



# Sex as a Biological Variable in Preclinical Modeling of Blast-Related Traumatic Brain Injury

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Approaches to furthering our understanding of the bioeffects, behavioral changes, and treatment options following exposure to blast are a worldwide priority. Of particular need is a more concerted effort to employ animal models to determine possible sex differences, which have been reported in the clinical literature. In this review, clinical and preclinical reports concerning blast injury effects are summarized in relation to sex as a biological variable (SABV). The review outlines approaches that explore the pertinent role of sex chromosomes and gonadal steroids for delineating sex as a biological independent variable. Next, underlying biological factors that need exploration for blast effects in light of SABV are outlined, including pituitary, autonomic, vascular, and inflammation factors that all have evidence as having important SABV relevance. A major second consideration for the study of SABV and preclinical blast effects is the notable lack of consistent model design - a wide range of devices have been employed with questionable relevance to real-life scenarios-as well as poor standardization for reporting of blast parameters. Hence, the review also provides current views regarding optimal design of shock tubes for approaching the problem of primary blast effects and sex differences and outlines a plan for the regularization of reporting. Standardization and clear description of blast parameters will provide greater comparability across models, as well as unify consensus for important sex difference bioeffects.

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## INTRODUCTION

Traumatic brain injury (TBI) is a significant military health problem, with the Defense and Veterans Brain Injury Center reporting  $\sim$ 384,000 worldwide cases from the years 2000 to 2018 in the US forces (1), supporting many studies suggesting a TBI incidence rate of  $\sim$ 20% in service members in Operation Iraqi Freedom (OIF) and Operation Enduring Freedom (OEF) (2, 3). The incidence of blast-related TBI, specifically, rose in military personnel during OIF/OEF compared to previous conflicts due to the increased use of explosive materials [e.g., improvised explosive devices (IEDs), rocket-propelled grenades], and explosions have been determined to be the leading causal agent of TBI in Iraq and/or Afghanistan (4–7). Blast-related injuries have also increased in civilian populations worldwide; conservative estimates from the RAND<sup>®</sup> Memorial Institute for the Prevention of Terrorism state a fourfold increase in the number of terrorist incidents employing explosive devices between 1999 and 2006, with the number of injuries resulting from those acts increasing eightfold (8, 9).

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The increasing participation of women in the US military and the lifting of the Combat Exclusion Policy in 2013, officially giving women eligibility to participate in full combat operations, have placed women at greater risk of sustaining a military-related TBI, including blast-related TBI. The number of women using the Veterans Administration in the United States increased by 46% between 2005 and 2015 (10), confirming female veterans are a rapidly growing patient population. Furthermore, many countries enforce mandatory conscription for women (e.g., Israel, Norway), with women occupying combat positions globally, making TBI in female military populations an international health concern. Although there have been a substantial number of studies comparing outcomes following TBI between men and women in clinical populations, particularly in sports-related contexts [for recent reviews, see (11-13)], specific attention to the potentially special needs of women who have sustained TBI in the military is a growing concern (14, 15). There is substantial evidence that women may be more at risk than men for many neurological and psychiatric conditions following military-acquired TBI [e.g., (16-20)], and it has been recently shown that up to 50% of older female veterans (>55 years old) with a diagnosed TBI also had a psychiatric diagnosis of depression or posttraumatic stress disorder (PTSD) (21). Furthermore, the authors reported that a diagnosed TBI increased the risk of dementia by 50%, and the risk increased twofold for women who suffered from any two of those conditions (21).

Animal models of TBI, including models of blast injuries, have aided in our understanding of the pathophysiology and symptomology of brain injuries for decades [for reviews, see (22-25)]. Since the National Institutes of Health (NIH) mandate requiring inclusion of both sexes in biomedical research (26), the number of preclinical TBI studies including females has increased. There have been several recent reviews on sex differences following TBI in animal models (13, 27–29). However, the majority of translational TBI work employing both male and female animals has been performed with more severe and/or surgically invasive TBI models such as controlled cortical impact [CCI; e.g., (30-35)], fluid percussion injury [FPI; e.g., (36, 37)], or repetitive concussive brain injury [CBI; e.g., (38, 39)]. Although blast injury models have been studied in male animals of many species (23, 40), there is a near absence of female inclusion in blast models of neurotrauma.

The purpose of this review is to discuss TBI, particularly as inflicted by a blast event, in the context of sex as a biological variable (SABV). What are known about the functional consequences of military-acquired blast TBI are discussed, followed by a description of the present experimental approaches that have had utility in manipulating sex chromosomes and gonadal steroids as independent variables. Dependent variables, including pituitary, autonomic, vascular, and inflammation factors, are then discussed because these focus on the most reported systems that are perturbed by blast. Finally, relevant to the study of blast and SABV in preclinical studies, the review makes an appeal for investigators to apply the highestquality experimental principles, because the study of SABV in this field is complex and requires the derivation of the uppermost-quality information for asking further questions and laying the groundwork for translational relevance. Suggestions and guidelines are provided for the use and reporting of sufficient information about blast animal models that will aid in the interpretation of data and generation of conclusions.

## CONSEQUENCES OF MTBI IN MILITARY POPULATIONS

## **Mild TBI Symptoms**

Mild TBI (mTBI) as a result of an explosion often leads to symptoms that are well-studied in military populations. The symptoms are most often short-term, resolving within 7–10 days, and often include physical (e.g., headache, dizziness, nausea), cognitive (e.g., memory and concentration problems), and behavioral (e.g., anxiety, irritability) complaints (41, 42). However, a small percentage of patients ( $\sim$ 10–25%) will have symptoms persisting >3 months and will be diagnosed with postconcussive syndrome (PCS), which can include a variety of symptoms ranging from psychiatric (anxiety and depression) to physical (headache, fatigue, dizziness) and sleep disturbances, among others (43, 44).

In addition to potential long-term symptoms following mTBI, there is a clear link between military-acquired TBI and PTSD [e.g., (4, 45, 46)]. PTSD has many overlapping symptoms with mTBI and PCS (e.g., irritability, fatigue, poor sleep, memory, and attention problems), but PTSD is often referred to as an "abnormally sustained stress response" with added symptoms of nightmares, hyperarousal, avoidance, and re-experiencing phenomena (47). Fully understanding the relationship between TBI and PTSD, and whether blast-related TBI carries a higher risk of a PTSD development than TBI incurred by other mechanisms, has been a challenge for researchers, and discussion of this complex topic is beyond the scope of this article (47). However, blast exposure clearly puts individuals at risk of the development of PTSD (4, 48–52) and other psychiatric conditions such as depression (4, 48–50).

## **Comparisons of Blast vs. Non-blast TBI**

It has been noted that the study of head trauma in military populations is difficult. Although blast has been the most common cause of mTBI in recent conflicts, military personnel are simultaneously at risk of TBI from other causes such as motor vehicle accidents, rigorous training exercise and sports, falls, fights, etc. (53), making it possible or even likely that a subject in a study has sustained more than one TBI of different types (blast, concussion, etc.) over their deployment, or earlier in their lifetime. Greer et al. (54) recently conducted a meta-analysis of the literature comparing clinical and functional outcomes in blast and non-blast TBI in US OIF/OEF service members and veterans. For most outcome measures studied (i.e., vision loss, vestibular dysfunction, depression, sleep disorders, alcohol abuse), there were no differences between blast and non-blast TBI groups. For other outcome measures (i.e., PTSD diagnosis and symptom severity, headache, hearing loss, and neurocognitive function), results were inconsistent (54). Thus, there were no functional measures that could be definitively linked to blast-induced TBI. Importantly, the authors also reported that the majority of the studies used varying definitions of "blast" and "non-blast" injury, and there was often little information about the blast injury, including how close the individual was to the explosion, if he/she was in a vehicle or dismounted, whether there was a loss of consciousness, additional trauma, etc. (54). Indeed, it has been noted that the majority of sustained blast injuries include a mixture of secondary and tertiary injuries (sometimes called "blast-plus" see *Blast Events*), making the contributions of the primary blast wave to the subsequent outcomes difficult to clearly establish (55, 56).

Several publications, however, employed in vivo imaging and discovered morphological or functional differences between blast-exposed cases and other forms of TBI. An initial key observation was evidence of white matter tract changes. Davenport et al. (57) examined white matter tract integrity with diffusion tensor imaging (DTI) in service members with a reported blast-related TBI and with no reported blast exposure or signs of mTBI. Twenty subcortical white matter tracts were evaluated for differences in fractional anisotropy (FA; a measure of white matter integrity). Ten white matter tracts had significantly lower FA measures in the cases with blastrelated TBI. The authors noted that the analysis required a close assessment of the regions of interest, and the differences were diffuse and widespread, and the specific tracts with alterations varied across the cases. Levin et al. (58) performed DTI assessments and an extensive characterization of veterans and service members with exposure to blast and a group with no TBI or blast-exposure history. No differences were found between the groups, but FA measures in some brain regions were associated with impairments of verbal memory. DTI was employed by MacDonald et al. (59) to compare alterations in service members who had a history of blast exposure, as well an additional blast-related trauma (e.g., impact with objects, a fall, or motor vehicle crash). The control group in this study comprised individuals with blast exposure and other injuries, but who had not received a TBI diagnosis (59). DTI changes were seen in the service members with a diagnosis of TBI. Alterations in DTI were employed by Bazarian et al. (60) to assess the relationship of white matter tract alterations and mTBI with the severity of PTSD symptoms. FA measures were associated with blast exposure, and PTSD severity was associated with stress symptoms and abnormal DTI, but not with an assessment of mTBI, suggesting DTI changes were observed in "subclinical TBI" cases (60). Taber et al. (61) compared white matter changes in veterans with primary blast exposure (but no TBI symptoms), individuals with reported primary blast exposure consistent with no TBI symptoms or a lack of signs to indicate mTBI, or no exposure to blast. Compared to veterans who had no history of blast exposure, veterans who had sustained a blast event, with or without a diagnosis of TBI, were found to have lower FA and higher radial diffusivity [a general correlate of myelin damage; (62)]. Similar to the observations of Davenport et al. (57), the changes were heterogeneous and widely dispersed. A significant observation from this report is that the changes were seen even in participants with no presenting TBI complaints. Trotter et al. (63) also examined white matter integrity in veterans with a history of blast exposure or with no reported incidence. The participants were 19–62 years of age and had sustained severe blast injury and were compared with service members who had no exposure to blast. In each group, some of the participants had a diagnosis of TBI (69 and 53%, respectively). Alterations in white matter integrity were associated with the intensity of blast exposure, and decreased degree of FA was associated with the number of years since the most severe blast injury.

Several studies have employed functional magnetic resonance imaging (fMRI) to assess cerebral blood flow as a correlate of cerebral activity. Han et al. (64) found blast-related TBI disrupted resting-state cortical network function compared to participants who also had experienced blast exposure but were not diagnosed with TBI. Robinson et al. (65) described a difference between functional connectivity within components of the default mode network when service members were close to a blast (<10 m) compared to individuals located at a site that was farther from the blast. fMRI was used by Fischer et al. (66) while participants attended to the Stop Signal Task, a measure of response inhibition/impulse control (67, 68). Participants included individuals who sustained blast-related TBI, control (uninjured) military personnel, civilians with no TBI, and civilians with non-blast TBI. fMRI activation was lower in service personnel during correct inhibition responses compared to military controls personnel in brain regions associated with response inhibition and the default mode network. Interestingly, the service members with blast-related TBI exhibited greater activation than controls in trials where the respondent failed to appropriately inhibit their response during the Stop Signal Task. In contrast, non-blast civilians displayed an opposite process where TBI civilians had less activation compared to civilian controls. As noted later (see Vascular Alterations From TBI), vascular changes are noted in clinical and preclinical studies of blast effects. Sullivan et al. (69) applied arterial spin labeling to assess possible changes after blast. An increase in the total number of blast exposures was associated with increased cerebral perfusion, but there was no noted relationship to blast proximity or a diagnosis of mTBI or PTSD (69).

Several trends, then, are gained from MRI. First, alterations in white matter integrity have been observed, and the findings suggest these cases exhibit diffuse changes and variability in the location of changes (60, 63, 65, 66), although there is perhaps overlap with impact-related mTBI cases [e.g., (61, 70, 71)]. Second, there are intriguing hints of differential metabolic, gray matter, alterations, particularly for fMRI analyses where the injuries from blast are associated with milder impairments on some performance tests. The finding of a variance in the default mode network response pattern in blast and non-blast TBI cases may point the way to differences in mechanism (66). Lastly, some observations suggest MRI differences are observed in cases where clinical diagnoses of mTBI are not reported (60, 61). Relative to sex differences (most of the cited studies had few or no female participants), white matter alterations and fMRI changes may provide important clues, including potential differences related to activity during performance tasks (66).

## Women and Military-Related TBI

Research on military health and blast-induced brain injuries has largely focused on males, as they have historically made up the majority of service members of the US military and have been more likely to occupy combat roles. However,  $\sim$ 300,000 female service members were deployed to Iraq and/or Afghanistan between 2001 and 2013 (72), and they currently make up 15% of active-duty armed forces. The incidence rate of TBI in deployed women is estimated at  $\sim 10\%$ , about half that of deployed men (16, 20, 45, 73-75). It should be noted that the causes of TBI in military women differ substantially from those of men; intimate partner violence (including physical and sexual assault) is recognized as a significant risk factor for TBI in military women compared to their nonmilitary peers (14, 19, 76). However, the increasing participation of women in combat operations during deployment in recent years continues to put them at greater risk of combat-related TBI (77), and like male service members, blast events have been identified as the greatest cause of combatrelated injuries, including mTBI, in women during OIF/OEF (16, 78).

Overall, following combat-related mTBI, women are likely to suffer the same symptom clusters as their male military peers, such as PCS, PTSD, psychiatric complaints (i.e., anxiety and depression), and somatic symptoms (e.g., vestibular and somatosensory dysfunction) (16-19, 45). However, several studies comparing the outcomes of male and female service members following mTBI have reported differences in the frequencies of specific diagnoses and symptoms between men and women. In a recent scoping study describing the literature addressing gender differences in outcomes following TBI in military populations, Cogan et al. (19) identified 29 relevant articles from 2000 to 2018. One clear conclusion was that women are very underrepresented; most of the studies were not specifically focused on gender differences, and women represented <20% of the sample. The most consistent finding to date was that following a TBI, females in the military are subsequently more susceptible to depression than male service members and veterans (16, 19, 45, 79, 80).

In addition to depression, there is evidence that female service members may have increased susceptibility to anxiety disorders and/or PTSD following mTBI. The literature describing gender differences in PTSD symptoms in military personnel is relatively broad and reports mixed results, possibly as a result of variations in the definition of TBI or methodological differences (18). In an earlier study, Iverson et al. (16) reported that although men were more likely to be diagnosed with PTSD alone following mTBI, women were more likely to have PTSD with comorbid depression. Women were also more likely to suffer from a non-PTSD anxiety disorder and/or to have more than one psychiatric diagnosis compared to men. By subsequently adjusting the model for blast exposure, the authors were able to provide some insights into the potential specific contributions of blast injury to sex differences in outcomes following TBI; there were no longer differences in the likelihood of a PTSD diagnosis alone, frequency of non-PTSD anxiety disorders, or diagnosis of more than one psychiatric condition (16). These results suggest that blast may uniquely contribute to the female susceptibility to anxiety disorders and PTSD with comorbid depression and to the diagnosis of multiple psychiatric diagnoses.

Because of the complexity of ascertaining relevant variables for blast mTBI etiologies, it is important to supplement the clinical literature by applying preclinical animal research. To further understand the role of sex-related variables as proximate causes for sex differences, animal modeling enables greater control of conditions and the ability to more invasively explore cellular response mechanisms from blast exposure. Following an overview of approaches to the study of SABV as an independent variable for blast TBI preclinical work, there is a summary of what is presently known concerning blast bioeffects on pituitary and the hypothalamic–pituitary–adrenal (HPA) axis, the autonomic nervous system, the vasculature, and inflammation.

In addition to the NIH mandate regarding consideration of SABV in clinical and preclinical research, a second policy relates to scientific rigor by employment of preclinical experimental practices that derive valid and reliable findings to adequately address research gaps, set the stage for discovering important mechanisms underlying sex difference and properly modeling translational testing (81–83). Accordingly, there is discussion for a second important feature of preclinical blast research related to principles for application of shock tubes, the most common approach for preclinical modeling.

## SABV IN BLAST MTBI RESEARCH

## SABV

Approaches to the study of SABV in animals has been clearly articulated in several reviews (84–87). With respect to TBI, data summarized by Gupta from 43 studies that examined sex differences, using many different outcome criteria following a variety of TBI models (CCI, FPI, CBI), concluded that females fared better in 55% of the studies, and none indicated males had a better outcome (13). Their Table 2 included a single preclinical blast paper by Russell and colleagues; reviewed below in *HPA Axis Dysfunction in Laboratory Animals After Blast*. The review by Rubin and Lipton (29) of 50 articles found high variability in outcomes, but they too concluded that generally females fared better after injury by FPI, CCI, and weight drop.

For preclinical study of SABV and blast effects, **Table 1** summarizes the main dimensions for investigation of sex as an independent variable. The aforementioned publications regarding experimental design are excellent summaries, and the most salient issues related to sex chromosomes and steroid hormone status are discussed. Subsequently, what are perhaps the most relevant bioeffects of blast exposure, as dependent variables, are outlined, with particular attention to previous preclinical findings in blast TBI experiments.

## **Gonadal Hormone Effects**

Some evidence suggests there is no relationship for estrous phase as a significant impact on outcomes after TBI (88– 90). However, potentially subtle endocrine factors that have important mechanistic ramifications may be overlooked when studies do not account for potential differences related

TABLE 1	Approaches	to sex as a	biological	variable in	preclinical	blast research	n.
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Sex-related variable	Experimental approach	Relevancy
Sex chromosomes	Male testis-determining gene, Sry mouse model, X* mouse strains	Permit study of the impact of <i>Sry</i> and possibly other Y chromosome–encoded genes; X chromosome (single X, XX, models) permit study of the genetic load of the X chromosome
Estrous cycle factors	Assessment and comparison of endocrine status	Ascertainment of ovarian cycle effects with injuries sustained at a particular stage of the ovarian cycle may lead to insights regarding differential effects on outcome
Gonadal hormone status	"Endocrine ablation" by gonadectomy; hormone replacement	Assess gonadal steroid effects upon dependent variables. Other factors include reproductive status, possible relevancy to contraceptives, hormone replacement therapies, steroid or anabolic steroid use, exposure to endocrine disrupting chemicals (e.g., phthalates, bisphenol), menopause, eating disorders, intense physical activities

to estrous cycle stage in females (84). There is strong evidence supporting the neuroprotective roles of estrogen and progesterone, suggesting that female animals may be more resistant to the deleterious effects of injury during the proestrous phase of the cycle, when levels of hormones are at their highest. If there is specific interest in estrous cycle effects, initial studies to evaluate SABV related to blast can be directed to the basic hormone status of laboratory animals by assessment of menstrual cycle. Becker et al. (84) suggest the experimental design could compare male rodents with four groups of females, one group at each stage of the estrous cycle. This allows the researcher to determine if sex and/or the variable levels of steroid hormones across the estrous cycle affect the dependent variable(s) in question. To evaluate the estrous cycle stage, vaginal smear examination should be performed daily, and it has been suggested to perform the examination for at least 8 days immediately prior to an experiment (91). Likewise, for better assignment to hormone status, it is suggested that animals be excluded should they not exhibit regular cyclicity (91). When experimental questions relate to the estrous cycle, these are important considerations, and care should be taken in defining estrous cycle stage, as hormone levels change very rapidly during the day, particularly during proestrous when progesterone levels are peaking (84).

There has been speculation concerning the significance of estrous status in laboratory animals, and some have warned that this is challenging in rodents with shorter cycles where there is inherent variability, even across time of day. Disregarding cycle effects was considered problematic because females may exhibit greater data variability, perhaps complicating interpretation. Alternatively, it is argued that employing female animals at random/cycling stages of estrous more accurately represents the clinical condition. Nonetheless, comparisons of measures in female and male mice and rats suggest variability may not be a significant factor (92-94). Shansky (94) has pointed out other related factors that affect hormone status should be considered, including housing conditions, which was found to affect variability and that group housing of male rodents can alter testosterone levels. Circadian or seasonal factors may also come into play as a variable (95, 96). In addition, some reports relate changes due to female hormonal status, and findings from an initial study of sex differences may suggest the need for closer examination of estrous cycle as an important variable.

### Sex Chromosomes

The pioneering observations of Nettie Maria Stevens documented the spermatozoa of Tenebrio molitor mealworms contain nine similarly sized chromosomes and a smaller chromatin element related to male offspring; in contrast to spermatozoa with 10 chromosomes of equal size associated with female progeny (97). Thus began the intriguing pursuit of sex chromatin differences, subsequent XY nomenclature, and attention to their potential significance in sex-linked disorders (98). The genetic sex of neurons, glia, the cerebral vasculature, and other support cells of the central nervous system and the response of peripheral organ systems to blast injuries are important variables for investigation. Potential differences attributable to sex chromosome effects relate to X chromosome exclusion, where in female progeny the maternal X chromosome (X<sub>M</sub>) or the paternal X chromosome (X<sub>P</sub>) is silenced by X chromosome inactivation (XCI) to partially rebalance the level of expression (99). XCI leads to a mosaic expression pattern in females where the cells in an organ express X<sub>M</sub> or X<sub>P</sub>, although across the female population there is further complexity related to the degree of mosaicism and that a proportion of genes on the "silenced" X chromosome escape inactivation (99). Genes encoded on the male Y chromosome may also have differential effects on cell phenotype and responses to injuries if the pathways are not also homologously encoded on X chromosomes (84).

The mammalian Y chromosome encodes the testisdetermining gene, Sry, which initiates testes formation and spermatogenesis, as well as a small number of additional genes with X-linked homologs that, in females, escape XCI (99). One approach to understand the differential contributions of hormone effects and sex chromosome effects employs the "four core genotype" design in mice (100). The four genome design includes deletion of Sry from the male Y chromosome and insertion of the gene in an autosome. This allows the creation of four genomes: (1) an XY complement with the Sry gene for XY mice with testes; (2) an XY complement without Sry, resulting in XY mice with ovaries; (3) an Sry mouse with the gene incorporated into an autosome resulting in an XX mouse with testes; and (4) an XX mouse with no copy of Sry, resulting in XX mice with ovaries. The mice with similar gonadal forms then permit investigation of the sex chromosome complementation (XX vs. XY) in the context of gonad-related hormonal status (101). To date, this paradigm has not been employed in preclinical blast studies. However, sex chromosome differences have been associated with pathological effects. Li et al. (102) used a cardiac ischemia/reperfusion model and found infarct size was greater with two X chromosomes, independent of gonadal status, compared to XY mice. A second study in this report employed the XY\* mouse model that allows comparisons for the number of X chromosomes and likewise found XX mice exhibited poorer recovery than 1X females (102). Other sex chromosome–related models available, and more complex genomic analyses can be applied (98, 103, 104). No preclinical studies have examined sex chromosome effects after blast injuries.

### **Gonadal Steroid Effects and Steroid Receptors**

Despite some debate regarding menstrual cycle status as a significant factor in TBI outcome (see Gonadal Hormone Effects), gonadal steroid action has been a main, classic focus for the study of sex differences. This is particularly relevant because there is strong evidence of neuroprotective roles of estrogen and progesterone after a range of brain injuries (105-108). As an initial procedure for the study of SABV, Becker et al. (84) describe a standard "two-step approach" for the study of SABV that comprises an initial effort to determine steroid action by gonadectomy, followed by procedures to provide replacement of the hormone. In the first procedure, male and female animals receive a gonadectomy as a comparison to endocrinologically intact animals. Separate groups of animals receive a sham procedure where the identical surgery is performed to externalize the gonads followed by replacement in situ. At specified times after the procedure, animals are utilized in the study. If the gonadectomy resulted in an experimental change for the variables under study, the second step is undertaken where gonadal steroids are administered to gonadectomized animals, whereas a control group receives similar treatment by administration of the vehicle diluent for the hormone(s). Becker et al. (84) note that a third group can be incorporated in this step by including gonadally intact animals as a comparison. Differences are preliminarily interpreted as indication of a gonadal steroid contribution to the biological process under study.

There are additional variables to consider for the twostep approach, including the timing of testing (e.g., surgical treatment or hormone replacement following brain injury) following gonadectomy in the first step as well as after hormone replacement in the second phase of the study. Experienced investigators suggest that the administration of gonadal steroids should be monitored to ensure the replacement procedure provides levels of steroid within the physiological range. Further studies can employ the same approach with compounds that block steroid synthesis or that disrupt steroid receptor effects, or to determine the role of clinically relevant intervening effects on steroid action such as contraceptives. Maintenance on the contraceptives, desogestrel and drospirenone, e.g., was found to reduce the severity of stroke neuropathology in ovariectomized mice (109). Finally, further experiments can be performed to determine the role of specific steroid receptors subtypes. In one of the first articles to explore the role of the two estrogen receptors (ERs), Dubal et al. (110) employed a stroke model that occluded the anterior cerebral artery in ovariectomized mice. Some of the mice were given estrogen replacement in Silastic capsules or the vehicle alone, sesame oil. In the ER $\alpha$  knockout mice provided with physiological levels of 17 $\beta$ -estradiol, level of injury was equivalent to what was observed in wild-type mice or ER $\beta$ knockout mice that were not provided with estradiol, indicating the  $\alpha$  receptor mediates the neuroprotective effects of estrogen.

## **Bioeffects of Blast Exposure**

A second level of inquiry relates to blast-related mechanismsthe dependent variables-that may be differentially affected by sex differences. Outlined below are the most salient biological effects known to date for the impact of blast exposure relevant to SABV. The discussion for some effects begins with clinical descriptions, but some of the reportage concerns findings with non-blast-related methods that may help point to relevant effects, including sex-relevant differences of pituitary and HPA axis function, and blast effects on the autonomic nervous system function, the vasculature, and inflammation. Although researchers often focus on a single dimension of outcome, investigators have recognized that TBI consequences demonstrate it manifests as a systemic condition (111, 112). Likewise, it is probably a significant truism that blast exposures should be considered a polytrauma. High-energy shock wave exposure injures, or at least perturbs, all organ systems, leading to complex, reciprocal interaction between peripheral organs and tissues and central nervous system networks.

# Clinical HPA Axis Dysfunction After TBI and Blast Exposure

Although more studies of military-acquired, particularly blastrelated, mTBI in women are required, a picture is emerging of a gender dichotomy in the stress response following mTBI. There are clear sex differences in non-TBI civilian populations in the lifetime susceptibility to depression and anxiety disorders (113, 114), as well as evidence from the civilian literature that women may be more susceptible to psychiatric disorders following mTBI [e.g., (115–118)], although data are not entirely consistent (13). Anxiety and depression, as well as PTSD, are linked to the HPA axis, the major neuroendocrine system that controls responses to stress (119–122).

The primary stress hormone is cortisol (CORT; corticosterone in laboratory rodents), which is released by the HPA axis when activated by a physical or psychological stressor. The stress response is characterized by release of corticotropinreleasing factor (CRF) from the paraventricular nucleus (PVN) of the hypothalamus, which binds to CRF receptors on the anterior pituitary gland. The anterior pituitary gland secretes adrenocorticotropic hormone (ACTH) into the bloodstream, where it reaches the adrenal cortex and binds to receptors to stimulate the synthesis and release of the steroid hormones, glucocorticoids (e.g., CORT), and mineralocorticoids (e.g., aldosterone). Steroid hormone receptors are located throughout the brain [see (123) for more detailed review], including limbic regions involved in emotion and responses to stressful stimuli.

Clinical studies have demonstrated HPA axis dysfunction in a proportion of individuals several months to years following mild to moderate TBI (119, 120, 124). The pituitary gland is especially

vulnerable to damage, with multiple potential syndromes resulting from hormonal deficiencies (e.g., hypogonadism, hypothyroidism, central diabetes insipidus) (125, 126). Little is known about HPA axis disruption following blast injury, although there are reports indicating decreases in pituitary function up to 2 years following blast-related mTBI (127) or moderate to severe blast TBI (128). A follow-up study found that pituitary dysfunction following blast-related mTBI was associated with increased neuropsychiatric symptoms (i.e., anxiety, irritability) compared to individuals with mTBI and normal pituitary hormone levels (129).

# HPA Axis Dysfunction in Laboratory Animals After Blast

In addition to the insight provided by clinical studies, translational studies employing animal models have allowed further probing of the pathological underpinnings of TBIinduced HPA axis dysfunction (122, 130-136), as well as the use of validated and controlled behavioral paradigms for measuring anxiety- and depressive-like symptoms following experimental TBI (137, 138). Serum levels of ACTH have been shown to decrease 1 month following blast injury in male rats, followed by an increase at 3 months postinjury, suggesting a biphasic blastinduced hypothalamic-pituitary dysfunction (139). Recently, Zuckerman et al. (140) evaluated the CORT response in male rats at more acute time points following blast exposure. Animals exposed to blast had elevated CORT levels 3 h following blast that returned to baseline within 5 h. However, rats with a PTSDlike phenotype, as assessed by their behavior 1 week following injury in the elevated plus maze (EPM; a test for anxiety) and the acoustic startle response (tests for heightened responses to a sensory stimulus), had blunted CORT responses compared to blast-exposed rats with a "well-adjusted" phenotype (140).

Although investigators have recently turned their attention to sex factors in a variety of TBI models [for reviews, see (13, 27– 29)], preclinical studies of blast effects, and specifically on the effects of blast on HPA axis function and/or the development of anxiety and depressive disorders, remain essentially nonexistent. In fact, Russell et al. (136, 141) are the only investigators to date to assess sex differences in the effects of blast-induced TBI in an animal model. In two publications, they reported the effects of mild blast TBI on central and HPA axis function (136) and on CRF receptor gene expression and anxiety-like behaviors (141). Sex differences following exposure to blast overpressure in the advanced blast simulator (ABS; described in more detail below) were reported in both studies.

First, the authors employed a restraint-induced stress model and demonstrated that while blast injury increased the restraintinduced rise in CORT levels in males, the opposite effect was observed in female mice, with blast attenuating CORT levels in restrained animals compared to sham-treated mice. Blast did not alter CORT suppression in the dexamethasone-suppression test or affect the expression of pituitary or adrenal genes involved in ACTH or CORT synthesis or secretion, suggesting a central disruption in feedback, rather than a peripheral effect, as the more likely source of the sexually dimorphic response to injury. Examining potential central nervous system sources, it was first

determined that there were no effects of blast injury in either males or females on mRNA expression of mineralocorticoid and glucocorticoid receptors at central feedback regulation sites: the PVN or other brain limbic structures [e.g., amygdala, hippocampus, bed nucleus of the stria terminalis (BNST)]. However, a restraint-induced increase in CRF neuron activation was differentially altered by blast injury in male and female mice: in males with restraint treatment, blast (compared to sham treatment) reduced CRF neuron activation in the PVN; in females, restraint-treated mice receiving blast treatment had increased levels of CRF neuron activation in the PVN. Retrograde tracing determined that there was a TBI-related decrease of CRF neurons in female mice primarily in preautonomic (nonneuroendocrine) neurons in the PVN, suggesting a decreased use of the preautonomic system in dealing with stressors, leading to a possible blast-induced disruption in CRF outputs to brainstem structures regulating autonomic function. There were no blastinduced changes in the percentage of activated CRF neurons that were endocrine projecting or preautonomic projecting in male mice, and the authors hypothesized that disruption in limbic structures of the HPA axis may result from blast-induced TBI.

A second study was designed to measure changes in the expression of CRF receptor subtypes 1 and 2 (CRFR1, CRFR2, respectively) in limbic structures following blast-induced brain injury in male and female mice, as well as to assess the sexdependent effects of blast on anxiety-like behaviors (141). CRFR1 is widely distributed throughout the brain, and blocking these receptors reduces psychiatric symptoms, whereas expression of CRFR2 is more localized, and activation of these receptors dampens stress responses (142, 143). Blast did not affect CRFR1 expression in either male or female mice, but the injury altered CRFR2 expression in limbic structures in a sexually dimorphic way. The restraint-induced increase in CRFR2 expression was reduced by blast injury in the dorsal hippocampus in females, and in the amygdala and anterior BNST of male mice. In addition, in males, blast injury increased basal CRFR2 (non-restraintinduced) expression in the ventral hippocampus. These changes in CRFR2 expression were paralleled by decreased time spent in the open arms of the elevated plus maze by both males and females, indicating elevated levels of anxiety. The authors suggest that the increase in anxiety following blast injury results from the downregulation of CRFR2 and reduced compensation for the angiogenic effects of the CRFR1 (141). This hypothesis is supported by the observation that CRFR2 knockout mice have increased anxiety-like behaviors (144). Furthermore, the sex differences observed in regional changes in CRFR2 expression post-TBI suggest that male and female mice employ different limbic circuits to cope with the effects of TBI.

### Autonomic Nervous System Function After TBI

During the acute period following TBI, systemic effects appear to result from excessive catecholamine release and subsequent autonomic dysfunction. In more severe cares, autonomic dysfunction leads to transient episodes of paroxysmal sympathetic hyperactivity (PSH), which includes tachycardia, hypertension, hyperthermia, spasticity, and tachypnea (145– 147). A recent review by Baguley and colleagues (148) provides

support for an excitatory:inhibitory ratio model. TBI that includes damage to the mesencephalon results in the loss of descending inhibitory inputs to spinal pathways, resulting in acute, non-nociceptive stimulatory, autonomic overreactivity. Fernandez-Ortega et al. (149) studied 179 severe TBI patients and found  $\sim 10\%$  of the sample exhibited PSH; all were male patients. For blast wave exposure, acute autonomic responses are elicited from pulmonary injuries ("blast lung"), an organ particularly susceptible to damage, resulting in cardiorespiratory distress [c.f., (150)]. Blast lung symptoms include bradycardia and prolonged hypotension, as well apnea episodes followed by rapid, shallow breathing, where bradycardia and hypotension are a result of vagal reflex responses, whereas the hypotension results from autonomic changes, direct heart damage, and the acute release of the potent vasodilator, nitric oxide [cf., (151, 152)]. Pulmonary hemorrhage and edema, as well as later proinflammatory mediators, are activated, which further compromise pulmonary function (152, 153). To date, there appear to be no publications that have explored PSH after blast injuries, as well as no studies of PSH and more severe cases of blast lung that compared the sexes.

Evidence for persistent cardiovascular changes after milder cases of TBI has been reported, with alterations in cardiac rhythm variability providing an overall, integrated indicator of autonomic function (154-156). In milder insults, it is hypothesized that injury results in subtle anatomical lesions in central autonomic networks that give rise to functional changes seen in potentially unhealthful or lethal cardiac irregularities (154). Manifestation of dysregulation may only be evident with close physiological monitoring of autonomic response challenges, such as standing, but less conspicuous changes are also reported during the resting, supine state (154). The six studies reviewed by Bishop et al. (156) appear to have focused on male athletes. "However, Hilz et al. (154) reported on three females and 17 males. La Fountaine et al. (157) studied three subjects (two females, one male), and Senthinathan et al. (158) studied seven females and four males, but none of the reports analyzed sex differences. For blast injury, there appear to be no studies that have examined SABV for cardiac variability or other autonomic changes. However, SABV for autonomic differences is important. In general, females exhibit greater vagal activity, whereas males generally manifest higher sympathetic activity [cf., (155) for review], and uninjured females exhibit a greater baseline of heart rate variability (159). Likewise, gonadal hormones are known to modulate autonomic nervous system networks, where, e.g., estrogen administration to male and ovariectomized rats increased cardiac baroreflex response (160, 161).

## Vascular Alterations From TBI

### Evidence of physical damage

Perhaps on par with reports of significant changes in neuroinflammation after blast exposure (see below), vascular alterations from blast exposure have received the greatest research attention. A particularly vulnerable organ to blast exposure is the pulmonary system, where more energetic shock waves result in significant lung contusions and accompanying autonomic dysregulation (see *Autonomic Nervous System*) *Function After TBI*) and further trauma with leukocyte recruitment and the release of proinflammatory signals [(153) and see *Inflammatory Factors*]. However, in addition to lung response, other effects are observed throughout the vasculature.

An oft-cited hypothesis for the initial physical effects for brain injury relates to "hydrodynamic pulse through venous vasculature," a mechanism purported mainly by Cernak (162, 163). Briefly, the energy from a blast exposure is transferred to the body causing a rapid alteration in abdominal, thoracic, and central venous pressure. Cernak (163) cites Gelman's (164) report that  $\sim$ 70% of blood volume in humans is in the venous compartment compared to 18% in arteries and the remaining 3% in terminal arteries and arterioles. The abrupt pressure change in the arterial and venous vasculatures further contributes to rapid pressure changes in the common carotid artery and inferior vena cava, inducing fluid sheer stress that may result in platelet-activating factor-induced neutrophil activation (163, 165), as well as additional complex interactions (163). Some reports have described peripheral organ damage for endothelial barriers (166). However, a blastmimicking pulse to the thorax of anesthetized rats also causes widespread neuroinflammation, evident by tumor necrosis factor  $\alpha$  in perivenular regions in the brain and activated microglia and macrophages adjacent to veins (167). Investigations in rodents also have described cases of blast exposure resulting in signs of minor cerebral injury, including instances of tears of penetrating cortical vessels, microhemorrhages, swelling, and end-feet degeneration of perivascular astrocytes (168-170). All of the aforementioned studies have employed male laboratory rodents. Finally, the injury effects of blast exposure, having an impact on central functions, including central autonomic networks and immunomodulation, are potentially complex interactions where peripheral injuries affect cerebral functions and reciprocal links from brain to peripheral organs (112).

### Cerebral vasospasm

A common sequelae to blast exposure is cerebral vasospasm (171). The publication by Armonda and colleagues was one of the first clinical reports to describe this phenomenon, most frequently evident in more severe cases (172). Of interest was that the vasospasm occurred as a delayed phenomenon, peaking about 2 weeks after blast injury and lasting for at least a month, the length of the study (172). In the report of Armonda et al., the sex of the casualties was not reported, but  ${\sim}50\%$  of the injured patients exhibited vasospasm. Although there have been no descriptions of sex differences in vascular reactivity after blast exposure, young females admitted to hospitals after impact-related TBI were found to show vulnerabilities. Czosnyka et al. (173) observed that after accidents young women exhibited greater cerebral hypertension and reactivity. In severely injured patients [requiring intubation, mechanical ventilation, intracranial pressure (ICP) monitoring], Sorrentino et al. (174) likewise reported a vulnerability, where a more favorable outcome was observed if younger female patients had lower ICPs and lower pressure-reactivity index (PRx; a measure of cerebral autoregulation), perhaps in line with reports of higher vulnerability in females (and older patients) with ICP. Hamer et al. (175) recently reported observations in young athletes (19–21 years of age) who had sustained single or multiple concussions. Males were found to exhibit lower cerebral blood flow in temporal regions, whereas female athletes with a history of concussions were not different from uninjured females. However, females who had sustained multiple concussions, compared to women who sustained a single concussion, exhibited lower cerebral blood flow in the left anterior cingulum and right cerebellum and middle occipital gyrus. The role of vasospasm was not addressed, but the authors speculate the alterations may relate to long-term central metabolic activity changes or perhaps a loss of cerebral volume from injuries.

Alterations in vascular reactivity have been reported in preclinical studies, but studies are skewed to males. As noted by Mollayeva et al. (176), there appears to be a discrepancy in the preclinical literature, where more frequently female laboratory animals exhibited better cerebral hemodynamics after TBI. A study by Armstead et al. (177), e.g., studied pial microvascular responses after fluid percussion injury in piglets. Following injury, male piglets were observed to exhibit greater reductions in pial artery diameter, cortical cerebral blood flow, and cerebral perfusion pressure, as well as greater elevation of ICP after injury (177).

### **Inflammatory Factors**

In addition to signs of vasculature damage, inflammation is often observed acutely with proximate tissue damage, as well as over the long term as a secondary consequence (27). Cernak et al. (178) pioneered in describing acute systemic inflammation after blast exposure. "Local" effects of blast exposure were observed, including the activation of eicosanoids-bioactive, locally released immune system signals (179). This group sampled plasma from 65 blast-injured male personnel, using an inclusion criteria of signs of lung injury, and found higher blood levels of thromboxane A2, prostacyclin (PGI2), and sulfidopeptide leukotrienes, in comparison with 62 patients who sustained similar levels of injury severity, but had not sustained blast exposure. Subsequent studies using whole-body imaging in mice found elevated myeloperoxidase activity, a measure of activated phagocytes, throughout the gastrointestinal tract, lungs, and brain that persisted for at least 1 month, with central nervous system response suggesting a higher expression at 1 month, the last time point assessed (180). Similar observations have been reported from blast trauma in the lungs and brains of male rats (181). Gorbunov et al. (153) described pulmonary contusions from shock wave exposure (alveolar rupture and blood extravasation) and the release of proinflammatory signals, including macrophage inflammatory protein-2, interleukin 6 (IL-6), monocyte chemoattractant protein-1, and cytokine-induced neutrophil chemoattractant-2 [summarized in Gorbunov et al. (153)].

Central nervous system inflammation is a key variable after TBI. Investigators hypothesize brain injury leads to chronic, lower-level neuroinflammation that results in insidious neurodegeneration. Johnson et al. (182), e.g., observed evidence of neuroinflammation in 28% of their TBI patients at more than 1 year after injury and up to 18 years after insult. In preclinical blast

injury studies, microgliosis, usually assessed by alterations in Iba-1 staining, is regularly observed (168, 181, 183-188). Likewise, reactive astrocytes, which may mediate proinflammatory and anti-inflammatory effects, are a common benchmark (181, 189-191). In all of these reports, male laboratory animals were used exclusively. Other evidence of inflammatory signals following TBI is commonly reported. For example, Späni et al. (27) recently summarized their findings from a number of their studies that levels of several cytokines and chemokines were elevated in the brain after closed head injuries, including IL-1β, IL-6, tumor necrosis factor  $\alpha$  (TNF- $\alpha$ ), IL-10, CXCL1, and CCL2, and sex differences were noted where the concentrations IL-6, TNFα, and CCL2 levels were higher in female mice after injury, compared with males. Blast exposure likewise results in cytokine responses, which includes IL-1β, IL-6, IL-12, IL-18, IFN-γ, and TNF- $\alpha$ , and chemokines, monocyte chemoattractant protein-1, GRO, and RANTES [e.g., (185, 189, 192-195)]. However, no studies to date that evaluated protein or mRNA changes have examined sex differences in expression in animal models.

# PRECLINICAL MODELING OF BLAST FOR THE STUDY OF SEX DIFFERENCES

## The Challenge of Modeling Blast Events

As just reviewed, interpretation of the patient literature on blast effects is a challenge and can at best be viewed as "unsettled" regarding bioeffects and potential differences based on sex. Likewise, there is a dearth of preclinical reports that have investigated SABV. However, for preclinical research of sex differences, this can be viewed as a unique opportunity to get things right. Likewise, there are compelling reasons for getting things right for investigators to recognize the relevance of matching, as best as possible, in-laboratory blast experiments to real-world scenarios. Experimental approaches to preclinical modeling of blast effects then relate not only to present efforts and mandates to evaluate sex differences (26), but also for recognition of potentially important bioeffects from blast. A clear understanding of blast exposure effects has extraordinary relevance to how to direct efforts to treatment, requiring rigorously established models. This section reviews important parameters that have been recognized for their role in reaching valid conclusions for preclinical research studies. Previous approaches, which have not to date so much addressed blast research, have shown how complex sex difference studies can be, and-consequently-strong experimental design is critical to what may be small but significant experimental effects. The majority of publications related to neurobiological effects have employed laboratory shock tubes using compressed gases (40, 196). This will be the emphasis in the discussion below.

Modeling blast effects is a task. Over five decades ago, White (197) summarized the state of the science for understanding "shock and blast biology." He recognized the vast challenges of outlining the relevant physical and biological parameters for delineating the hazards to man. White emphasized the need for closer collaboration especially between physicists and biologists—although he makes note of critical additional

collaboration with engineers, architects, and physicians—for each expert to bring their discipline to bear on this problem. The need for integration continues to be echoed by experts, where only through collaborative efforts between blast physicists and biologists (198), military-relevant and academic researchers (199), and "surgical engineers" (200) and that there can be progress by learning from clinical cases that elucidate what symptoms require mimicking in animal studies (199). The blast neuroscience or neuroendocrinology investigator, then, should seek collaboration and ongoing consultation with the appropriate experts who can immensely improve the quality of the research effort.

## **Blast Events**

In an explosion, the rapid expansion of detonation products drives a supersonic shock wave into the surrounding air. The ambient air is compressed in microseconds as the shock front passes a location, after which the pressure falls rapidly to pressures below ambient levels over the timescale of milliseconds. The shock front is also associated with an immediate jump in air-flow velocity, or "blast wind," which can be of hurricane strength, although this also decays rapidly along with the overpressure. The majority of TBIs sustained by blast are classified as mTBI, defined by the Department of Defense as a loss of consciousness <30 min, posttraumatic amnesia for 24 h or less, and alteration of consciousness for a duration <24 h (201). Brain injury resulting from explosive blast occurs as a result of several mechanisms: (1) primary-direct impact on bodily tissues caused by the abrupt variation in air pressure resulting from the blast overpressure wave, (2) secondarypenetrating or blunt injuries as a result of debris set in motion from the blast, (3) tertiary-coup/countercoup injuries resulting from acceleration and deceleration of the body and head or the head/body striking the ground or other object (202-204). Although most blast-induced brain injuries result from primary through tertiary mechanisms, also spoken of are quaternary injuries that result from intense heat (burns) and quinary injuries such as infections, radiation illness, tetanus, and poisoning that are varied and are the result of other injurious factors that are released at the time of the explosion (199).

In a free-field setting, the Friedlander curve (Figure 1A) is used as the model for an ideal blast wave, and with specific design, this waveform can be replicated by especially designed laboratory shock tubes (206, 207). The key feature of the blast wave is the shock front, causing a nearly instantaneous change in the gasdynamic properties of the air such as the static pressure, flow velocity, density, and temperature. The shock front thickness is less than a micron translating to a rise time of the order of a nanosecond; this shock front itself is capable of tissue disruption due to the extreme rate of loading. While the human body can endure extremes of pressure (300 psi in the case of "free-divers"), tissue is highly sensitive to rate of change of pressure, in this case in the form of a supersonic wavefront. Following the shock front, the gas-dynamic conditions decay uniformly to below ambient levels (the negative phase) before gradually returning to ambient. The duration of the positive phase is dependent on the scale of the blast being several milliseconds in the case of a typical roadside IED. Simplistically, the static overpressure of the wave causes crushing action, whereas the combination of high-flow velocity and high air density represents the "blast wind" effect causing displacement action and the tertiary blast injury effects described earlier.

Blast physics experts have emphasized the extreme complexity of real-scenario explosions and that while it is one means for setting experimental conditions, including well-designed shock tube studies, the Friedlander wave has been adapted as a model for free-field explosions, but the waveform in no way mimics the high variation and complexity of conditions (208). In real-world scenarios, the target and surrounding objects have a great influence on the blast waveform, and shock wave interactions with surroundings lead to complex reflected waves that can amplify intensity and be followed by secondary shocks and variable negative shock wave phases (208). An explosion above ground, e.g., will cause a complex shock wave due to the effect of the ground reflection. A compound wave structure develops involving a Mach stem with shock wave properties of much greater severity than the incident blast (209). Likewise, when a shock wave encounters a wall or traverses an enclosed space, the reflected wave can be 2-14 times the magnitude of the incident wave (209, 210). For an IED, the shock wave characteristics are altered by a number of interacting factors such as charge shape (e.g., IEDs designed for penetration of vehicles), the encasement of the charge, and the subsurface location that adds tertiary effects from the highvelocity ejecta (dirt, casement, additional components of the IED such as metal shard, toxic and exothermic chemicals). All of these components add immeasurably to the complexity of the injuries; laboratory conditions are simplifications. Nevertheless, the idealized Friedlander-type waveform remains an important reference standard for "free-field" blast exposures for the purposes of laboratory research studies. Although a conventional shock tube was never intended to generate the specially tailored waveform distinctive of explosive blast, within certain important constraints a good approximation can be achieved.

Factors associated with biomechanical differences related to scaling, sex, and age are also of relevance. For example, blast effect sex differences for human males and females have received little attention, but—while there is significant overlap—there are reported average differences in size and skull thickness that can have different consequences on skull flexure during shock wave loading (211–213). Likewise, the skull shape differences are significantly dissimilar for different species used in preclinical study (214), for the determination of sex differences in primates, but mouse differences appear to be trivial (215). Lastly, the focus in blast research has been on younger women and men. The impact of hormone status in older adults and laboratory animals, chronic disease conditions, and aging, as predominant and overriding contributors to morbidity, has not been investigated in blast studies.

## Modeling Preclinical Blast for SABV

For several years now outstanding—and edifying—publications outline details for proper design of shock wave studies (198,



206-208, 216-219). Several of the noted publications, e.g., emphasize the important issue of specimen placement. Most experts indicated that placement of specimens just outside a shock tube is problematic, because either the nature of the shock wave in this position is extremely difficult to characterize, or the shock wave has components that diverge significantly from the conventional Friedlander profile. Specimen position near the exterior of a shock tube results in exposures that are significantly different from a free-field waveform, where the "exit jet" exhibits anomalies (Figure 1B), including multiple peaks, rarefaction waves, and unclear combinations of sonic blast and subsonic effects, including large gradients in flow (198, 220). A second important consideration is the size of the specimen relative to the dimensions of the tube. Referred to as the "presented area," the specimen should not occupy more than 5-10% of the crosssectional area of the tube to not impede the steady flow of the shock wave (Figure 1C), where blockage alters the free-field flow of the shock wave and can cause ancillary tertiary effects from specimen acceleration (198, 219). Optimal design of the shock tube can provide a location inside the apparatus (Figure 1D) that minimizes end-tube perturbations, including a strong reflection wave generated when the tube has an open end, and can further control the waveform by "tailoring" to reduce transverse and longitudinal waves inherent in tubes (208). Likewise, when attempting to mimic primary shock wave conditions, securing the specimen is an important additional factor. The restraint

system may contribute to test injuries resulting from blast wind effects that result in impacts with the holder (198). Sawyer et al. (221), e.g., emphasize how dynamic pressure can cause head movements, leading to increased staining for glial fibrillary acidic protein, which is a common "confirmatory" injury observation in shock tube publications. In their model, elevations in staining were observed when the head was not restrained, while head fixation, limiting effects to primary shock wave effects, showed no change (222). In addition to the constraints described previously, efforts should be directed to not solely apply a strong air blast as a model for research without ensuring it meets characteristics that are relevant to actual conditions.

## Research Guidelines and Standards for Results Reporting

Related to the need for proper design of shock studies, there has been a recent convergence of views regarding a critical aspect of blast research progress with the paramount need for standardization of results reporting for blast studies (23, 206, 208, 210, 216, 223–227). Clear description of static, dynamic, stagnation, and reflected pressure and how these properties are measured and interpreted are critical. The necessity of standardization of reporting was especially emphasized by experts with concerns regarding the proliferation of blast devices with questionable relevancy to real-life scenarios and the potential for misleading interpretations of biological effects that

Title	Description
Blast-induced delivery device	Device used to induce blast injury
Pressure wave type	Friedlander wave is an instantaneous rise in pressure immediately followed by a decay curve; idealized blast in open space; can be reproduced in tube
Detonation type	Material for open field explosions, blast tube explosions
Detonation material quantity	Quantity of material used for open field explosions, blast tube explosions
Driver gas	Gas used to generate overpressure in shock tube
Pressure wave medium	Medium through which blast wave travels to reach target
Distance from detonation	For open-field exposures
Blast tube or column area	Area of the distal end of the blast tube/column or shock tube/column
Blast tube length	Length of the blast tube; use when no membrane is used
Shock tube driven section length	Length of the shock tube driven section; use when membrane present
Membrane/diaphragm thickness	Thickness of membrane between driver and driven sections of shock tube
Membrane/diaphragm burst method	Indicate whether membrane is punctured or allowed to rupture by gas pressure buildup in driver section of shock tube
Membrane/diaphragm burst pressure (shock tube)	Pressure at which the membrane/diaphragm within the shock tube bursts
Tube end configuration	Is the tube end "open" or "closed"
Placement of animal relative to shock tube	Inside or outside the shock tube
Distance between the animal and the tube end	Indicate how far animal is from the end of the shock or blast tube
Animal orientation to the blast wave	Describe positioning of the animal relative to the blast wavefront
Overpressure peak (blast or shock)	Incident pressure
Overpressure rise time	A measure of how rapidly pressure changes from the ambient level to the maximum positive value, defined as the time required for pressure to increase from 10% to 90% of the maximum positive value
Overpressure wave duration (pulse width)	Full width at half maximum amplitude
Impulse	Integration of overpressure with respect to time
Reflective wave overpressure	Pressure measured following reflection or dampening; overpressure following interference
Blast wind pressure	The post-shock or blast wind is important in describing the complete blast wave
Pressure sensor orientation	Location of pressure gauge needed to assess temporal, spatial characteristics of measured pressure
Pressure sensor type	Indicate type of pressure sensor used to characterize, calibrate, and/or record pressure
Pressure sensor sampling frequency	Pressure sensor sampling frequency
Incident pressure time history (image)	Incident pressure time history (image)
Body exposure	Designates whether whole body is exposed to pressure or is partially shielded
Protective shielding	Location
Protective shielding type	Nature of material used for shielding
Reflective surfaces (where and type)	Indicates the presence and nature of reflective or dampening surfaces integrated into blast wave path
Primary blast effects	Methodology employed to isolate primary blast effects from secondary, tertiary, or quaternary effects
Secondary blast effects type	Secondary blast effects include the effects of any projectile, including fragments of debris, propelled by the blast that penetrates the skin. This may be modeled with a blast (primary blast effect) or in isolation to mimic the secondary blast effects associated with a blast. Cross reference with penetrating models of brain injury as appropriate
Secondary blast effects specifications	Entered to further explain "secondary blast effect type."
Tertiary blast effects type	Tertiary blast effects describe when explosion propels body and brain is injured due to acceleration and/or impacts the ground or a surrounding object. For animal models, could be used to describe the head hitting the ground or object, or ground or object hitting head. For small objects, use secondary blast effects
Tertiary blast effects specifications	Provide further explanation of methods used to induce tertiary injury and/or methodology to measure resultant forces or accelerations. Cross reference with blunt force and/or acceleration model CDEs as necessary. For head impact only (i.e., no blast), use appropriate CDE (e.g., weight drop model)
Quaternary blast effects	Quaternary blast effects include toxic gas inhalation, thermal exposure, flash burns, microwave heating, electromagnetic fields
Systemic injury	Measures of systemic inflammation/stress as a result of the blast (including primary, secondary, tertiary, quaternary effects)
Extracranial injuries	Injuries other than brain injury that occurs as a result of the blast (including primary, secondary, tertiary, quaternary effects)
Blast-induced specific preinjury surgical procedures Blast-induced specific postinjury surgical procedures	Description of any presurgical procedures specific to the blast-induced neurotrauma model Description of any postsurgical procedures specific to the blast-induced neurotrauma model

\*From https://fitbir.nih.gov/content/preclinical-common-data-elements.

do not reflect the actualities of blast biology and the difficulties for synthesis and summarizing findings from such laboratories with uncommon shock devices (23, 198, 199, 216, 217, 225).

Indubitably, this challenge of comparability from TBI research endeavors is not restricted to preclinical models of blast. Clinical TBI investigators have been formalizing data reporting since 2008 (228-230), with the formation of the Interagency Common Data Elements Project for TBI, with an updated version described in Hicks et al. (231). A website for clinical study registration and data storage, as a repository permitting eventual secondary meta-analyses by the TBI community, was established by the Federal Interagency Traumatic Brain Injury Research Informatics System for TBI Research (FITBIR; https://fitbir.nih. gov/). The initiative got underway from a Workshop for the Classification of TBI for Targeted Therapies held in October 2007, by the National Institute of Neurological Disorders and Stroke, with the participation of representatives from other groups, including the Defense and Veterans Brain Injury Center and the National Institute on Disability and Rehabilitation Research. The working group initially focused on the limitations of diagnostic criteria and that a pathoanatomical classification system could be the springboard for addressing the heterogeneity of TBIs and for improved systemization for clinical studies and trials (232). A commentary by Dr. John Povlishock, editor of the Journal of Neurotrauma, emphasized the importance of this enterprise to basic scientists for their assessment of pathobiology in preclinical research (233). In 2012, FITBIR initiated the effort for a data recording system that employs common data element terminologies.

A subsequent meeting, the Traumatic Brain Injury Preclinical Working Group, was convened to develop a dictionary of common data elements for preclinical studies (234). This group emphasized the importance of the initiative for further 'enhancing rigor, reproducibility, and transparency in study performance" in preclinical studies. The CDEs are available as a Preclinical TBI CDE Zip File in Excel format at https://fitbir. nih.gov/content/preclinical-common-data-elements. The Excel files list 61 "Core, Module 1" descriptors (species, animal age, vendor, treatment conditions and outcome measures, etc.) and 41 elements in "Module 6," specific to blast/shock studies (Table 2). The recent publication of Rodriguez et al. (235) is an excellent example following this scheme. Finally, there are a number of efforts to encourage open data sharing (236), including unpublished data, dubbed "dark data" (237), and efforts to promote preregistration of studies for peer-centered review of studies (238). For good progress in determination of sex differences and blast effects, these initiatives may move the field forward.

Of added high relevance for preclinical blast research is the framework of the NATO Task Group, HFM-234 (220). This document resulted in the dissemination of useful guidelines, including rules for more detailed description of the blast (or shock) exposure device (219) and the specifics of animal modeling (239). In addition to allowing comparisons across laboratories, improved standardization and description of conditions can lead to improvement of data quality where the guidelines permit funding bodies to better evaluate the

TABLE 3 | Checklist for experimental planning of preclinical blast studies\*.

- 1) Start with a clearly stated question you wanted to answer
- 2) What was the rationale for selecting the model you did?
- 3) The model must be a valid model for the question
- 4) What parameters will be measured (both biomechanical and biological) and how are they related to real-life conditions or other published work?
- 5) Can you vary the parameters accurately within field-relevant range, so you can examine the range of observed injuries?
- 6) Have recognition that there are limits to your model so that results are not overinterpreted
- Need to ask if these changes you see in the animal model are changes we would see in humans
- 8) Rationale for using the animal model, the species, weight, gender, age, etc., a description of all the things that matter, i.e., 20- vs. 60-kg pig is important as well as how firmly they are fixed
- 9) Expected kinetic therefore the rationale for choosing specific time points. Justification of your end points. This may be species specific?
- 10) Where are the animals placed in a test field? Show clearly in a diagram with respect to loading source. Rationale for this. In the guide will describe drawbacks or issues with placing an animal in certain areas of the tube
- 11) Have to give the relevant exposure for the question they are answering, not overexposing or underexposing the animals for the problem they are trying to answer
- 12) Can you relate observed pathophysiological changes as a function of external loading and different time points?
- 13) Justification for the use of a certain technique, e.g., use of explosives instead of compressed gas for primary blast experiments
- 14) Justify the specific placement and binding of the animal in the experimental model through direct pressure, acceleration, and strain measurements on the animal or animal surrogates
- 15) A plan for the statistics, and where possible a power calculation, and estimation of n numbers
- 16) Can rodents be used or would gyrencephalic species, such as ferrets or pigs, be needed?
- 17) Will the skull thickness, head shape, and orientation of the animals affect the result when translated to an erect human with face forward to blast?

\*Table 1 in Appendix J1 from NATO Health Factors and Medicine (HFM) Research Task Group (RTG) HFM-234 (220).

experimental plan and design of proposals, and journal reviewers and editors to have a better sense of the quality of reported findings (239).

The original publications should be consulted for full discussion, but some of the most salient challenges are outlined here. **Table 3** is a summary checklist for experimental planning; many of the queries in the checklist overlap with the Common Data Elements in **Table 2**, but it is included here because there is additional emphasis on investigator review of study rationale and description of the shock/blast-generating apparatus. Investigators of blast effects on preclinical models of sex differences should first consider the details for inducing blast overpressure [cf., (219)].

The application of blast, whether using free-field exposure or a laboratory-based apparatus that employs blast (explosion) or compressed gas as the driver, must be recognized for its complexity of model application, the inherent pitfalls in each model, and the onus for understanding and communicating exposure metrics. A detailed description of the design of the experimental setting should be documented, including the dimensions of the free-field conditions or the shock/blast

tube, for investigators to have a clear sense of the exposure conditions. A published description of the environment becomes a permanent record of intervening effects permitting metaanalyses through standard reporting. Full reportage also allows documentation of existing or potential experimental artifacts, such as reflective and blockage effects from the surroundings (including gauge or animal holder interference) and potential constraining factors such as shock tube dimensions, the size, location, orientation, fixation/restraint conditions of the study specimen(s), and exposure conditions of the test specimen(s) in relation to the overpressure source (209, 217, 219). The guidelines address additional considerations. Is the stimulus reproducible and controllable, and how have the conditions been quantified? What were the conditions pertaining to reflection? What is the intended nature of the injury? If the focus is primary blast, what conditions are in place to mitigate secondary and tertiary effects (207, 239)? Finally, other considerations must be heeded, including choice of recording devices that accurately allow spatial assessments of static pressure for aboveambient pressure (207) and total pressure from the motion of gas (dynamic pressure) with the static pressure, assessed by Pitot tube (219). Do measurement devices have adequate sensor bandwidth to accurately record changes (219, 240)? The aforementioned publications offer excellent overviews for experimental design and emphasize the point of the studiestranslation of results-which demands validation of findings and their relevance to real-event scenarios and accurate and correct identification of underlying pathology as an entrée to therapeutic translation.

For the role of sex in underlying biological responses, the field of laboratory blast studies *is ripe to get things right*. As noted by the Traumatic Brain Injury Preclinical Working Group, preclinical blast research studies are presently an "immature research area" where the lack of more substantial clinical information as a guide to basic hypothesis-driven research is a challenge; the causal effects following blast are emerging

## REFERENCES

- Defense and Veterans Brain Injury Center (DVBIC). DoD Numbers for Traumatic Brain Injury. (2018). Available online at: https://dvbic.dcoe.mil/ system/files/tbi-numbers/worldwide-totals-2000-2018Q1-total\_jun-21-2018\_v1.0\_2018-07-26\_0.pdf
- Tanielian T, Jaycox LH. Invisible wounds of war: psychological and cognitive injuries, their consequences, and services to assist recovery. *RAND Corporation Monograph Series*. (2008). Available online at: http://www.rand. org/pubs/monographs/MG720/.
- Swanson TM, Isaacson BM, Cyborski CM, French LM, Tsao JW, Pasquina PF. Traumatic brain injury incidence, clinical overview, and policies in the US military health system since 2000. *Public Health Rep.* (2017) 132:251– 9. doi: 10.1177/0033354916687748
- Hoge CW, McGurk D, Thomas JL, Cox AL, Engel CC, Castro CA. Mild traumatic brain injury in U.S. Soldiers returning from Iraq. N Engl J Med. (2008) 358:453–63. doi: 10.1056/NEJMoa072972
- Wojcik BE, Stein CR, Bagg K, Humphrey RJ, Orosco J. Traumatic brain injury hospitalizations of U.S. Army soldiers deployed to Afghanistan and Iraq. Am J Prev Med. (2010) 38:S108–16. doi: 10.1016/j.amepre.2009.10.006

but still uncertain, and there are no commonly accepted injury devices (234), as is seen in other models employed in SABV with hundreds of publications. Challenges in the future related to blast and SABV will demand clarification of potential "differences in metabolism, pharmacokinetics, receptor distribution and activity, enzyme activity, and [how] ongoing hormonal interactions may affect whether a particular intervention exhibits important neurological properties" (241). Rigor and standardization are critical for furthering our understanding of sex differences, which are complex and potentially of smaller effect sizes, but critical for translating findings for clinical relevance.

## **AUTHOR CONTRIBUTIONS**

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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- Betthauser LM, Adams RS, Hostetter TA, Scher AI, Schwab K, Brenner LA. Characterization of lifetime TBIs in a cohort of recently deployed soldiers: the warrior strong study. *Rehabil Psychol.* (2019) 64:398– 406. doi: 10.1037/rep0000286
- Regasa LE, Agimi Y, Stout KC. Traumatic brain injury following military deployment: evaluation of diagnosis and cause of injury. *J Head Trauma Rehabil.* (2019) 34:21–9. doi: 10.1097/htr.000000000 000417
- Wolf SJ, Bebarta VS, Bonnett CJ, Pons PT, Cantrill SV. Blast injuries. *Lancet.* (2009) 374:405–15. doi: 10.1016/S0140-6736(09)60257-9
- Mathews ZR, Koyfman A. Blast injuries. J Emerg Med. (2015) 49:573– 87. doi: 10.1016/j.jemermed.2015.03.013
- Office of Data Governance and Analytics. Women veterans report: the past, present, and future of women veterans. Washington DC: National Center for Veterans Analysis and Statistics, Department of Veterans Affairs. (2017). Available online at: https://www.va.gov/womenvet/docs/womenvet\_history. pdf
- Covassin T, Savage JL, Bretzin AC, Fox ME. Sex differences in sportrelated concussion long-term outcomes. *Int J Psychophysiol.* (2018) 132:9– 13. doi: 10.1016/j.ijpsycho.2017.09.010

- Mollayeva T, El-Khechen-Richandi G, Colantonio A. Sex & gender considerations in concussion research. *Concussion*. (2018) 3:Cnc51. doi: 10.2217/cnc-2017-0015
- Gupte R, Brooks W, Vukas R, Pierce J, Harris J. Sex differences in traumatic brain injury: what we know and what we should know. *J Neurotrauma*. (2019) 36:3063–91. doi: 10.1089/neu.2018.6171
- Amoroso T, Iverson KM. Acknowledging the risk for traumatic brain injury in women veterans. J Nerv Ment Dis. (2017) 205:318–23. doi: 10.1097/NMD.00000000000621
- Kim LH, Quon JL, Sun FW, Wortman KM, Adamson MM, Harris OA. Traumatic brain injury among female veterans: a review of sex differences in military neurosurgery. *Neurosurg Focus.* (2018) 45:E16. doi: 10.3171/2018.9.focus18369
- Iverson KM, Hendricks AM, Kimerling R, Krengel M, Meterko M, Stolzmann KL, et al. Psychiatric diagnoses and neurobehavioral symptom severity among OEF/OIF VA patients with deployment-related traumatic brain injury: a gender comparison. *Womens Health Issues*. (2011) 21:S210– 7. doi: 10.1016/j.whi.2011.04.019
- Brickell TA, Lippa SM, French LM, Kennedy JE, Bailie JM, Lange RT. Female service members and symptom reporting after combat and noncombat-related mild traumatic brain injury. *J Neurotrauma*. (2017) 34:300– 12. doi: 10.1089/neu.2016.4403
- Lippa SM, Brickell TA, Bailie JM, French LM, Kennedy JE, Lange RT. Postconcussion symptom reporting after mild traumatic brain injury in female service members: impact of gender, posttraumatic stress disorder, severity of injury, and associated bodily injuries. *J Head Trauma Rehabil.* (2018) 33:101–12. doi: 10.1097/HTR.00000000000353
- Cogan AM, McCaughey VK, Scholten J. Gender differences in outcomes after traumatic brain injury among service members and veterans. *PM R*. (2019) 12:301–14. doi: 10.1002/pmrj.12237
- Gray M, Adamson MM, Thompson RC, Kapphahn KI, Han S, Chung JS, et al. Sex differences in symptom presentation and functional outcomes: a pilot study in a matched sample of veterans with mild TBI. *Brain Inj.* (2020) 34:535–47. doi: 10.1080/02699052.2020.1725979
- Yaffe K, Lwi SJ, Hoang TD, Xia F, Barnes DE, Maguen S, et al. Militaryrelated risk factors in female veterans and risk of dementia. *Neurology*. (2019) 92:e205–11. doi: 10.1212/WNL.00000000006778
- O'Connor WT, Smyth A, Gilchrist MD. Animal models of traumatic brain injury: a critical evaluation. *Pharmacol Ther.* (2011) 130:106– 13. doi: 10.1016/j.pharmthera.2011.01.001
- Risling M, Davidsson J. Experimental animal models for studies on the mechanisms of blast-induced neurotrauma. *Front Neurol.* (2012) 3:30. doi: 10.3389/fneur.2012.00030
- Johnson VE, Meaney DF, Cullen DK, Smith DH. Animal models of traumatic brain injury. *Handb Clin Neurol.* (2015) 127:115–28. doi: 10.1016/B978-0-444-52892-6.00008-8
- Povlishock J. The history and evolution of experimental traumatic brain injury models. *Methods Mol Biol.* (2016) 1462:3–7. doi: 10.1007/978-1-4939-3816-2\_1
- 26. Clayton JA, Collins FS. Policy: NIH to balance sex in cell and animal studies. *Nature*. (2014) 509:282–3. doi: 10.1038/509282a
- Späni CB, Braun DJ, Van Eldik LJ. Sex-related responses after traumatic brain injury: considerations for preclinical modeling. *Front Neuroendocrinol.* (2018) 50:52–66. doi: 10.1016/j.yfrne.2018.03.006
- Kim T, Chelluboina B, Chokkalla AK, Vemuganti R. Age and sex differences in the pathophysiology of acute CNS injury. *Neurochem Int.* (2019) 127:22– 8. doi: 10.1016/j.neuint.2019.01.012
- Rubin T, Lipton M. Sex differences in animal models of traumatic brain injury. J Exp Neurosci. (2019) 13:117906951984402. doi: 10.1177/1179069519844020
- Wagner AK, Kline AE, Ren D, Willard LA, Wenger MK, Zafonte RD, et al. Gender associations with chronic methylphenidate treatment and behavioral performance following experimental traumatic brain injury. *Behav Brain Res.* (2007) 181:200–9. doi: 10.1016/j.bbr.2007.04.006
- Tucker LB, Fu AH, McCabe JT. Performance of male and female C57BL/6J mice on motor and cognitive tasks commonly used in preclinical traumatic brain injury research. J Neurotrauma. (2016) 33:880– 94. doi: 10.1089/neu.2015.3977

- Villapol S, Loane DJ, Burns MP. Sexual dimorphism in the inflammatory response to traumatic brain injury. *Glia.* (2017) 65:1423–38. doi: 10.1002/glia.23171
- 33. Jullienne A, Salehi A, Affeldt B, Baghchechi M, Haddad E, Avitua A, et al. Male and female mice exhibit divergent responses of the cortical vasculature to traumatic brain injury. *J Neurotrauma*. (2018) 35:1646– 58. doi: 10.1089/neu.2017.5547
- Doran SJ, Ritzel RM, Glaser EP, Henry RJ, Faden AI, Loane DJ. Sex differences in acute neuroinflammation after experimental traumatic brain injury are mediated by infiltrating myeloid cells. *J Neurotrauma*. (2019) 36:1040– 53. doi: 10.1089/neu.2018.6019
- 35. Gölz C, Kirchhoff FP, Westerhorstmann J, Schmidt M, Hirnet T, Rune GM, et al. Sex hormones modulate pathogenic processes in experimental traumatic brain injury. J Neurochem. (2019) 150:173– 87. doi: 10.1111/jnc.14678
- 36. Avcu P, Sinha S, Pang KCH, Servatius RJ. Reduced avoidance coping in male, but not in female rats, after mild traumatic brain injury: implications for depression. *Behav Brain Res.* (2019) 373:112064. doi: 10.1016/j.bbr.2019.112064
- 37. Saber M, Giordano KR, Hur Y, Ortiz JB, Morrison H, Godbout JP, et al. Acute peripheral inflammation and post-traumatic sleep differ between sexes after experimental diffuse brain injury. *Eur J Neurosci.* (2019) 52:2791– 814. doi: 10.1111/ejn.14611
- Velosky AG, Tucker LB, Fu AH, Liu J, McCabe JT. Cognitive performance of male and female C57BL/6J mice after repetitive concussive brain injuries. *Behav Brain Res.* (2017) 324:115–24. doi: 10.1016/j.bbr.2017. 02.017
- Tucker LB, Velosky AG, Fu AH, McCabe JT. Chronic neurobehavioral sex differences in a murine model of repetitive concussive brain injury. *Front Neurol.* (2019) 10:509. doi: 10.3389/fneur.2019.00509
- Skotak M, Townsend MT, Ramarao KV, Chandra N. A comprehensive review of experimental rodent models of repeated blast TBI. *Front Neurol.* (2019) 10:1015. doi: 10.3389/fneur.2019.01015
- Blennow K, Brody DL, Kochanek PM, Levin H, McKee A, Ribbers GM, et al. Traumatic brain injuries. *Nat Rev Dis Primers*. (2016) 2:16084. doi: 10.1038/nrdp.2016.84
- Giza C, Greco T, Prins ML. Concussion: pathophysiology and clinical translation. *Handb Clin Neurol.* (2018) 158:51– 61. doi: 10.1016/b978-0-444-63954-7.00006-9
- Dwyer B, Katz DI. Postconcussion syndrome. Handb Clin Neurol. (2018) 158:163–78. doi: 10.1016/b978-0-444-63954-7.00017-3
- 44. Polinder S, Cnossen MC, Real RGL, Covic A, Gorbunova A, Voormolen DC, et al. A multidimensional approach to post-concussion symptoms in mild traumatic brain injury. *Front Neurol.* (2018) 9:1113. doi: 10.3389/fneur.2018.01113
- 45. Iverson KM, Pogoda TK, Gradus JL, Street AE. Deployment-related traumatic brain injury among operation enduring freedom/operation iraqi freedom veterans: associations with mental and physical health by gender. J Womens Health (Larchmt). (2013) 22:267–75. doi: 10.1089/jwh.20 12.3755
- Yurgil KA, Barkauskas DA, Vasterling JJ, Nievergelt CM, Larson GE, Schork NJ, et al. Association between traumatic brain injury and risk of posttraumatic stress disorder in active-duty Marines. *JAMA Psychiatry*. (2014) 71:149–57. doi: 10.1001/jamapsychiatry.2013.3080
- Elder GA, Mitsis EM, Ahlers ST, Cristian A. Blast-induced mild traumatic brain injury. *Psychiatr Clin North Am.* (2010) 33:757–81. doi: 10.1016/j.psc.2010.08.001
- Michael AP, Stout J, Roskos PT, Bolzenius J, Gfeller J, Mogul D, et al. Evaluation of cortical thickness after traumatic brain injury in military veterans. J Neurotrauma. (2015) 32:1751–8. doi: 10.1089/neu.2015.3918
- Walker WC, Franke LM, McDonald SD, Sima AP, Keyser-Marcus L. Prevalence of mental health conditions after military blast exposure, their co-occurrence, and their relation to mild traumatic brain injury. *Brain Inj.* (2015) 29:1581–8. doi: 10.3109/02699052.2015.1075151
- Mac Donald CL, Barber J, Jordan M, Johnson AM, Dikmen S, Fann JR, et al. Early clinical predictors of 5-year outcome after concussive blast traumatic brain injury. *JAMA Neurol.* (2017) 74:821– 9. doi: 10.1001/jamaneurol.2017.0143

- Yeh PH, Guan Koay C, Wang B, Morissette J, Sham E, Senseney J, et al. Compromised neurocircuitry in chronic blast-related mild traumatic brain injury. *Hum Brain Mapp.* (2017) 38:352–69. doi: 10.1002/hbm.23365
- Ryan-Gonzalez C, Kimbrel NA, Meyer EC, Gordon EM, DeBeer BB, Gulliver SB, et al. Differences in post-traumatic stress disorder symptoms among post-9/11 veterans with blast- and non-blast mild traumatic brain injury. J Neurotrauma. (2019) 36:1584–90. doi: 10.1089/neu.2017.5590
- Robinson ME, McKee AC, Salat DH, Rasmusson AM, Radigan LJ, Catana C, et al. Positron emission tomography of tau in Iraq and afghanistan veterans with blast neurotrauma. *Neuroimage Clin.* (2019) 21:101651. doi: 10.1016/j.nicl.2019.101651
- Greer N, Sayer N, Koeller E, Velasquez T, Wilt TJ. Outcomes associated with blast versus nonblast-related traumatic brain injury in US military service members and veterans: a systematic review. *J Head Trauma Rehabil.* (2018) 33:E16–29. doi: 10.1097/HTR.000000000000304
- 55. Warden D. Military TBI during the Iraq and afghanistan wars. J Head Trauma Rehabil. (2006) 21:398–402. doi: 10.1097/00001199-200609000-00004
- Elder GA, Cristian A. Blast-related mild traumatic brain injury: mechanisms of injury and impact on clinical care. *Mt Sinai J Med.* (2009) 76:111– 8. doi: 10.1002/msj.20098
- 57. Davenport ND, Lim KO, Armstrong MT, Sponheim SR. Diffuse and spatially variable white matter disruptions are associated with blast-related mild traumatic brain injury. *Neuroimage*. (2012) 59:2017–24. doi: 10.1016/j.neuroimage.2011.10.050
- Levin HS, Wilde E, Troyanskaya M, Petersen NJ, Scheibel R, Newsome M, et al. Diffusion tensor imaging of mild to moderate blast-related traumatic brain injury and its sequelae. *J Neurotrauma*. (2010) 27:683– 94. doi: 10.1089/neu.2009.1073
- Mac Donald CL, Johnson AM, Cooper D, Nelson EC, Werner NJ, Shimony JS, et al. Detection of blast-related traumatic brain injury in U.S. military personnel. N Engl J Med. (2011) 364:2091–100. doi: 10.1056/NEJMoa1008069
- Bazarian JJ, Donnelly K, Peterson DR, Warner GC, Zhu T, Zhong J. The relation between posttraumatic stress disorder and mild traumatic brain injury acquired during operations enduring freedom and iraqi freedom. J Head Trauma Rehabil. (2013) 28:1–12. doi: 10.1097/HTR.0b013e318256d3d3
- Taber KH, Hurley RA, Haswell CC, Rowland JA, Hurt SD, Lamar CD, et al. White matter compromise in veterans exposed to primary blast forces. *J Head Trauma Rehabil.* (2015) 30:E15–25. doi: 10.1097/HTR.000000000000030
- Winklewski PJ, Sabisz A, Naumczyk P, Jodzio K, Szurowska E, Szarmach A. Understanding the physiopathology behind axial and radial diffusivity changes—what do we know? *Front Neurol.* (2018) 9:92. doi: 10.3389/fneur.2018.00092
- Trotter BB, Robinson ME, Milberg WP, McGlinchey RE, Salat DH. (2015) Military blast exposure, ageing and white matter integrity. *Brain.* 138(Pt 8):2278–92. doi: 10.1093/brain/awv139
- 64. Han K, Mac Donald CL, Johnson AM, Barnes Y, Wierzechowski L, Zonies D, et al. Disrupted modular organization of resting-state cortical functional connectivity in U.S. military personnel following concussive 'mild' blast-related traumatic brain injury. *Neuroimage*. (2014) 84:76– 96. doi: 10.1016/j.neuroimage.2013.08.017
- Robinson ME, Lindemer ER, Fonda JR, Milberg WP, McGlinchey RE, Salat DH. Close-range blast exposure is associated with altered functional connectivity in veterans independent of concussion symptoms at time of exposure. *Hum Brain Mapp*. (2015) 36:911–22. doi: 10.1002/hbm.22675
- 66. Fischer BL, Parsons M, Durgerian S, Reece C, Mourany L, Lowe MJ, et al. Neural activation during response inhibition differentiates blast from mechanical causes of mild to moderate traumatic brain injury. J Neurotrauma. (2014) 31:169–79. doi: 10.1089/neu.2013.2877
- Logan GD, Cowan WB, Davis KA. On the ability to inhibit simple and choice reaction time responses: a model and a method. *J Exp Psychol Hum Percept Perform.* (1984) 10:276–91. doi: 10.1037//0096-1523.10.2.276
- Aron AR, Poldrack RA. Cortical and subcortical contributions to Stop signal response inhibition: role of the subthalamic nucleus. J Neurosci. (2006) 26:2424–33. doi: 10.1523/JNEUROSCI.4682-05.2006

- Sullivan DR, Miller MW, Wolf EJ, Logue MW, Robinson ME, Fortier CB, et al. Cerebral perfusion is associated with blast exposure in military personnel without moderate or severe TBI. J Cereb Blood Flow Metab. (2020) 271678X20935190. doi: 10.1177/0271678X20935190. [Epub ahead of print].
- Koerte IK, Ertl-Wagner B, Reiser M, Zafonte R, Shenton ME. White matter integrity in the brains of professional soccer players without a symptomatic concussion. *JAMA*. (2012) 308:1859–61. doi: 10.1001/jama.2012.13735
- Lipton ML, Kim N, Zimmerman ME, Kim M, Stewart WF, Branch CA, et al. Soccer heading is associated with white matter microstructural and cognitive abnormalities. *Radiology*. (2013) 268:850–7. doi: 10.1148/radiol.13130545
- Burrelli DF. Women in combat: issues for congress. Congressional Research Service. (2013). Available online at: http://www.ncdsv.org/ images/CongResearchService\_WomenInCombatIssuesForCongress\_5-9-2013.pdf (accessed February 28, 2020).
- Hendricks AM, Amara J, Baker E, Charns MP, Gardner JA, Iverson KM, et al. Screening for mild traumatic brain injury in OEF-OIF deployed US military: an empirical assessment of VHA's experience. *Brain Inj.* (2013) 27:125–34. doi: 10.3109/02699052.2012.729284
- Jackson CE, Green JD, Bovin MJ, Vasterling JJ, Holowka DW, Ranganathan G, et al. Mild traumatic brain injury, PTSD, and psychosocial functioning among male and female U.S. OEF/OIF veterans. J Trauma Stress. (2016) 29:309–16. doi: 10.1002/jts.22110
- Lindquist LK, Love HC, Elbogen EB. Traumatic brain injury in Iraq and afghanistan veterans: new results from a national random sample study. J Neuropsychiatry Clin Neurosci. (2017) 29:254–9. doi: 10.1176/appi.neuropsych.16050100
- Gerber MR, Iverson KM, Dichter ME, Klap R, Latta RE. Women veterans and intimate partner violence: current state of knowledge and future directions. J Womens Health (Larchmt). (2014) 23:302– 9. doi: 10.1089/jwh.2013.4513
- Street AE, Gradus JL, Giasson HL, Vogt D, Resick PA. Gender differences among veterans deployed in support of the wars in Afghanistan and Iraq. J Gen Intern Med. (2013) 2:S556–62. doi: 10.1007/s11606-013-2333-4
- Dye JL, Eskridge SL, Tepe V, Clouser MC, Galarneau M. Characterization and comparison of combat-related injuries in women during OIF and OEF. *Mil Med.* (2016) 181:92–8. doi: 10.7205/milmed-d-15-00237
- Pugh MJ, Finley EP, Wang CP, Copeland LA, Jaramillo CA, Swan AA, et al. A retrospective cohort study of comorbidity trajectories associated with traumatic brain injury in veterans of the Iraq and Afghanistan wars. *Brain Inj.* (2016) 30:1481–90. doi: 10.1080/02699052.2016.1219055
- Kennedy JE, Lu LH, Reid MW, Leal FO, Cooper DB. Correlates of depression in U.S. military service members with a history of mild traumatic brain injury. *Mil Med.* (2019) 184:148–54. doi: 10.1093/milmed/usy321
- Landis SC, Amara SG, Asadullah K, Austin CP, Blumenstein R, Bradley EW, et al. A call for transparent reporting to optimize the predictive value of preclinical research. *Nature*. (2012) 490:187–91. doi: 10.1038/nature11556
- 82. McNutt M. Reproducibility. Science. (2014) 343:229. doi: 10.1126/science.1250475
- Steward O, Balice-Gordon R. Rigor or mortis: best practices for preclinical research in neuroscience. *Neuron.* (2014) 84:572-81. doi: 10.1016/j.neuron.2014.10.042
- Becker JB, Arnold AP, Berkley KJ, Blaustein JD, Eckel LA, Hampson E, et al. Strategies and methods for research on sex differences in brain and behavior. *Endocrinology*. (2005) 146:1650–73. doi: 10.1210/en.2004-1142
- Miller VM, Kaplan JR, Schork NJ, Ouyang P, Berga SL, Wenger NK, et al. Strategies and methods to study sex differences in cardiovascular structure and function: a guide for basic scientists. *Biol Sex Differ*. (2011) 2:14. doi: 10.1186/2042-6410-2-14
- McCarthy MM. Multifaceted origins of sex differences in the brain. *Philos Trans R Soc Lond B Biol Sci.* (2016) 371:20150106. doi: 10.1098/rstb.2015.0106
- Clayton JA. Applying the new SABV (sex as a biological variable) policy to research and clinical care. *Physiol Behav.* (2018) 187:2– 5. doi: 10.1016/j.physbeh.2017.08.012
- Mihalik JP, Ondrak KS, Guskiewicz KM, McMurray RG. The effects of menstrual cycle phase on clinical measures of concussion in healthy college-aged females. J Sci Med Sport. (2009) 12:383–7. doi: 10.1016/j.jsams.2008.05.003

- Maghool F, Khaksari M, Siahposht Khachki A. Differences in brain edema and intracranial pressure following traumatic brain injury across the estrous cycle: involvement of female sex steroid hormones. *Brain Res.* (2013) 1497:61–72. doi: 10.1016/j.brainres.2012.12.014
- 90. Wunderle K, Hoeger KM, Wasserman E, Bazarian JJ. Menstrual phase as predictor of outcome after mild traumatic brain injury in women. *J Head Trauma Rehabil.* (2014) 29:E1–8. doi: 10.1097/HTR.00000000000006
- Becker JB, Robinson TE, Lorenz KA. Sex differences and estrous cycle variations in amphetamine-elicited rotational behavior. *Eur J Pharmacol.* (1982) 80:65–72. doi: 10.1016/0014-2999(82)90178-9
- Prendergast BJ, Onishi KG, Zucker I. Female mice liberated for inclusion in neuroscience and biomedical research. *Neurosci Biobehav Rev.* (2014) 40:1–5. doi: 10.1016/j.neubiorev.2014.01.001
- Becker JB, Prendergast BJ, Liang JW. Female rats are not more variable than male rats: a meta-analysis of neuroscience studies. *Biol Sex Differ*. (2016) 7:34. doi: 10.1186/s13293-016-0087-5
- 94. Shansky RM. Are hormones a "female problem" for animal research? *Science*. (2019) 364:825–6. doi: 10.1126/science.aaw7570
- Galea LA. Gonadal hormone modulation of neurogenesis in the dentate gyrus of adult male and female rodents. *Brain Res Rev.* (2008) 57:332– 41. doi: 10.1016/j.brainresrev.2007.05.008
- Smith RP, Coward RM, Kovac JR, Lipshultz LI. The evidence for seasonal variations of testosterone in men. *Maturitas*. (2013) 74:208– 12. doi: 10.1016/j.maturitas.2012.12.003
- 97. Brush SG. Nettie Stevens M, and the discovery of sex determination by chromosomes. *Isis.* (1978) 69:163–72. doi: 10.1086/352001
- Cox KH, Bonthuis PJ, Rissman EF. Mouse model systems to study sex chromosome genes and behavior: relevance to humans. Front Neuroendocrinol. (2014) 35:405–19. doi: 10.1016/j.yfrne.2013.12.004
- 99. Snell DM, Turner JMA. Sex chromosome effects on male-female differences in mammals. *Curr Biol.* (2018) 28:R1313–24. doi: 10.1016/j.cub.2018.09.018
- 100. De Vries GJ, Rissman EF, Simerly RB, Yang LY, Scordalakes EM, Auger CJ, et al. A model system for study of sex chromosome effects on sexually dimorphic neural and behavioral traits. J Neurosci. (2002) 22:9005–14. doi: 10.1523/JNEUROSCI.22-20-09005.2002
- 101. Itoh Y, Mackie R, Kampf K, Domadia S, Brown JD, O'Neill R, et al. Four core genotypes mouse model: localization of the Sry transgene and bioassay for testicular hormone levels. *BMC Res Notes.* (2015) 8:69. doi: 10.1186/s13104-015-0986-2
- 102. Li J, Chen X, McClusky R, Ruiz-Sundstrom M, Itoh Y, Umar S, et al. The number of X chromosomes influences protection from cardiac ischaemia/reperfusion injury in mice: one X is better than two. *Cardiovasc Res.* (2014) 102:375–84. doi: 10.1093/cvr/cvu064
- 103. Arnold AP, van Nas A, Lusis AJ. Systems biology asks new questions about sex differences. *Trends Endocrinol Metab.* (2009) 20:471–6. doi: 10.1016/j.tem.2009.06.007
- 104. Liu S, Seidlitz J, Blumenthal JD, Clasen LS, Raznahan A. Integrative structural, functional, and transcriptomic analyses of sex-biased brain organization in humans. *Proc Natl Acad Sci U S A*. (2020) 117:18788– 98. doi: 10.1073/pnas.1919091117
- Bryant DN, Sheldahl LC, Marriott LK, Shapiro RA, Dorsa DM. Multiple pathways transmit neuroprotective effects of gonadal steroids. *Endocrine*. (2006) 29:199–207. doi: 10.1385/ENDO:29:2:199
- Brown CM, Suzuki S, Jelks KA, Wise PM. Estradiol is a potent protective, restorative, and trophic factor after brain injury. *Semin Reprod Med.* (2009) 27:240–9. doi: 10.1055/s-0029-1216277
- 107. Deutsch ER, Espinoza TR, Atif F, Woodall E, Kaylor J, Wright DW. Progesterone's role in neuroprotection, a review of the evidence. *Brain Res.* (2013) 1530:82–105. doi: 10.1016/j.brainres.2013.07.014
- Siddiqui AN, Siddiqui N, Khan RA, Kalam A, Jabir NR, Kamal MA, et al. Neuroprotective role of steroidal sex hormones: an overview. CNS Neurosci Ther. (2016) 22:342–50. doi: 10.1111/cns.12538
- 109. El Amki M, Binder N, Steffen R, Schneider H, Luft AR, Weller M, et al. Contraceptive drugs mitigate experimental stroke-induced brain injury. *Cardiovasc Res.* (2019) 115:637–46. doi: 10.1093/cvr/cvy248
- 110. Dubal DB, Zhu H, Yu J, Rau SW, Shughrue PJ, Merchenthaler I, et al. Estrogen receptor alpha, not beta, is a critical link in estradiol-mediated

protection against brain injury. Proc Natl Acad Sci U S A. (2001) 98:1952–7. doi: 10.1073/pnas.041483198

- Masel BE, DeWitt DS. Traumatic brain injury: a disease process, not an event. J Neurotrauma. (2010) 27:1529–40. doi: 10.1089/neu.2010.1358
- 112. McDonald SJ, Sharkey JM, Sun M, Kaukas LM, Shultz SR, Turner RJ, et al. Beyond the brain: peripheral interactions after traumatic brain injury. J Neurotrauma. (2020) 37:770–81. doi: 10.1089/neu.2019.6885
- 113. Kessler RC, McGonagle KA, Zhao S, Nelson CB, Hughes M, Eshleman S, et al. Lifetime and 12-month prevalence of DSM-III-R psychiatric disorders in the United States. Results from the national comorbidity survey. *Arch Gen Psychiatry*. (1994) 51:8–19. doi: 10.1001/archpsyc.1994.03950010008002
- Altemus M, Sarvaiya N, Neill Epperson C. Sex differences in anxiety and depression clinical perspectives. *Front Neuroendocrinol.* (2014) 35:320– 30. doi: 10.1016/j.yfrne.2014.05.004
- 115. Levin HS, Brown SA, Song JX, McCauley SR, Boake C, Contant CF, et al. Depression and posttraumatic stress disorder at three months after mild to moderate traumatic brain injury. *J Clin Exp Neuropsychol.* (2001) 23:754– 69. doi: 10.1076/jcen.23.6.754.1021
- Bay E, Sikorskii A, Saint-Arnault D. Sex differences in depressive symptoms and their correlates after mild-to-moderate traumatic brain injury. *J Neurosci Nurs*. (2009) 41:298–309. doi: 10.1097/jnn.0b013e3181b6be81
- 117. Ahman S, Saveman BI, Styrke J, Bjornstig U, Stalnacke BM. Long-term follow-up of patients with mild traumatic brain injury: a mixed-method study. J Rehabil Med. (2013) 45:758–64. doi: 10.2340/16501977-1182
- 118. Gabrys RL, Dixon K, Holahan MR, Anisman H. Self-reported mild traumatic brain injuries in relation to rumination and depressive symptoms: moderating role of sex differences and a brain-derived neurotrophic factor gene polymorphism. *Clin J Sport Med.* (2019) 29:494– 9. doi: 10.1097/jsm.00000000000550
- 119. Lieberman SA, Oberoi AL, Gilkison CR, Masel BE, Urban RJ. Prevalence of neuroendocrine dysfunction in patients recovering from traumatic brain injury. J Clin Endocrinol Metab. (2001) 86:2752–6. doi: 10.1210/jcem.86.6.7592
- 120. Aimaretti G, Ambrosio MR, Di Somma C, Fusco A, Cannavo S, Gasperi M, et al. Traumatic brain injury and subarachnoid haemorrhage are conditions at high risk for hypopituitarism: screening study at 3 months after the brain injury. *Clin Endocrinol (Oxf)*. (2004) 61:320– 26. doi: 10.1111/j.1365-2265.2004.02094.x
- Hoffman AN, Taylor AN. Stress reactivity after traumatic brain injury: implications for comorbid post-traumatic stress disorder. *Behav Pharmacol.* (2019) 30:115–21. doi: 10.1097/fbp.000000000000461
- Tapp ZM, Godbout JP, Kokiko-Cochran ON. A tilted axis: maladaptive inflammation and HPA axis dysfunction contribute to consequences of TBI. *Front Neurol.* (2019) 10:345. doi: 10.3389/fneur.2019.00345
- Srinivasan S, Shariff M, Bartlett SE. The role of the glucocorticoids in developing resilience to stress and addiction. *Front Psychiatry.* (2013) 4:68. doi: 10.3389/fpsyt.2013.00068
- 124. Zhou D, Zhao Y, Wan Y, Wang Y, Xie D, Lu Q, et al. Neuroendocrine dysfunction and insomniain in mild traumatic brain injury patients. *Neurosci Lett.* (2016) 610:154–9. doi: 10.1016/j.neulet.2015.10.055
- 125. Glynn N, Agha A. The frequency and the diagnosis of pituitary dysfunction after traumatic brain injury. *Pituitary*. (2019) 22:249– 60. doi: 10.1007/s11102-019-00938-y
- 126. Sav A, Rotondo F, Syro LV, Serna CA, Kovacs K. Pituitary pathology in traumatic brain injury: a review. *Pituitary*. (2019) 22:201–11. doi: 10.1007/s11102-019-00958-8
- 127. Wilkinson CW, Pagulayan KF, Petrie EC, Mayer CL, Colasurdo EA, Shofer JB, et al. High prevalence of chronic pituitary and target-organ hormone abnormalities after blast-related mild traumatic brain injury. *Front Neurol.* (2012) 3:11. doi: 10.3389/fneur.2012.00011
- Baxter D, Sharp DJ, Feeney C, Papadopoulou D, Ham TE, Jilka S, et al. Pituitary dysfunction after blast traumatic brain injury: the UK BIOSAP study. Ann Neurol. (2013) 74:527–36. doi: 10.1002/ana.23958
- 129. Undurti A, Colasurdo EA, Sikkema CL, Schultz JS, Peskind ER, Pagulayan KF, et al. Chronic hypopituitarism associated with increased postconcussive symptoms is prevalent after blast-induced mild traumatic brain injury. *Front Neurol.* (2018) 9:72. doi: 10.3389/fneur.2018.00072

- 130. Roe SY, McGowan EM, Rothwell NJ. Evidence for the involvement of corticotrophin-releasing hormone in the pathogenesis of traumatic brain injury. *Eur J Neurosci.* (1998) 10:553–9. doi: 10.1046/j.1460-9568.1998.00064.x
- 131. Grundy PL, Harbuz MS, Jessop DS, Lightman SL, Sharples PM. The hypothalamo-pituitary-adrenal axis response to experimental traumatic brain injury. J Neurotrauma. (2001) 18:1373–81. doi: 10.1089/08977150152725669
- McCullers DL, Sullivan PG, Scheff SW, Herman JP. Traumatic brain injury regulates adrenocorticosteroid receptor mRNA levels in rat hippocampus. *Brain Res.* (2002) 947:41–9. doi: 10.1016/s0006-8993(02)02904-9
- 133. Taylor AN, Rahman SU, Sanders NC, Tio DL, Prolo P, Sutton RL. Injury severity differentially affects short- and long-term neuroendocrine outcomes of traumatic brain injury. J Neurotrauma. (2008) 25:311– 23. doi: 10.1089/neu.2007.0486
- 134. Osterstock G, El Yandouzi T, Romano N, Carmignac D, Langlet F, Coutry N, et al. Sustained alterations of hypothalamic tanycytes during posttraumatic hypopituitarism in male mice. *Endocrinology.* (2014) 155:1887–98. doi: 10.1210/en.2013-1336
- Rowe RK, Rumney BM, May HG, Permana P, Adelson PD, Harman SM, et al. Diffuse traumatic brain injury affects chronic corticosterone function in the rat. *Endocr Connect.* (2016) 5:152–66. doi: 10.1530/EC-16-0031
- 136. Russell AL, Richardson MR, Bauman BM, Hernandez IM, Saperstein S, Handa RJ, et al. Differential responses of the HPA axis to mild blast traumatic brain injury in male and female mice. *Endocrinology*. (2018) 159:2363– 75. doi: 10.1210/en.2018-00203
- Malkesman O, Tucker LB, Ozl J, McCabe JT. Traumatic brain injury - modeling neuropsychiatric symptoms in rodents. *Front Neurol.* (2013) 4:157. doi: 10.3389/fneur.2013.00157
- Tucker LB, Burke JF, Fu AH, McCabe JT. Neuropsychiatric symptom modeling in male and female C57BL/6J mice after experimental traumatic brain injury. J Neurotrauma. (2017) 34:890–905. doi: 10.1089/neu.2016.4508
- 139. VandeVord PJ, Sajja VS, Ereifej E, Hermundstad A, Mao S, Hadden TJ. Chronic hormonal imbalance and adipose redistribution is associated with hypothalamic neuropathology following blast exposure. J Neurotrauma. (2016) 33:82–8. doi: 10.1089/neu.2014.3786
- 140. Zuckerman A, Ram O, Ifergane G, Matar MA, Kaplan Z, Hoffman JR, et al. Role of endogenous and exogenous corticosterone on behavioral and cognitive responses to low-pressure blast wave exposure. *J Neurotrauma*. (2019) 36:380–94. doi: 10.1089/neu.2018.5672
- 141. Russell AL, Handa RJ, Wu TJ. Sex-dependent effects of mild blast-induced traumatic brain injury on corticotropin-releasing factor receptor gene expression: potential link to anxiety-like behaviors. *Neuroscience*. (2018) 392:1–12. doi: 10.1016/j.neuroscience.2018.09.014
- 142. Henckens M, Printz Y, Shamgar U, Dine J, Lebow M, Drori Y, et al. CRF receptor type 2 neurons in the posterior bed nucleus of the stria terminalis critically contribute to stress recovery. *Mol Psychiatry.* (2017) 22:1691–700. doi: 10.1038/mp.2016.133
- Deussing JM, Chen A. The corticotropin-releasing factor family: physiology of the stress response. *Physiol Rev.* (2018) 98:2225–86. doi: 10.1152/physrev.00042.2017
- 144. Bale TL, Contarino A, Smith GW, Chan R, Gold LH, Sawchenko PE, et al. Mice deficient for corticotropin-releasing hormone receptor-2 display anxiety-like behaviour and are hypersensitive to stress. *Nat Genet.* (2000) 24:410–4. doi: 10.1038/74263
- 145. Perkes I, Baguley IJ, Nott MT, Menon DK. A review of paroxysmal sympathetic hyperactivity after acquired brain injury. Ann Neurol. (2010) 68:126–35. doi: 10.1002/ana.22066
- 146. Meyfroidt G, Baguley IJ, Menon DK. Paroxysmal sympathetic hyperactivity: the storm after acute brain injury. *Lancet Neurol.* (2017) 16:721– 9. doi: 10.1016/s1474-4422(17)30259-4
- 147. Hasen M, Almojuela A, Zeiler FA. Autonomic dysfunction and associations with functional and neurophysiological outcome in moderate/severe traumatic brain injury: a scoping review. J Neurotrauma. (2019) 36:1491– 504. doi: 10.1089/neu.2018.6073
- Baguley IJ, Heriseanu RE, Cameron ID, Nott MT, Slewa-Younan S. A critical review of the pathophysiology of dysautonomia following traumatic brain injury. *Neurocrit Care.* (2008) 8:293–300. doi: 10.1007/s12028-007-9021-3

- 149. Fernandez-Ortega JF, Prieto-Palomino MA, Garcia-Caballero M, Galeas-Lopez JL, Quesada-Garcia G, and Baguley IJ. Paroxysmal sympathetic hyperactivity after traumatic brain injury: clinical and prognostic implications. J Neurotrauma. (2012) 29:1364–70. doi: 10.1089/neu.2011.2033
- Mackenzie IM, Tunnicliffe B. Blast injuries to the lung: epidemiology and management. *Philos Trans R Soc Lond B Biol Sci.* (2011) 366:295– 9. doi: 10.1098/rstb.2010.0252
- 151. Kirkman E, Watts S, Sapford W, Sawdon M. Effects of blast injury on the autonomic nervous system and the response to resuscitation. In: Elsayed NM, Atkins JL, editors. *Explosion and Blast-Related Injuries*. Burlington, MA: Elsevier Academic Press (2008). p. 105–42.
- Kirkman E, Watts S. Characterization of the response to primary blast injury. *Philos Trans R Soc Lond B Biol Sci.* (2011) 366:286– 90. doi: 10.1098/rstb.2010.0249
- 153. Gorbunov NK, Asher LV, Elsayed NM, Atkins JL. Inflammatory response in primary blast injury. In: Elsayed NM, Atkins JL, editors. *Explosion and Blast-Related Injuries*. Burlington, MA: Elsevier Academic Press (2008). p. 298–303.
- Hilz MJ, DeFina PA, Anders S, Koehn J, Lang CJ, Pauli E, et al. Frequency analysis unveils cardiac autonomic dysfunction after mild traumatic brain injury. J Neurotrauma. (2011) 28:1727–38. doi: 10.1089/neu.2010.1497
- Pothineni NV, Shirazi LF, Mehta JL. Gender differences in autonomic control of the cardiovascular system. *Curr Pharm Des.* (2016) 22:3829– 34. doi: 10.2174/1381612822666160518125810
- 156. Bishop SA, Dech RT, Guzik P, Neary JP. Heart rate variability and implication for sport concussion. *Clin Physiol Funct Imaging*. (2018) 38:733– 42. doi: 10.1111/cpf.12487
- La Fountaine MF, Heffernan KS, Gossett JD, Bauman WA, De Meersman RE. Transient suppression of heart rate complexity in concussed athletes. *Auton Neurosci.* (2009) 148:101–3. doi: 10.1016/j.autneu.2009.03.001
- Senthinathan A, Mainwaring LM, Hutchison M. Heart rate variability of athletes across concussion recovery milestones: a preliminary study. *Clin J* Sport Med. (2017) 27:288–95. doi: 10.1097/JSM.00000000000337
- 159. Abhishekh HA, Nisarga P, Kisan R, Meghana A, Chandran S, Trichur R, et al. Influence of age and gender on autonomic regulation of heart. J Clin Monit Comput. (2013) 27:259–64. doi: 10.1007/s10877-012-9424-3
- 160. Saleh MC, Connell BJ, Saleh TM. Medullary and intrathecal injections of 17β-estradiol in male rats. *Brain Res.* (2000) 867:200–9. doi: 10.1016/s0006-8993(00)02313-1
- 161. Saleh TM, Connell BJ, Saleh MC. Acute injection of 17β-estradiol enhances cardiovascular reflexes and autonomic tone in ovariectomized female rats. Auton Neurosci. (2000) 84:78–88. doi: 10.1016/s1566-0702(00) 00196-x
- 162. Cernak I. Blast injuries and blast-induced neurotrauma: overview of pathophysiology and experimental knowledge models and findings. In: Kobeissy FH, editor. Brain Neurotrauma: Molecular, Neuropsychological, Rehabilitation Aspects. Boca Raton, FL (2015), p. 631–44.
- 163. Cernak I. Blast-induced neurotrauma. In: Winn RH, editor. Youmans and Winn Neurological Surgery. 7th ed. Philadelphia, PA: Elsevier (2017). p. 2933–42.
- Gelman S. Venous function and central venous pressure: a physiologic story. Anesthesiology. (2008) 108:735–48. doi: 10.1097/ALN.0b013e3181672607
- 165. Mitchell MJ, Lin KS, King MR. Fluid shear stress increases neutrophil activation via platelet-activating factor. *Biophys J.* (2014) 106:2243– 53. doi: 10.1016/j.bpj.2014.04.001
- 166. Wang JM, Chen J. Damage of vascular endothelial barrier induced by explosive blast and its clinical significance. *Chin J Traumatol.* (2016) 19:125– 8. doi: 10.1016/j.cjtee.2016.03.001
- 167. Simard JM, Pampori A, Keledjian K, Tosun C, Schwartzbauer G, Ivanova S, et al. Exposure of the thorax to a sublethal blast wave causes a hydrodynamic pulse that leads to perivenular inflammation in the brain. *J Neurotrauma*. (2014) 31:1292–304. doi: 10.1089/neu.2013.3016
- 168. Gama Sosa MA, De Gasperi R, Paulino AJ, Pricop PE, Shaughness MC, Maudlin-Jeronimo E, et al. Blast overpressure induces shear-related injuries in the brain of rats exposed to a mild traumatic brain injury. *Acta Neuropathol Commun.* (2013) 1:51. doi: 10.1186/2051-5960-1-51
- Gama Sosa MA, De Gasperi R, Janssen PL, Yuk FJ, Anazodo PC, Pricop PE, et al. Selective vulnerability of the cerebral vasculature to blast injury in a

rat model of mild traumatic brain injury. *Acta Neuropathol Commun.* (2014) 2:67. doi: 10.1186/2051-5960-2-67

- 170. Gama Sosa MA, De Gasperi R, Perez Garcia GS, Perez GM, Searcy C, Vargas D, et al. Low-level blast exposure disrupts gliovascular and neurovascular connections and induces a chronic vascular pathology in rat brain. Acta Neuropathol Commun. (2019) 7:6. doi: 10.1186/s40478-018-0647-5
- 171. Ling G, Bandak F, Armonda R, Grant G, Ecklund J. Explosive blast neurotrauma. J Neurotrauma. (2009) 26:815–25. doi: 10.1089/neu.2007.0484
- 172. Armonda RA, Bell RS, Vo AH, Ling G, DeGraba TJ, Crandall B, et al. Wartime traumatic cerebral vasospasm: recent review of combat casualties. *Neurosurgery.* (2006) 59:1215– 25. doi: 10.1227/01.NEU.0000249190.46033.94
- 173. Czosnyka M, Radolovich D, Balestreri M, Lavinio A, Hutchinson P, Timofeev I, et al. Gender-related differences in intracranial hypertension and outcome after traumatic brain injury. *Acta Neurochir Suppl.* (2008) 102:25–8. doi: 10.1007/978-3-211-85578-2\_5
- 174. Sorrentino E, Diedler J, Kasprowicz M, Budohoski KP, Haubrich C, Smielewski P, et al. Critical thresholds for cerebrovascular reactivity after traumatic brain injury. *Neurocrit Care.* (2012) 16:258–66. doi: 10.1007/s12028-011-9630-8
- 175. Hamer J, Churchill NW, Hutchison MG, Graham SJ, Schweizer TA. Sex differences in cerebral blood flow associated with a history of concussion. *J Neurotrauma*. (2020) 37:1197–203. doi: 10.1089/neu.2019.6800
- 176. Mollayeva T, Mollayeva S, Colantonio A. Traumatic brain injury: sex, gender and intersecting vulnerabilities. *Nat Rev Neurol.* (2018) 14:711– 22. doi: 10.1038/s41582-018-0091-y
- 177. Armstead WM, Kiessling JW, Kofke WA, Vavilala MS. SNP improves cerebral hemodynamics during normotension but fails to prevent sex dependent impaired cerebral autoregulation during hypotension after brain injury. *Brain Res.* (2010) 1330:142–50. doi: 10.1016/j.brainres.2010. 03.024
- 178. Cernak I, Savic J, Ignjatovic D, Jevtic M. Blast injury from explosive munitions. J Trauma Acute Care Surg. (1999) 47:96–103.
- 179. Esser-von Bieren J. Immune-regulation and -functions of eicosanoid lipid mediators. *Biol Chem.* (2017) 398:1177–91. doi: 10.1515/hsz-2017-0146
- Cernak I. The importance of systemic response in the pathobiology of blast-induced neurotrauma. *Front Neurol.* (2010) 1:151. doi: 10.3389/fneur.2010.00151
- 181. Toklu HZ, Yang Z, Oktay S, Sakarya Y, Kirichenko N, Matheny MK, et al. Overpressure blast injury-induced oxidative stress and neuroinflammation response in rat frontal cortex and cerebellum. *Behav Brain Res.* (2018) 340:14–22. doi: 10.1016/j.bbr.2017.04.025
- 182. Johnson VE, Stewart JE, Begbie FD, Trojanowski JQ, Smith DH, Stewart W. Inflammation and white matter degeneration persist for years after a single traumatic brain injury. *Brain.* (2013) 136(Pt 1):28–42. doi: 10.1093/brain/aws322
- 183. Kwon SK, Kovesdi E, Gyorgy AB, Wingo D, Kamnaksh A, Walker J, et al. Stress and traumatic brain injury: a behavioral, proteomics, histological study. *Front Neurol.* (2011) 2:12. doi: 10.3389/fneur.2011.00012
- 184. Goldstein LE, Fisher AM, Tagge CA, Zhang XL, Velisek L, Sullivan JA, et al. Chronic traumatic encephalopathy in blast-exposed military veterans and a blast neurotrauma mouse model. *Sci Transl Med.* (2012) 4:134ra160. doi: 10.1126/scitranslmed.3003716
- 185. Cho HJ, Sajja VS, Vandevord PJ, Lee YW. Blast induces oxidative stress, inflammation, neuronal loss and subsequent short-term memory impairment in rats. *Neuroscience*. (2013) 253:9–20. doi: 10.1016/j.neuroscience.2013.08.037
- Sajja VS, Ereifej ES, VandeVord PJ. Hippocampal vulnerability and subacute response following varied blast magnitudes. *Neurosci Lett.* (2014) 570:33– 7. doi: 10.1016/j.neulet.2014.03.072
- 187. Sajja VS, Hubbard WB, VandeVord PJ. Subacute oxidative stress and glial reactivity in the amygdala are associated with increased anxiety following blast neurotrauma. *Shock.* (2015) 1:71–8. doi: 10.1097/SHK.000000000000311
- Huber BR, Meabon JS, Hoffer ZS, Zhang J, Hoekstra JG, Pagulayan KF, et al. Blast exposure causes dynamic microglial/macrophage responses and microdomains of brain microvessel dysfunction. *Neuroscience*. (2016) 319:206–20. doi: 10.1016/j.neuroscience.2016.01.022

- 189. Cernak I, Merkle AC, Koliatsos VE, Bilik JM, Luong QT, Mahota TM, et al. The pathobiology of blast injuries and blast-induced neurotrauma as identified using a new experimental model of injury in mice. *Neurobiol Dis.* (2011) 41:538–51. doi: 10.1016/j.nbd.2010.10.025
- 190. Svetlov SI, Prima V, Glushakova O, Svetlov A, Kirk DR, Gutierrez H, et al. Neuro-glial and systemic mechanisms of pathological responses in rat models of primary blast overpressure compared to "composite" blast. *Front Neurol.* (2012) 3:15. doi: 10.3389/fneur.2012.00015
- 191. Sajja VS, Perrine SA, Ghoddoussi F, Hall CS, Galloway MP, VandeVord PJ. Blast neurotrauma impairs working memory and disrupts prefrontal myo-inositol levels in rats. *Mol Cell Neurosci.* (2014) 59:119–26. doi: 10.1016/j.mcn.2014.02.004
- 192. Tompkins P, Tesiram Y, Lerner M, Gonzalez LP, Lightfoot S, Rabb CH, et al. Brain injury: neuro-inflammation, cognitive deficit, and magnetic resonance imaging in a model of blast induced traumatic brain injury. *J Neurotrauma*. (2013) 30:1888–97. doi: 10.1089/neu.2012.2674
- 193. Valiyaveettil M, Alamneh Y, Wang Y, Arun P, Oguntayo S, Wei Y, et al. Contribution of systemic factors in the pathophysiology of repeated blast-induced neurotrauma. *Neurosci Lett.* (2013) 539:1–6. doi: 10.1016/j.neulet.2013.01.028
- 194. Valiyaveettil M, Alamneh YA, Miller SA, Hammamieh R, Arun P, Wang Y, et al. Modulation of cholinergic pathways and inflammatory mediators in blast-induced traumatic brain injury. *Chem Biol Interact.* (2013) 203:371– 5. doi: 10.1016/j.cbi.2012.10.022
- 195. Li Y, Yang Z, Liu B, Valdez C, Chavko M, Cancio LC. (2019) Low-level primary blast induces neuroinflammation and neurodegeneration in rats. *Mil Med.* 184 (Suppl. 1):265–72. doi: 10.1093/milmed/usy330
- 196. Kobeissy F, Mondello S, Tümer N, Toklu HZ, Whidden MA, Kirichenko N, et al. Assessing neuro-systemic & behavioral components in the pathophysiology of blast-related brain injury. *Front Neurol.* (2013) 4:186. doi: 10.3389/fneur.2013.00186
- 197. White CS. The scope of blast and shock biology and problem areas in relating physical and biological parameters. Ann N Y Acad Sci. (1968) 152:89–102. doi: 10.1111/j.1749-6632.1968.tb11969.x
- Needham CE, Ritzel D, Rule GT, Wiri S, Young L. Blast testing issues and TBI: experimental models that lead to wrong conclusions. *Front Neurol.* (2015) 6:72. doi: 10.3389/fneur.2015.00072
- Cernak I. Understanding blast-induced neurotrauma: how far have we come? Concussion. (2017) 2:CNC42. doi: 10.2217/cnc-2017-0006
- Pearce AP, Clasper J. Improving survivability from blast injury: 'shifting the goalposts' and the need for interdisciplinary research. J R Army Med Corps. (2019) 165:5–6. doi: 10.1136/jramc-2018-000968
- Casscells SW. Traumatic Brain Injury: Definition and Reporting. HA Department of Defense: Washington, DC (2007). Available online at: http:// mhs.osd.mil/Content/docs/pdfs/policies/2007/07-030.pdf
- 202. de Candole CA. Blast injury. Can Med Assoc J. (1967) 96:207-14.
- Cernak I, Noble-Haeusslein LJ. Traumatic brain injury: an overview of pathobiology with emphasis on military populations. J Cereb Blood Flow Metab. (2010) 30:255–66. doi: 10.1038/jcbfm.2009.203
- Westrol MS, Donovan CM, Kapitanyan R. Blast physics and pathophysiology of explosive injuries. *Ann Emerg Med.* (2017) 69:S4–9. doi: 10.1016/j.annemergmed.2016.09.005
- 205. Vu PA, Tucker LB, Liu J, McNamara EH, Tran T, Fu AH, et al. Transient disruption of mouse home cage activities and assessment of orexin immunoreactivity following concussive- or blast-induced brain injury. *Brain Res.* (2018) 1700:138–51. doi: 10.1016/j.brainres.2018.08.034
- 206. VandeVord PJ, Dal Cengio Leonardi A, Ritzel D. Bridging the gap of standardized animals models for blast neurotrauma: methodology for appropriate experimental testing. In: Kobeissy FH, Dixon CE, Hayes RL, Mondello S, editors. *Injury Models of the Central Nervous System*. Boca Raton: CRC Press/Taylor & Francis (2016). p. 101–18.
- 207. Cernak I. Utilization of shock tubes in blast injury research. In: Risling M, Davidsson J, editors. *Animal Models of Neurotrauma*. New York, NY: Spring Science+Business Media, LLC (2019). p. 93–115.
- Ritzel DV, Parks SA, Roseveare J, Rude G, Sawyer TW. Experimental blast simulation for injury studies. In: *RTO Human Factors and Medicine Panel (HFM) Symposium*. Halifax, Canada: NATO Science and Technology Organization (2011), 11-1–11-20.

- Philippens MMGM, Ouellet S. Introduction to blast in the context of blast-induced TBI. In: Risling M, Davidsson J, editors. *Animal Models of Neurotrauma*. New York, NY: Spring Science+Business Media, LLC (2019). p. 117–50.
- Champion HR, Holcomb JB, Young LA. Injuries from explosions: physics, biophysics, pathology, and required research focus. J Trauma. (2009) 66:1468–77. doi: 10.1097/TA.0b013e3181a27e7f
- 211. Bolander R, Mathie B, Bir C, Ritzel D, VandeVord P. Skull flexure as a contributing factor in the mechanism of injury in the rat when exposed to a shock wave. *Ann Biomed Eng.* (2011) 39:2550– 9. doi: 10.1007/s10439-011-0343-0
- Lillie EM, Urban JE, Lynch SK, Weaver AA, Stitzel JD. Evaluation of skull cortical thickness changes with age and sex from computed tomography scans. J Bone Miner Res. (2016) 31:299–307. doi: 10.1002/jbmr.2613
- 213. Garimella HT, Kraft RH, Przekwas AJ. Do blast induced skull flexures result in axonal deformation? *PLoS ONE.* (2018) 13:e0190881. doi: 10.1371/journal.pone.0190881
- 214. Jean A, Nyein MK, Zheng JQ, Moore DF, Joannopoulos JD, Radovitzky R. An animal-to-human scaling law for blast-induced traumatic brain injury risk assessment. *Proc Natl Acad Sci U S A.* (2014) 111:15310– 5. doi: 10.1073/pnas.1415743111
- Maga AM, Navarro N, Cunningham ML, Cox TC. Quantitative trait loci affecting the 3D skull shape and size in mouse and prioritization of candidate genes in-silico. *Front Physiol.* (2015) 6:92. doi: 10.3389/fphys.2015.00092
- 216. Masel BE, Bell RS, Brossart S, Grill RJ, Hayes RL, Levin HS, et al. Galveston brain injury conference 2010: clinical and experimental aspects of blast injury. J Neurotrauma. (2012) 29:2143–71. doi: 10.1089/neu.2011.2258
- Chandra N, Sundaramurthy A, Gupta RK. Validation of laboratory animal and surrogate human models in primary blast injury studies. *Mil Med.* (2017) 182:105–13. doi: 10.7205/MILMED-D-16-00144
- 218. Needham CE. Blast loads on animals. In: *Blast Waves*. 2nd ed. Cham, Switzerland: Springer International Publishing (2018). p. 383-92.
- Josey T, Ouellet S, Bieler D, Cernak I, Franke A, Gupta R, et al. Guidelines for reproducing blast exposures in the laboratory. J R Army Med Corps. (2019) 165:10–4. doi: 10.1136/jramc-2018-000954
- 220. NATO Health Factors and Medicine (HFM) Research Task Group (RTG) HFM-234. Environmental Toxicology of Blast Exposures: Injury Metrics, Modelling, Methods and Standards. Neuilly-sur-Seine Cedex, France: Science and Technology Organization, North Atlantic Treaty Organization (2018), p. 1–318.
- 221. Sawyer TW, Josey T, Wang Y, Villanueva M, Ritzel DV, Nelson P, et al. Investigations of primary blast-induced traumatic brain injury. *Shock Waves*. (2017) 28:85–99. doi: 10.1007/s00193-017-0756-2
- 222. Sawyer TW, Wang Y, Ritzel DV, Josey T, Villanueva M, Shei Y, et al. High-fidelity simulation of primary blast: direct effects on the head. *J Neurotrauma*. (2016) 33:1181–93. doi: 10.1089/neu.2015.3914
- 223. Bell MK. A standardized model is needed to study the neurological effects of primary blast wave exposure. *Mil Med.* (2008) 173:v-viii.
- 224. Brix KA, Brody DL, Grimes JB, Yitzhak A, Agoston D, Aldag M, et al. Military blast exposure and chronic neurodegeneration: summary of working groups and expert panel findings and recommendations. *J Neurotrauma*. (2017) 34:S-18–S-25. doi: 10.1089/neu.2017.5222
- 225. Cernak I, Stein DG, Elder GA, Ahlers S, Curley K, DePalma RG, et al. Preclinical modelling of militarily relevant traumatic brain injuries: challenges and recommendations for future directions. *Brain Inj.* (2017) 31:1168–76. doi: 10.1080/02699052.2016.1274779
- 226. DeWitt DS, Hawkins BE, Dixon CE, Kochanek PM, Armstead W, Bass CR, et al. Pre-clinical testing of therapies for traumatic brain injury. J Neurotrauma. (2018) 35:2737–54. doi: 10.1089/neu.2018.5778
- 227. Ma X, Aravind A, Pfister BJ, Chandra N, Haorah J. Animal models of traumatic brain injury and assessment of injury severity. *Mol Neurobiol.* (2019) 56:5332–45. doi: 10.1007/s12035-018-1454-5

- 228. Maas AI, Harrison-Felix CL, Menon D, Adelson PD, Balkin T, Bullock R, et al. Common data elements for traumatic brain injury: recommendations from the interagency working group on demographics and clinical assessment. Arch Phys Med Rehabil. (2010) 91:1641–9. doi: 10.1016/j.apmr.2010.07.232
- 229. Manley GT, Diaz-Arrastia R, Brophy M, Engel D, Goodman C, Gwinn K, et al. Common data elements for traumatic brain injury: recommendations from the biospecimens and biomarkers working group. *Arch Phys Med Rehabil.* (2010) 91:1667–72. doi: 10.1016/j.apmr.2010. 05.018
- 230. Thurmond VA, Hicks R, Gleason T, Miller AC, Szuflita N, Orman J, et al. Advancing integrated research in psychological health and traumatic brain injury: common data elements. *Arch Phys Med Rehabil.* (2010) 91:1633– 6. doi: 10.1016/j.apmr.2010.06.034
- 231. Hicks R, Giacino J, Harrison-Felix C, Manley G, Valadka A, Wilde EA. Progress in developing common data elements for traumatic brain injury research: version two-the end of the beginning. J Neurotrauma. (2013) 30:1852–61. doi: 10.1089/neu.2013.2938
- Saatman KE, Duhaime AC, Bullock R, Maas AI, Valadka A, Manley GT, et al. Classification of traumatic brain injury for targeted therapies. J Neurotrauma. (2008) 25:719–38. doi: 10.1089/neu.2008.0586
- Povlishock JT. Journal of neurotrauma. Editorial. J Neurotrauma. (2008) 25:1133. doi: 10.1089/neu.2008.9961
- 234. Smith DH, Hicks RR, Johnson VE, Bergstrom DA, Cummings DM, Noble LJ, et al. Pre-clinical traumatic brain injury common data elements: toward a common language across laboratories. J Neurotrauma. (2015) 32:1725–35. doi: 10.1089/neu.2014.3861
- 235. Rodriguez UA, Zeng Y, Deyo D, Parsley MA, Hawkins BE, Prough DS, et al. Effects of mild blast traumatic brain injury on cerebral vascular, histopathological, and behavioral outcomes in rats. *J Neurotrauma*. (2018) 35:375–92. doi: 10.1089/neu.2017.5256
- 236. Wilkinson MD, Dumontier M, Aalbersberg IJ, Appleton G, Axton M, Baak A, et al. The FAIR guiding principles for scientific data management and stewardship. *Sci Data*. (2016) 3:160018. doi: 10.1038/sdata.2016.18
- Hawkins BE, Huie JR, Almeida C, Chen J, Ferguson AR. Data dissemination: shortening the long tail of traumatic brain injury dark data. *J Neurotrauma*. (2019) doi: 10.1089/neu.2018.6192. [Epub ahead of print].
- 238. Kabitzke P, Cheng KM, Altevogt B. Guidelines and initiatives for good research practice. In: *Handbook of Experimental Pharmacology* (Berlin: Springer) (2019) p. 19–34.
- 239. Watts S, Kirkman E, Bieler D, Bjarnason S, Franke A, Gupta R, et al. Guidelines for using animal models in blast injury research. J R Army Med Corps. (2019) 165:38–40. doi: 10.1136/jramc-2018-000956
- 240. Skotak M, Alay E, Chandra N. On the accurate determination of shock wave time-pressure profile in the experimental models of blastinduced neurotrauma. *Front Neurol.* (2018) 9:52. doi: 10.3389/fneur.2018. 00052
- 241. Wright DW, Espinoza TR, Merck LH, Ratcliff JJ, Backster A, Stein DG. Gender differences in neurological emergencies part II: a consensus summary and research agenda on traumatic brain injury. *Acad Emerg Med.* (2014) 21:1414–20. doi: 10.1111/acem.12532

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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