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RESEARCH ARTICLE

Reproductive trade-offs maintain bract color polymorphism in Scarlet Indian paintbrush (*Castilleja coccinea*)

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Abstract

Populations of scarlet Indian paintbrush (Castilleja coccinea) in the Midwestern United States exhibit a bract color polymorphism, with each population having predominantly yellow or scarlet bracts. We investigated a possible mechanism for this maintenance of bract color polymorphism in C. coccinea by conducting hand-pollination experiments in two nearby populations, one predominantly yellow and one predominantly scarlet. The handpollination treatments were either self-pollination or cross pollination using pollen from within and between populations. Both color morphs were used as pollen donors for the within and between crosses. We found that both color morphs of C. coccinea were self-compatible. When the scarlet morph was the maternal plant it had higher seed set. When pollinators were excluded, the yellow morph outperformed the scarlet morph in fruit set and seed set. The apparent trade-offs between a higher reproductive output in the scarlet morph and a reproductive assurance advantage in the vellow morph may explain the maintenance of the polymorphism in C. coccinea. While many previous studies have provided evidence for pollinator preference playing a role in floral color polymorphism, the results of the current study indicate that reproductive assurance, which would be important for fluctuations in pollinator abundance or colonizing new areas, may act as a selective agent to maintain such polymorphisms.

Introduction

Polymorphisms for floral traits occur in many angiosperm species, and the underlying evolutionary forces maintaining these polymorphisms have long been the subject of interest and debate among evolutionary biologists. Floral traits reported to vary intraspecifically include corolla length and corolla flare [1], calyx length [2], flower size and style length [3], and floral color [4–6]. Among these traits, floral color polymorphisms are the most visually striking and thus have drawn many researchers to investigate the cause and maintenance of intraspecific variation [7–10]. Floral color polymorphisms vary both within [5,11–14] and between populations [4,15–17] and a variety of selective agents have been implicated in their maintenance.

Numerous studies have demonstrated that pollinators are often the primary selective agent maintaining floral color polymorphisms both within and between populations [11,18–23]. Pollinator preference and constancy may result in assortative mating, limiting gene flow between the morphs within a population [24,25]. For *Ipomoea purpurea*, pollinator constancy by bumble bees resulted in assortative mating within a population [26], while in *Clarkia xantiana*, floral color polymorphism is maintained via a combination of positive frequency-dependent pollinator preference by one bee species and negative frequency-dependent pollinator preferences by two other bee species [13]. In *Mimulus aurantiacus*, where red and yellow ecotypes inhabit different habitats, hummingbirds and hawkmoths show strong preference for red and yellow morphs, respectively, hence both pollinator preferences and ecogeographic isolation has led to assortative mating, thereby maintaining the flower color polymorphism between populations [27].

Selection by non-pollinator agents can also lead to floral color polymorphism [28,29]. Differences in seed set, seed weight, and seed predation under different environmental conditions have been documented between color morphs [4,30–32]. Anthocyanins, a primary floral pigment [33], are related to tolerance against abiotic stresses such as UV-B radiation [34], heat [35], and drought [36], as well as non-pollinator biotic pressures such as herbivore defense [6,37]. Such pleiotropic effects will interact with the pollinator community to either maintain or enforce floral color polymorphism [38].

In theory, floral color polymorphisms associated with differences in breeding system could also be maintained by selection. For example, autogamous selfing (within the same flower) provides reproductive assurance when vector-mediated cross pollination is insufficient, but the advantage is offset by pollen and/or seed discounting [39,40]. Color morphs associated with higher rates of selfing may therefore have a selective advantage when pollinators are limited but not when they are abundant. Numerous studies have demonstrated that intraspecific variation in other floral traits, such as herkogamy and protandry [41], flower size [42,43] and even scent [44] can influence reproductive assurance within and between populations. We are aware of only one report of differences in selfing rates associated with variation in flower color. In *Ipomoea purpurea*, when the relative frequency of the white morph is low compared to the darkly and lightly pigmented morphs, the white morph had higher selfing rates [26,45]. When morphs were at more similar frequencies, all three morphs had similar selfing rates, so the white morph seems to be maintained by negative frequency dependent selection on reproductive assurance [45–47].

Scarlet Indian paintbrush, Castilleja coccinea L. Sprengel (Orobanchaceae), is a hemiparasitic forb native to the Eastern United States. Showy bracts surround small, greenish flowers. Flowers are perfect with the style slightly exserted [48]. Individuals are annual or biennial and may produce multiple stems and inflorescences. A successful fertilization results in a capsule that contains an average of 150 seed. The bracts surrounding flowers display yellow or scarlet (orange-red) colors. Despite the common name, the yellow morph dominates some populations in the Midwestern United States. Populations in this region are predominantly one color or the other, with over 90% of the individuals typically having either yellow or scarlet bract colors [49]. Although the basis of bract color has not been studied, seeds collected from natural populations and sown in a common garden grew into plants exhibiting maternal bract colors, suggesting that bract color is a heritable trait [49]. Further, we have hand pollination data that indicates bract color shows simple Mendelian inheritance, with yellow dominant over scarlet (in prep). Castilleja coccinea has been reported to attract ruby-throated hummingbirds, Archi*lochus colubris* [50–54] and insect pollinators such as bees and butterflies. It is tempting to hypothesize that pollinator preference might cause positive assortative mating and thus maintains the bract color polymorphism in C. coccinea, but there are no published studies

demonstrating different rates of pollinator visitations and effectiveness to color morphs in this species.

We investigated the possible role of the breeding system in the maintenance of flower color polymorphism in *C. coccinea*. We used hand-pollination experiments at a site in northeastern Illinois (Illinois Beach State Park) where a yellow population and a scarlet population are found approximately 500 m apart. The color morphs grow on the same sandy dune-swale complex under similar abiotic conditions, and likely share a pollinator community. Our overall goal was to characterize the breeding system of the species, and to identify differences between color morphs, if they occurred. We used pollinator exclusion and hand-pollination experiments to compare the color morphs with regard to 1) self-compatibility, 2) response to pollinator exclusion, 3) cross-compatibility between the color morphs, and 4) relative female fertility and male fitness.

Materials and methods

Study populations

A hand-pollination experiment was conducted at Illinois Beach State Park from May 29th to July 6th in 2013. Two populations in Illinois Beach State Park, separated by an oak savannah and approximately 500 m apart, differ in bract color. Population 1 (hereafter, the yellow population) is predominantly yellow (87% yellow) whereas population 2 (hereafter, the scarlet population) is predominantly scarlet (99.6% scarlet) [49]. A limited pollinator observation study was conducted in both populations to determine the presence or absence of floral visitors to *C. coccinea.* A total of 24 observation sessions, each lasting 15 minutes, were conducted in the yellow population, and 17 observation sessions were conducted in the scarlet population from morning to late afternoon.

Hand-pollinations

We conducted hand-pollinations to study the breeding system of C. coccinea and to compare female fertility under different treatments. To exclude animal pollinators, we used nylon mesh bags (17.8 cm by 11.4 cm) to cover entire inflorescences for all six treatments. The pollen donors varied by bract color and population. There were six pollination treatments: 1) bagged, no hand pollination (BN); 2) self-pollination (SP); 3) same color, same population (SS); 4) same color, different populations (SD); 5) different color, same population (DS); and 6) different color, different populations (DD) (Fig 1). For self-pollination, the pollen was transferred to the stigma of the same flower (autogamous selfing). For all "same population" treatments, pollen donors were chosen at least 5 m apart from the pollen recipients to decrease chances of biparental inbreeding. Eighteen randomly selected individuals from each population were chosen as pollen recipients, providing three replicates for all six treatments. All 18 individuals were defined by the predominant color of the population, hence the "yellow morph" refers to a pollen recipient with yellow bract color from the yellow population and the "scarlet morph" refers to a pollen recipient with scarlet bract color from the scarlet population. For the treatment assigned "same color, between populations", the pollen donor was the same color as the predominate color of the recipient population but from the other population. Toothpicks and small plastic containers were used to remove and transfer pollen grains between individuals and populations. New pollen grains were collected for each day's hand-pollination. Leftover pollen grains were discarded. In addition to the experimental treatments, three open control plants were followed in each population.

Flowers that were open prior to bagging were counted and marked by threads. Hand-pollinated flowers were marked with a black permanent marker. On each visit, the number of



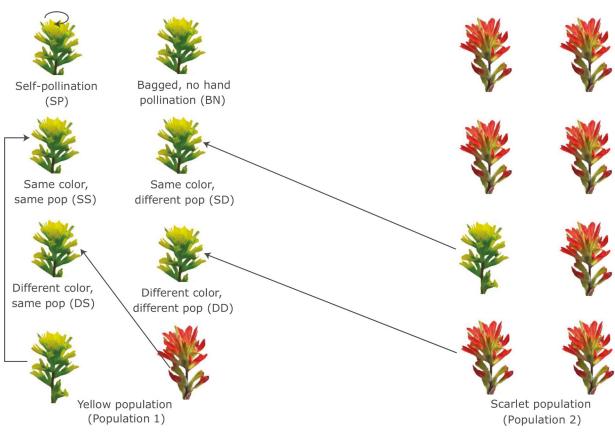


Fig 1. Illustration of six hand-pollination treatments for the yellow morph. All pollen recipients were bagged. The arrows move away from pollen donors and point to pollen recipients. The scarlet morph also received the corresponding six treatments but are not illustrated in this figure. There were three replicates for each treatment.

fertilized flowers was recorded for each individual. For the open control, the number of fertilized flowers were divided by the total number of flowers to estimate the fruit set. When individuals receiving treatments were either removed or had a broken stem, they were replaced with other individuals to keep sample size consistent. A total of 35 individuals were used for the fruit set analyses. One yellow morph that received the SS treatment had a broken stem, but it was too late in the blooming season to replace it. After collecting the fruit set data, two additional individuals were found with broken stems, one yellow morph that received the DD treatment and one yellow morph that received the SD treatment. Excluding these two, a total of 33 individuals were used for the seed set analyses. To calculate seed set we collected mature capsules and counted the number of seeds for each capsule using a Contador seed counter (Pfeuffer GmbH, Kitzingen, Germany). After the seed count, the seeds were returned to the site.

Data analysis

Two variables were used as metrics of reproductive success, fruit set (the proportion of flowers that developed into fruits) and seed set (the number of seeds per fruit). Seed set excluded flowers that did not develop into fruits. We used linear mixed-effects models (LMM) for analyses involving seed set and generalized linear mixed-effects models (GLMM) for analyses involving fruit set. For GLMM relating to fruit set, fertilization of individual flowers was modeled as a

binary (Bernoulli) response variable. Mixed-effects modeling was implemented with the *lme4* package for R version 3.5.0 [55,56]. In all mixed-effects models, we used the individual plant as the random effect (random intercept), and maternal color as one of the fixed effects.

To address our questions regarding self-compatibility, we compared the reproductive success of plants that were self-pollinated to those that were cross-pollinated with plants of the same color and the same population (i.e., the SP and SS treatments, respectively). Self-compatibility indices were calculated for each population using the average seed set where the average seed set from SP was divided by the average seed set from SS [57]. To investigate the effects of pollinator exclusion, we compared the reproductive success of plants that were bagged and not hand-pollinated to those that were self-pollinated (i.e., the BN and SP treatments). For each model we included the interaction of treatment and bract color. Thus, the maximal model had two fixed effects (maternal color, pollination treatment) with two levels each, an interaction of the fixed effects, and a random effect of the individual.

Additionally, we compared the open control treatment (unmanipulated, unbagged plants) in each population. This analysis only included maternal color as a fixed effect (same as the source population).

We investigated cross-compatibility between color morphs with regard to the relative fitness of the sexes using a larger model, that added 1) whether pollen came from the same or different population, (i.e., the "population" fixed effect), and 2) whether pollen came from individuals of the same bract color, (i.e., the "color" fixed effect), in addition to maternal color. Both of these had two levels (same or different). These two fixed effects combined to describe four of the pollination treatments described earlier (SS, SD, DS, and DD). The interactions of maternal color with each of the other two fixed effects were also included in model testing.

For each test, we compared candidate models with every combination of fixed effects using the Akaike information criterion corrected for small sample size (AICc). Akaike weights were calculated and the best model, assuming one correct model exists in the candidate set, was determined by the maximum Akaike weight value. Estimates of mean values and 95% confidence intervals for a given factor in our mixed-effects models were presented as estimated marginal means calculated using the *lsmeans* package [58].

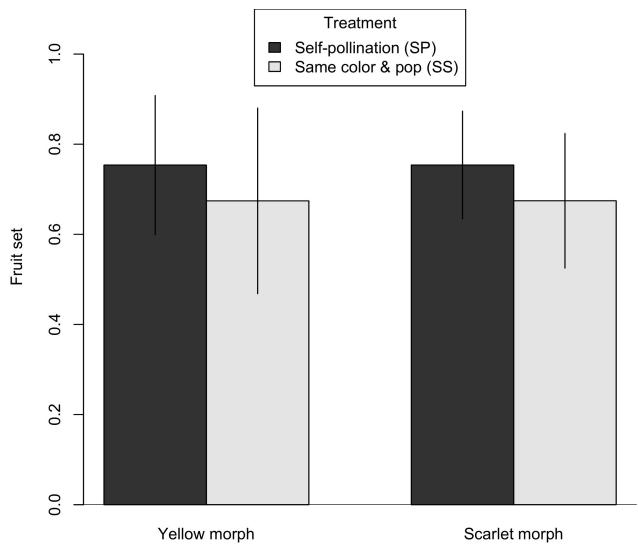
We determined the relative influence of fixed effects and random effects by comparing the marginal and conditional R², following Nakagawa and Schielzeth [59]. Calculation of the coefficients of determination was completed with the *MuMIn* package [60].

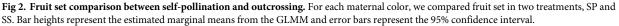
Results

Our limited pollinator observations confirmed the presence of insect floral visitors. In the yellow population, we observed black swallowtails (*Papilio polyxeneson*) and bumble bees (*Bombus* sp.) on the yellow morph. In the scarlet population, we observed only sweat bees on the scarlet morph. Bumble bees and butterflies were present in the scarlet population, but they did not visit scarlet individuals during our observation sessions. We did not observe ruby-throated hummingbirds at our study sites.

Self-compatibility

Both color morphs were self-compatible with no evidence of self-sterility. The average seed set for SP was 110.43 and 144.43, for the yellow morph and the scarlet morph, respectively. The average seed set for SS was 121.89 and 162.40, for the yellow morph and the scarlet morph, respectively. The self-compatibility indices were 0.91 for the yellow and 0.89 for the scarlet. The self-pollination (SP) treatment had similar fruit set to the same color, same population (SS) treatment (Fig 2, S1A Table). The average fruit set was 60% in all treatment-color





combinations and 70% for the self-pollination treatment (Fig 2). In our statistical models for fruit set (Table 1), the null model had the greatest support (Akaike weight = 0.447; Table 1). There was little support for a difference between treatments (Δ AICc = 1.31, Akaike weight = 0.232), or color morphs (Δ AICc = 2.11, Akaike weight = 0.155).

Seed set was about 18% lower in the self-pollination treatment (SP) compared to the same color, same population (SS) treatment (Fig 3, S1B Table), but there was little statistical support for including treatment in the best model (Table 2; for the best model with treatment, $\Delta AICc = 1.51$, Akaike weight = 0.198). There was a more pronounced difference between color morphs, with scarlet individuals having 33% to 42% greater seed set for the cross-pollination (SS) and self-pollination (SP) treatments, respectively. The best model did include maternal color as a fixed effect (Akaike weight = 0.423), while the null model excluding maternal color and treatment was the next best model ($\Delta AICc = 1.37$, Akaike weight = 0.213).

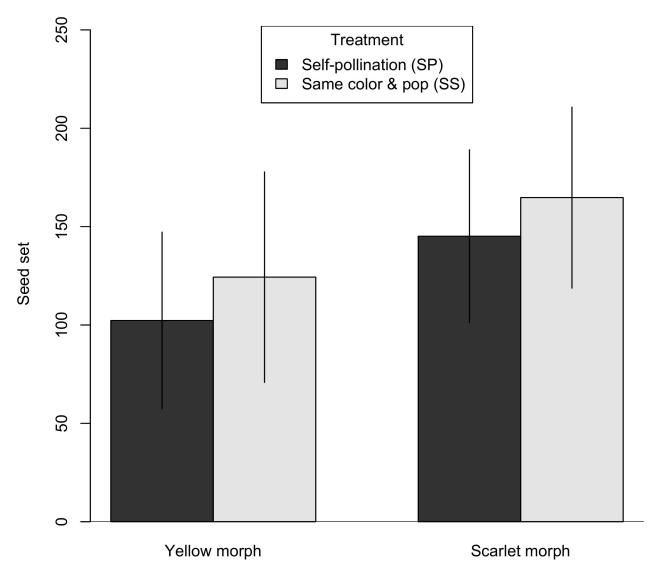
Among open control plants, one yellow plant had a broken stem and one scarlet plant could not be located after collecting the fruit set data (S1G and S1H Table). For the remaining

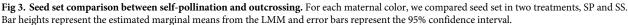
Model	df	AICc	ΔAICc	Weight
Null	2	131.7	0	0.447
Treatment	3	133	1.31	0.232
Maternal color	3	133.8	2.11	0.155
Maternal color + Treatment + Maternal color:Treatment	5	135	3.27	0.087
Maternal color + Treatment	4	135.2	3.47	0.079

Table 1. Summary of model selection results for the relationship between fruit set and self-compatibility.

Fruit set was predicted by two fixed effects and their interaction. This set of models includes the SP and SS treatments.

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https://doi.org/10.1371/journal.pone.0209176.g003

Model	df	AICc	ΔAICc	Weight
Maternal color	4	540.8	0	0.423
Null	3	542.2	1.37	0.213
Maternal color + Treatment	5	542.4	1.51	0.198
Treatment	4	543.5	2.67	0.111
Maternal color + Treatment + Maternal color:Treatment	6	545	4.11	0.054

Table 2. Summary of model selection results for the relationship between seed set and self-compatibility.

Seed set was predicted by two fixed effects and their interaction. This set of models includes the SP and SS treatments.

https://doi.org/10.1371/journal.pone.0209176.t002

plants, fruit set in the open control was not different between the two populations, as the null model was preferred (Δ AICc = 2.08, weight = 0.739). Fruit set for open control plants was high in both populations: 80.6% in the yellow population (95% confidence interval, 66.7%-94.6%) and 82.8% in the scarlet population (73.0%-92.5%). When testing seed set, model selection showed a slight preference for the model that included population over the null model (Δ AICc = 0.45, weight = 0.556). Seed set in the scarlet population was estimated as 188.8 seeds per fruit (95% C.I., 117.3–260.4), much greater than the 96.3 seeds per fruit (27.40086–165.2658) estimated for the yellow population.

Pollinator exclusion

Both morphs were capable of self-fertilization; both the bagged, no hand pollination treatments (BN) and the self-pollination (SP) treatments yielded fruits and seeds. Fruit and seed production in the BN treatment indicates either autonomous self-pollination, a bag effect (accidental pollination when bags were placed or removed), or pollination by "squatters" [57] (small, long-staying insects such as aphids and thrips already present when bags were placed). Interestingly, the color morphs differed markedly in comparisons of the BN and SP treatments. For the scarlet morph, the BN plants had a 43% reduction in fruit set (Fig 4, S1C Table) and a 66% decline in seed set compared to the SP plants (Fig 5, S1D Table). For the yellow morph, the BN plants had no reduction in fruit set (Fig 4), and only a 12% reduction in seed set relative to the SP plants (Fig 5). Thus, pollinator exclusion had a greater effect on the scarlet morph with respect to both fruit set and seed set (Figs 4 and 5). Statistical modeling provides the most support for difference between morphs with respect to reliance on a pollen vector. For both fruit set (Table 3) and seed set (Table 4), the best candidate model included the interaction between treatment and maternal color (fruit set, Akaike weight = 0.598; seed set, Akaike weight = 0.483).

Cross-compatibility and relative fitness

There was some evidence that fruit set was greatest for pollination between the same colors from same population (Fig 6, S1E Table). The manipulated movement of pollen between plants readily yielded fruits with a large number of seeds (Fig 7, S1F Table). In all combinations, pollination with same source population but different color saw a 6–15% reduction in fruit set. However, no reduction in seed set for pollinations using different population of different color was observed for maternal plants with yellow bracts, and the reduction was less than 3% for maternal plants with scarlet bracts (Fig 7). The "color" fixed effect did appear in the best candidate model for describing fruit set (Akaike weight = 0.417; Table 5), and the lack of interaction with maternal color suggests a consistent effect in both color morphs. The

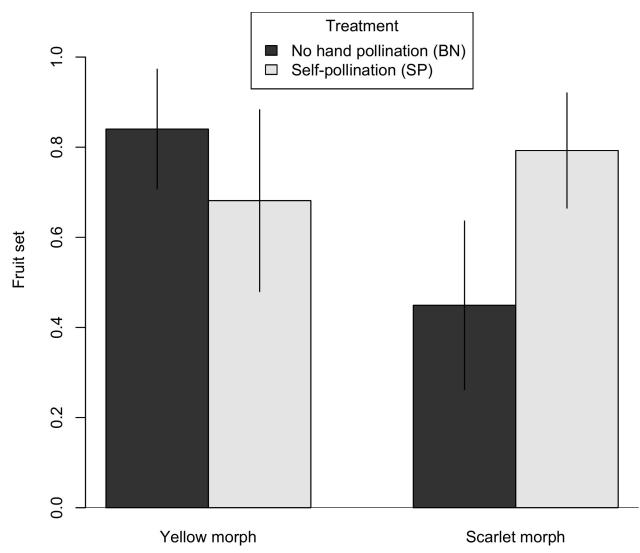


Fig 4. Fruit set comparison between bagged, no hand pollination and self-pollination. For each maternal color, we compared fruit set in two treatments, BN and SP. Bar heights represent the estimated marginal means from the GLMM and error bars represent the 95% confidence interval.

"color" fixed effect was not present in any of the top candidate models for seed set (best model that included "color", $\Delta AICc = 2.16$, Akaike weight = 0.113; Table 6).

Fruit set was lower when pollen donors came from the scarlet population, whether the pollen donors were scarlet or yellow. The decline was 12–18% for yellow maternal plants ("different population" pollen donors) and 29–41% for scarlet maternal plants ("same population" pollen donors). The differential success of pollen from the two populations was reflected in the strong support for models that included the interaction between "population" and maternal color (top three combined Akaike weights = 0.905; Table 5).

Seed set was consistently 24–57% greater in maternal plants of the scarlet morph compared to the yellow morph (Fig 7), with seed set being slightly higher (by 6–16%) when the maternal plant and pollen donor came from the same population (Fig 7). Mixed-effects models provided little support for a role of pollen source in predicting seed set. Also, there is almost no support for an interaction between "population" and maternal color, as was found with fruit set. The variable with the strongest explanatory power for seed set was maternal color; the best

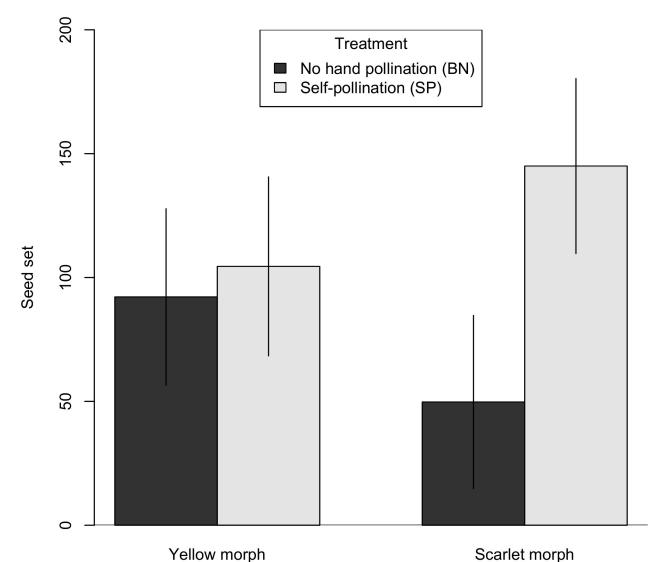


Fig 5. Seed set comparison between bagged, no hand pollination and self-pollination. For each maternal color, we compared seed set in two treatments, BN and SP. Bar heights represent the estimated marginal means from the LMM and error bars represent the 95% confidence interval.

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Model	df	AICc	ΔAICc	Weight
Maternal color + Treatment + Maternal color:Treatment	5	155	0	0.598
Null	2	157.7	2.7	0.155
Maternal color	3	158.4	3.41	0.109
Treatment	3	159	4.08	0.078
Maternal color + treatment	4	159.5	4.58	0.061

Table 3. Summary of model selection results for the relationship between fruit set and pollinator exclusion.

Fruit set was predicted by two fixed effects and their interaction. This set of models includes the BN and SP treatments.

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Model	df	AICc	ΔAICc	Weight
Maternal color + Treatment + Maternal color: Treatment	6	619.4	0	0.483
Treatment	4	620.2	0.85	0.316
Maternal color + Treatment	5	622.6	3.25	0.095
Null	3	622.9	3.56	0.081
Maternal color	4	625.2	5.87	0.026

Table 4. Summary of model selection results for the relationship between seed set and pollinator exclusion.

Seed set was predicted by two fixed effects and their interaction. This set of models includes the BN and SP treatments.

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performing candidate model had only maternal color as an explanatory variable (Akaike weight = 0.332, <u>Table 6</u>).

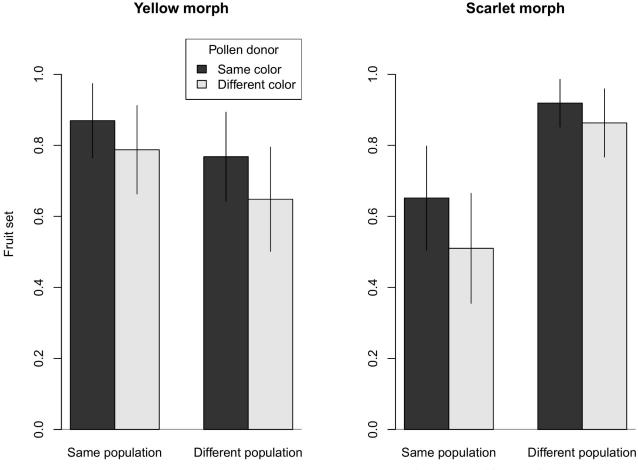
Relative influence of fixed and random effects

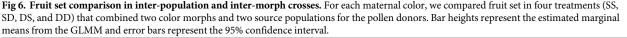
For each statistical model, we estimated the amount of variance explained by the fixed effects (marginal R^2) and the combination of fixed and random effects (conditional R^2). The same random effect, a random intercept for individual plant, was present in all the models. When there is a large discrepancy between the marginal R^2 and conditional R^2 , we expect a large influence of individual plant on the response variable. We found that the two R^2 values were similar for all three of the fruit set models (Table 7), suggesting that the variance observed in fruit set was not explained by differences between individual plants. In the model testing the effect of pollinator exclusion on seed set (Table 4, Fig 5) the marginal R^2 was more than half of the conditional R^2 (Table 7); the fixed effects alone accounted for more than half of the variance observed in seed set. For the seed set models that tested self-compatibility (Table 2, Fig 3) and cross-compatibility (Table 6, Fig 7), the conditional R^2 was much greater than the marginal R^2 (Table 7), indicating a large influence of individual plants in the performance of the model.

Discussion

Castilleja coccinea populations in the Midwestern region show intraspecific bract color polymorphism. Braum [49] reported that the color morphs were also associated with morphological differences, with the scarlet morph consistently larger in several bract and flower measurements including stamen and style length. Differences in both floral color and morphology could impact the breeding system of *C. coccinea* in ways that might involve reproductive trade-offs under pollen or pollinator limitation. Hence, we chose to investigate whether factors related to the breeding system might play a role in maintaining the floral color polymorphism. We found that both color morphs were self-compatible, and fruit set and seed set did not differ between selfed (SP) and outcrossed (SS) pollinations (Figs 2 and 3). Both color morphs are also inter-morph crosses (Fig 6). Two notable differences were found between the color morphs. First, they differed in their response to pollinator exclusion. In the control treatments (bagged, no hand pollination), the scarlet morph showed reduced fruit and seed set, whereas the yellow morph did not (Figs 4 and 5). Second, the scarlet morph set more seed than the yellow morph (Fig 7).

The genus *Castilleja* includes both self-incompatible (*C. levisecta*, *C. linariaefolia*, *C. min-iata*, *C. rhexiifolia*, and *C. sulphurea*) [61–63] and self-compatible (*C. attenuata*) [64] species,

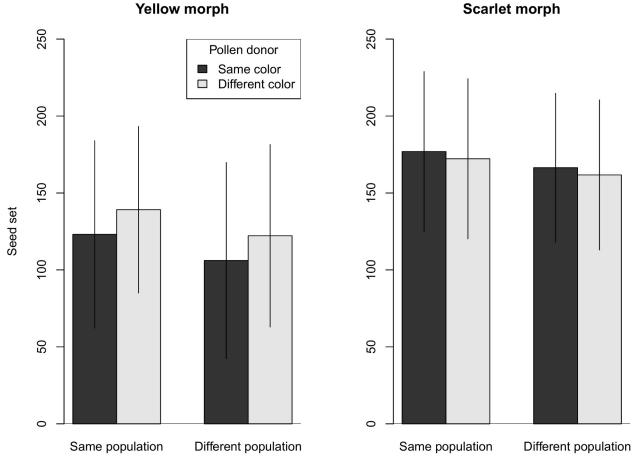


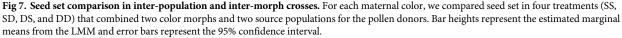


but breeding system had not been previously assessed in *C. coccinea*. Results of this study show that *C. coccinea* is highly self-compatible because fruit set and seed set were not reduced in individuals that received self-pollen compared to individuals that received outcross pollen (Figs 2 and 3). This pattern is true in both color morphs as shown by the self-compatibility indices which are above the threshold of 0.75 to be described as self-compatible, following Lloyd and Schoen [57]. While self-incompatibility assures the genetic and evolutionary bene-fits of outcrossing [65], self-compatible species have the advantage of reproductive assurance when pollen is limited [66], especially when inbreeding depression is low.

Comparison of the self-pollination treatment (SP) and the negative control (BN) showed that, surprisingly, the pollinator exclusion treatment had little effect on the yellow morph, which showed only slight or no reduction in either fruit set and seed set (Figs 4 and 5). The scarlet morph showed reductions in both measures of female fertility under the pollinator exclusion treatment. Thus, the yellow morph would likely experience an advantage of reproductive assurance in cases of pollinator limitation, and perhaps even pollinator absence. Fruit set from the negative control might have been the result of true autonomous self-pollination. In the greenhouse, *C. coccinea* did not set fruits or seeds (J. Fant, personal observation). This suggests that the autogamous selfing we observed might be due to accidental transfer of self-







pollen to stigma ("bag-effect"). Alternatively, predatory insects such as aphids and thrips (squatters) that dwell in flowers may have caused "quasi-autonomous" pollination [57]. Whatever the mechanism of autogamy, the yellow morph outperformed the scarlet morph in the negative control treatment, indicating the yellow morph's ability to tolerate limited pollen delivery. While there are many cases where different color flower morphs attract different pollinators [8,27,63], to our knowledge this is the first report of color morphs differing in their dependence on pollinators for self-fertilization. The abundance of pollinators appears to act as a selective agent in *C. coccinea*, with the yellow morphs being favored when pollinator abundance is low.

There are different modes of self-pollination that offer different levels of reproductive assurance. Geitonogamy, where pollen is transferred between flowers, does not offer reproductive assurance because it relies upon the same pollinator activity as cross-pollination. Bagging experiments like ours investigate autogamous selfing (within flowers), but we did not investigate the precise timing and mechanism, factors that are important for determining the level of benefits provided by reproductive assurance [57].

Greater seed set was observed for the scarlet morph compared to the yellow morph, suggesting that the two morphs also differ in potential reproductive output. Regardless of the bract color of the pollen donor or which population the pollen came from, the scarlet

Model	df	AICc	ΔAICc	Weight
Maternal color + Color + Pop + Maternal color: Pop	6	276.9	0	0.417
Maternal color + Pop + Maternal color: Pop	5	277.6	0.73	0.29
Maternal color + Color + Pop + Maternal color: Color + Maternal color: Pop	7	278.4	1.49	0.198
Рор	3	282.8	5.88	0.022
Null	2	283	6.12	0.02
Color + Pop	4	283.8	6.91	0.013
Color		284	7.07	0.012
Maternal color + Pop		284.8	7.93	0.008
Maternal color		285.1	8.17	0.007
Maternal color + Color + Pop	5	285.9	8.99	0.005
Maternal color + Color		286	9.12	0.004
Maternal color + Color + Maternal color: Color	5	287.4	10.47	0.002
Maternal color + Color + Pop + Maternal color: Color	6	287.5	10.62	0.002

Table 5. Summary of model selection results for fruit set and cross-compatibility between color morphs and populations.

Fruit set was predicted by three fixed effects and two of their interactions. The data for this set of models resulted from the SS, SD, DS, and DD treatments.

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individuals consistently produced more seeds per capsule ($\underline{Fig 7}$). This difference was also observed for individuals that were self-pollinated ($\underline{Fig 3}$) (except for the negative control as already noted).

Our study was not ideally designed to distinguish the effects of genotypic differences among individual plants from the effects of treatment, since we could not apply every

Model	df	AICc	ΔAICc	Weight
Maternal color	4	1267	0	0.332
Null	3	1268	1.73	0.14
Maternal color + Same pop	5	1269	1.81	0.134
Maternal color + Same color	5	1269	2.16	0.113
Same pop	4	1271	3.74	0.051
Same color	4	1271	3.88	0.048
Maternal color + Same color + Same Pop	6	1271	4.02	0.045
Maternal color + Same pop + Maternal color: Same pop	6	1271	4.02	0.044
Maternal color + Same color + Maternal color: Same color	6	1271	4.17	0.041
Same color + Same pop	5	1273	5.93	0.017
Maternal color + Same color + Same Pop + Maternal color: Same color		1273	6.06	0.016
Maternal color + Same color + Same pop + Maternal color: Same pop		1273	6.27	0.014
Maternal color + Same color + Same pop + Maternal color: Same color + Maternal color: Same pop	8	1275	8.36	0.005

Table 6. Summary of model selection results for seed set and cross-compatibility between color morphs and populations.

Seed set was predicted by three fixed effects and two of their interactions. The data for this set of models resulted from the SS, SD, DS, and DD treatments.

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	Response variable	Figure	R ² m	R ² c
Self-compatibility	Fruit set	Fig 2	0.0444	0.0444
	Seed set	Fig 3	0.151	0.441
Pollinator exclusion	Fruit set	Fig 4	0.136	0.145
	Seed set	Fig 5	0.286	0.464
Cross-compatibility and relative fitness	Fruit set	Fig 6	0.142	0.159
	Seed set	Fig 7	0.0758	0.456

Table 7. Comparison of the marginal (R^2m) and conditional (R^2c) coefficient of determination values.

 R^2 m and R^2 c are marginal R^2 and conditional R^2 , respectively.

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treatment to every individual. The small number of individuals in each treatment compounds this limitation. However, we made efforts to statistically assess the relative influence of fixed effects (experimental treatments, maternal plant color) and random effects (individual plant) in the models using marginal and conditional R^2 [59]. Where the difference between these values is large, there is the potential that genotypic differences between individual plants may be confounding our experimental findings. We found that differences in individual plants explained almost none of the variance observed in the fruit set analyses, as the marginal R^2 and conditional R^2 were nearly equal (Table 7). Additionally, for our analysis of pollinator exclusion and seed set, the marginal R^2 was more than half the value of the conditional R^2 . These findings reinforce our conclusions regarding the differences in fruit set between bract colors and among experimental treatments. Also, the influence of pollinator exclusion on seed set (which differs between color morphs) is largely confirmed. However, the conditional R^2 was much greater for the other two analyses of seed set. Conclusions regarding seed set and the self-compatibility experiment, or the cross-compatibility experiment, must be made with caution. We cannot rule out the possibility that genotypic differences between plants randomly assigned to treatment were the primary drivers of the patterns we observed for the latter two tests.

Our study was conducted over a single flowering season, so we cannot say whether the higher seed set for the red morph would be maintained over multiple years or varying conditions. Differences in seed set were observed between color morphs when five floral color polymorphic species (*Cirsium palustris, Digitalis purpurea, Holcus lanatus, Polygonum persicaria, and Vicia sepium*) were under drought and well-watered treatments [36]. Under a drought treatment, pink/purple morphs had greater seed set, but under a well-watered treatment, white morphs had greater seed set. The year of our study (2013) was extremely wet from April through June but had near average temperature and precipitation from June through August (National Temperature and Precipitations Maps obtained from NOAA).

We also observed differences in male fertility between populations, though not between color morphs. Plants from the scarlet population were poor pollen donors; hand-pollinations using pollen from the scarlet population individuals consistently resulted in lower fruit set (Fig 6). The lower fruit set was not related to bract color but rather related to the source population of the pollen donor because both color morphs were tested as pollen donors. The poor quality or low quantity pollen of the scarlet population likely reduces the overall male fertility of the scarlet population. We did not directly test pollen viability or pollen competition in this study, and we do not know the cause of the reduced pollen performance. Different pollen viability among color morphs has been observed in *Claytonia virginica* [67]. The reduced male fertility in the scarlet population might be due to higher inbreeding depression in that population [68],

although we have no other evidence that suggests inbreeding levels or levels of genetic variability differ between the two populations.

While the color morphs were cross-compatible, there was slight evidence of reduced fruit set for inter-morph crosses (Fig 6). Reduced intermorph compatibility may lead to reproductive isolation and genetic divergence of the color morphs. For seed set, there was no evidence that inter-morph crosses produced fewer seeds per fruit (Fig 7). Further studies of pollinator behavior, mating system, and population genetics could reveal more about the reproductive interaction between the two morphs. We are currently conducting a genetic study, using a double digest Restriction-Site Associated DNA sequencing (ddRADseq) approach [69], to address gene flow between color morphs, and compare genetic structure and inbreeding across morphs and populations.

This study was limited to two populations that differed strikingly in bract color frequency. The populations were very close to each other, but undetected site-specific effects might exist. Further investigations at additional sites would be needed to confirm that the reproductive differences we observed between scarlet and yellow bract colored *C. coccinea* extend across the species range. Based on findings from these two sites, we posit that these reproductive trade-offs maintain the bract color polymorphism in *C. coccinea*, where pollinators are selective agents. The scarlet morph has greater potential reproductive output, but the yellow morph has greater reproductive assurance when pollinators are limited. In the absence of pollen limitation, both bract colors develop fruit equally well, but the scarlet morph would yield greater seed set. This appeared to be the situation for our open control, where scarlet plants produced many more seeds per fruit (though fruit set was similar between morphs). When pollinators are limited, the yellow morph may self-pollinate at a higher rate. The differences we observed between the color morphs might also have conservation implications, with the scarlet morph more susceptible to the negative consequences of pollinator declines due to pollution, habitat loss, or habitat degradation [70,71].

Supporting information

S1 Table. Sample sizes for fruit set and seed set analyses. We provide the sample sizes for our study as number of individuals, number of flowers for fruit set analyses, and number of fruits for seed set analyses. (DOCX)

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References

- 1. Galen C. Why Do Flowers Vary?: The functional ecology of variation in flower size and form within natural plant populations. Bioscience. 1999; 49: 631–640. https://doi.org/10.2307/1313439
- Cariveau D, Irwin RE, Brody AK, Garcia-Mayeya LS, von der Ohe A. Direct and indirect effects of pollinators and seed predators to selection on plant and floral traits. Oikos. 2004; 104: 15–26. <u>https://doi.org/10.2307/3548311</u>
- Elmqvist T, Liu D, Carlsson U, Giles BE. Anther-smut infection in *Silene dioica*: variation in floral morphology and patterns of spore deposition. Oikos. 1993; 68: 207. https://doi.org/10.2307/3544832
- Schemske DW, Bierzychudek P. Spatial differentiation for flower color in the desert annual *Linanthus* parryae: was Wright right? Evolution (N Y). 2007; 61: 2528–2543. https://doi.org/10.1111/j.1558-5646. 2007.00219.x PMID: 17894812
- Levin DA, Brack ET. Natural Selection Against White Petals in *Phlox*. Evolution (N Y). 1995; 49: 1017. https://doi.org/10.2307/2410423
- Strauss SY, Irwin RE, Lambrix VM. Optimal defence theory and flower petal colour predict variation in the secondary chemistry of wild radish. J Ecol. 2004; 92: 132–141. <u>https://doi.org/10.1111/j.1365-2745.</u> 2004.00843.x
- Wright S. An analysis of local variability of flower color in *Linanthus Parryae*. Genetics. 1943; 28: 139– 56. PMID: 17247075
- Kay Q. The role of preferential and assortative pollination in the maintenance of flower colour polymorphisms. In: Richards AJ, editor. The pollination of flowers by insects. London: Academic Press; 1978. pp. 175–190.
- Epling C, Lewis H, Ball FM. The breeding group and seed storage: a study in population dynamics. Evolution (N Y). 1960; 14: 238. https://doi.org/10.2307/2405830
- Epling C, Dobzhansky T. Genetics of Natural Populations. VI. Microgeographic Races in *Linanthus parryae*. Genetics. 1942; 27: 317–32. PMID: 17247043
- Gigord LD, Macnair MR, Smithson A. Negative frequency-dependent selection maintains a dramatic flower color polymorphism in the rewardless orchid *Dactylorhiza sambucina* (L.) Soo. Proc Natl Acad Sci U S A. 2001; 98: 6253–5. https://doi.org/10.1073/pnas.111162598 PMID: 11353863
- Takahashi Y, Takakura K, Kawata M. Flower color polymorphism maintained by overdominant selection in *Sisyrinchium* sp. J Plant Res. 2015; 128: 933–939. https://doi.org/10.1007/s10265-015-0750-7 PMID: 26354759
- 13. Eckhart VM, Rushing NS, Hart GM, Hansen JD. Frequency-dependent pollinator foraging in polymorphic *Clarkia xantiana* ssp. *xantiana* populations: implications for flower colour evolution and pollinator interactions. Oikos. 2006; 112: 412–421. https://doi.org/10.1111/j.0030-1299.2006.14289.x
- 14. Irwin RE, Strauss SY. Flower color microevolution in wild radish: evolutionary response to pollinatormediated selection. Am Nat. 2005; 165: 225–237. https://doi.org/10.1086/426714 PMID: 15729652
- Arista M, Talavera M, Berjano R, Ortiz PL. Abiotic factors may explain the geographical distribution of flower colour morphs and the maintenance of colour polymorphism in the scarlet pimpernel. J Ecol. 2013; 101: 1613–1622. https://doi.org/10.1111/1365-2745.12151
- Streisfeld MA, Kohn JR. Contrasting patterns of floral and molecular variation across a cline in *Mimulus aurantiacus*. Evolution (N Y). 2005; 59: 2548. https://doi.org/10.1554/05-514.1
- Miller RB. Hawkmoths and the geographic patterns of floral variation in Aquilegia caerulea. Evolution (N Y). 1981; 35: 763–774. https://doi.org/10.1111/j.1558-5646.1981.tb04936.x PMID: 28563131
- Fenster CB, Armbruster WS, Wilson P, Dudash MR, Thomson JD. Pollination syndromes and floral specialization. Annu Rev Ecol Evol Syst. 2004; 35: 375–403. <u>https://doi.org/10.1146/annurev.ecolsys.</u> 34.011802.132347
- Sobral M, Veiga T, Domínguez P, Guitián JA, Guitián P, Guitián JM. Selective pressures explain differences in flower color among *Gentiana lutea* populations. PLoS One. 2015; 10: e0132522. https://doi.org/10.1371/journal.pone.0132522 PMID: 26172378

- Waser NM, Price M V. Pollinator choice and stabilizing selection for flower color in *Delphinium nelsonii*. Evolution (N Y). 1981; 35: 376. https://doi.org/10.2307/2407846
- Waser NM, Price M V. Pollinator behaviour and natural selection for flower colour in *Delphinium nelso-nii*. Nature. 1983; 302: 422–424. https://doi.org/10.1038/302422a0
- Smithson A. Pollinator preference, frequency dependence, and floral evolution. In: Chittka L, Thomson JD, editors. Cognitive Ecology of Pollination. Cambridge: Cambridge University Press; 2001. pp. 237–258. https://doi.org/10.1017/CBO9780511542268.013
- Schemske DW, Bradshaw HD. Pollinator preference and the evolution of floral traits in monkeyflowers (*Mimulus*). Proc Natl Acad Sci U S A. National Academy of Sciences; 1999; 96: 11910–5. <u>https://doi.org/10.1073/PNAS.96.21.11910 PMID: 10518550</u>
- Stanton ML. Reproductive biology of petal color variants in wild populations of *Raphanus sativus*: I. Pollinator response to color morphs. Am J Bot. 1987; 74: 178. https://doi.org/10.2307/2444019
- 25. Niovi Jones K, Reithel JS. Pollinator-mediated selection on a flower color polymorphism in experimental populations of *Antirrhinum* (Scrophulariaceae). Am J Bot. 2001; 88: 447–54. PMID: <u>11250822</u>
- Brown BA, Clegg MT. Influence of flower color polymorphism on genetic transmission in a natural population of the common morning glory, *Ipomoea purpurea*. Evolution (N Y). 1984; 38: 796. <u>https://doi.org/10.2307/2408391</u>
- Streisfeld MA, Kohn JR. Environment and pollinator-mediated selection on parapatric floral races of *Mimulus aurantiacus*. J Evol Biol. 2007; 20: 122–132. https://doi.org/10.1111/j.1420-9101.2006.01216. x PMID: 17210005
- Strauss SY, Whittall JB. Non-pollinator agents of selection on floral traits. In: Harder LD, Barrett SC., editors. Ecology and Evolution of Flowers. Oxford: Oxford University Press; 2006. pp. 120–138.
- Armbruster WS. Can indirect selection and genetic context contribute to trait diversification? A transition-probability study of blossom-colour evolution in two genera. J Evol Biol. 2002; 15: 468–486. https://doi.org/10.1046/j.1420-9101.2002.00399.x
- **30.** Schemske DW, Bierzychudek P. Perspective: evolution of flower color in the desert annual *Linanthus parryae*: Wright revisited. Evolution (N Y). 2001; 55: 1269–1282. https://doi.org/10.1111/j.0014-3820. 2001.tb00650.x
- Carlson JE, Holsinger KE. Natural selection on inflorescence color polymorphisms in wild *Protea* populations: the role of pollinators, seed predators, and intertrait correlations. Am J Bot. 2010; 97: 934–44. https://doi.org/10.3732/ajb.0900348 PMID: 21622464
- Carlson JE, Holsinger KE. Extrapolating from local ecological processes to genus-wide patterns in colour polymorphism in South African *Protea*. Proc R Soc London B Biol Sci. 2015; 282: 20150583. <u>https:// doi.org/10.1098/rspb.2015.0583 PMID: 25876847</u>
- **33.** Grotewold E. The genetics and biochemistry of floral pigments. Annu Rev Plant Biol. 2006; 57: 761– 780. https://doi.org/10.1146/annurev.arplant.57.032905.105248 PMID: 16669781
- Steyn WJ, Wand SJE, Holcroft DM, Jacobs G. Anthocyanins in vegetative tissues: a proposed unified function in photoprotection. New Phytol. 2002; 155: 349–361. <u>https://doi.org/10.1046/j.1469-8137</u>. 2002.00482.x
- Coberly LC, Rausher MD. Analysis of a chalcone synthase mutant in *Ipomoea purpurea* reveals a novel function for flavonoids: amelioration of heat stress. Mol Ecol. 2003; 12: 1113–1124. <u>https://doi.org/10.1046/j.1365-294X.2003.01786.x</u> PMID: 12694276
- **36.** Warren J, Mackenzie S. Why are all colour combinations not equally represented as flower-colour polymorphisms? New Phytol. 2001; 151: 237–241. https://doi.org/10.1046/j.1469-8137.2001.00159.x
- Irwin RE, Strauss SY, Storz S, Emerson A, Guibert G. The role of herbivores in the maintenance of a flower color polymorphism in wild radish. Ecology. 2003; 84: 1733–1743. https://doi.org/10.1890/0012-9658(2003)084[1733:TROHIT]2.0.CO;2
- Johnson SD. The pollination niche and its role in the diversification and maintenance of the southern African flora. Philos Trans R Soc Lond B Biol Sci. 2010; 365: 499–516. https://doi.org/10.1098/rstb. 2009.0243 PMID: 20047876
- Lloyd DG. Self- and Cross-Fertilization in Plants. II. The Selection of Self- Fertilization. Int J Plant Sci. The University of Chicago Press; 1992; 153: 370–380. https://doi.org/10.1086/297041
- Schoen DJ, Lloyd DG. Self- and Cross-Fertilization in Plants. III. Methods for Studying Modes and Functional Aspects of Self-Fertilization [Internet]. International Journal of Plant Sciences. The University of Chicago Press; pp. 381–393. https://doi.org/10.2307/2995678
- Moeller DA. Geographic structure of pollinator communities, reproductive assurance, and the evolution of self-pollination. Ecology. 2006; 87: 1510–1522. <u>https://doi.org/10.1890/0012-9658(2006)87[1510:</u> GSOPCR]2.0.CO;2 PMID: 16869427

- 42. Elle E, Carney R. Reproductive assurance varies with flower size in *Collinsia parviflora* (Scrophulariaceae). Am J Bot. 2003; 90: 888–896. https://doi.org/10.3732/ajb.90.6.888 PMID: 21659183
- 43. Dart SR, Samis KE, Austen E, Eckert CG. Broad geographic covariation between floral traits and the mating system in *Camissoniopsis cheiranthifolia* (Onagraceae): multiple stable mixed mating systems across the species" range?. Ann Bot. 2012; 109: 599–611. <u>https://doi.org/10.1093/aob/mcr266</u> PMID: 22028462
- 44. Gervasi DDL, Schiestl FP. Real-time divergent evolution in plants driven by pollinators. Nat Commun. 2017; 8: 14691. https://doi.org/10.1038/ncomms14691 PMID: 28291771
- 45. Fry JD, Rausher MD. Selection on a floral color polymorphism in the tall morning glory (*Ipomoea purpurea*): transmission success of the alleles through pollen. Evolution (N Y). 1997; 51: 66–78. <u>https://doi.org/10.1111/j.1558-5646.1997.tb02389.x PMID: 28568811</u>
- Epperson BK, Clegg MT. Frequency-dependent variation for outcrossing rate among flower-color morphs of *Ipomoea purpurea*. Evolution (N Y). 1987; 41: 1302. https://doi.org/10.2307/2409095
- Rausher MD, Augustine D, VanderKooi A. Absence of pollen discounting in a genotype of *Ipomoea purpurea* exhibiting increased selfing. Evolution (N Y). 1993; 47: 1688–1695. https://doi.org/10.1111/j.1558-5646.1993.tb01261.x PMID: 28567997
- 48. Torrey J. A flora of the state of New York, vol. II,. Albany: Carroll and Cook; 1843.
- Braum A. Investigating the drivers of floral trait polymorphism in *Castilleja Coccinea* (L.) Sprengel (Orobanchaceae) [Internet]. Northwestern University. 2014. Available: https://arch.library.northwestern.edu/concern/generic_works/1n79h4338
- 50. Robertson C. Flowers and insects: Asclepiadaceae To Scrophulariaceae. Trans Acad Sci Saint Louis. 1891; 5: 569–598.
- Bertin RI. The Ruby-throated Hummingbird and its major food plants: ranges, flowering phenology, and migration. Can J Zool. NRC Research Press Ottawa, Canada; 1982; 60: 210–219. <u>https://doi.org/10. 1139/z82-029</u>
- **52.** Williamson S. A field guide to hummingbirds of North America. Roger Tory Peterson, editor. Boston: Houghton Mifflin; 2001.
- 53. Spira T. Wildflowers and Plant Communities of the Southern Appalachian Mountains and Piedmont: A Naturalist's Guide to the Carolinas, Virginia, Tennessee, and Georgia. Chapel Hill, North Carolina: The University of North Carolina Press; 2011.
- 54. Eastman J. Wildflowers of the eastern United States. 1st ed. Mechanicsburg, Pennsylvania, USA: Stackpole Books; 2014.
- Bates D, Mächler M, Bolker B, Walker S. Fitting Linear Mixed-Effects Models Using Ime4. J Stat Softw. 2015; 67: 1–48. https://doi.org/10.18637/jss.v067.i01
- R Core Team. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing; 2018. Available: https://www.r-project.org/
- Lloyd DG, Schoen DJ. Self- and Cross-Fertilization in Plants. I. Functional Dimensions. International Journal of Plant Sciences. The University of Chicago Press; pp. 358–369.
- Lenth R V. Least-Squares Means: The R Package Ismeans. J Stat Softw. 2016; 69: 1–33. <u>https://doi.org/10.18637/jss.v069.i01</u>
- Nakagawa S, Schielzeth H. A general and simple method for obtaining R² from generalized linear mixed-effects models. O'Hara RB, editor. Methods Ecol Evol. Wiley/Blackwell (10.1111); 2013; 4: 133– 142. https://doi.org/10.1111/j.2041-210x.2012.00261.x
- Barton K. MuMIn: Multi-Model Inference. R Package. version 1.42.1. [Internet]. 2018. Available: https://cran.r-project.org/package=MuMIn
- Kaye TN, Lawrence B. Fitness effects of inbreeding and outbreeding on golden paintbrush (Castilleja levisecta): Implications for recovery and reintroduction. 2003;
- Carpenter FL. Pollination energetics in avian communities: simple concepts and complex realities. In: Jones C. E., Little R. J., editors. Handbook of experimental pollination biology. New York: Van Nostrand Reinhold; 1983. pp. 215–234.
- Hersch-Green EI. Polyploidy in Indian paintbrush (*Castilleja*; Orobanchaceae) species shapes but does not prevent gene flow across species boundaries. Am J Bot. Botanical Society of America; 2012; 99: 1680–90. https://doi.org/10.3732/ajb.1200253 PMID: 23032815
- Chuang TI, Heckard LR. New species of bee-pollinated *Castilleja* from Peru, with a taxonomic revision of South American members of subg. *Colacus*. Syst Bot. American Society of Plant Taxonomists; 1992; 17: 417. https://doi.org/10.2307/2419482
- Igic B, Lande R, Kohn JR. Loss of self-incompatibility and its evolutionary consequences. Int J Plant Sci. 2008; 169: 93–104. https://doi.org/10.1086/523362

- 66. Kalisz S, Vogler DW, Hanley KM. Context-dependent autonomous self-fertilization yields reproductive assurance and mixed mating. Nature. 2004; 430: 884–887. https://doi.org/10.1038/nature02776 PMID: 15318220
- Frey FM. Phenotypic integration and the potential for independent color evolution in a polymorphic spring ephemeral. Am J Bot. 2007; 94: 437–444. https://doi.org/10.3732/ajb.94.3.437 PMID: 21636413
- Krebs SL, Hancock JF. Early-acting inbreeding depression and reproductive success in the highbush blueberry, *Vaccinium corymbosum* L. Theor Appl Genet. 1990; 79: 825–832. https://doi.org/10.1007/ BF00224252 PMID: 24226746
- Peterson BK, Weber JN, Kay EH, Fisher HS, Hoekstra HE. Double digest RADseq: an inexpensive method for de novo SNP discovery and genotyping in model and non-model species. Orlando L, editor. PLoS One. Molecular Biology and Evolution; 2012; 7: e37135. <u>https://doi.org/10.1371/journal.pone.</u> 0037135 PMID: 22675423
- Steffan-Dewenter I, Tscharntke T. Effects of habitat isolation on pollinator communities and seed set. Oecologia. 1999; 121: 432–440. https://doi.org/10.1007/s004420050949 PMID: 28308334
- Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE. Global pollinator declines: trends, impacts and drivers. Trends Ecol Evol. 2010; 25: 345–53. https://doi.org/10.1016/j.tree.2010. 01.007 PMID: 20188434