



Clinical value of procalcitonin for suspected nosocomial bloodstream infection

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Background/Aims: Procalcitonin (PCT) may prove to be a useful marker to exclude or predict bloodstream infection (BSI). However, the ability of PCT levels to differentiate BSI from non-BSI episodes has not been evaluated in nosocomial BSI.

Methods: We retrospectively reviewed the medical records of patients ≥ 18 years of age with suspected BSI that developed more than 48 hours after admission.

Results: Of the 785 included patients, 105 (13.4%) had BSI episodes and 680 (86.6%) had non-BSI episodes. The median serum PCT level was elevated in patients with BSI as compared with those without BSI (0.65 ng/mL vs. 0.22 ng/mL, $p = 0.001$). The optimal PCT cut-off value of BSI was 0.27 ng/mL, with a corresponding sensitivity of 74.6% (95% confidence interval [CI], 66.4% to 81.7%) and a specificity of 56.5% (95% CI, 52.7% to 60.2%). The area under curve of PCT (0.692) was significantly larger than that of C-reactive protein (CRP; 0.526) or white blood cell (WBC) count (0.518). However, at the optimal cut-off value, PCT failed to predict BSI in 28 of 105 cases (26.7%). The PCT level was significantly higher in patients with an eGFR < 60 mL/min/1.73 m² than in those with an eGFR ≥ 60 mL/min/1.73 m² (0.68 vs. 0.17, $p = 0.01$).

Conclusions: PCT was more useful for predicting nosocomial BSI than CRP or WBC count. However, the diagnostic accuracy of predicting BSI remains inadequate. Thus, PCT is not recommended as a single diagnostic tool to avoid taking blood cultures in the nosocomial setting.

Keywords: Procalcitonin; Nosocomial bloodstream infection; Renal function

Received: April 18, 2016

Revised: September 9, 2016

Accepted: September 13, 2016

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INTRODUCTION

Bloodstream infection (BSI) in hospitalized patients is a common and deadly problem. Between 0.5% and 1.3% of patients develop BSI during hospitalization [1]. In-hospital mortality estimates among patients with BSI vary between 30% and 40% [2,3]. Nosocomial BSI is associated with significantly higher mortality rates than community-onset BSI [4-6]. Early administration of effective antimicrobial therapy to patients with BSI has

repeatedly been shown to improve mortality in patients with community-acquired and nosocomial BSI [3,7-9]. Therefore, early diagnosis and appropriate treatment is important especially in patients with nosocomial BSI.

Accurate and timely diagnosis of BSI remains challenging to both clinicians and laboratories. The most specific diagnostic modality for BSI is blood culture; however, it usually takes more than 24 hours to obtain results [10]. Many indirect diagnostic modalities have been used to indicate suspected BSI, including fever,

white blood cell (WBC) count, and C-reactive protein (CRP) level. However, these approaches have poor sensitivity and specificity. More accurate diagnostic tools are needed [11-13].

Elevation in serum procalcitonin (PCT) levels above normal values in patients with sepsis and other clinically significant bacterial infections was first described approximately 25 years ago [14]. PCT level is not elevated in viral infections. PCT has recently been proposed as a useful marker to rule out BSI in the emergency department and to guide antimicrobial therapy in patients hospitalized for community-acquired BSI [8,15,16]. Patients with community-onset BSI have been reported to have higher PCT levels as compared to those with nosocomial BSI [4]. Although mortality is much higher for patients with nosocomial BSI, PCT has not yet been studied in a large nosocomial BSI patient population. Accordingly, we investigated the usefulness of PCT as a diagnostic marker in patients with suspected nosocomial BSI, and assessed the relationship between PCT and renal function.

METHODS

Patients and population

We retrospectively reviewed medical records taken from January 2013 to November 2014 in Dongguk University Ilsan Hospital. Patients fulfilling the criteria of systemic inflammatory response (SIRS) with positive peripheral blood cultures were enrolled by clinicians 48 hours after admission. SIRS was defined in patients with two or more of the following conditions: (1) fever (oral temperature $> 38^{\circ}\text{C}$) or hypothermia ($< 36^{\circ}\text{C}$); (2) tachypnea (> 24 breaths/min); (3) tachycardia (heart rate > 90 beats/min); and (4) leukocytosis ($> 12,000/\mu\text{L}$), leukopenia ($< 4,000/\mu\text{L}$), or $> 10\%$ bands [17]. We excluded patients under 18 years of age. This study was approved by the Ethics Review Board of Dongguk University Ilsan Hospital, Korea (2015-110). The need for informed consent was waived because of the retrospective and observational nature of this study.

Basic characteristics, medical history, laboratory data including WBC count, CRP, PCT, and serum creatinine level were reviewed. Estimated glomerular filtration rate (eGFR) was determined by the Chronic Kidney Disease

Epidemiology Collaboration (CKD-EPI) formula [18]. Records were obtained from the previous administration of antibiotics at the time of blood culture in patients with positive blood cultures.

Laboratory examinations

We performed two or more blood cultures and a PCT test concomitantly on all enrolled patients with SIRS. If the results of the blood culture were positive, the microorganisms were identified and recorded. Blood cultures utilized the BACTEC FX automated system (BD, Sparks, MD, USA). Microorganisms were identified using standard procedures. Antimicrobial susceptibility tests were performed using a Phoenix automated system (BD). Blood samples were obtained from each patient such that samples fulfilled the SIRS criteria and blood cultures and levels of serum PCT and CRP were obtained at the first blood sample obtained for culture. CRP and PCT were tested using a Cobas e602 apparatus (Roche Diagnostics, Mannheim, Germany). The analytical reportable range of the PCT assay was 0.02 to 100 ng/mL. A BSI episode was defined as a positive blood culture with appropriate clinical correlation. Coagulase-negative staphylococci, *Bacillus* species, *Corynebacteria* that were isolated in only one blood culture were considered contaminants [15,19,20]. Contaminants and negative blood cultures were defined as non-BSI episodes.

Statistical analyses

Statistical analyses were performed with IBM SPSS version 20 (IBM Co., Armonk, NY, USA). Demographic data, PCT level, CRP, WBC count, renal function, and patient outcomes were compared between the BSI episode and non-BSI episode groups by chi-square test or Mann-Whitney *U* test. We plotted the receiver operating characteristic curves of PCT, CRP, WBC level, and the diagnostic accuracy of assessed by area under receiver operating characteristic curves (AUCs). The optimal cut-off value of PCT was identified by maximizing the AUC.

RESULTS

A total of 814 patients were identified during the study period. Of these, 163 patients displayed positive blood cultures. Among them, 29 cases were regarded as con-

tamination and 29 patients were already receiving antimicrobial treatment before blood culture. We excluded 58 patients from the BSI group. Thus, 105 patients were allocated to the BSI group and 680 patients were allocated

to the non-BSI group.

The underlying disease and origin of infection of the 105 patients in the BSI group are presented in Table 1. The median age of the study patients was 71 years (interquartile range [IQR], 58 to 79). The median Charlson comorbidity index was 3 (IQR, 2 to 6) and 47 patients (44.8%) had solid tumors, while 28 patients (26.7%) had respiratory infections. The baseline characteristics of each group are presented in Table 2. WBC count was not elevated in the BSI group compared to the non-BSI group. In contrast, elevation was evident in the median level of PCT (0.65 ng/mL vs. 0.22 ng/mL, $p = 0.001$). Table 3 presents the identification data of the isolated microorganisms. In the BSI group, gram-positive bacteria, gram-negative bacteria, and fungi were isolated from 44 (32.8%), 48 (35.8%), and nine patients (6.7%), respectively. Four patients (3.0%) had multiple types of isolated bacteria.

Table 4 presents comparative data of PCT, CRP, WBC count stratified by the isolated microorganisms. A statistical difference was evident between gram-negative bacteria and gram-positive bacteria (0.90 [IQR, 0.32 to 4.09] vs. 0.44 [IQR, 0.16 to 1.83], $p = 0.01$).

We compared the AUC of PCT, CRP, and WBC count to determine diagnostic accuracy. The AUCs of PCT, CRP, and WBC count were 0.69, 0.53, and 0.52, respectively (Fig. 1). Table 5 presents data concerning the number of cases under the cut-off value, sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV) for eight PCT cut-off values. The optimal cut-off value was 0.27 ng/mL. The sensitivity, specificity, PPV, and NPV of the optimal cut-off value (0.27 ng/mL)

Table 1. Characteristics of patients with blood stream infections

Characteristic	Patients (n = 105)
Age, yr	71 (58–79)
Male sex	45 (57.1)
McCabe Jackson criteria	
Nonfatal	46 (43.8)
Ultimately fatal	59 (56.2)
Rapidly fatal disease	0 (0)
Charlson comorbidity index	3 (2–6)
Underlying disease	
Solid tumor	47 (44.8)
Neurologic disease	38 (36.2)
Diabetes mellitus	36 (34.3)
Chronic kidney disease	20 (19.0)
Chronic lung disease	14 (13.3)
Liver cirrhosis	13 (12.4)
Heart disease	11 (10.4)
Portal of entry	
Respiratory infection	28 (26.7)
Unknown primary	20 (19.0)
Gastrointestinal infection	18 (17.9)
Genitourinary infection	17 (16.2)
Catheter related blood stream infection	16 (15.2)
Musculoskeletal infection	5 (4.8)
Severe sepsis or septic shock	54 (40.3)

Values are presented as median (interquartile range) or number (%).

Table 2. Baseline characteristics of the study population in relation to the presence of bloodstream infection

Characteristic	Bloodstream infection group (n = 105)	Non-bloodstream infection group (n = 680)	p value
Age, yr	71.0 (58.0–79.0)	70.0 (56.0–78.0)	0.16
Male sex	45 (42.8)	368 (54.1)	0.03
Creatinine, mg/dL	0.81 (0.64–1.35)	0.73 (0.56–1.12)	0.87
eGFR, mL/min/1.73 m ²	78.0 (41.5–98.4)	89.0 (56.0–107.0)	0.01
PCT, ng/mL	0.65 (0.23–2.76)	0.22 (0.10–0.70)	0.001
CRP, mg/dL	5.44 (2.02–9.73)	4.59 (1.79–9.36)	0.30
WBC count, /μL	10,180 (5,535–14,730)	9,795 (6,788–13,060)	0.25
30-Day mortality	26 (24.8)	48 (7.1)	< 0.01

Values are presented as median (interquartile range) or number (%).

eGFR, estimated glomerular filtration rate; PCT, procalcitonin; CRP, C-reactive protein; WBC, white blood cell.

were 74.6%, 56.5%, 25.3%, and 91.9%, respectively. At the time of blood culture sampling, PCT was < 0.1 ng/mL in seven patients with BSI. PCT was < 0.27 ng/mL in 28 patients with BSI.

The PCT level was higher in patients with renal insufficiency than in those with normal renal function, as shown in Table 6. The PCT level was significantly higher in patients with an eGFR < 60 mL/min/1.73 m² than in those with an eGFR ≥ 60 mL/min/1.73 m² (0.68 [IQR, 0.28 to 2.58] vs. 0.17 [IQR, 0.09 to 0.49], *p* = 0.01). There

was no statistically significant difference in PCT levels between patients with eGFR < 30 and 30 ≤ eGFR < 60 mL/min/1.73 m².

DISCUSSION

Our study demonstrates a higher PCT level in patients with nosocomial BSI, and shows that an elevated PCT level was more useful for predicting nosocomial BSI than were CRP or WBC count. However, BSI could not be excluded, even if the PCT level was low (0.1 ng/mL). In addition, PCT was significantly correlated with eGFR,

Table 3. Microorganisms isolated from blood culture

Microorganism	Isolates
Gram-positive bacteria	44 (32.8)
<i>Staphylococcus aureus</i>	25
<i>Streptococcus</i> species	1
<i>Enterococcus</i> species	9
Other gram-positive bacteria	9
Gram-negative bacteria	48 (35.8)
<i>Escherichia coli</i>	16
<i>Klebsiella</i> species	13
<i>Acinetobacter</i> species	8
<i>Pseudomonas aeruginosa</i>	3
Other gram-negative bacteria	8
Fungi	9 (6.7)
<i>Candida albicans</i>	3
Other <i>Candida</i> species	6
Multiple microbes	4 (3.0)
Contaminant bacteria	29 (21.6)
Coagulase-negative staphylococci	23
<i>Bacillus</i> species	3
Other contaminant bacteria	3

Values are presented as number (%).

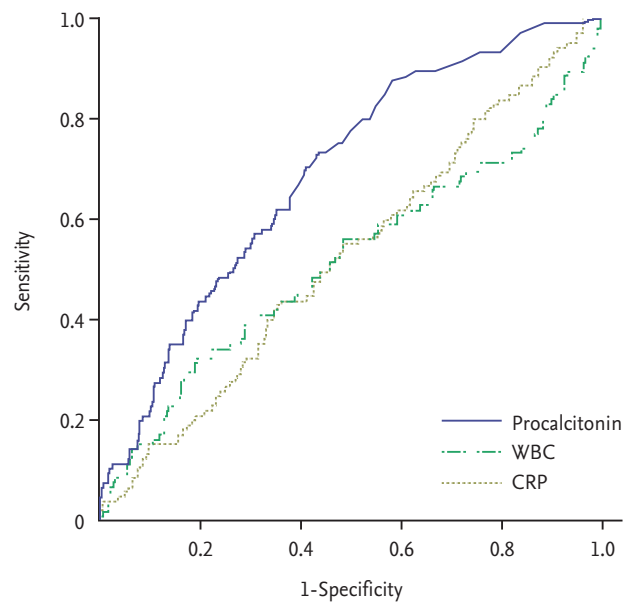


Figure 1. Receiver operating characteristic curves of procalcitonin (PCT), C-reactive protein (CRP), and white blood cell (WBC) count. PCT, CRP, and WBC had areas under the curve of 0.692, 0.526, and 0.518, respectively.

Table 4. Comparisons of biochemical data from 785 blood tests according to blood culture results

Variable	Non-bloodstream infection		Bloodstream infection			
	Negative (n = 651)	Contamination (n = 29)	Gram-positive (n = 44)	Gram-negative (n = 48)	Multiple microbes (n = 4)	Fungi (n = 9)
PCT, ng/mL	0.21 (0.10–0.70)	0.26 (0.09–0.61)	0.44 (0.16–1.83)	0.90 (0.32–4.09)	0.24 (0.12–0.49)	1.01 (0.34–2.74)
CRP, mg/dL	4.67 (1.81–9.48)	2.93 (1.22–8.13)	5.92 (2.19–10.22)	4.97 (1.58–9.30)	3.31 (1.24–8.56)	5.44 (4.10–11.30)
WBC count, /μL	9,820 (6,780–13,060)	9,160 (6,780–11,245)	10,950 (8,020–16,480)	9,850 (5,000–13,555)	8,470 (3,490–14,447)	9,870 (4,765–17,840)

Values are presented as median (interquartile range).

PCT, procalcitonin; CRP, C-reactive protein; WBC, white blood cell.

and the optimum cut-off value of PCT for predicting a nosocomial BSI increased along with the deterioration of renal function.

Hoeningl et al. [21] reported a PCT sensitivity of 70% and 60% for community-acquired and nosocomial BSI, respectively. However, the authors could not demonstrate positive or NPV owing to limited study population. Previous studies reported a significantly lower PCT level in patients with nosocomial BSI than in those with community-acquired BSI [21,22]. The significantly higher levels of PCT in patients with community-onset BSI might derive from a time delay between the onset of the first symptoms and drawing of blood for laboratory

analysis [23]. Other suggestions include the contributions of devices and interventions [22]. Thus, the diagnostic value of PCT might be different between community-acquired BSI and nosocomial BSI.

PCT was superior to CRP and WBC in predicting nosocomial BSI, but its diagnostic accuracy remained modest (AUC, 0.69). The optimal cut-off value of PCT was 0.27 ng/mL. However, even at the lowest evaluated cut-off value of 0.1 ng/mL, PCT could not predict BSI in 6% of cases, and the optimal cut-off value of PCT could not predict BSI in 26% of cases. This finding is consistent with a previous report [21]. Most previous studies including on community-onset BSI concluded that a low

Table 5. Sensitivity, specificity, PPV, and NPV of serum procalcitonin at different cut-off values of PCT

Cut-off value of PCT, ng/mL	No. of cases under the cut-off value	Sensitivity, % (95% CI)	Specificity, % (95% CI)	PPV, % (95% CI)	NPV, % (95% CI)
0.10	7/105	93.3 (86.8–97.3)	24.4 (21.2–27.8)	16.0 (13.2–19.2)	95.9 (91.8–98.3)
0.27	28/105	74.6 (66.4–81.7)	56.5 (52.7–60.2)	25.3 (21.1–29.8)	91.9 (88.8–94.3)
0.40	43/105	61.2 (52.4–69.5)	65.7 (62.0–69.3)	26.0 (21.3–31.3)	89.6 (86.6–92.1)
0.50	46/105	56.2 (46.2–65.9)	69.6 (65.9–73.0)	22.2 (17.3–27.7)	91.1 (88.3–93.4)
0.60	50/105	52.4 (42.0–64.2)	72.5 (69.0–75.8)	22.9 (17.6–28.5)	90.8 (88.0–93.1)
0.90	58/105	45.5 (37.0–54.4)	78.8 (75.6–81.8)	29.8 (23.6–36.5)	88.0 (85.2–90.5)
2.0	72/105	31.4 (22.7–41.2)	86.8 (84.0–89.2)	26.8 (19.2–35.6)	89.1 (86.5–91.4)
5.0	105/105	14.3 (8.2–22.5)	93.5 (91.4–95.3)	25.4 (15.0–38.4)	87.6 (85.0–90.0)

PCT, procalcitonin; CI, confidence interval; PPV, positive predictive value; NPV, negative predictive value.

Table 6. Procalcitonin value for predicting positive blood culture according to renal function

Characteristic	eGFR ≥ 60 (n = 564)	30 ≤ eGFR < 60 (n = 113)	eGFR < 30 (n = 108)
Creatinine, mg/dL	0.64 (0.51–0.79)	1.34 (1.16–1.56)	3.73 (2.55–5.58)
eGFR, mL/min/1.73 m ²	98.3 (85.0–111.0)	44.0 (38.5–52.5)	13.0 (8.0–21.0)
PCT, ng/mL	0.17 (0.09–0.49)	0.38 (0.16–1.79)	0.99 (0.45–3.21)
Optimal cut-off value of PCT, ng/mL	0.12	0.84	1.71
Sensitivity, %	85.3 (74.6–92.7)	61.1 (35.7–82.7)	63.16 (38.4–83.7)
Specificity, %	38.5 (34.2–43.0)	86.0 (80.8–90.3)	69.7 (59.0–79.0)
PPV, %	16.0 (12.4–20.2)	26.2 (13.9–42.0)	30.8 (17.0–47.6)
NPV, %	95.0 (91.0–97.6)	96.4 (92.8–98.6)	89.9 (80.2–95.8)
AUC	0.691	0.705	0.661
CRP, mg/dL	4.31 (1.63–9.35)	5.02 (2.05–8.83)	6.41 (2.91–12.14)
WBC count, /μL	9,510 (6,432–13,055)	10,480 (7,510–13,680)	10,590 (7,630–13,825)
30-Day mortality	35 (16.1)	19 (16.8)	20 (18.5)

Values are presented as median (interquartile range) or number (%).

eGFR, estimated glomerular filtration rate; PCT, procalcitonin; PPV, positive predictive value; NPV, negative predictive value; AUC, area under curve; CRP, C-reactive protein; WBC, white blood cell.

PCT level can be used to rule out the presence of BSI [8,15,16,20,24,25]. In one such study, a PCT cut-off value ≤ 0.1 ng/mL seemed to be a useful marker to rule out bacteremia in the emergency department in patients with community-acquired BSI [15]. Two meta-analyses on the diagnostic accuracy of PCT for BSI also suggested that low PCT was effective at excluding BSI [24,25]. Hoeboer et al. [24] reported that a PCT cut-off value of 0.5 ng/mL corresponded with a 71% to 89% sensitivity and 95% to 98% NPV for different hospital settings. In the present study, a 0.5 ng/mL PCT cut-off value produced a sensitivity of 55.2% and NPV of 88.8% (Table 5). Even at the lowest cut-off value (0.1 ng/mL), NPV was only 94.5%. Although PCT was elevated in patients with nosocomial BSI, its diagnostic accuracy as a biomarker remained inadequate, as reported for intensive care unit patients [26]. Several factors may explain this poor performance. First, PCT increases with a 24- to 48-hour time lag after infection onset, which reduces the effectiveness of crude PCT measured when BSI is suspected [27,28]. Second, PCT remains elevated for up to several weeks after infection. Because hospitalized patients may be subject to several bacterial insults during their hospital stay, PCT may still be elevated due to previous episodes, thereby lowering its ability to detect a new BSI. Lastly, many conditions associated with hospital stay (profound circulatory failure, major surgery, trauma, pancreatitis, etc.) trigger systemic release of inflammatory mediators responsible for a nonspecific PCT increase [29-32]. Therefore, an approach where blood cultures are guided only by PCT cannot be recommended in the nosocomial setting.

We found that the PCT level was especially high in cases involving gram-negative bacteria rather than gram-positive bacteria. Gram-negative bacteria tend to induce higher levels of blood PCT compared with gram-positive bacteria [25,33]. This may reflect different interactions of gram-positive and gram-negative bacteria with host cells, involving lipoteichoic acids or lipopolysaccharide [34]. In particular, gram-positive bacteria activate the Toll-like receptor 2 (TLR2) pathway [35,36], whereas gram-negative bacteria are involved in the TLR4 pathway [37], resulting in different production of inflammatory cytokines, which ultimately stimulate ubiquitous transcription of calcitonin-mRNA and release of PCT from multiple tissues.

The mechanism of metabolism of PCT is not well-known; some studies have reported elevation of PCT in patients with impaired renal function [38-41]. A significant correlation between PCT and eGFR has been demonstrated [16,41]. Similarly, in our study, impaired renal function influenced the PCT level (Table 6). The cause of PCT elevation in patients with renal dysfunction could be impaired renal or hepatic elimination or increased production. Peripheral blood mononuclear cells release more PCT in patients with impaired renal function and in those receiving renal replacement therapy [42]. In addition, patients with severe renal dysfunction often show evidence of a systemic inflammatory response, which leads to PCT production [38]. In the present study, the PCT level tended to increase as eGFR fell, although there was no significant statistical difference between the $30 \leq \text{eGFR} < 60$ mL/min/1.73 m² group and the $\text{eGFR} < 30$ mL/min/1.73 m² group. Sitter et al. [43] proposed that loss of renal function does not affect the PCT; however, they set the normal PCT value < 0.5 ng/mL, which is higher than our cut-off values of 0.84 ng/mL in the $30 \leq \text{eGFR} < 60$ mL/min/1.73 m² group and 1.71 ng/mL in the $\text{eGFR} < 30$ mL/min/1.73 m² group.

Our study has some limitations. It is a retrospective single center study done on a selected population. There may have been some selection bias because of the study's retrospective design. The patient population might be heterogenous because of various hospitalization settings, including wards and intensive care units. Second, unexpected factors that could affect PCT levels were not concurrently analyzed in this study. It is well established that such conditions as profound circulatory failure, major surgery, trauma, and pancreatitis diminish the diagnostic value of PCT [29-32]. Third, our study design established the diagnostic value of PCT for predict nosocomial BSI, so blood culture-confirmed infections included only a BSI group. There is a possibility that patients with a local infection with a negative blood culture might be included in the non-BSI group [44]. Another factor to consider is the possibility of patients receiving antibiotic treatment before blood cultures were obtained, which would result in false negative blood culture results in individuals with high PCT levels, in turn reducing the PPV of PCT for predicting BSI. However, when we investigated the administration of antibiotics before blood collection for culture in the

group of patients with BSI, the level of PCT was not different between those who had received prior antibiotic administration and those who had not.

In conclusion, PCT level was higher in patients with nosocomial BSI and was more useful for predicting nosocomial BSI than was CRP or WBC count. However, even at the lowest cut-off value, PCT could not rule out nosocomial BSI. Therefore, an approach for blood cultures guided by PCT only does not seem reasonable.

KEY MESSAGE

1. Procalcitonin (PCT) might be a useful marker to exclude or predict bloodstream infection (BSI). However, the ability of PCT levels to differentiate BSI from non-BSI episodes has not been evaluated in nosocomial BSI.
2. PCT level was higher in patients with nosocomial BSI and was more useful for predicting nosocomial BSI than C-reactive protein or white blood cell count.
3. However, even at the lowest cut-off value, PCT could not rule out nosocomial BSI. Therefore, an approach for blood cultures guided by PCT only does not seem reasonable.

Conflict of interest

No potential conflict of interest relevant to this article was reported.

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