

REVIEW

Challenges of HIV diagnosis and management in the context of pre-exposure prophylaxis (PrEP), post-exposure prophylaxis (PEP), test and start and acute HIV infection: a scoping review

Tamara Elliott^{1,2} (D), Eduard J Sanders^{3,4}, Meg Doherty⁵, Thumbi Ndung'u^{6,7,8,9}, Myron Cohen¹⁰ (D), Pragna Patel¹¹, Gus Cairns^{12,13}, Sarah E Rutstein¹⁰, Jintanat Ananworanich^{14,15} (D), Colin Brown^{16,17} and Sarah Fidler^{1,18,5} (D)

[§]Corresponding author: Sarah Fidler, Faculty of Medicine, Imperial College London, Jefferiss Research Trust Laboratories, 2nd Floor, Medical School Building, St Mary's Campus, Norfolk Place, Paddington, London W2 1PG, United Kingdom. Tel +44 (0)20 7594 3910/+44 7912 205671. (s.fidler@imperial.ac.uk)

Abstract

Introduction: Knowledge of HIV status relies on accurate HIV testing, and is the first step towards access to HIV treatment and prevention programmes. Globally, HIV-status unawareness represents a significant challenge for achieving zero new HIV infections and deaths. In order to enhance knowledge of HIV status, the World Health Organisation (WHO) recommends a testing strategy that includes the use of HIV-specific antibody point-of-care tests (POCT). These POCTs do not detect acute HIV infection, the stage of disease when viral load is highest but HIV antibodies are undetectable. Complicating things further, in the presence of antiretroviral therapy (ART) for pre-exposure prophylaxis (PrEP) or post-exposure prophylaxis (PEP), other currently available testing technologies, such as viral load detection for diagnosis of acute HIV infection, may yield false-negative results. In this scoping review, we evaluate the evidence and discuss alternative HIV testing algorithms that may mitigate diagnostic dilemmas in the setting of increased utilization of ART for immediate treatment and prevention of HIV infection.

Discussion: Missed acute HIV infection prevents people living with HIV (PLHIV) from accessing early treatment, increases likelihood of onward transmission, and allows for inappropriate initiation or continuation of PrEP, which may result in HIV drug resistance. While immediate ART is recommended for all PLHIV, studies have shown that starting ART in the setting of acute HIV infection may result in a delayed or complete absence of development of HIV-specific antibodies, posing a diagnostic challenge that is particularly pertinent to resource-limited, high HIV burden settings where HIV-antibody POCTs are standard of care. Similarly, ART used as PrEP or PEP may supress HIV RNA viral load, complicating current HIV testing algorithms in resource-wealthy settings where viral detection is included. As rollout of PrEP continues, HIV testing algorithms may need to be modified

Conclusions: With increasing use of PrEP and ART in acute infection we anticipate diagnostic challenges using currently available HIV testing strategies. Research and surveillance are needed to determine the most appropriate assays and optimal testing algorithms that are accurate, affordable and sustainable.

Keywords: Acute HIV infection; pre-exposure prophylaxis; post-exposure prophylaxis; immediate antiretroviral therapy; HIV testing algorithms; indeterminate HIV test

Received 11 June 2019; Accepted 22 October 2019

Copyright © 2019 The Authors. Journal of the International AIDS Society published by John Wiley & Sons Ltd on behalf of the International AIDS Society. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

1 | INTRODUCTION

In the current era of immediate antiretroviral therapy (ART), and pre- or post-exposure prophylaxis, confidently diagnosing HIV is becoming increasingly complex. Cases of diagnostic uncertainty can be confusing and distressing to both clinicians and patients, and can lead to difficult management decisions, particularly in regard to initiation of ART for treatment or prophylaxis.

ART has dramatically improved survival for people living with HIV (PLHIV) and globally there has been an improvement in treatment coverage [1]. Viral suppression on ART confers

an individual health benefit [2,3], and a significant reduction in the risk of onward transmission [4–6], with an impact on HIV incidence at a population level [7–10]. In 2014, the Joint United Nations Programme on HIV and AIDS (UNAIDS) set the 90-90-90 targets, whereby 90% of people with HIV will know their status, 90% diagnosed will be on ART and 90% on ART will be virally supressed by 2020 [11]. By the end of 2017, globally 75% of PLHIV knew their status, but there were still 1.8 million new HIV infections [1].

Oral ART agents tenofovir disoproxil fumarate (TDF) and emtricitabine (FTC) as pre-exposure prophylaxis (PrEP) for individuals at risk of HIV are highly effective at reducing HIV acquisition when taken correctly [12–14]. In addition, the World Health Organisation (WHO) recommends one month of triple ART as post-exposure prophylaxis (PEP) following HIV exposure [15]. UNAIDS re-set targets to reduce the global numbers of new HIV infections to less than 500,000 through ensuring that 90% of people at risk of HIV infection have access to comprehensive HIV prevention services by 2020, including PEP and PrEP [16].

Access to accurate HIV testing is crucial in order to direct PLHIV to treatment programmes and those who are HIV negative to appropriate prevention strategies. Tests to determine HIV status include those that measure HIV-specific antibody and those that detect viral genetic material or proteins. Rapid HIV point-of-care tests (POCTs) are often used, particularly in resource-limited settings, and also increasingly for self-testing [17]. These tests, which rely on the detection of HIV-specific antibodies, do not diagnose acute HIV infection (AHI) [18]. AHI is defined as the period prior to the development of detectable antibodies, and can be characterized by Fiebig staging [19-21], where in Fiebig I only HIV RNA is detectable, and in Fiebig II p24 antigen becomes detectable, a transient viral core protein. AHI plays a disproportionate role in HIV transmission events, and epidemiologic, phylogenetic and mathematical modelling studies suggest that AHI may be responsible for between 10% and 50% of all HIV transmissions [22-27], varying by risk group and stage of the epidemic. Ideally, individuals at high risk or presenting with symptoms suggestive of AHI should have further diagnostic tests in addition to HIV-antibody testing, such as nucleic acid amplification testing (NAAT) and/or HIV p24 gag viral core protein. However, most of these tests require venous blood sampling, sophisticated laboratory infrastructure and advanced personnel training, which are costly, time consuming and unavailable in many settings.

This scoping review was originally based on an invited symposium entitled "Strategies for diagnosing and managing AHI in the context of PrEP and immediate ART" at the 22nd International AIDS conference, July 2018. It has since been supplemented with evidence from clinical trials, observational studies, systematic reviews and international best practice guidelines, as well as updates from a similar session "HIV testing and management in the era of PrEP" at IAS 2019. In this scoping review, we aim to consider the difficulties in confirming HIV status using current testing strategies, and the reported challenges in confirming HIV status among people receiving PrEP or PEP, or those starting immediate ART in AHI using currently approved test kits and testing algorithms.

2 | DISCUSSION

2.1 | HIV testing algorithms

WHO HIV testing guidelines recommend that specimens are first tested with the most sensitive rapid antibody POCT available. If this test is non-reactive, individuals are considered HIV negative, whereas if this test is reactive, a second distinct assay is used [28]. The United States (US) [29], European [30] and United Kingdom (UK) [31] guidelines all recommend an HIV diagnostic algorithm consisting of a laboratory-based antigen/antibody (Ag/Ab) combination immunoassay followed by a confirmatory HIV-1/HIV-2 differentiation assay if positive.

These guidelines also recognize that there are certain situations when a POCT may be recommended, such as settings where a rapid turnaround is desirable, or if venepuncture is unavailable or refused.

2.2 Diagnosis of AHI and immediate ART

Accurate HIV testing is necessary to allow timely identification of AHI and facilitate immediate ART initiation. However, identifying those with AHI is challenging, particularly since symptoms can be non-specific or absent [32]. Symptom and sexual behaviour risk scores have been validated in multiple settings across sub-Saharan Africa to direct higher risk individuals to more intensive HIV testing with HIV RNA or p24 antigen [33-35]. Targeted rather than non-selective screening in this way gives the potential for substantial cost saving. Data from Lilongwe. Malawi have shown that rates of AHI were higher in symptomatic patients presenting to sexually transmitted infection (STI) clinics (1.0% of HIV-seronegative patients) compared to HIV testing centres (0.3%)[36]. Among these patients, implementing a risk score-based screening criteria would have identified 80% of patients with AHI by only screening 50% of the presenting patient population with HIV RNA, thereby conserving scarce screening resources [36].

Point-of-care Ag/Ab diagnostics that meet the WHO ASSURED (affordable, sensitive, specific, user-friendly, rapid and robust, equipment free, delivered) criteria [37] are under development, but not widely available. These rapid tests use a lateral flow cassette to separately assay for both HIV antibodies and p24 antigen. Field studies of the US Food and Drug Administration (FDA)-approved rapid Alere Determine™ HIV-1/2 Ag/Ab Combo [38-42] have shown that antigen is rarely detected in whole blood specimens, with poor sensitivity for detection of AHI. The re-formulated AlereTM HIV Combo [43] has shown much improved sensitivity for detection of p24 antigen [44-47], however, further evaluation of its use in clinical practice is required. Other options include diagnostic platforms for RNA testing, such as the Alere™ q HIV-1/2 Detect [48] which has been validated for RNA detection among infants and children, the Cepheid GeneXpert[®], the technology for which already has an existing presence in sub-Saharan Africa where it is predominantly used for tuberculosis testing [49], and SAMBA [50] which is a dipstick-based nucleic acid assay for the detection of HIV in whole blood developed for monitoring and diagnostic use in LMIC settings.

While there are clear benefits for all PLHIV to start ART irrespective of CD4 count, there are less randomized data on the urgency of starting ART among those with AHI. Early ART can increase the chance of CD4 count recovery [51], and there are also data that very early ART in AHI may limit viral reservoir seeding and confer enhanced likelihood of posttreatment viral control of ART [52] although this remains rare. In the prospective RV254/Search010 study in Thailand, samples from clients with a non-reactive Ag/Ab immunoassay were screened for AHI by pooled NAAT [53]. Of the 112 participants with AHI (40% in Fiebig I), 111 initiated ART on the day of enrolment. The median time from HIV exposure to enrolment was 19 days [53]. In this cohort there has been a trend towards participants treated in Fiebig I having a better clinical phenotype after two years on ART compared to later in acute infection or in chronic infection [54,55]. Despite

encouraging observations in this cohort, interruption of ART among eight individuals followed for 24 weeks did not confer post-treatment viral control [56].

The FRESH study in South Africa [57] follows initially HIVnegative young women at very high risk of HIV acquisition with twice weekly HIV RNA testing. More than 1600 women have been enrolled, 71 of whom had incident HIV infection identified during the acute phase (> 70% in Fiebig I), with a median of four days since the last negative HIV RNA [58]. Fifty-seven of those with AHI initiated rapid ART within a median of one day from detection. A majority of those starting rapid ART (87%) did not develop detectable HIV-specific antibodies on western blot, and nearly half (48%) of participants were consistently Western blot antibody negative up to 340 days of follow-up [57]. These data demonstrate that in the context of rapid ART initiation during Fiebig I, antibody tests for the confirmation of HIV infection are unreliable. Similar findings were reported in Thai participants initiating ART during AHI [59,60], as well as among infants with perinatally acquired HIV receiving ART early in infection, where subsequent antibody testing remained negative at almost two years of age [61].

Understanding dissonant HIV test results in the setting of starting ART during AHI is a key community message to share with advocates and community leaders. In the era of rapid ART start there may need to be a new narrative around "testing positive" for HIV versus "being infected" or "living with" HIV. One patient from the RV254/Search010 cohort commented "although I have HIV, my blood test result is still normal just like a normal person... I still continue living my life as normal" [55,62]. There is a complex dialogue about "normal" and being HIV negative, and management of negative HIV-antibody test results for PLHIV can be difficult. The long-term implications of remaining HIV seronegative maybe far reaching and if future ART interruption studies are explored amongst these cohorts there is a potential chance of seroconversion.

2.3 HIV diagnosis in the context of PrEP and PEP

It is important to rule out AHI prior to initiating PrEP, as starting dual ART with TDF/FTC in undiagnosed AHI may not control viraemia and carries the potential for emergence of drug-resistance [63-65]. In a meta-analysis of 18 PrEP studies, six reported cases of TDF or FTC resistance, and the risk of developing resistance mutations was significantly higher in PrEP users who initiated PrEP during AHI [63]. In another review exploring HIV resistance in PrEP studies, 3% of participants infected with HIV post enrolment had TDF or FTC resistance compared to 41% of those who had undetected AHI at enrolment [66]. When the long-acting injectable cabotegravir, which is currently in development for PrEP, was given to macaques in acute infection there were high levels of integrase resistance [67]. These data highlight the importance of accurate testing to exclude AHI prior to initiation of PrEP. In some reported cases of HIV infection in the context of PrEP, resistance mutations may have been transmitted, that is, occurred secondary to exposure to resistant virus rather than selected for post transmission [68,69].

International guidelines have recognized the need to adjust HIV testing algorithms in the context of PrEP. WHO guidelines state that it is critical to rule out HIV prior to starting PrEP

and recommend a serial testing strategy which, in most LMIC, will involve a combination of POCTs. If the initial HIV serology test is negative without any clinical suggestion of AHI then PrEP can be initiated [70], and HIV testing is recommended every three months for the duration that the client remains on PrEP. If an HIV-antibody test is inconclusive then a repeat is recommended 14 days later and discontinuation of PrEP should be considered. In the Centers for Disease Control (CDC) algorithm for determining HIV status for PrEP provision, a negative HIV-antibody test (which may be an antibody POCT) with no signs or symptoms of AHI in the past four weeks means PrEP can be initiated [71]. If there is concern about AHI then a laboratory Ag/Ab assay is preferred, with alternative options of HIV RNA or to retest for antibody in one month and defer the decision on whether PrEP can be started [71]. The IAS-USA guidelines recommend an Ag/Ab assay to determine PrEP eligibility, and if there is any suspicion of AHI then an HIV RNA assay should also be sent, and a repeat Ag/Ab test should be performed in one month [72].

Once PrEP has been initiated, frequent monitoring for HIV is important, with most guidelines recommending re-testing at least every three months. This minimizes the risk of inappropriate dual therapy with TDF/FTC among PLHIV, and the subsequent risk of development of HIV drug resistance. However, it is unclear how well currently available HIV tests perform in the presence of PrEP. Case reports have detailed atypical HIV-antibody test results in those taking PrEP [73-75], with the possibility of a delayed or reduced antibody response to HIV infection and suppressed viral replication. In the ADAPT trial (HIV Prevention Trials Network (HPTN) 067) that evaluated TDF/FTC PrEP in women in South Africa and men who have sex with men (MSM) in Thailand and USA, there were 12 new HIV infections [76]. In 9 of the 12 cases, both of two POCTs were non-reactive at the first HIV-positive visit, including eight cases of AHI. Of these eight cases, five also had a negative laboratory Ag/Ab test. The viral load was ≤ 400 copies/mL in four of the eight AHI cases, but all were positive on a sensitive HIV RNA qualitative assay. In eight cases there was continuation of PrEP, including one case where PrEP was continued for three to four months post HIV infection; there were three cases of drug resistance to ART [76]. In an assessment of HIV diagnostic tests in the ANRS-IPERGAY study, the Abbott Architect® Ag/Ab Combo test detected 85% of AHI but failed to detect two early infections, both which had HIV RNA of < 500 copies/mL [77]. The BioRad Bioplex 2200 Ag/ Ab assay missed three more infections, and the antibody POCTs performed poorly in AHI [77]. In the Partners PrEP study, PrEP delayed the time to detect seroconversion but this delay was not associated with developing resistant virus [78]. Eleven percent of seroconverters in the PrEP arm had an undetectable HIV RNA compared to 3% in the placebo arm [78]. Overall, interpretation of HIV RNA and antibody test results can be more challenging in the context of PrEP, and confirming or ruling out a diagnosis can be difficult [79].

At the screening visit in HPTN PrEP studies, a combination of a POCT, instrumented Ag/Ab test, and an HIV RNA test are recommended for HIV diagnosis [80]. At all follow-up visits, one or two POCTs are performed as well as an instrumented Ag/Ab test. In any cases of discrepant results, confirmatory samples are taken and additionally tested for proviral HIV DNA from cell pellet samples. If needed,

additional testing is performed at a third visit after a four-week product hold to help determine HIV status [80]. Presently, total HIV nucleic acid and proviral DNA testing are used only in research algorithms and are not feasible or available in routine care.

In large ongoing PrEP trials including the use of long-acting injectables as PrEP (HPTN 083 [81] and HPTN 084 [82]), broadly neutralizing antibodies [83] or vaccines [84], we would expect to see similar diagnostic challenges. These trials will help inform appropriate HIV testing algorithms pre- and post-PrEP initiation, as well as preferred HIV diagnostic platforms and management decisions in cases of diagnostic uncertainty.

In situations where there are indeterminate results, Smith et al. have proposed three potential management options [85,86]. The first is to continue PrEP while confirming HIV status. This has the advantage of ongoing protection against HIV infection if negative, but would be inadequate therapy if HIV positive, with a risk of drug resistance. The second option is to discontinue PrEP to aid diagnosis with recommendation to use condoms, however, this may put HIV-negative individuals at risk of acquiring HIV. The final option would be to intensify PrEP to triple therapy while performing additional tests to confirm status. If HIV positive, this would allow for the benefits of immediate ART, but would be unnecessarily exposing the individual to a third agent if HIV negative, and may make it more difficult to confirm or rule out HIV. Similar considerations on whether to continue or discontinue ART may be needed in those taking PEP with ambiguous HIV testing results.

HIV testing in the context of PrEP is particularly challenging in LMIC settings where there is limited access to advanced laboratory tests and infrastructure. In these settings sensitive and affordable rapid tests to detect acute infection are desperately needed to supplement current WHO guidelines, which only require an antibody POCT to rule out HIV [70]. Additionally, relying on antibody tests to detect HIV during PrEP follow-up is also likely to be inadequate. Cases of HIV drug resistance either occurring as a result of PrEP initiation in missed AHI, or due to the increasing rates of transmitted ART resistance in LMIC [87], could be particularly problematic in these settings as ART choice is likely to be limited, and alternative therapy options may not be available.

3 | CONCLUSIONS

We can anticipate that with increasing use of PEP, PrEP and immediate ART, the currently available HIV diagnostic tests may be inadequate to reliably make clinical assessments and treatment decisions. Current routine rapid POCTs are unable to detect AHI, and some individuals starting ART in AHI may never have detectable antibody with these tests. There is no one easy solution to this, especially in resource-poor settings where there is limited choice and availability of diagnostic tests, and the costs of implementing widespread additional HIV testing maybe unsustainable. Optimal testing algorithms to allow safe use of ART for prevention still need to be established, and future guidelines are likely to incorporate the use of sensitive rapid tests to detect AHI, including whole blood nucleic acid assays or rapid Ag/Ab tests. Information gained from ongoing large-scale HIV prevention

studies will provide insights into solutions for these diagnostic challenges.

AUTHORS' AFFILIATIONS

¹Imperial College London, London, United Kingdom; ²Imperial College Healthcare NHS Trust, London, United Kingdom; ³Kenya Medical Research Institute-Wellcome Trust Research Programme, Kilifi, Kenya; ⁴Nuffield Department of Medicine, University of Oxford, Oxford, United Kingdom; ⁵Department of HIV and Global Hepatitis Programme, WHO, Geneva, Switzerland; ⁶Africa Health Research Institute, Durban, South Africa; ⁷HIV Pathogenesis Programme, Doris Duke Medical Research Institute, University of KwaZulu-Natal, Durban, South Africa; ⁸The Ragon Institute of Massachusetts General Hospital, Massachusetts Institute of Technology and Harvard University, Cambridge, MA, USA; ⁹Max Planck Institute for Infection Biology, Berlin, Germany; ¹⁰Department of Internal Medicine, Division of Infectious Diseases, UNC School of Medicine, University of North Carolina At Chapel Hill, Chapel Hill, NC, USA; 11 Division of Global HIV and TB, Centers for Disease Control and Prevention, Atlanta, GA, USA; ¹²NAM Aidsmap, London, United Kingdom; ¹³PrEP in Europe Initiative, London, United Kingdom; ¹⁴U.S. Military HIV Research Program, Walter Reed Army Institute of Research, Silver Spring, MD, USA; ¹⁵Henry M. Jackson Foundation for the Advancement of Military Medicine, Bethesda, MD, USA; ¹⁶National Infection Service, Public Health England, London, United Kingdom; ¹⁷Department of Infection, Royal Free London NHS Foundation Trust, London, United Kingdom; ¹⁸Imperial College NIHR BRC, London, United Kingdom

COMPETING INTERESTS

None: TE, EJS, MD, TN, MC, PP, GC, SER, JA, CB, SF.

AUTHORS' CONTRIBUTIONS

All authors contributed to the content and preparation of the manuscript, and approved the final draft. Specific contributions: TE drafted the manuscript, and performed a literature review; EJS, JA and CB provided expert opinion and revision of the manuscript for important content, MD provided expert opinion, specifically on WHO guidelines; TN and SER provided expert opinion, preparation and revision of the manuscript, specifically on immediate ART in AHI and experience in LMIC; MC provided expert opinion and revision of the manuscript, including in regards to HPTN study protocols; PP provided expert opinion and revision of the manuscript, specifically in regards to CDC guidelines; GC revised the manuscript with inclusion of patient perspectives; SF contributed to the preparation and revision of the manuscript, and provided expert opinion.

ACKNOWLEDGEMENTS

EJS receives research funding from International AIDS Vaccine Initiative, NIH (grant R01AI124968), and the Wellcome Trust. SF receives support from Imperial College NIHR BRC. This work was also supported through the Sub-Saharan African Network for TB/HIV Research Excellence (SANTHE), a DELTAS Africa Initiative [grant # DEL-15-006]. The DELTAS Africa Initiative is an independent funding scheme of the African Academy of Sciences (AAS)'s Alliance for Accelerating Excellence in Science in Africa (AESA) and supported by the New Partnership for Africa's Development Planning and Coordinating Agency (NEPAD Agency) with funding from the Wellcome Trust [grant # 107752/Z/15/Z] and the UK government. This summary has been supported by the President's Emergency Plan for AIDS Relief (PEPFAR) through the Centers for Disease Control and Prevention (CDC) by participation of subject matter experts. The views expressed in this publication are those of the author(s) and not necessarily those of AAS, NEPAD Agency, Wellcome Trust, UK government, USAID, NIH, CDC or the US Government. This report was published with permission from Kenya Medical Research Institute.

FUNDING

No funding was received for the preparation of this manuscript.

DISCLAIMER

The findings and conclusions in this report are those of the author(s) and do not necessarily represent the official position of the funding agencies. Use of trade names and commercial sources is for identification only and does not constitute endorsement by the Centers for Disease Control and Prevention.

REFERENCES

- 1. UNAIDS. Fact sheet Latest global and regional statistics on the status of the AIDS epidemic, World AIDS Day. [Internet]. [cited 2019 Mar 5]. 2018. Available from: http://www.unaids.org/sites/default/files/media_asset/UNAIDS_FactSheet_en.pdf
- 2. The INSIGHT START Study Group. Initiation of antiretroviral therapy in early asymptomatic HIV infection. N Engl J Med. 2015;373(9):795–807.
- 3. The TEMPRANO ANRA 12136 Study Group. A Trial of Early Antiretrovirals and Isoniazid Preventive Therapy in Africa. N Engl J Med. 2015;373(9):808–22.
- 4. Cohen MS, Chen YQ, McCauley M, Gamble T, Hosseinipour MC, Kumarasamy N, et al. Prevention of HIV-1 Infection With Early Antiretroviral Therapy. N Engl J Med. 2011;365(6):493–505.
- 5. Rodger AJ, Cambiano V, Bruun T, Vernazza P, Collins S, van Lunzen J, et al. Sexual activity without condoms and risk of HIV transmission in serodifferent couples when the HIV-positive partner is using suppressive antiretroviral therapy. JAMA. 2016;316(2):171.
- 6. Rodger AJ, Cambiano V, Bruun T, Vernazza P, Collins S, Degen O, et al. Risk of HIV transmission through condomless sex in MSM couples with suppressive ART: the PARTNER2 Study extended results in gay men. AIDS. 2018, 23–27 July 2018, Amsterdam. Late breaker oral abstract WEAX0104LB. [Internet]. [cited 2019 Mar 19]. Available from: http://programme.aids2018.org/Abstract/Abstract/13470
- 7. Granich RM, Gilks CF, Dye C, De Cock KM, Williams BG. Universal voluntary HIV testing with immediate antiretroviral therapy as a strategy for elimination of HIV transmission: a mathematical model. Lancet (London, England). 2009;373 (9657):48–57.
- 8. Eaton JW, Johnson LF, Salomon JA, Bärnighausen T, Bendavid E, Bershteyn A, et al. HIV treatment as prevention: systematic comparison of mathematical models of the potential impact of antiretroviral therapy on HIV incidence in South Africa. PLoS Med. 2012;9:e1001245.
- 9. Tanser F, Vandormael A, Cuadros D, Phillips AN, de Oliveira T, Tomita A, et al. Effect of population viral load on prospective HIV incidence in a hyperendemic rural African community. Sci Transl Med. 2017;9(420):eaam8012.
- 10. Hayes RJ, Donnell D, Floyd S, Mandla N, Bwalya J, Sabapathy K, et al. Effect of Universal Testing and Treatment on HIV Incidence HPTN 071 (PopART), N Engl J Med. 2019;381(3):207–18.
- 11. UNAIDS. 90–90-90: An ambitious treatment target to help end the AIDS epidemic [Internet]. 2014 [cited 2019 Mar 5], Available from: http://www.unaids.org/sites/default/files/media_asset/90-90-90_en.pdf
- 12. McCormack S, Dunn DT, Desai M, Dolling DI, Gafos M, Gilson R, et al. Pre-exposure prophylaxis to prevent the acquisition of HIV-1 infection (PROUD): effectiveness results from the pilot phase of a pragmatic open-label randomised trial. Lancet. 2016;387(10013):53–60.
- 13. Grant RM, Lama JR, Anderson PL, McMahan V, Liu AY, Vargas L, et al. Pre-exposure chemoprophylaxis for HIV prevention in men who have sex with men. N Engl J Med. 2010;363(27):2587–99.
- 14. Molina J-M, Capitant C, Spire B, Pialoux G, Cotte L, Charreau I, et al. On-Demand preexposure prophylaxis in men at high risk for HIV-1 infection. N Engl J Med. 2015;373(23):2237–46.
- 15. WHO. Guidelines on post-exposure prophylaxis for HIV and the use of cotrimoxazole prophylaxis for HIV-related infections among adults, adolescents and children: recommendations for a public health approach. Geneva: World Health Organisation: 2014.
- 16. UNAIDS. HIV Prevention 2020 Road Map Accelerating HIV prevention to reduce new infections by 75% [Internet]. [cited 2019 Mar 5]. Available from: http://www.unaids.org/sites/default/files/media_asset/hiv-prevention-2020-road-map_en.pdf
- 17. Unitaid. HIV rapid diagnostic tests for self-testing, 4th edition, July 2018 [Internet]. [cited 2019 May 3]. Available from: https://unitaid.org/assets/HIVST-landscape-report.pdf
- 18. Patel P, Bennett B, Sullivan T, Parker MM, Heffelfinger JD, Sullivan PS, et al. Rapid HIV screening: missed opportunities for HIV diagnosis and prevention. J Clin Virol. 2012;54(1):42–7.
- 19. Fiebig EW, Wright DJ, Rawal BD, Garrett PE, Schumacher RT, Peddada L, et al. Dynamics of HIV viremia and antibody seroconversion in plasma donors: implications for diagnosis and staging of primary HIV infection. AIDS. 2003;17 (13):1871–9
- 20. McMichael AJ, Borrow P, Tomaras GD, Goonetilleke N, Haynes BF. The immune response during acute HIV-1 infection: clues for vaccine development. Nat Rev Immunol. 2010;10(1):11–23.
- 21. Cohen MS, Gay CL, Busch MP, Hecht FM. The detection of acute HIV infection. J Infect Dis. 2010;202(S2):S270–7.
- 22. Powers KA, Ghani AC, Miller WC, Hoffman IF, Pettifor AE, Kamanga G, et al. The role of acute and early HIV infection in the spread of HIV and

- implications for transmission prevention strategies in Lilongwe, Malawi: a modelling study. Lancet2011;378(9787):256–68.
- 23. Volz EM, Ionides E, Romero-Severson EO, Brandt M-G, Mokotoff E, Koopman JS. HIV-1 transmission during early infection in men who have sex with men: a phylodynamic analysis. Hallett TB, editor. PLoS Med. 2013;10:e1001568; discussion e1001568.
- 24. Bellan SE, Dushoff J, Galvani AP, Meyers LA. Reassessment of HIV-1 acute phase infectivity: accounting for heterogeneity and study design with simulated cohorts. Newell M-L, editor. PLoS Med. 2015;12:e1001801.
- 25. Hollingsworth TD, Pilcher CD, Hecht FM, Deeks SG, Fraser C. High transmissibility during early HIV infection among men who have sex with men-San Francisco, California. J Infect Dis. 2015;211(11):1757–60.
- 26. Hollingsworth TD, Anderson RM, Fraser C. HIV-1 transmission, by stage of infection. J Infect Dis. 2008;198(5):687–93.
- 27. Wawer MJ, Gray RH, Sewankambo NK, Serwadda D, Li X, Laeyendecker O, et al. Rates of HIV-1 transmission per coital act, by stage of HIV-1 infection, in Rakai, Uganda. J Infect Dis. 2005;191(9):1403–9.
- 28. WHO. Consolidated guidelines on HIV testing services. Geneva: World Health Organisation; 2015.
- 29. Branson BM, Owen SM, Wesolowski LG, Bennett B, Werner BG, Wroblewski KE,, et al. Laboratory testing for the diagnosis of HIV infection: updated recommendations. Atlanta, GA: 2014.
- 30. Gökengin D, Geretti AM, Begovac J, Palfreeman A, Stevanovic M, Tarasenko O, et al. 2014 European guideline on HIV testing. Int J STD AIDS. 2014;25 (10):695–704.
- 31. BHIVA. UK National Guidelines for HIV Testing 2008 [Internet]. 2008 [cited 2019 Mar 24]. Available from: www.bhiva.org
- 32. Robb ML, Eller LA, Kibuuka H, Rono K, Maganga L, Nitayaphan S, et al. Prospective Study Of Acute HIV-1 infection in adults in East Africa and Thailand. N Engl J Med. 2016;374(22):2120–30.
- 33. Powers KA, Miller WC, Pilcher CD, Mapanje C, Martinson FEA, Fiscus SA, et al. Improved detection of acute HIV-1 infection in sub-Saharan Africa: development of a risk score algorithm. AIDS. 2007;21(16):2237–42.
- 34. Sanders EJ, Wahome E, Powers KA, Werner L, Fegan G, Lavreys L, et al. Targeted screening of at-risk adults for acute HIV-1 infection in sub-Saharan Africa. AIDS. 2015;29:S221–30.
- 35. Rutstein SE, Ananworanich J, Fidler S, Johnson C, Sanders EJ, Sued O, et al. Clinical and public health implications of acute and early HIV detection and treatment: a scoping review. J Int AIDS Soc. 2017;20(1):21579.
- 36. Rutstein SE, Pettifor AE, Phiri S, Kamanga G, Hoffman IF, Hosseinipour MC, et al. Incorporating acute HIV screening into routine HIV testing at sexually transmitted infection clinics, and HIV testing and counseling centers in Lilongwe, Malawi. J Acquir Immune Defic Syndr. 2016;71(3):272–80.
- 37. Wu G, Zaman MH. Low-cost tools for diagnosing and monitoring HIV infection in low-resource settings. Bull World Health Organ. 2012;90 (12):914–20.
- 38. Conway DP, Holt M, McNulty A, Couldwell DL, Smith DE, Davies SC, et al. Multi-centre evaluation of the determine hiv combo assay when used for point of care testing in a high risk clinic-based population. Abrams WR, editor. PLoS ONE. 2014;9:e94062.
- 39. Duong YT, Mavengere Y, Patel H, Moore C, Manjengwa J, Sibandze D, et al. Performance of the determine HIV-1/2 Ag/Ab combo fourth-generation rapid test for detection of acute infections in a national household survey in Swaziland. J Clin Microbiol. 2014;52(10):3743–8.
- 40. Jones CB, Kuldanek K, Muir D, Phekoo K, Black A, Sacks R, et al. Clinical Evaluation of the Determine HIV-1/2 Ag/Ab Combo test. J Infect Dis. 2012;206 (12):1947–9.
- 41. Lewis JM, Macpherson P, Adams ER, Ochodo E, Sands A, Taegtmeyer M. Field accuracy of fourth-generation rapid diagnostic tests for acute HIV-1: a systematic review. AIDS. 2015;29(18):2465–71.
- 42. Rosenberg NE, Kamanga G, Phiri S, Nsona D, Pettifor A, Rutstein SE, et al. Detection of Acute hiv infection: a field evaluation of the determine® HIV-1/2 Ag/Ab combo test. J Infect Dis. 2012;205(4):528–34.
- 43. Abbott Rapid Diagnostics. Alere HIV Combo [Internet]. 2018. [cited 2019 Sep 27]. Available from: https://www.alere.com/en/home/product-details/alere-hiv-combo.html
- 44. Fitzgerald N, Cross M, O'Shea S, Fox J. Diagnosing acute HIV infection at point of care: a retrospective analysis of the sensitivity and specificity of a fourth-generation point-of-care test for detection of HIV core protein p24. Sex Transm Infect. 2017;93(2):100–1.
- 45. van Tienen C, Rugebregt S, Scherbeijn S, Götz H. Geurts van Kessel C. The performance of the Alere HIV combo point-of-care test on stored serum samples; useful for detection of early HIV-1 infections? Sex Transm Infect. 2018;94 (5):331–3.

- 46. Livant E, Heaps A, Kelly C, Maharaj R, Samsunder N, Nhlangulela L, et al. The fourth generation AlereTM HIV Combo rapid test improves detection of acute infection in MTN-003 (VOICE) samples. J Clin Virol. 2017;94:15–21.
- 47. Ottiger C, Huber AR. Comparison of the new alere HIV combo with alere determine HIV-1/2 Ag/Ab combo in acute primo and established HIV infections. Ann Clin Lab Res. 2015;3(3):100022.
- 48. Jani IV, Meggi B, Vubil A, Sitoe NE, Bhatt N, Tobaiwa O, et al. Evaluation of the whole-blood alere Q NAT point-of-care RNA assay for HIV-1 viral load monitoring in a primary health care setting in Mozambique. J Clin Microbiol. 2016;54(8):2104–8.
- 49. Steingart KR, Schiller I, Horne DJ, Pai M, Boehme CC, Dendukuri N. Xpert[®] MTB/RIF assay for pulmonary tuberculosis and rifampicin resistance in adults. Cochrane Database Syst Rev. 2014:CD009593.
- 50. Ondiek J, Namukaya Z, Mtapuri-Zinyowera S, Balkan S, Elbireer A, Ushiro Lumb I, et al. Multicountry Validation of SAMBA A Novel Molecular Point-of-Care Test for HIV-1 Detection in Resource-Limited Setting. J Acquir Immune Defic Syndr. 2017;76(2):e52–7.
- 51. Le T, Wright EJ, Smith DM, He W, Catano G, Okulicz JF, et al. Enhanced CD4+ T-cell recovery with earlier HIV-1 antiretroviral therapy. N Engl J Med. 2013;368(3):218–30.
- 52. Sáez-Cirión A, Bacchus C, Hocqueloux L, Avettand-Fenoel V, Girault I, Lecuroux C, et al. Post-Treatment HIV-1 controllers with a long-term virological remission after the interruption of early initiated antiretroviral therapy ANRS VISCONTI study. Lifson J, editor. PLoS Pathog. 2013;14;9:e1003211.
- 53. De Souza MS, Phanuphak N, Pinyakorn S, Trichavaroj R, Pattanachaiwit S, Chomchey N, et al. Impact of nucleic acid testing relative to antigen/antibody combination immunoassay on the detection of acute HIV infection. AIDS. 2015;29(7):793–800
- 54. Ananworanich J, Pinyakorn S, Avihingsanon A, Sophonphan J, Sacdalan C, Kroon E, et al.Poster discussion abstract: Favorable clinical phenotype reached in less than half of people treated in acute HIV infection [Internet]. 2018 [cited 2019 Sep 27]. Available from: https://programme.aids2018.org/Abstract/Abstract/9868 55. Ananworanich J.Plenary session: Early treatment: what is early enough for individual benefit? Strategies for diagnosing and managing acute HIV infection in the context of PrEP and immediate ART The 22nd International AIDS conference. Amsterdam, the Netherlands, 23–27.
- 56. Colby DJ, Trautmann L, Pinyakorn S, Leyre L, Pagliuzza A, Kroon E, et al. Rapid HIV RNA rebound after antiretroviral treatment interruption in persons durably suppressed in Fiebig I acute HIV infection. Nat Med. 2018;24(7):923–6. 57. Dong KL, Moodley A, Kwon DS, Ghebremichael MS, Dong M, Ismail N, et al. Detection and treatment of Fiebig stage I HIV-1 infection in young at-risk women in South Africa: a prospective cohort study. lancet. HIV. 2018;5(1):e35–44.
- 58. Ndung'u T. Plenary session: Acute HIV infection and HIV cure. Strategies for diagnosing and managing acute HIV infection in the context of PrEP and immediate ART The 22nd International AIDS conference. Amsterdam, the Netherlands. 23-27 July 2018.
- 59. de Souza MS, Pinyakorn S, Akapirat S, Pattanachaiwit S, Fletcher JLK, Chomchey N, et al. Initiation of antiretroviral therapy during acute HIV-1 infection leads to a high rate of nonreactive HIV serology. Clin Infect Dis. 2016;63 (4):555–61.
- 60. Manak MM, Jagodzinski LL, Shutt A, Malia JA, Leos M, Ouellette J, et al. Decreased seroreactivity in individuals initiating antiretroviral therapy during acute HIV. Infection. 2019;57(10):19.
- 61. Payne H, Mkhize N, Otwombe K, Lewis J, Panchia R, Callard R, et al. Reactivity of routine HIV antibody tests in children who initiated antiretroviral therapy in early infancy as part of the Children with HIV Early Antiretroviral Therapy (CHER) trial: a retrospective analysis. Lancet Infect Dis. 2015;15 (7):803–9.
- 62. Henderson GE, Peay HL, Kroon E, Cadigan RJ, Meagher K, Jupimai T, et al. Ethics of treatment interruption trials in HIV cure research: addressing the conundrum of risk/benefit assessment. J Med Ethics. 2018;44(4):270–6.
- 63. Fonner VA, Dalglish SL, Kennedy CE, Baggaley R, O'Reilly KR, Koechlin FM, et al. Effectiveness and safety of oral HIV preexposure prophylaxis for all populations. AIDS. 2016;30(12):1973–83.
- 64. Thigpen MC, Kebaabetswe PM, Paxton LA, Smith DK, Rose CE, Segolodi TM, et al. Antiretroviral preexposure prophylaxis for heterosexual HIV transmission in Botswana. N Engl J Med. 2012;367(5):423–34.
- 65. Van Damme L, Corneli A, Ahmed K, Agot K, Lombaard J, Kapiga S, et al. Preexposure prophylaxis for HIV infection among African women. N Engl J Med. 2012;367(5):411–22.

- 66. Parikh UM, Mellors JW. Should we fear resistance from tenofovir/emtricitabine preexposure prophylaxis? Curr Opin HIV AIDS. 2016;11(1):49–55.
- 67. Radzio-Basu J, Council O, Cong M-E, Ruone S, Newton A, Wei X, et al. Drug resistance emergence in macaques administered cabotegravir long-acting for pre-exposure prophylaxis during acute SHIV infection. Nat Commun. 2019;10(1):2005.
- 68. Knox DC, Anderson PL, Harrigan PR, Tan DHS. Multidrug-Resistant HIV-1 infection despite preexposure prophylaxis. N Engl J Med. 2017;376(5):501–2.
- 69. Markowitz M, Grossman H, Anderson PL, Grant R, Gandhi M, Horng H, et al. Newly acquired infection with multidrug-resistant HIV-1 in a patient adherent to preexposure prophylaxis. J Acquir Immune Defic Syndr. 2017;76(4): e104–6.
- 70. WHO. WHO implementation tool for pre-expsure prophylaxis (PrEP) of HIV infection. Module 10: testing providers, July 2017.
- 71. CDC. Pre-exposure prophylaxis for the prevention of HIV infection in the United States 2017 update. A clinical practice guideline [Internet]. [cited 2019 Mar 24]. Available from: https://www.cdc.gov/std/tg2015/tg-2015
- 72. Saag MS, Benson CA, Gandhi RT, Hoy JF, Landovitz RJ, Mugavero MJ, et al. Antiretroviral drugs for treatment and prevention of HIV infection in adults. JAMA. 2018;320(4):379.
- 73. Hoornenborg E, Prins M, Achterbergh RCA, Woittiez LR, Cornelissen M, Jurriaans S, et al. Acquisition of wild-type HIV-1 infection in a patient on pre-exposure prophylaxis with high intracellular concentrations of tenofovir diphosphate: a case report. Lancet HIV. 2017;4(11):e522–8.
- 74. Colby DJ, Kroon E, Sacdalan C, Gandhi M, Grant RM, Phanuphak P, et al. Acquisition of multidrug-resistant human immunodeficiency virus type 1 infection in a patient taking preexposure prophylaxis. Clin Infect Dis. 2018;67 (6):962–4.
- 75. Zucker J, Carnevale C, Rai AJ, Gordon P, Sobieszczyk ME. Positive or not, that is the question: HIV testing for individuals on pre-exposure prophylaxis. J Acquir Immune Defic Syndr. 2018;78(2):e11–3.
- 76. Sivay MV, Li M, Piwowar-Manning E, Zhang Y, Hudelson SE, Marzinke MA, et al. Characterization of HIV seroconverters in a TDF/FTC PrEP study. J Acquir Immune Defic Syndr. 2017;75(3):271–9.
- 77. Delaugerre C, Antoni G, Mahjoub N, Pialoux G, Cua E, Pasquet A, et al. Assessment of HIV Screening Tests for Use in Preexposure Prophylaxis Programs. J Infect Dis. 2017;216(3):382-6.
- 78. Donnell D, Ramos E, Celum C, Baeten J, Dragavon J, Tappero J, et al. The effect of oral preexposure prophylaxis on the progression of HIV-1 seroconversion. AIDS. 2017;31(14):2007–16.
- 79. Stekler JD, Violette LR, Niemann L, McMahan VM, Katz DA, Baeten JM, et al. Repeated false-positive HIV test results in a patient taking HIV pre-exposure prophylaxis. Open Forum Infect Dis. 2018;5(9):ofy197.
- 80. Landovitz RJ, Grinsztejn B. HPTN 083 protocol v2.0. A phase 2b/3 double blind safety and efficacy study of injectable cabotegravir compared to daily oral tenofovir disoproxil fumarate/emtricitabine (TDF/FTC), For pre-exposure prophylaxis in HIV-uninfected cisgender men and transgend [Internet]. 2018 [cited 2019 Nov 18]. Available from: https://www.hptn.org/sites/default/files/inlinefile s/HPTN083_FinalVersion2.0_25July2018.pdf
- 81. The HIV Prevention Trials Network.Prevention Now. HPTN 083 [Internet]. [cited 2019 May 4]. Available from: https://www.hptn.org/research/studies/hpt n083
- 82. The HIV Prevention Trials Network.Prevention Now. HPTN 084 [Internet]. [cited 2019 May 4]. Available from: https://www.hptn.org/research/studies/hpt n084
- 83. The HIV Prevention Trials Network. Prevention Now. HVTN 704/HPTN 085 [Internet]. [cited 2019 May 4]. Available from: https://www.hptn.org/research/studies/704 085
- 84. PrEPVacc. AVAC [Internet]. [cited 2019 May 4]. Available from: https://www.avac.org/trial/prepvacc
- 85. Smith DK, Switzer WM, Peters P, Delaney KP, Granade TC, Masciotra S, et al. A strategy for PrEP clinicians to manage ambiguous HIV test results during follow-up visits. Open Forum. Infect Dis. 2018;5(8):ofy180.
- 86. Jean-Michel M. Tough questions in antiretroviral therapy management. PrEP Failures: Diagnosis, resistance and treatment CROI 2019 [Internet]. [cited 2019 Mar 24]. Available from: http://www.croiwebcasts.org/console/player/41366?mediaType=audio&&crd_fl=1&ssmsrq=1553452684080&ctms=5000&csmsrq=943
- 87. Hamers RL, Rinke de Wit TF, Holmes CB. HIV drug resistance in low-income and middle-income countries. Lancet HIV. 2018;5(10):e588–96.