhang X, Elliott V, et al. Caffeine protects against experimental acute pancreatitis by inhibition of inositol 1,4,5-trisphosphate receptor-mediated Ca2 + release. Gut. 2017;66(2):301–13.
- Chen X, Zhong R, Hu B. Mitochondrial dysfunction in the pathogenesis of acute pancreatitis. Hepatobiliary Pancreat Dis Int. 2023. S1499-3872(23)00246-1.
- Du W, Liu G, Shi N, Tang D, Ferdek PE, Jakubowska MA, et al. A microRNA checkpoint for ca(2+) signaling and overload in acute pancreatitis. Mol Ther. 2022;30(4):1754–74.
- Petersen OH, Gerasimenko JV, Gerasimenko OV, Gryshchenko O, Peng S. The roles of calcium and ATP in the physiology and pathology of the exocrine pancreas. Physiol Rev. 2021;101(4):1691–744.
- Hu Z, Wang D, Gong J, Li Y, Ma Z, Luo T, et al. MSCs deliver hypoxia-treated Mitochondria Reprogramming Acinar metabolism to alleviate severe Acute Pancreatitis Injury. Adv Sci (Weinh). 2023;10(25):e2207691.
- Armstrong JA, Cash NJ, Ouyang Y, Morton JC, Chvanov M, Latawiec D, et al. Oxidative stress alters mitochondrial bioenergetics and modifies pancreatic cell death independently of cyclophilin D, resulting in an apoptosis-to-necrosis shift. J Biol Chem. 2018;293(21):8032–47.
- 50. Shen Y, Wen L, Zhang R, Wei Z, Shi N, Xiong Q, et al. Dihydrodiosgenin protects against experimental acute pancreatitis and associated lung injury

through mitochondrial protection and PI3Kgamma/Akt inhibition. Br J Pharmacol. 2018;175(10):1621–36.

- Zhang R, Wen L, Shen Y, Shi N, Xing Z, Xia Q, et al. One compound of saponins from Disocorea zingiberensis protected against experimental acute pancreatitis by preventing mitochondria-mediated necrosis. Sci Rep. 2016;6:35965.
- Mareninova OA, Hermann K, French SW, O'Konski MS, Pandol SJ, Webster P, et al. Impaired autophagic flux mediates acinar cell vacuole formation and trypsinogen activation in rodent models of acute pancreatitis. J Clin Invest. 2009;119(11):3340–55.
- Wang Y, Wang X, Zhang X, Zhang B, Meng X, Qian D, et al. Inflammation and Acinar Cell Dual-Targeting nanomedicines for Synergistic Treatment of Acute Pancreatitis via ca(2+) homeostasis regulation and pancreas autodigestion inhibition. ACS Nano. 2024;18(18):11778–803.
- Lopez-Blazquez C, Lacalle-Gonzalez C, Sanz-Criado L, Ochieng'Otieno M, Garcia-Foncillas J, Martinez-Useros J. Iron-dependent cell death: a New Treatment Approach against Pancreatic Ductal Adenocarcinoma. Int J Mol Sci. 2023;24:19.
- Mareninova OA, Sendler M, Malla SR, Yakubov I, French SW, Tokhtaeva E, et al. Lysosome associated membrane proteins maintain pancreatic acinar cell homeostasis: LAMP-2 deficient mice develop pancreatitis. Cell Mol Gastroenterol Hepatol. 2015;1(6):678–94.
- Wang MJ, Wang YC, Masson E, Wang YH, Yu D, Qian YY, et al. SEC16A variants predispose to chronic pancreatitis by impairing ER-to-Golgi transport and inducing ER stress. Adv Sci (Weinh). 2024;11(38):e2402550.
- Tong J, Wang Q, Gao Z, Liu Y, Lu C. VMP1: a multifaceted regulator of cellular homeostasis with implications in disease pathology. Front Cell Dev Biol. 2024;12:1436420.
- Giacomello M, Pyakurel A, Glytsou C, Scorrano L. The cell biology of mitochondrial membrane dynamics. Nat Rev Mol Cell Biol. 2020;21(4):204–24.
- Lewis S, Evans DL, Tsugorka TT, Peng S, Stauderman K, Gerasimenko O et al. Combination of the CRAC Channel Inhibitor CM4620 and galactose as a potential therapy for Acute Pancreatitis. Function (Oxf). 2024;5(4).
- Waldron RT, Chen Y, Pham H, Go A, Su HY, Hu C, et al. The Orai ca(2+) channel inhibitor CM4620 targets both parenchymal and immune cells to reduce inflammation in experimental acute pancreatitis. J Physiol. 2019;597(12):3085–105.
- Bruen C, Miller J, Wilburn J, Mackey C, Bollen TL, Stauderman K, et al. Auxora for the treatment of patients with Acute Pancreatitis and Accompanying systemic inflammatory response syndrome: Clinical Development of a calcium release-activated Calcium Channel inhibitor. Pancreas. 2021;50(4):537–43.
- 62. Xi H, Shan W, Li M, Wang Z, Li Y. Trehalose attenuates testicular aging by activating autophagy and improving mitochondrial quality. Andrology. 2024.
- 63. Zhao M, Li C, Shen F, Wang M, Jia N, Wang C. Naringenin ameliorates LPS-induced acute lung injury through its anti-oxidative and anti-inflammatory activity and by inhibition of the PI3K/AKT pathway. Exp Ther Med. 2017;14(3):2228–34.
- 64. Dou W, Zhang J, Sun A, Zhang E, Ding L, Mukherjee S, et al. Protective effect of naringenin against experimental colitis via suppression of toll-like receptor 4/NF-kappaB signalling. Br J Nutr. 2013;110(4):599–608.
- 65. Li WS, Lin SC, Chu CH, Chang YK, Zhang X, Lin CC et al. The gastroprotective effect of naringenin against ethanol-Induced gastric ulcers in mice through inhibiting oxidative and inflammatory responses. Int J Mol Sci. 2021;22(21).
- Li Y, Pan Y, Gao L, Zhang J, Xie X, Tong Z, et al. Naringenin protects against Acute Pancreatitis in two experimental models in mice by NLRP3 and Nrf2/ HO-1 pathways. Mediators Inflamm. 2018;2018:3232491.
- 67. Yan X, Lin T, Zhu Q, Zhang Y, Song Z, Pan X. Naringenin protects against acute pancreatitis-associated intestinal injury by inhibiting NLRP3 inflammasome activation via AhR signaling. Front Pharmacol. 2023;14:1090261.
- Wu X, Yao J, Hu Q, Kang H, Miao Y, Zhu L, et al. Emodin ameliorates Acute Pancreatitis-Associated Lung Injury through inhibiting the alveolar macrophages pyroptosis. Front Pharmacol. 2022;13:873053.
- Guo F, Zhou Q, Wu Y, Chen M, Zhao L, Xiang H. Emodin alleviates Sodium Taurocholate-Induced pancreatic ductal cell damage by inhibiting the S100A9/VNN1 signaling pathway. Pancreas. 2022;51(7):739–46.
- Fan J, Duan L, Wu N, Xu X, Xin J, Jiang S, et al. Baicalin ameliorates pancreatic fibrosis by inhibiting the activation of pancreatic stellate cells in mice with chronic pancreatitis. Front Pharmacol. 2020;11:607133.
- Zhao ZF, Zhang Y, Sun Y, Zhang CH, Liu MW. Protective effects of baicalin on caerulein-induced AR42J pancreatic acinar cells by attenuating oxidative stress through mir-136-5p downregulation. Sci Prog. 2021;104(2):368504211026118.

- Qian Y, Chen Y, Wang L, Tou J. Effects of baicalin on inflammatory reaction, oxidative stress and PKDI and NF-kB protein expressions in rats with severe acute pancreatitis1. Acta Cir Bras. 2018;33(7):556–64.
- 73. Sheu SS, Nauduri D, Anders MW. Targeting antioxidants to mitochondria: a new therapeutic direction. Biochim Biophys Acta. 2006;1762(2):256–65.
- Dong X, Fu J, Yin X, Cao S, Li X, Lin L, et al. Emodin: a review of its Pharmacology, Toxicity and Pharmacokinetics. Phytother Res. 2016;30(8):1207–18.
- Lykkesfeldt J, Tveden-Nyborg P. The pharmacokinetics of vitamin C. Nutrients. 2019;11(10).
- Lodge JK, Hall WL, Jeanes YM, Proteggente AR. Physiological factors influencing vitamin E biokinetics. Ann N Y Acad Sci. 2004;1031:60–73.
- 77. Ancuceanu R, Dinu M, Dinu-Pirvu C, Anuta V, Negulescu V. Pharmacokinetics of B-Ring Unsubstituted flavones. Pharmaceutics. 2019;11(8).
- Garcia-Rayado G, Cardenas-Jaen K, de-Madaria E. Towards evidence-based and personalised care of acute pancreatitis. United Eur Gastroenterol J. 2020;8(4):403–9.
- 79. Cao X, Hu Y, Luo S, Wang Y, Gong T, Sun X, et al. Neutrophil-mimicking therapeutic nanoparticles for targeted chemotherapy of pancreatic carcinoma. Acta Pharm Sin B. 2019;9(3):575–89.
- Brandl T, Simic O, Skaanderup PR, Namoto K, Berst F, Ehrhardt C, et al. Trypsin inhibitors for the treatment of pancreatitis. Bioorg Med Chem Lett. 2016;26(17):4340–4.
- Navya PN, Kaphle A, Srinivas SP, Bhargava SK, Rotello VM, Daima HK. Current trends and challenges in cancer management and therapy using designer nanomaterials. Nano Converg. 2019;6(1):23.
- Demirturk N, Bilensoy E. Nanocarriers targeting the diseases of the pancreas. Eur J Pharm Biopharm. 2022;170:10–23.
- Wan Z, Mao H, Guo M, Li Y, Zhu A, Yang H, et al. Highly efficient hierarchical micelles integrating photothermal therapy and singlet oxygen-synergized chemotherapy for cancer eradication. Theranostics. 2014;4(4):399–411.
- Peng J, Xiao Y, Li W, Yang Q, Tan L, Jia Y, et al. Photosensitizer Micelles together with IDO inhibitor enhance Cancer Photothermal Therapy and Immunotherapy. Adv Sci (Weinh). 2018;5(5):1700891.
- Zhou X, Cao X, Tu H, Zhang ZR, Deng L. Inflammation-targeted delivery of Celastrol via Neutrophil membrane-coated nanoparticles in the management of Acute Pancreatitis. Mol Pharm. 2019;16(3):1397–405.
- Pan W, Li Z, Qiu S, Dai C, Wu S, Zheng X, et al. Octahedral Pt-MOF with au deposition for plasmonic effect and Schottky junction enhanced hydrogenothermal therapy of rheumatoid arthritis. Mater Today Bio. 2022;13:100214.
- Kou L, Sun R, Jiang X, Lin X, Huang H, Bao S, et al. Tumor Microenvironment-Responsive, Multistaged Liposome induces apoptosis and ferroptosis by amplifying oxidative stress for enhanced Cancer Therapy. ACS Appl Mater Interfaces. 2020;12(27):30031–43.
- Yao Q, Kou L, Tu Y, Zhu L. MMP-Responsive 'Smart' drug delivery and Tumor Targeting. Trends Pharmacol Sci. 2018;39(8):766–81.
- Taguchi K, Nagao S, Maeda H, Yanagisawa H, Sakai H, Yamasaki K, et al. Biomimetic carbon monoxide delivery based on hemoglobin vesicles ameliorates acute pancreatitis in mice via the regulation of macrophage and neutrophil activity. Drug Deliv. 2018;25(1):1266–74.
- 90. Hassanzadeh P, Arbabi E, Rostami F. Coating of ferulic acid-loaded silk fibroin nanoparticles with neutrophil membranes: a promising strategy against the acute pancreatitis. Life Sci. 2021;270:119128.
- Kou L, Sun R, Xiao S, Zheng Y, Chen Z, Cai A, et al. Ambidextrous Approach to disrupt Redox Balance in Tumor cells with increased ROS production and decreased GSH synthesis for Cancer Therapy. ACS Appl Mater Interfaces. 2019;11(30):26722–30.
- 92. Cruz MEM, Corvo ML, Martins MB, Simoes S, Gaspar MM. Liposomes as tools to improve therapeutic enzyme performance. Pharmaceutics. 2022;14(3).
- Allen TM, Cullis PR. Liposomal drug delivery systems: from concept to clinical applications. Adv Drug Deliv Rev. 2013;65(1):36–48.
- Ren Y, Liu W, Zhang L, Zhang J, Bi J, Wang T, et al. Milk fat globule EGF factor 8 restores mitochondrial function via integrin-medicated activation of the FAK-STAT3 signaling pathway in acute pancreatitis. Clin Transl Med. 2021;11(2):e295.
- Kim TW, Lee SY, Kim M, Cheon C, Ko SG. Kaempferol induces autophagic cell death via IRE1-JNK-CHOP pathway and inhibition of G9a in gastric cancer cells. Cell Death Dis. 2018;9(9):875.
- Qu Y, Li X, Xu F, Zhao S, Wu X, Wang Y et al. Kaempferol alleviates murine experimental colitis by restoring gut microbiota and inhibiting the LPS-TLR4-NF-кВ Axis. Front Immunol. 2021;12.

- Crocetto F, di Zazzo E, Buonerba C, Aveta A, Pandolfo SD, Barone B et al. Kaempferol, Myricetin and fisetin in prostate and bladder Cancer: a systematic review of the literature. Nutrients. 2021;13(11).
- 98. Wen E, Cao Y, He S, Zhang Y, You L, Wang T, et al. The mitochondria-targeted Kaempferol nanoparticle ameliorates severe acute pancreatitis. J Nanobio-technol. 2024;22(1):148.
- Deng Y, Zhang X, Shen H, He Q, Wu Z, Liao W, et al. Application of the Nano-Drug Delivery System in Treatment of Cardiovascular diseases. Front Bioeng Biotechnol. 2019;7:489.
- Ren T, Mi Y, Wei J, Han X, Zhang X, Zhu Q et al. Advances in Nano-Functional materials in targeted thrombolytic drug delivery. Molecules. 2024;29(10).
- 101. Yildirim AB, Gol M, Yigin A, Cimen L, Dinc H, Yildiz H et al. Therapeutic use of fisetin and pirfenidone combination in bleomycin-induced pulmonary fibrosis in adult male albino rats. Naunyn Schmiedebergs Arch Pharmacol. 2024.
- 102. Zamanian MY, Taheri N, Ramadan MF, Mustafa YF, Alkhayyat S, Sergeevna KN, et al. A comprehensive view on the fisetin impact on colorectal cancer in animal models: focusing on cellular and molecular mechanisms. Animal Model Exp Med. 2024;7(5):591–605.
- 103. Elsallabi O, Patruno A, Pesce M, Cataldi A, Carradori S, Gallorini M. Fisetin as a Senotherapeutic Agent: Biopharmaceutical properties and Crosstalk between Cell Senescence and Neuroprotection. Molecules. 2022;27(3).
- Sun Y, Shen X, Yang J, Tan C. Hyaluronic Acid-Coated nanoliposomes as Delivery systems for Fisetin: Stability, membrane fluidity, and Bioavailability. Foods. 2024;13:15.
- 105. Awadeen RH, Boughdady MF, Zaghloul RA, Elsaed WM, Abu H, Meshali II. Formulation of lipid polymer hybrid nanoparticles of the phytochemical Fisetin and its in vivo assessment against severe acute pancreatitis. Sci Rep. 2023;13(1):19110.
- Zhang Y, Sun T, Jiang C. Biomacromolecules as carriers in drug delivery and tissue engineering. Acta Pharm Sin B. 2018;8(1):34–50.
- Mitchell MJ, Billingsley MM, Haley RM, Wechsler ME, Peppas NA, Langer R. Engineering precision nanoparticles for drug delivery. Nat Rev Drug Discov. 2021;20(2):101–24.
- Fu Z, Wang D, Zheng C, Xie M, Chen Y, Zhou Y et al. Elimination of intracellular ca(2+) overload by BAPTA–AM liposome nanoparticles: a promising treatment for acute pancreatitis. Int J Mol Med. 2024;53(4).
- 109. Wang Y, Pu M, Yan J, Zhang J, Wei H, Yu L, et al. 1,2-Bis(2-aminophenoxy) ethane-N,N,N',N'-tetraacetic acid Acetoxymethyl Ester Loaded reactive oxygen species responsive hyaluronic acid-bilirubin nanoparticles for acute kidney Injury Therapy via alleviating calcium overload mediated endoplasmic reticulum stress. ACS Nano. 2023;17(1):472–91.
- Quarato G, Llambi F, Guy CS, Min J, Actis M, Sun H, et al. Ca(2+)-mediated mitochondrial inner membrane permeabilization induces cell death independently of Bax and Bak. Cell Death Differ. 2022;29(7):1318–34.
- 111. Raraty M, Ward J, Erdemli G, Vaillant C, Neoptolemos JP, Sutton R, et al. Calcium-dependent enzyme activation and vacuole formation in the apical granular region of pancreatic acinar cells. Proc Natl Acad Sci U S A. 2000;97(24):13126–31.
- Xiao Y, Xu RH, Dai Y. Nanoghosts: harnessing mesenchymal stem cell membrane for construction of Drug Delivery platforms Via Optimized Biomimetics. Small. 2024;20(1):e2304824.
- 113. Fan L, Wei A, Gao Z, Mu X. Current progress of mesenchymal stem cell membrane-camouflaged nanoparticles for targeted therapy. Biomed Pharmacother. 2023;161:114451.
- Wang Y, Qian D, Wang X, Zhang X, Li Z, Meng X, et al. Biomimetic trypsinresponsive structure-bridged Mesoporous Organosilica Nanomedicine for Precise Treatment of Acute Pancreatitis. ACS Nano. 2024;18(29):19283–302.
- Zhang R, Wu S, Ding Q, Fan Q, Dai Y, Guo S, et al. Recent advances in cell membrane-camouflaged nanoparticles for inflammation therapy. Drug Deliv. 2021;28(1):1109–19.
- Luk BT, Zhang L. Cell membrane-camouflaged nanoparticles for drug delivery. J Control Release. 2015;220(Pt B):600–7.
- 117. Hussain Z, Rahim MA, Jan N, Shah H, Rawas-Qalaji M, Khan S, et al. Cell membrane cloaked nanomedicines for bio-imaging and immunotherapy of cancer: improved pharmacokinetics, cell internalization and anticancer efficacy. J Control Release. 2021;335:130–57.
- Hu CM, Zhang L, Aryal S, Cheung C, Fang RH, Zhang L. Erythrocyte membrane-camouflaged polymeric nanoparticles as a biomimetic delivery platform. Proc Natl Acad Sci U S A. 2011;108(27):10980–5.
- Parodi A, Quattrocchi N, van de Ven AL, Chiappini C, Evangelopoulos M, Martinez JO, et al. Synthetic nanoparticles functionalized with biomimetic leukocyte membranes possess cell-like functions. Nat Nanotechnol. 2013;8(1):61–8.

- 121. Mayerle J, Dummer A, Sendler M, Malla SR, van den Brandt C, Teller S, et al. Differential roles of inflammatory cells in pancreatitis. J Gastroenterol Hepatol. 2012;27(Suppl 2):47–51.
- 122. Wang J, Tang W, Yang M, Yin Y, Li H, Hu F, et al. Inflammatory tumor microenvironment responsive neutrophil exosomes-based drug delivery system for targeted glioma therapy. Biomaterials. 2021;273:120784.
- 123. Ye B, Zhao B, Wang K, Guo Y, Lu Q, Zheng L, et al. Neutrophils mediated multistage nanoparticle delivery for prompting tumor photothermal therapy. J Nanobiotechnol. 2020;18(1):138.
- 124. Yuan S, Hu Q. Convergence of nanomedicine and neutrophils for drug delivery. Bioact Mater. 2024;35:150–66.
- 125. Chen KH, Nguyen N, Huang TY, Lin YJ, Yu YT, Song HL, et al. Macrophagehitchhiked orally administered beta-glucans-Functionalized nanoparticles as Precision-guided Stealth Missiles for targeted pancreatic Cancer therapy. Adv Mater. 2023;35(40):e2304735.
- 126. Shi Y, Yang J, Gao F, Zhang Q. Covalent Organic frameworks: recent progress in Biomedical Applications. ACS Nano. 2023;17(3):1879–905.
- Zhang Y, Fang F, Li L, Zhang J. Self-assembled Organic nanomaterials for Drug Delivery, Bioimaging, and Cancer Therapy. ACS Biomater Sci Eng. 2020;6(9):4816–33.
- 128. Alshammari BH, Lashin MMA, Mahmood MA, Al-Mubaddel FS, Ilyas N, Rahman N, et al. Organic and inorganic nanomaterials: fabrication, properties and applications. RSC Adv. 2023;13(20):13735–85.
- 129. Natesan V, Kim SJ. The Trend of Organic Based nanoparticles in the treatment of diabetes and its perspectives. Biomol Ther (Seoul). 2023;31(1):16–26.
- 130. Wang X, Zhong X, Li J, Liu Z, Cheng L. Inorganic nanomaterials with rapid clearance for biomedical applications. Chem Soc Rev. 2021;50(15):8669–742.
- Lenders V, Koutsoumpou X, Sargsian A, Manshian BB. Biomedical nanomaterials for immunological applications: ongoing research and clinical trials. Nanoscale Adv. 2020;2(11):5046–89.
- Ma X, Tian Y, Yang R, Wang H, Allahou LW, Chang J, et al. Nanotechnology in healthcare, and its safety and environmental risks. J Nanobiotechnol. 2024;22(1):715.
- Khurana A, Tekula S, Saifi MA, Venkatesh P, Godugu C. Therapeutic applications of selenium nanoparticles. Biomed Pharmacother. 2019;111:802–12.
- 134. Hosnedlova B, Kepinska M, Skalickova S, Fernandez C, Ruttkay-Nedecky B, Peng Q, et al. Nano-Selenium and its nanomedicine applications: a critical review. Int J Nanomed. 2018;13:2107–28.
- 135. El-Ghazaly MA, Fadel N, Rashed E, El-Batal A, Kenawy SA. Anti-inflammatory effect of selenium nanoparticles on the inflammation induced in irradiated rats. Can J Physiol Pharmacol. 2017;95(2):101–10.
- 136. Khurana A, Anchi P, Allawadhi P, Kumar V, Sayed N, Packirisamy G, et al. Yttrium oxide nanoparticles reduce the severity of acute pancreatitis caused by cerulein hyperstimulation. Nanomedicine. 2019;18:54–65.
- 137. Tan X, Jia F, Wang P, Zhang K. Nucleic acid-based drug delivery strategies. J Control Release. 2020;323:240–52.
- Buddolla AL, Kim S. Recent insights into the development of nucleic acid-based nanoparticles for tumor-targeted drug delivery. Colloids Surf B Biointerfaces. 2018;172:315–22.
- 139. Zhang T, Tian T, Lin Y. Functionalizing Framework Nucleic-Acid-based nanostructures for Biomedical Application. Adv Mater. 2022;34(46):e2107820.
- 140. Zhou M, Gao S, Zhang X, Zhang T, Zhang T, Tian T, et al. The protective effect of tetrahedral framework nucleic acids on periodontium under inflammatory conditions. Bioact Mater. 2021;6(6):1676–88.

- 141. Gao S, Wang Y, Li Y, Xiao D, Lin Y, Chen Y, et al. Tetrahedral Framework nucleic acids reestablish Immune Tolerance and restore saliva secretion in a Sjogren's Syndrome Mouse Model. ACS Appl Mater Interfaces. 2021;13(36):42543–53.
- 142. Gao S, Zhou M, Li Y, Xiao D, Wang Y, Yao Y, et al. Tetrahedral Framework nucleic acids reverse new-onset type 1 diabetes. ACS Appl Mater Interfaces. 2021;13(43):50802–11.
- 143. Wang Y, Li Y, Gao S, Yu X, Chen Y, Lin Y. Tetrahedral Framework nucleic acids can alleviate Taurocholate-Induced severe Acute Pancreatitis and its subsequent Multiorgan Injury in mice. Nano Lett. 2022;22(4):1759–68.
- 144. Xue S, Schlosburg JE, Janda KD. A New Strategy for Smoking Cessation: characterization of a bacterial enzyme for the degradation of Nicotine. J Am Chem Soc. 2015;137(32):10136–9.
- 145. Huang Y, Ren J, Qu X, Nanozymes. Classification, Catalytic mechanisms, Activity Regulation, and applications. Chem Rev. 2019;119(6):4357–412.
- 146. Wang D, Jana D, Zhao Y. Metal-Organic Framework Derived nanozymes in Biomedicine. Acc Chem Res. 2020;53(7):1389–400.
- 147. Zhou HC, Long JR, Yaghi OM. Introduction to metal-organic frameworks. Chem Rev. 2012;112(2):673–4.
- Wang C, Gao J, Cao Y, Tan H. Colorimetric logic gate for alkaline phosphatase based on copper (II)-based metal-organic frameworks with peroxidase-like activity. Anal Chim Acta. 2018;1004:74–81.
- 149. Zhang J, Guo M, He Q, Zhang Z, Wu B, Wu H, et al. Precise Control of Metal Active Sites of Metal-Organic Framework Nanozymes for achieving excellent enzyme-like activity and efficient pancreatitis therapy. Small. 2024;20(32):e2310675.
- 150. Charkiewicz AE. Is copper still safe for us? What do we know and what are the latest literature statements? Curr Issues Mol Biol. 2024;46(8):8441–63.
- 151. Wu SJ, Yuk H, Wu J, Nabzdyk CS, Zhao X. A multifunctional Origami Patch for minimally invasive tissue sealing. Adv Mater. 2021;33(11):e2007667.
- 152. Kim SR, Yi HJ, Lee YN, Park JY, Hoffman RM, Okano T, et al. Engineered mesenchymal stem-cell-sheets patches prevents postoperative pancreatic leakage in a rat model. Sci Rep. 2018;8(1):360.
- 153. Li J, Moeinzadeh S, Kim C, Pan CC, Weale G, Kim S, et al. Development and systematic characterization of GelMA/alginate/PEGDMA/xanthan gum hydrogel bioink system for extrusion bioprinting. Biomaterials. 2023;293:121969.
- 154. Ibne Mahbub MS, Bae SH, Gwon JG, Lee BT. Decellularized liver extracellular matrix and thrombin loaded biodegradable TOCN/Chitosan nanocomposite for hemostasis and wound healing in rat liver hemorrhage model. Int J Biol Macromol. 2023;225:1529–42.
- Liu Y, Li H, Shu XZ, Gray SD, Prestwich GD. Crosslinked hyaluronan hydrogels containing mitomycin C reduce postoperative abdominal adhesions. Fertil Steril. 2005;83(Suppl 1):1275–83.
- Chen H, Wang S, Sun Y, Wang J. Mitomycin C induces fibroblast apoptosis and reduces intra-articular scar adhesion by regulating miR-21 and its target programmed cell death 4. Fitoterapia. 2020;142:104392.
- Shanto PC, Fahad MAA, Jung HI, Park M, Kim H, Bae SH, et al. Multi-functional dual-layer nanofibrous membrane for prevention of postoperative pancreatic leakage. Biomaterials. 2024;307:122508.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.