

# Article Synthesis of Demissidine Analogues from Tigogenin via Imine Intermediates <sup>†</sup>

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Dedicated to Prof. Dr. Ludger Wessjohann on the occasion of his 60th birthday.

**Abstract:** A five-step transformation of a spiroketal side chain of tigogenin into an indolizidine system present in solanidane alkaloids such as demissidine and solanidine was elaborated. The key intermediate in the synthesis was spiroimine **3** readily obtained from tigogenin by its RuO<sub>4</sub> oxidation to 5,6-dihydrokryptogenin followed by amination with aluminum amide generated in situ from DIBAlH and ammonium chloride. The mild reduction of spiroimine to a 26-hydroxy-dihydropyrrole derivative and subsequent mesylation resulted in the formation of 25-epidemissidinium salt or 23-sulfone depending on reaction conditions.

Keywords: steroidal alkaloids; solanidane alkaloids; demissidine; solanidine

## 1. Introduction

Demissidine and solanidine are the main representatives of the solanidane alkaloids that occur mainly as glycosides in potato species including *Solanum tuberosum*, *Solanum demissum*, and *Solanum acaule* (Figure 1) [1,2]. The various biological properties of these cholestane alkaloids have been reported in the literature [3]. Among these,  $\alpha$ -solanine and  $\alpha$ -chaconine, two main solanidine glycosides, are potent enough to inhibit proliferation and induce apoptosis in various types of cancer cells including cervical, liver, lymphoma, and stomach cancer cells [4]. The effectiveness of  $\alpha$ -chaconine against hepatocellular cancer HepG2 cells is higher than the common anticancer agents doxorubicin and camptothecin [5]. Additionally, demissidine and its natural glycoside, commersonine, inhibit the growth of human colon and liver cancer cells in culture [5]. Apart from showing antitumor activity, solanidane-type alkaloids are known to act as natural insect deterrents, have antimicrobial and anti-inflammatory properties, inhibit acetylcholinesterase, and disrupt cell membranes [3,6–9]. Additionally, studies of solanidine and demissidine analogues confirm their potency for the design of new pharmacologically active agents [10–12].



Figure 1. Steroidal alkaloids of solanidane type.

So far, eight syntheses of solanidine and demissidine have been described, four of them in the last decade, and the latest one was reported last year [13–20]. Although recently invented methods brought a significant improvement, the described methods suffer from several drawbacks, such as multi-step procedures or unsatisfactory yields.



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**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). Moreover, they cannot be easily adapted to the synthesis of demissidine or solanidine analogues. Therefore, the elaboration of an efficient route to demissidine congeners is still needed. An improved approach to the synthesis of different demissidine stereoisomers has been recently reported [21]. Here, we propose an alternative strategy toward demissidine analogues from an easily available steroid sapogenin—tigogenin.

#### 2. Results and Discussion

We found that a convenient intermediate for the transformation of the spiroketal system present in steroidal sapogenins, e.g., tigogenin, into the solanidane framework of demissidine was spiroimine **3**, shown in Scheme 1. This novel spirostane aza-analogue was obtained from tigogenin by a two-step protocol involving tigogenin oxidation to a 5,6-dihydrokryptogenin derivative and its reaction with aluminum amide as an aminating agent.



Scheme 1. Synthesis of spiroimine 3 from tigogenin 3-TBS ether (1).

The most convenient method for the oxidative cleavage of sapogenin spiroketal to hydroxy-diketone was chosen first. After perusing the known literature protocols [22–25], we employed the RuO<sub>4</sub>/NaIO<sub>4</sub> catalytic system. The desired 5,6-dihydrokryptogenin derivative was obtained by the oxidation of tigogenin 3-TBS ether (1) as a mixture of two tautomers 2a and 2b in 71% yield. In the next step, the obtained product was subjected to a reaction with aluminum amide generated in situ from diisobutylaluminum hydride (DIBAlH) and ammonium salt. The use of various aminoalanes as aminating agents for such compounds as epoxides, ketones, carboxylic acids, and their derivatives (chlorides, esters) has previously been widely reported in the literature [26–31]. Our previous investigations have shown that the desired aminoalane might be readily synthesized by the treatment of DIBAlH with ammonium chloride under mild reaction conditions  $(0 \circ C - room temperature, THF, up to 2 h)$  [32]. However, the reagent proved to be unstable and its structure was not definitely determined. The reaction of the 2a/2b mixture with aminoalane prepared as described above was carried out in refluxing THF/toluene (Scheme 1). Spiroimine **3** was obtained as the main reaction product (44%) when using aluminum amide prepared in situ from 40 equivalents of DIBAlH and 42 equivalents of ammonium chloride. It is worth noting that this compound was not formed in the absence of DIBAIH. Compound 3 was accompanied by two minor products, imine 4 (12%) and enone 5 (5%). Ketone 5 was produced as a result of an aldol condensation probably due to enolization caused by aluminum amide playing a role as a Lewis acid. The formation of imine 4 in the experiment was unexpected and difficult to explain in terms of the substrate 2a/2b reaction with prepared aminoalane. It seems that some unreacted diisobutylaluminum hydride was still present in the reaction mixture, resulting in the reduction of initially produced spiroimine 3 to 4 (vide infra). The reaction of compounds 2a and 2b with aminoalane prepared from a lower amount of DIBAIH led to the incomplete conversion. For example, employing the aminating reagent produced from 20 equivalents of DIBAlH, imine **3** was obtained in 26% yield only, while the  $\alpha_{\lambda}\beta$ -unsaturated ketone **5** was produced in 25% yield. In this case, compound 4 was not isolated. Both imines, 3 and 4, appeared to be convenient substrates for the synthesis of solasodine or solanidine derivatives. The

mild reduction of spiroimine should provide hitherto unknown 'reverse' spirosolanes with the nitrogen atom in the pyrrolidine E-ring and the oxygen atom in the 'pyranose' F-ring. Moreover, the reductive cleavage of the spiroimine F-ring may open a direct way to solanidane alkaloids possessing an indolizidine moiety.

First, the reduction of compound **3** under mild conditions was attempted. Interestingly, the expected hemiaminal **6** (Scheme 2) was not obtained, though various reducing agents were examined. Using an equimolar amount of various borohydrides, such as NaBH<sub>4</sub>, NaBH<sub>4</sub>/I<sub>2</sub>, and NaBH<sub>3</sub>CN, under different conditions (temperature, reaction times), the main isolated product was always imine **4** accompanied by small amounts of pyrrolidine **7**. The other examined reducing agents (DIBAlH, H<sub>2</sub>/PtO<sub>2</sub>, H<sub>2</sub>/Pd, Hantzsch ester/TFA [33], TESH/acid) proved less effective.



Scheme 2. Reduction of imine 3 with borohydrides.

The above-described results of the reduction experiments pointed out that compound 6 is less stable than its open-chain isomer 4 (confirmed by calculations). This explains unsuccessful attempts of imine 4 cyclization to 6 in the presence of acids. The observed behavior of imine 4 is clearly different from that of 'pseudosapogenins', which readily cyclize to spiroketals. The latter are relatively stable compounds, though their F-ring opening occurs when they are treated with strong Lewis acids. The natural aza-analogues of spirostanes (spirosolanes) containing the nitrogen atom in ring F, e.g., solasodine or tomatidine, are even more susceptible to an electrophilic attack than spiroketals. However, in the case of spirosolanes, the 'furanose' E-ring is readily opened [34]. This is because the cation resulting from the C–O bond cleavage is stabilized by electrons of the neighboring nitrogen atom. It seems that the isomeric compounds containing the nitrogen atom in the E-ring undergo the opening of the 'pyranose' F-ring for the same reason. The cleavage of the oxygen-containing ring in spirosolanes was also observed under the reducing conditions [35,36]. Despite the failure to obtain a 'reverse' spirosolane analogue from imine 3, it still seemed to be a convenient intermediate for the synthesis of solanidane alkaloids. A derivative of imine 4 was previously employed by Uhle and Tian to build an indolizidine system. In the solanidine analogue synthesis reported by Uhle [37], the imine was obtained in 20% yield from kryptogenin 16-(2,4-dinitrophenyl)hydrazone and transformed into 25-episolanidine by refluxing with KOH in ethylene glycol in 65% yield. In 2016, Tian and coworkers [18] developed a new way to synthesize solanidine and demissidine using diosgenin or tigogenin as a starting material, respectively. In the method proposed by the Chinese group, 26-methyl ester 22-imine was prepared in five steps and further transformed into the desired alkaloid by the selective reduction of the imine moiety to pyrrolidine, spontaneous intramolecular aminolysis of the obtained amino-ester to lactam, and reduction. The use of spiroimine 3 as an intermediate for the construction of an indolizidine unit allowed us to shorten the solanidane synthesis from tigogenin. The approach explored in our study involved the reduction of spiroimine 3 to dihydropyrrole 4 followed by its cyclization and reduction. As our initial studies on the imine reduction showed that only complex borohydrides were effective for this transformation, we went on to optimize the reduction reaction conditions using NaBH<sub>4</sub> and NaBH<sub>3</sub>CN as reducing agents. Our results are summarized in Table 1. Apart from compound 4, in most cases a small amount of amine 7 was formed. Imine 4 was obtained in the best yield in reaction with a NaBH $_4/I_2$  system (entry 5). With 1 equivalent of NaBH $_4$  (without any additives or with AcONa) at a low temperature and controlling the reaction time, we restrained imine over-reduction and obtained compound 4 in good yield (entry 2, 3, 4). NaBH<sub>3</sub>CN was less

efficient (entry 6, 7). Additionally, when NaBH<sub>3</sub>CN was used in the presence of AcOH, an imine–cyanoborane complex **8** was formed (Figure 2).

**Table 1.** The optimization of imine 3 reduction conditions.

	H H H 3	sodium borohydrides		H NH	ЮН
Entry	Reagent (equiv.)	Conditions	Product 4 Yield (%)	Product 7 Yield (%)	Substrate Recovery (%)
1	NaBH <sub>4</sub> (2)	MeOH/CH <sub>2</sub> Cl <sub>2</sub> , -18 °C, 1 h	64	30	<1
2	NaBH <sub>4</sub> (1)	MeOH/CH <sub>2</sub> Cl <sub>2</sub> , -18 °C, 1 h	58	<5	21
3	NaBH <sub>4</sub> (1)	MeOH/CH <sub>2</sub> Cl <sub>2</sub> , 0 °C, 1 h	67	13	<5
4	NaBH <sub>4</sub> (1)	NaOAc (1 equiv.), MeOH/CH <sub>2</sub> Cl <sub>2</sub> , 0 °C–room temp., 2 h	65	<5	30
5	NaBH <sub>4</sub> (4)	I <sub>2</sub> (4 equiv.), MeOH/ CH <sub>2</sub> Cl <sub>2</sub> , 0 °C–reflux, 16 h	78	9	<1
6	NaBH <sub>3</sub> CN (2)	AcOH (2 equiv.), THF, room temp., 1 h	complex 8 (48)	nd *	<5
7	NaBH <sub>3</sub> CN (2)	THF/MeOH, room temp., 2 h	28	nd *	30

\* nd—not detected.



Figure 2. Complex 8 formed during the imine 3 reduction with NaBH<sub>3</sub>CN/AcOH.

We envisaged that the activation of the 26-hydroxyl group in compound 4 would result in spontaneous ring closing to iminium salt. Therefore, we subjected compound 4 to a reaction with mesyl chloride. As examples of the successful chemoselective mesylation of the primary hydroxyl group in the presence of amine function could be found in the literature [38–40], we supposed that the chemoselective mesylation of hydroxy-imine should be reached under similar conditions. The initial mesylation of hydroxy-imine 4 carried out with 1.2 equivalents of mesyl chloride in the presence of triethylamine at -15 °C resulted mainly in a less polar product (26-mesyloxy-imine), which spontaneously cyclized after work-up to the desired iminium salt 9 (Scheme 3). Under mesylation conditions, TBS protection of the 3-OH group was also removed and the indolizinium salt 9 was isolated in 45% yield. Compound 9 was readily transformed into 25-epidemissidine (10) by reduction with NaBH<sub>4</sub>.



Scheme 3. Synthesis of 25-epidemissidine (10).

Conducting the mesylation under slightly harsher conditions (1.2 equiv. of MsCl,  $Et_3N$ , 0 °C or 2 equiv. of MsCl, Py, DMAP(cat.), 0 °C–room temp.) led to a complex mixture of products. The iminium salt was formed only in 5% yield, while the main reaction product was identified as an enamine *N*,*O*-dimesyl derivative **11a** or **11b** (Figure 3).



**11a**: R = Ms (for MsCl (2 equiv.),  $CH_2Cl_2/Py$ , DMAP cat.) **11b**: R = TBS (for MsCl (1.2 equiv.),  $Et_3N$ ,  $CH_2Cl_2$ , 0 °C)

Figure 3. Major products of imine 4 mesylation under harsh conditions.

As the changes made did not result in the yield improvement of the desired indolizinium salt, we also attempted to improve the chemoselectivity of *O*-mesylation by deactivating the imine nitrogen. For this purpose, hydroxy-imine **4** was reacted with hydrogen chloride (generated in situ from AcCl and MeOH) to obtain imine hydrochloride before mesylation. The crude salt without isolation was subjected to mesylation with 2 equivalents of MsCl in the presence of Et<sub>3</sub>N at 0 °C–room temp. To our surprise, after basic work-up sulfone **12** (Scheme 4) was isolated, instead of the expected indolizinium salt **9**. The obtained solanidane seems to be a valuable intermediate for the synthesis of leptinidine analogues.



Scheme 4. Mesylation of imine 4 preceded by protonation with HCl.

The hypothetical mechanism of sulfone formation is outlined in Scheme 5. An addition of HCl caused the tautomerization of imine to enamine (I) via the in situ formation of iminium salt and simultaneous deprotection of TBS ether. The enamine (I) possessing three nucleophilic sites, primary OH group, secondary OH group, and enamine carbon atom, reacted further with mesyl chloride. Apart from alcohol mesylation (II), the mesylation of an enamine electron-rich carbon occurred, leading to sulfone formation with the reconstruction of imine in ring E (III). In the final step, the cyclization to indolizine took place via an intramolecular nucleophilic substitution of 26-mesylate with the imine nitrogen. The sequence of the last-mentioned transformations (the sulfone formation followed by the ring closing) is not obvious. The reverse order of transformations (with the cyclization first) is less likely but could not be excluded. It should be mentioned that a small amount of sulfone was also formed in the mesylation of imine **4** without pre-addition of HCl.



Scheme 5. Tentative mechanism of sulfone 12 formation from imine 4.

The novel compounds prepared within the study, including the imine intermediates that frequently show antibiotic activity [41], will be subjected to biological activity evaluation in due course.

#### 3. Materials and Methods

#### 3.1. General

NMR spectra were recorded with Bruker Avance II 400 spectrometer operating at 400 MHz, using CDCl<sub>3</sub> solutions with TMS as the internal standard (only selected signals in the <sup>1</sup>H NMR spectra are reported). Coupling constants (*J*) are given in Hz. The spectra of compounds 3–10 and 12 are included in the Supplementary Materials. The FTIR spectra were obtained using Nicolet<sup>™</sup> 6700 spectrometer (Thermo Scientific, Waltham, MA, USA). The spectra were recorded in the range between 4000 and 500 cm<sup>-1</sup> with a resolution of 4 cm<sup>-1</sup> and 32 scans using Attenuated Total Reflectance (ATR) techniques. ESI and ESI-HRMS spectra were obtained on the Agilent 6530 Accurate-Mass Q-TOF ESI and LC/MS system. Melting points were determined using MP70 Melting Point System (Mettler Toledo, Greifensee, Switzerland). Thin-layer chromatography (TLC) was performed on aluminum plates coated with silica gel 60 F254 (Merck, Darmstadt, Germany), by spraying with ceric ammonium molybdate (CAM) solution, followed by heating. The reaction products were isolated by column chromatography, performed using 70–230 mesh silica gel (J. T. Baker).

## 3.2. Chemical Synthesis

#### 3.2.1. Oxidation of 3-TBS Tigogenin (1) with RuO<sub>4</sub>/NaIO<sub>4</sub>

Solution of NaIO<sub>4</sub> (1.8 g, 8.4 mmol) and RuO<sub>2</sub> (23 mg, 0.17 mmol) in the mixture of water (20 mL), acetone (10 mL), and tetrachloride (20 mL) was vigorously stirred until the yellow color of RuO<sub>4</sub> appeared. Then, a solution of 3-TBS tigogenin (1, 0.3 g, 0.57 mmol) in 8 mL of CCl<sub>4</sub> was added in three portions and the reaction mixture was stirred for 10 h at room temperature. After that time, the TLC control showed that no starting material remained. A few drops of isopropanol were added to quench RuO<sub>4</sub> and the resulting slurry was stirred for an additional 10 min at room temperature (yellow RuO<sub>4</sub> turned into black RuO<sub>2</sub>). The reaction mixture was poured into water and product was extracted with CHCl<sub>3</sub>. The extract was dried over anhydrous sodium sulfate, and the solvent was evaporated. Silica gel column chromatography afforded the product as an equilibrium mixture of two tautomers **2a** and **2b**, identical to that described in reference [42] in 73% total yield.

Compound **2a**/**2b**, eluted with 7.5% to 25% AcOEt/hexane: for main tautomer: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.58 (m, 2H), 3.47 (m, 1H), 2.57 (m, 1H), 2.62 (m, 1H), 1.02 (d, *J* = 7.0, 3H), 0.95 (d, *J* = 6.6, 3H), 0.89 (s, 9H), 0.81 (s, 3H), 0.74 (s, 3H), 0.06 (s, 6H); ESI-MS 547 [M+H]<sup>+</sup>. HRMS calculated for C<sub>33</sub>H<sub>59</sub>O<sub>4</sub>Si (M+H)<sup>+</sup>, 547.4177; found 547.4230.

#### 3.2.2. Synthesis of (25R)- $3\beta$ -*t*-butyldimethylsililoxy-16-aza-spirost-16(N)-ene (3)

#### Preparation of the aminoalane reagent from DIBALH and NH<sub>4</sub>Cl

A solution of DIBAlH in toluene (1 M, 22 mL, 22 mmol, 40 equiv. relative to compounds **2a** and **2b**) was added to a cooled (0–5 °C) suspension of NH<sub>4</sub>Cl (1.23 g, 23 mmol, 42 equiv.) in anhydrous THF (15 mL) under argon. The reaction was stirred for 15 min in an ice bath and then 1.5 h at room temperature. After this time, the obtained reagent solution was used directly for the reaction with compound **2a**/**2b**.

Synthesis of imine 3

The solution of aminoalane reagent (prepared from 40 equiv. of DIBAlH) was added dropwise to a solution of compound **2a** and **2b** (0.3 g, 0.549 mmol) in anhydrous THF (ca 6 mL) at room temperature. Then, stirring was continued for 16 h at reflux. After this time, the reaction mixture was cooled, quenched with aqueous solution of KHSO<sub>4</sub>, and the product was extracted with ether. The extract was washed with water, dried over anhydrous sodium sulfate, and the solvent was evaporated. Silica gel column chromatography afforded three products: spiroimine **3** (44%) eluted with 10% AcOEt/hexane,  $\alpha$ , $\beta$ -unsaturated ketone **5** (5%) eluted with 15% AcOEt/hexane, and dihydropyrrole **4** (12%) eluted with 70% AcOEt/hexane.

Compound 3: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.56 (m, 2H), 3.45 (dd, *J* = 11.0, 10.9, 1H), 2.56 (m, 1H), 2.45 (m, 1H), 1.01 (d, *J* = 6.9, 3H), 0.89 (s, 9H), 0.84 (d, *J* = 6.6, 3H), 0.83 (s, 3H), 0.61 (s, 3H), 0.06 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  193.2 (C), 107.6 (C), 72.1 (CH), 70.6 (CH), 69.1 (CH<sub>2</sub>), 56.8 (CH), 54.5 (CH), 45.0 (CH), 42.7 (CH), 39.8 (C), 38.6 (CH<sub>2</sub>), 38.3 (CH<sub>2</sub>), 37.0 (CH<sub>2</sub>), 35.7 (C), 35.1 (CH), 33.7 (CH<sub>2</sub>), 32.3 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 30.8 (CH), 29.3 (CH<sub>2</sub>), 28.7 (CH<sub>2</sub>), 28.5 (CH<sub>2</sub>), 25.9 (3xCH<sub>3</sub>), 20.9 (CH<sub>2</sub>), 18.3 (C), 17.2 (CH<sub>3</sub>), 13.5 (CH<sub>3</sub>), 12.43 (CH<sub>3</sub>), 12.39 (CH<sub>3</sub>), -4.6 (2xCH<sub>3</sub>); ESI-MS 528 [M+H]<sup>+</sup>. HRMS calculated for C<sub>33</sub>H<sub>59</sub>NO<sub>2</sub>Si (M+H)<sup>+</sup>, 528.4231; found 528.4297; IR ATR,  $\nu_{max}$  (cm<sup>-1</sup>): 1728, 1667, 1457, 1373, 1248, 1173, 1063.

Compound 4: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  4.43 (m, 1H), 3.55 (m, 1H), 3.43 (dd, J = 11.1, 4.0, 1H), 3.30 (dd, J = 11.1, 6.0, 1H), 2.61 (q, J = 7.3, 1H), 2.29 (m, 2H), 2.22 (m, 1H), 1.08 (d, J = 7.3, 3H), 0.92 (d, J = 6.3, 3H), 0.89 (s, 9H), 0.79 (s, 3H), 0.51 (s, 3H), 0.05 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  182.0 (C), 74.8 (CH), 72.1 (CH), 65.9 (CH<sub>2</sub>), 61.6 (CH), 54.9 (CH), 54.6 (CH), 45.0 (CH), 44.5 (CH), 41.4 (C), 39.2 (CH<sub>2</sub>), 38.6 (CH<sub>2</sub>), 37.2 (CH<sub>2</sub>), 35.9 (CH), 35.6 (C), 35.1 (CH), 32.4 (CH<sub>2</sub>), 32.0 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 28.7 (CH<sub>2</sub>), 27.6 (CH<sub>2</sub>), 27.5 (CH<sub>2</sub>), 25.9 (3xCH<sub>3</sub>), 20.8 (CH<sub>2</sub>), 18.8 (CH<sub>3</sub>), 18.3 (C), 17.0 (CH<sub>3</sub>), 14.0 (CH<sub>3</sub>), 12.4 (CH<sub>3</sub>), -4.6 (2xCH<sub>3</sub>); ESI-MS 530 [M+H]<sup>+</sup>. HRMS calculated for C<sub>33</sub>H<sub>60</sub>NO<sub>2</sub>Si (M+H)<sup>+</sup>, 530.4388; found 530.4398; IR ATR,  $\nu_{max}$  (cm<sup>-1</sup>): 3235, 1631, 1454, 1372, 1250, 1095, 1062.

Compound 5: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.57 (m, 1H), 3.47 (m, 1H), 3.37 (m, 1H), 3.12 (m, 1H), 2.58 (m, 1H), 2.39 (dd, *J* = 13.5, 5.6, 1H), 2.24 (bs, 1H), 2.17 (q, *J* = 7.2, 1H), 1.19 (d, *J* = 7.2, 3H), 0.89 (s, 9H), 0.83 (s, 3H), 0.80 (d, *J* = 6.8, 3H), 0.54 (s, 3H), 0.06 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  214.9 (C), 183.9 (C), 134.1 (C), 72.0 (CH), 66.0 (CH<sub>2</sub>), 65.8 (CH), 55.9 (CH), 54.7 (CH), 45.0 (CH), 42.8 (CH), 41.2 (C), 38.6 (CH<sub>2</sub>), 37.9 (CH<sub>2</sub>), 37.1 (CH<sub>2</sub>), 35.7 (C), 35.2 (CH), 35.0 (CH), 32.2 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 28.5 (CH<sub>2</sub>), 28.0 (CH<sub>2</sub>), 26.2 (CH<sub>2</sub>), 25.9 (3xCH<sub>3</sub>), 20.9 (CH<sub>2</sub>), 18.3 (C), 16.4 (CH<sub>3</sub>), 14.5 (CH<sub>3</sub>), 12.4 (CH<sub>3</sub>), 12.3 (CH<sub>3</sub>), -4.6 (2xCH<sub>3</sub>); ESI-MS 529 [M+H]<sup>+</sup>. HRMS calculated for C<sub>33</sub>H<sub>57</sub>O<sub>3</sub>Si (M+H)<sup>+</sup>, 529.4071; found 529.4062; IR ATR,  $\nu_{max}$  (cm<sup>-1</sup>): 3431, 1697, 1654, 1456, 1373, 1248, 1080, 834, 772.

#### 3.2.3. General Procedure for Imine 3 Reduction with Complex Sodium Hydride

To the stirred solution of imine **3** (1 equiv.) in the proper solvent, reducing agents (NaBH<sub>4</sub>, NaBH<sub>3</sub>CN) and additives (NaOAc, I<sub>2</sub>, AcOH) were added. The detailed reaction conditions are indicated in Table 1. The reaction mixture was monitored by TLC. The reaction mixture was poured into water and extracted with CHCl<sub>3</sub>. The extract was washed with water, dried over anhydrous sodium sulfate, and the solvent was evaporated. The crude products (**4**, **7**, **8**) were isolated by silica gel column chromatography.

Compound 7, eluted with 8% MeOH/CHCl<sub>3</sub>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  3.75 (m, 1H), 3.51 (m, 2H), 3.37 (m, 1H), 2.88 (m, 1H), 2.01 (m, 1H), 1.01 (d, *J* = 6.4, 3H), 0.88 (s, 9H), 0.864 (s, 3H), 0.859 (d, *J* = 6.3, 3H), 0.80 (s, 3H), 0.05 (s, 6H); <sup>13</sup>C NMR (100 MHz, 100 MHz).

CDCl<sub>3</sub>)  $\delta$  72.1 (CH), 70.4 (CH), 67.3 (CH<sub>2</sub>), 63.2 (CH), 62.2 (CH), 57.6 (CH), 54.3 (CH), 45.0 (CH), 41.2 (C), 40.0 (CH<sub>2</sub>), 38.6 (CH<sub>2</sub>), 38.4 (CH), 37.1 (CH<sub>2</sub>), 35.6 (C), 34.8 (CH), 34.7 (CH), 32.3 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 30.8 (CH<sub>2</sub>), 29.6 (CH<sub>2</sub>), 28.6 (CH<sub>2</sub>), 27.9 (CH<sub>2</sub>), 25.9 (3xCH<sub>3</sub>), 20.9 (CH<sub>2</sub>), 18.2 (CH<sub>3</sub>), 18.1 (C), 17.0 (CH<sub>3</sub>), 16.0 (CH<sub>3</sub>), 12.3 (CH<sub>3</sub>), -4.6 (2xCH<sub>3</sub>); ESI-MS 532 [M+H]<sup>+</sup>. HRMS calculated for C<sub>33</sub>H<sub>62</sub>NO<sub>2</sub>Si (M+H)<sup>+</sup>, 532.4544; found 532.4559; IR ATR,  $\nu_{max}$  (cm<sup>-1</sup>): 3288, 1454, 1368, 1247, 1092.

Compound 8 (obtained by reduction with NaBH<sub>3</sub>CN, Table 1, entry 6), eluted with 45% AcOEt/hexane: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  4.66 (m, 1H), 3,62–3.50 (m, 3H), 3.07 (q, *J* = 7.4, 1H), 2.91 (dd, *J* = 12.4, 4.3, 1H), 2.48–2.35 (m, 2H), 1.83 (d, *J* = 8.4, 1H), 1.22 (d, *J* = 7.4, 3H), 1.00 (d, *J* = 6.6, 3H), 0.89 (s, 9H), 0.79 (s, 3H), 0.57 (s, 3H), 0.05 (s, 6H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  191. 4 (C), 76.9 (CH), 72.0 (CH), 66.7 (CH<sub>2</sub>), 57.6 (CH), 54.4 (CH), 54.0 (CH), 44.9 (CH), 43.6 (CH), 42.3 (C), 38.52 (CH<sub>2</sub>), 38.49 (CH<sub>2</sub>), 37.2 (CH<sub>2</sub>), 35.8 (CH), 35.6 (C), 34.9 (CH), 32.2 (CH<sub>2</sub>), 31.9 (CH<sub>2</sub>), 31.2 (CH<sub>2</sub>), 28.7 (CH<sub>2</sub>), 28.4 (CH<sub>2</sub>), 27.6 (CH<sub>2</sub>), 25.9 (3xCH<sub>3</sub>), 20.5 (CH<sub>2</sub>), 18.5 (CH<sub>3</sub>), 18.2 (C), 16.3 (CH<sub>3</sub>), 14.6 (CH<sub>3</sub>), 12.3 (CH<sub>3</sub>), -4.57 (CH<sub>3</sub>), -4.59 (CH<sub>3</sub>); <sup>11</sup>B NMR (128 MHz, CDCl<sub>3</sub>)  $\delta$  -24.52; ESI-MS 1159 [2M+Na]<sup>+</sup>. IR ATR,  $\nu_{max}$  (cm<sup>-1</sup>): 3468, 2401, 1636, 1458, 1249, 1093, 1056.

## 3.2.4. Synthesis of Compound 9

To a solution of 4 (19 mg, 0.036 mmol) in dichloromethane (2 mL) at -15 °C, Et<sub>3</sub>N (0.01 mL, 7.3 mg, 0.072 mmol) and 0.22 mL of solution of MsCl (0.03 mL) in dichloromethane (2 mL) were added, successively. The reaction mixture was continuously stirred at -15 °C for 1.5 h and quenched by adding aqueous NaHCO<sub>3</sub>, and the layers were separated and the aqueous layer was extracted with chloroform. The organic layers were combined, dried over Na<sub>2</sub>SO<sub>4</sub>, and evaporated under reduced pressure. The crude product was purified by column chromatography (20% MeOH/CHCl<sub>3</sub>) to obtain compound **9** (45%).

Compound 9: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  5.45 (m, 1H), 4.25 (m, 1H), 3.61 (m, 1H), 3.13-3.00 (m, 3H), 2.75 (s, 3H), 2.72 (m, 1H), 2.62 (m, 1H), 2.32 (m, 1H), 2.18 (d, *J* = 6.1, 1H), 1.57 (d, *J* = 7.1, 3H), 1.08 (d, *J* = 5.9, 3H), 0.81 (s, 3H), 0.62 (s, 3H);  $\delta$  <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  190.6 (C), 75.7 (CH), 71.0 (CH), 57.0 (CH), 54.3 (CH), 54.0 (CH), 52.7 (CH<sub>2</sub>), 44.6 (CH), 44.5 (CH), 42.0 (C), 39.4 (CH<sub>3</sub>), 38.2 (CH<sub>2</sub>), 38.0 (CH<sub>2</sub>), 36.9 (CH<sub>2</sub>), 35.5 (C), 35.0 (CH), 32.0 (CH<sub>2</sub>), 31.3 (CH<sub>2</sub>), 29.2 (CH<sub>2</sub>), 28.3 (CH<sub>2</sub>), 25.84 (CH), 25.82 (CH<sub>2</sub>), 24.6 (CH<sub>2</sub>), 20.5 (CH<sub>2</sub>), 18.2 (CH<sub>3</sub>), 18.1 (CH<sub>3</sub>), 14.6 (CH<sub>3</sub>), 12.3 (CH<sub>3</sub>); ESI-MS 398 [M<sup>+</sup>]; IR ATR,  $\nu_{max}$  (cm<sup>-1</sup>): 3377, 1664, 1628, 1456, 1195, 1043.

#### 3.2.5. Synthesis of 25-epidemissidine (10)

To the stirred ice-cooled solution of compound **9** (20 mg, 0.04 mmol) in MeOH (2 mL)/ DCM (2 mL), NaBH<sub>4</sub> (4.6 mg, 0.12 mmol) was added. The stirring of the reaction mixture was continued at -10 - 0 °C for 0.5 h. The reaction mixture was poured into water and extracted with CHCl<sub>3</sub>. The extract was washed with water, dried over anhydrous sodium sulfate, and the solvent was evaporated. The crude product was purified by column chromatography (20% AcOEt/hexane) to obtain compound **10** (71%), identical to that described in ref. [21].

Compound **10**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 3.60 (m, 1H), 2.59 (m, 2H), 1.03 (d, *J* = 7.0, 3H), 0.92 (d, *J* = 6.7, 3H), 0.85 (s, 3H), 0.82 (s, 3H).

## 3.2.6. Synthesis of Compound 12

Acetyl chloride (0.027 mL, 29 mg, 0.38 mmol) was added to the stirred, ice-cold solution of compound 4 (20 mg, 0.038 mmol) in dry MeOH (3 mL). The reaction mixture was stirred for 3 h and allowed to warm up to room temperature. Then, the solvent was evaporated under reduced pressure and the residue was dissolved in dichloromethane (2 mL) and THF (2 mL). To the obtained suspension, triethylamine (0.02 mL) and 0.58 mL of a solution of MsCl (0.02 mL) in  $CH_2Cl_2$  (2 mL) were added. The reaction mixture was stirred overnight, allowing it to warm up to room temperature. After this time, the mixture was poured into aqueous NaHCO<sub>3</sub> and extracted with CHCl<sub>3</sub>. The extract was washed

with water, dried over anhydrous sodium sulfate, and the solvent was evaporated. The crude product was purified by column chromatography (35% AcOEt/hexane) to afford compound **12** (53%).

Compound **12**: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  4.62 (m, 1H), 4.04 (m, 1H), 3.61 (q, *J* = 7.0, 1H), 3.02 (m, 1H), 3.00 (s, 3H), 2.83 (s, 3H), 2.74 (m, 1H), 2.50 (d, *J* = 12.5, 1H), 1.24 (d, *J* = 7.0, 3H), 1.04 (d, *J* = 6.1, 3H), 0.83 (s, 3H), 0.61 (s, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  160.9 (C), 88.9 (C), 81.6 (CH), 65.4 (CH), 60.5 (CH), 55.0 (CH), 54.1 (CH), 49.0 (CH<sub>2</sub>), 44.8 (CH), 43.0 (CH<sub>3</sub>), 41.8 (C), 38.9 (CH<sub>3</sub>), 38.2 (CH<sub>2</sub>), 36.7 (CH<sub>2</sub>), 36.6 (CH), 35.3 (C), 35.1 (CH<sub>2</sub>), 34.9 (CH), 32.0 (CH<sub>2</sub>), 30.8 (CH<sub>2</sub>), 30.7 (CH<sub>2</sub>), 28.6 (CH<sub>2</sub>), 28.3 (CH<sub>2</sub>), 26.8 (CH), 23.0 (CH<sub>3</sub>), 20.5 (CH<sub>2</sub>), 18.7 (CH<sub>3</sub>), 13.5 (CH<sub>3</sub>), 12.1 (CH<sub>3</sub>); ESI-MS 554 [M+H]<sup>+</sup>, 1129 [2M+Na]<sup>+</sup>. HRMS calculated for C<sub>29</sub>H<sub>48</sub>NO<sub>5</sub>S<sub>2</sub> (M+H)<sup>+</sup> 554.2968; found 554.2965; IR ATR,  $\nu_{max}$  (cm<sup>-1</sup>): 1658, 1453, 1333, 1212, 1163, 1036, 925.

#### 4. Conclusions

In summary, we developed a novel, concise synthesis of solanidanes from the spirostane sapogenin tigogenin. The indolizidine moiety present in solanidane-type alkaloids was constructed from spirostane in five steps involving tigogenin oxidation, amination, reduction, mesylation, and reduction again. The key intermediate for the proposed approach was spiroimine obtained in the reaction of a 5,6-dihydrokrytogenin derivative with aminoalane generated in situ from DIBAlH and NH<sub>4</sub>Cl. Depending on mesylation conditions, two different solanidanes were obtained: the indolizinium salt **9a**, which was readily converted into 25-epidemissidine (**10**), and the 23-sulfone derivative **12**, a convenient intermediate for further derivatization.

**Supplementary Materials:** The following are available online at https://www.mdpi.com/article/10.3390/ijms221910879/s1, <sup>1</sup>H NMR, <sup>13</sup>C NMR spectra of compounds **3–10** and **12**.

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#### References

- 1. Attaur, R.; Choudhary, M.I. Diterpenoid and steroidal alkaloids. Nat. Prod. Rep. 1997, 14, 191–203. [CrossRef]
- Li, H.J.; Jiang, Y.; Li, P. Chemistry, bioactivity and geographical diversity of steroidal alkaloids from the *Liliaceae* family. *Nat. Prod. Rep.* 2006, 23, 735–752. [CrossRef]
- 3. Jiang, Q.-W.; Chen, M.-W.; Cheng, K.-J.; Yu, P.-Z.; Wei, X.; Shi, Z. Therapeutic potential of steroidal alkaloids in cancer and other diseases. *Med. Res. Rev.* 2016, *36*, 119–143. [CrossRef] [PubMed]
- 4. Friedman, M.; Lee, K.R.; Kim, H.J.; Lee, I.S.; Kozukue, N. Anticarcinogenic effects of glycoalkaloids from potatoes against human cervical, liver, lymphoma, and stomach cancer cells. *J. Agric. Food Chem.* **2005**, *53*, 6162–6169. [CrossRef] [PubMed]
- Lee, K.R.; Kozukue, N.; Han, J.S.; Park, J.H.; Chang, E.Y.; Baek, E.J.; Chang, J.S.; Friedman, M. Glycoalkaloids and metabolites inhibit the growth of human colon (HT29) and liver (HepG2) cancer cells. J. Agric. Food. Chem. 2004, 52, 2832–2839. [CrossRef]
- 6. Tingey, W.M. Glycoalkaloids as pest resistance factors. *Am. J. Potato. Res.* **1984**, *61*, 157–167. [CrossRef]
- Schreiber, K. Steroid alkaloids: The Solanum group. In The Alkaloids: Chemistry and Physiology; Manske, R.H.F., Ed.; Academic Press: New York, NY, USA, 1968; Volume 10, pp. 1–192.
- 8. Roddick, J.G. The acetylcholinesterase-inhibitory activity of steroidal glycoalkaloids and their aglycones. *Phytochemistry* **1989**, *28*, 2631–2634. [CrossRef]
- 9. Roddick, J.G.; Weissenberg, M.; Leonard, A.L. Membrane disruption and enzyme inhibition by naturally-occurring and modified chacotriose-containing *Solanum* steroidal glycoalkaloids. *Phytochemistry* **2001**, *56*, 603–610. [CrossRef]

- Mótyán, G.; Kovács, F.; Wölfling, J.; Gyovai, A.; Zupkó, I.; Frank, É. Microwave-assisted stereoselective approach to novel steroidal ring D-fused 2-pyrazolines and an evaluation of their cell-growth inhibitory effects in vitro. *Steroids* 2016, 112, 36–46. [CrossRef] [PubMed]
- 11. Abdalla, M.M.; Al-Omar, M.A.; Bhat, M.A.; Amr, A.-G.E.; Al-Mohizeae, A.M. Steroidal pyrazolines evaluated as aromatase and quinone reductase-2 inhibitors for chemoprevention of cancer. *Int. J. Biol. Macromol.* **2012**, *50*, 1127–1132. [CrossRef]
- Schuster, D.; Wolber, G. Identification of bioactive natural products by pharmacophore-based virtual screening. *Curr. Pharm. Des.* 2010, 16, 1666–1681. [CrossRef] [PubMed]
- 13. Kuhn, R.; Low, I.; Trischmann, H. Uberfuhrung von tomatidin in demissidin. Angew. Chem. 1952, 64, 397–397. [CrossRef]
- 14. Sato, Y.; Latham, H.G. Chemistry of dihydrotomatidines. J. Am. Chem. Soc. 1956, 78, 3146–3150. [CrossRef]
- 15. Schreiber, K.; Ronsch, H. Solanum-alkaloide-LIV. Synthese von solanidin und 22-iso-solanidin aus tomatid-5-en-3β-ol. *Tetrahedron* **1965**, *21*, 645–650. [CrossRef]
- 16. Adam, G.; Schreiber, K. Synthese des steroid alkaloids demissidin aus 3β-acetoxy-pregn-5-en-20-on; aufbau des solanidinegerustes durch Hofmann-Loffler-Freytag-cyclisierung. *Tetrahedron Lett.* **1963**, *4*, 943–948. [CrossRef]
- 17. Zhang, Z.; Giampa, G.M.; Draghici, C.; Huang, Q.; Brewer, M. Synthesis of demissidine by a ring fragmentation 1,3-dipolar cycloaddition approach. *Org. Lett.* **2013**, *15*, 2100–2103. [CrossRef]
- 18. Zhang, Z.-D.; Shi, Y.; Wu, J.-J.; Lin, J.-R.; Tian, W.-S. Synthesis of demissidine and solanidine. *Org. Lett.* **2016**, *18*, 3038–3040. [CrossRef]
- 19. Hou, L.-L.; Shi, Y.; Zhang, Z.-D.; Wu, J.-J.; Yang, Q.-X.; Tian, W.-S. Divergent synthesis of solanidine and 22-epi-solanidine. *J. Org. Chem.* **2017**, *82*, 7463–7469. [CrossRef]
- 20. Wang, Y.; Huang, G.; Shi, Y.; Tian, W.-S.; Zhuang, C.; Chen, E.-F. Asymmetric synthesis of (–)-solanidine and (–)-tomatidenol. *Org. Biomol. Chem.* **2020**, *18*, 3169–3176. [CrossRef]
- Rivas-Loaiza, J.A.; Baj, A.; López, Y.; Witkowski, S.; Wojtkielewicz, A.; Morzycki, J.W. Synthesis of solanum alkaloid demissidine stereoisomers and analogues. J. Org. Chem. 2021, 86, 1575–1582. [CrossRef]
- 22. Cheng, M.S.; Wang, Q.L.; Tian, Q.; Song, Y.H.; Liu, Y.X.; Li, Q.; Xu, X.; Miao, H.D.; Yao, X.S.; Yang, Z. Total synthesis of methyl protodioscin: A potent agent with antitumor activity. *J. Org. Chem.* **2003**, *68*, 3658–3662. [CrossRef] [PubMed]
- Lee, J.S.; Cao, H.; Fuchs, P.L. Ruthenium-catalyzed mild C–H oxyfunctionalization of cyclic steroidal ethers. J. Org. Chem. 2007, 72, 5820–5823. [CrossRef] [PubMed]
- 24. Yu, B.; Liao, J.; Zhang, J.; Hui, Y. The first synthetic route to furostan saponins. Tetrahedron Lett. 2001, 42, 77–79. [CrossRef]
- 25. Bovicelli, P.; Lupettelli, P.; Fracas, D. Sapogenins and dimethyldioxirane: A new entry to cholestanes functionalized at the side chain. *Tetrahedron Lett.* **1994**, *35*, 935–938. [CrossRef]
- 26. Overman, L.E.; Sugai, S. A Convenient Method for obtaining *trans*-2-aminocyclohexanol and *trans*-2-aminocyclopentanol in enantiomerically pure form. *J. Org. Chem.* **1985**, *50*, 4154–4155. [CrossRef]
- 27. Overman, L.E.; Flippin, L.A. Facile aminolysis of epoxides with diethylaluminum amides. *Tetrahedron Lett.* **1981**, 22, 195–198. [CrossRef]
- 28. Ashworth, I.W.; Bowden, M.C.; Dembofsky, B.; Levin, D.; Moss, W.; Robinson, E.; Szczur, N.; Virica, J. A new route for manufacture of 3-cyano-1-naphthalenecarboxylic acid. *Org. Process. Res. Dev.* **2003**, *7*, 74–81. [CrossRef]
- Veerasamy, N.; Carlson, E.C.; Collett, N.D.; Saha, M.; Carter, R.G. Enantioselective approach to quinolizidines: Total synthesis of cermizine D and formal syntheses of senepodine G and cermizine C. J. Org. Chem. 2013, 78, 4779–4800. [CrossRef]
- 30. Bakunova, S.M.; Bakunov, S.A.; Wenzler, T.; Barszcz, T.; Werbovetz, K.A.; Brun, R.; Hall, J.E.; Tidwell, R.R. Synthesis and in vitro antiprotozoal activity of bisbenzofuran cations. *J. Med. Chem.* 2007, *50*, 5807–5823. [CrossRef]
- Huang, S.-Q.; Zheng, X.; Deng, X.-M. DIBAL-H-H<sub>2</sub>NR and DIBAL-H-HNR<sub>1</sub>R<sub>2</sub>·HCl complexes for efficient conversion of lactones and esters to amides. *Tetrahedron Lett.* 2001, 42, 9039–9041. [CrossRef]
- 32. Wojtkielewicz, A.; Łotowski, Z.; Morzycki, J.W. One-step synthesis of nitriles from acids, esters and amides using DIBAL-H and ammonium chloride. *Synlett* 2015, *26*, 2288–2292. [CrossRef]
- 33. Bechara, W.S.; Charette, A.B.; Na, R.; Wang, W.; Zheng, C. Diethyl 1,4-dihydro-2,6-dimethyl-3,5-pyridinedicarboxylate. In *Encyclopedia of Reagents for Organic Synthesis* 2020; John Wiley & Sons, Ltd.: Hoboken, NJ, USA, 2020. [CrossRef]
- 34. Czajkowska-Szczykowska, D.; Jastrzebska, I.; Rode, J.E.; Morzycki, J.W. Revision of the structure of *N*,*O*-diacetylsolasodine. Unusual epimerization at the spiro carbon atom during acetylation of solasodine. *J. Nat. Prod.* **2019**, *82*, 59–65. [CrossRef]
- 35. Chagnon, F.; Guay, I.; Bonin, M.-A.; Mitchell, G.; Bouarab, K.; Malouin, F.; Marsault, É. Unraveling the structure-reactivity relationship of tomatidine, a steroid alkaloid with unique antibiotic properties against persistent forms of *Staphylococcus aureus*. *Eur. J. Med. Chem.* **2014**, *80*, 605–620. [CrossRef]
- 36. Zha, X.M.; Zhang, F.R.; Shan, J.Q.; Zhang, Y.H.; Liu, J.O.; Sun, H.B. Synthesis and evaluation of in vitro anticancer activity of novel solasodine derivatives. *Chin. Chem. Lett.* **2010**, *21*, 1087–1090. [CrossRef]
- 37. Uhle, F.C.; Sallmann, F. The synthesis and transformations of a steroid pyrroline Derivative. J. Am. Chem. Soc. **1960**, 82, 1190–1199. [CrossRef]
- 38. Reddy, C.R.; Ramesh, P.; Latha, B. Formal syntheses of 5,8-disubstituted indolizidine alkaloids (–)-205A, (–)-207A, and (–)-235B. *Synlett* **2016**, 27, 481–484. [CrossRef]

- 39. Donohoe, T.J.; Sintim, H.O.; Hollinshead, J. A Noncarbohydrate Based Approach to Polyhydroxylated. *J. Org. Chem.* 2005, 70, 7297–7304. [CrossRef]
- 40. Mátyus, P.; Szilágyi, G.; Kasztreiner, E.; Sohár, P. Studies on pyridazine derivatives. IV. Mesylation reaction of pyridazinylpyrazoles. *J. Heterocycl. Chem.* **1980**, 17, 781–783. [CrossRef]
- 41. Back, T.G.; Hu, N.-X. Synthesis of Antibiotic A25822 B. Tetrahedron 1993, 49, 337–348. [CrossRef]
- 42. Cheng, S.; Du, Y.; Mac, B.; Tanc, D. Total synthesis of a furostan saponin, timosaponin BII. Org. Biomol. Chem. 2009, 7, 3112–3118. [CrossRef]