REVIEW



The pathogenesis and potential therapeutic targets in sepsis

Wendan Zhang 1,2 | Honghong Jiang 1,2 | Gaosong Wu 1 | Pengli Huang 1 | Haonan Wang^{1,*} | Huazhasng An^{3,*} | Sanhong Liu^{1,*} | Weidong Zhang^{1,4,5,6,*}

*Correspondence

Weidong Zhang, Shanghai Frontiers Science Center of TCM Chemical Biology, Institute of Interdisciplinary Integrative Medicine Research, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, China; Department of Phytochemistry, School of Pharmacy, Second Military Medical University, Shanghai 200433, China; The Research Center for Traditional Chinese Medicine, Shanghai Institute of Infectious Diseases and Biosecurity, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, China.

Email: wdzhangy@hotmail.com

Sanhong Liu, Shanghai Frontiers Science Center of TCM Chemical Biology, Institute of Interdisciplinary Integrative Medicine Research, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, China.

Email: liush@shutcm.edu.cn

Huazhang An, Shandong Provincial Key Laboratory for Rheumatic Disease and Translational Medicine, The First

Abstract

Sepsis is defined as "a life-threatening organ dysfunction caused by dysregulated host systemic inflammatory and immune response to infection." At present, sepsis continues to pose a grave healthcare concern worldwide. Despite the use of supportive measures in treating traditional sepsis, such as intravenous fluids, vasoactive substances, and oxygen plus antibiotics to eradicate harmful pathogens, there is an ongoing increase in both the morbidity and mortality associated with sepsis during clinical interventions. Therefore, it is urgent to design specific pharmacologic agents for the treatment of sepsis and convert them into a novel targeted treatment strategy. Herein, we provide an overview of the molecular mechanisms that may be involved in sepsis, such as the inflammatory response, immune dysfunction, complement deactivation, mitochondrial damage, and endoplasmic reticulum stress. Additionally, we highlight important targets involved in sepsis-related regulatory mechanisms, including GSDMD, HMGB1, STING, and SQSTM1, among others. We summarize the latest advancements in potential therapeutic drugs that specifically target these signaling pathways and paramount targets, covering both preclinical studies and clinical trials. In addition, this review provides a detailed description of the crosstalk and function between signaling pathways and vital targets, which provides more opportunities for the clinical development of new treatments for sepsis.

Wendan Zhang, Honghong Jiang, Gaosong Wu and Pengli Huang contributed equally to this work.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2023 The Authors. MedComm published by Sichuan International Medical Exchange & Promotion Association (SCIMEA) and John Wiley & Sons Australia, Ltd.

MedComm. 2023;4:e418. wileyonlinelibrary.com/journal/mco2 1 of 37

¹Shanghai Frontiers Science Center of TCM Chemical Biology, Institute of Interdisciplinary Integrative Medicine Research, Shanghai University of Traditional Chinese Medicine, Shanghai, China

²Faculty of Pediatrics, National Engineering Laboratory for Birth defects prevention and control of key technology, Beijing Key Laboratory of Pediatric Organ Failure, the Chinese PLA General Hospital, Beijing, China

³Shandong Provincial Key Laboratory for Rheumatic Disease and Translational Medicine, The First Affiliated Hospital of Shandong First Medical University & Shandong Provincial Qianfoshan Hospital, Jinan, Shandong, China

⁴Department of Phytochemistry, School of Pharmacy, Second Military Medical University, Shanghai, China

⁵The Research Center for Traditional Chinese Medicine, Shanghai Institute of Infectious Diseases and Biosecurity, Shanghai University of Traditional Chinese Medicine, Shanghai, China

⁶Institute of Medicinal Plant Development, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China



Affiliated Hospital of Shandong First Medical University & Shandong Provincial Qianfoshan Hospital, Jingshi Road 16766, Jinan, Shandong 250014, China.

Email: anhz@immunol.org

Haonan Wang, Shanghai Frontiers Science Center of TCM Chemical Biology, Institute of Interdisciplinary Integrative Medicine Research, Shanghai University of Traditional Chinese Medicine, Shanghai 201203, China.

Email: wanghaonan0611@126.com

Funding information

National Natural Science Foundation of China, Grant/Award Numbers: 82141203, 81772798; Shanghai Municipal Science and Technology Major Project, Grant/Award Number: ZD2021CY001; Three-year Action Plan for Shanghai TCM Development and Inheritance Program, Grant/Award Number: ZY (2021-2023)-0401; Innovation Team and Talents Cultivation Program of National Administration of Traditional Chinese Medicine, Grant/Award Number: ZYYCXTDD-202004

KEYWORDS

molecular mechanism, pathogenesis, sepsis, signaling pathways, therapeutic drugs

1 | INTRODUCTION

Throughout history, the term "sepiton" was utilized by Hippocrates to express the possibility of harmful biological decay transpiring within the body.^{1,2} In earlier periods, sepsis was defined as a syndrome of systemic inflammation in reaction to microbial infection. This syndrome is characterized by extensive tissue damage and systemic inflammation.3 However, recent advancements in understanding the pathophysiology of sepsis have unveiled its true nature, indicating that the definition of sepsis cannot solely rely on it being an inflammatory syndrome.4 Clearly, the concept of systemic inflammatory response syndrome (SIRS) is too broadly employed in critically ill patients, failing to encompass the clinical heterogeneity and dynamics of real-life scenarios. In 2016, international guidelines were revised to introduce the concept of sepsis 3.0. Sepsis 3.0 defines sepsis as life-threatening organ dysfunction caused by an impaired host response to various infections, whether bacterial, fungal, protozoan, or viral, which includes the recent severe acute respiratory syndrome coronavirus 2 responsible for coronavirus disease 2019 (COVID-19).⁵ Sepsis is a multifaceted syndrome that manifests in different ways as the host's regulation is disrupted in response to infection. Traditionally, sepsis has been mainly attributed to the host's overly inflammatory immune response to infection. Consequently, most clinical studies in the 1990s focused on therapies aimed at limiting excessive inflammation, without any substantial success.^{6,7}

The morbidity and mortality of sepsis are tremendous, and the continuous growth of this condition has caused a substantial burden on global healthcare.^{8,9} World Sepsis Day was created on September 13, 2012, with the main goal of raising public awareness about the severity of this perilous illness. Shockingly, in 2017 alone, approximately 48.9 million individuals across the globe were afflicted by sepsis, leading to a staggering 11.0 million deaths. These sepsis-related fatalities accounted for nearly one-fifth of all deaths worldwide. The immense scale and devastating impact of sepsis cannot be underestimated. 10 Therefore, sepsis is a critical public health issue, causing considerable economic consequences around the world. The World Health Organization has called on member states to enhance their efforts in preventing, diagnosing, and managing sepsis. 11,12 This can only be accomplished by gaining a deeper understanding of the underlying mechanisms of sepsis and implementing more precise and efficient treatment strategies.¹³ Sepsis is a multifaceted condition that arises as a result of an unbalanced immune response to infection. It is characterized not only by an overwhelming inflammatory reaction but also by excessive suppression

of the immune system. To effectively address sepsis, it is crucial to comprehend the complexities of sepsis development and the intricate interplay between inflammation and immunosuppression. 14,15 Despite significant advancements in medical technologies, particularly in the field of anti-infective treatments, there is still no conclusive remedy to address the prevalent issue of sepsis. Furthermore, individuals afflicted with sepsis encounter enduring and severe complications encompassing physical, psychological, and cognitive impairments, even following therapeutic interventions.¹⁶ To address these issues, new criteria for sepsis recognition and intervention should focus more on regulating key targets in signaling pathways rather than determining signs of inflammation. Therefore, it is extremely urgent to develop prevention methods and effective treatment measures for sepsis to solve human health problems.

An improved understanding of sepsis has revealed that it is not just a systemic inflammatory response or immune disease process but also entails changes in the function of multiple organs in the body.¹⁷ Essentially, sepsis can be seen as a battle between the pathogen and the host immune system, where the outcome is determined by the delicate balance between pro- and anti-inflammatory pathways. This imbalance can result in multiple organ dysfunction and ultimately decide the fate of the individual. 18,19 Significant advancements have been made in understanding how sepsis causes organ injury at the molecular, cellular, and organ levels. The pathogenesis of sepsis is an extremely complex and diverse pathophysiological process that leads to an imbalance in homeostasis at the molecular, cellular, and organ levels, eventually leading to organ dysfunction and even death. 13,20-22 Herein, a comprehensive review is presented concerning the dysregulated/altered pathways and molecules implicated in sepsis, along with a potential molecular mechanism for the development of sepsis. Additionally, detailed explanations were given regarding the interactions among these pathways. We also delineated novel candidate therapeutic targets and drugs that might contribute to the treatment of sepsis.

2 | REGULATORY MECHANISMS INVOLVED IN SEPSIS

In the process of basic and clinical complementary and progressive research, many new mechanisms, targets, and therapeutic molecules of sepsis have been discovered and validated.^{23–25} The findings have attracted increasing attention, attempting to translate them into clinical outcomes. Expanding from the macro level to micro details, dissecting the essence of sepsis, transforming from a simple definition of septic shock in the past to a hetero-

geneous lesion with multiple organ dysfunction caused by infection.^{26,27} Variations of this nature result in the inability to identify a specific cause in the clinical presentation of patients with sepsis and thus to implement interventions.^{3,28} In the past, the clinical management of sepsis mainly took the form of the administration of interferon, vasopressors, or intravenous fluids. However, these interventions have some potential risks and safety issues, such as antibiotic resistance. Therefore, further evaluation is needed to determine the safety and long-term impact of these applications on human health. 19,22,29 With the continuous progress of science and technology and the rapid development of clinical medicine, the understanding of the development of sepsis has risen from the initial excessive inflammatory reaction leading to immunosuppression, mitochondrial dysfunction, endoplasmic reticulum (ER) emergencies, cell necrosis (pyroptosis, apoptosis, etc.), neural network damage, complement system disorder leading to coagulation abnormalities, and so on. 4,30 In this section, we review the main regulatory molecular mechanisms involved in the occurrence and development of sepsis and emphasize potential new candidate therapeutic targets and drugs for treating sepsis.

2.1 | Inflammation and immune

The improper presence of microorganisms (bacteria, etc.) and their products can trigger a host immune response that is critical for maintaining and restoring homeostasis but can lead to tissue damage if excessive.³¹ In practice, the greater the inflammatory response is, the stronger the cellular damage and thus the higher the risk of organ dysfunction.¹⁸ At its onset, sepsis manifests as an overwhelming release of inflammatory mediators (cytokine storms) in response to infection. 32,33 The immune response to infection can work with innate immunization defense, components of the innate immune system at the site of pathogen exposure that activate and recruit circulating immune cells.^{34,16} Immune cells have pathogenrecognition receptors (PRRs) on their surface that are activated by binding to pathogen-associated molecular patterns (PAMPs) or damage-associated molecular patterns (DAMPs), such as bacteria, burn, trauma, and so on. Receptor binding triggers intracellular signaling cascades, which in turn leads to the production of associated inflammatory mediators, thus initiating an immunoinflammatory cascade.^{35,36} The ensuing chain reaction will eventually lead to the activation of an even stronger "armed force," which is the adaptive immune response.³⁷ This explosive activation and the resulting immune "cytokine storm" have been identified as the pathogenic pathway for sepsis.^{33,38}

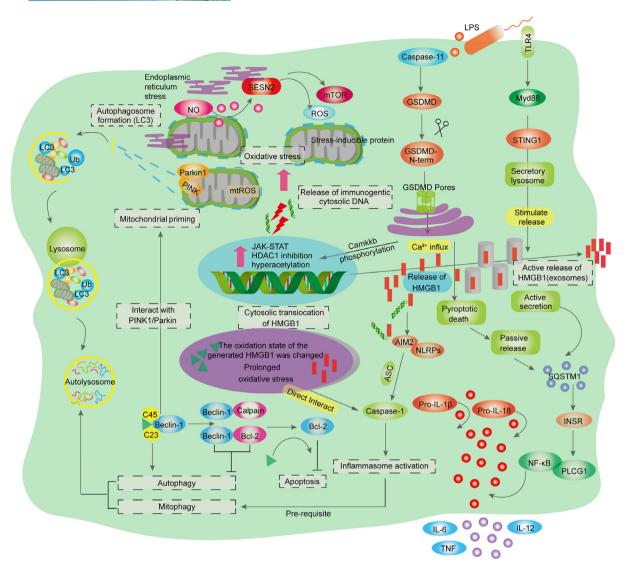


FIGURE 1 Altered innate immune function and cellular homeostasis after sepsis. In immune cells, the complementarity and regulation of inflammasomes, cell death pathways, and oxidative stress contribute greatly to the maintenance of host cell survival and homeostasis. MtROS and mtDNA activate the JAK–STAT pathway and trigger histone deacetylase 1 (HDAC1) inhibition, which is required for HMGB1 to play a regulatory role (hyperacetylation and cytosolic translocation). The massive release of all stimuli-induced HMGB1 by exosomes is an inflammatory mediator that marks the prelude of sepsis. AIM2 or NLRPs and dsDNA commence the inflammasome, which in turn triggers caspase-1-driven responses. These responses act as prerequisites for the induction of autophagy/mitophagy through pathways mediated by beclin1. In addition, HMGB1 induces autophagy by releasing Beclinp1 from Bcl2 after binding with Beclinp1 and then removing hazardous oxidative stress stimuli. PINK1 promotes the recruitment of Parkin to mitochondria, and subsequently, beclin-1 interacts with pink/Parkin to further initiate mitochondrial priming and autophagosome generation, which then interact with lysosomes to induce mitophagy. Autophagy and mitophagy are the basic lines of defense that protect the body from injuries and regulate innate immune responses. During sepsis, macrophages secrete or release SQSTM1 actively or passively in response to extracellular or intracellular LPS, respectively. The released SQSTM1 then functions as an inflammatory mediator by activating the NF-κB pathway, which is dependent on the INSR, contributing to the inflammatory response. Therefore, failure to activate protective autophagy/mitophagy may lead to accelerated death in sepsis.

In the sepsis-induced inflammatory response, both external factors derived from pathogens and internal factors released by damaged cells (such as lipopolysaccharide [LPS] and high-mobility group box-1 [HMGB-1] protein) can recognize PRRs, including Toll-like receptors (TLRs), C-type lectin receptors (CLRs), retinoic acid inducible gene I (RIG-I)-like receptors, and NOD-like

receptors (NLRs), leading to the activation of downstream pathways.³⁹ The activation of different receptors results in the translocation of transcription factors, such as nuclear factor- κ B (NF- κ B), into the nucleus and subsequent activation of target genes, which encode proinflammatory cytokines such as tumor necrosis factor (TNF), interleukin (IL)–6, IL-12, and interferons (IFNs) (Figure 1).⁴⁰ The

secretion of proinflammatory factors progresses to a lifethreatening cytokine storm. Moreover, soluble cytosolic NLRs also contribute to the immune imbalance associated with sepsis.41,42 The activation of specific NLRs is regulated by adaptor receptor-interacting protein kinase 2 (RIP2) (also known as RICK), leading to NF-kB activation and adaptor protein 1 activation. However, certain other NLRs (e.g., NLRP and NLRC4) participate in the assembly of distinct inflammasome protein complexes.⁴³ The inflammasome cleaves procaspase-1 into active caspase-1, which then cleaves pro-IL-1 β and pro-IL-18, resulting in the release of the highly inflammatory cytokines IL- 1β and IL-18 and triggering a form of programmed cell death (known as pyroptosis) characterized by intense inflammation.44 When these mechanisms are driven by sepsis, they can become catastrophic and lead to a lifethreatening inflammatory cascade. 45,46 Dectin is one of the main CLRs that plays a significant role in various biological processes. This particular Dectin has the ability to induce the production of reactive oxygen species (ROS) and activate inflammatory responses through the action of SRC and SYK kinases.⁴⁷ Interestingly, in addition to the PPRs mentioned above, some double-stranded RNA receptors have been found, mainly including RIG-I, melanoma differentiation-associated gene 5, and laboratory of genetics and physiology 2, and are also associated with sepsis induced immune dysfunction.^{48–50} It has been noted that PRR can be activated by both exogenous PAMPs and endogenous DAMPs. 36,51 In relation to the induction of sepsis from within the body, it has been found that hepatocytes release a significant amount of HMGB-1, which binds to bacterial endotoxin known as LPS (Figure 2).^{52,53} This bacterial endotoxin is then transported to the cytoplasm through receptors called receptor for advanced glycation end products (RAGE), which are expressed in vascular endothelial cells (ECs) and macrophages.⁵⁴ As a result of this process, caspase-11-mediated cell death, or apoptosis, occurs, ultimately leading to sepsis, multiple organ failure, and death.55,56

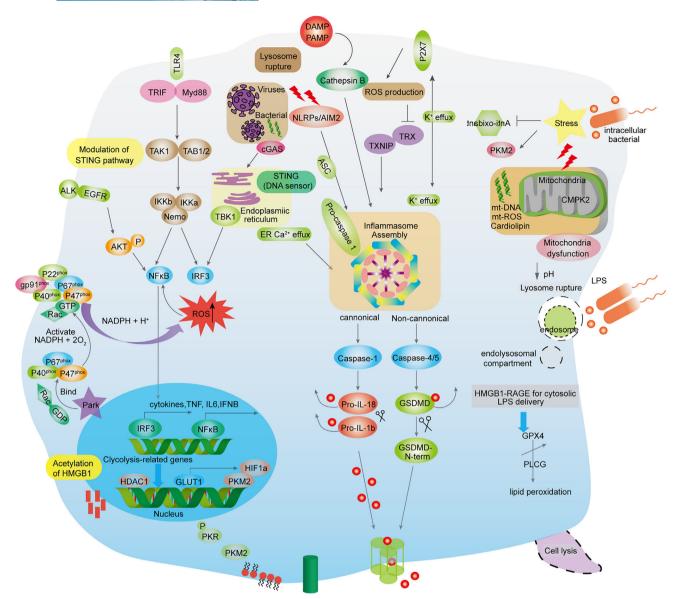
2.2 | Inflammation and coagulopathy

Disseminated intravascular coagulation (DIC) is a life-threatening syndrome that is commonly considered an organ dysfunction that is often present in sepsis, with severity positively correlated with mortality.^{57,58} DIC overactivation is characterized by an intravenous coagulation cascade, exhaustion of anticoagulants, and inhibition of fibrinolysis.⁵⁹ The dysregulation of inflammation has been found to trigger the coagulation response in sepsis, while the activation of the coagulation reaction exacerbates the inflammatory response.²⁹ This highlights the crucial

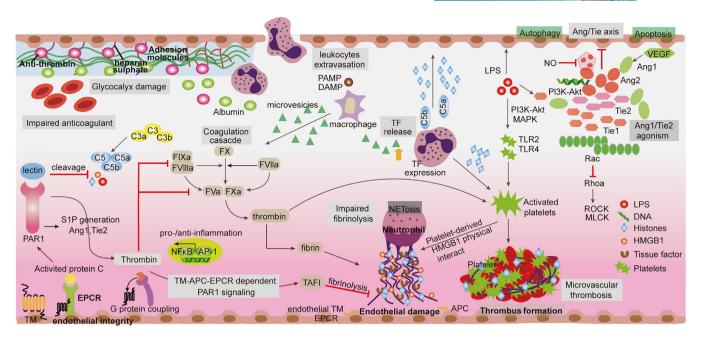
interaction between inflammation and coagulation, which is seen as the primary driver of sepsis pathogenesis.⁶⁰ Throughout the onset and progression of sepsis, the generation of cytokines not only facilitates the activation of coagulation factors and platelets but also initiates the mechanism of anticoagulation (such as the system of antithrombin, the system of activated protein C (APC), and the inhibitor of tissue factor [TF] pathway). This can result in the leakage of blood vessels, DIC, and extensive accumulation of fibrin in either the blood vessels or tissues (Figure 3).61,62 In general, coagulation activation is controlled by three vital physiological anticoagulant pathway systems, including the TF pathway inhibitor system (TFPI), APC system, and antithrombotic system, which complement each other to modulate coagulation activation. In the early stages of sepsis, insufficient balance of TFPI in TF-dependent coagulation events supports impaired physiological functions of anticoagulants identified during sepsis.⁶³ In addition, hyperinflammation and innate immunity work together in coordination to amplify the coagulation cascade, in which bacterial-derived products and DAMPs (such as neutrophil extracellular traps [NETs], HMGB1 and cell-free DNA [cfDNAs]) contribute to the development of DIC through multiple pathways. 25,64 In the advanced phases of sepsis, immune cell apoptosis (such as dendritic cells [DCs] and lymphocytes) results in immune stagnation by diminishing the host's capacity to eliminate intruding microorganisms and enhancing susceptibility to subsequent infections.^{65,66} DIC, a key factor in multiple organ failure during sepsis, manifests as the formation of numerous microvascular blood clots, subsequently depleting platelets and coagulation factors, ultimately resulting in impaired hemostasis.⁶⁷ In contrast, these thrombin and coagulation factors exhibit proinflammatory activity by cleaving protease-activated receptors (PARs) to form a vicious cycle in which increased inflammation and coagulation further enhance the mortality rate of sepsis. 25,57,68

2.3 | Complement system

In the late 19th century, scientists made an interesting discovery—a plasma element known as the complement system. This component, which was found to be thermally unstable, served as a complement to antibodies in the process of eliminating microorganisms. This is why it was named "complement." However, it is important to recognize that the complement system can be both beneficial and detrimental. When dysregulated or excessively activated, it can turn against its host, causing harm to tissues, resulting in organ failure, and, in severe cases, leading to mortality. 71,72 The function of complement is



Aggregation of signaling pathways in innate immune cells. Inflammation is initiated at the initiation of sepsis by the sensing of PAMPs and DAMPs. Subsequently, multiple pathways activate and regulate innate immune responses through danger signals sensed by intracellular and membrane-bound receptors. These signaling pathways toward IRF3 and NF-κB initiate early phase inflammatory responses. In addition, LPS or HMGB1 provides an essential priming step of inflammasome activation. An activation step for AIM2/NLRP3 inflammasome assembly necessitates the presence of stimuli called PAMPs and DAMPs, which include crystals and ATP. This step leads to the cleavage of caspase, GSDMD, and pro-IL-1 β /18, ultimately triggering canonical inflammasome activation and pyroptosis. Then, HMGB1 interacts with RAGE to deliver cytoplasmic LPS, which in turn triggers pyroptosis via a caspase-11-dependent pathway of noncanonical inflammasome pathway (equivalent to caspase-4 and caspase-5 in humans). AIM2 and cGAS-STING detect cytosolic DNA and intracellular pathogens, triggering the assembly of the inflammasome and phosphorylation of IRF3. This activation leads to the induction of type I interferon (IFN) responses and inflammasome activation. In the context of sepsis, mitochondrial dysfunction, the generation of reactive oxygen species (ROS), and metabolic reprogramming further augment the production of HMGB1, a key proinflammatory cytokine. The inflammasome plays a pivotal role in the pathogenesis of sepsis, as it is intricately linked to stress signaling, activation of immune cells, and maintenance of cellular homeostasis. In the later stages of sepsis, the impaired process of macrophage activation can occur when the TLR/NF-kB and/or TLR/MARKs signaling pathway is inactivated. This inactivation is induced by various stimuli such as LPS/LTA/PGN and proinflammatory cytokines. P47phox, a component of the NADPH oxidase proenzyme, interacts with Park 7, leading to the phosphorylation and translocation of P47Phox to the membrane. This translocation allows the formation of a holoenzyme complex. The subsequent activation of NADPH oxidase produces reactive oxygen species (ROS). These ROS molecules then trigger the downstream MAPK and NF-kB signaling pathways that are involved in TLR signaling, ultimately resulting in macrophage activation. The activation of macrophages serves multiple functions, including the prevention of sepsis-induced immunosuppression. This prevention is achieved through the release of proinflammatory cytokines, the elimination of pathogens, polarization into the M1 phenotype, and the enhancement of autophagy.



Activation of coagulation, complement, and endothelial cells during sepsis. The loss of antithrombogenicity and exposure of adhesion molecules are outcomes caused by the endothelial glycocalyx. These outcomes enable leukocyte adhesion, platelet recruitment, and the formation of a thrombus. Increased vascular permeability initiates the extravasation of leukocytes, the leakage of plasma proteins, and tissue edema. In response to immunogenic stimulation, macrophages and neutrophils express procoagulation tissue factors in the form of microvesicles that regulate coagulation cascades and prothrombin cleavage. The formation of FXa, thrombin, and fibrin occurs as a consequence of the activation of the cascade of proteolytic reactions. Fibroblasts, pericytes and epithelial cells constitutively express tissue factors that protect hemostasis and vascular integrity. During sepsis, NETosis interacts with platelet-mediated responses to modulate innate immune responses and bacterial clearance. The occurrence of sepsis involves vessel injury induced by inflammation, thereby exposing the blood coagulation factors and leading to the formation of blood clots. The production of APC is mediated by a combination of thrombin and thrombomodulin (TM) and amplified by endothelial protein C receptor (EPCR). During sepsis, APC deactivates the coagulation cofactors FVa and FVIIa, thus impeding the coagulation process. The recruitment of activated platelets and RBCs leads to thrombocytopenia and the presence of platelet-rich thrombus. In addition, HMGBI and efDNA behave as procoagulants, expediting the formation of blood clots. The APC-EPCR interaction also allows for the conversion of PAR-1 signaling into an anti-inflammatory mode through ANG/TIE axis mediated activity and enhances endothelial integrity. In addition, TM/thrombin directly mediates immune-coagulation, and ANG/TIE axis dysregulation is associated with decreased vascular stability during severe infection. Finally, disorders of coagulation, complement and endothelial function lead to microcirculation dysfunction and multiple organ damage, which are closely related to the extensive formation of inflammatory thrombi in sepsis.

beyond infectious defense, which promotes tissue regeneration and repair, facilitates organ development, and can be transformed and coordinated with plentiful processes, including hemostasis leading to inflammation that causes thrombosis. During the treatment phase of patients with sepsis, chemical reperfusion injuries (myocardial infarction, stroke, and transplant dysfunction) trigger (DAMPs) that trigger complement activation in chronic nervous system and rheumatic diseases, thereby contributing to inflammatory and immune disorders. Dysregulation of the complement system marks a disruption in host recognition of defense signaling pathways associated with inflammation, coagulation, and bacterial cell lysis, and is a prelude to multiorgan dysfunction in sepsis. To

The activation of complement cascades is a widely acknowledged phenomenon that occurs via three distinct pathways. These pathways include the classical pathway

(triggered by the complex formed by antigens and antibodies), the alternative pathway (initiated by factors B and D), and the lectin pathway (detected via the recognition of mannose derived from pathogens). After the complement system was activated, these three pathways gathered at the C3 level to produce cleavage products C3a and C5a, as well as the terminal membrane-attacking complex C5b, which forms pores in the cell membranes of bacteria and cells, eventually leading to lysis (Figure 3).⁷⁷ C3a, C5a, and C5b mediate antibacterial responses and proinflammatory effects by interacting with cell membrane receptors and triggering crosstalk among multiple signaling pathways. In sepsis, the innate immune functions of neutrophils are impaired due to excessive C5a generation, for example, chemotaxis, phagocytosis, and H₂O₂ production are blocked. 78,79 Thus, complement-mediated activation of neutrophils is not only responsible for diverse significant



impacts in sepsis but also assumes a contradictory function in innate defense.

Furthermore, sepsis is characterized by an early excessive production of C5a, which leads to an unregulated inflammatory response, ultimately resulting in tissue damage and the failure of multiple organs. 80 Sustained complement activation can cause a massive release of C5a, which may impair the innate immune response of neutrophils, resulting in reduced inflammation and impaired ability to kill bacteria. 81,82 Moreover, elevated levels of C5a can impede the targeted migration of neutrophils. Notably, the secretion of TNF by neutrophils can inhibit the transcription of NF-kB, as induced by the C5a-mediated increase in IκBα levels. Furthermore, C5a disrupts the C3a-C3aR axis and promotes the premature expulsion of granulocytes and hematopoietic stem cells from the bone marrow, leading to a less targeted but more progressive inflammatory response.⁸³ Through C5a signaling, CXCR4 levels on granulocytes are concurrently reduced, while proteases are released, resulting in the degradation of matrix proteins and inhibition of the effect of stromal cell-derived factor 1, which causes phenotypic changes in neutrophils.⁸⁴ Moreover, activation of the complement system triggers the cross-activation of the NOD-like receptor protein 3 (NLRP3) inflammasome and prethrombotic pathways (Figure 2).85 Pentraxins (such as CRP, SAP, PTX3) released in response can initiate the classical pathway by interacting with C1q.86 Apart from pentraxins, the complement system can be directly activated by coagulation proteases through the extrinsic protease pathway (Figure 2).87 The uncontrolled activation of the complement system during sepsis contributes to tissue damage and dysfunction in organs.

2.4 | Mitochondrial dysfunction

Mitochondria play a vital role in energy generation, protein synthesis, regulation of cell growth and cycle, and catabolism.^{88,89} Nevertheless, in cases of sepsis, the mitochondrial damage or malfunction can disrupt cellular metabolism, 41,90-92 impede energy production, and trigger oxidative stress. 93,94 Consequently, this may provoke apoptosis in both organ and immune cells, ultimately leading to immune disorders, multiple organ failure, and potentially fatal outcomes. 95,96 It has been proven that the involvement of specific NLRP3 inflammasome activators could expand mitochondrial instability, resulting in pyroptotic cell death and enhancing the formation of pores in the plasma membrane, which then release inflammatory factors through caspase-1 dependent mechanisms. However, excessive ROS due to electron transport chain damage, Ca²⁺ overload, or reduced endogenous antioxidants can also trigger patterns of cell death, such as apoptosis and

autophagy. 97-99 In the pathogenesis of sepsis, mitochondrial function is disorganized, mainly manifested by a decrease in oxidative phosphorylation, an increase in ROS production, the effects of hormonal changes, downregulation of the genes encoding mitochondrial proteins and an increase in apoptosis, which eventually leads to altered mitochondrial biogenesis. 100-102 Damaged mitochondria release a cascade of DAMPs, further activating and enhancing the immune response. 103,104 The changes in mitochondrial function that occur early in sepsis are an adaptive mechanism by which mitochondria protect cells. For example, the production of harmful ROS is regulated by oxidative phosphorylation. 105,106 In addition, in the early stages of sepsis, mitochondrial autophagy and mitochondrial biogenesis are increased to limit mitochondrial dysfunction. 107,108 Therefore, the development of sepsis is accompanied by changes in different mitochondrial functions.

Furthermore, molecular events resulting from mitochondrial dysfunction in sepsis trigger the activation of caspase-1, which intensifies the response to inflammatory responses, such as perturbations of membrane permeability and damage to the mitochondrial network. 109,110 Although serum proinflammatory factors and antiapoptotic proteins have been shown to play important roles in the activation and inhibition of the NLRP3 inflammasome, the pathogenesis of sepsis-induced acute respiratory distress syndrome is primarily an elevated intracellular oxidative stress-mediated apoptotic event. 111-113 The alterations in the stage of organ function and how persistent organ dysfunction further affects mitochondrial performance remain puzzling, but the confusion is still the subject of translational medicine research because these periodic responses illustrate the potential of targeted therapy to alleviate the organ dysfunction induced by sepsis.

2.5 | Pyroptosis

Pyroptosis is a lytic type of programmed necrosis that is linked to the secretion of proinflammatory cytokines. This process has pivotal functions in facilitating the defensive response of the innate immune system against microbial infections and invading pathogens. ^{114–117} These signature genes in pyroptosis are NLRP3, apoptotic specklike protein containing caspase activation and recruitment domains (CARD) (ASC), cleaved Caspase-1, Gasdermin-D (GSDMD) p30, IL-1 β , and IL-18, which play a prominent role in the response to infection, ultimately fueling inflammation (Figure 2). ^{21,109,110,118,119} The expression levels of inflammatory genes show complex correlations to a certain extent, ultimately leading to the rampant

proliferation of inflammatory gene profiles (such as NLRP3, NLRC4, NOD, IL-1β, and IL-18), while in sepsis studies, the same gene regulation pattern was observed with a higher amplitude of change, revealing its clinical correlation with the severity of sepsis. 39,120,121 It has been established that NLRP3 inflammasome activation requires "prime" and "activation." The NF-κB signaling pathway is an essential priming event that ensures accurate response of the inflammasome to LPS stimulation and prevents inappropriate activation of NLRP3. 124-126 Recent research has suggested that the initiation of NLRP3 deubiquitination may occur through a nontranscriptional mechanism. Specifically, these studies have proposed that mitochondrial ROS are responsible for inducing priming by causing the deubiquitination of the NLRP3 inflammasome. This finding challenges the traditional belief that NLRP3 deubiquitination is primarily regulated by transcriptional processes. Instead, it suggests that a direct interaction between mitochondrial ROS and the NLRP3 inflammasome is involved in this important regulatory step. Further investigation is needed to fully understand the exact molecular mechanisms underlying this nontranscriptional initiation of NLRP3 deubiquitination. 126-128 In addition, LPS-induced cytidine/uridine monophosphate kinase 2 (CMPK2), which is rate limiting for mtDNA synthesis, reduced NLRP3 inflammasome assembly (Figure 2). 129,130 In the absence of NLRP3 activators, prime-signaling provided by NF-κB or CMPK2 is insufficient for activating inflammasomes, suggesting that NLRP3 inflammasome activators play a pivotal role in triggering inflammation.¹³¹⁻¹³³ NLRP3 inflammasome activators include several physically and chemically diverse extracellular stimuli, such as extracellular adenosine triphosphate (ATP) or uric acid crystals, exogenous particulates such as aluminum salts and titanium dioxide. pathogens and pore-forming toxins. 134-138 During sepsis, ATP activates the P2 \times 7 receptor and NLRP3 signal transduction in an autocrine manner after activation of the inflammasome, leading to changes in metabolic status and convergence of inflammasome signals and is associated with increased lethality. 139,140 Mechanistically, these inflammasome activators function to increase downstream binding of NEK7 with NLRP3^{141–143} and regulate NLRP3 oligomerization, inflammasome assembly, and catalytic cleavage of procaspases.

Recently, a novel TLR4-independent mechanism for triggering pyroptosis was discovered. 144,145 Intracellular LPS directly binds to the procaspase-11 recruitment CARD domain, which then oligomerizes and activates procaspase-11. 146,147 The induction of pyroptosis, which is a critical event in septic mice, is dependent on Caspase-11 activity. 44,148-150 Additionally, pyroptosis in human mononuclear cell lines is mediated by homolo-

gous caspase-4/5. The activation of NLRP3 is regulated by Caspase-11, and it is important to note that the cleavage of GSDMD by active caspase-1/4/5/11 leads to the release of the functional gasdermin-N domain. This released domain subsequently forms pores in the membrane, facilitating the active release of inflammatory cytokines and intracellular components (Figure 2).^{151–156} The ability of the gasdermin family to form pores plays a significant role in the unique molecular and structural mechanisms underlying pyroptosis. 157,158 Interestingly, the pore formed by GSDMD-N enables nonselective ion diffusion without increasing osmotic pressure, unlike membrane blebbing and cell swelling. 159,160 Mitochondrial dysfunction may trigger the release of oxidized mtDNA and cardiolipin into the cytosol, consequently activating the NLRP3 inflammasome and leading to the production of IL-1\beta. Furthermore, the activation of inflammasomes can be driven by the significant efflux of potassium induced by toxins such as α -toxin, which form potassium channels. ^{161–164}

2.6 | Neuroinflammatory reflex

Homeostasis of neuroinflammatory reflex system interactions is also considered to be a crucial part of the host response during sepsis. 165 The neuroinflammatory response to sepsis is not due to infection of the brain but rather systemic inflammation associated with innate immunity to the central nervous system (CNS) through various pathways.¹⁶⁶ LPS can activate TLR2 and TLR4 surrounding the cerebrum, choroid plexus and leptomeninges, thus profoundly inducing the transcription of inflammatory mediators throughout the brain parenchyma.¹⁶⁷ However, the neuroinflammatory reflex immune system can suppress inflammation by activating the splenic nerve in the celiac nerve plexus, which causes the spleen to release norepinephrine (NA) and CD4⁺ T cell subsets to secrete acetylcholine. The characteristics of cerebral dysfunction in sepsis are altered mental states, including disorientation and confusion. 168,169 Patients with severe formal sepsis related to cerebral dysfunction may develop coma. 170,171 Importantly, sepsis-associated cerebral dysfunction can be present without other organ dysfunctions (such as liver failure) that might be associated with encephalopathy. 40 As a result, patients experiencing cerebral dysfunction associated with sepsis face a significant mortality risk and commonly endure prolonged cognitive and functional aftereffects. The activation/dysfunction of the vagus nerve and endothelium triggers the release of cytokines and neurotoxic mediators. This mechanism exemplifies the brain's immune system operations by transmitting distinct signals along various pathways to the brain. These incoming signals subsequently prompt the

CNS's outgoing response, activating both the sympathetic and parasympathetic branches of the autonomic nervous system. Consequently, the activation of the adrenal axis is regulated, which counteracts the inflammatory process and restores cardiovascular homeostasis. 172,173 Additionally, the production of cytokines is primarily regulated by a specific division of the autonomic nervous system, particularly the sympathetic branch. 174 Existing data suggest that NA can react to LPS, and elevated levels of NA were observed in close proximity to immune cells, inhibiting the expression of TNF- α and IL-12 (proinflammatory cytokines), while promoting the expression of IL-10 (an anti-inflammatory cytokine).¹⁷⁵ Furthermore, acetylcholine signaling, as a neurotransmitter, also plays a crucial role in regulating the inflammatory response of sepsis, thereby introducing novel avenues for sepsis treatment. 176,177

2.7 | Necroptosis

In contrast to caspase-dependent programmed cell death, necroptosis is an unprogrammed and autonomous form of cell death that can be activated by various virulence factors discharged by the pathogen. However, sepsis-induced changes in organ perfusion and hypovolemia can also contribute to necrosis.¹⁷⁸ This process, in turn, can bolster the release of extracellular "alarm proteins" such as HMGB1, thereby exacerbating local inflammation. Therefore, necroptosis plays an essential role in cell death during sepsis. Additionally, necroptosis triggers inflammation and immunogenic events in different diseases by interacting with death receptors and diverse cytoplasmic protein kinases. Unlike necrosis, which usually results in irreversible pathological damage, necroptosis is employed by organisms to acquire appropriate levels of energy supply and promote cell regeneration. 179,180 Initially, defined as a type of TNF-induced necrosis, necroptosis was subsequently recognized as a specific kinase-dependent process after being effectively subdued by RIPK1 inhibitors. 181-183 Necroptosis is specifically regulated by signal transduction proteins known as RIPK1 and RIPK3, which operate in complexes of membraneassociated proteins. These complexes are activated by TLR (TLR3, TLR4) via rhomboid-like (RHIM) domaincontaining proteins (TRIF, DAI). 184-186 Successive stimuli induce interactions between coupling proteins and kinases and then initiate necroptosis cytoplasmic complexes, thereby responding by inducing signal transduction associated with cell death/survival outcomes. 187-189 Necroptosis can be induced by various stimuli other than TNF/TNFR1, including immune death signals (FASL/TRAIL), bacterial and cellular stress signals (LPS/TLR4, poly(I:C)/TLR3),

and type I/II IFNs. 190-192 These stimuli trigger the formation of necrosomes by activating death receptors. Furthermore, the recruitment of RIPK1 to FAS without an inhibitor of apoptosis leads to the assembly of necrosomes and necroptosis. 193 On the other hand, activation of the cytosolic complex promotes cell survival and proinflammatory signaling through NF-kB and MAPK activation. 194,195 However, the mechanisms that determine whether a cell undergoes apoptosis, necroptosis, or cell survival are still not fully understood. Genetic studies have shown that deficiencies in FADD or Caspase-8 can result in RIPK3-mediated necroptosis, leading to embryonic lethality and inflammation. 196-198 In clinical sepsis, levels of RIPK3 were found to be significantly elevated at different time points. 199,200 Moreover, higher levels of RIPK3 were associated with an increased incidence of organ dysfunction and septic shock, highlighting the pathological effects of RIPK3-mediated pathways.

Collaboration between necroptosis and pyroptosis can expand the transmission of inflammatory signals, cause tissue injury, and exacerbate organ failure, both of which play pivotal roles in the progression of sepsis. 201,202 Although both processes can be triggered by similar stimuli, different pathways are involved in intracellular signaling, targeting distinct death regulatory proteins, specifically RIPK3 or GSDMD.^{203–205} The release of intracellular components differs in the active stage of sepsis development; necroptosis leads to the release of HMGB1, while pyroptosis results in the release of the proinflammatory cytokines IL-1 β and IL-18. ^{26,206} To explore potential synergistic effects in sepsis-induced injury, studies have investigated the protective effects of dual deletions of RIPK3 and GSDMD. Knockout (KO) of RIPK3/GSDMD or mixed lineage kinase domain-like/GSDMD has demonstrated protective effects against septic shock, systemic coagulation, and multiple organ injury caused by necroptosis and pyroptosis double obstruction. 207,208 Both RIPK3 and GSDMD contribute to lytic cell death, which collaborates to amplify necrotic inflammation and the release of TF in macrophages and ECs, resulting in significant tissue damage (Figure 3). Hence, RIPK inhibition is proposed to play a protective role in the sepsis model. A recent in vivo study using LPS-stimulated human kidney tubular epithelial cells showed increased RIPK3 expression, with enhanced insights into the unique role of RIPK3 in regulating mitochondrial function during sepsis, as evidenced by observations of mitochondrial depolarization in vitro.

2.8 | Endothelial barrier dysfunction

The endothelium forms the inner layer of cells from blood vessels and lymphatics and plays a vital role in controlling

blood flow and vascular tone as well as participating in immune responses.²⁰⁹ The destruction of the endothelial barrier is the most prominent phenomenon in the pathogenesis of sepsis. The integrity of the endothelial barrier is maintained by intracellular junction molecules, which can regulate the anticoagulant and anti-inflammatory immune properties of the endothelial intima in the process of sepsis.²¹⁰ Oxidative stress and inflammation lead to decreased vascular permeability, hypotension, and inadequate organ perfusion and eventually result in organ failure.⁶⁸ These signature features play a key role in the pathogenesis of sepsis-induced multiple organ failure.

Altered endothelial function was activated by disruption of intracellular junction molecule expression and dynamics of cytoskeleton contraction. 17,211,212 In sepsis, a large number of novel therapeutic targets have been discovered, such as sphingosine 1-phosphate 1, PAR1, vascular endothelial growth factor (VEGF), and angiopoietin 1 (Ang-1), which have been found to play a regulatory role in endothelial dysfunction (Figure 3).213-215 The antiinflammatory properties of Ang-1 derive from its ability to reduce leukocyte transmigration and cytokine production via Tie receptor 2 (Tie2) receptor signaling at the endothelial barrier. 47,216,217 Inflammation-induced release of Ang-2 disrupts constitutive Ang1-Tie2 signaling by preventing Ang-1 from binding to its receptor in sepsis, thereby reducing physiologic efforts to suppress ongoing inflammation. 218-220 Therefore, the Ang/Tie2 pathway has broad impacts on vascular remodeling, inflammation, and EC survival. 221,222 In addition, Ang/Tie2 activation induces the expression of KLF2 via the phosphoinositide-3 kinase/Akt pathway and counteracts VEGF-mediated vascular permeability (Figure 3).²²³

2.9 | ER stress

The ER is an intracellular organelle that is involved in protein translocation, folding, posttranslational modification, and further transport to the Golgi apparatus. 224 Moreover, the ER provides a place for calcium storage, lipid synthesis and carbohydrate metabolism. 225,226 Under stress and inflammation conditions, the ER loses homeostasis in its function, which is termed ER stress. Unfolded or misfolded proteins accumulate in the ER during sepsis, altering its homeostasis and leading to oxidative stress and severe calcium disorders that result in ER stress. 227,228 Recent studies on ER stress signaling have revealed intriguing interactions between ER stress and sepsis-related cell death. 229–231 Recently, many studies have demonstrated that inhibition of ER stress can stabilize protein conformation, promote the transport of mutant proteins and improve ER folding

ability.²³² Therefore, ER stress is suggested as a potential therapeutic target for a variety of diseases, including diabetes, cystic fibrosis, sepsis and ischemic brain injury.²³³ In sepsis animal models, markers of increased ER stress (such as glucose-regulated protein 94 [GRP94], CEBP homologous protein [CHOP], and caspase-12) are detected in several organs, including the heart and liver, and these markers are directly connected with the extent of organ dysfunction, which may be a major cause of sepsis-induced multiple organ failure. 234-236 Studies have demonstrated that caspase-12 deficiency confers protection in septic mice and that the presence of caspase-12 leads to enhanced vulnerability to bacterial infection and septic mortality, suggesting a detrimental role of caspase-12 in sepsis.²³⁷ These effects have been demonstrated in cecal ligation and puncture (CLP) models of sepsis, where ER stress leads to abnormal lymphocyte apoptosis in mice during sepsis, suggesting that the ER stress-mediated apoptosis pathway may be a new target for the clinical prevention and treatment of sepsis-induced lymphocyte apoptosis. 15 During sepsis, the transcription factor CHOP is a major inducer of apoptosis in response to ER stress; however, recent evidence suggests an inflammatory role of CHOP as a mediator of the inflammatory response in sepsis.²³⁸ A major role for CHOP, which acts as an amplifier of the inflammatory response in the pathogenesis of sepsis, and the ability of H₂S treatment to counter CHOP signaling via upregulation of NF-E2-related factor 2 are highlighted.²³⁹ ER stress causes abnormal apoptosis in septic animals, suggesting that ER stress-mediated apoptosis represents a potential new target for the clinical prevention and treatment of sepsis. 240,241 However, these observations increase our knowledge of the biological mechanisms in the context of ER stress and sepsis and simultaneously shed light on new targets and suggest novel strategies for the treatment of this condition. Further research is warranted to elucidate the exact mechanism of how ER stress contributes to sepsis-associated cell death.

2.10 | Selective autophagy

Autophagy is a vital innate immune defense and self-protective mechanism against microbial challenges. It is a self-degradation process of basic metabolic mechanisms that degrades intracellular proteins and dysfunctional organelles to maintain cellular homeostasis and survival. ^{242,243} Increasing evidence shows that autophagy malfunction is associated with many human diseases, such as cancer, neurodegenerative diseases, liver diseases, and inflammatory diseases. ²⁴⁴ It is noteworthy that autophagy can be mobilized in various organs in early sepsis, manifested by increased accumulation of autophagic

vesicles and enhanced expression of autophagy-related proteins. 245,246 Autophagy is likely to have a protective effect in sepsis through the mechanisms described below: elimination of pathogens, neutralization of microbial toxins, regulation of cytokine release, reduction in apoptosis, and promotion of antigen expression.^{247–249} Importantly. mitochondria-produced ROS then oxidize HMGB1, which is released by apoptotic cells, resulting in the elimination of its immunogenicity and the inability to activate innate immune cells. ^{233,250,251} However, during sepsis, the reduction of oxidized HMGB1 by thioredoxin is not efficient. 252 HMGB1 directly interacts with the autophagy protein Beclin-1, displacing Bcl-2, which leads to the formation of autophagy initiation complexes and the elimination of harmful oxidative stress triggers. 243,252-255 When binding to AIM2, all forms of intracellular HMGB1 that have been reduced by thiol display the highest affinity and subsequently initiate inflammasome signaling during redox stress (Figure 1).²⁵⁶ Therefore, activation of the inflammasome pathway is a critical prerequisite for promoting protective autophagy and enhancing cell survival.

Sepsis-induced autophagy is initiated by binding PAMPs within the microbial structure to PRRs, such as TLRs, which further trigger different intracellular events and lead to increased autophagic activity by promoting the conversion of LC3-I (microtubule-associated protein 1A/1B-light chain 3-I) to LC3-II.^{257,258} The binding of LPS from gram-negative pathogens to TLR4 activates autophagy through the MAPK/P38 (mitogen-activated protein kinase) signaling axis, while the binding of lipoteichoic acid to TLR2 induces autophagy via the MAPK1/ERK2-MAPK3/ERK1 pathway.²⁵⁹⁻²⁶¹ Thus, different TLR ligands exhibit variable autophagy-inducing abilities.^{262,263} Although autophagy levels were regulated by the severity of sepsis, mitochondrial biogenesis was successfully restored by injection of cell-permeable TATI-beclin-1 through the PINK1/Parkin and AMPactivated protein kinase (AMPK)/Unc-51-like protein kinase 1 signaling pathways and maintained sepsis cardiac function.²⁶⁴ Beclin-1 plays a vital role in supporting PINK1/Parkin-mediated mitophagy by interacting with Parkin. This interaction helps localize mitochondrialrelated membranes, leading to the tethering of ER mitochondria and the formation of mitochondrial autophagosome precursors. 233,265 The recruitment and activation of Parkin by the PINK1 protein on the mitochondrial outer membrane is the initial step in this process. Parkin then constructs ubiquitin chains on damaged mitochondria, facilitating their binding with LC3 on autophagosomes to induce mitophagy (Figure 1). 227,266,267 Furthermore, crosstalk can occur between autophagy and apoptosis. 268,269 In a CLP mouse model of sepsis, T cell apoptosis was induced, while autophagy in CD4⁺ spleen

cells and CD8⁺ spleen cells was downregulated.^{3,270} In addition, mice with lymphocyte-specific deletion of the Atg5 or Atg7 genes, which are needed for autophagy, had increased mortality, immune dysfunction, and T-cell apoptosis compared with control mice. For example, T cell-specific deletion of Atg5 in mice leads to increased Il-10 production by T cells after CLP.^{271,272} These findings suggest that inadequate autophagy results in sepsis-induced immunosuppression.

3 | POTENTIAL THERAPEUTIC TARGETS: SIGNALING MODULATORS

Due to sepsis being a complex disease condition, multiple targets and target external environment interactions are necessary for the occurrence and clinical manifestations of sepsis. Despite substantial improvements in care delivery, intensive drug therapy and immunosuppression continue to lead to multidrug-resistant organisms and opportunistic infections, as well as long-term outcomes such as organ failure, immunosuppression, and disability. Various candidate target studies have identified polymorphisms, but these results have not been consistently confirmed. To further understand the pathogenesis of sepsis and the results of new interventions, nine targets, such as GSDMD, HMGB1, and STING, were evaluated according to the important molecular signaling pathways of sepsis, which play vital regulatory roles in the occurrence and development of sepsis.

The nine crucial targets in dangerous liaisons act as linchpins in the events that lead to sepsis. One of the signature events in mediating protective innate immune responses against invading pathogens and microbial infections is the cleavage and pore formation of GSDMD. This event plays a decisive role in regulating inflammation, pyroptosis, and coagulation.²⁷³ Regarding the coagulation process, extracellular HMGB1 has been shown to enhance TF procoagulant activity by promoting the externalization of PS to the outer plasma membrane. Intracellular HMGB1 binds to bacterial endotoxin and transmits it to the cytoplasm through the RAGE receptor on vascular ECs and macrophages.²⁷⁴ This transmission triggers caspase-11mediated pyroptosis, eventually resulting in shock, multiorgan failure, and death. Although CASP-11 activation converges as an upstream signal for GSDMD-dependent PS exposure, other mechanisms, such as NETosis, platelet activation, and disruption of the endothelial Tie2 axis, may contribute to the coagulation cascade. STING, a classical innate immune sensor that stimulates the expression of IFN in response to DAMP (cytoplasmic DNA), has been identified to be involved in coagulation by triggering ER stress-induced activation of GSDMD. 275,276 STING

interacts with the major calcium channel ITPR1 to promote ER calcium efflux, which is necessary for CASP-1/11/8-induced cleavage and activation of GSDMD. Eventually, ER stress-mediated pore formation in GSDMD triggers pyroptosis and subsequent release of TF. Persistent innate immune activation by STING continuously promotes the process of inflammasome coagulation present in GSDMD pores. 277,278 SQSTM1 is released into the extracellular environment by macrophages through two mechanisms (active secretion or passive release). Extracellular LPS induces SQSTM1 lysosomal secretion through activation of the TLR4-MYD88-STING1 pathway, while intracellular LPS triggers the release of SQSTM1 through pyroptosis via activation of the CASP11-GSDMD-GSDMDN pathway.²⁷⁹ After release, SOSTM1 binds to the receptor insulin receptor (INSR) to activate the NF-kB pathway, leading to the polarization of proinflammatory macrophages and ultimately mediating septic death in mice through hyperinflammation and coagulation.²⁸⁰ Recent studies have shown the important role of regulating immune checkpoints in sepsis-induced immunosuppression. HMGB1 is secreted by melanocytes, which in turn activates the RAGE and promotes NF-κB and IRF3-dependent PD-L1 transcription in melanocytes. In addition, PD-1 and PD-L1 are important regulators that inhibit T cell receptor-induced activation signals, and regulation of the PD-1/PD-L1 axis can correct the immunosuppression of sepsis.²⁸¹⁻²⁸³ Oxidative stress is caused by an imbalance between the production of ROS and the body's ability to remove these toxic intermediates. Park 7 is an unparalleled antioxidant that independently functions in cellular defense against ROS, without relying on other antioxidant pathways. It plays a vital role in regulating the generation of ROS through its interaction with p47phox, a subunit of NADPH oxidase. 284-286 ROS act as a pivotal mediator, initiating the TLR signaling pathway to activate macrophages. The mounting evidence strongly suggests that Park 7 acts as an antagonist of sepsis-induced immunosuppression, indicating its potential as a novel therapeutic target for reversing impaired immunity caused by sepsis. The activation of complement leads to the discharge of C3a and C5a, which possess immense proinflammatory properties, such as the recruitment and activation of leukocytes, ECs, and platelets. While complement activation plays a crucial role in the initial defense mechanisms of the immune system, uncontrolled activation leads to septic shock.⁸⁶ SESN2, a stress-inducible protein, inhibits the long-lasting activation of the NLRP3 inflammasome by inducing mitophagy in macrophages to eliminate damaged mitochondria. SESN2 performs a dual function in the induction of mitophagy activated by inflammasomes. First, SESN2 initiates "mitochondrial priming" by marking mitochondria for recognition by

the autophagy machinery. To prepare the mitochondria for this process, SESN2 causes the aggregation of mitochondria near the nucleus, mediated by the aggregation of SOSTM1 and its binding to the ubiquitin linked to lysine 63 (Lys63) on the mitochondrial surface. Second, SESN2 stimulates specific autophagy mechanisms to breakdown the targeted mitochondria by increasing the levels of ULK1 (UNC-51-like kinase 1) protein. ²⁸⁷ Furthermore, prolonged LPS stimulation induces an increase in SESN2 expression in macrophages by mediating NO (nitric oxide) through NOS2 (nitric oxide synthase 2, inducible).²⁸⁸ Therefore, mice deficient in SESN2 display impaired mitophagy in two distinct sepsis models, leading to hyperactivation of the inflammasome and ultimately elevated mortality. Targeting JAKs and/or STATs could become an important approach to reducing mortality in septic shock, as these proteins are clearly associated with immune dysfunction and MOF. Original JAK-STAT-based therapies should be designed to treat sepsis-induced immunosuppression and may reach novel targets in the field.^{289,290} Furthermore, phosphorylated STAT may provide creative therapeutic markers to drive such therapy. Therefore, we selected these nine targets for a detailed description of their major regulatory molecular pathways and potential in the development of sepsis.

3.1 | GSDMD: promoters of inflammation and pyroptosis

Recent studies have found that gasdermin (GSDMs) can induce the transition from apoptosis to pyroptosis, and induce inflammatory and antitumor immunity. Activation of GSDMs can occur through caspases and granzymes, resulting in apoptosis in various scenarios. 150,151,291,292 For instance, when the level of GSDM expression is insufficient to induce pyroptosis, caspase/granzyme-driven apoptosis can be converted into pyroptosis by expressing GSDMs.²⁹³⁻²⁹⁵ The occurrence of pyroptosis is primarily associated with inflammation, which arises from the assembly of a multiprotein complex in response to PAMPs, DAMPs, or environmental stress. Notably, the regulatory network of GSDMD, which plays a significant role in inflammatory diseases, has demonstrated its ability to safeguard animals against septic shock or lethal endotoxemia. 296,297 Hence, GSDMD is regarded as a promising target for sepsis treatment.

The pore-forming properties of the gasdermin family play a crucial role in pyroptosis, which is a form of cell death with inflammation.²⁹⁸ When active caspase-1/4/5/11 cleaves GSDMD (gasdermin protein), it releases the functional gasdermin-N domains. These domains have the ability to induce liposome permeation and form intrinsic

pores in the cell membrane. This process ultimately leads to the formation of membrane pores and the release of inflammatory cytokines, specifically IL-18 and IL-1β. Studies have shown that deleting GSDMD can block pyroptosis and provide protection against sepsis, systemic blood coagulation, and multiorgan damage. 279,299,300 GSDMD not only induces cell death but also contributes to necroinflammation by releasing TF in macrophages and ECs, resulting in significant tissue damage. 121,301 Moreover, evidence suggests that GSDMD is involved in coagulation responses during sepsis. It promotes the release of coagulation factor III (F3) into the bloodstream, leading to systemic coagulation. Activation of GSDMD through caspase 11 (CASP11) is needed for this process. 60,302 Interestingly, GSDMD-independent pyroptosis relies on the activation of the purinergic receptor $P2 \times 7$ (P2RX7), but disrupting P2RX7 does not affect the coagulation cascade in septic mice. 302,303 This emphasizes the unique role of GSDMD in promoting coagulation.^{304,305} Additionally, experiments with intraperitoneal injections of nuclear sulfonamides, such as pseudokinase, have shown that they can directly inhibit GSDMD. This inhibition offers protection against sepsis in mice compared with control groups. 273,304 Based on these findings, the GSDMD system is considered a potential therapeutic target for treating sepsis and warrants further investigation.

3.2 | HMGB1: crucial intermediators between inflammation and coagulation

HMGB1 was initially identified as a protein that binds to the nucleus and enhances the transcription of genes by ensuring the stability of nucleosome formation. Follow-up investigations have validated the involvement of HMGB1 in various cellular processes, including DNA recombination, repair, replication, and gene transcription. These activities are facilitated by the presence of repeated positively charged domains (HMG boxes) within the Nterminal region of HMGB1.306 Thus, human or mouse HMGB1 protein not only functions as a nuclear factor but also functions as a vital cytokine mediating responses to infection, injury, and inflammation.³⁰⁷ Once it is actively secreted by immune cells or the epithelium or released by damaged cells, extracellular HMGB1 can serve as a DAMP and lethal mediator in critical diseases such as sepsis and COVID-19 in both mice and humans. 308,309 Recent research has indicated that HMGB1 may potentially trigger inflammasome activation through cell interactions in mouse sepsis models.^{55,310} As a late-phase alarmin in sepsis, HMGB1 interacts with RAGE to deliver cytosolic LPS, consequently activating the NLRP3 inflammasome in murine macrophages and lung ECs, leading to caspase11-dependent pyroptosis. 274,310,311 HMGB1 also binds to cellular receptors for RAGE that enter the lysosomes of macrophages and ECs and trigger activation of the transcription factors NF- κ B and MAPK. 312 In addition, HMGB1 destabilizes lysosomal membranes and promotes LPS release into the cytoplasm, leading to pyroptosis, and thus promoting lethal coagulation. $^{313-315}$ These results establish a pathway dependent on DAMP for the transportation of LPS into the cytoplasm, triggering the activation of CASP11 and coagulation. This pathway operates independently from the non-DAMP pathways previously documented, which involve the release of outer membrane vesicles by bacteria or the involvement of human guanylate binding protein 1 (GBP1). $^{316-318}$

Note that, although the expression of CASP11 induced by LPS requires TLR4, it is not necessary for sepsis death induced by poly(I:C), which may help explain the unsatisfactory results of clinical trials attempting to treat sepsis with TLR4 inhibitors. 121,319,320 These findings suggest that in mice, the activation and formation of pores by HMGB1 are necessary for F3 activation through the exposure of phosphatidylserine (PS). 306,313 As a result, PS-binding proteins such as lactadherin and MFG-E8 can restrict cytoplasmic F3 activation induced by LPS in murine macrophages. 321,322 An unexpected discovery was that glycine, an osmoprotectant, inhibits the release of F3 from infected BMDMs but does not affect cytoplasmic F3 activation induced by LPS in murine macrophages. 302,303,323-325 Several possible reasons exist for these seemingly contradictory outcomes: the disruption of the plasma membrane by Ninjurin 1 during lytic cell death, including pyroptosis, might also contribute to HMGB1 release and F3 activation during lethal infection in mice. 278,324,326,327 In vitro studies have concluded that GSDMD-mediated influx of Ca²⁺ promotes F3 activation, partly through anoctamin 6 (TMEM16F). 328,329 In contrast to pyroptosis mediated by GSDMD-induced release of HMGB1 in macrophages, GSDMD-deficient mice typically release HMGB1 during endotoxemia. 243,299,330,331 Consequently, these findings suggest the possibility of overlapping and diverse functions of GSDMD and inflammasomes in regulating HMGB1 release, which may play a role in feedback mechanisms controlling inflammation and coagulation.

3.3 | STING: multiple regulatory functions of STING in sepsis

Stimulator of interferon response, cGAMP interactor 1 (STING1 or TMEM173), is a transmembrane adaptor protein that is commonly found in the ER of both human and mouse cells.³³² Recently, the activation of the STING1

pathway in myeloid cells, such as macrophages and monocytes, has been associated with inflammation, coagulation, and tissue damage in mouse models of sepsis. These models include polymicrobial infections or bacteremia induced by *Escherichia coli* or *Streptococcus pneumoniae*. To better understand the role of STING in regulating the innate immune response to sepsis, extensive research has been conducted to identify key regulators of the STING pathway. Among the potential targets, anapestic lymphoma receptor tyrosine kinase (ALK) has been proposed as a novel regulatory factor for the STING-mediated innate response. ALK, a receptor tyrosine kinase commonly associated with tumors, exhibits relatively low expression levels in healthy individuals.

The binding of ALK/EGFR has been theoretically demonstrated to activate STING in innate immune cells dependent on Akt. This finding provides supporting evidence for a novel signaling pathway implicated in the development of sepsis pathogenesis and septic shock.³³⁷ Therefore, the ALK-EGFR-Akt pathway plays a critical regulatory role in the STING-mediated innate immune response, which has been proven to be another important pathogenesis of fatal sepsis. In previous studies, STING mutations were shown to perturb ER calcium homeostasis and drive T cells to overreact to ER stress induced by TCR signaling.338 This chronic increase in STING-mediated ER stress can effectively trigger the death of T cells due to apoptosis.339 Additionally, the severity of DIC and mortality in sepsis patients is linked to the expression of STING and GSDMD. 279,292,303 Systemic coagulation mediated by STING is not dependent on the classical STING-induced pathway. It is noteworthy that during endotoxemia in mice, STING1-induced apoptosis of CD4⁺ T cells promotes inflammation-induced immunosuppression, indicating the potential of STING as a target for treating T cell-mediated diseases. 278,324 Moreover, when cytosolic DNA is recognized by cyclic GMP-AMP synthase (CGAS), activated STING1 can be moved to lysosomes, causing lysosomal rupture and subsequent K⁺ efflux, ultimately leading to the activation of the NLRP3 inflammasome. 340,341 GSDMD has been shown to either limit or induce CGAS-mediated STING1 activation by measuring the production of type I interferon genes in murine macrophages and ECs, respectively. This suggests that GSDMD has a dual function in the feedback control of STING1 activation. 277,342 In summary, the activity of STING1 is regulated by various binding proteins or posttranslational modifications associated with immunity, autophagy, and cell death, highlighting the complex contribution of STING1 to the development of diseases, particularly sepsis and coagulation-related conditions.^{275,343} Recently, the STING pathway has been

proposed as another viable therapeutic target for sepsis due to its multiple effects on immune homeostasis, coagulation, and inflammation.

3.4 | C3a/C5a: the double-edged sword of the innate immune system

In the complement system, the most crucial molecule to inhibit is C3 because it obstructs all connections of the entire system to C3 and downstream. 71,78,344,345 Hence, there is a significant potential to decelerate all complement-mediated functions by obstructing C3.73,346 It was verified that a synthetic inhibitor of C3 convertase (Compstatin) not only impeded complement activation during E. coli-induced sepsis in baboons but also mitigated other inflammatory responses, activation of coagulation, and multiple organ failure. 71,347 It is noteworthy that while blocking C3, the cascade is not completely intercepted by the other constituents, since activation of the classical and lectin pathways produces some C5 convertase, and some activation of the terminal pathway transpires. 130,348 Nevertheless, blocking C3a or C3aR might also serve as an alternative to inhibiting the anaphylactic toxin activity of C3 cleavage, although this approach has not been tested in clinical trials. C5a demonstrates the highest potency as an inflammatory mediator of the complement system. 69,79,349 Blocking C5a signaling improved outcomes in multiple E. coli-induced sepsis animal models, including monkeys, mice, and rats with a variety of types of microbial sepsis.^{350,351} According to a recent investigation, baboon survival was enhanced, coagulopathy was decreased, and endothelial and barrier function were maintained in an E. coli-induced sepsis model through the utilization of RA101295, a C5 peptide inhibitor. This study observed a noteworthy reduction in consecutive identical words, thereby demonstrating the effective role of RA101295 in combating sepsis-related challenges. 71,352 Therefore, C3a and C5a receptors may have different regulatory functions in sepsis. Nevertheless, recent research has illustrated that the utilization of a C5a receptor blocker resulted in improved survival rates across various forms of microbial sepsis. Conversely, the administration of a C3a receptor blocker led to diminished survival rates. 353-355 Consequently, it can be concluded that complement activation plays a significant role in the development of multiorgan dysfunction syndrome during sepsis. Consequently, the inhibition of complement fragments and/or their receptors could be a valuable approach in sepsis treatment. Despite the successful development of agents specifically designed to inhibit complement activation, their complexity and, in some instances, unfavorable effects hinder their



therapeutic applicability. Further investigation into the alterations in complement activation throughout sepsis may aid in identifying patients and interventions that are most likely to yield positive outcomes.

3.5 | Park 7: park7 is a unique antioxidant that blocks inflammation and ROS

Macrophages, as crucial cells in the innate immune system, have a significant impact on inflammation and immune processes. 356-358 In the initial phases of sepsis, macrophages commonly exhibit proinflammatory characteristics. However, an excessive inflammatory response by macrophages can trigger macrophage apoptosis, disrupt macrophage polarization, and ultimately lead to immunosuppression. 359,360 ROS can activate various TLR signaling pathways, subsequently governing macrophage function. These ROS are generated through the activation of NADPH oxidase. 361,362 Currently, the indispensable role of Parkinson's disease protein 7 (Park 7) in regulating the production of ROS by interacting with P47PHOx, a subunit of NADPH oxidase, is well known.³⁶²⁻³⁶⁴ This interaction activates NADPH oxidase, leading to an increase in ROS levels in macrophages, thereby initiating TRL signaling and enhancing macrophage function to counteract sepsis-induced immune suppression. 365,366 Consequently, mice with Park7 KO can serve as an ideal model for studying advanced sepsis.^{367,368} More specifically, during the later stages of sepsis, the impaired activation of macrophages can be attributed to the dampened TLR/NF-κB and/or TLR/MARKs signaling pathways induced by LPS/LTA/PGN/proinflammatory cytokines. 285,286,369 P47phox, which serves as the proenzyme subunit of NADPH oxidase, plays a critical role in the assembly of NADPH oxidase.370,371 The interaction between Park 7 and p47phox facilitates the phosphorylation and membrane translocation of p47phox, ultimately leading to the formation of the holoenzyme complex. 372,373 Subsequent activation of NADPH oxidase results in the generation of ROS, which, in turn, activate the MAPKs and NF-kB signaling pathways downstream of TLR signaling, thereby promoting macrophage activation.^{374–376} Activated macrophages play a crucial role in preventing sepsis-induced immune suppression by releasing proinflammatory cytokines, eliminating pathogens, polarizing toward the M1 phenotype, and enhancing autophagy capacity.^{377–379} Based on this knowledge, targeting the Park 7/P47PHOX/ROS axis could prove to be an effective therapeutic strategy against sepsis-induced immunosuppression.

3.6 | SQSTM1: 'Jack of all trades' in health and septic shock

Sequestosome 1 (SQSTM1/P62) serves not only as a receptor for macroautophagy/autophagy but also as a versatile protein engaged in signaling cascades during inflammation and oxidative stress. 380,381 In vitro and in vivo experiments have emphasized the role of extracellular SQSTM1 as a deadly inflammatory mediator leading to sepsis and septic shock mortality, according to recent research. 280,382 In brief, SQSTM1 release occurs during tissue damage or microbial invasion, mainly in two ways: passively and actively. Passive release of SQSTM1 from human or mouse macrophages and monocytes is induced by the activation of TLR4-mediated transactivation of the Sqstm1 gene and STING1-mediated phosphorylation of the SQSTM1 protein at Ser403 in the active mode. 383,384 In vitro, cytoplasmic LPS also triggers GSDMD-dependent pyroptosis, which further facilitates the passive release of SQSTM1 from macrophages and monocytes. 279,280 Following its release, extracellular SOSTM1 binds to the INSR located on the membrane, resulting in the activation of glycolysis. This activation subsequently leads to the production of proinflammatory cytokines in a manner dependent on the transcription factor NF-kB. 385,386 Functionally, the pathway involving SQSTM1 and INSR can have a significant impact on tissue damage, systemic inflammation, organ failure, and mortality in experimental sepsis mouse models when either genetically deleted or pharmaceutically inhibited. 387,388 Moreover, the severity of sepsis in patients has been found to be associated with the activation of the SQSTM1-INSR pathway.

The SQSTM1-INSR axis activates NFKB and triggers inflammation, which occurs in macrophages via a pathway involving phospholipase C gamma 1 (PLCG1)-dependent lipid peroxidation. This process leads to elevated pyroptosis, a form of cell death. 389,390 Blocking SQSTM1 by administering neutralizing monoclonal antibodies or deleting the INSR or SQSTM1 genes in bone marrow cells (BMCs) effectively safeguards mice from fatal sepsis. 279,280 Patients with bacterial sepsis show increased levels of SQSTM1 and INSR mRNA in peripheral blood mononuclear cells, along with elevated serum SQSTM1 concentrations. This finding further supports the idea that the SQSTM1-INSR pathway may have a detrimental role in human sepsis. 391,392 Additionally, higher serum SQSTM1 concentrations have been found to be an independent risk factor for patients with steatosis and liver inflammation.36,398 Therefore, revealing its proinflammatory potential would help shed light on the regulatory role of extracellular SQSTM1 in the pathogenesis of various inflammatory diseases.

3.7 | PD-1/PD-L1: regulatory immune checkpoints in sepsis

PD-1/PD-L1 signaling pathways play a crucial role in the immunosuppression caused by sepsis. These pathways act through both the innate and adaptive immune systems, leading to various detrimental effects. These effects include the depletion of T cells, lymphopenia, apoptosis, impaired proliferation, decreased production of proinflammatory and anti-inflammatory factors, compromised secretion of antigen-presenting cells, and reduced functionality of BMCs. 282,393 Specifically, PD-1/PD-L not only play critical roles in tumor-infiltrating T lymphocytes but are also highly expressed in nonimmune cells, and the expression level is correlated with organ injury.^{27,394} In addition to inhibiting the number and functional activity of T cells, PD-1 can also bind to its receptor PD-1 to deliver a coinhibitory signal to negatively regulate T cell activation and mediate apoptosis. 395-398 In short, PD-1 binding to PD-L inhibited T cell activation and cytokine production, which may provide new guidance for sepsis defense. 399,400

Enhanced PD-L1 expression has been documented in various cell types in sepsis. 283,400-403 During sepsis, PD-L1 expression is increased on both stromal cells and DCs. 404,405 Splenic ECs obtained from sepsis patients exhibited higher levels of PD-L1 expression compared with the spleens obtained from patients who experienced brain death or needed emergent splenectomy due to trauma. 401,402,406-409 However, multiple other studies have demonstrated a higher expression of PD-1 in memory subsets of B cells and CD4⁺ T cells among individuals suffering from sepsis. 410,411 Experimental investigations conducted on septic mice have confirmed the reprogramming of monocytes or macrophages during sepsis, resulting in decreased expression of human leukocyte antigen-DR (HLA-DR) and the release of proinflammatory cytokines. This is then followed by a decline in antigen presentation capability and phagocytosis. 33,412-414 Elevated levels of PD-1/PD-L1 observed on the surface of peripheral blood monocytes or macrophages have been associated with reduced phagocytosis, a decrease in the secretion of proinflammatory cytokines such as IL-6 and TNF-a, and an increase in the release of anti-inflammatory cytokines such as IL-10.398,415 Furthermore, clinical trials have revealed a decrease in the expression of HLA-DR and CD86, which are costimulatory molecules, on monocytes/macrophages in sepsis patients. Conversely, the expression of PD-1 and PD-L1 is elevated, indicating a correlation with cellular dysfunction. These biomarkers may serve as indicators of monocyte or macrophage dysfunction in patients with sepsis.⁴⁰⁸ In vitro studies have shown that PD-1-mediated immune dysfunction of macrophages during sepsis occurs through the alteration

of macrophage migratory ability. However, the administration of an anti-PD-1/PD-L1 antibody can reverse these dysfunctions in sepsis patients. 416,417 Similarly, mice deficient in PD-1 have demonstrated an enhanced phagocytic capacity in macrophages. Kupffer cells, which act as macrophages in the liver and remove bacteria from the bloodstream, also exhibit increased cytokine production during sepsis. 403,417–420 The results presented here reinforce the concept that the abnormal activation of the PD-1/PD-L1 pathway is the main reason behind immunotherapy in individuals suffering from sepsis.

3.8 | JAK/STAT: JAK-STAT signaling blocks sepsis-induced immunosuppression

Janus kinases and signal transducers and activators of transcription (JAKs-STATs) are pivotal signaling components downstream of cytokine receptors.²⁸⁹ During the onset of sepsis, the two main causes of death are SIRS-induced multiple organ failure and sepsis-induced immune rejection responsible for late infections. 421,422 The signal conduction of these two lethal processes is involved in the regulation of JAK-STAT pathways. 423 SIRS is mainly mediated by STAT1 and STAT4, whereas compensatory antiinflammatory response syndrome is predominantly mediated by STAT3 and STAT6, with the involvement of JAK1-2 and tyrosine kinase (TYK) 2 for both cases. 424,425 Moreover, the JAK-STAT pathway also plays an essential role in cell proliferation and apoptosis, mainly mediating sepsistriggered emergency hematopoiesis and sepsis-induced organ dysfunction.^{290,426} Thus, targeting the JAK-STAT signaling pathway in sepsis may reduce sepsis-induced MOF and sepsis-induced immunosuppression. The recent successful development of JAK-STAT targeted therapy in oncology and hematology has provided new insights for the treatment of sepsis patients.

3.9 | SESN2: the protector of the immune system

Sestrin2 (SESN2) is a highly evolutionarily conserved protein associated with cellular responses to various stresses; it protects against oxidative stress, DNA damage, hypoxia, nutritional stress, ER stress, autophagy, metabolism and inflammation. SESN2 functions as an antioxidant, activating AMPK, and suppressing mTORC1 signaling. Initially, discovered as a downstream mediator of p53, it is induced by various detrimental environmental strains, including oxidative stress, ER stress, energy stress, and age- and obesity-related metabolic disorders. SESN2, regulates autophagy, ER stress, and inflammasome

activity under various conditions, exhibiting pleiotropic biological functions in cell homeostasis and metabolic homeostasis. 430 Particularly, when facing ER stress, SESN2 is upregulated and exerts a protective effect against ER stress-induced cell death. In situations of high stress, the activation of ER stress occurs as a result of the build-up of proteins that are unfolded or improperly folded within the lumen of the ER. This activation is then followed by the initiation of three branches within the pathway for ER stress. SESN2 expression is upregulated by PKR-like ER kinase (via eukaryotic translation initiation factor 2α and activating transcription factor 4), inositol-requiring enzyme 1 (via Xbox binding protein 1, TRAF2 and c-Jun N-terminal kinase [JNK]) and activating transcription factor 6. SESN2 functions as a regulatory mechanism to mitigate the cellular response to ER stress or ER stress-induced apoptosis.²⁴ The results showed that hypoxia and NO strongly induced the expression of SESN2 in hypoxia-induced factor- 1α , and the activation of SESN2 could prevent the peroxidation of peroxiredoxin, thus also acting as a cell protector. 287 SESN2 expression has been detected in various immune cells, including monocytes, macrophages, natural killer (NK) cells, and T lymphocytes.²⁸⁸ Overexpression of SESN2 reduced ER stress-related cell death, and SESN2 knockdown exacerbated the extent of ER stress, resulting in enhanced ER stress-mediated apoptosis.²⁴ SESN2 expression may exert beneficial effects on immune cell function by activating AMPK, inhibiting mTORC1 signaling, inhibiting JNK pathway activation, reducing the degree of ER stress, activating autophagy, or attenuating the NLRP3 inflammasome. 288 Therefore, SESN2 has multiple regulatory effects and could be a promising therapeutic target and play a protective role in various inflammatory diseases, such as sepsis.

4 | SEPSIS-RELATED BIOLOGICS: PRECLINICAL STUDIES AND CLINICAL TRIALS

In recent years, a large number of emerging studies have provided increasing theoretical support for the process of pathological response to sepsis. Management interventions that are customary, such as the administration of fluids to restore proper hydration and the provision of vasopressors for hemodynamic support, have proven to be successful in the initial resuscitation of sepsis. These interventions bring about a noteworthy enhancement in the overall clinical results of patients. However, targeting the underlying causes that trigger disruptive manifestations remains unclear. Numerous clinical investigations have noted indications of immune cell activation and dysregulation of the host response in individuals experiencing severe

sepsis. These findings align with multiple well-known molecular pathways.²⁷ Therefore, the development of new agents targeting molecular mechanisms and targets provides a strong experimental basis for further exploration of the treatment of sepsis. Here, we review novel molecular modulators targeting the main regulatory mechanisms of sepsis as well as therapeutic targets in the case of fatal infection, as shown in Table 1. Table 1 mainly summarizes the target proteins that this part of the regulator regulates when it exerts its function and the regulatory mechanism involved. In addition, the dosage of the modulator used, the time of publication, etc., are summarized.

Several large phase II and III trials are currently underway (Table 2). Advances in supportive care have significantly improved the clinical outcomes of patients with sepsis, and new clinical trials are seeking further advantages over infusion, hemodynamic, and sedation therapies.³¹ However, due to differences in the clinical definition of sepsis and the multifaceted manifestations observed during the clinical course, the inclusion criteria for patients with sepsis and the observed clinical end points varied over the course of the trial. 431 Some of the metrics of basic experiments do not seem to match the significance of clinical monitoring and cannot cover its impact on clinical efficacy, which is an inherent limitation of basic research. After the failure of therapeutic strategies for the inflammatory cascade in the early stage of sepsis development, the research focus of immunomodulatory therapy has shifted to enhancing the immune response in the late stage of immune paralysis. Recognizing that multiorgan failure is a major contributor to the clinical burden of systemic infections, early studies increasingly emphasized strategies to improve endothelial and epithelial-cell barrier function,⁶⁸ biofunctionalism, ^{68,432} and active inflammation resolution pathways.³²⁴ The use of cell therapy, such as allogeneic stromal cells or mesenchymal stem cells, shows promise as a treatment option. These cells possess powerful functions in immunomodulation, antimicrobial activity, bioenergetics, and enhancement of barriers (Figure 4).

Recently, encouraging results have been observed in clinical randomized controlled trials (Table 2). Until then, monitoring a large number of targets and biomarkers related to innate immune responses, receptor expression, vascular barrier integrity, and tissue/organ damage can provide circumstantial evidence informing the extent of host response dysregulation, which is closely linked to the state of immune homeostasis associated with the clinical course and is critical for elucidating the pathophysiology of sepsis.^{3,433} Therapeutic strategies aiming to remove and inhibit the respective molecular stimuli can be guided by biomarkers related to the innate immune response, including DAMPs, PAMPs, chemokines, and cytokines (Figure 4). These methods could potentially have a

TABLE 1 Potential inhibitors and their impact on sepsis in preclinical studies.

Therapeutic molecules	Target	Targeted mechanism	Usage	Model	Publication time	Sources
Glycyrrhizin	HMGB1	Inhibits HMGB1 release and activity	In vitro: $5-50 \mu M$ In vivo: 10-50 mg/kg	In vitro: BMDM, THP1, NR8383 In vivo: endotoxemia, CLP (mouse or rat)	2020, 2016, 2017	284,309,434
MCC950	NLRP3	Attenuates NLRP3 activation and inhibits HMGB1 release	In vitro: $0.1-10 \mu M$ In vivo: $10-50 \text{ mg/kg}$	In vitro: BMDM, HMDM In vivo: CLP (mouse or rat)	2018, 2016, 2019, 2020	284,310,435,436
LDK-378	ALK	Inhibits ALK-STING pathway and NLRP3 inflammasome activation	In vitro: 1–10 μ M In vivo: 20 mg/kg	In vitro: BMDM; THP1; RAW264.7; J774A.1 In vivo: endotoxemia, CLP (mouse)	2017,2018	437,438
Ac-FLTD-CMK	CASP1/4/5/11	Inhibits GSDMD cleavage by caspases-1, -4, -5, and -11	In vitro: $10 \mu\text{M}$	In vitro: BMDM	2018	439
Z-IETD-FMK	CASP8	Inhibits CASP8 to decrease GSDMD-N production	In vitro: $10 \mu\mathrm{M}$	In vitro: BMDM	2020	303
U73122	PLCG1	Inhibits GSDMD-N to relieve pyroptosis	In vitro: 10 μM In vivo: 30 mg/kg	In vitro: BMDM In vivo: CLP (mouse)	2020, 2018	385,303
H-151	STING1	Covalently binds to STING at the transmembrane cysteine residue site 91 to inhibit STING1	In vitro: $2 \mu M$ In vivo: 750 nM/mice	In vitro: primary human or mouse macrophages In vivo: Trex1 ^{-/-} mice	2020, 2018	280,440
YQ128	NLRP3	Inhibits the NLRP3 inflammasome and brain penetration	In vitro: $10-100 \mu M$ In vivo: 10-20 mg/kg	In vitro: BMDM, J774A.1 In vivo: endotoxemia (mouse)	2019	441
FPS-ZM1	AGER	A high-affinity AGER-specific inhibitor	In vitro: 0.1 – $1 \mu M$ In vivo: 10 mg/kg, $75 \mu \text{g/day}$	In vitro: BMDM In vivo: endotoxemia, Acinetobacter baumannii infection (mouse)	2019, 2017	442,443
Zileuton	ALOX5	Inhibits ALOX5 to relieve lipid peroxidation	In vitro: 5 μM In vivo: 30 mg/kg	In vitro: BMDM In vivo: endotoxemia (mouse)	2019	442
Disulfiram	GSDMD	Inhibits pyroptosis by blocking gasdermin D pore formation	In vitro: 1–30 μ M In vivo: 15–50 mg/kg	In vitro: BMDM, THP1 In vivo: endotoxemia (mouse)	2020	444
TUDCA	ER stress	F3 release was rescued by inhibiting ER stress	In vitro: $50 \mu M$ In vivo: 200 mg/kg	In vitro: THP1 In vivo: CLP (mouse)	2020	303
4PBA	ER stress	F3 release was rescued by inhibiting ER stress	In vitro: 1 mM In vivo: 20 mg/kg	In vitro: THP1 In vivo: CLP (mouse)	2020	303,445
Ruxolitinib	JAK1, JAK2	Reduce NO production by interfering with the NF-κB pathway	In vivo: 0.67 mg/kg	LPS-induced sepsis model	2020	445

(Continues)

TABLE 1 (Continued)

Therapeutic molecules	Target	Targeted mechanism	Usage	Model	Publication time	Sources
Tofacitinib	JAK1, JAK2, JAK3	Restrain differentiation of T-cells by inhibiting JAK–STAT pathway	In vivo: 15 mg/kg/day	Staphylococcus aureus-induced sepsis	2020	446
STC3141	NETs	Neutralization of extracellular histones and NETs to reverse organ damage caused by excessive immune responses	In vivo: 100 mg/kg In vivo: 50 mg/kg	LPS-induced acute respiratory distress syndrome/CLP (rat)	2023	

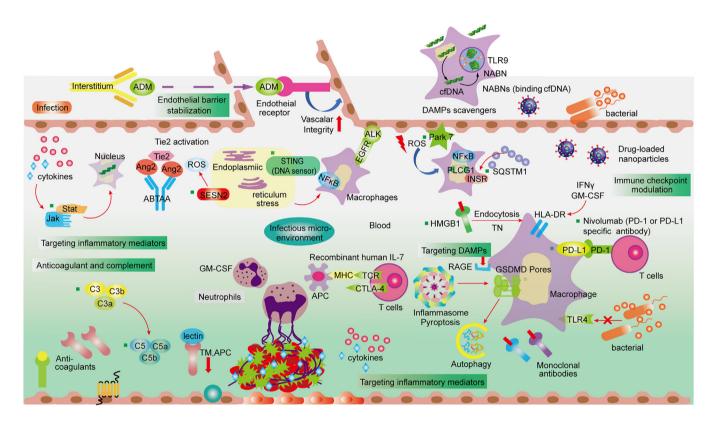


FIGURE 4 Potential therapeutic targets in sepsis intervention. Concerning the signaling pathways aimed at, the proposed therapeutic target for sepsis primarily encompasses the following pathway categories: (1) targeting DAMPs (including host cell stress), SESN2, and Park 7; (2) targeting inflammatory mediators (anti-inflammatory), JAK–STAT, HMGB1, GSDMD, and SQSTM1; (3) immune checkpoint modulation, PD-1/PD-L1; (4) restoration of complement and anticoagulant properties, C3a/C5a; (5) endothelial barrier stabilization, Ang/Tie2; (6) targeting the endoplasmic reticulum, STING. The discovery of therapeutic targets is a promising new strategy that has a wide range of therapeutic effects in experimental studies of sepsis.

positive impact on promoting the early reversal of harmful occurrences. By employing alternative novel biological substances (nano-sized particles filled with medications), the ongoing surveillance of immune cell activity levels can be utilized to detect individuals experiencing immune paralysis or abnormal proinflammatory host reactions. These measures provide the basis for future target-specific guided interventions and personalized treatment.

Sepsis causes numerous immune response defects, leading to immune suppression and increased susceptibility to

infection and death. In recent decades, great strides have been made in comprehending the mechanisms underlying the emergence of diverse pathogeneses associated with sepsis. Regrettably, these discoveries have not yet translated into efficacious therapies for bacterial sepsis. However, the mouse model of sepsis used, including injection of endotoxin, colonic ligation and perforation, and exogenous injection of pathogenic bacteria, does not fully mimic septic patients. Almost invariably, these models have been studied with the use of young, healthy, syngeneic

TABLE 2 Novel therapeutic molecules and their clinical impact on sepsis in clinical trials.

	4		•					
Therapeutic molecules	Mechanism	Population	Publication Study phase time and design	Study phase and design	Usage	Conclusion	Comment	Sources
Adrecizumab	ADM bounds to a non-neutralizing antibody, interacts with receptors on endothelial cells and reduces vascular leakage and tissue oedema	300	2019	Phase II RCT 2 or 4 mg.	2 or 4 mg/kg	Adrecizumab represents an effective approach to the treatment of fatal syndrome	The patients with concentration of circulating bio-ADM levels >70 pg/mL are recruited	
Human recombinant alkaline phosphatase	The inflammatory response was attenuated as a result of alkaline phosphatase dephosphorylating endotoxin and adenosine triphosphate	301	2018	RCT	1.6 mg/kg	In patients with severe sepsis-associated All patients were men acute kidney injury, the administration of recombinant human alkaline phosphatase did not result in improvement in renal function during the initial week of treatment	All patients were men	
Recombinant human IL-7 (CYT107)	It reverses the dramatic loss of CD4 ⁺ and CD8 ⁺ immune effector cells	27	2018	RCT	10 µg/kg	CYTIO7 is a promising approach for treating sepsis by restoring adaptive immunity. These immune-based therapies have the potential to provide comprehensive protection against various pathogens, with a specific focus on multidrug-resistant bacteria in immunocompromised patients	1	
Nivolumab	The expression of PD-1 and PD-L1 is increased in patients with sepsis, which is associated with a decrease in T cells. Nivolumab reverses T cell exhaustion by blocking PD-L1 to restore immune cell function, thereby improving immunosuppression	13	2020	Phase I/II study	480 or 960 mg	Over time, 480 mg and 960mg of nivolumab appeared to improve sepsis-induced immune system injury	Japanese patients with immunosuppressive sepsis	452
								(Continues)

_	
ŕ	ż
ď	í
-	ż
-	4
٠,=	į
7	4
>	٦
ځ	₹
_	,
0	1
7	1
	1
<u></u>	1
T T	1

Sources				456,457		
Comment	Patients with sepsis-associated coagulopathy	Patients ages ≥12 years with a surgical confirmation of NSTI, and organ dysfunction (mSOFA score≥3)		Patients with sepsis, organ dysfunction, absolute lymphocyte count ≤1100 cells/µL		
Conclusion	The 28-day all-cause mortality was not significantly reduced when administering a human recombinant thrombomodulin, as compared with the placebo	In patients with severe NSTIs, early administration of reltecimod significantly improved the primary composite endpoint in the PP population	The 96-h vitamin C infusion did not significantly improve organ dysfunction scores or change markers of inflammation and vascular damage compared with the placebo group	BMS-936559 exhibited good tolerability, no drug-induced cytokine release, and immune status was restored within 28 days at high doses		
Usage	0.06 mg/ kg/d	0.5 mg/kg	50 mg/kg	10-900 mg		
Study phase and design	RCT	Phase III RCT	RCT	Phase 1b	Phase II	Phase I
Publication Study phase time and design	2019	2020	2019	2019		
Population	008	290	167	24		
Mechanism	rhsTM binds circulating thrombin molecules and converts protein C to APC as an activation complex. And, rhsTM inhibits inflammation and organ damage caused by damage-associated molecular patterns	Interaction of superantigens with dimers of the costimulatory receptor CD28 expressed on T cells mediates T helper type 1 cytokine responses	1	Restore or enhance T-cell function		
Therapeutic molecules	Recombinant human thrombomod- ulin (ART-123)	Reltecimod, AB103	Ascorbic acid	BMS-936559	STC3141	APAD

ZHANG ET AL.

animals, often without sophisticated monitoring, and with little or no supportive therapy (antibiotics, respiratory support including oxygen, fluid and vasopressor resuscitation, and renal replacement therapy). In addition to the significant differences in the composition and treatment of sepsis patients, the important species differences also lay an enormous potential for the inconsistency between preclinical and clinical trials in the development of drug release. To that end, many effective molecules (small molecule modulators and biologics), targets and related interventions for sepsis are being studied as hotspots (Figure 4). It will be imperative to further define the roles they play in experimental sepsis to facilitate the development of novel therapeutic approaches for clinical sepsis. Given that these inhibitors and targets have demonstrated the ability to impede disease progression in laboratory or animal models of sepsis, the anticipation for their assessment in human trials is pressing.

5 | CONCLUSIONS AND PERSPECTIVES

The development of sepsis is characterized by hyperinflammation, immunosuppression, EC injury, DIC, multiple organ failure, and limited treatment options, which continues to be a formidable challenge for basic and clinical research. As of now, the attempts made to target only one mediator or target, such as IL-1\beta or APC, have proven unsuccessful in diminishing mortality in sepsis patients. This distressing outcome warns us that the evolution of sepsis is a heterogeneous development process with multiple interfering factors and multiple regulatory pathways interacting. However, in recent years, basic research using cell experiments and mouse models has revealed that many key molecules exist in the complex regulatory network of organismal systems, and there are regulatory and regulated relationships between these key molecules, which play a decisive role in the development of the disease, thereby enhancing our comprehension of the essence, qualities, and outcomes of the sepsis response. Nonetheless, the significant genetic variations between humans and mice considerably restrict their utilization in sepsis patients. Despite this complexity, our objective is to enhance the understanding of the initial signaling, intermediate associations, final effectors, and feedback loops involved in sepsis. This will enable the identification of novel therapeutic targets and the development of treatments that can significantly benefit patients suffering from life-threatening infections.

As noted previously, sepsis presents itself as a diverse syndrome that exhibits noteworthy distinctions in pathophysiology, clinical presentations, and consequences. The primary emphasis in many clinical trials examining inno-

vative sepsis treatments has been on the preimmune status and pathophysiology exclusive to infection. These approaches generate high signal-to-noise ratios that do not result in favorable clinical outcomes. Past responses to sepsis-induced homeostasis imbalances, such as complement, coagulation, mitochondrial damage, and endothelial dysfunction, have been improved. However, the history of clinical trials in sepsis has demonstrated that improvements in a single or a few physiologic markers in septic patients do not reverse clinical patient mortality, and the results do not necessarily translate into the clinical setting. Nevertheless, the primary hurdle lies in the fact that nearly every immune mediator assumes varied and interconnected functions in the pathophysiology of sepsis. This implies that the interception of these mediators can potentially disrupt the balance of homeostasis and accentuate disease-amplifying networks. As a result, most clinical trials generally do not improve overall outcomes. A number of explanations have been proposed, including underdosing of interventional agents due to rapid clearance or rapid metabolic inactivation. The discovery of novel therapeutic targets targeting vital signaling pathways offers new hope for the treatment of sepsis, but a thoughtful preclinical approach will be essential. Effective interventions can be found to limit organ failure as quickly and effectively as possible in the treatment of sepsis. The complexity of sepsis-associated organ dysfunction poses a challenge in achieving this goal, but the possibility of recovery underscores the need for identifying and implementing efficient measures.

In the past few years, studies examining the imbalance of inflammation, dysfunction in the immune system, damage to mitochondria, dysfunction in blood clotting, abnormalities in the network between the nervous and endocrine systems, stress in the ER, autophagy, and other mechanisms have greatly enhanced our understanding of how sepsis develops and uncovered potential targets for treatment as well as markers for prognosis. These findings hold promise for the prevention, diagnosis, and management of sepsis. Given the complexity of host responses and the diversity of pathophysiological pathways involved in sepsis patients, current "one-target" and "one-size-fits-all" approaches are unlikely to succeed. Since sepsis is a systemic disease involving multiple organs, patients currently die upon admission. However, clinical guidelines have implemented efficient management strategies to promptly control the spread of the source and ensure adequate cell perfusion through early and appropriate administration of antimicrobial therapy. Despite being costly and time consuming, this approach exhibits limited sensitivity and selectivity due to its inherent limitations. Therefore, there is an urgent need to develop alternatives to sepsis diagnostic systems. Currently, there is an absence of both a



United States Food and Drug Administration-approved medication and efficient, speedy tools for diagnosing sepsis in clinical settings. The direction that sepsis diagnosis is heading toward involves early detection and risk evaluation, based on the clinical traits and biomarkers of patients. Furthermore, more targeted investigations are needed to enhance our comprehension of the fundamental pathophysiology underlying sepsis mortality. Categorizing sepsis into distinct groups in the future might aid in identifying superior and more potent treatments, surpassing the efficacy of current approaches employed for sepsis management.

AUTHOR CONTRIBUTION

Wendan Zhang wrote the paper. Honghong Jiang made the figures. Gaosong Wu, Pengli Huang, and Haonan Wang made the tables. Huazhang An revised the article. Weidong Zhang performed the supervision, project administration, and funding acquisition. Sanhong Liu contributed with the project administration, review, and editing. All authors have read and approved the final manuscript.

ACKNOWLEDGMENTS

This work was funded by National Natural Science Foundation of China (No. 82141203, 81772798), Shanghai Municipal Science and Technology Major Project (ZD2021CY001), Three-year Action Plan for Shanghai TCM Development and Inheritance Program [ZY (2021-2023)-0401], and Innovation Team and Talents Cultivation Program of National Administration of Traditional Chinese Medicine (ZYYCXTDD-202004).

CONFLICT OF INTEREST STATEMENT

The authors declare no conflicts of interest.

ETHICS STATEMENT

Not applicable.

DATA AVAILABILITY STATEMENT

Not applicable.

ORCID

Sanhong Liu https://orcid.org/0000-0002-8695-0343

REFERENCES

- Tang D, Wang H, Billiar TR, Kroemer G, Kang R. Emerging mechanisms of immunocoagulation in sepsis and septic shock. *Trends Immunol.* 2021;42(6):508-522.
- 2. Kumar V. Targeting macrophage immunometabolism: dawn in the darkness of sepsis. *Int Immunopharmacol*. 2018;58(April):173-185.

- 3. Venet F, Monneret G. Advances in the understanding and treatment of sepsis-induced immunosuppression. *Nat Rev Nephrol.* 2018;14(2):121-137.
- Gotts JE, Matthay MA. Sepsis: pathophysiology and clinical management. BMJ (Online). 2016;353:1585.
- 5. Zhou P, Yang XL, Wang XG, et al. Erratum: Addendum: a pneumonia outbreak associated with a new coronavirus of probable bat origin (Nature (1988) 81 11 (18-19)). *Nature*. 2020;588(7836):E6.
- Lever A, Mackenzie I. Sepsis: definition, epidemiology, and diagnosis. Br Med J. 2007;335(7625):879-883.
- Rello J, Valenzuela-Sánchez F, Ruiz-Rodriguez M, Moyano S. Sepsis: a review of advances in management. *Adv Ther*. 2017;34(11):2393-2411.
- 8. Tiru B, DiNino EK, Orenstein A, et al. The economic and humanistic burden of severe sepsis. *Pharmacoeconomics*. 2015;33(9):925-937.
- 9. Zhu Z, Chambers S, Zeng Y, Bhatia M. Gases in sepsis: novel mediators and therapeutic targets. *Int J Mol Sci.* 2022;23(7):3669.
- Rudd KE, Johnson SC, Agesa KM, et al. Global, regional, and national sepsis incidence and mortality, 1990–2017: analysis for the Global Burden of Disease Study. *Lancet North Am Ed.* 2020;395(10219):200-211.
- Martischang R, Pires D, Masson-Roy S, Saito H, Pittet D. Promoting and sustaining a historical and global effort to prevent sepsis: The 2018 World Health Organization SAVE LIVES: Clean Your Hands campaign. *Crit Care*. 2018;22(1):7-9.
- 12. Husabø G, Nilsen RM, Flaatten H, et al. Early diagnosis of sepsis in emergency departments, time to treatment, and association with mortality: an observational study. *PLoS One*. 2020;15(1):1-15.
- Mao K, Chen S, Chen M, et al. Nitric oxide suppresses NLRP3 inflammasome activation and protects against LPS-induced septic shock. *Cell Res.* 2013;23(2):201-212.
- Lelubre C, Vincent JL. Mechanisms and treatment of organ failure in sepsis. Nat Rev Nephrol. 2018;14(7):417-427.
- Rosen DA, Seki SM, Fernández-Castañeda A, et al. Modulation of the sigma-1 receptor-IRE1 pathway is beneficial in preclinical models of inflammation and sepsis. *Sci Transl Med*. 2019;11(478):eaau5266.
- Rocheteau P, Chatre L, Briand D, et al. Sepsis induces long-term metabolic and mitochondrial muscle stem cell dysfunction amenable by mesenchymal stem cell therapy. *Nat Commun*. 2015;6(May):1-12.
- 17. Zhang YY, Ning BT. Signaling pathways and intervention therapies in sepsis. Signal Transduct Target Ther. 2021;6(1):407.
- Schneidewind L. Reducing mortality in adults with sepsis, severe sepsis, or septic shock: effectiveness and safety of procalcitonin. *Urologe* . 2021;60(5):624-627.
- McBride MA, Patil TK, Bohannon JK, Hernandez A, Sherwood ER, Patil NK. Immune checkpoints: novel therapeutic targets to attenuate sepsis-induced immunosuppression. *Front Immunol*. 2021;11(February):1-14.
- Pan B, Wang X, Kojima S, et al. The fifth epidermal growth factor like region of thrombomodulin alleviates LPS-induced sepsis through interacting with GPR15. *Thromb Haemost*. 2017;117(3):570-579.



- Moon JS, Lee S, Park MA, et al. UCP2-induced fatty acid synthase promotes NLRP3 inflammasome activation during sepsis. *J Clin Invest*. 2015;125(2):665-680.
- Bergmann CB, Beckmann N, Salyer CE, Hanschen M, Crisologo PA, Caldwell CC. Potential targets to mitigate trauma- or sepsis-induced immune suppression. Front Immunol. 2021;12(February):1-19.
- Blasco RB, Patrucco E, Mota I, Tai WT, Chiarle R. Comment on "ALK is a therapeutic target for lethal sepsis". Sci Transl Med. 2018;10(471):10-13.
- 24. Luo L, Wu J, Qiao L, Lu G, Li J, Li D. Sestrin 2 attenuates sepsis-associated encephalopathy through the promotion of autophagy in hippocampal neurons. *J Cell Mol Med*. 2020;24(12):6634-6643.
- Iba T, Levy JH. Sepsis-induced coagulopathy and disseminated intravascular coagulation. *Anesthesiology*. 2020;(5):1238-1245.
- Deng C, Zhao L, Yang Z, et al. Targeting HMGB1 for the treatment of sepsis and sepsis-induced organ injury. Acta Pharmacol Sin. 2022;43(3):520-528.
- Patil NK, Bohannon JK, Sherwood ER. Immunotherapy: A promising approach to reverse sepsis-induced immunosuppression. *Pharmacol Res.* 2016;111:688-702.
- Sureshbabu A, Patino E, Ma KC, et al. RIPK3 promotes sepsisinduced acute kidney injury via mitochondrial dysfunction. *JCI Insight*. 2018;3(11):1-17.
- Vincent JL, Francois B, Zabolotskikh I, et al. Effect of a recombinant human soluble thrombomodulin on mortality in patients with sepsis-associated coagulopathy: the SCARLET randomized clinical trial. *JAMA*. 2019;321(20):1993-2002.
- Iskander KN, Osuchowski MF, Stearns-Kurosawa DJ, et al. Sepsis: multiple abnormalities, heteroge-neous responses, and evolving understanding. *Physiol Rev.* 2013;93:1247-1288.
- 31. Martín-Fernández M, Heredia-Rodríguez M, González-Jiménez I, et al. Hyperoxemia in postsurgical sepsis/septic shock patients is associated with reduced mortality. *Crit Care*. 2022;26(1):1-9.
- 32. Cheng Y, Marion TN, Cao X, Wang W, Cao Y. Park 7: A novel therapeutic target for macrophages in sepsis-induced immunosuppression. *Front Immunol.* 2018;9(NOV):1-8.
- 33. van der Poll T, van de Veerdonk FL, Scicluna BP, Netea MG. The immunopathology of sepsis and potential therapeutic targets. *Nat Rev Immunol*. 2017;17(7):407-420.
- Cecconi M, Evans L, Levy M, Rhodes A. Sepsis and septic shock. Lancet North Am Ed. 2018;392(10141):75-87.
- Deng W, Zhu S, Zeng L, et al. The Circadian clock controls immune checkpoint pathway in sepsis. *Cell Rep.* 2018;24(2):366-378
- 36. Schneck E, Edinger F, Hecker M, et al. Blood levels of freecirculating mitochondrial dna in septic shock and postsurgical systemic inflammation and its influence on coagulation: a secondary analysis of a prospective observational study. *J Clin Med*. 2020;9(7):1-14.
- 37. De Backer D, Cortes DO, Donadello K, Vincent JL. Pathophysiology of microcirculatory dysfunction and the pathogenesis of septic shock. *Virulence*. 2014;5(1):73-79.
- 38. Kaukonen KM, Bailey M, Pilcher D, Cooper DJ, Bellomo R. Systemic inflammatory response syndrome criteria in defining severe sepsis. *N Engl J Med*. 2015;372(17):1629-1638.

- 39. Martinon F, Mayor A, Tschopp J. The inflammasomes: guardians of the body. *Annu Rev Immunol*. 2009;27:229-265.
- Sharshar T, Annane D, De La Grandmaison GL, Brouland JP, Hopkinson NS, Gray F. The neuropathology of septic shock. *Brain Pathol.* 2004;14(1):21-33.
- 41. Chauhan D, Vande Walle L, Lamkanfi M. Therapeutic modulation of inflammasome pathways. *Immunol Rev.* 2020;297(1):123-138.
- Corcoran SE, Halai R, Cooper MA. Pharmacological inhibition of the nod-like receptor family pyrin domain containing 3 inflammasome with mcc950. *Pharmacol Rev.* 2021;73(3):968-1000.
- 43. Huang M, Cai S, Su J. The pathogenesis of sepsis and potential therapeutic targets. *Int J Mol Sci.* 2019;20(21):5376.
- Baker PJ, Boucher D, Bierschenk D, et al. NLRP3 inflammasome activation downstream of cytoplasmic LPS recognition by both caspase-4 and caspase-5. *Eur J Immunol*. 2015;45(10):2918-2926.
- Li Z, Liu W, Fu J, et al. Shigella evades pyroptosis by arginine ADP-riboxanation of caspase-11. *Nature*. 2021;599(7884):290-295
- Broz P, Newton K, Lamkanfi M, Mariathasan S, Dixit VM, Monack DM. Redundant roles for inflammasome receptors NLRP3 and NLRC4 in host defense against Salmonella. *J Exp Med*. 2010;207(8):1745-1755.
- Xing K, Murthy S, Liles WC, Singh JM. Clinical utility of biomarkers of endothelial activation in sepsis-a systematic review. *Crit Care*. 2012;16(1):R7.
- Moser J, Heeringa P, Jongman RM, et al. Intracellular RIG-I signaling regulates TLR4-independent endothelial inflammatory responses to endotoxin. *J Immunol*. 2016;196(11):4681-4691.
- Yang N, Luna JM, Dai P, Wang Y, Rice CM, Deng L. Lung type II alveolar epithelial cells collaborate with CCR2+ inflammatory monocytes in host defense against poxvirus infection. *Nat Commun*. 2022;13(1):1671.
- Thoresen D, Wang W, Galls D, Guo R, Xu L, Pyle AM. The molecular mechanism of RIG-I activation and signaling. *Immunol Rev.* 2021;304(1):154-168.
- Shao S, Chen C, Shi G, et al. Therapeutic potential of the target on NLRP3 inflammasome in multiple sclerosis. *Pharmacol Ther*. 2021;227:107880.
- 52. Boyer MJ, Kimura Y, Akiyama T, et al. Endothelial cell-derived extracellular vesicles alter vascular smooth muscle cell phenotype through high-mobility group box proteins. *J Extracell Vesicles*. 2020;9(1):1-19.
- 53. Haratani K, Yonesaka K, Takamura S, et al. U3-1402 sensitizes HER3-expressing tumors to PD-1 blockade by immune activation. *J Clin Invest*. 2020;130(1):374-388.
- Leisman DE, Mehta A, Thompson BT, et al. Alveolar, endothelial, and organ injury marker dynamics in severe COVID-19.
 Am J Respir Crit Care Med. 2022;205(5):507-519.
- 55. Tang Y, Wang X, Li Z, et al. Heparin prevents caspase-11-dependent septic lethality independent of anticoagulant properties. *Immunity*. 2021;54(3):454-467. e6.
- Lu A, Magupalli VG, Ruan J, et al. Unified polymerization mechanism for the assembly of ASC-dependent inflammasomes. *Cell*. 2014;156(6):1193-1206.
- 57. Tsao CM, Ho ST, Wu CC. Coagulation abnormalities in sepsis. *Acta Anaesthesiol Taiwan*. 2015;53(1):16-22.



- 58. Yuan C, Wu M, Xiao Q, et al. Blocking Msrl by berberine alkaloids inhibits caspase-11-dependent coagulation in bacterial sepsis. *Signal Transduct Target Ther*. 2021;6(1):2020-2022.
- Iba T, Ito T, Maruyama I, et al. Potential diagnostic markers for disseminated intravascular coagulation of sepsis. *Blood Rev.* 2016;30(2):149-155.
- Takayama W, Endo A, Morishita K, Otomo Y. Dielectric blood coagulometry for the early detection of sepsis-induced disseminated intravascular coagulation: a prospective observational study. Crit Care Med. 2022;50(1):E31-E39.
- Gando S, Saitoh D, Ishikura H, et al. A randomized, controlled, multicenter trial of the effects of antithrombin on disseminated intravascular coagulation in patients with sepsis. *Crit Care*. 2013;17(6):R297.
- McDonald B, Davis RP, Kim SJ, et al. Platelets and neutrophil extracellular traps collaborate to promote intravascular coagulation during sepsis in mice. *Blood*. 2017;129(10):1357-1367.
- Kohli S, Shahzad K, Jouppila A, Holthöfer H, Isermann B, Lassila R. Thrombosis and inflammation—a dynamic interplay and the role of glycosaminoglycans and activated protein C. Front Cardiovasc Med. 2022;9(March):1-13.
- 64. Warren BL, Eid A, Singer P, et al. High-dose antithrombin III in severe sepsis. *JAMA*. 2001;286(15):1869.
- 65. Gando S, Saitoh D, Ogura H, et al. A multicenter, prospective validation study of the Japanese association for acute medicine disseminated intravascular coagulation scoring system in patients with severe sepsis. Crit Care. 2013;17(3):R111.
- Creasey AA, Reinhart K. Tissue factor pathway inhibitor activity in severe sepsis. Crit Care Med. 2001;29(7 SUPPL):S126-S129.
- Delabranche X, Boisramé-Helms J, Asfar P, et al. Microparticles are new biomarkers of septic shock-induced disseminated intravascular coagulopathy. *Intens Care Med.* 2013;39(10):1695-1703.
- Schouten M, Wiersinga WJ, Levi M, van der Poll T. Inflammation, endothelium, and coagulation in sepsis. *J Leukoc Biol.* 2008;83(3):536-545.
- 69. Karasu E, Nilsson B, Köhl J, Lambris JD, Huber-Lang M. Targeting complement pathways in polytrauma- and sepsis-induced multiple-organ dysfunction. *Front Immunol*. 2019;10(Mar):1-14.
- Lupu F, Keshari RS, Lambris JD, Mark Coggeshall K. Crosstalk between the coagulation and complement systems in sepsis. *Thromb Res.* 2014;133(Suppl 1):S28-S31.
- Silasi-Mansat R, Zhu H, Popescu NI, et al. Complement inhibition decreases the procoagulant response and confers organ protection in a baboon model of Escherichia coli sepsis. *Blood*. 2010;116(6):1002-1010.
- Sprong T, Brandtzaeg P, Fung M, et al. Inhibition of C5ainduced inflammation with preserved C5b-9-mediated bactericidal activity in a human whole blood model of meningococcal sepsis. *Blood*. 2003;102(10):3702-3710.
- Napier BA, Brubaker SW, Sweeney TE, et al. Complement pathway amplifies caspase-11-dependent cell death and endotoxin-induced sepsis severity. *J Exp Med.* 2016;213(11):2365-2382.
- Zhao L, Ohtaki Y, Yamaguchi K, et al. LPS-induced platelet response and rapid shock in mice: contribution of O-antigen region of LPS and involvement of the lectin pathway of the complement system. *Blood*. 2002;100(9):3233-3239.

- Ward PA. The dark side of C5a in sepsis. Nat Rev Immunol. 2004;4(2):133-142.
- Nahm MH, Yu J, Calix JJ, Ganaie F. Ficolin-2 lectin complement pathway mediates capsule-specific innate immunity against invasive pneumococcal disease. Front Immunol. 2022;13(March):1-10.
- Iwasaki A, Medzhitov R. Control of adaptive immunity by the innate immune system. *Nat Immunol*. 2015;16(4):343-353.
- Keshari RS, Popescu NI, Silasi R, et al. Complement C5 inhibition protects against hemolytic anemia and acute kidney injury in anthrax peptidoglycan-induced sepsis in baboons. *Proc Natl Acad Sci USA*. 2021;118(37):e2104347118.
- Keshari RS, Silasi R, Popescu NI, et al. Inhibition of complement C5 protects against organ failure and reduces mortality in a baboon model of Escherichia coli sepsis. *Proc Natl Acad Sci USA*. 2017;114(31):E6390-E6399.
- 80. Hoehlig K, Maasch C, Shushakova N, et al. A novel C5aneutralizing mirror-image (l-)aptamer prevents organ failure and improves survival in experimental sepsis. *Mol Ther*. 2013;21(12):2236-2246.
- 81. Riedemann NC, Guo RF, Hollmann TJ, et al. Regulatory role of C5a in LPS-induced IL-6 production by neutrophils during sepsis 1. *FASEB J.* 2004;18(2):370-372.
- 82. Gilmore AC, Zhang Y, Cook HT, et al. Complement activity is regulated in C3 glomerulopathy by IgG–factor H fusion proteins with and without properdin targeting domains. *Kidney Int*. 2021;99(2):396-404.
- Bermejo-Martin JF, Andaluz-Ojeda D, Almansa R, et al. Defining immunological dysfunction in sepsis: a requisite tool for precision medicine. *J Infect*. 2016;72(5):525-536.
- Riedemann NC, Guo RF, Bernacki KD, et al. Regulation by C5a of neutrophil activation during sepsis. *Immunity*. 2003;19(2):193-202.
- 85. Huber-Lang M, Sarma JV, Zetoune FS, et al. Generation of C5a in the absence of C3: a new complement activation pathway. *Nat Med*. 2006;12(6):682-687.
- Jackson WD, Gulino A, Fossati-Jimack L, et al. C3 drives inflammatory skin carcinogenesis independently of C5. *J Invest Dermatol.* 2021;141(2):404-414.e6.
- 87. Chan KW, Yu KY, Yiu WH, et al. Potential therapeutic targets of rehmannia formulations on diabetic nephropathy: a comparative network pharmacology analysis. *Front Pharmacol*. 2022;13(March):1-14.
- Zhen Y, Yuan Z, Zhang J, et al. Flubendazole induces mitochondrial dysfunction and DRP1-mediated mitophagy by targeting EVA1A in breast cancer. *Cell Death Dis.* 2022;13(4):1-14.
- Kabir MF, Karami AL, Cruz-Acuña R, et al. Single cell transcriptomic analysis reveals cellular diversity of murine esophageal epithelium. *Nat Commun*. 2022;13(1):2167.
- Carty M, Kearney J, Shanahan KA, et al. Cell survival and cytokine release after inflammasome activation is regulated by the Toll-IL-1R protein SARM. *Immunity*. 2019;50(6):1412-1424. e6.
- 91. Ridker PM, MacFadyen JG, Thuren T, et al. Effect of interleukin-1β inhibition with canakinumab on incident lung cancer in patients with atherosclerosis: exploratory results from a randomised, double-blind, placebo-controlled trial. *Lancet North Am Ed.* 2017;390(10105):1833-1842.



- 92. Oláhová M, Yoon WH, Thompson K, et al. Biallelic mutations in ATP5F1D, which encodes a subunit of ATP synthase, cause a metabolic disorder. *Am J Hum Genet*. 2018;102(3):494-504.
- 93. Adam C, Paolini L, Gueguen N, et al. Acetoacetate protects macrophages from lactic acidosis-induced mitochondrial dysfunction by metabolic reprograming. *Nat Commun*. 2021;12(1):1-13.
- 94. Xia H, Scholtes C, Dufour CR, Ouellet C, Ghahremani M, Giguère V. Insulin action and resistance are dependent on a GSK3β-FBXW7-ERRα transcriptional axis. *Nat Commun*. 2022:13(1):2105.
- 95. Wu KKL, Long KK, Lin H, et al. The APPL1-Rab5 axis restricts NLRP3 inflammasome activation through early endosomal-dependent mitophagy in macrophages. *Nat Commun*. 2021;12(1):1-17.
- Yim DB, Lee DE, So Y, et al. Sustainable nanosheet antioxidants for sepsis therapy via scavenging intracellular reactive oxygen and nitrogen species. ACS Nano. 2020;14(8):10324-10336.
- Deo P, Chow SH, Han ML, et al. Mitochondrial dysfunction caused by outer membrane vesicles from Gram-negative bacteria activates intrinsic apoptosis and inflammation. *Nat Microbiol.* 2020;5(11):1418-1427.
- 98. Esteves AR, Munoz-Pinto MF, Nunes-Costa D, et al. Footprints of a microbial toxin from the gut microbiome to mesencephalic mitochondria. *Gut.* 2023;72(1):73-89. Published online 2021:gutjnl-2021-326023.
- 99. Giorgi C, Danese A, Missiroli S, Patergnani S, Pinton P. Calcium dynamics as a machine for decoding signals. *Trends Cell Biol*. 2018;28(4):258-273.
- 100. MacGarvey NC, Suliman HB, Bartz RR, et al. Activation of mitochondrial biogenesis by heme oxygenase-1-mediated NF-E2-related factor-2 induction rescues mice from lethal Staphylococcus aureus sepsis. Am J Respir Crit Care Med. 2012;185(8):851-861.
- 101. Bartz RR, Suliman HB, Fu P, et al. Staphylococcus aureus sepsis and mitochondrial accrual of the 8-oxoguanine DNA glycosylase DNA repair enzyme in mice. *Am J Respir Crit Care Med*. 2011;183(2):226-233.
- 102. Lourbopoulos AI, Mourouzis IS, Trikas AG, Tseti IK, Pantos CI. Effects of thyroid hormone on tissue hypoxia: relevance to sepsis therapy. *J Clin Med*. 2021;10(24):5855.
- Haden DW, Suliman HB, Carraway MS, et al. Mitochondrial biogenesis restores oxidative metabolism during Staphylococcus aureus sepsis. Am J Respir Crit Care Med. 2007;176(8):768-777.
- 104. Misheva M, Kotzamanis K, Davies LC, et al. Oxylipin metabolism is controlled by mitochondrial β -oxidation during bacterial inflammation. *Nat Commun*. 2022;13(1):1-20.
- Souza ACP, Yuen PST, Star RA. Microparticles: Markers and mediators of sepsis-induced microvascular dysfunction, immunosuppression, and AKI. Kidney Int. 2015;87(6):1100-1108.
- 106. Dear JW, Leelahavanichkul A, Aponte A, et al. Liver proteomics for therapeutic drug discovery: inhibition of the cyclophilin receptor CD147 attenuates sepsis-induced acute renal failure. Crit Care Med. 2007;35(10):2319-2328.
- 107. Yi Z, Wu Y, Zhang W, et al. Activator-mediated pyruvate kinase M2 activation contributes to endotoxin tolerance by promoting

- mitochondrial biogenesis. *Front Immunol.* 2021;11(January):1-14.
- Li Y, Feng YF, Liu XT, et al. Songorine promotes cardiac mitochondrial biogenesis via Nrf2 induction during sepsis. *Redox Biol.* 2021:38
- 109. Xia Y, Cao Y, Sun Y, et al. Calycosin alleviates sepsisinduced acute lung injury via the inhibition of mitochondrial ROS-mediated inflammasome activation. Front Pharmacol. 2021;12(October):1-14.
- Wang LX, Ren C, Yao RQ, et al. Sestrin2 protects against lethal sepsis by suppressing the pyroptosis of dendritic cells. *Cell Mol Life Sci.* 2021;78(24):8209-8227.
- zhang Z, Han N, Shen Y. S100A12 promotes inflammation and cell apoptosis in sepsis-induced ARDS via activation of NLRP3 inflammasome signaling. *Mol Immunol*. 2020;122(March):38-48.
- Thomas H. Sepsis: bile acids promote inflammation in cholestasis-associated sepsis. Nat Rev Gastroenterol Hepatol. 2017;14(6):324-325.
- Garcia-Irigoyen O, Moschetta A. A novel protective role for FXR against inflammasome activation and endotoxemia. *Cell Metab.* 2017;25(4):763-764.
- 114. Kim HM, Kim YM. HMGB1: LPS delivery vehicle for caspase-11-mediated pyroptosis. *Immunity*. 2018;49(4):582-584.
- 115. Russo AJ, Vasudevan SO, Méndez-Huergo SP, et al. Intracellular immune sensing promotes inflammation via gasdermin D-driven release of a lectin alarmin. *Nat Immunol*. 2021;22(2):154-165.
- 116. Ding X, Kambara H, Guo R, et al. Inflammasome-mediated GSDMD activation facilitates escape of Candida albicans from macrophages. *Nat Commun.* 2021;12(1):6699.
- 117. Angosto-Bazarra D, Alarcón-Vila C, Hurtado-Navarro L, Baños MC, Rivers-Auty J, Pelegrín P. Evolutionary analyses of the gasdermin family suggest conserved roles in infection response despite loss of pore-forming functionality. *BMC Biol*. 2022;20(1):1-18.
- 118. Xia S, Zhang Z, Magupalli VG, et al. Gasdermin D pore structure reveals preferential release of mature interleukin-1. *Nature*. 2021;593(7860):607-611.
- Liu L, Li H, Hu D, et al. Insights into N6-methyladenosine and programmed cell death in cancer. Mol Cancer. 2022;21(1):1-16.
- 120. Liu L, Sun B. Neutrophil pyroptosis: new perspectives on sepsis. *Cell Mol Life Sci.* 2019;76(11):2031-2042.
- Yang X, Cheng X, Tang Y, et al. Bacterial endotoxin activates the coagulation cascade through gasdermin D-dependent phosphatidylserine exposure. *Immunity*. 2019;51(6):983-996. e6.
- 122. Di A, Xiong S, Ye Z, et al. The TWIK2 potassium efflux channel in macrophages mediates NLRP3 inflammasome-induced inflammation. *Immunity*. 2018;49(1):56-65. e4.
- 123. Stunden HJ, Latz E. PKR stirs up inflammasomes. *Cell Res.* 2013;23(2):168-170.
- Tang SCW, Yiu WH. Innate immunity in diabetic kidney disease. Nat Rev Nephrol. 2020;16(4):206-222.
- 125. Suetomi T, Willeford A, Brand CS, et al. Inflammation and NLRP3 inflammasome activation initiated in response to pressure overload by Ca2+/calmodulin-dependent protein kinase II δ signaling in cardiomyocytes are essential for adverse cardiac remodeling. *Circulation*. 2018;138(22):2530-2544.



- Afonina IS, Zhong Z, Karin M, Beyaert R. Limiting inflammation—the negative regulation of NF-B and the NLRP3 inflammasome. *Nat Immunol*. 2017;18(8):861-869.
- 127. Yu X, Lan P, Hou X, et al. HBV inhibits LPS-induced NLRP3 inflammasome activation and IL-1 β production via suppressing the NF- κ B pathway and ROS production. *J Hepatol*. 2017;66(4):693-702.
- 128. Bruder-Nascimento T, Ferreira NS, Zanotto CZ, et al. NLRP3 inflammasome mediates aldosterone-induced vascular damage. *Circulation*. 2016;134(23):1866-1880.
- 129. Zhong Z, Liang S, Sanchez-Lopez E, et al. New mitochondrial DNA synthesis enables NLRP3 inflammasome activation. *Nature*. 2018;560(7717):198-203.
- 130. Xian H, Liu Y, Rundberg Nilsson A, et al. Metformin inhibition of mitochondrial ATP and DNA synthesis abrogates NLRP3 inflammasome activation and pulmonary inflammation. *Immunity*. 2021;54(7):1463-1477.e11.
- O'Neill LAJ. Cardiolipin and the Nlrp3 inflammasome. Cell Metab. 2013;18(5):610-612.
- 132. Muñoz-Planillo R, Kuffa P, Martínez-Colón G, Smith BL, Rajendiran TM, Núñez G. K+ efflux is the common trigger of NLRP3 inflammasome activation by bacterial toxins and particulate matter. *Immunity*. 2013;38(6):1142-1153.
- Compan V, Baroja-Mazo A, López-Castejón G, et al. Cell volume regulation modulates NLRP3 inflammasome activation. *Immunity*. 2012;37(3):487-500.
- 134. Hafner-Bratkovič I, Sušjan P, Lainšček D, et al. NLRP3 lacking the leucine-rich repeat domain can be fully activated via the canonical inflammasome pathway. *Nat Commun*. 2018;9(1):5182.
- 135. Gross O, Thomas CJ, Guarda G, Tschopp J. The inflammasome: an integrated view. *Immunol Rev*. 2011;243(1):136-151.
- 136. Bauernfeind FG, Horvath G, Stutz A, et al. Cutting edge: NF-κB activating pattern recognition and cytokine receptors license NLRP3 inflammasome activation by regulating NLRP3 expression. *J Immunol.* 2009;183(2):787-791.
- 137. Lamkanfi M, Dixit VM. Inflammasomes and their roles in health and disease. *Annu Rev Cell Dev Biol*. 2012;28(May):137-161
- Luo XQ, Duan JX, Yang HH, et al. Epoxyeicosatrienoic acids inhibit the activation of NLRP3 inflammasome in murine macrophages. *J Cell Physiol.* 2020;235(12):9910-9921.
- 139. Palazon-Riquelme P, Lopez-Castejon G. The inflammasomes, immune guardians at defence barriers. *Immunology*. 2018;155(3):320-330.
- 140. Cantin AM. The P2×7 receptor in cystic fibrosis monocytes: linking CFTR deficiency to inflammation. *Am J Respir Crit Care Med*. 2022;205(7):740-742.
- 141. Juliana C, Fernandes-Alnemri T, Kang S, Farias A, Qin F, Alnemri ES. Non-transcriptional priming and deubiquitination regulate NLRP3 inflammasome activation. *J Biol Chem.* 2012;287(43):36617-36622.
- 142. Mariathasan S, Weiss DS, Newton K, et al. Cryopyrin activates the inflammasome in response to toxins and ATP. *Nature*. 2006;440(7081):228-232.
- 143. He Y, Zeng MY, Yang D, Motro B, Núñez G. NEK7 is an essential mediator of NLRP3 activation downstream of potassium efflux. *Nature*. 2016;530(7590):354-357.

- 144. Walev I, Reske K, Palmer M, Valeva A, Bhakdi S. Potassium-inhibited processing of IL-1 β in human monocytes. *EMBO J*. 1995;14(8):1607-1614.
- 145. Iyer SS, He Q, Janczy JR, et al. Mitochondrial cardiolipin is required for Nlrp3 inflammasome activation. *Immunity*. 2013;39(2):311-323.
- Kayagaki N, Wong MT, Stowe IB, et al. Noncanonical inflammasome activation by intracellular LPS *independent of TLR4*. 2013;341(6151):1246-1249.
- 147. Broz P, Ruby T, Belhocine K, et al. Caspase-11 increases susceptibility to Salmonella infection in the absence of caspase-1. Nature. 2012;490(7419):288-291.
- 148. Case CL, Kohler LJ, Lima JB, et al. Caspase-11 stimulates rapid flagellin-independent pyroptosis in response to Legionella pneumophila. *Proc Natl Acad Sci USA*. 2013;110(5):1851-1856.
- 149. Rühl S, Broz P. Caspase-11 activates a canonical NLRP3 inflammasome by promoting K+ efflux. *Eur J Immunol*. 2015:45(10):2927-2936.
- Kayagaki N, Stowe IB, Lee BL, et al. Caspase-11 cleaves gasdermin D for non-canonical inflammasome signalling. *Nature*. 2015;526(7575):666-671.
- Shi J, Zhao Y, Wang K, et al. Cleavage of GSDMD by inflammatory caspases determines pyroptotic cell death. *Nature*. 2015;526(7575):660-665.
- 152. Chen X, He WT, Hu L, et al. Pyroptosis is driven by non-selective gasdermin-D pore and its morphology is different from MLKL channel-mediated necroptosis. *Cell Res.* 2016;26(9):1007-1020.
- Yu J, Nagasu H, Murakami T, et al. Inflammasome activation leads to Caspase-1-dependent mitochondrial damage and block of mitophagy. *Proc Natl Acad Sci USA*. 2014;111(43):15514-15519.
- 154. Gupta SC, Hevia D, Patchva S, Park B, Koh W, Aggarwal BB. Upsides and downsides of reactive oxygen species for cancer: the roles of reactive oxygen species in tumorigenesis, prevention, and therapy. *Antioxid Redox Signal*. 2012;16(11):1295-1322.
- Choi YJ, Kim S, Choi Y, et al. SERPINB1-mediated checkpoint of inflammatory caspase activation. *Nat Immunol*. 2019;20(3):276-287.
- Li Q, Shi N, Cai C, et al. The role of mitochondria in pyroptosis. Front Cell Dev Biol. 2021;8(January):1-5.
- Bernard NJ. Mitochondria control pyroptosis. Nat Immunol. 2021;22(9):1071.
- Liu Z, Xiao TS. Partners with a killer: metabolic signaling promotes inflammatory cell death. Cell. 2021;184(17):4374-4376.
- Vora SM, Lieberman J, Wu H. Inflammasome activation at the crux of severe COVID-19. Nat Rev Immunol. 2021;21(11):694-703.
- 160. Evavold CL, Hafner-Bratkovič I, Devant P, et al. Control of gasdermin D oligomerization and pyroptosis by the ragulator-Rag-mTORC1 pathway. *Cell*. 2021;184(17):4495-4511. e19.
- 161. Huang Y, Zhou JH, Zhang H, et al. Brown adipose TRX2 deficiency activates mtDNA-NLRP3 to impair thermogenesis and protect against diet-induced insulin resistance. *J Clin Invest*. 2022;132(9):e148852.
- 162. Wang X, Li X, Liu S, et al. PCSK9 regulates pyroptosis via mtDNA damage in chronic myocardial ischemia. *Basic Res Cardiol.* 2020;115(6):1-14.
- 163. Tapia-Abellán A, Angosto-Bazarra D, Alarcón-Vila C, et al. Sensing low intracellular potassium by NLRP3 results in a



- stable open structure that promotes inflammasome activation. *Sci Adv.* 2021;7(38):eabf4468.
- 164. Camilli G, Bohm M, Piffer AC, et al. β-Glucan-induced reprogramming of human macrophages inhibits NLRP3 inflammasome activation in cryopyrinopathies. *J Clin Invest*. 2020:130(9):4561-4573.
- Andersson U, Tracey KJ. Reflex principles of immunological homeostasis. Annu Rev Immunol. 2012;30:313-335.
- 166. Torres-Rosas R, Yehia G, Peña G, et al. Dopamine mediates vagal modulation of the immune system by electroacupuncture. Nat Med. 2014;20(3):291-295.
- Semmler A, Hermann S, Mormann F, et al. Sepsis causes neuroinflammation and concomitant decrease of cerebral metabolism. J Neuroinflammation. 2008;5:1-10.
- 168. Steckert AV, Comim CM, Mina F, et al. Late brain alterations in sepsis-survivor rats. *Synapse*. 2013;67(11):786-793.
- Manabe T, Heneka MT. Cerebral dysfunctions caused by sepsis during ageing. *Nat Rev Immunol*. 2021;0123456789(Table 1).
- 170. Orhun G, Tüzün E, Bilgiç B, et al. Brain volume changes in patients with acute brain dysfunction due to sepsis. *Neurocrit Care*. 2020;32(2):459-468.
- 171. Sharshar T, Gray F, De La Grandmaison GL, et al. Apoptosis of neurons in cardiovascular autonomic centres triggered by inducible nitric oxide synthase after death from septic shock. *Lancet*. 2003;362(9398):1799-1805.
- 172. Naito Y, Yoshioka K, Tanaka K, Tatsumi K, Kimura S, Kasuya Y. Endothelin B receptor-mediated encephalopathic events in mouse sepsis model. *Life Sci.* 2014;118(2):340-346.
- 173. Stare J, Siami S, Trudel E, Prager-Khoutorsky M, Sharshar T, Bourque CW. Effects of peritoneal sepsis on rat central osmoregulatory neurons mediating thirst and vasopressin release. *J Neurosci.* 2015;35(35):12188-12197.
- 174. Kanczkowski W, Sue M, Zacharowski K, Reincke M, Bornstein SR. The role of adrenal gland microenvironment in the HPA axis function and dysfunction during sepsis. *Mol Cell Endocrinol.* 2015;408:241-248.
- 175. Hoover DB. Cholinergic modulation of the immune system presents new approaches for treating inflammation. *Pharmacol Ther*. 2017;179:1-16.
- 176. Salama M, Akan A, Mueller MR. Transcutaneous stimulation of auricular branch of the vagus nerve attenuates the acute inflammatory response after lung lobectomy. *World J Surg*. 2020;44(9):3167-3174.
- 177. Kohoutova M, Horak J, Jarkovska D, et al. Vagus nerve stimulation attenuates multiple organ dysfunction in resuscitated porcine progressive sepsis. *Crit Care Med.* 2019;47(6):e461-e469.
- 178. Deng B, Yang D, Wu H, et al. Ketamine inhibits TNF-α-induced cecal damage by enhancing RIP1 ubiquitination to attenuate lethal SIRS. *Cell Death Discov*. 2022;8(1):1-12.
- 179. Huang WY, Lai YL, Liu KH, et al. TNFα-mediated necroptosis in brain endothelial cells as a potential mechanism of increased seizure susceptibility in mice following systemic inflammation. *J Neuroinflammation*. 2022;19(1):1-14.
- 180. Li J, Zhou L, Li Z, Yang S, Tang L, Gong H. Identification of crucial genes and infiltrating immune cells underlying sepsis-induced cardiomyopathy via weighted gene co-expression network analysis. *Front Genet*. 2021;12(December):1-13.

- 181. Silke J, Rickard JA, Gerlic M. Erratum: the diverse role of RIP kinases in necroptosis and inflammation. *Nat Immunol*. 2015;16(8):889.
- 182. Lu Z, van Eeckhoutte HP, Liu G, et al. Necroptosis signaling promotes inflammation, airway remodeling, and emphysema in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med.* 2021;204(6):667-681.
- 183. Holler N, Zaru R, Micheau O, et al. Fas triggers an alternative, caspase-8-independent cell death pathway using the kinase RIP as effector molecule. *Nat Immunol.* 2000;1(6):489-495.
- 184. Alvarez-Diaz S, Preaudet A, Samson AL, et al. Necroptosis is dispensable for the development of inflammation-associated or sporadic colon cancer in mice. *Cell Death Differ*. 2021;28(5):1466-1476.
- 185. Khoury MK, Gupta K, Franco SR, Liu B. Necroptosis in the pathophysiology of disease. *Am J Pathol*. 2020;190(2):272-285.
- Zhang H, Zhou X, McQuade T, Li J, Chan FKM, Zhang J. Functional complementation between FADD and RIP1 in embryos and lymphocytes. *Nature*. 2011;471(7338):373-377.
- 187. Cho YS, Challa S, Moquin D, et al. Phosphorylation-driven assembly of the RIP1-RIP3 complex regulates programmed necrosis and virus-induced inflammation. *Cell*. 2009;137(6):1112-1123.
- 188. Menon MB, Gropengießer J, Fischer J, et al. P38 MAPK /MK2-dependent phosphorylation controls cytotoxic RIPK1 signalling in inflammation and infection. *Nat Cell Biol*. 2017;19(10):1248-1259.
- 189. He S, Wang L, Miao L, et al. Receptor interacting protein kinase-3 determines cellular necrotic response to TNF-α. Cell. 2009;137(6):1100-1111.
- 190. He S, Liang Y, Shao F, Wang X. Toll-like receptors activate programmed necrosis in macrophages through a receptorinteracting kinase-3-mediated pathway. *Proc Natl Acad Sci* USA. 2011;108(50):20054-20059.
- Kaiser WJ, Sridharan H, Huang C, et al. Toll-like receptor 3mediated necrosis via TRIF, RIP3, and MLKL. *J Biol Chem*. 2013;288(43):31268-31279.
- 192. McComb S, Cessford E, Alturki NA, et al. Type-I interferon signaling through ISGF3 complex is required for sustained Rip3 activation and necroptosis in macrophages. *Proc Natl Acad Sci USA*. 2014;111(31):3206-3213.
- 193. Thapa RJ, Nogusa S, Chen P, et al. Interferon-induced RIP1/RIP3-mediated necrosis requires PKR and is licensed by FADD and caspases. *Proc Natl Acad Sci USA*. 2013;110(33):3109-3118
- 194. Li D, Wu M. Pattern recognition receptors in health and diseases. Signal Transduct Target Ther. 2021;6(1):1-24.
- 195. Huang L, Chen J, Li X, et al. Polydatin improves sepsisassociated encephalopathy by activating Sirt1 and reducing p38 phosphorylation. *J Surg Res.* 2022;276:379-393.
- 196. Welz PS, Wullaert A, Vlantis K, et al. FADD prevents RIP3-mediated epithelial cell necrosis and chronic intestinal inflammation. *Nature*. 2011;477(7364):330-334.
- Upton JW, Kaiser WJ, Mocarski ES. DAI/ZBP1/DLM-1 complexes with RIP3 to mediate virus-induced programmed necrosis that is targeted by murine cytomegalovirus vIRA. *Cell Host Microbe*. 2012;11(3):290-297.



- 198. Geserick P, Hupe M, Moulin M, et al. Cellular IAPs inhibit a cryptic CD95-induced cell death by limiting RIP1 kinase recruitment. *J Cell Biol*. 2009;187(7):1037-1054.
- 199. Kaiser WJ, Upton JW, Long AB, et al. RIP3 mediates the embryonic lethality of caspase-8-deficient mice. *Nature*. 2011;471(7338):368-373.
- 200. Tan S, Tan S, Zhao J, et al. Hepatocyte-specific TAK1 deficiency drives RIPK1 kinase-dependent inflammation to promote liver fibrosis and hepatocellular carcinoma. *Proc Natl Acad Sci USA*. 2020;117(25):14231-14242.
- He L, Huang Z, Huang K, et al. Optogenetic control of nonapoptotic cell death. Adv Sci. 2021;8(13):1-15.
- 202. Moujalled D, Strasser A, Liddell JR. Molecular mechanisms of cell death in neurological diseases. *Cell Death Differ*. 2021;28(7):2029-2044.
- 203. Takeuchi Y, Ohara D, Watanabe H, et al. Dispensable roles of Gsdmd and Ripk3 in sustaining IL-1β production and chronic inflammation in Th17-mediated autoimmune arthritis. *Sci Rep.* 2021:11(1):1-11.
- Chen H, Li Y, Wu J, et al. RIPK3 collaborates with GSDMD to drive tissue injury in lethal polymicrobial sepsis. *Cell Death Differ*. 2020;27(9):2568-2585.
- 205. Sharma A, Matsuo S, Yang WL, Wang Z, Wang P. Receptor-interacting protein kinase 3 deficiency inhibits immune cell infiltration and attenuates organ injury in sepsis. *Crit Care*. 2014;18(4):1-13.
- 206. Bolognese AC, Yang WL, Hansen LW, et al. Inhibition of necroptosis attenuates lung injury and improves survival in neonatal sepsis. *Surgery (United States)*. 2018;164(1): 110-116.
- 207. Hansen LW, Jacob A, Yang WL, et al. Deficiency of receptorinteracting protein kinase 3 (RIPK3) attenuates inflammation and organ injury in neonatal sepsis. *J Pediatr Surg*. 2018;53(9):1699-1705.
- Wang W, Gong YN. MLKL ubiquitylation: more than a makeover. Cell Death Differ. 2022;29(2):269-271.
- 209. Aird WC. The role of the endothelium in severe sepsis and multiple organ dysfunction syndrome. *Blood*. 2003;101(10):3765-3777
- 210. Opal SM, van der Poll T. Endothelial barrier dysfunction in septic shock. *J Intern Med.* 2015;277(3):277-293.
- 211. Tressel SL, Kaneider NC, Kasuda S, et al. A matrix metalloprotease-PAR1 system regulates vascular integrity, systemic inflammation and death in sepsis. *EMBO Mol Med*. 2011;3(7):370-384.
- 212. Joffre J, Hellman J, Ince C, Ait-Oufella H. Endothelial responses in sepsis. *Am J Respir Crit Care Med.* 2020;202(3):361-370.
- 213. Wang Z, Sims CR, Patil NK, Gokden N, Mayeux PR. Pharmacologic targeting of sphingosine-1-phosphate receptor 1 improves the renal microcirculation during sepsis in the mouse. *J Pharmacol Exp Ther*. 2015;352(1):61-66.
- 214. Asehnoune K, Moine P. Protease-activated receptor-1: Key player in the sepsis coagulation-inflammation crosstalk. *Crit Care*. 2013;17(1):1-3.
- 215. Yano K, Liaw PC, Mullington JM, et al. Vascular endothelial growth factor is an important determinant of sepsis morbidity and mortality. *J Exp Med*. 2006;203(6):1447-1458.

- 216. Parikh SM, Mammoto T, Schultz A, et al. Excess circulating angiopoietin-2 may contribute to pulmonary vascular leak in sepsis in humans. *PLoS Med.* 2006;3(3):356-370.
- 217. Stiehl T, Thamm K, Kaufmann J, et al. Lung-targeted RNA interference against angiopoietin-2 ameliorates multiple organ dysfunction and death in sepsis. *Crit Care Med*. 2014;42(10):e654-e662.
- 218. Pizurki L, Zhou Z, Glynos K, Roussos C, Papapetropoulos A. Angiopoietin-1 inhibits endothelial permeability, neutrophil adherence and IL-8 production. *Br J Pharmacol*. 2003:139(2):329-336.
- 219. Mammoto T, Jiang A, Jiang E, Mammoto A. Plateletrich plasma extract prevents pulmonary edema through angiopoietin-Tie2 signaling. Am J Respir Cell Mol Biol. 2015;52(1):56-64.
- 220. Kümpers P, Gueler F, David S, et al. The synthetic Tie2 agonist peptide vasculotide protects against vascular leakage and reduces mortality in murine abdominal sepsis. *Crit Care*. 2011;15(5):1-14.
- 221. Kim M, Allen B, Korhonen EA, et al. Opposing actions of angiopoietin-2 on Tie2 signaling and FOXO1 activation. *J Clin Invest*. 2016;126(9):3511-3525.
- 222. Kümpers P, van Meurs M, David S, et al. Time course of angiopoietin-2 release during experimental human endotoxemia and sepsis. *Crit Care*. 2009;13(3):1-9.
- 223. Korhonen EA, Lampinen A, Giri H, et al. Tiel controls angiopoietin function in vascular remodeling and inflammation. *J Clin Invest*. 2016;126(9):3495-3510.
- 224. Chiang MD, Chang CY, Shih HJ, Le VL, Huang YH, Huang CJ. Exosomes from human placenta choriodecidual membrane-derived mesenchymal stem cells mitigate endoplasmic reticulum stress, inflammation, and lung injury in lipopolysaccharide-treated obese mice. *Antioxidants*. 2022;11(4):615.
- Sano R, Reed JC. ER stress-induced cell death mechanisms.
 Biochim Biophys Acta Mol Cell Res. 2013;1833(12):3460-3470.
- 226. Jian B, Hsieh CH, Chen J, et al. Activation of endoplasmic reticulum stress response following trauma-hemorrhage. *Biochim Biophys Acta Mol Basis Dis.* 2008;1782(11):621-626.
- 227. Senft D, Ronai ZA. UPR, autophagy, and mitochondria crosstalk underlies the ER stress response. *Trends Biochem Sci.* 2015;40(3):141-148.
- 228. Kim I, Xu W, Reed JC. Cell death and endoplasmic reticulum stress: disease relevance and therapeutic opportunities. *Nat Rev Drug Discov*. 2008;7(12):1013-1030.
- 229. Li Y, Guo Y, Tang J, Jiang J, Chen Z. Erratum: New insights into the roles of CHOP-induced apoptosis in ER stress (Acta Biochimica et Biophysica Sinica (2014) 46 (629-640) h tt ps://doi.org/10.1093/abbs/gmu048). *Acta Biochim Biophys Sin (Shanghai)*. 2015;47(2):146-147.
- García de la Cadena S, Massieu L. Caspases and their role in inflammation and ischemic neuronal death. Focus on caspase-12. Apoptosis. 2016;21(7):763-777.
- Bettigole SE, Glimcher LH. Endoplasmic reticulum stress in immunity. Annu Rev Immunol. 2015;33(*):107-138.
- 232. Endo M, Oyadomari S, Suga M, Mori M, Gotoh T. The ER stress pathway involving CHOP is activated in the lungs of LPS-treated mice. *J Biochem.* 2005;138(4):501-507.



- 233. Wang H, Zhang G. Endoplasmic reticulum stress-mediated autophagy protects against β , β -dimethylacrylshikonin-induced apoptosis in lung adenocarcinoma cells. *Cancer Sci.* 2018:109(6):1889-1901.
- 234. Jiao G, Hao L, Wang M, et al. Upregulation of endoplasmic reticulum stress is associated with diaphragm contractile dysfunction in a rat model of sepsis. *Mol Med Rep.* 2017;15(1):366-374.
- Minamino T, Komuro I, Kitakaze M. Endoplasmic reticulum stress as a therapeutic target in cardiovascular disease. *Circ Res.* 2010;107(9):1071-1082.
- 236. Ghosh S, Adhikary A, Chakraborty S, et al. Cross-talk between endoplasmic reticulum (ER) stress and the MEK/ERK pathway potentiates apoptosis in human triple negative breast carcinoma cells: role of a dihydropyrimidone, nifetepimine. *J Biol Chem.* 2015;290(7):3936-3949.
- 237. Saleh M, Mathison JC, Wolinski MK, et al. Enhanced bacterial clearance and sepsis resistance in caspase-12-deficient mice. *Nature*. 2006;440(7087):1064-1068.
- 238. Ma T, Han L, Gao Y, et al. The endoplasmic reticulum stress-mediated apoptosis signal pathway is involved in sepsis-induced abnormal lymphocyte apoptosis. *Eur Surg Res.* 2008;41(2):219-225.
- 239. Ferlito M, Wang Q, Fulton WB, et al. H 2 S increases survival during sepsis: protective effect of CHOP inhibition. *J Immunol*. 2014;192(4):1806-1814.
- Ron D, Walter P. Signal integration in the endoplasmic reticulum unfolded protein response. Nat Rev Mol Cell Biol. 2007:8(7):519-529.
- 241. Hetz C. The unfolded protein response: Controlling cell fate decisions under ER stress and beyond. *Nat Rev Mol Cell Biol.* 2012;13(2):89-102.
- 242. Racanelli AC, Kikkers SA, Choi AMK, Cloonan SM. Autophagy and Inflammation in Chronic Respiratory Disease. Vol 14. Taylor & Francis; 2018.
- 243. Tang D, Kang R, Zeh HJ, Lotze MT. High-mobility group box 1, oxidative stress, and disease. *Antioxid Redox Signal*. 2011;14(7):1315-1335.
- 244. Jo EK, Yuk JM, Shin DM, Sasakawa C. Roles of autophagy in elimination of intracellular bacterial pathogens. *Front Immunol.* 2013;4(MAY):1-10.
- 245. Mizumura K, Cloonan SM, Haspel JA, Choi AMK. The emerging importance of autophagy in pulmonary diseases. *Chest.* 2012;142(5):1289-1299.
- 246. Singer M. The role of mitochondrial dysfunction in sepsisinduced multi-organ failure. *Virulence*. 2014;5(1):66-72.
- Shintani T, Klionsky DJ. Autophagy in health and disease: a double-edged sword. *Science* (1979). 2004;306(5698):990-995.
- 248. Ren C, Zhang H, Wu TT, Yao YM. Autophagy: a potential therapeutic target for reversing sepsis-induced immunosuppression. *Front Immunol.* 2017;8(DEC):1-10.
- 249. Gkikas I, Palikaras K, Tavernarakis N. The role of mitophagy in innate immunity. *Front Immunol.* 2018;9(JUN):1-15.
- Oh JE, Lee HK. Pattern recognition receptors and autophagy. Front Immunol. 2014;5(JUN):1-7.
- Delgado MA, Elmaoued RA, Davis AS, Kyei G, Deretic V. Tolllike receptors control autophagy. EMBO J. 2008;27(7):1110-1121.
- 252. Tang D, Kang R, Livesey KM, et al. Endogenous HMGB1 regulates autophagy. *J Cell Biol*. 2010;190(5):881-892.

- Choi ME. Autophagy in kidney disease. Annu Rev Physiol. 2020;82:297-322.
- 254. Kang R, Livesey KM, Zeh HJ, Lotze MT, Tang D. HMGB1: a novel Beclin 1-binding protein active in autophagy. *Autophagy*. 2010;6(8):1209-1211.
- 255. Zhu X, Messer JS, Wang Y, et al. Cytosolic HMGB1 controls the cellular autophagy/apoptosis checkpoint during inflammation. *J Clin Invest.* 2015;125(3):1098-1110.
- 256. Liu L, Yang M, Kang R, et al. HMGB1-DNA complex-induced autophagy limits AIM2 inflammasome activation through RAGE. Biochem Biophys Res Commun. 2014;450(1):851-856.
- 257. Padman BS, Nguyen TN, Uoselis L, Skulsuppaisarn M, Nguyen LK, Lazarou M. LC3/GABARAPs drive ubiquitin-independent recruitment of Optineurin and NDP52 to amplify mitophagy. Nat Commun. 2019;10(1):1-13.
- Lee YK, Lee JA. Role of the mammalian ATG8/LC3 family in autophagy: differential and compensatory roles in the spatiotemporal regulation of autophagy. BMB Rep. 2016;49(8):424-430
- 259. Zhong W, Zhu H, Sheng F, et al. Activation of the MAPK11/12/13/14 (p38 MAPK) pathway regulates the transcription of autophagy genes in response to oxidative stress induced by a novel copper complex in HeLa cells. *Autophagy*. 2014;10(7):1285-1300.
- 260. Hirota Y, Yamashita SI, Kurihara Y, et al. Mitophagy is primarily due to alternative autophagy and requires the MAPK1 and MAPK14 signaling pathways. *Autophagy*. 2015;11(2):332-343.
- 261. Webber JL, Tooze SA. Coordinated regulation of autophagy by p38a MAPK through mAtg9 and p38IP. *EMBO J.* 2010;29(1):27-40
- 262. Lazarou M, Sliter DA, Kane LA, et al. The ubiquitin kinase PINK1 recruits autophagy receptors to induce mitophagy. *Nature*. 2015;524(7565):309-314.
- Koyano F, Okatsu K, Kosako H, et al. Ubiquitin is phosphorylated by PINK1 to activate parkin. *Nature*. 2014;510(7503):162-166
- 264. Kim J, Kundu M, Viollet B, Guan KL. AMPK and mTOR regulate autophagy through direct phosphorylation of Ulk1. *Nat Cell Biol.* 2011;13(2):132-141.
- Narendra DP, Jin SM, Tanaka A, et al. PINK1 is selectively stabilized on impaired mitochondria to activate Parkin. *PLoS Biol*. 2010;8(1):e1000298.
- 266. Wang Y, Tang C, Cai J, et al. PINK1/Parkin-mediated mitophagy is activated in cisplatin nephrotoxicity to protect against kidney injury. *Cell Death Dis.* 2018;9(11):1113.
- 267. Gomez-Suaga P, Paillusson S, Stoica R, Noble W, Hanger DP, Miller CCJ. The ER-mitochondria tethering complex VAPB-PTPIP51 regulates autophagy. *Curr Biol.* 2017;27(3):371-385.
- 268. Chauhan S, Jena KK, Mehto S, et al. Innate immunity and inflammophagy: balancing the defence and immune homeostasis. *FEBS J*. Published online 2021:1-20.
- Ho J, Yu J, Wong SH, et al. Autophagy in sepsis: degradation into exhaustion? *Autophagy*. 2016;12(7):1073-1082.
- 270. Hsieh YC, Athar M, Chaudry IH. When apoptosis meets autophagy: deciding cell fate after trauma and sepsis. *Trends Mol Med*. 2009;15(3):129-138.
- Lin CW, Lo S, Hsu C, et al. T-cell autophagy deficiency increases mortality and suppresses immune responses after sepsis. *PLoS One*. 2014;9(7):e102066.



- 272. Oami T, Watanabe E, Hatano M, et al. Suppression of T cell autophagy results in decreased viability and function of T cells through accelerated apoptosis in a murine sepsis model. *Crit Care Med*. 2017;45(1):e77-e85.
- 273. Rathkey JK, Xiao TS, Abbott DW. Human polymorphisms in GSDMD alter the inflammatory response. *J Biol Chem.* 2020;295(10):3228-3238.
- 274. Wang G, Jin S, Huang W, et al. LPS-induced macrophage HMGB1-loaded extracellular vesicles trigger hepatocyte pyroptosis by activating the NLRP3 inflammasome. *Cell Death Discov*, 2021;7(1):1-11.
- 275. Hu Q, Ren H, Li G, et al. STING-mediated intestinal barrier dysfunction contributes to lethal sepsis. *EBioMedicine*. 2019;41:497-508.
- 276. Long J, Yang C, Zheng Y, et al. Notch signaling protects CD4 T cells from STING-mediated apoptosis during acute systemic inflammation. Sci Adv. 2020;6(39):1-13.
- 277. Wu J, Chen YJ, Dobbs N, et al. STING-mediated disruption of calcium homeostasis chronically activates ER stress and primes T cell death. *J Exp Med*. 2019;216(4):867-883.
- Vandewalle J, Vanderhaeghen T, Beyaert R, Libert C. Taking the STING out of sepsis? *Cell Host Microbe*. 2020;27(4):491-493.
- Zou B, Liu J, Klionsky DJ, Tang D, Kang R. Extracellular SQSTM1 as an inflammatory mediator. *Autophagy*. 2020;16(12):2313-2315.
- Zhou B, Liu J, Zeng L, et al. Extracellular SQSTM1 mediates bacterial septic death in mice through insulin receptor signalling. *Nat Microbiol.* 2020;5(12):1576-1587.
- 281. Karunarathne DS, Horne-Debets JM, Huang JX, et al. Programmed death-1 ligand 2-mediated regulation of the PD-L1 to PD-1 axis is essential for establishing CD4+ T Cell immunity. *Immunity*. 2016;45(2):333-345.
- 282. Monneret G, Gossez M, Venet F. Sepsis in PD-1 light. *Crit Care*. 2016;20(1):1-2.
- 283. Young JS, Heffernan DS, Chung CS, et al. Effect of PD-1:PD-L1 in invariant natural killer T-cell emigration and chemotaxis following sepsis. *Shock.* 2016;45(5):534-549.
- 284. Kang R, Zeng L, Xie Y, et al. A novel PINK1- and PARK2-dependent protective neuroimmune pathway in lethal sepsis. *Autophagy*. 2016;12(12):2374-2385.
- 285. Billia F, Hauck L, Grothe D, et al. Parkinson-susceptibility gene DJ-1/PARK7 protects the murine heart from oxidative damage in vivo. *Proc Natl Acad Sci USA*. 2013;110(15):6085-6090.
- 286. Hijioka M, Inden M, Yanagisawa D, Kitamura Y. Brain damage and potential therapeutic interventions DJ-1 /PARK7: a new therapeutic target for neurodegenerative disorders. *Biol Pharm Bull.* 2017;40(5):548-552.
- Essler S, Dehne N, Brüne B. Role of sestrin2 in peroxide signaling in macrophages. FEBS Lett. 2009;583(21):3531-3535.
- 288. Kim MJ, Bae SH, Ryu JC, et al. SESN2/sestrin2 suppresses sepsis by inducing mitophagy and inhibiting NLRP3 activation in macrophages. *Autophagy*. 2016;12(8):1272-1291.
- 289. Villarino AV., Kanno Y, O'Shea JJ. Mechanisms and consequences of Jak-STAT signaling in the immune system. *Nat Immunol*. 2017;18(4):374-384.
- 290. Cai B, Cai JP, Luo YL, Chen C, Zhang S. The specific roles of JAK/STAT signaling pathway in sepsis. *Inflammation*. 2015;38(4):1599-1608.

- 291. Sun P, Zhong J, Liao H, et al. Hepatocytes are resistant to cell death from canonical and non-canonical inflammasome-activated pyroptosis. *Cmgh.* 2022;13(3):739-757.
- Burdette BE, Esparza AN, Zhu H, Wang S. Gasdermin D in pyroptosis. Acta Pharm Sin B. 2021;11(9):2768-2782.
- 293. Tsujimoto H, Kobayashi M, Sugasawa H, Ono S, Kishi Y, Ueno H. Potential mechanisms of tumor progression associated with postoperative infectious complications. *Cancer Metastasis Rev.* 2021;40(1):285-296.
- Vignon P, Laterre PF, Daix T, François B. New agents in development for sepsis: any reason for hope? *Drugs*. 2020;80(17):1751-1761
- 295. Karbian N, Abutbul A, el-Amore R, et al. Apoptotic cell therapy for cytokine storm associated with acute severe sepsis. *Cell Death Dis*. 2020;11(7):535.
- 296. Kumari P, Russo AJ, Wright SS, Muthupalani S, Rathinam VA. Hierarchical cell-type-specific functions of caspase-11 in LPS shock and antibacterial host defense. *Cell Rep.* 2021;35(3):109012.
- 297. Linder A, Hornung V. Irgm2 and Gate-16 put a break on caspase-11 activation. *EMBO Rep.* 2020;21(11):10-12.
- Rathkey JK, Zhao J, Liu Z, et al. Chemical disruption of the pyroptotic pore-forming protein gasdermin D inhibits inflammatory cell death and sepsis. Sci Immunol. 2018;3(26):eaat2738.
- Volchuk A, Ye A, Chi L, Steinberg BE, Goldenberg NM. Indirect regulation of HMGB1 release by gasdermin D. Nat Commun. 2020;11(1):1-11.
- Xia S. Biological mechanisms and therapeutic relevance of the gasdermin family. Mol Aspects Med. 2020:76(May):100890.
- 301. Xu S, Zhou Z, Li H, et al. BMSCs ameliorate septic coagulopathy by suppressing inflammation in cecal ligation and puncture-induced sepsis. *J Cell Sci.* 2018;131(3):jcs211151.
- Wu C, Lu W, Zhang Y, et al. Inflammasome activation triggers blood clotting and host death through pyroptosis. *Immunity*. 2019;50(6):1401-1411.e4.
- 303. Zhang H, Zeng L, Xie M, et al. TMEM173 drives lethal coagulation in sepsis. *Cell Host Microbe*. 2020;27(4):556-570.e6.
- 304. Yang D, He Y, Muñoz-Planillo R, Liu Q, Núñez G. Caspase-11 requires the pannexin-1 channel and the purinergic P2×7 pore to mediate pyroptosis and endotoxic shock. *Immunity*. 2015;43(5):923-932.
- 305. Pawlinski R, Pedersen B, Schabbauer G, et al. Role of tissue factor and protease-activated receptors in a mouse model of endotoxemia. *Blood*. 2004;103(4):1342-1347.
- 306. Wang Y, Zhang X, Guo Y, et al. Type 1 interferon aggravates lipopolysaccharide-induced sepsis through upregulating Caspase-11 and Gasdermin D. *J Physiol Biochem*. 2021;77(1):85-92.
- 307. Huang H, Nace GW, Mcdonald KA, et al. Hepatocyte-specific high-mobility group box 1 deletion worsens the injury in liver ischemia/reperfusion: a role for intracellular high-mobility group box 1 in cellular protection. *Hepatology*. 2014;59(5):1984-1997.
- Kang R, Chen R, Zhang Q, et al. HMGB1 in health and disease.
 Mol Aspects Med. 2014;40(July):1-116.
- Chen R, Huang Y, Quan J, et al. HMGB1 as a potential biomarker and therapeutic target for severe COVID-19. *Heliyon*. 2020;6(12):e05672.



- 310. Deng M, Tang Y, Li W, et al. The endotoxin delivery protein HMGB1 mediates caspase-11-dependent lethality in sepsis. *Immunity*. 2018;49(4):740-753.e7.
- 311. Li Z, Fu WJ, Chen XQ, et al. Autophagy-based unconventional secretion of HMGB1 in glioblastoma promotes chemosensitivity to temozolomide through macrophage M1-like polarization. *J Exp Clin Cancer Res.* 2022;41(1):1-20.
- 312. Camacho-Mejorado R, Gómez R, Torres-Sánchez LE, et al. ALOX5, LPA, MMP9 and TPO gene polymorphisms increase atherothrombosis susceptibility in middle-aged Mexicans. *R Soc Open Sci.* 2020;7(1):190735.
- 313. Yang X, Cheng X, Tang Y, et al. The role of type 1 interferons in coagulation induced by gram-negative bacteria. *Blood*. 2020;135(14):1087-1100.
- 314. Bustin M. Regulation of DNA-dependent activities by the functional motifs of the high-mobility-group chromosomal proteins. *Mol Cell Biol.* 1999;19(8):5237-5246.
- 315. Wang H, Yang H, Czura CJ, Sama AE, Tracey KJ. HMBG1 as a late mediator of lethal systemic inflammation. *Am J Respir Crit Care Med*. 2001;164(10 I):1768-1773.
- 316. Peng Y, Gao M, Liu Y, et al. Bacterial outer membrane vesicles induce disseminated intravascular coagulation through the caspase-11-gasdermin D pathway. *Thromb Res.* 2020;196(138):159-166.
- 317. Santos JC, Boucher D, Schneider LK, et al. Human GBP1 binds LPS to initiate assembly of a caspase-4 activating platform on cytosolic bacteria. *Nat Commun.* 2020;11(1):3276.
- 318. Fisch D, Bando H, Clough B, et al. Human GBP 1 is a microbe-specific gatekeeper of macrophage apoptosis and pyroptosis. *EMBO J.* 2019;38(13):1-19.
- 319. Deng M, Scott MJ, Loughran P, et al. Lipopolysaccharide clearance, bacterial clearance, and systemic inflammatory responses are regulated by cell type–specific functions of TLR4 during sepsis. *J Immunol*. 2013;190(10):5152-5160.
- 320. Karmakar M, Minns M, Greenberg EN, et al. N-GSDMD trafficking to neutrophil organelles facilitates IL-1 β release independently of plasma membrane pores and pyroptosis. *Nat Commun*. 2020;11(1):1-14.
- 321. Matsuda A, Jacob A, Wu R, et al. Novel therapeutic targets for sepsis: regulation of exaggerated inflammatory responses. *J Nippon Med Sch.* 2012;79(1):4-18.
- 322. Kamińska A, Enguita FJ, Stępień E. Lactadherin: an unappreciated haemostasis regulator and potential therapeutic agent. *Vascul Pharmacol.* 2018;101(September 2017):21-28.
- 323. Schulman S, El-Darzi E, Florido MHC, et al. A coagulation defect arising from heterozygous premature termination of tissue factor. *J Clin Invest*. 2020;130(10):5302-5312.
- Wu R, Wang N, Comish PB, Tang D, Kang R. Inflammasomedependent coagulation activation in sepsis. Front Immunol. 2021;12(March):1-8.
- 325. Pawlinski R, Wang JG, Owens AP, et al. Hematopoietic and nonhematopoietic cell tissue factor activates the coagulation cascade in endotoxemic mice. *Blood*. 2010;116(5):806-814.
- 326. Kayagaki N, Kornfeld OS, Lee BL, et al. NINJ1 mediates plasma membrane rupture during lytic cell death. *Nature*. 2021;591(7848):131-136.
- 327. Evavold CL, Ruan J, Tan Y, Xia S, Wu H, Kagan JC. The poreforming protein gasdermin D regulates interleukin-1 secretion from living macrophages. *Immunity*. 2018;48(1):35-44.e6.

- 328. Ousingsawat J, Wanitchakool P, Schreiber R, Kunzelmann K. Contribution of TMEM16F to pyroptotic cell death. *Cell Death Dis.* 2018;9(3):300.
- 329. Mahaut-Smith MP. A role for platelet TRPC channels in the Ca2+ response that induces procoagulant activity. *Sci Signal*. 2013;6(281):2013-2015.
- Agnew A, Nulty C, Creagh EM. Regulation, activation and function of caspase-11 during health and disease. *Int J Mol Sci.* 2021;22(4):1-20.
- 331. Sun Z, Nyanzu M, Yang S, et al. VX765 attenuates pyroptosis and HMGB1/TLR4/NF-κ B pathways to improve functional outcomes in TBI mice. *Oxid Med Cell Longev*. 2020;2020:7879629.
- 332. Barber GN. STING: infection, inflammation and cancer. *Nat Rev Immunol*. 2015;15(12):760-770.
- Heipertz EL, Harper J, Walker WE. STING and TRIF contribute to mouse sepsis, depending on severity of the disease model. *Shock*. 2017;47(5):621-631.
- 334. Saukkonen K, Lakkisto P, Pettilä V, et al. Cell-free plasma DNA as a predictor of outcome in severe sepsis and septic shock. Clin Chem. 2008;54(6):1000-1007.
- 335. Prabakaran T, Bodda C, Krapp C, et al. Attenuation of c GAS STING signaling is mediated by a p62/SQSTM 1-dependent autophagy pathway activated by TBK1 . *EMBO J*. 2018;37(8):e97858.
- 336. Rhodes A, Cecconi M. Cell-free DNA and outcome in sepsis. *Crit Care*. 2012;16(6):170.
- 337. Knoop KA, Coughlin PE, Floyd AN, et al. pathogenic Escherichia coli in a model of late-onset neonatal sepsis. *Proc Natl Acad Sci USA*. 2020;117(14):7941-7949.
- 338. Liu N, Pang X, Zhang H, Ji P. The cGAS-STING pathway in bacterial infection and bacterial immunity. *Front Immunol*. 2022;12(January):1-18.
- 339. Vieira R de S, Nascimento MS, Noronha IH, et al. STING signaling drives production of innate cytokines, generation of CD8+ T cells and enhanced protection against Trypanosoma cruzi infection. *Front Immunol.* 2022;12(January):1-13.
- 340. Gaidt MM, Ebert TS, Chauhan D, et al. The DNA inflammasome in human myeloid cells is initiated by a STING-cell death program upstream of NLRP3. *Cell*. 2017;171(5):1110-1124.e18.
- 341. Chunmei M, Yang D, Wang B, et al. Gasdermin d in macrophages restrains colitis by controlling cGAS-mediated inflammation. *Sci Adv.* 2020;6(21):eaaz6717.
- 342. Hopfner KP, Hornung V. Molecular mechanisms and cellular functions of cGAS–STING signalling. *Nat Rev Mol Cell Biol.* 2020;21(9):501-521.
- 343. Ge W, Hu Q, Fang X, et al. LDK378 improves micro- and macro-circulation via alleviating STING-mediated inflammatory injury in a Sepsis rat model induced by Cecal ligation and puncture. *J Inflamm (United Kingdom)*. 2019;16(1):1-10.
- 344. Garred P, Tenner AJ, Mollnes TE. Therapeutic targeting of the complement system: from rare diseases to pandemics. *Pharmacol Rev.* 2021;73(2):792-827.
- Merle NS, Noe R, Halbwachs-Mecarelli L, Fremeaux-Bacchi V, Roumenina LT. Complement system part II: role in immunity. Front Immunol. 2015;6(MAY):1-26.
- 346. Gustavsen A, Nymo S, Landsem A, et al. Combined inhibition of complement and CD14 attenuates bacteria-induced inflammation in human whole blood more efficiently than



- antagonizing the Toll-like receptor 4-MD2 complex. *J Infect Dis.* 2016;214(1):140-150.
- Koch MA. Sex bias in sepsis. *Cell Host Microbe*. 2018;24(5):613-615.
- 348. Carvelli J, Demaria O, Vély F, et al. Association of COVID-19 inflammation with activation of the C5a–C5aR1 axis. *Nature*. 2020;588(7836):146-150.
- 349. Ward PA, Guo RF, Riedemann NC. Manipulation of the complement system for benefit in sepsis. *Crit Care Res Pract.* 2012:2012:427607.
- 350. Shao Z, Nishimura T, Leung LLK, Morser J. Carboxypeptidase B2 deficiency reveals opposite effects of complement C3a and C5a in a murine polymicrobial sepsis model. *J Thromb Haemost*. 2015;13(6):1090-1102.
- 351. Ren J, Zhao Y, Yuan Y, et al. Complement depletion deteriorates clinical outcomes of severe abdominal sepsis: a conspirator of infection and coagulopathy in crime? PLoS One. 2012:7(10):1-9.
- 352. Muhlfelder TW, Niemetz J, Kreutzer D, Beebe D, Ward PA, Rosenfeld SI. C5 chemotactic fragment induces leukocyte production of tissue factor activity. A link between complement and coagulation. *J Clin Invest*. 1979;63(1):147-150.
- 353. Tedesco F, Pausa M, Nardon E, Introna M, Mantovani A, Dobrina A. The cytolytically inactive terminal complement complex activates endothelial cells to express adhesion molecules and tissue factor procoagulant activity. *J Exp Med*. 1997;185(9):1619-1627.
- 354. Platt JL, Dalmasso AP, Lindman BJ, Ihrcke NS, Bach FH. The role of C5a and antibody in the release of heparan sulfate from endothelial cells. *Eur J Immunol*. 1991;21(11):2887-2890.
- 355. Maekawa T, Krauss JL, Abe T, et al. Porphyromonas gingivalis manipulates complement and TLR signaling to uncouple bacterial clearance from inflammation and promote dysbiosis. *Cell Host Microbe*. 2014;15(6):768-778.
- 356. Edsfeldt A, Swart M, Singh P, et al. Interferon regulatory factor-5-dependent CD11c+ macrophages contribute to the formation of rupture–prone atherosclerotic plaques. *Eur Heart J*. Published online 2022:1-14.
- 357. Sun X, Wu J, Liu L, et al. Transcriptional switch of hepatocytes initiates macrophage recruitment and T cell suppression in endotoxemia. *J Hepatol*. Published online 2022:1-17.
- 358. McGettrick AF, O'Neill LAJ. The role of HIF in immunity and inflammation. *Cell Metab.* 2020;32(4):524-536.
- Peace CG, O'Neill LAJ. The role of itaconate in host defense and inflammation. J Clin Invest. 2022;132(2):1-10.
- 360. Castegna A, Gissi R, Menga A, et al. Pharmacological targets of metabolism in disease: opportunities from macrophages. *Pharmacol Ther*. 2020;210:107521.
- 361. van der Post S, Birchenough GMH, Held JM. NOXI-dependent redox signaling potentiates colonic stem cell proliferation to adapt to the intestinal microbiota by linking EGFR and TLR activation. Cell Rep. 2021;35(1):108949.
- 362. Holterman CE, Boisvert NC, Thibodeau JF, et al. Podocyte NADPH oxidase 5 promotes renal inflammation regulated by the Toll-like receptor pathway. *Antioxid Redox Signal*. 2019;30(15):1817-1830.
- 363. Shen ZY, Sun Q, Xia ZY, et al. Overexpression of DJ-1 reduces oxidative stress and attenuates hypoxia/reoxygenation injury

- in NRK-52E cells exposed to high glucose. *Int J Mol Med*. 2016;38(3):729-736.
- 364. Nagakubo D, Taira T, Kitaura H, et al. DJ-1, a novel oncogene which transforms mouse NIH3T3 cells in cooperation with ras. *Biochem Biophys Res Commun*. 1997;231(2):509-513.
- 365. Guo JF, Xiao B, Liao B, et al. Mutation analysis of Parkin, PINK1, DJ-1 and ATP13A2 genes in Chinese patients with autosomal recessive early-onset Parkinsonism. Mov Disord. 2008;23(14):2074-2079.
- 366. Clements CM, McNally RS, Conti BJ, Mak TW, Ting JPY. DJ-1, a cancer- and Parkinson's disease-associated protein, stabilizes the antioxidant transcriptional master regulator Nrf2. Proc Natl Acad Sci USA. 2006;103(41):15091-15096.
- Honbou K, Suzuki NN, Horiuchi M, et al. The crystal structure of DJ-1, a protein related to male fertility and Parkinson's disease. *J Biol Chem.* 2003;278(33):31380-31384.
- Dongworth RK, Mukherjee UA, Hall AR, et al. DJ-1 protects against cell death following acute cardiac ischemiareperfusion injury. Cell Death Dis. 2014;5(2):1-7.
- 369. Kim W, Kim DW, Jeong HJ, et al. Tat-DJ-1 protects neurons from ischemic damage in the ventral horn of rabbit spinal cord via increasing antioxidant levels. *Neurochem Res.* 2014;39(1):187-193.
- 370. Njau F, Haller H. Calcium dobesilate modulates pkcδ-nadph oxidase-mapk-nf-κb signaling pathway to reduce cd14, tlr4, and mmp9 expression during monocyte-to-macrophage differentiation: potential therapeutic implications for atherosclerosis. *Antioxidants*. 2021;10(11):1798.
- 371. Akoumianaki T, Vaporidi K, Diamantaki E, et al. Uncoupling of IL-6 signaling and LC3-associated phagocytosis drives immunoparalysis during sepsis. *Cell Host Microbe*. 2021;29(8):1277-1293. e6.
- 372. Jiang J, Huang K, Xu S, Garcia JGN, Wang C, Cai H. Targeting NOX4 alleviates sepsis-induced acute lung injury via attenuation of redox-sensitive activation of CaMKII/ERK1/2/MLCK and endothelial cell barrier dysfunction. *Redox Biol*. 2020:36:101638.
- 373. Kim YR, Hwang J, Koh HJ, et al. The targeted delivery of the c-Src peptide complexed with schizophyllan to macrophages inhibits polymicrobial sepsis and ulcerative colitis in mice. *Biomaterials*. 2016;89:1-13.
- 374. Simon F, Fernández R. Early Iipopolysaccharide-induced reactive oxygen species production evokes necrotic cell death in human umbilical vein endothelial cells. *J Hypertens*. 2009;27(6):1202-1216.
- 375. Wu J, Xu H, Yang M, Martin CM, Kvietys PR, Rui T. NADPH oxidase contributes to conversion of cardiac myocytes to a proinflammatory phenotype in sepsis. *Free Radic Biol Med*. 2009;46(10):1338-1345.
- 376. Yu HH, Xu Q, Chen HP, et al. Stable overexpression of DJ-1 protects H9c2 cells against oxidative stress under a hypoxia condition. *Cell Biochem Funct*. 2013;31(8):643-651.
- 377. Wu F, Schuster DP, Tyml K, Wilson JX. Ascorbate inhibits NADPH oxidase subunit p47phox expression in microvascular endothelial cells. Free Radic Biol Med. 2007;42(1):124-131.
- Wu F, Tyml K, Wilson JX. iNOS expression requires NADPH oxidase-dependent redox signaling in microvascular endothelial cells. *J Cell Physiol*. 2008;217(1):207-214.



- 379. Yanagida T, Tsushima J, Kitamura Y, et al. Oxidative stress induction of DJ-1 protein in reactive astrocytes scavenges free radicals and reduces cell injury. *Oxid Med Cell Longev*. 2009;2(1):36-42.
- 380. Sánchez-Martín P, Saito T, Komatsu M. p62/SQSTM1: 'Jack of all trades' in health and cancer. *FEBS J.* 2019;286(1):8-23.
- 381. Pietrocola F, Lachkar S, Enot DP, et al. Spermidine induces autophagy by inhibiting the acetyltransferase EP300. *Cell Death Differ*. 2015;22(3):509-516.
- 382. Biasizzo M, Trstenjak-prebanda M, Dolinar K, et al. Cystatin C deficiency increases LPS-induced sepsis and nlrp3 inflammasome activation in mice. *Cells*. 2021;10(8):1-18.
- 383. Mallarpu CS, Ponnana M, Prasad S, et al. Distinct cell death markers identified in critical care patient survivors diagnosed with sepsis. *Immunol Lett.* 2021;231(March 2020):1-10.
- 384. Prajsnar TK, Serba JJ, Dekker BM, et al. The autophagic response to Staphylococcus aureus provides an intracellular niche in neutrophils. *Autophagy*. 2021;17(4):888-902.
- 385. Kang R, Zeng L, Zhu S, et al. Lipid peroxidation drives gasdermin D-mediated pyroptosis in lethal polymicrobial sepsis. *Cell Host Microbe*. 2018;24(1):97-108.e4.
- 386. Samie M, Lim J, Verschueren E, et al. Selective autophagy of the adaptor TRIF regulates innate inflammatory signaling article. *Nat Immunol.* 2018;19(3):246-254.
- 387. Mitroi DN, Karunakaran I, Gräler M, et al. SGPL1 (sphingosine phosphate lyase 1) modulates neuronal autophagy via phosphatidylethanolamine production. *Autophagy*. 2017;13(5):885-899.
- Vasconcellos LRC, Dutra FF, Siqueira MS, et al. Protein aggregation as a cellular response to oxidative stress induced by heme and iron. *Proc Natl Acad Sci USA*. 2016;113(47):E7474-E7482.
- Giegerich AK, Kuchler L, Sha LK, et al. Autophagy-dependent PELI3 degradation inhibits proinflammatory IL1B expression. Autophagy. 2014;10(11):1937-1952.
- 390. Lu X, Fu H, Chen R, et al. Phosphoinositide specific phospholipase Cγl inhibitiondriven autophagy caused cell death in human lung adenocarcinoma A549 cells in vivo and in vitro. *Int J Biol Sci.* 2020;16(8):1427-1440.
- López-Grueso MJ, Lagal DJ, García-Jiménez ÁF, et al. Knockout of PRDX6 induces mitochondrial dysfunction and cell cycle arrest at G2/M in HepG2 hepatocarcinoma cells. *Redox Biol*. 2020;37:101737.
- 392. Yang H, Ni HM, Ding WX. The double-edged sword of MTOR in autophagy deficiency induced-liver injury and tumorigenesis. *Autophagy*. 2019;15(9):1671-1673.
- 393. Kumar V. T cells and their immunometabolism: a novel way to understanding sepsis immunopathogenesis and future therapeutics. *Eur J Cell Biol.* 2018;97(6):379-392.
- 394. Sun C, Mezzadra R, Schumacher TN. Regulation and function of the PD-L1 checkpoint. *Immunity*. 2018;48(3):434-452.
- Nakamori Y, Park EJ, Shimaoka M. Immune deregulation in sepsis and septic shock: reversing immune paralysis by targeting PD-1/PD-L1 pathway. Front Immunol. 2021;11(February):1-14.
- 396. Chen H, Huang N, Tian H, et al. Splenectomy provides protective effects against CLP-induced sepsis by reducing TRegs and PD-1/PD-L1 expression. *Int J Biochem Cell Biol.* 2021;136(157):105970.

- 397. Avendanõ-Ortiz J, Maroun-Eid C, Martín-Quirós A, et al. PD-L1 overexpression during endotoxin tolerance impairs the adaptive immune response in septic patients via HIF1α. *J Infect Dis.* 2018;217(3):393-404.
- 398. Patera AC, Drewry AM, Chang K, Beiter ER, Osborne D, Hotchkiss RS. Frontline science: defects in immune function in patients with sepsis are associated with PD-1 or PD-L1 expression and can be restored by antibodies targeting PD-1 or PD-L1. *J Leukoc Biol.* 2016;100(6):1239-1254.
- 399. Ruan WS, Feng MX, Xu J, et al. Early activation of myeloid-derived suppressor cells participate in sepsis-induced immune suppression via PD-L1/PD-1 axis. Front Immunol. 2020;11(July):1-15.
- 400. Sun S, Chen Y, Liu Z, et al. Serum-soluble PD-L1 may be a potential diagnostic biomarker in sepsis. *Scand J Immunol*. 2021;94(1):1-9.
- 401. Tan CL, Kuchroo JR, Sage PT, et al. PD-1 restraint of regulatory T cell suppressive activity is critical for immune tolerance. *J Exp Med*. 2021;218(1):e20182232.
- 402. Akçay G, Belmonte MA, Aquila B, et al. Inhibition of Mcl-1 through covalent modification of a noncatalytic lysine side chain. *Nat Chem Biol*. 2016;12(11):931-936.
- 403. Huang X, Venet F, Wang YL, et al. PD-1 expression by macrophages plays a pathologic role in altering microbial clearance and the innate inflammatory response to sepsis. *Proc Natl Acad Sci USA*. 2009;106(15):6303-6308.
- 404. Ayala A, Elphick GF, Kim YS, et al. Sepsis-induced potentiation of peritoneal macrophage migration is mitigated by programmed cell death receptor-1 gene deficiency. *J Innate Immun*. 2014;6(3):325-338.
- Zhu W, Bao R, Fan X, et al. PD-L1 blockade attenuated sepsisinduced liver injury in a mouse cecal ligation and puncture model. *Mediat Inflamm*. 2013;2013:361501.
- 406. Zhen J, Chen W. MiR-142 inhibits cecal ligation and puncture (CLP)-induced inflammation via inhibiting PD-L1 expression in macrophages and improves survival in septic mice. *Biomed Pharmacother*. 2018;97(July 2017):1479-1485.
- 407. Zheng J, Liang H, Xu C, et al. An unbalanced PD-L1/CD86 ratio in CD14 ++ CD16 + monocytes is correlated with HCV viremia during chronic HCV infection. *Cell Mol Immunol*. 2014;11(3):294-304.
- 408. Wilson JK, Zhao Y, Singer M, Spencer J, Shankar-Hari M. Lymphocyte subset expression and serum concentrations of PD-1/PD-L1 in sepsis pilot study. *Crit Care*. 2018;22(1):1-7.
- 409. Hutchins NA, Wang F, Wang Y, Chung CS, Ayala A. Kupffer cells potentiate liver sinusoidal endothelial cell injury in sepsis by ligating programmed cell death ligand-1. *J Leukoc Biol*. 2013;94(5):963-970.
- 410. Guignant C, Lepape A, Huang X, et al. Programmed death-1 levels correlate with increased mortality, nosocomial infection and immune dysfunctions in septic shock patients. *Crit Care*. 2011;15(2):1-11.
- Von Knethen A, Schäfer A, Kuchler L, et al. Tolerizing CTL by sustained hepatic PD-L1 expression provides a new therapy approach in mouse sepsis. *Theranostics*. 2019;9(7):2003-2016.
- 412. Jiang W, Li X, Wen M, et al. Increased percentage of PD-L1+ natural killer cells predicts poor prognosis in sepsis patients: a prospective observational cohort study. *Crit Care*. 2020;24(1):1-10.



- 413. Chen R, Zhou L. PD-1 signaling pathway in sepsis: does it have a future? *Clin Immunol*. 2021;229:108742.
- 414. Zhang Y, Zhou Y, Lou J, et al. PD-L1 blockade improves survival in experimental sepsis by inhibiting lymphocyte apoptosis and reversing monocyte dysfunction. *Crit Care*. 2010;14(6):1-9.
- Shalova IN, Lim JY, Chittezhath M, et al. Human monocytes undergo functional re-programming during sepsis mediated by hypoxia-inducible factor-1α. *Immunity*. 2015;42(3):484-498.
- 416. Zhang Y, Li J, Lou J, et al. Upregulation of programmed death-1 on T cells and programmed death ligand-1 on monocytes in septic shock patients. *Crit Care*. 2011;15(1):1-9.
- 417. Avendaño-Ortiz J, Maroun-Eid C, Martín-Quirós A, et al. Oxygen saturation on admission is a predictive biomarker for PD-L1 expression on circulating monocytes and impaired immune response in patients with sepsis. *Front Immunol*. 2018;9(SEP):1-10.
- 418. Wang F, Huang X, Chung CS, Chen Y, Hutchins NA, Ayala A. Contribution of programmed cell death receptor (PD)-1 to kupffer cell dysfunction in murine polymicrobial sepsis. Am J Physiol Gastrointest Liver Physiol. 2016;311(2):G237-G245.
- Liang X, Li T, Zhou Q, et al. Mesenchymal stem cells attenuate sepsis-induced liver injury via inhibiting M1 polarization of Kupffer cells. *Mol Cell Biochem*. 2019;452(1-2):187-197.
- 420. Chang K, Svabek C, Vazquez-Guillamet C, et al. Targeting the programmed cell death 1: programmed cell death ligand 1 pathway reverses T cell exhaustion in patients with sepsis. *Crit Care*. 2014;18(1):1-15.
- Elke G, Bloos F, Wilson DC, Meybohm P. Identification of developing multiple organ failure in sepsis patients with low or moderate SOFA scores. Crit Care. 2018;22(1):1-3.
- 422. Van Vught LAV, Klouwenberg PMCK, Spitoni C, et al. Incidence, risk factors, and attributable mortality of secondary infections in the intensive care unit after admission for sepsis. *JAMA*. 2016;315(14):1469-1479.
- 423. Mukherjee S, Allen RM, Lukacs NW, Kunkel SL, Carson WF. STAT3-mediated IL-17 production by postseptic T cells exacerbates viral immunopathology of the lung. *Shock*. 2012;38(5):515-523.
- 424. Herzig D, Fang G, Toliver-Kinsky TE, Guo Y, Bohannon J, Sherwood ER. Stat1-deficient mice are resistant to cecal ligation and puncture-induced septic shock. *Shock.* 2012;38(4): 395-402.
- 425. Lentsch AB, Kato A, Davis B, Wang W, Chao C, Edwards MJ. STAT4 and STAT6 regulate systemic inflammation and protect against lethal endotoxemia. *J Clin Invest.* 2001;108(10):1475-1482.
- 426. O'Shea JJ, Holland SM, Staudt LM. JAKs and STATs in immunity, immunodeficiency, and cancer. *N Engl J Med*. 2013;368(2):161-170.
- 427. Wang LX, Zhu XM, Yao YM. Sestrin2: its potential role and regulatory mechanism in host immune response in diseases. *Front Immunol.* 2019;10(December):1-15.
- 428. Budanov AV, Karin M. p53 target genes sestrin1 and sestrin2 connect genotoxic stress and mTOR signaling. *Cell*. 2008;134(3):451-460.
- 429. Chantranupong L, Wolfson RL, Orozco JM, et al. The sestrins interact with gator2 to negatively regulate the amino-acid-sensing pathway upstream of mTORC1. *Cell Rep.* 2014;9(1):1-8.

- Lanna A, Gomes DCO, Muller-Durovic B, et al. A sestrindependent Erk-Jnk-p38 MAPK activation complex inhibits immunity during aging. *Nat Immunol*. 2017;18(3):354-363.
- 431. De Castro REV, Medeiros DNM, Prata-Barbosa A, De Magalhães-Barbosa MC. Surviving sepsis campaign international guidelines for the management of septic shock and sepsis-associated organ dysfunction in children. *Pediatr Crit Care Med*. 2020;46:924-925.
- 432. Ince C, Mayeux PR, Nguyen T, et al. The endothelium in sepsis. *Shock*. 2016;45(3):259-270.
- 433. Langley RJ, Tsalik EL, Van Velkinburgh JC, et al. Sepsis: an integrated clinico-metabolomic model improves prediction of death in sepsis. *Sci Transl Med*. 2013;5(195):195ra95.
- 434. Zhao F, Fang Y, Deng S, et al. Glycyrrhizin protects rats from sepsis by blocking HMGB1 signaling. *Biomed Res Int.* 2017;2017:9719647.
- Cornelius DC, Baik CH, Travis OK, et al. NLRP3 inflammasome activation in platelets in response to sepsis. *Physiol Rep.* 2019;7(9):e14073.
- 436. Cornelius DC, Travis OK, Tramel RW, et al. NLRP3 inflammasome inhibition attenuates sepsis-induced platelet activation and prevents multi-organ injury in cecal-ligation puncture. *PLoS One*. 2020;15(6 June):1-15.
- 437. Zeng L, Kang R, Zhu S, et al. ALK is a therapeutic target for lethal sepsis. *Sci Transl Med*. 2017;9(412):1-16.
- 438. Zhang B, Wei W, Qiu J. ALK is required for NLRP3 inflammasome activation in macrophages. *Biochem Biophys Res Commun*. 2018;501(1):246-252.
- 439. Yang J, Liu Z, Wang C, et al. Mechanism of gasdermin D recognition by inflammatory caspases and their inhibition by a gasdermin D-derived peptide inhibitor. *Proc Natl Acad Sci USA*. 2018:115(26):6792-6797.
- Haag SM, Gulen MF, Reymond L, et al. Targeting STING with covalent small-molecule inhibitors. *Nature*. 2018;559(7713):269-273.
- Jiang Y, He L, Green J, et al. Discovery of second-generation NLRP3 inflammasome inhibitors: design, synthesis, and biological characterization. *J Med Chem.* 2019;62(21):9718-9731.
- 442. Chen R, Zhu S, Zeng L, et al. AGER-mediated lipid peroxidation drives caspase-11 inflammasome activation in sepsis. *Front Immunol.* 2019;10(AUG):1904.
- 443. Nielsen TB, Pantapalangkoor P, Yan J, et al. Diabetes exacerbates infection via hyperinflammation by signaling through TLR4 and RAGE. *mBio*. 2017;8(4):1-15.
- 444. Hu JJ, Liu X, Xia S, et al. FDA-approved disulfiram inhibits pyroptosis by blocking gasdermin D pore formation. *Nat Immunol.* 2020;21(7):736-745.
- 445. Li L, He X, Wang X, et al. Ruxolitinib protects lipopolysaccharide (LPS)-induced sepsis through inhibition of nitric oxide production in mice. *Ann Transl Med.* 2020;8(8):546-546.
- 446. Jarneborn A, Mohammad M, Engdahl C, et al. Tofacitinib treatment aggravates Staphylococcus aureus septic arthritis, but attenuates sepsis and enterotoxin induced shock in mice. *Sci Rep.* 2020;10(1):1-9.
- 447. Ge Y, Wang C, Yao C, et al. STC3141 improves acute lung injury through neutralizing circulating histone in rat with experimentally-induced acute respiratory distress syndrome. *Front Pharmacol.* 2023;14:1166814.



- 448. Meara CHO, Coupland LA, Kordbacheh F, et al. Neutralizing the pathological effects of extracellular histones with small polyanions. *Nat Commun.* 2020;11(1):6408.
- 449. Geven C, Blet A, Kox M, et al. A double-blind, placebocontrolled, randomised, multicentre, proof-of-concept and dose-finding phase II clinical trial to investigate the safety, tolerability and efficacy of adrecizumab in patients with septic shock and elevated adrenomedullin concentrat. *BMJ Open*. 2019:9(2):1-11.
- 450. Pickkers P, Mehta RL, Murray PT, et al. Effect of human recombinant alkaline phosphatase on 7-day creatinine clearance in patients with sepsis-associated acute kidney injury a randomized clinical trial. *JAMA*. 2018;320(19):1998-2009
- 451. Francois B, Jeannet R, Daix T, et al. Interleukin-7 restores lymphocytes in septic shock: the IRIS-7 randomized clinical trial. *JCI Insight*. 2018;3(5):e98960.
- 452. Watanabe E, Nishida O, Kakihana Y, et al. Pharmacokinetics, pharmacodynamics, and safety of nivolumab in patients with sepsis-induced immunosuppression: a multicenter, open-label phase 1/2 study. *Shock.* 2020;53(6):686-694.
- 453. Vincent JL, Francois B, Zabolotskikh I, et al. Effect of a recombinant human soluble thrombomodulin on mortality in patients with sepsis-associated coagulopathy: the SCARLET randomized clinical trial. *JAMA*. 2019;321:1993-2002.

- 454. Bulger EM, May AK, Robinson BRH, et al. A novel immune modulator for patients with necrotizing soft tissue infections (NSTI): results of a multicenter, phase 3 randomized controlled trial of reltecimod (AB 103). *Ann Surg.* 2020;272(3):469-478.
- 455. Fowler AA, Truwit JD, Hite RD, et al. Effect of vitamin C infusion on organ failure and biomarkers of inflammation and vascular injury in patients with sepsis and severe acute respiratory failure: the CITRIS-ALI randomized clinical trial. *JAMA*. 2019;322:1261-1270.
- 456. Hotchkiss RS, Colston E, Yende S, et al. Immune check-point inhibition in sepsis: a phase 1b randomized, placebo-controlled, single ascending dose study of antiprogrammed cell death-ligand 1 antibody (BMS-936559). *Crit Care Med*. 2019;47(5):632-642.
- 457. Schlosser K, Wang JP, Dos Santos C, et al. Effects of mesenchymal stem cell treatment on systemic cytokine levels in a phase 1 dose escalation safety trial of septic shock patients. *Crit Care Med.* 2019;47(7):918-925.

How to cite this article: Zhang W, Jiang H, Wu G, et al. The pathogenesis and potential therapeutic targets in sepsis. *MedComm*. 2023;4:e418.

https://doi.org/10.1002/mco2.418