

RESEARCH ARTICLE

High *SLC4A11* expression is an independent predictor for poor overall survival in grade 3/4 serous ovarian cancer

Lianzhi Qin¹, Ting Li², Yuhua Liu^{3*}

1 Delivery room, Linyi Central Hospital, Linyi, Shandong, China, **2** Department of Gynecology, Zoucheng People's Hospital, Zoucheng, Shandong, China, **3** Department of Obstetrics, Anqiu People's Hospital, Anqiu, Shandong, China

* yuhualiu1@foxmail.com



Abstract

In this study, we aimed to examine the expression of *SLC4A11* in ovarian cancer and in normal ovarian tissues, its prognostic value and the possible mechanism of its dysregulation. Bioinformatic analysis was performed by using data from the GEO datasets, the Cancer Genome Atlas-Ovarian Cancer (TCGA-OV) and the Human Protein Atlas (HPA). Results showed that *SLC4A11* was upregulated in ovarian cancer compared with normal ovarian epithelial tissues. In patients with primary serous ovarian cancer in TCGA-OV, the cases with lymphatic invasion (N = 133) had significantly higher *SLC4A11* expression than those without lymphatic invasion (N = 77) ($p = 0.0069$). High *SLC4A11* expression was consistently associated with worse overall survival (OS). Univariate and multivariate analysis confirmed that high *SLC4A11* expression was an independent prognostic factor for poor OS in grade 3/4 (G3/G4) tumors (HR = 1.416, 95%CI: 1.098–1.824, $p = 0.007$). 320 out of 578 (55.4%) ovarian cancer cases had *SLC4A11* amplification. High methylation group had a significantly lower level of *SLC4A11* expression. Based on these findings, we infer that high *SLC4A11* expression is an independent predictor for poor OS in grade 3/4 serous ovarian cancer. Both DNA amplification and hypomethylation contribute to its upregulation in ovarian cancer.

OPEN ACCESS

Citation: Qin L, Li T, Liu Y (2017) High *SLC4A11* expression is an independent predictor for poor overall survival in grade 3/4 serous ovarian cancer. PLoS ONE 12(11): e0187385. <https://doi.org/10.1371/journal.pone.0187385>

Editor: Hiromu Suzuki, Sapporo Ika Daigaku, JAPAN

Received: September 21, 2017

Accepted: October 18, 2017

Published: November 1, 2017

Copyright: © 2017 Qin et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: The authors received no specific funding for this work.

Competing interests: The authors have declared that no competing interests exist.

Introduction

Intracellular and extracellular pH (pH_i and pH_e) homeostasis is an important constitution of cellular microenvironment and is a prerequisite for normal cell function [1]. Studies in the past decade revealed that pH_i homeostasis is often dramatically altered in cancer [2, 3]. Cancer cells usually keep a pH_i that is equal to or even more alkaline than the surrounding normal counterparts, suggesting that they upregulate net acid extrusion [2, 3]. This alteration leads to at least two fundamental differences to normal physiology: firstly, it is supportive to maintain pH_i homeostasis by eliminating excessive production of acid equivalents in cancer cells due to

hypoxia and/or oncogene-induced changes in glycolytic metabolism [4]; secondly, prolonged extracellular acidification (pH_e), which favors tumor invasion and metastasis [5, 6].

Solute linked cotransporter 4 (SLC4) family is comprised of ten members (SLC4A1-5; SLC4A7-11), which have critical roles in pHi buffering [7, 8]. SLC4A1-3 are $\text{Cl}^-/\text{HCO}_3^-$ exchangers and function as cellular acid loaders [9]. SLC4A4, -4A5, -4A7, -4A8 and -4A10 are Na^+ -coupled HCO_3^- transporters and function as cellular acid extruders [9]. SLC4A11 is the most divergent member of this family and has recently been characterized as a Na^+/OH^- and NH_4^+ transporter [10]. Therefore, SLC4A11 can mediate H^+ efflux and can be considered as a cellular acid extruder. Dysregulated SLC4 family members have been implied in pathological development of some cancers. For example, SLC4A7 regulates pHi and tumor cell progression in breast cancers [11, 12]. Upregulated *SLC4A4* contributes to growth and migration of colon and breast cancer cells [13]. *SLC4A9* disruption by either genetic or pharmaceutical approaches results in pHi acidification and reduced cell growth of breast cancer and glioma cells [14].

Ovarian cancer cells have an elevated H^+ efflux compared with non-tumor cells [15]. One recent study found that the basal pHi is higher in ovarian cancer A2780 cells than in normal ovarian HOSE cells [16]. *SLC9A1* amplification is a mechanism of the high basal pHi and is associated with unfavorable overall patient survival [16]. These findings suggest that pHi regulation is closely related to ovarian cancer cell behaviors and prognosis of the patients.

In this study, we examined the expression of *SLC4A11* in ovarian cancer/normal ovarian tissues and further assessed its prognostic value and the possible mechanism of its dysregulation.

Materials and methods

Bioinformatic data mining in GEO

The normalized data of one previous microarray (GDS3592) [17] that explored the dysregulated genes in ovarian cancer epithelial cells (CEPIs) from patients with primary serous ovarian cancer compared with normal ovarian surface epithelia (OSE) was downloaded from GEO dataset for secondary analysis.

Data mining in the Human Protein Atlas

SLC4A11 expression at the protein level in normal ovarian tissues and in serous ovarian cancer tissues was compared by using the immunohistochemistry (IHC) staining data provided by the Human Protein Atlas (HPA) (<http://www.proteinatlas.org/>) [18, 19].

Bioinformatic analysis using data from TCGA-Ovarian Cancer (TCGA-OV)

SLC4A11 expression, its copy number alteration and its DNA methylation in patients with ovarian cancer were examined using data from TCGA-OV. Original data downloaded from the UCSC Xena Browser (<https://xenabrowser.net/>) were given in S1 Table. The association between *SLC4A11* expression and overall survival (OS) or recurrence-free survival (RFS) was assessed by generating Kaplan-Meier survival curves, with median *SLC4A11* expression as the cutoff. The analysis was performed using the UCSC Xena Browser or by using GraphPad Prism 6.0. Among 595 patients with primary serous ovarian cancer in TCGA-OV, 540 patients had *SLC4A11* expression measured by AgilentG4502A_07_3. This proportion of patients was included in the univariate and multivariate analysis of the association between *SLC4A11* expression and OS/RFS.

Statistical analysis

The association between clinicopathological characteristics and *SLC4A11* expression was assessed by using χ^2 tests. The significance of the difference between the survival curves was assessed by log-rank test. Univariate and multivariate Cox regression models were used to evaluate prognostic significance. Welch's t-test was conducted to compare *SLC4A11* expression between patients with or without lymphatic invasion and between groups with high/low methylation. $p < 0.05$ was considered statistically significant.

Results

SLC4A11 is upregulated in ovarian cancer compared with normal ovarian epithelial tissues

By re-analysis of the normalized data of GDS3592, we examined the expression of SLC4A family members in CEPIs and OSE (Fig 1A). *SLC4A11* was the most significantly upregulated gene among the 10 SLC4A family members (Fig 1A, red arrow). The bar chart of the array signaling value further indicated that *SLC4A11* was significantly upregulated in the CEPI samples

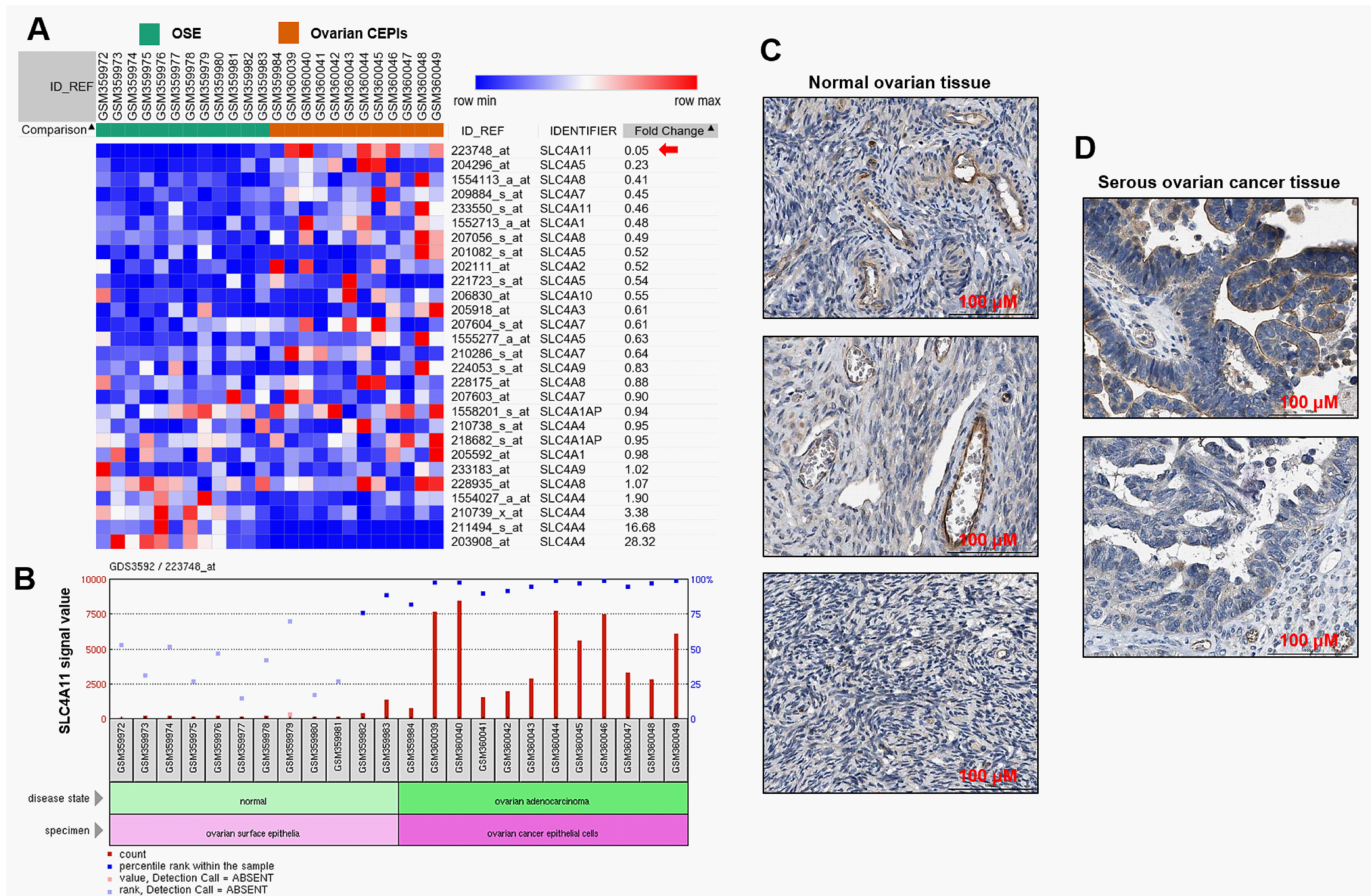


Fig 1. *SLC4A11* is upregulated in ovarian cancer compared with normal ovarian epithelial tissues. **A.** Heat map of the expression of SLC4A family members in 12 cases of ovarian CEPIs compared with 12 cases of OSE. Red: up-regulation; Blue: down-regulation. The image was generated by re-analysis of the raw microarray data of GDS3592. **B.** *SLC4A11* microarray signal values in 12 CEPIs and 12 OSE cases. Data were analyzed by using the tool provided by GEO datasets. **C-D.** Representative images of IHC staining of SLC4A11 in normal ovarian tissues (C) and serous ovarian cancer tissues (D). Data were obtained from the HPA: <http://www.proteinatlas.org/ENSG0000088836-SLC4A11/pathology/tissue/ovarian+cancer>.

<https://doi.org/10.1371/journal.pone.0187385.g001>

compared with the OSE samples (Fig 1B). By data mining in the HPA, we found that in normal ovarian tissues, follicle cells had medium SLC4A11 staining (Fig 1C). But SLC4A11 expression was not detectable in ovarian stroma cells (Fig 1C). In comparison, the serous ovarian cancer tissues usually had low to medium SLC4A11 staining in both cytoplasm and cell membrane (Fig 1D).

High *SLC4A11* expression is associated with lymphatic invasion

Using data from TCGA-OV, we compared *SLC4A11* expression in ovarian cancer patients with or without lymphatic invasion. Results revealed that the cases with lymphatic invasion (N = 133) had significantly higher *SLC4A11* expression ($p = 0.0069$) than those without lymphatic invasion (N = 77) (Fig 2A and 2B).

High *SLC4A11* expression is an independent predictor of poor OS in ovarian cancer patients

The association between clinicopathological features and *SLC4A11* expression in patients with primary serous ovarian cancer was summarized in Table 1. High *SLC4A11* expression was significantly associated with lower grade tumors (GB/G1/G2) ($p = 0.038$), and a larger proportion of lymphatic invasion ($p = 0.027$) and deceased cases ($p = 0.036$) (Table 1). In addition, the high *SLC4A11* expression group also had a higher ratio of tumor residual disease at the margin level of significance ($p = 0.050$) (Table 1). However, no significant difference was observed in

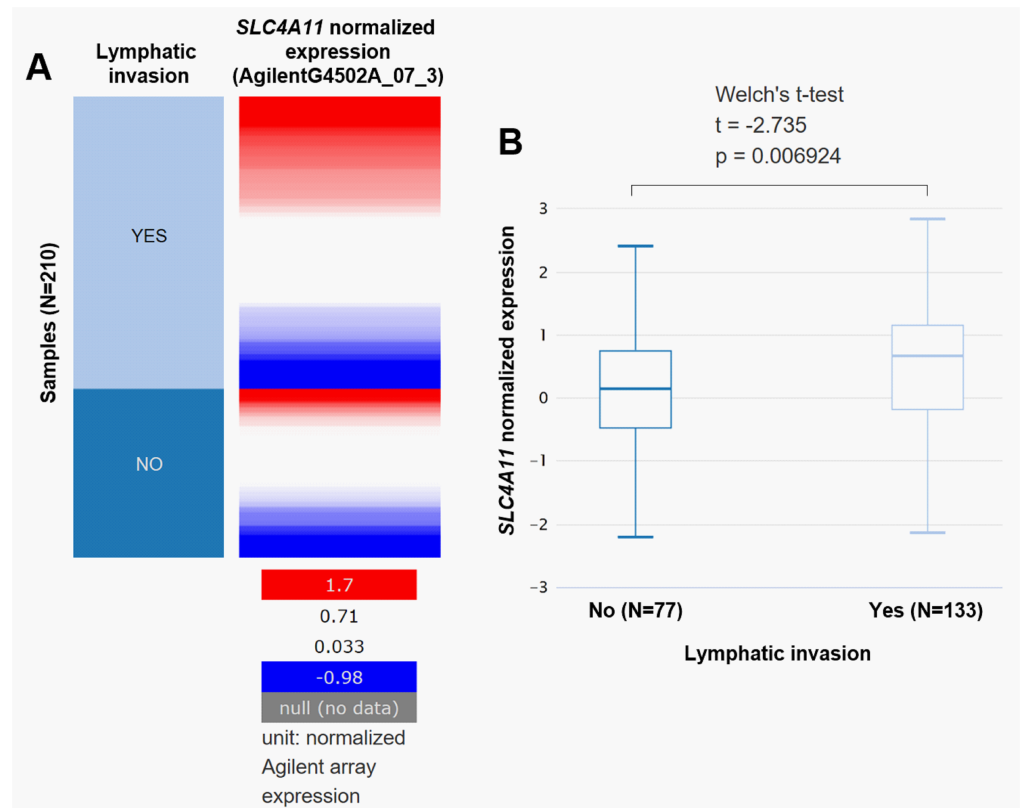


Fig 2. High *SLC4A11* expression is associated with lymphatic invasion. A-B. Heat map (A) and box plots (B) of *SLC4A11* expression in patients with or without lymphatic invasion.

<https://doi.org/10.1371/journal.pone.0187385.g002>

Table 1. Demographic and clinicopathological parameters of patients with primary ovarian cancer in TCGA-OV.

Parameters		SLC4A11 expression RNAseq		χ^2	p Value
		High (N = 270)	Low (N = 270)		
Age (Mean \pm SD)	≥ 60	131	122	0.54	0.46
	<60	139	147		
	Null	0	1		
Grade	GB/G1/G2	46	30	4.30	0.038
	G3/G4	215	236		
	GX + Null	9	4		
Clinical stage	I/II	18	24	0.93	0.33
	III/IV	249	243		
	Null	3	3		
Venous invasion	No	30	37	2.05	0.15
	Yes	48	37		
	Null	192	196		
Lymphatic invasion	No	33	44	4.88	0.027
	Yes	78	55		
	Null	159	171		
Tumor residual disease	No	44	62	3.83	0.050
	Yes	194	177		
	Null	32	31		
Recurrence status	No	5	10	0.24	0.62
	Yes	21	31		
	Null	244	229		
Living Status	Living	99	122	4.38	0.036
	Dead	169	144		
	Null	2	4		

GB: Border line malignancy; G1: Well differentiated; G2: Moderately differentiated; G3-G4: Poorly differentiated or Undifferentiated; GX: Grade cannot be assessed. Null: no data.

<https://doi.org/10.1371/journal.pone.0187385.t001>

recurrence rate between the two groups (Table 1). Then, Kaplan-Meier survival analysis was performed between the high and low *SLC4A11* expression groups in TCGA-OV. In the database, *SLC4A11* expression was quantified by RNAseq (AgilentG4502A_07_3 and IlluminaHiSeq respectively) and exon RNAseq (polyA+ IlluminaHiSeq). Among 540 patients with *SLC4A11* measured by AgilentG4502A_07_3, 534 patients had OS data. Both IlluminaHiSeq and RNAseq data included 302 patients with OS data (Fig 2A–2C). Log-rank test showed that in all measurements, high *SLC4A11* expression was consistently associated with worse OS compared with low *SLC4A11* expression ($p = 0.00025, 0.0051$ and 0.014 respectively, Fig 2A–2C). Recent studies suggest that GB/G1/G2 carcinoma belong to type I, while G3/G4 carcinomas belong to type II tumors, which have significant clinicopathologic and molecular differences [20, 21]. We then performed subgroup analysis in GB/G1/G2 and G3/G4 tumors respectively. In both groups, we found that high *SLC4A11* expression was associated with unfavorable OS (Fig 3D and 3E). In univariate analysis, high *SLC4A11* expression was associated with poor prognosis in terms of OS in both GB/G1/G2 and G3/G4 tumors ($p = 0.011$ and $p < 0.001$ respectively) (Table 2). However, following multivariate analysis only confirmed the independent prognostic value of high *SLC4A11* expression in G3/G4 tumors (HR = 1.416, 95%CI: 1.098–1.824, $p = 0.007$; Table 2), but not in GB/G1/G2 tumors (HR = 1.832, 95%CI: 0.987–3.402, $p = 0.055$; Table 2). Due to insufficient data of recurrence, we only assessed the

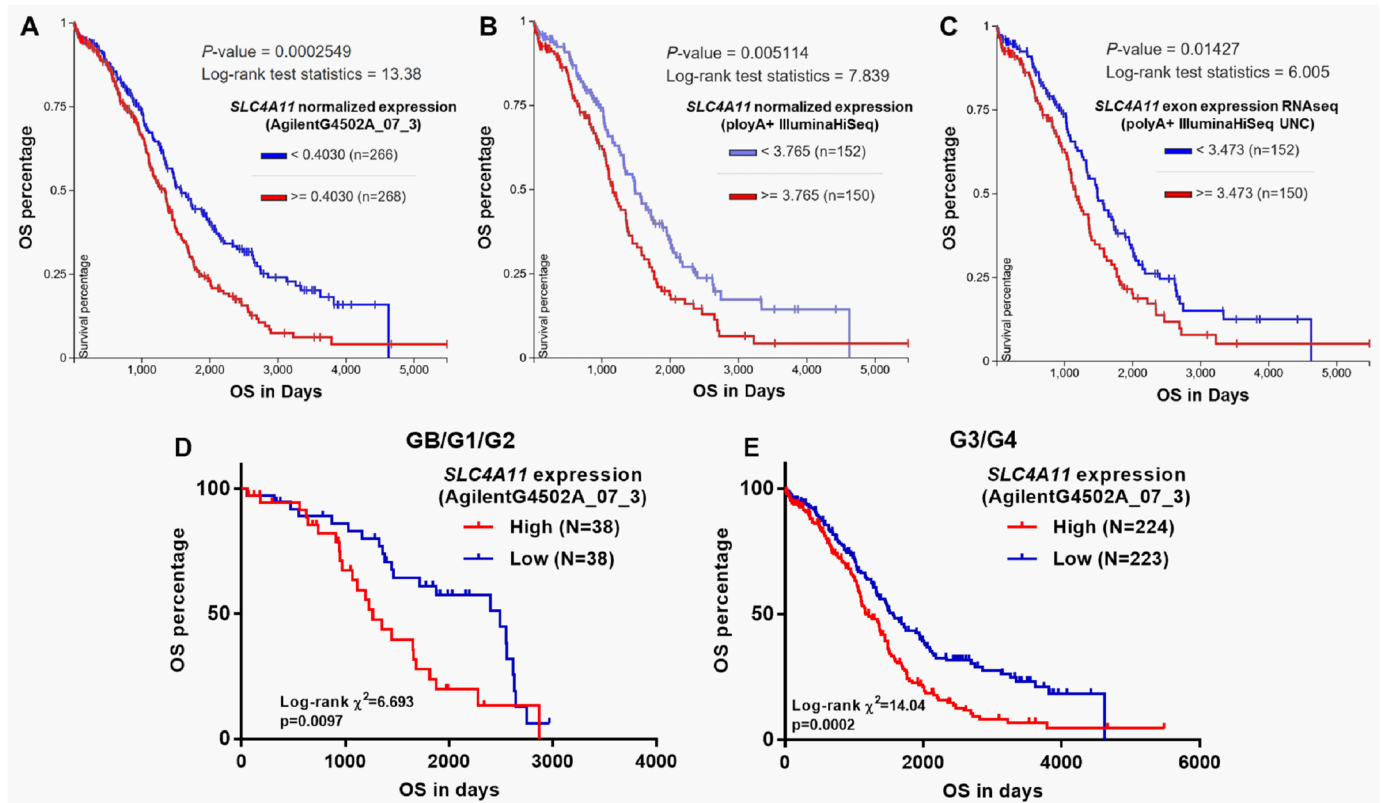


Fig 3. High *SLC4A11* expression is associated with poor survivals in ovarian cancer patients. A-E. Kaplan-Meier curves of OS in ovarian cancer patients with high and low *SLC4A11* expression. Data were obtained from TCGA-OV. *SLC4A11* expression was quantified by RNAseq (AgilentG4502A_07_3 (A) and IlluminaHiSeq respectively (B) and exon RNAseq (polyA+ IlluminaHiSeq) (C). Kaplan-Meier curves of OS in GB/G1/G2 patients (D) and in G3/G4 patients (E).

<https://doi.org/10.1371/journal.pone.0187385.g003>

association between *SLC4A11* expression and RFS in patients with G3/G4 diseases. No independent prognostic value of *SLC4A11* expression was observed in terms of RFS (Table 2).

SLC4A11 expression is regulated by DNA amplification and methylation in ovarian cancer

Via analyzing the deep sequencing data in TCGA-OV, we further explored the mechanisms of *SLC4A11* dysregulation. Among 578 patients with copy number measured, 320 (55.4%) cases had DNA amplification (Fig 4A). By comparing *SLC4A11* expression and its DNA methylation status, we also confirmed that the high methylation group had a significantly lower level of *SLC4A11* expression (Fig 4B–4D).

Discussion

As a Na⁺-coupled base transporter capable of Na⁺-H⁺ exchange in mammalian cells, *SLC4A11* mutation is associated with a series of corneal endothelial diseases characterized by dysfunction of the endothelial cells in the inner surface of the cornea [22–24]. These diseases finally result in the apoptosis of corneal endothelial cells, and subsequent edema and impaired vision [25]. In fact, the normal corneal endothelial function is dependent on the balanced regulation of bicarbonate concertation and pHi/pHe. Based on these findings, we infer that the putative

Table 2. Univariate and multivariate analyses of OS/RFS in patients with primary ovarian cancer in TCGA-OV.

Parameters	Univariate analysis				Multivariate analysis			
	p	HR	95%CI (lower/upper)		p	HR	95%CI (lower/upper)	
OS (GB/G1/G2) Age ≥60 vs. < 60	0.011	2.171	1.192	3.956	0.999	1.00	0.549	1.824
Clinical stage III/IV vs. I/II	0.018	5.557	1.338	23.07	0.0367	4.63	1.099	19.504
Venous invasion No vs. Yes	0.112	0.392	0.123	1.244				
Lymphatic invasion No vs. Yes	0.999	1.001	0.27	3.712				
Tumor residual disease No vs. Yes	0.208	1.693	0.746	3.841				
SLC4A11 expression High vs. Low	0.011	2.171	1.192	3.956	0.055	1.832	0.987	3.402
OS (G3/G4) Age ≥60 vs. < 60	<0.001	1.616	1.266	2.062	0.001	1.516	1.187	1.937
Clinical stage III/IV vs. I/II	0.123	1.743	0.861	3.529				
Venous invasion No vs. Yes	0.533	1.2	0.676	2.132				
Lymphatic invasion No vs. Yes	0.055	0.613	0.373	1.01	0.563	0.859	0.514	1.437
Tumor residual disease No vs. Yes	<0.001	2.46	1.647	3.675	<0.001	2.252	1.497	3.39
SLC4A11 expression High vs. Low	<0.001	1.595	1.247	2.04	0.007	1.416	1.098	1.824
RFS (G3/G4) Age ≥60 vs. < 60	0.3004	1.366	0.757	2.465				
Clinical stage III/IV vs. I/II	0.146	0.22	0.029	1.692				
Tumor residual disease No vs. Yes	0.017	2.421	1.175	4.989	0.041	2.167	1.031	4.553
SLC4A11 expression High vs. Low	0.052	1.769	0.996	3.14	0.179	1.511	0.828	2.757

GB: Border line malignancy; G1: Well differentiated; G2: Moderately differentiated; G3-G4: Poorly differentiated or Undifferentiated. OS: overall survival; RFS: recurrence-free survival.

<https://doi.org/10.1371/journal.pone.0187385.t002>

bicarbonate transport or Na⁺-H⁺ exchange activity of SLC4A11 is essential for maintaining normal cellular physiological functions.

In this study, we found that *SLC4A11* expression is significantly upregulated in ovarian cancer tissues than in normal tissues. The alkaline intracellular environment of cancer cells is usually associated with the upregulation of the expression and/or activity of acid extruders or downregulation of the expression and/or activity of acid loading transporters [1]. Given the pHi regulatory activity demonstrated in previous studies [10, 26], SLC4A11 naturally owns a strong cellular buffering capacity that enables the highly glycolytic cancer cells to remove lactic acid rapidly. Its upregulation can be considered as an adaptive response of the cancer cells to metabolic changes. In addition, upregulation of the acid extruders leads to extracellular acidification, thereby generating a favorable environment for tumor invasion and metastasis [5, 6]. In this study, we observed that the patients with metastasis had significantly higher *SLC4A11* expression than their counterparts without metastasis. Therefore, we infer that *SLC4A11* upregulation is an important mechanism leading to malignant cellular behaviors in ovarian cancer.

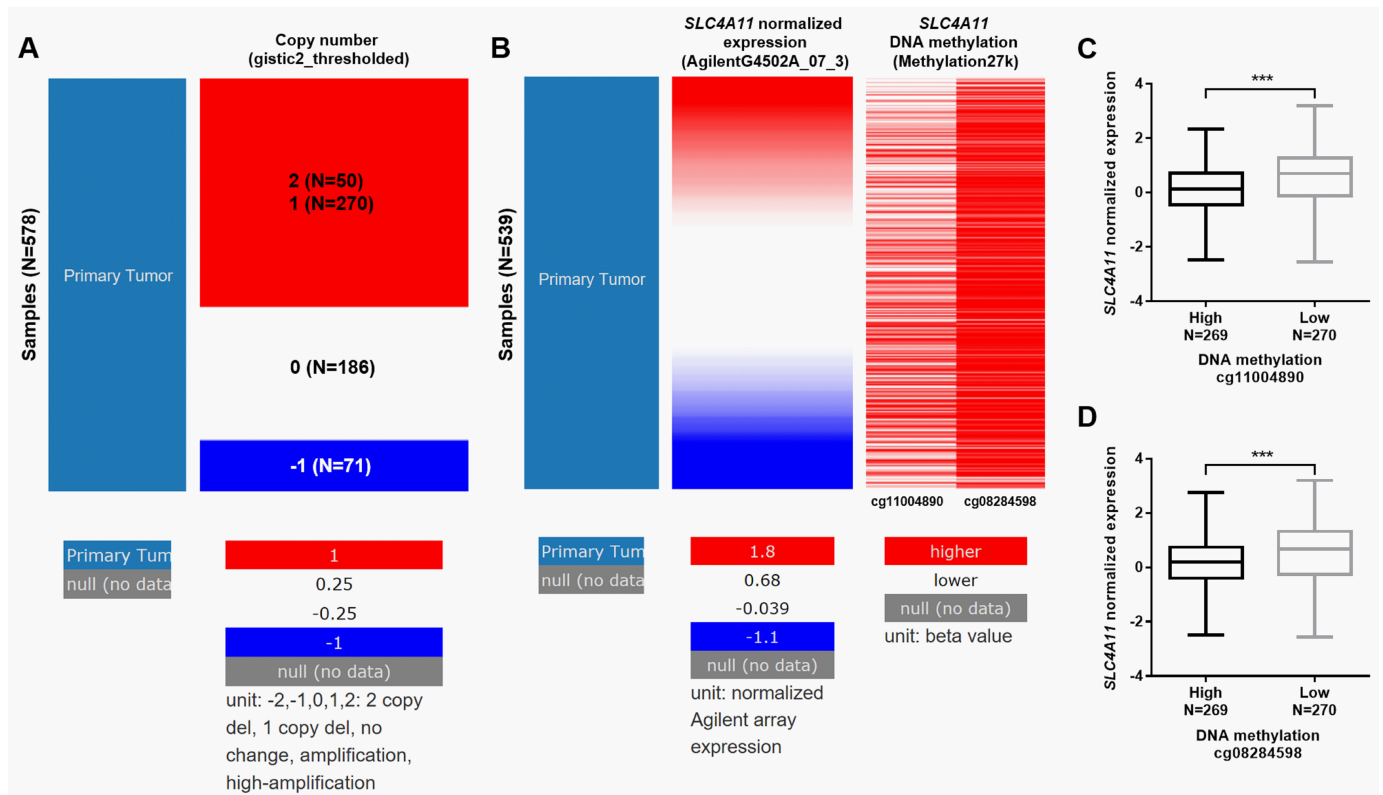


Fig 4. *SLC4A11* expression is regulated by DNA amplification and methylation in ovarian cancer. **A.** Heat map of *SLC4A11* copy number changes in TCGA-OV. -1: deletion; 0: no change; 1: amplification; 2: high-amplification. **B-D.** Heat map (B) and box plots (C-D) of *SLC4A11* expression and DNA methylation in TCGA-OV. Patients were divided into high and low methylation groups according to the median value of the two probes (cg11004890 and cg08284598) respectively.

<https://doi.org/10.1371/journal.pone.0187385.g004>

By comparing the clinicopathological features between high and low *SLC4A11* expression groups, we found that the high *SLC4A11* expression group had a significantly higher ratio of lymphatic invasion and deceased cases. However, we also observed that the high *SLC4A11* expression group had a substantially larger proportion of low-grade tumors (GB/G1/G2) (46/261, 17.6% vs. 30/261, 11.3%) compared with the low *SLC4A11* group. Actually, G1/G2 serous ovarian carcinomas belong to type I tumors, while G3/G4 serous ovarian carcinomas belong to type II tumors, which have different molecular and biological profiles [20, 21]. We hypothesized that the difference in tumor grade might be related to the genetic difference between type I and type II tumors. However, more studies are required to confirm this hypothesis.

Currently, a series of predictors such as clinical stage, CA125 levels, age, response to chemotherapy and residual disease after debulking surgery are used to predict prognosis of ovarian cancer patients. However, there is still some discrepancy in the prognosis of similar tumors characterized by these clinical variables [27]. Ovarian cancers actually are a heterogeneous group of neoplasias derived from the ovarian surface epithelium, inclusion cysts, or the fallopian tube [28, 29]. The differences at the molecular level may be important sources of the variations. Therefore, it is necessary to explore other molecular parameters for better prediction of prognosis. In this study, based on the survival data of patients in TCGA-OV, we found that high *SLC4A11* expression was robustly associated with poor OS. Following univariate and multivariate analysis confirmed that high *SLC4A11* expression was an independent prognostic

factor for poor OS in G3/G4 tumors. These findings suggest that *SLC4A11* might be a potential clinical marker in this group of patients. Although the association between *SLC4A11* expression and OS in GB/G1/G2 patients was not significant, it showed a trend toward significance ($p = 0.055$). Considering the small number of GB/G1/G2 patients included in this study ($N = 76$), it is meaningful to further explore its prognostic value with a large sample base in the future.

SLC4A11 locates in chromosome 20p in the human genome, a region with a relatively high frequency of genetic amplification in ovarian cancer [30, 31]. By using deep sequencing data in TCGA-OV, we found that 320 out of 578 ovarian cancer cases had *SLC4A11* amplification. Therefore, DNA amplification is one important mechanism of upregulated *SLC4A11* in ovarian cancer. Epigenetic regulation, such as methylation and histone modification are also important mechanisms of dysregulated genes in ovarian cancer [32, 33]. Thus, we also investigated whether methylation status influences *SLC4A11* expression. By using the results from Illumina 27k methylation array, we found that the group with high DNA methylation had significantly lower *SLC4A11* expression, suggesting that epigenetic alterations also contribute to *SLC4A11* upregulation in ovarian cancer.

Conclusion

Based on findings above, we infer that high *SLC4A11* expression is an independent predictor for poor OS in grade 3/4 serous ovarian cancer. Both DNA amplification and hypomethylation contribute to its upregulation in ovarian cancer.

Supporting information

S1 Table. Original data downloaded from the UCSC Xena Browser.
(XLSX)

Author Contributions

Conceptualization: Lianzhi Qin, Yuhua Liu.

Data curation: Ting Li, Yuhua Liu.

Formal analysis: Ting Li, Yuhua Liu.

Investigation: Lianzhi Qin, Ting Li, Yuhua Liu.

Methodology: Lianzhi Qin, Ting Li, Yuhua Liu.

Project administration: Yuhua Liu.

Resources: Yuhua Liu.

Software: Ting Li, Yuhua Liu.

Supervision: Lianzhi Qin, Yuhua Liu.

Validation: Lianzhi Qin, Ting Li, Yuhua Liu.

Visualization: Lianzhi Qin, Ting Li.

Writing – original draft: Lianzhi Qin, Ting Li.

Writing – review & editing: Yuhua Liu.

References

1. Gorbatenko A, Olesen CW, Boedtkjer E, Pedersen SF. Regulation and roles of bicarbonate transporters in cancer. *Front Physiol*. 2014; 5:130. <https://doi.org/10.3389/fphys.2014.00130> PMID: 24795638
2. Parks SK, Chiche J, Pouyssegur J. Disrupting proton dynamics and energy metabolism for cancer therapy. *Nat Rev Cancer*. 2013; 13(9):611–23. <https://doi.org/10.1038/nrc3579> PMID: 23969692.
3. Pedersen SF, Stock C. Ion channels and transporters in cancer: pathophysiology, regulation, and clinical potential. *Cancer Res*. 2013; 73(6):1658–61. <https://doi.org/10.1158/0008-5472.CAN-12-4188> PMID: 23302229.
4. Andersen AP, Moreira JM, Pedersen SF. Interactions of ion transporters and channels with cancer cell metabolism and the tumour microenvironment. *Philos Trans R Soc Lond B Biol Sci*. 2014; 369(1638):20130098. <https://doi.org/10.1098/rstb.2013.0098> PMID: 24493746
5. Estrella V, Chen T, Lloyd M, Wojtkowiak J, Cornell HH, Ibrahim-Hashim A, et al. Acidity generated by the tumor microenvironment drives local invasion. *Cancer Res*. 2013; 73(5):1524–35. <https://doi.org/10.1158/0008-5472.CAN-12-2796> PMID: 23288510
6. Gatenby RA, Gawlinski ET, Gmitro AF, Kaylor B, Gillies RJ. Acid-mediated tumor invasion: a multidisciplinary study. *Cancer Res*. 2006; 66(10):5216–23. <https://doi.org/10.1158/0008-5472.CAN-05-4193> PMID: 16707446.
7. Romero MF, Chen AP, Parker MD, Boron WF. The SLC4 family of bicarbonate (HCO₃⁻) transporters. *Mol Aspects Med*. 2013; 34(2–3):159–82. <https://doi.org/10.1016/j.mam.2012.10.008> PMID: 23506864
8. Thornell IM, Bevensee MO. Regulators of Slc4 bicarbonate transporter activity. *Front Physiol*. 2015; 6:166. <https://doi.org/10.3389/fphys.2015.00166> PMID: 26124722
9. Shei W, Liu J, Htoon HM, Aung T, Vithana EN. Differential expression of the Slc4 bicarbonate transporter family in murine corneal endothelium and cell culture. *Mol Vis*. 2013; 19:1096–106. PMID: 23734078
10. Ogando DG, Jalimarada SS, Zhang W, Vithana EN, Bonanno JA. SLC4A11 is an EIPA-sensitive Na⁺ permeable pH_i regulator. *Am J Physiol Cell Physiol*. 2013; 305(7):C716–27. <https://doi.org/10.1152/ajpcell.00056.2013> PMID: 23864606
11. Lee S, Axelsen TV, Andersen AP, Vahl P, Pedersen SF, Boedtkjer E. Disrupting Na⁺, HCO₃⁻ cotransporter NBCn1 (Slc4a7) delays murine breast cancer development. *Oncogene*. 2016; 35(16):2112–22. <https://doi.org/10.1038/onc.2015.273> PMID: 26212013.
12. Boedtkjer E, Moreira JM, Mele M, Vahl P, Wielenga VT, Christiansen PM, et al. Contribution of Na⁺, HCO₃⁻ cotransport to cellular pH control in human breast cancer: a role for the breast cancer susceptibility locus NBCn1 (SLC4A7). *Int J Cancer*. 2013; 132(6):1288–99. <https://doi.org/10.1002/ijc.27782> PMID: 22907202.
13. Parks SK, Pouyssegur J. The Na⁺/HCO₃⁻ Co-Transporter SLC4A4 Plays a Role in Growth and Migration of Colon and Breast Cancer Cells. *J Cell Physiol*. 2015; 230(8):1954–63. <https://doi.org/10.1002/jcp.24930> PMID: 25612232.
14. McIntyre A, Hulikova A, Ledaki I, Snell C, Singleton D, Steers G, et al. Disrupting Hypoxia-Induced Bicarbonate Transport Acidifies Tumor Cells and Suppresses Tumor Growth. *Cancer Res*. 2016; 76(13):3744–55. <https://doi.org/10.1158/0008-5472.CAN-15-1862> PMID: 27197160.
15. Sanhueza C, Araos J, Naranjo L, Villalobos R, Westermeier F, Salomon C, et al. Modulation of intracellular pH in human ovarian cancer. *Curr Mol Med*. 2016; 16(1):23–32. PMID: 26695697.
16. Sanhueza C, Araos J, Naranjo L, Toledo F, Beltran AR, Ramirez MA, et al. Sodium/proton exchanger isoform 1 regulates intracellular pH and cell proliferation in human ovarian cancer. *Biochim Biophys Acta*. 2017; 1863(1):81–91. <https://doi.org/10.1016/j.bbadis.2016.10.013> PMID: 27773735.
17. Bowen NJ, Walker LD, Matyunina LV, Logani S, Totten KA, Benigno BB, et al. Gene expression profiling supports the hypothesis that human ovarian surface epithelia are multipotent and capable of serving as ovarian cancer initiating cells. *BMC Med Genomics*. 2009; 2:71. <https://doi.org/10.1186/1755-8794-2-71> PMID: 20040092
18. Thul PJ, Akesson L, Wiking M, Mahdessian D, Geladaki A, Ait Blal H, et al. A subcellular map of the human proteome. *Science*. 2017; 356(6340). <https://doi.org/10.1126/science.aal3321> PMID: 28495876.
19. Uhlen M, Oksvold P, Fagerberg L, Lundberg E, Jonasson K, Forsberg M, et al. Towards a knowledge-based Human Protein Atlas. *Nat Biotechnol*. 2010; 28(12):1248–50. <https://doi.org/10.1038/nbt1210-1248> PMID: 21139605.
20. Kurman RJ, Shih Ie M. The Dualistic Model of Ovarian Carcinogenesis: Revisited, Revised, and Expanded. *Am J Pathol*. 2016; 186(4):733–47. <https://doi.org/10.1016/j.ajpath.2015.11.011> PMID: 27012190.

21. Koshiyama M, Matsumura N, Konishi I. Recent concepts of ovarian carcinogenesis: type I and type II. *Biomed Res Int*. 2014; 2014:934261. <https://doi.org/10.1155/2014/934261> PMID: 24868556
22. Vithana EN, Morgan PE, Ramprasad V, Tan DT, Yong VH, Venkataraman D, et al. SLC4A11 mutations in Fuchs endothelial corneal dystrophy. *Hum Mol Genet*. 2008; 17(5):656–66. <https://doi.org/10.1093/hmg/ddm337> PMID: 18024964.
23. Vithana EN, Morgan P, Sundaresan P, Ebenezer ND, Tan DT, Mohamed MD, et al. Mutations in sodium-borate cotransporter SLC4A11 cause recessive congenital hereditary endothelial dystrophy (CHED2). *Nat Genet*. 2006; 38(7):755–7. <https://doi.org/10.1038/ng1824> PMID: 16767101.
24. Vilas GL, Loganathan SK, Liu J, Riau AK, Young JD, Mehta JS, et al. Transmembrane water-flux through SLC4A11: a route defective in genetic corneal diseases. *Hum Mol Genet*. 2013; 22(22):4579–90. <https://doi.org/10.1093/hmg/ddt307> PMID: 23813972
25. Patel SP, Parker MD. SLC4A11 and the Pathophysiology of Congenital Hereditary Endothelial Dystrophy. *Biomed Res Int*. 2015; 2015:475392. <https://doi.org/10.1155/2015/475392> PMID: 26451371
26. Kao L, Azimov R, Abuladze N, Newman D, Kurtz I. Human SLC4A11-C functions as a DIDS-stimulatable H⁽⁺⁾(OH⁽⁻⁾) permeation pathway: partial correction of R109H mutant transport. *Am J Physiol Cell Physiol*. 2015; 308(2):C176–88. <https://doi.org/10.1152/ajpcell.00271.2014> PMID: 25394471
27. Shimada T, Saito T, Shimokawa M, Shimamoto K, Matsushita S, Yamaguchi S, et al. Improvement in the prognosis of ovarian cancer in the era before addition of molecular targeting therapy. *Jpn J Clin Oncol*. 2017; 47(6):494–8. <https://doi.org/10.1093/jjco/hyx026> PMID: 28334884.
28. McCluggage WG. Morphological subtypes of ovarian carcinoma: a review with emphasis on new developments and pathogenesis. *Pathology*. 2011; 43(5):420–32. <https://doi.org/10.1097/PAT.0b013e328348a6e7> PMID: 21716157.
29. Cancer Genome Atlas Research N. Integrated genomic analyses of ovarian carcinoma. *Nature*. 2011; 474(7353):609–15. <https://doi.org/10.1038/nature10166> PMID: 21720365
30. Watanabe T, Imoto I, Kosugi Y, Ishiwata I, Inoue S, Takayama M, et al. A novel amplification at 17q21-23 in ovarian cancer cell lines detected by comparative genomic hybridization. *Gynecol Oncol*. 2001; 81(2):172–7. <https://doi.org/10.1006/gyno.2001.6132> PMID: 11330945.
31. Lambros MB, Fiegler H, Jones A, Gorman P, Roylance RR, Carter NP, et al. Analysis of ovarian cancer cell lines using array-based comparative genomic hybridization. *J Pathol*. 2005; 205(1):29–40. <https://doi.org/10.1002/path.1681> PMID: 15586366.
32. Kwon MJ, Shin YK. Epigenetic regulation of cancer-associated genes in ovarian cancer. *Int J Mol Sci*. 2011; 12(2):983–1008. <https://doi.org/10.3390/ijms12020983> PMID: 21541038
33. Asadollahi R, Hyde CA, Zhong XY. Epigenetics of ovarian cancer: from the lab to the clinic. *Gynecol Oncol*. 2010; 118(1):81–7. <https://doi.org/10.1016/j.ygyno.2010.03.015> PMID: 20421130.