



# Grand Challenge Animal Reproduction-Theriogenology: From the Bench to Application to Animal Production and Reproductive Medicine

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### INTRODUCTION

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Tibary A (2017) Grand Challenge Animal Reproduction-Theriogenology: From the Bench to Application to Animal Production and Reproductive Medicine. Front. Vet. Sci. 4:114. doi: 10.3389/fvets.2017.00114 Reproductive physiology and procreation has always fascinated human kind. Therefore, it is not surprising that scientific research in reproduction is one of the oldest and most established field in biology. Advancement in reproductive sciences has been possible because of the curiosity of scientists of various backgrounds (biologists, animal scientists, and veterinarians). At the turn of the twentieth century, advances in reproductive research were mostly driven by needs for improved animal production and prevention of venereal diseases. The body of knowledge in animal reproduction has seen an exponential growth in the last 50 years. In recent years, the field of study expanded beyond laboratory species and production animals to include wildlife conservation and management.

As this field of research grew, scientists felt the importance of organizing in international societies dedicated to this area. One of the oldest of these societies is the Society for the Study of Reproduction. In the veterinary field, reproductive physiology and pathology became known as "Theriogenology" thanks to the efforts of the founding members of a veterinary specialty under the name of the American College of Theriogenologists, recognized in 1971 by the American Veterinary Medical Association as an integral part of the veterinary curriculum (1). Similar specialty colleges were also started in Europe (European College of Animal Reproduction). In addition to these specialty colleges, other international societies have emerged including Society for Theriogenology, International Embryo Transfer Society, European Society for Domestic Animal Reproduction, and European Society for Small Animal Reproduction. All these society have now well-established regular meetings to provide a forum for communication of recent research and their application to the health and welfare of animals. This paper attempts to highlight some of the major milestones and challenges in reproductive research.

### **REPRODUCTIVE PHYSIOLOGY AND PATHOLOGY IN THE MALE**

Tremendous progress has been achieved in understanding of sexual differentiation, testicular differentiation, and function (2–4). Early endocrine studies shed light on early testicular development and puberty. Clinical studies in the domestic animal led to the development of the field of applied clinical andrology (5). Studies incorporating clinical methods for prediction of fertility proved to be useful in the screening of males for optimal fertility in production animals and helped identify potential problems in individual animals with high genetic value. Breeding soundness examinations are now standardized techniques used in veterinary practice in almost all domestic animals. However, the prediction of fertility on a case-by-case basis remains challenging despite the development of a large number of clinical, histological, and molecular techniques (6, 7).

One the most important areas of research in male reproduction is the understanding of factors affecting spermatogenesis and sperm production. Traditionally, this has been approached through experimental designs involving live animals. The development of techniques such as specific molecular probes and xenografting of testicular tissues opened a new era of study on factors affecting spermatogenesis (8–11).

## MALE GAMETE PRESERVATION AND ARTIFICIAL INSEMINATION

Artificial insemination is recognized as the reproductive technology that had the most impact on animal reproduction. Although semen collection and preservation has long been established in several species, challenges still exist in others (camelids, wildlife). Sperm preservation in liquid (chilled) or frozen (cryopreservation) form continues to be a challenge because of tremendous individual male variation particularly in some species (i.e., equine, camelids). Molecular techniques have allowed scientists to detect changes during preservation which affect fertility (5). Recently, a focus was put on reactive oxygen species and how they alter sperm function (12).

Advances in sperm technologies include the introduction of sperm sexing for gender selection and the preservation of epididymal sperm. The limitation of use of sex-sorted semen imposed reconsideration of established artificial insemination parameters such as required number of sperm per insemination (13). Preservation of epididymal sperm is one of the most important aspects in preservation of genetic material from valuable terminally ill or deceased animals (wild and domestic species) (14–18). This new approach to breeding raises fundamental physiological questions on the role of seminal plasma in fertility (19–21).

Other significant advances in sperm technologies include cryopreservation by vitrification (22, 23), refreezing of spermatozoa (24–27), and production of spermatozoa from frozen testicular tissue (28, 29). Development in spermatogonia stem cell (SSC) culture opens a new era in the understanding of testis function and male fertility (2, 30–32). Transfer of SSC will eventually revolutionize our breeding strategies and preservation of genetics (33).

### REPRODUCTIVE PHYSIOLOGY IN THE FEMALE

Without doubt one the most important achievements in the study of reproductive physiology in the female is the development of protocols for synchronization of estrus and ovulation. These protocols proved to be of great values in production animals. Fixed-time artificial insemination is now a standard procedure for dairy and beef production as well as for large-scale sheep and goat operations (34–36). Estrous and ovulation synchronization protocols are also an integral part of embryo transfer programs. These techniques have become possible through advances in our understanding of the endocrinology of the female reproductive cycle and also through the introduction of *in vivo* monitoring of follicular dynamics through the use of ultrasonography (37). The characterization of follicular wave dynamics used initially in the bovine has become a standard approach to any study on the reproductive pattern in the female and has been adopted to study other species (i.e., camelids) (38). As ultrasonography technology advances, its uses in reproductive biology multiply. The development of Doppler ultrasonography provided a better characterization of ovulation, corpus luteum development, and blood flow to the uterus (39, 40).

Another area of advancement in female reproductive biology is the study of maternal recognition of pregnancy (MRP) (41, 42). Major advances have been realized in ruminants, shedding light on mechanisms involved in embryo elongation and trophoblast/ endometrium cross talk during MRP (43, 44). Molecular and genomics techniques provided insights on this mechanism and its implication in infertility and early embryonic loss (45). However, challenges still exist in deciphering MRP mechanisms in several species including equine (46, 47) and camelids (38).

# FEMALE GAMETE PRESERVATION

Oocyte preservation has been lagging behind comparatively to sperm. However, in recent years and with the development of ultrasound-guided follicular aspiration, oocytes are now routinely and repeatedly collected from valuable females and either stored or fertilized. Fertilization in vivo (oocyte transfer) or in vitro through techniques such as intracytoplasmic sperm injection or in vitro fertilization (IVF) is now commonplace in ruminants (48). Development of these technologies has seen a surge in the equine recently (49, 50). Oocyte cryopreservation has been challenging; however, newer techniques of verification show great promise (51). In vivo and in vitro production of oocytes from preserved/transplanted ovarian tissue is a novel technique used for the preservation of reproductive ability and could be an asset for the preservation of endangered species or genetic material from animals (52, 53). The effect of all these technologies on oocyte activation and capacity to develop into a viable embryo after fertilization is a subject of study using advanced molecular and genomic techniques (54).

# **EMBRYO TECHNOLOGY**

*In vivo* and *in vitro* production has become commonplace for several species. In ruminants, and more recently in camelids, advancement in protocols for ovarian superstimulation allowed reduction of cost and increased use of these technologies in production systems. In cattle, embryo transfer activity is progressively moving toward the use of *in vitro* produced embryo *via* IVF or somatic cell nuclear transfer (SCNT). Pregnancies obtained from *in vitro* produced embryos and particularly those by SCNT have been challenging due to increased pregnancy loss and increased rate of abnormalities of the placenta (hydrops) or fetus (abnormal

large offspring syndrome). SCNT has been reported in almost all domestic animal species. However, adoption of this technique for large-scale production remains challenging due to the great variability of results due to increased early pregnancy loss and abnormal pregnancies. Genomic studies showed a great difference in gene expression between embryos derived *in vivo* and those produced *in vitro* which could explain these abnormalities (55–57).

Embryo cryopreservation has become common place in the majority of ruminant production systems but still faces challenges in other species (equine, camelid). New techniques such as vitrification and dehydration prior to freezing are being developed and show some promise particularly in the equine and camelid species (58, 59).

Embryo manipulation allowed development of power tools such as testing for genetic disorders on embryo biopsies prior to transfer. Genetically engineered animals have been the goal of several studies for various reasons. This technology progressively moved from relatively crude techniques of production of transgenic animals (60, 61) to more sophisticated genome editing techniques such as clustered regularly interspaced short palindromic repeats (62, 63). These techniques will be great tools for the production of animals with specific genes of interest.

#### **REPRODUCTIVE MEDICINE**

Reproductive medicine research has focused on two paradoxical goals: contraception and enhanced fertility.

Effective non-surgical contraception has been a goal for many domestic and feral or wild species. Several approaches have been considered including immunization against zona pellucida and GnRH. However, these techniques proved to be ineffective in some species and not practical for large-scale use particularly in wild or feral animals (64–66). Another challenge in population control in the wild is the need for an effective method of contraception without alteration of normal reproductive behavior that often regulates the herd social structure. A challenge grant has been offered to find a safe and effective (permanent), single-dose non-surgical sterilant for both genders of cats and dogs (http:// www.michelsonprizeandgrants.org/). This has generated new interest and development of new approaches to contraception.

Enhanced fertility continues to be one of the most important aspects of production animal clinical services. The cattle dairy industry is the most concerned, as there is a trend toward poor reproductive efficiency with increased production (67). The first challenge in this industry was the inability to adequately detect estrus and reinseminate cows in a timely manner. This was in part due to human error in management and also due to changes in the reproductive biology of the high-producing cow. The interaction between metabolic activity and circulating hormone patterns is responsible for poor expression of estrus, increased rate of anovulatory cycles, and increased early embryonic loss. Strategies to improve the hormonal profiles are possible, but they are always limited by regulations on the use of hormones in foodproducing animals. The interaction between metabolic disorders, reproductive function, and susceptibility to uterine infection has been studied extensively in dairy cattle (68-72).

Other factors involved in reduced fertility in production animals that remains a challenge to the scientist and practitioner include environment and interaction with systemic disease. Heat stress has long been identified as a major limiting factor in reproduction. Molecular and genomics studies have shown a profound effect of heat stress on oocyte and embryo quality (68, 73). These studies represent a model of study as climatic changes experienced by the planet are bound to continue to have an effect on reproduction of both wild and domestic species.

Reproductive medicine/Theriogenology is also an individual animal practice. Several clinical problems have been studied to provide the best care for subfertile animals or animals with high-risk pregnancies. In addition to the common use of imaging in reproduction, new surgical and non-surgical techniques are being developed to diagnose and treat causes of infertility. The main species that have benefited from such techniques are small animals, equine, and camelid. The combination of established techniques such as endometrial biopsy, culture, and cytology with molecular techniques has allowed substantial advances in our understanding of the pathophysiology of endometritis and led to the development of methods for diagnosis, prevention, and treatment (74, 75).

The effect of age on follicular dynamics and endometrial degenerative changes has been studied primarily in mares (76–78). Stem cell therapy for these degenerative changes is being investigated (79, 80).

Advances in our understanding of the endocrinology of pregnancy and ultrasonographic evaluation of the fetus and placenta allowed a more efficient way for the evaluation of high-risk pregnancy in mares. An experimental model for ascending placentitis in this species allowed scientists to establish protocols for the diagnosis, monitoring, and treatment of this common cause of abortion and premature delivery (81–84).

Another area of critical importance in reproductive medicine is the diagnosis and prevention of abortion. Several infectious causes of abortion in ruminants are zoonotic and present serious health risk for humans (85). Strategies for rapid diagnosis of infectious causes of abortion have become available with the introduction of highly specific and sensitive molecular techniques (86). Studies on the pathophysiology of some of these diseases allowed a better understanding of the host–pathogen interaction and the development of congenital abnormalities. This has been useful recently in the discovery and study of a new disease "Schmallenberg" (87).

Reproductive toxicology has generated tremendous interest from various researchers. In addition to the traditional toxins (i.e., mycotoxin, plants) known to cause reproductive disorders (infertility or abortion), steroid disruptors have been a great concern for both animal and human health (88, 89).

### CONCLUSION

The primary goal of this article was to highlight the complexity and variety of areas of research in reproductive physiology and medicine. As methods for the study of reproduction have increased in sophistication, a huge amount of information has been generated. This creates a challenge for the scientific community and the practitioner as it becomes very difficult to translate some of the discoveries into application to medicine. This concern has already been stated almost a decade ago by Sirard who wrote "...too many publications are now reporting observations with little understanding of their value or how they fit in the big picture..." (90). It is important that scientists keep this in mind when developing training programs for future reproductive physiologists and theriogenologists. Multidisciplinary collaborations between bench scientists and clinical researchers will be more and more

### REFERENCES

- 1. Roberts S. The evaluation of the art and science of theriogenology. *Theriogenology* (1986) 25:618–38. doi:10.1016/0093-691X(86)90120-2
- Griswold MD. Spermatogenesis: the commitment to meiosis. *Physiol Rev* (2016) 96:1–17. doi:10.1152/physrev.00013.2015
- Makiyan Z. Studies of gonadal sex differentiation. Organogenesis (2016) 12:42–51. doi:10.1080/15476278.2016.1145318
- Kidder GM, Cyr DG. Roles of connexins in testis development and spermatogenesis. Semin Cell Dev Biol (2016) 50:22–30. doi:10.1016/j.semcdb.2015. 12.019
- 5. Chenoweth P, Lorton S. Animal Andrology: Theories and Applications. Wallingford: CABI (2014).
- Jean-Louis D, Francoise D, Xavier D. Epididymal protein markers and fertility. *Anim Reprod Sci* (2016) 169:76–87. doi:10.1016/j.anireprosci.2016.02.034
- Kaya A, Memili E. Sperm macromolecules associated with bull fertility. *Anim Reprod Sci* (2016) 169:88–94. doi:10.1016/j.anireprosci.2016.02.015
- Mirzapour T, Movahedin M, Koruji M, Nowroozi MR. Xenotransplantation assessment: morphometric study of human spermatogonial stem cells in recipient mouse testes. *Andrologia* (2015) 47:626–33. doi:10.1111/and. 12310
- Campos PHA, Costa GMJ, Avelar GF, Lacerda SMSN, da Costa NN, Ohashi OM, et al. Derivation of sperm from xenografted testis cells and tissues of the peccary (*Tayassu tajacu*). *Reproduction* (2014) 147:291–9. doi:10.1530/ REP-13-0581
- Mitchell RT, Anderson RA, Kelnar CJH, Wallace WHB, McKinnell C, Sharpe RM. Endocrine disruption in the human fetal testis: use of a xenograft system to assess effects of exposure to environmental agents and pharmaceutical drugs. *Lancet* (2013) 381:77. doi:10.1016/S0140-6736(13)60517-6
- Herrid M, McFarlane JR. Application of testis germ cell transplantation in breeding systems of food producing species: a review. *Anim Biotechnol* (2013) 24:293–306. doi:10.1080/10495398.2013.785431
- Aitken RJ, Gibb Z, Baker MA, Drevet J, Gharagozloo P. Causes and consequences of oxidative stress in spermatozoa. *Reprod Fert Dev* (2016) 28:1–10. doi:10.1071/RD15325
- Mikkola M, Taponen J. Quality and developmental rate of embryos produced with sex-sorted and conventional semen from superovulated dairy cattle. *Theriogenology* (2017) 87:135–40. doi:10.1016/j.theriogenology.2016. 08.013
- Roth TL, Stoops MA, Robeck TR, O'Brien JK. Factors impacting the success of post-mortem sperm rescue in the rhinoceros. *Anim Reprod Sci* (2016) 167:22–30. doi:10.1016/j.anireprosci.2016.01.019
- Prieto-Pablos MT, Sanchez-Calabuig MJ, Hildebrandt TB, Goritz F, Ortmann S, Eder S, et al. Cryopreservation of captive roe deer (*Capreolus capreolus*) semen. *Theriogenology* (2016) 86:695–703. doi:10.1016/j.theriogenology.2016. 02.023
- Pukazhenthi BS. Saving wild ungulate diversity through enhanced management and sperm cryopreservation. *Reprod Fert Dev* (2016) 28:1133–44. doi:10.1071/RD15412
- Vilela CG, Marquez JM, Graham JK, Barfield JP. Cryopreservation of bison epididymal sperm: a strategy for improving post-thaw quality when collecting sperm in field conditions. *Theriogenology* (2017) 89:155–61. doi:10.1016/j. theriogenology.2016.09.044
- Stawicki RJ, McDonnell SM, Giguere S, Turner RM. Pregnancy outcomes using stallion epididymal sperm stored at 5 degrees C for 24 or 48

important. One aim of the Frontiers in Veterinary Science specialty section on Animal Reproduction/Theriogenology will be to bridge the gap between fundamental science and application to animal health and reproduction.

### **AUTHOR CONTRIBUTIONS**

The author confirms being the sole contributor of this work and approved it for publication.

hours before harvest. *Theriogenology* (2016) 85:698–702. doi:10.1016/j. theriogenology.2015.10.009

- Neuhauser S, Dorfel S, Handler J. Dose-dependent effects of homologous seminal plasma on motility and kinematic characteristics of post-thaw stallion epididymal spermatozoa. *Andrology* (2015) 3:536–43. doi:10.1111/andr.12003
- Yoon SJ, Rahman MS, Kwon WS, Ryu DY, Park YJ, Pang MG. Proteomic identification of cryostress in epididymal spermatozoa. J Anim Sci Biotechnol (2016) 7:67. doi:10.1186/s40104-016-0128-2
- Yoon SJ, Rahman MS, Kwon WS, Park YJ, Pang MG. Addition of cryoprotectant significantly alters the epididymal sperm proteome. *PLoS One* (2016) 11:e0152690. doi:10.1371/journal.pone.0152690
- Pradiee J, Esteso MC, Lopez-Sebastian A, Toledano-Diaz A, Castano C, Carrizosa JA, et al. Successful ultrarapid cryopreservation of wild Iberian ibex (*Capra pyrenaica*) spermatozoa. *Theriogenology* (2015) 84:1513–22. doi:10.1016/j.theriogenology.2015.07.036
- Jimenez-Rabadan P, Garcia-Alvarez O, Vidal A, Maroto-Morales A, Iniesta-Cuerda M, Ramon M, et al. Effects of vitrification on ram spermatozoa using free-egg yolk extenders. *Cryobiology* (2015) 71:85–90. doi:10.1016/j. cryobiol.2015.05.004
- Gonzalez-Castro R, Sanches FA, Graham J, Carnevale E. Comparison of extenders for equine semen refreezing for intracytoplasmic sperm injection. *Anim Reprod Sci* (2016) 169:131. doi:10.1016/j.anireprosci.2016.03.084
- Abdussamad AM, Gauly M, Holtz W. Temporary storage of bovine semen cryopreserved in liquid nitrogen on dry ice and refreezing of frozen-thawed semen. *Cryo Lett* (2015) 36:278–84.
- Alvarez-Rodriguez M, Alvarez M, Lopez-Uruena E, Martinez-Rodriguez C, Borragan S, Anel-Lopez L, et al. Brown bear sperm double freezing: effect of elapsed time and use of PureSperm (R) gradient between freeze-thaw cycles. *Cryobiology* (2013) 67:339–46. doi:10.1016/j.cryobiol.2013.10.001
- Choi YH, Love CC, Varner DD, Hinrichs K. Equine blastocyst development after intracytoplasmic injection of sperm subjected to two freeze-thaw cycles. *Theriogenology* (2006) 65:808–19. doi:10.1016/j.theriogenology.2005. 04.035
- Yokonishi T, Sato T, Komeya M, Katagiri K, Kubota Y, Nakabayashi K, et al. Offspring production with sperm grown in vitro from cryopreserved testis tissues. *Nat Commun* (2014) 5:4320. doi:10.1038/ncomms5320
- Onofre J, Baert Y, Faes K, Goossens E. Cryopreservation of testicular tissue or testicular cell suspensions: a pivotal step in fertility preservation. *Hum Reprod Update* (2016) 22:744–61. doi:10.1093/humupd/dmw029
- Oatley MJ, Kaucher AV, Yang QE, Waqas MS, Oatley JM. Conditions for long-term culture of cattle undifferentiated spermatogonia. *Biol Reprod* (2016) 95:14. doi:10.1095/biolreprod.116.139832
- Oatley JM. Spermatogonial stem cell biology in the bull: development of isolation, culture, and transplantation methodologies and their potential impacts on cattle production. Soc Reprod Fertil Suppl (2010) 67:133–43.
- 32. de Rooij DG. Recent developments in the spermatogonial stem cell field. *Anim Reprod* (2017) 14:82–8. doi:10.21451/1984-3143-AR890
- Kanatsu-Shinohara M, Morimoto H, Shinohara T. Fertility of male germline stem cells following spermatogonial transplantation in infertile mouse models. *Biol Reprod* (2016) 94:112. doi:10.1095/biolreprod.115.137869
- Chebel RC, Ribeiro ES. Reproductive systems for North American dairy cattle herds. Vet Clin North Am Food Anim Pract (2016) 32:267. doi:10.1016/j. cvfa.2016.01.002
- 35. Dolecheck KA, Silvia WJ, Heersche G, Wood CL, McQuerry KJ, Bewley JM. A comparison of timed artificial insemination and automated

activity monitoring with hormone intervention in 3 commercial dairy herds. *J Dairy Sci* (2016) 99:1506–14. doi:10.3168/jds.2015-9914

- Lamb GC, Mercadante VRG. Synchronization and artificial insemination strategies in beef cattle. Vet Clin North Am Food Anim Pract (2016) 32:335. doi:10.1016/j.cvfa.2016.01.006
- Adams GP, Singh J, Baerwald AR. Large animal models for the study of ovarian follicular dynamics in women. *Theriogenology* (2012) 78:1733–48. doi:10.1016/j.theriogenology.2012.04.010
- Tibary A, Anouassi A, Sghiri A, Khatir H. Current knowledge and future challenges in camelid reproduction. Soc Reprod Fertil Suppl (2007) 64: 297–313.
- Ginther OJ, Rakesh HB, Hoffman MM. Blood flow to follicles and CL during development of the periovulatory follicular wave in heifers. *Theriogenology* (2014) 82:304–11. doi:10.1016/j.theriogenology.2014.04.009
- Honig H, Ofer L, Kaim M, Jacobi S, Shinder D, Gershon E. The effect of cooling management on blood flow to the dominant follicle and estrous cycle length at heat stress. *Theriogenology* (2016) 86:626–34. doi:10.1016/j. theriogenology.2016.02.017
- Bazer FW. Pregnancy recognition signaling mechanisms in ruminants and pigs. J Anim Sci Biotechnol (2013) 4:23. doi:10.1186/2049-1891-4-23
- Bazer FW, Wu GY, Johnson GA. Pregnancy recognition signals in mammals: the roles of interferons and estrogens. *Anim Reprod* (2017) 14:7–29. doi:10.21451/1984-3143-AR888
- Brooks K, Burns G, Spencer TE. Conceptus elongation in ruminants: roles of progesterone, prostaglandin, interferon tau and cortisol. J Anim Sci Biotechnol (2014) 5:53. doi:10.1186/2049-1891-5-53
- Dorniak P, Bazer FW, Spencer TE. Physiology and endocrinology symposium: biological role of interferon tau in endometrial function and conceptus elongation. J Anim Sci (2013) 91:1627–38. doi:10.2527/jas.2012-5845
- Geary TW, Burns GW, Moraes JGN, Moss JI, Denicol AC, Dobbs KB, et al. Identification of beef heifers with superior uterine capacity for pregnancy. *Biol Reprod* (2016) 95:47. doi:10.1095/biolreprod.116.141390
- Klohonatz KM, Hess AM, Hansen TR, Squires EL, Bouma GJ, Bruemmer JE. Equine endometrial gene expression changes during and after maternal recognition of pregnancy. *J Anim Sci* (2015) 93:3364–76. doi:10.2527/jas. 2014-8826
- Aurich C, Budik S. Early pregnancy in the horse revisited does exception prove the rule? J Anim Sci Biotechnol (2015) 6:50. doi:10.1186/s40104-015-0048-6
- Kassens A, Held E, Salilew-Wondim D, Sieme H, Wrenzycki C, Tesfaye D, et al. Intrafollicular oocyte transfer (IFOT) of abattoir-derived and in vitromatured oocytes results in viable blastocysts and birth of healthy calves. *Biol Reprod* (2015) 92:150. doi:10.1095/biolreprod.114.124883
- Galli C, Duchi R, Colleoni S, Lagutina I, Lazzari G. Ovum pick up, intracytoplasmic sperm injection and somatic cell nuclear transfer in cattle, buffalo and horses: from the research laboratory to clinical practice. *Theriogenology* (2014) 81:138–51. doi:10.1016/j.theriogenology.2013.09.008
- Hinrichs K. Assisted reproduction techniques in the horse. *Reprod Fert Dev* (2013) 25:80–93. doi:10.1071/RD12263
- Spricigo JFW, Netto SBS, Muterlle CV, Rodrigues SDD, Leme LO, GuimaraesAL,etal.Intrafolliculartransferoffreshandvitrifiedimmaturebovine oocytes. *Theriogenology* (2016)86:2054–62. doi:10.1016/j.theriogenology.2016. 07.003
- Silber S. Ovarian tissue cryopreservation and transplantation: scientific implications. J Assist Reprod Genet (2016) 33:1595–603. doi:10.1007/ s10815-016-0814-1
- 53. Meirow D, Ra'anani H, Shapira M, Brenghausen M, Chaim SD, Aviel-Ronen S, et al. Transplantations of frozen-thawed ovarian tissue demonstrate high reproductive performance and the need to revise restrictive criteria. *Fertil Steril* (2016) 106:467–74. doi:10.1016/j.fertnstert.2016.04.031
- Fernandes CB, Devito LG, Martins LR, Blanco IDP, Neto JFD, Tsuribe PM, et al. Artificial activation of bovine and equine oocytes with cycloheximide, roscovitine, strontium, or 6-dimethylaminopurine in low or high calcium concentrations. *Zygote* (2014) 22:387–94. doi:10.1017/S0967199412000627
- Niemann H. Epigenetic reprogramming in mammalian species after SCNT-based cloning. *Theriogenology* (2016) 86:80–90. doi:10.1016/j. theriogenology.2016.04.021
- Loi P, Toschi P, Zacchini F, Ptak G, Scapolo PA, Capra E, et al. Synergies between assisted reproduction technologies and functional genomics. *Genet Sel Evol* (2016) 48:53. doi:10.1186/s12711-016-0231-z

- 57. Loi P, Iuso D, Czernik M, Ogura A. A new, dynamic era for somatic cell nuclear transfer? *Trends Biotechnol* (2016) 34:791–7. doi:10.1016/j.tibtech.2016.03.008
- Herrid M, Vajta G, Skidmore JA. Current status and future direction of cryopreservation of camelid embryos. *Theriogenology* (2017) 89:20–5. doi:10.1016/j.theriogenology.2016.10.005
- Choi YH, Hinrichs K. Vitrification of in vitro-produced and in vivo-recovered equine blastocysts in a clinical program. *Theriogenology* (2017) 87:48–54. doi:10.1016/j.theriogenology.2016.08.005
- Proudfoot C, Carlson DF, Huddart R, Long CR, Pryor JH, King TJ, et al. Genome edited sheep and cattle. *Transgenic Res* (2015) 24:147–53. doi:10.1007/ s11248-014-9832-x
- Menchaca A, Anegon I, Whitelaw CBA, Baldassarre H, Crispo M. New insights and current tools for genetically engineered (GE) sheep and goats. *Theriogenology* (2016) 86:160–9. doi:10.1016/j.theriogenology.2016.04.028
- Park KE, Kaucher AV, Powell A, Waqas MS, Sandmaier SES, Oatley MJ, et al. Generation of germline ablated male pigs by CRISPR/Cas9 editing of the NANOS2 gene. Sci Rep (2017) 7:40176. doi:10.1038/srep40176
- Josa S, Seruggia D, Fernandez A, Montoliu L. Concepts and tools for gene editing. *Reprod Fert Dev* (2017) 29:1–7. doi:10.1071/RD16396
- Wilker M, Pearson LK, Campbell A, Tibary A. Non-surgical methods of contraception and sterilization in select domestic and wildlife species. *Clinical Theriogenol* (2014) 6:93–104.
- Asa C. Weighing the options for limiting surplus animals. Zoo Biol (2016) 35:183–6. doi:10.1002/zoo.21293
- Massei G, Koon KK, Benton S, Brown R, Gomm M, Orahood DS, et al. Immunocontraception for managing feral cattle in Hong Kong. *PLoS One* (2015) 10:e0121598. doi:10.1371/journal.pone.0121598
- Bradford BJ, Yuan K, Ylioja C. Managing complexity: dealing with systemic crosstalk in bovine physiology. J Dairy Sci (2016) 99:4983–96. doi:10.3168/ jds.2015-10271
- Baruselli PS, Batista EOS, Vieira LM, Sales JNDS, Gimenes LU, Ferreira RM. Intrinsic and extrinsic factors that influence ovarian environment and efficiency of reproduction in cattle. *Anim Reprod* (2017) 14:48–60. doi:10.21451/1984-3143-AR907
- Zebeli Q, Ghareeb K, Humer E, Metzler-Zebeli BU, Besenfelder U. Nutrition, rumen health and inflammation in the transition period and their role on overall health and fertility in dairy cows. *Res Vet Sci* (2015) 103:126–36. doi:10.1016/j.rvsc.2015.09.020
- Vukasinovic N, Bacciu N, Przybyla CA, Boddhireddy P, DeNise SK. Development of genetic and genomic evaluation for wellness traits in US Holstein cows. J Dairy Sci (2017) 100:428–38. doi:10.3168/jds.2016-11520
- Bicalho MLS, Marques EC, Gilbert RO, Bicalho RC. The association of plasma glucose, BHBA, and NEFA with postpartum uterine diseases, fertility, and milk production of Holstein dairy cows. *Theriogenology* (2017) 88:270–82. doi:10.1016/j.theriogenology.2016.09.036
- Baithalu RK, Singh SK, Kumaresan A, Mohanty AK, Mohanty TK, Kumar S, et al. Transcriptional abundance of antioxidant enzymes in endometrium and their circulating levels in Zebu cows with and without uterine infection. *Anim Reprod Sci* (2017) 177:79–87. doi:10.1016/j.anireprosci.2016.12.008
- Slimen IB, Najar T, Ghram A, Abdrrabba M. Heat stress effects on livestock: molecular, cellular and metabolic aspects, a review. J Anim Physiol Anim Nutr (Berl) (2016) 100:401–12. doi:10.1111/jpn.12379
- Troedsson MHT, Woodward EM. Our current understanding of the pathophysiology of equine endometritis with an emphasis on breeding-induced endometritis. *Reprod Biol* (2016) 16:8–12. doi:10.1016/j.repbio.2016.01.003
- Woodward EM, Troedsson MHT. Inflammatory mechanisms of endometritis. Equine Vet J (2015) 47:384–9. doi:10.1111/evj.12403
- Hendriks WK, Colleoni S, Galli C, Paris DBBP, Colenbrander B, Roelen BAJ, et al. Maternal age and in vitro culture affect mitochondrial number and function in equine oocytes and embryos. *Reprod Fert Dev* (2015) 27:957–68. doi:10.1071/RD14450
- Esteller-Vico A, Liu IKM, Vaughan B, Steffey EP, Brosnan RJ. Effects of vascular elastosis on uterine blood flow and perfusion in anesthetized mares. *Theriogenology* (2015) 83:988–94. doi:10.1016/j.theriogenology.2014.11.032
- Ferreira-Dias GM, Rebordao MR, Galvao AM, Mateus LM, Szostek A, Skarzynski DI. Pathways from mare endometritis to endometrosis. *Reprod Domest Anim* (2015) 50:27.
- 79. Mambelli LI, Mattos RC, Winter GHZ, Madeiro DS, Morais BP, Malschitzky E, et al. Changes in expression pattern of selected endometrial proteins

following mesenchymal stem cells infusion in mares with endometrosis. *PLoS One* (2014) 9:e97889. doi:10.1371/journal.pone.0097889

- Mambelli LI, Winter GHZ, Kerkis A, Malschitzky E, Mattos RC, Kerkis I. A novel strategy of mesenchymal stem cells delivery in the uterus of mares with endometrosis. *Theriogenology* (2013) 79:744–50. doi:10.1016/j. theriogenology.2012.11.030
- Canisso IF, Ball BA, Scoggin KE, Squires EL, Williams NM, Troedsson MH. Alpha-fetoprotein is present in the fetal fluids and is increased in plasma of mares with experimentally induced ascending placentitis. *Anim Reprod Sci* (2015) 154:48–55. doi:10.1016/j.anireprosci.2014.12.019
- 82. Macpherson ML, Giguere S, Hatzel JN, Pozor M, Benson S, Diaw M, et al. Disposition of desfuroylceftiofur acetamide in serum, placental tissue, fetal fluids, and fetal tissues after administration of ceftiofur crystalline free acid (CCFA) to pony mares with placentitis. *J Vet Pharmacol Ther* (2013) 36:59–67. doi:10.1111/j.1365-2885.2012.01392.x
- LeBlanc MM, Giguere S, Lester GD, Brauer K, Paccamonti DL. Relationship between infection, inflammation and premature parturition in mares with experimentally induced placentitis. *Equine Vet J* (2012) 44:8–14. doi:10.1111/j.2042-3306.2011.00502.x
- dos Santos RS, Correa MN, de Araujo LO, Pazinato FM, Feijo LS, Curcio BR, et al. Hematological and hemogasometric evaluation of foals born from mares with ascending placentitis. *Arq Bras Med Vet Zoo* (2017) 69:48–56.
- Ganter M. Zoonotic risks from small ruminants. Vet Microbiol (2015) 181:53–65. doi:10.1016/j.vetmic.2015.07.015
- 86. Clothier K, Anderson M. Evaluation of bovine abortion cases and tissue suitability for identification of infectious agents in California

diagnostic laboratory cases from 2007 to 2012. *Theriogenology* (2016) 85:933-8. doi:10.1016/j.theriogenology.2015.11.001

- Lievaart-Peterson K, Luttikholt S, Peperkamp K, Van den Brom R, Vellema P. Schmallenberg disease in sheep or goats: past, present and future. *Vet Microbiol* (2015) 181:147–53. doi:10.1016/j.vetmic.2015.08.005
- Ferreira-Dias G, Botelho M, Zagrajczuk A, Rebordao MR, Galvao AM, Bravo PP, et al. Coumestrol and its metabolite in mares' plasma after ingestion of phytoestrogen-rich plants: potent endocrine disruptors inducing infertility. *Theriogenology* (2013) 80:684–92. doi:10.1016/j.theriogenology.2013.06.002
- Guerrero-Bosagna C, Savenkova M, Haque MM, Nilsson E, Skinner MK. Environmentally induced epigenetic transgenerational inheritance of altered sertoli cell transcriptome and epigenome: molecular etiology of male infertility. *PLoS One* (2013) 8:e59922. doi:10.1371/journal.pone.0059922
- 90. Sirard MA. From biological fundamentals to practice, and back. *Theriogenology* (2007) 68:S250–6. doi:10.1016/j.theriogenology.2007.05.042

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