



Review Sirtuins as Mediator of the Anti-Ageing Effects of Calorie Restriction in Skeletal and Cardiac Muscle

Alberto Zullo ^{1,2,*,†}, Emanuela Simone ^{1,†}, Maddalena Grimaldi ³, Vincenzina Musto ¹ and Francesco Paolo Mancini ^{1,*}

- ¹ Department of Sciences and Technologies, University of Sannio, 82100 Benevento, Italy; emanuela.simone1@gmail.com (E.S.); vincenzina.musto@icloud.com (V.M.)
- ² CEINGE Biotecnologie Avanzate s.c.ar.l., 80145 Naples, Italy
- ³ Department of Pediatric Oncology and Hematology, Charité University Hospital, 13353 Berlin, Germany; grimaldi.madda@gmail.com
- * Correspondence: albzullo@unisannio.it (A.Z.); mancini@unisannio.it (F.P.M.); Tel.: +39-0824-305107 (A.Z. & F.P.M.)
- + These authors contributed equally to this work.

Received: 18 February 2018; Accepted: 20 March 2018; Published: 21 March 2018



Abstract: Fighting diseases and controlling the signs of ageing are the major goals of biomedicine. Sirtuins, enzymes with mainly deacetylating activity, could be pivotal targets of novel preventive and therapeutic strategies to reach such aims. Scientific proofs are accumulating in experimental models, but, to a minor extent, also in humans, that the ancient practice of calorie restriction could prove an effective way to prevent several degenerative diseases and to postpone the detrimental signs of ageing. In the present review, we summarize the evidence about the central role of sirtuins in mediating the beneficial effects of calorie restriction in skeletal and cardiac muscle since these tissues are greatly damaged by diseases and advancing years. Moreover, we entertain the possibility that the identification of sirtuin activators that mimic calorie restriction could provide the benefits without the inconvenience of this dietary style.

Keywords: sirtuins; calorie restriction; ageing; nutrient deprivation; skeletal muscle; cardiac muscle

1. Introduction

Environmental factors profoundly affect the fate of living organisms and nutrition is one of the most influential factors. Nowadays longevity is a major goal of medical science and has always been a chimera for the human being since ancient times. In particular, efforts are aimed at achieving successful ageing, namely a long life in the absence of serious diseases, with a good level of physical and mental independence and adequate social relationships [1].

Accumulating data clearly demonstrates that it is possible to influence the signs of ageing. Indeed, nutritional interventions can promote health and longevity. A tribute must be given to Ancel Keys, who was the first one to provide solid scientific evidence about the role of nutrition in the health/disease balance at the population level, specifically in relation to cardiovascular disease, still the leading cause of death worldwide [2–4]. It is generally appreciated that the type of diet can profoundly influence the quality and quantity of life and the Mediterranean diet is paradigmatic of a beneficial dietary pattern [5–7].

The growing consciousness of the beneficial effects of a specific dietary pattern on health and longevity in the second half of the last century generated a powerful thrust toward designing diets that could reduce the risk of chronic diseases, thus resulting in healthy ageing. Thus, in the 1990s the Dietary Approaches to Stop Hypertension (DASH) diet was devised in order to evaluate whether it

was possible to treat hypertension non pharmacologically [8]. Indeed, the DASH diet was quite similar to the Mediterranean Diet, being rich in fruit and vegetable, whole grains, and fibers, while poor in animal saturated fats and cholesterol. The very good news coming out of the study was that not only did the DASH diet lower blood pressure, but it also reduced the risk of cardiovascular disease, type 2 diabetes, some types of cancer, and other aging-associated diseases [9,10]. To further improve the health benefits of plant food-rich, animal fat-poor diets, particularly in hypercholesterolemic individuals, the Portfolio Diet was designed [11]. This diet, besides being largely vegetarian, with only small amounts of saturated fats, recommends also a high intake of functional foods, including viscous fibers, plant stanols, soy proteins, and almonds. Interestingly, participants on the Portfolio Diet exhibited a reduction of coronary heart disease risk associated with lower plasma cholesterol and inflammatory indexes in comparison to participants on a healthy, mainly vegetarian diet [12].

However, also the amount of ingested food has been attracting the interest of the scientific community as a potential modifier of the balance between health and disease in many different living species. In particular, calorie restriction (CR) has been demonstrated to be an emerging nutritional intervention that stimulates the anti-ageing mechanisms in the body [13–22]. Therefore, the diet of the people living on the Japanese island of Okinawa has been extensively analyzed because these islanders are well-known for their longevity and increased health span, resulting in the greatest frequency of centenarians in the world [23]. Very interestingly, the traditional Okinawan diet resulted to be very similar to the Mediterranean Diet and the DASH diet in terms of food types [23]. However, the energy intake of Okinawans, at the time of the initial scientific observations, was about 20% lower than the average energy intake of the Japanese, thus determining a typical condition of CR [24].

CR causes life extension through mechanisms that involve different molecular players among which sirtuins, a family of nutrient-sensing proteins, take center stage. Sirtuins have different, NAD⁺-dependent enzymatic activities, among which the deacylating one is common to all the isoforms [25]. Sirtuins are expressed in several tissues, including skeletal muscle, which is particularly relevant in ageing, due to its influence on metabolism and its gradual loss as organisms get older. In this review, we focus on the role of sirtuins in mediating the anti-ageing effects of CR in skeletal and cardiac muscle (Table 1).

Skeletal Muscle	Cardiac Muscle
\uparrow Mass and function	Altered lipid metabolism
\uparrow Satellite cell activity	Hypertrophy
↓ Inflammation	↓ Inflammation
\uparrow Resistance to stress	\uparrow Resistance to stress
\uparrow Insulin sensitivity	↑ Insulin sensitivity
↑ Autophagy	↑ Autophagy
Mitochondrial dysfunction	Mitochondrial dysfunction
↓ DNA damage	-

Table 1. Sirtuin-mediated anti-ageing effects of Calorie Restriction in cardiac and skeletal muscle.

2. Calorie Restriction, Sirtuins, and Ageing

CR is a dietary regimen with a 20–40% reduction of the normal daily calorie intake without malnutrition [26]. The effect of CR on ageing was first shown by McCay and colleagues in an animal model over 80 years ago [27]. They observed that calorie-resticted mice lived longer compared to control mice on a regular diet. These early results have been confirmed later by many other studies in different species, from unicellular yeast to non-human primates [15–22,28–31].

However, while two of the three available studies demonstrated increased survival in calorie-restricted rhesus monkeys, the third study did not report any significant effect of CR on the survival of the experimental animals [15,32,33]. Interestingly, the same authors of two out of the three contrasting papers reassessed their studies together. As results, they identified the differences in

experimental design that could explain, at least in part, the diverging outcomes and concluded that CR was indeed able to improve both health and survival in monkeys [34]. At this point, it is conceivable to speculate that the same mechanisms and effects on health and survival could also take place in humans practicing CR. Yet, there are no long-term human studies to validate this hypothesis. Indirect evidence comes from historical episodes, such as the forced reduction of food intake in Danish people during the World War 1 that led to a reduction of 34% of death rates, and, similarly, a forced 20% CR in the population of Oslo, during World War 2 that brought about a 30% reduction of mortality [35,36]. A natural experiment is provided by the already mentioned population of the Japanese island of Okinawa, where 4–5 times higher incidence of centenarians compared to any other industrialized country was registered, in association with a spontaneous calorie-restricted diet due to the frugal nutritional habits of the Okinawans [37].

As firstly demonstrated in animal models, CR delays the detrimental consequences of the signs of ageing, and this is a key point underlying the health effects of CR. In particular, CR is inversely correlated with the onset of age-related diseases, such as diabetes, atherosclerosis, cardiomyopathy, kidney disease, sarcopenia, respiratory disease and cancer [14,38]. In rodents and monkeys, CR reduces insulin resistance, glucose intolerance, cognitive decline and immune dysfunction [34,39–41]. Moreover, it has been demonstrated that CR also attenuates the age-related neurodegeneration, sarcopenia, and auditory loss [42]. Importantly, these metabolic and functional effects of CR are long-lasting [34].

So far, some observational and randomized clinical trials have been performed also in humans. They confirmed the anti-ageing effects of CR, including a delayed onset of the most common chronic diseases, such as type 2 diabetes, cardiovascular diseases, and cancer. In particular, the metabolic and molecular changes induced by CR in humans are similar to those promoting health and prolonging life of CR-treated animals [42].

At a molecular level (Figure 1), the effects of CR involve nutrient-signaling pathways, such as those of the mammalian target of rapamycin (mTOR), the insulin-like growth factor 1 (IGF-1), and a family of seven proteins named sirtuins [31]. Sirtuins target proteins involved in different processes, such as DNA repair, DNA stability, epigenetic modification of chromatin, stress resistance, cell cycle progression, reactive oxygen species (ROS) production and metabolism, mitochondrial function and autophagy [43]. Several pathways can activate sirtuins following CR. For instance, the upregulation of endothelial nitric oxide synthase (eNOS), which is a very important effector of CR, can activate sirtuin 1 (SIRT1) [44–46]. Then, sirtuin activation modifies the acetylation/deacetylation balance, a crucial mechanism for mediating the metabolic responses to changes in nutrient availability [47]. Many of the proteins targeted by acetylation are localized in the mitochondria and are involved in the tricarboxylic acid cycle, β -oxidation, and oxidative phosphorylation [47]. Nutritional stimuli not only influence mitochondrial biochemical pathways, but also affect the number and life cycle of these organelles. Indeed, CR, via inhibition of IGF-1 and mTOR pathways and activation of AMP-activated protein kinase (AMPK) and sirtuins, modulates oxidative metabolism as well as the biogenesis and turnover of mitochondria. AMPK and peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1 α) are key players in these pathways. CR also lowers ROS production through enhanced mitochondrial aerobic metabolism and increased activity of antioxidant enzymes [48].



Cardiac Muscle

Skeletal Muscle

Figure 1. Molecular mechanisms underlying the sirtuin-mediated anti-ageing effects of Calorie Restriction in cardiac and skeletal muscle. CR can modulate sirtuin activity in different cell compartments, either cytoplasm, or mitochondria, or nucleus. Abbreviations: CR, calorie restriction; PI3K, phosphatidylinositol-3-kinase; AKT, protein kinase B; TOR, target of rapamycin; AMPK, AMP-activated protein kinase; FOXO, forkhead box O transcription factor; GLUT-4, glucose transporter type 4; PEPCK1, phosphoenolpyruvate carboxykinase 1; PGC1 α , proliferator-activated receptor-gamma coactivator-1 α ; IR, insulin receptor; IGF1, insulin-like growth factor1; IGFR, insulin-like growth factor receptor.

Aerobic metabolism is the most efficient energy-producing process in living organisms, thus, CR improves metabolic efficiency, an effect that fits very well with the CR-associated lower metabolic rate. Moreover, CR enhances fatty acid oxidation, while reducing carbohydrate oxidation [49,50].

Beside regulating cellular activities at the cytoplasmic and organelle levels, sirtuins also mediate the response to CR at the nuclear level, although some issues remain still open, such as the relationship between CR and telomere length. Telomere shortening is an important biomarker of cell senescence and CR has been demonstrated to decrease the rate of telomere shortening in various mouse tissues during ageing [44]. In particular, CR enhances the activity of telomerase, probably through the regulation of telomere-associated proteins [51–53]. On the contrary, the effects of CR on telomere length has been questioned by other studies in humans and rhesus monkeys [54,55]. DNA integrity is another target of the nuclear activity of sirtuins, following CR. DNA integrity is fundamental for the health of cells and organisms and is guaranteed by the protection from DNA damaging agents and the activity of DNA repair systems [56]. CR both reduces DNA damage from oxidative stress and stimulates DNA repair by

supporting the expression of genes involved in the anti-oxidant defense and in the nucleotide excision repair process [57–60]. Therefore, experimental evidence so far collected supports the hypothesis that CR hinders the age-related accumulation of DNA damage and reduction in DNA repair capacity.

The effects of ageing are exacerbated also by the progressive reduction of stem cell activity and, thus, by the reduced regeneration potential of tissues. Tissue regeneration is a delicate and complex process allowing the maintenance of body integrity, wellness and suitable lifespan [61–65]. In this regard, it has been shown that CR promotes also the activity of stem cells, thus preserving their regenerative potential. Studies on mice suggest that CR sustains the health and survival of stem cells, allowing the maintenance of adequate stem cell pools in different tissues [34,66–68]. Indeed, CR reduces hematopoietic stem cell (HSC) ageing and increases HSC quiescence, thus enhancing the repopulation capacity of HSCs [69]. Moreover, CR augments the number of neural stem cells, improves their function and their niche. These effects lead to an enhanced regeneration potential of the intestine [71]. In skeletal muscle, CR improves satellite stem cell availability and activity, thus promoting a more effective recovery from injury [72]. The molecular mechanisms involved in the stem cell response to CR include mTOR, AMPK/LKB1, FOXOs, sirtuins, IGF-I, PI3K/AKT; PTEN and the authophagic machinery [68,73].

The comprehension of the molecular pathways promoting both health span and lifespan, has led to the identification of natural and synthetic compounds with anti-ageing effects. This is the case of resveratrol, a polyphenol from grapes and the anti-diabetic drug metformin. These molecules have been demonstrated to be sirtuin activators and to hinder the deleterious effects of ageing by modulating some of the same molecular pathways activated by CR [74–84].

3. CR and Sirtuins in Skeletal Muscle

In mammals, skeletal muscle undergoes a progressive age-dependent loss of mass, function and regenerative potential [85]. This process contributes to a continued metabolic derangement, a decay in the quality of life and a consequent reduction of health- and lifespan. The function of skeletal muscle in the whole body extends far beyond the locomotion. In fact, this tissue acts also as an amino acid stock for protein synthesis, contributes to glucose uptake from the bloodstream, and secretes cytokines and growth factors (myokines) into the bloodstream, thus regulating the activity of other tissues [85]. The progressive loss of strength, which is typical of the ageing muscle, is also related to the compromised function and reduced number of satellite, stem, and progenitor cells. This also leads to a reduced muscle repair and regeneration potential in aged organisms [48,68]. Indeed, skeletal muscle is very susceptible to changes in nutrient availability and in vitro, the availability of glucose and amino acid strongly influences the myogenic potential of skeletal muscle satellite cells [86,87]. CR delays the age-dependent reduction in skeletal muscle mass, as demonstrated in animal models, and this effect is mediated by the protection of DNA and mitochondria from damage, stimulation of autophagy, and inhibition of inflammation [88–91]. During CR, cells undergo a transcriptional shift from the activation of genes promoting growth to those supporting maintenance/repair [92]. In this regard, the analysis of gene expression in skeletal muscle of aged mice under CR revealed an upregulation of genes involved protein turnover with a concomitant lower level of macromolecular damages [93].

Interestingly, some muscular effects of CR are also depending on age, sex and genetics [82,93–95]. In particular, dietary condition has been seen to influence ageing-related changes in gene expression of mouse skeletal muscle only if it occurs at the juvenile, but not the adult stage [82,93,94]. Furthermore, CR counteracts the age-dependent increase in systemic chronic inflammation and insulin resistance by modulating the glutathione redox status, nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB), SIRT1, peroxisome proliferator-activated receptors (PPARs), and forkhead box O (FOXOs) [96,97]. This is particularly relevant because age-related chronic inflammation negatively influences the regenerative potential of skeletal muscle and skeletal muscle mass [48,98]. Beside reduced inflammation, CR-treated mice had increased plasma cortisol level, and enhanced

levels of chaperones and autophagy complexes involved in protein folding and turnover [47]. Although elevated cortisol is associated with adverse effects, including increased protein catabolism, this hormone is also a powerful anti-inflammatory agent. Moreover, humans do not respond to CR with a biologically relevant increase of serum cortisol [99]. In skeletal muscle, ageing is also associated with increased DNA damage, inhibition of mitochondrial function and reduced energy metabolism and muscle performances. In particular, the increased activity of the DNA-dependent protein kinase in aged organisms is responsible for the metabolic dysregulation and the decay of physical performances [100].

Intriguingly, CR can synergize with physical exercise in reducing the negative effects of ageing on muscular mass and efficiency. Physical workout is the major stimulus for muscle trophism and must go together with adequate supply of nutrients, particularly essential aminoacids and vitamins in order to maintain or increase muscular efficiency. Therefore, it is important to recall the significance of CR in the present context, which is a reduction of calorie intake in the absolute absence of any deficiency of essential nutrients. Having said that, CR, together with physical exercise, can be effective in hindering the age-related changes in sarcolemmal proteins, and therefore, contrasting the decline in muscle performances of rats. In particular, this observation refers to the membrane repair proteins MG53, dysferlin, annexin A6, and annexin A2 and Nox2 subunits [101]. However, the effects of CR and exercise on muscle are different. Indeed, CR was able to increase insulin-stimulated glucose uptake in the soleus muscle of old rats, whereas physical exercise did not [102]. A similar effect of CR in rat skeletal muscle was observed in another study, without changes in the content of mitochondrial hexokinase II, or electron transport chain and oxidative phosphorylation proteins [103].

There are several other pathways that can be influenced by CR in the muscle and that can positively affect this tissue. A major one recognizes as central player mTOR, a kinase that regulates skeletal muscle growth, by phosphorylating P70S6 kinase 1 (S6K1) [104–106]. It is also worth mentioning IGF-1, since in aged skeletal muscle circulating IGF-1 and its downstream intracellular signaling are reduced. In particular, there is a reduction in the activity of phosphoinositide 3-kinase, AKT, mTOR, p70S6K1, 4E-BP1 and eukaryotic translation initiation factor 2B [55,107–110]. Interestingly, this reduction of IGF-1 activity is associated with both reduction of skeletal muscle mass and with increased life- and health-spans [111].

The tapering muscular function, which characterizes ageing and chronic diseases, is influenced by nutrient availability, redox state, and the ratio of NAD⁺ to NADH. Therefore, SIRT1 plays a crucial role in the balance between healthy and defective skeletal muscle. Indeed, in vitro studies demonstrated that SIRT1 expression prompts an anti-oxidative response in skeletal muscle cells after mechanical stress [112]. Moreover, SIRT1 and AMPK are crucial mediators of the effects of CR in mouse skeletal muscle [113].

Exposure of cultured myoblasts to glucose restriction blocks their differentiation into myotubes and increases the activity of SIRT1 by AMPK-dependent regulation of nicotinamide phosphoribosyltransferase (NAMPT), the enzyme responsible for NAD⁺ turnover [114,115]. In myotubes cultured in the presence of low glucose concentration, SIRT1 is necessary for the metabolic switch from glucose utilization to fatty acid oxidation; this important response is present during CR and is reduced during metabolic diseases [116].

SIRT1 produces these and other effects by acting on/with its substrates and transcriptional/epigenetic co-factors, including PGC-1 α , hepatocyte nuclear factor 4 alpha, p53, FOXOs, sterol regulatory element-binding protein 1 (SREBP-1C) PPAR γ , NF- κ B, Ku70, P300/CBP-associated factor, myogenic differentiation factor (MYOD), myocyte enhancer factor-2 (MEF2), signal transducer and activator of transcription 3, heat shock transcription factor 1, Smad7, suppressor of variegation 3-9 homolog 1, enhancer of zeste 2 polycomb repressive complex 2 subunit, nucleomethylin, eNOSC and various histones [117–120]. SIRT1 deacetylates SREBP-1C, thus enhancing its activity and the expression of its target genes in mouse skeletal muscle [121]. Moreover, SIRT1 participates in the response to injury in muscle, by modulating both NF- κ B and FOXOs and regulating MYOD and MEF2. In fact,

NF- κ B and FOXOs regulate the muscle-specific RING finger protein 1 and the muscle atrophy F-box protein/atrogin-1, which are E3 ubiquitin ligases involved in proteasome-mediated proteolysis of muscle proteins [122].

Gene expression in skeletal and heart muscle is regulated in a circadian fashion by the CLOCK:BMAL1 transcription factor, that controls the expression of other clock genes. This endogenous rhythm can be influenced by external stimuli, such as light and food availability that act on the hypothalamic suprachiasmatic nucleus [123]. SIRT1 deacetylates CLOCK and BMAL1 in a circadian fashion in mouse embryonic fibroblasts and is an important component of the circadian clock. NAD⁺ plays a central role in both metabolism and circadian rhythm and controls the NAMPT production through SIRT1-mediated CLOCK and BMAL1 activation. In the same way, the circadian clock controls intracellular NAD⁺ levels by a transcriptional feedback loop [123]. Moreover, SIRT1 is also able to promote proliferation of muscle cell precursors by blocking the expression of cell cycle inhibitors and hindering their transformation into fully differentiated myocytes [111,124,125]. This observation has been made in cultured murine myotubes and has been associated with SIRT1-mediated repression of MYOD, a major muscle transcriptional regulator, and of MEF2 [114,126]. In turn, repression of MYOD downregulates the expression of myogenin and several contractile proteins [114].

SIRT1 could be important for the regeneration of skeletal muscle and heart, by inducing proliferation and differentiation of their respective adult stem cells. Moreover, the activity of SIRT1 on FOXO transcription factors regulates the catabolic pathways of proteins, blocks apoptosis and promotes oxidative stress resistance and DNA repair [127]. In chronic inflammation or physical inactivity, SIRT1 sustains skeletal muscle survival, thus counteracting muscular atrophy [111]. Most importantly, CR (particularly glucose restriction), but also physical exercise enhance SIRT1 expression through AMPK, FOXO3A and EGR1 activities, that, in turn, leads to activation of FOXO4 and PGC-1α. These factors induce the oxidative metabolism and an anti-oxidative response in muscle cells [126].

SIRT1 and 6 are important downregulators of IGF-1 and its target signaling complex AKT/mTOR [128]. This mechanism engages SIRT6 into the homeostasis of the muscle since it has been demonstrated that the alteration of the AKT-mTOR pathway is involved in the evolution of sarcopenia [129]. In addition, SIRT6, together with SIRT1 inhibits NF-κB, thus attenuating its inflammatory signaling cascade and contrasting DNA damage, cellular senescence and oxidative stress [130–132].

SIRT2 has been demonstrated to be responsible for the reduction of glucose transporter 4 translocation to the sarcolemma in response to insulin in transgenic mice [133]. The expression of SIRT2 is negatively correlated with insulin sensitivity, as demonstrated in vitro in C_2C_{12} skeletal muscle cells [134]. Interestingly, SIRT2 is also related to CR since it regulates metabolism during fasting by deacetylating phosphoenolpyruvate carboxykinase [135].

CR upregulates SIRT3 in skeletal muscle [136]. This sirtuin is localized within the mitochondria where it modulates the activity of several proteins, such as succinate-dehydrogenase, manganese-dependent superoxide dismutase (Mn-SOD) and FOXO3. The absence of SIRT3 leads to a downregulation of PGC-1 α [136]. SIRT3 is also involved in the regulation of acetate metabolism that is relevant during ageing and fasting or starvation [137–139]. In particular, the enzymes acetyl-CoA synthase type 1 and 2 are activated by SIRT3 [137–139]. This activation is associated with increased lifespan in yeast [140]. During fasting, in skeletal muscle, the expression of SIRT3 is increased and the SIRT1-dependent deacetylation of PGC-1 α is increased, thus, enhancing mitochondrial oxidative phosphorylation [116,136,141].

The evidence relating SIRT4 and 7 to muscle pathophysiology and CR is limited. It has been demonstrated in mouse skeletal muscle that AMPK regulates metabolism by modulating SIRT4 and 1 activity [142]. Another study analyzed the expression of SIRT7 in young and old rats and observed tissue-dependent changes in gene expression occurring during ageing and during periods of CR [143].

Interestingly, although several studies demonstrated a positive effect of CR on skeletal muscle, a recent study on mice provided contrasting data, thus, suggesting caution in considering the

advantages of CR on skeletal muscle health. Moreover, long-term, CR reduces the amount of protein in rat skeletal muscle through reduction of the mTORC1 pathway [144].

4. CR and Sirtuins in Cardiac Muscle

In the last decades, evidence has been provided about the beneficial effects of CR on the cardiovascular system. These effects, as in other tissues, are mediated by the activation of the sirtuin genes [48] However, the expression and activity of several sirtuins is decreased, and protein acetylation increased, in the presence of cardiovascular diseases [145]. Sirtuins can affect cardiac and endothelial cells either directly or indirectly by systemic regulation. Indeed, SIRT1 has been demonstrated to be protective against endothelial dysfunction, atherothrombosis, and myocardial infarction [146]. Interestingly, the interplay between sirtuins and beta-adrenergic receptors has been hypothesized, thus opening a new scenario for the molecular changes occurring in the heart during ageing and diseases [147].

SIRT1, 2, 3 and 7 regulate cell survival in stressing or unfavorable conditions, such as ischemia/reperfusion (I/R) injury, ageing, and increase of oxidative stress [148]. SIRT3 and 6 hinder cardiac hypertrophy and heart failure [148]. Short-term CR in rats increases the expression of SIRT1–4 and 7 [149].

CR activates the autophagic machinery, which is important for the survival and the resistance of cardiomyocytes to stress. However, the efficacy of SIRT1-induced autophagy progressively deteriorates with ageing leading to a progressive decline of cardiomyocyte health and ischemic resistance, and, consequently, to heart dysfunction and failure [150,151]. This important mechanism is regulated by the SIRT1/FOXO1 pathway following nutrient deprivation.

In general, oxidative stress is a very important factor in the onset of cardiovascular disease as a consequence of its detrimental effects on cardiac and vascular cells, and of the oxidative modifications of lipoproteins that potently accelerate atherosclerosis [152]. Therefore, it is worth mentioning the well-documented contribution of CR to cellular anti-oxidative activity. Moreover, CR exerts beneficial effects on the cardiovascular system by reducing inflammation, ameliorating the insulin sensitivity and preventing cardiomyocyte apoptotic death [152]. The connection among CR, reduced inflammation, and heart benefits is reinforced by stimulation of ghrelin signaling, which has a powerful anti-inflammatory activity and leads to increased SIRT1 protein expression in mouse heart, thus, ameliorating the conditions of aged hearts. Therefore, it has been proposed that CR exerts its beneficial effects also through ghrelin signaling [153].

CR may protect heart function by also controlling adverse rises of blood pressure and by empowering the resilience of cardiomyocytes to pro-apoptotic conditions. It has been reported that CR has antihypertensive and cardioprotective effects against I/R injury in mice and rats. These effects are mediated by the activity of nitric oxide which, in turn, activates SIRT1 and its pathway [152,154,155]. In particular, the protective effect against I/R injury depends on the activation of the cardiac complement component 3 [154].

The beneficial effects of CR against the deleterious effects of ageing are mediated by inhibition of mTOR signaling, and enhancement of the activity of sirtuins and KLOTHO [156]. It has been suggested that the interplay among these three pathways could be relevant in the contest of anti-ageing approaches [156]. There is a definite time window for the healthy effects of CR. Indeed, it has been recently demonstrated that CR is beneficial only when initiated in middle or old age in mice [157]. In mice under CR, FOXO1 mediates anti-oxidant, autophagic, and anti-apoptotic responses by cardiac cells [52,53]. More interestingly, FOXO1 has been suggested as a positive regulator of telomerase activity without telomere shortening [52,53]. The length of telomeres is a marker of cell senescence, and its attrition is associated with a decay in the function and the proliferative potential of cells. This phenomenon also occurs in cardiac cells, thus resulting in the development of heart diseases [53,158,159]. CR is able to modulate the activity of telomerase and to reduce the shortening of telomeres [53,158,159].

SIRT1 expression is sensitive to environmental changes and to stress, but its response varies according to different triggering factors [148]. Moreover, SIRT1 activity is also strongly dependent

on post-translational modifications [160,161]. Interestingly, the role of SIRT1 in mouse heart is strictly dose-dependent. Indeed, moderate cardiac SIRT1 expression is associated with protection from senescence, hypertrophy, apoptosis, fibrosis, heart dysfunction and oxidative stress; on the contrary, high SIRT1 expression leads to an increase of oxidative stress, apoptosis and hypertrophy and cardiac dysfunction [162].

It has been demonstrated (in vivo) that the expression and the activity of SIRT1 must be strictly controlled to end up with a specific cardiomyocyte response [163]. In fact, 2.5–7.5 cardiac overexpression of SIRT1 is beneficial for the heart, hindering the age-dependent hypertrophy and reducing the detrimental effects of I/R injury. The molecular mechanism underlying this effect is the activation of protective genes, such as Mn-SOD, thioredoxin 1, and BCL2-like protein and the downregulation of proapoptotic genes such as BCL2-associated X protein [164].

On the contrary, a marked overexpression of SIRT1 (12.5 times) in heart is associated with mitochondrial dysfunction, low ATP content, decreased citrate synthase activity, increased oxidative stress, and significantly reduced PGC-1 α expression, thus leading to cardiac dysfunction [162]. Thus, in heart, a controlled SIRT1 expression could attenuate the detrimental effects of ageing, although the total ablation of SIRT1 expression, as realized in SIRT1 knockout (KO) mice, could reduce the negative heart remodeling in response to hypertrophic stimuli [165]. The molecular mechanisms underlying the effects of SIRT1 on cardiac hypertrophic response to pressure overload implicate various molecules and pathways, including PPAR α , estrogen-related receptors, AKT and pyruvate dehydrogenase kinase 1 (PDK1) [165,166].

Besides SIRT1, SIRT2, 3, 6 and 7 have been involved in heart pathophysiology. Although the role of SIRT2 in the heart is not well characterized, some reports indicate that this protein is involved in the development of the heart and suggest that it may be also involved in the development of cardiac hypertrophy by activating AKT signaling [163].

SIRT3 has been demonstrated to be a key factor in the anti-oxidant effect of CR [146]. This protein exerts its deacetylase activity on mitochondrial proteins and regulates the turnover of mitochondria. Experiments on SIRT3 KO mice demonstrated that this protein is involved in the regulation of proteins of the ATP production machinery and in the age-dependent development of hypertrophy in the heart [163]. In this context, SIRT3 promotes resistance to the onset of hypertrophy, cardiomyopathy, oxidative stress damage, and metabolic dysregulation [146].

The anti-hypertrophic activity of SIRT3 is mediated by the Mn-SOD and catalase genes for their effect in diminishing the oxidative stress; by cyclophilin D, which triggers the mitochondrial permeability transition pore opening, and thus cell death; by liver kinase B1-dependent activation of AMPK [167–169]. Moreover, the anti-ageing activity of SIRT3 relies also on the glycogen synthase kinase 3 beta-mediated block of transforming growth factor beta 1 signaling and its fibrotic activity [170]. Collectively, all data seem to indicate that SIRT3 activity has beneficial effects on the heart by improving mitochondrial activity and health.

Differently from other sirtuins, the evidence relating SIRT4 and 5 with heart pathophysiology is scarce, mainly relies on in vitro studies, and suggests a protective role of these sirtuins in cardiac cells [171–173]. These sirtuins regulate mitochondrial metabolism and are involved in cellular energy and redox balance in cardiac cells [174].

Concerning SIRT6, it hinders dyslipidemia, cell senescence, and cardiac hypertrophy [146,175]. A very recent evidence indicates that SIRT6 regulates the transcription of PDK4 via FOXO1. This, in turn, has a detrimental effect on mitochondrial activity [176].

As shown in SIRT7 KO mice, this protein is implicated in the cardiac protection from hypertrophy and inflammation-mediated fibrosis [177]. The mechanism is based on the activation of AKT pathway [177]. Moreover, SIRT7 is involved in regulating lipid metabolism and contrasting the onset of cardiac diseases [146].

Apart from the experimental evidence supporting a positive role of CR on the health of the cardiovascular system, there are also some warnings in the scientific community regarding the

indiscriminate application of CR regimen in humans. In fact, it has been reported in a 29 years old woman that a CR regimen can exacerbate the myocardial stress after high-intensity exercise [178].

5. Conclusions

Sirtuins are critical mediators of the beneficial effect of CR on signs of ageing and diseases. In this context, SIRT1 plays a major role compared to the other members of the sirtuin family. So far, the mechanisms underlying the mechanisms associated with detrimental effects of ageing are not fully elucidated. However, the possibility to affect some negative consequences of ageing in different species, including humans, is seriously entertained within the scientific community and nutritional interventions are some of the most studied strategies to promote a healthy life. In particular, experimental evidence indicates a positive role of CR in affecting the decay of the organismal performances with ageing; thus, we believe that CR can contribute to increase health span in many species, and most probably also in humans. To reach this goal CR should preserve the efficiency of skeletal and heart muscles, which is fundamental for the wellbeing of vertebrates in general, and humans in particular, and is compromised during ageing and in age-related diseases. However, CR might not be a strategy that is easily applicable by many, especially in the long run, because it is very demanding for many. Therefore, the present and future knowledge about the role of sirtuins in muscle tissues, especially under conditions of CR, might offer alternative strategies to dietary restrictions since many compounds that stimulate sirtuins and the same anti-ageing pathways induced by CR, are being identified and developed.

Acknowledgments: This work was financially supported by University of Sannio.

Author Contributions: Alberto Zullo, Emanuela Simone, Maddalena Grimaldi, Vincenzina Musto and Francesco Paolo Mancini drafted and revised the manuscript. All authors read and approved the final manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

AE BD1	aIE/E hinding protoin1
4E-DF1	err4E-binding protein
AMPK	AMP-activated protein kinase
CR	Calorie restriction
eNOS	Endothelial nitric oxide synthase
FOXOs	Forkhead box O
GLUT-4	Glucose transporter type 4
HSC	Hematopoietic stem cell
I/R	Ischemia/reperfusion
IGF-1	Insulin growth factor type 1
КО	Knockout
MEF2	Myocyte enhancer factor-2
Mn-SOD	Manganese-dependent superoxide dismutase
mTOR	Mammalian target of rapamycin
MYOD	Myogenic differentiation factor
NAMPT	Nicotinamide phosphoribosyltransferase
NF-ĸB	Nuclear factor kappa-light-chain-enhancer of activated B cells
PDK	Pyruvate dehydrogenase kinase
PEPCK1	Phosphoenolpyruvate carboxykinase 1
PGC-1α	Peroxisome proliferator-activated receptor gamma coactivator 1-alpha
PPAR	Peroxisome proliferator-activated receptor
ROS	Reactive oxygen species
S6K1	P70S6 kinase 1
SIRT	Sirtuin
SREBP-1C	Sterol regulatory element-binding protein 1

References

- 1. De Nigris, F.; Balestrieri, M.L.; Napoli, C. Targeting C-Myc, Ras and IGF Cascade to treat cancer and Vascular Disorders. *Cell Cycle* **2006**, *5*, 1621–1628. [CrossRef] [PubMed]
- 2. Keys, A. Coronary Heart Disease in Seven Countries. Circulation 1970, 41, 1–211. [CrossRef]
- 3. Mancini, M.; Stamler, J. Diet for preventing cardiovascular diseases: Light from Ancel Keys, Distinguished Centenarian Scientist. *Nutr. Metab. Cardiovasc. Dis.* **2004**, *14*, 52–57. [CrossRef]
- 4. World Health Organization. Available online: http://www.Who.Int/Mediacentre/Factsheets/Fs317/En/ (accessed on 2 February 2018).
- 5. Keys, A. Mediterranean Diet and Public Health: Personal Reflections. *Am. J. Clin. Nutr.* **1995**, *61* (Suppl. 6), 1321s–1323s. [CrossRef] [PubMed]
- 6. Rubba, P.; Mancini, F.P.; Gentile, M.; Mancini, M. The Mediterranean Diet in Italy: An Update. *World Rev. Nutr. Diet.* **2007**, *97*, 85–113. [PubMed]
- Balestrieri, M.L.; Fiorito, C.; Crimi, E.; Felice, F.; Schiano, C.; Milone, L.; Casamassimi, A.; Giovane, A.; Grimaldi, V.; Del Giudice, V.; et al. Effect of Red Wine Antioxidants and Minor Polyphenolic Constituents on Endothelial Progenitor Cells after Physical Training in Mice. *Int. J. Cardiol.* 2008, *126*, 295–297. [CrossRef] [PubMed]
- Sacks, F.M.; Obarzanek, E.; Windhauser, M.M.; Svetkey, L.P.; Vollmer, W.M.; Mccullough, M.; Karanja, N.; Lin, P.H.; Steele, P.; Proschan, M.A. Rationale and Design of the Dietary Approaches to Stop Hypertension Trial (Dash). A Multicenter Controlled-Feeding Study of Dietary Patterns to Lower Blood Pressure. *Ann. Epidemiol.* 1995, 5, 108–118. [CrossRef]
- Fung, T.T.; Hu, F.B.; Wu, K.; Chiuve, S.E.; Fuchs, C.S.; Giovannucci, E. The Mediterranean and Dietary Approaches to Stop Hypertension (Dash) Diets and Colorectal Cancer. *Am. J. Clin. Nutr.* 2010, *92*, 1429–1435. [CrossRef] [PubMed]
- 10. Salehi-Abargouei, A.; Maghsoudi, Z.; Shirani, F.; Azadbakht, L. Effects of Dietary Approaches to Stop Hypertension (Dash)-Style Diet on Fatal or Nonfatal Cardiovascular Diseases—Incidence: A Systematic Review and Meta-Analysis on Observational Prospective Studies. *Nutrition* **2013**, *29*, 611–618. [CrossRef] [PubMed]
- Jenkins, D.J.A.; Kendall, C.W.C.; Marchie, A.; Faulkner, D.A.; Wong, J.M.W.; De Souza, R.; Emam, A.; Parker, T.L.; Vidgen, E.; Lapsley, K.G.; et al. Effects of a Dietary Portfolio of Cholesterol-Lowering Foods vs. Lovastatin on Serum Lipids and C-Reactive Protein. *JAMA* 2003, *290*, 502–510. [CrossRef] [PubMed]
- 12. Kendall, C.W.C.; Jenkins, D.J.A. A Dietary Portfolio: Maximal Reduction of Low-Density Lipoprotein Cholesterol with Diet. *Curr. Atheroscler. Rep.* **2004**, *6*, 492–498. [CrossRef] [PubMed]
- Sinclair, D.A. Toward a Unified Theory of Caloric Restriction and Longevity Regulation. *Mech. Ageing Dev.* 2005, 126, 987–1002. [CrossRef] [PubMed]
- 14. Fontana, L.; Partridge, L. Promoting Health and Longevity Through Diet: From Model Organisms to Humans. *Cell* **2015**, *161*, 106–118. [CrossRef] [PubMed]
- 15. Mattison, J.A.; Roth, G.S.; Beasley, T.M.; Tilmont, E.M.; Handy, A.M.; Herbert, R.L.; Longo, D.L.; Allison, D.B.; Young, J.E.; Bryant, M.; et al. Impact of Caloric Restriction on Health and Survival in Rhesus Monkeys from the Nia Study. *Nature* **2012**, *489*, 318–321. [CrossRef] [PubMed]
- Colman, R.J.; Beasley, T.M.; Kemnitz, J.W.; Johnson, S.C.; Weindruch, R.; Anderson, R.M. Caloric Restriction Reduces Age-Related and All-Cause Mortality in Rhesus Monkeys. *Nat. Commun.* 2014, *5*, 3557. [CrossRef] [PubMed]
- 17. Trepanowski, J.F.; Canale, R.E.; Marshall, K.E.; Kabir, M.M.; Bloomer, R.J. Impact of Caloric and Dietary Restriction Regimens on Markers of Health and Longevity in Humans and Animals: A Summary of Available Findings. *Nutr. J.* **2011**, *10*, 107. [CrossRef] [PubMed]
- Romey-Glüsing, R.; Li, Y.; Hoffmann, J.; Von Frieling, J.; Knop, M.; Pfefferkorn, R.; Bruchhaus, I.; Fink, C.; Roeder, T. Nutritional Regimens with Periodically Recurring Phases of Dietary Restriction Extend Lifespan in *Drosophila*. *FASEB J.* 2018. [CrossRef] [PubMed]
- 19. Min, K.; Flatt, T.; Kulaots, I.; Tatar, M. Counting Calories in *Drosophila* Diet Restriction. *Exp. Gerontol.* **2007**, 42, 247–251. [CrossRef] [PubMed]
- Lawler, D.F.; Larson, B.T.; Ballam, J.M.; Smith, G.K.; Biery, D.N.; Evans, R.H.; Greeley, E.H.; Segre, M.; Stowe, H.D.; Kealy, R.D. Diet Restriction and Ageing in the Dog: Major Observations over Two Decades. *Br. J. Nutr.* 2008, *99*, 793–805. [CrossRef] [PubMed]

- 21. Comfort, A. Effect of Delayed and Resumed Growth on the Longevity of a Fish (*Lebistes reticulatus*, Peters) in Captivity. *Gerontologia* **1963**, *49*, 150–155. [CrossRef] [PubMed]
- 22. Mair, W.; Piper, M.D.W.; Partridge, L. Calories Do Not Explain Extension of Life Span by Dietary Restriction in *Drosophila*. *PLoS Biol*. **2005**, *3*, E223. [CrossRef] [PubMed]
- 23. Willcox, D.C.; Willcox, B.J.; Todoriki, H.; Curb, J.D.; Suzuki, M. Caloric Restriction and Human Longevity: What Can We Learn from the Okinawans? *Biogerontology* **2006**, *7*, 173–177. [CrossRef] [PubMed]
- 24. Kagawa, Y. Impact of Westernization on the Nutrition of Japanese: Changes in Physique, Cancer, Longevity and Centenarians. *Prev. Med. (Baltim)* **1978**, *7*, 205–217. [CrossRef]
- 25. Feldman, J.L.; Dittenhafer-Reed, K.E.; Denu, J.M. Sirtuin Catalysis and Regulation. *J. Biol. Chem.* **2012**, *287*, 42419–42427. [CrossRef] [PubMed]
- 26. Bales, C.W.; Kraus, W.E. Caloric Restriction: Implications for Human Cardiometabolic Health. J. Cardiopulm. *Rehabil. Prev.* **2013**, *33*, 201–208. [CrossRef] [PubMed]
- 27. Mccay, C.M.; Crowell, M.F.; Maynard, L.A. The Effect of Retarded Growth upon the Length of Life Span and upon the Ultimate Body Size. *Nutrition* **1935**, *5*, 155–171. [CrossRef]
- 28. Ross, M.H. Length of Life and Nutrition in the Rat. J. Nutr. 1961, 75, 197–210. [CrossRef] [PubMed]
- 29. Yu, B.P.; Masoro, E.J.; Murata, I.; Bertrand, H.A.; Lynd, F.T. Life Span Study of SPF Fischer 344 Male Rats Fed ad Libitum or Restricted Diets: Longevity, Growth, Lean Body Mass and Disease. *J. Gerontol.* **1982**, *37*, 130–141. [CrossRef] [PubMed]
- 30. Weindruch, R.; Walford, R.L.; Fligiel, S.; Guthrie, D. The Retardation of Aging in Mice by Dietary Restriction: Longevity, Cancer, Immunity and Lifetime Energy Intake. J. Nutr. **1986**, 116, 641–654. [CrossRef] [PubMed]
- Kapahi, P.; Kaeberlein, M.; Hansen, M. Dietary Restriction and Lifespan: Lessons from Invertebrate Models. Ageing Res. Rev. 2017, 39, 3–14. [CrossRef] [PubMed]
- 32. Colman, R.J.; Anderson, R.M. Nonhuman Primate Calorie Restriction. *Antioxid. Redox Signal.* 2011, 14, 229–239. [CrossRef] [PubMed]
- Bodkin, N.L.; Alexander, T.M.; Ortmeyer, H.K.; Johnson, E.; Hansen, B.C. Mortality and Morbidity in Laboratory-Maintained Rhesus Monkeys and Effects of Long-Term Dietary Restriction. *J. Gerontol. A Biol. Sci. Med. Sci.* 2003, *58*, 212–219. [CrossRef] [PubMed]
- 34. Mattison, J.A.; Colman, R.J.; Beasley, T.M.; Allison, D.B.; Kemnitz, J.W.; Roth, G.S.; Ingram, D.K.; Weindruch, R.; De Cabo, R.; Anderson, R.M. Caloric Restriction Improves Health and Survival of Rhesus Monkeys. *Nat. Commun.* **2017**, *8*, 14063. [CrossRef] [PubMed]
- 35. Strom, A.; Jensen, R.A. Mortality from Circulatory Diseases in Norway 1940–1945. *Lancet (Lond. Engl.)* **1951**, 1, 126–129. [CrossRef]
- Malmros, H. The Relation of Nutrition to Health; A Statistical Study of the Effect of the War-Time on Arteriosclerosis, Cardiosclerosis, Tuberculosis and Diabetes. *Acta Med. Scand. Suppl.* 1950, 246, 137–153. [PubMed]
- 37. Suzuki, M.; Wilcox, B.J.; Wilcox, C.D. Implications from and for Food Cultures for Cardiovascular Disease: Longevity. *Asia Pac. J. Clin. Nutr.* **2001**, *10*, 165–171. [CrossRef] [PubMed]
- 38. Fontana, L.; Klein, S. Aging, Adiposity, and Calorie Restriction. JAMA 2007, 297, 986. [CrossRef] [PubMed]
- 39. Masoro, E.J.; Yu, B.P.; Bertrand, H.A. Action of Food Restriction in Delaying the Aging Process. *Proc. Natl. Acad. Sci. USA* **1982**, *79*, 4239–4241. [CrossRef] [PubMed]
- 40. Barger, J.L.; Walford, R.L.; Weindruch, R. The Retardation of Aging By Caloric Restriction: Its Significance in the Transgenic Era. *Exp. Gerontol.* **2003**, *38*, 1343–1351. [CrossRef] [PubMed]
- Selman, C.; Phillips, T.; Staib, J.L.; Duncan, J.S.; Leeuwenburgh, C.; Speakman, J.R. Energy Expenditure of Calorically Restricted Rats Is Higher Than Predicted from Their Altered Body Composition. *Mech. Ageing Dev.* 2005, 126, 783–793. [CrossRef] [PubMed]
- 42. Most, J.; Tosti, V.; Redman, L.M.; Fontana, L. Calorie Restriction in Humans: An Update. *Ageing Res. Rev.* **2017**, *39*, 36–45. [CrossRef] [PubMed]
- 43. Vassilopoulos, A.; Fritz, K.S.; Petersen, D.R.; Gius, D. The Human Sirtuin Family: Evolutionary Divergences and Functions. *Hum. Genomics* **2011**, *5*, 485–496. [CrossRef] [PubMed]
- 44. Ota, H.; Eto, M.; Kano, M.R.; Kahyo, T.; Setou, M.; Ogawa, S.; Iijima, K.; Akishita, M.; Ouchi, Y. Induction of Endothelial Nitric Oxide Synthase, SIRT1, and Catalase By Statins Inhibits Endothelial Senescence through the Akt Pathway. *Arterioscler. Thromb. Vasc. Biol.* **2010**, *30*, 2205–2211. [CrossRef] [PubMed]

- 45. Takizawa, Y.; Kosuge, Y.; Awaji, H.; Tamura, E.; Takai, A.; Yanai, T.; Yamamoto, R.; Kokame, K.; Miyata, T.; Nakata, R.; et al. Up-Regulation of Endothelial Nitric Oxide Synthase (Enos), Silent Mating Type Information Regulation 2 Homologue 1 (SIRT1) and Autophagy-Related Genes By Repeated Treatments with Resveratrol in Human Umbilical Vein Endothelial Cells. *Br. J. Nutr.* **2013**, *110*, 2150–2155. [CrossRef] [PubMed]
- 46. Xia, N.; Strand, S.; Schlufter, F.; Siuda, D.; Reifenberg, G.; Kleinert, H.; Förstermann, U.; Li, H. Role of SIRT1 and Foxo Factors in Enos Transcriptional Activation By Resveratrol. *Nitric Oxide* **2013**, *32*, 29–35. [CrossRef] [PubMed]
- 47. Yang, L.; Vaitheesvaran, B.; Hartil, K.; Robinson, A.J.; Hoopmann, M.R.; Eng, J.K.; Kurland, I.J.; Bruce, J.E. The Fasted/Fed Mouse Metabolic Acetylome: N6-Acetylation Differences Suggest Acetylation Coordinates Organ-Specific Fuel Switching. *J. Proteome Res.* **2011**, *10*, 4134–4149. [CrossRef] [PubMed]
- 48. López-Lluch, G.; Navas, P. Calorie Restriction as an Intervention in Ageing. *J. Physiol.* **2016**, 594, 2043–2060. [CrossRef] [PubMed]
- Selman, C.; Kerrison, N.D.; Cooray, A.; Piper, M.D.W.; Lingard, S.J.; Barton, R.H.; Schuster, E.F.; Blanc, E.; Gems, D.; Nicholson, J.K.; et al. Coordinated Multitissue Transcriptional and Plasma Metabonomic Profiles Following Acute Caloric Restriction in Mice. *Physiol. Genomics* 2006, 27, 187–200. [CrossRef] [PubMed]
- Jové, M.; Naudí, A.; Ramírez-Núñez, O.; Portero-Otín, M.; Selman, C.; Withers, D.J.; Pamplona, R. Caloric Restriction Reveals a Metabolomic and Lipidomic Signature in Liver of Male Mice. *Aging Cell* 2014, 13, 828–837. [CrossRef] [PubMed]
- 51. Vera, E.; Bernardes De Jesus, B.; Foronda, M.; Flores, J.M.; Blasco, M.A. Telomerase Reverse Transcriptase Synergizes with Calorie Restriction to Increase Health Span and Extend Mouse Longevity. *PLoS ONE* **2013**, *8*, E53760. [CrossRef] [PubMed]
- 52. Makino, N.; Oyama, J.; Maeda, T.; Koyanagi, M.; Higuchi, Y.; Tsuchida, K. Calorie Restriction Increases Telomerase Activity, Enhances Autophagy, and Improves Diastolic Dysfunction in Diabetic Rat Hearts. *Mol. Cell. Biochem.* **2015**, *403*, 1–11. [CrossRef] [PubMed]
- Makino, N.; Oyama, J.; Maeda, T.; Koyanagi, M.; Higuchi, Y.; Shimokawa, I.; Mori, N.; Furuyama, T. Foxo1 Signaling Plays a Pivotal Role in the Cardiac Telomere Biology Responses to Calorie Restriction. *Mol. Cell. Biochem.* 2016, 412, 119–130. [CrossRef] [PubMed]
- 54. Tomiyama, A.J.; Milush, J.M.; Lin, J.; Flynn, J.M.; Kapahi, P.; Verdin, E.; Sinclair, E.; Melov, S.; Epel, E.S. Long-Term Calorie Restriction in Humans Is Not Associated with Indices of Delayed Immunologic Aging: A Descriptive Study. *Nutr. Healthy Aging* **2017**, *4*, 147–156. [CrossRef] [PubMed]
- Cuthbertson, D.; Smith, K.; Babraj, J.; Leese, G.; Waddell, T.; Atherton, P.; Wackerhage, H.; Taylor, P.M.; Rennie, M.J. Anabolic Signaling Deficits Underlie Amino Acid Resistance of Wasting, Aging Muscle. *FASEB J.* 2005, 19, 422–424. [CrossRef] [PubMed]
- 56. Weraarchakul, N.; Strong, R.; Wood, W.G.; Richardson, A. The Effect of Aging and Dietary Restriction on DNA Repair. *Exp. Cell Res.* **1989**, *181*, 197–204. [CrossRef]
- 57. Matt, K.; Burger, K.; Gebhard, D.; Bergemann, J. Influence of Calorie Reduction on DNA Repair Capacity of Human Peripheral Blood Mononuclear Cells. *Mech. Ageing Dev.* **2016**, *154*, 24–29. [CrossRef] [PubMed]
- 58. Langie, S.A.S.; Knaapen, A.M.; Houben, J.M.J.; Van Kempen, F.C.; De Hoon, J.P.J.; Gottschalk, R.W.H.; Godschalk, R.W.L.; Van Schooten, F.J. The Role of Glutathione in the Regulation of Nucleotide Excision Repair during Oxidative Stress. *Toxicol. Lett.* 2007, 168, 302–309. [CrossRef] [PubMed]
- 59. Heydari, A.R.; Unnikrishnan, A.; Lucente, L.V.; Richardson, A. Caloric Restriction and Genomic Stability. *Nucleic Acids Res.* **2007**, *35*, 7485–7496. [CrossRef] [PubMed]
- 60. Vermeij, W.P.; Dollé, M.E.T.; Reiling, E.; Jaarsma, D.; Payan-Gomez, C.; Bombardieri, C.R.; Wu, H.; Roks, A.J.M.; Botter, S.M.; Van Der Eerden, B.C.; et al. Restricted Diet Delays Accelerated Ageing and Genomic Stress in DNA-Repair-Deficient Mice. *Nature* **2016**, *537*, 427–431. [CrossRef] [PubMed]
- 61. Yun, M.H. Changes in Regenerative Capacity through Lifespan. *Int. J. Mol. Sci.* **2015**, *16*, 25392–25432. [CrossRef] [PubMed]
- 62. Sommese, L.; Zullo, A.; Schiano, C.; Mancini, F.P.; Napoli, C. Possible Muscle Repair in the Human Cardiovascular System. *Stem Cell Rev. Rep.* **2017**, *13*, 170–191. [CrossRef] [PubMed]
- 63. Nödl, M.-T.; Fossati, S.M.; Domingues, P.; Sánchez, F.J.; Zullo, L. The Making of an Octopus Arm. *Evodevo* **2015**, *6*, 19. [CrossRef] [PubMed]

- 64. Fossati, S.M.; Candiani, S.; Nödl, M.-T.; Maragliano, L.; Pennuto, M.; Domingues, P.; Benfenati, F.; Pestarino, M.; Zullo, L. Identification and Expression of Acetylcholinesterase in Octopus Vulgaris Arm Development and Regeneration: A Conserved Role for Ache? *Mol. Neurobiol.* 2015, 52, 45–56. [CrossRef] [PubMed]
- 65. Grimaldi, V.; Schiano, C.; Casamassimi, A.; Zullo, A.; Soricelli, A.; Mancini, F.P.; Napoli, C. Imaging Techniques to Evaluate Cell Therapy in Peripheral Artery Disease: State of the Art and Clinical Trials. *Clin. Physiol. Funct. Imaging* **2016**, *36*, 165–178. [CrossRef] [PubMed]
- 66. Brandhorst, S.; Choi, I.Y.; Wei, M.; Cheng, C.W.; Sedrakyan, S.; Navarrete, G.; Dubeau, L.; Yap, L.P.; Park, R.; Vinciguerra, M.; et al. A Periodic Diet that Mimics Fasting Promotes Multi-System Regeneration, Enhanced Cognitive Performance, and Healthspan. *Cell Metab.* **2015**, *22*, 86–99. [CrossRef] [PubMed]
- Choi, I.Y.; Piccio, L.; Childress, P.; Bollman, B.; Ghosh, A.; Brandhorst, S.; Suarez, J.; Michalsen, A.; Cross, A.H.; Morgan, T.E.; et al. A Diet Mimicking Fasting Promotes Regeneration and Reduces Autoimmunity and Multiple Sclerosis Symptoms. *Cell Rep.* 2016, *15*, 2136–2146. [CrossRef] [PubMed]
- 68. Murphy, T.; Thuret, S. The Systemic Milieu as a mediator of dietary influence on stem cell function during ageing. *Ageing Res. Rev.* **2015**, *19*, 53–64. [CrossRef] [PubMed]
- 69. Tang, D.; Tao, S.; Chen, Z.; Koliesnik, I.O.; Calmes, P.G.; Hoerr, V.; Han, B.; Gebert, N.; Zörnig, M.; Löffler, B.; et al. Dietary Restriction Improves Repopulation But Impairs Lymphoid Differentiation Capacity of Hematopoietic Stem Cells in Early Aging. *J. Exp. Med.* **2016**, *213*, 535–553. [CrossRef] [PubMed]
- 70. Park, J.-H.; Glass, Z.; Sayed, K.; Michurina, T.V.; Lazutkin, A.; Mineyeva, O.; Velmeshev, D.; Ward, W.F.; Richardson, A.; Enikolopov, G. Calorie Restriction Alleviates the Age-Related Decrease in Neural Progenitor Cell Division in the Aging Brain. *Eur. J. Neurosci.* **2013**, *37*, 1987–1993. [CrossRef] [PubMed]
- 71. Yilmaz, Ö.H.; Katajisto, P.; Lamming, D.W.; Gültekin, Y.; Bauer-Rowe, K.E.; Sengupta, S.; Birsoy, K.; Dursun, A.; Yilmaz, V.O.; Selig, M.; et al. mTORC1 in the Paneth Cell Niche Couples Intestinal Stem-Cell Function to Calorie Intake. *Nature* **2012**, *486*, 490–495. [CrossRef] [PubMed]
- 72. Cerletti, M.; Jang, Y.C.; Finley, L.W.S.; Haigis, M.C.; Wagers, A.J. Short-Term Calorie Restriction Enhances Skeletal Muscle Stem Cell Function. *Cell Stem Cell* **2012**, *10*, 515–519. [CrossRef] [PubMed]
- 73. Mazzoccoli, G.; Tevy, M.F.; Borghesan, M.; Vergini, M.R.D.; Vinciguerra, M. Caloric Restriction and Aging Stem Cells: The Stick and the Carrot? *Exp. Gerontol.* **2014**, *50*, 137–148. [CrossRef] [PubMed]
- 74. Klimova, B.; Novotny, M.; Kuca, K. Anti-Aging Drugs–Prospect of Longer Life? *Curr. Med. Chem.* 2017, 25. [CrossRef] [PubMed]
- 75. Anisimov, V.N.; Berstein, L.M.; Popovich, I.G.; Zabezhinski, M.A.; Egormin, P.A.; Piskunova, T.S.; Semenchenko, A.V.; Tyndyk, M.L.; Yurova, M.N.; Kovalenko, I.G.; et al. If Started Early in Life, Metformin Treatment Increases Life Span and Postpones Tumors in Female SHR Mice. *Aging* 2011, *3*, 148–157. [CrossRef] [PubMed]
- 76. Cabreiro, F.; Au, C.; Leung, K.-Y.; Vergara-Irigaray, N.; Cochemé, H.M.; Noori, T.; Weinkove, D.; Schuster, E.; Greene, N.D.E.; Gems, D. Metformin Retards Aging in *C. elegans* By Altering Microbial Folate and Methionine Metabolism. *Cell* 2013, 153, 228–239. [CrossRef] [PubMed]
- 77. De Haes, W.; Frooninckx, L.; Van Assche, R.; Smolders, A.; Depuydt, G.; Billen, J.; Braeckman, B.P.; Schoofs, L.; Temmerman, L. Metformin Promotes Lifespan through Mitohormesis via the Peroxiredoxin PRDX-2. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, E2501-9. [CrossRef] [PubMed]
- Allard, J.S.; Perez, E.J.; Fukui, K.; Carpenter, P.; Ingram, D.K.; De Cabo, R. Prolonged Metformin Treatment Leads to Reduced Transcription of Nrf2 and Neurotrophic Factors without Cognitive Impairment in Older C57BL/6J Mice. *Behav. Brain Res.* 2016, 301, 1–9. [CrossRef] [PubMed]
- 79. Onken, B.; Driscoll, M. Metformin Induces a Dietary Restriction–Like State and the Oxidative Stress Response to Extend *C. elegans* Healthspan via AMPK, LKB1, and SKN-1. *PLoS ONE* **2010**, *5*, E8758. [CrossRef] [PubMed]
- Martin-Montalvo, A.; Mercken, E.M.; Mitchell, S.J.; Palacios, H.H.; Mote, P.L.; Scheibye-Knudsen, M.; Gomes, A.P.; Ward, T.M.; Minor, R.K.; Blouin, M.-J.; et al. Metformin Improves Healthspan and Lifespan in Mice. *Nat. Commun.* 2013, *4*, 2192. [CrossRef] [PubMed]
- 81. Li, Y.-R.; Li, S.; Lin, C.-C. Effect of Resveratrol and Pterostilbene on Aging and Longevity. *Biofactors* **2018**, *44*, 69–82. [CrossRef] [PubMed]
- Tung, B.T.; Rodriguez-Bies, E.; Thanh, H.N.; Le-Thi-Thu, H.; Navas, P.; Sanchez, V.M.; López-Lluch, G. Organ and Tissue-Dependent Effect of Resveratrol and Exercise on Antioxidant Defenses of Old Mice. *Aging Clin. Exp. Res.* 2015, *27*, 775–783. [CrossRef] [PubMed]

- Marchal, J.; Blanc, S.; Epelbaum, J.; Aujard, F.; Pifferi, F. Effects of Chronic Calorie Restriction or Dietary Resveratrol Supplementation on Insulin Sensitivity Markers in a Primate, *Microcebus murinus*. *PLoS ONE* 2012, 7, E34289. [CrossRef] [PubMed]
- Timmers, S.; Konings, E.; Bilet, L.; Houtkooper, R.H.; Van De Weijer, T.; Goossens, G.H.; Hoeks, J.; Van Der Krieken, S.; Ryu, D.; Kersten, S.; et al. Calorie Restriction-Like Effects of 30 Days of Resveratrol Supplementation on Energy Metabolism and Metabolic Profile in Obese Humans. *Cell Metab.* 2011, 14, 612–622. [CrossRef] [PubMed]
- 85. Demontis, F.; Piccirillo, R.; Goldberg, A.L.; Perrimon, N. Mechanisms of Skeletal Muscle Aging: Insights from *Drosophila* and Mammalian Models. *Dis. Model. Mech.* **2013**, *6*, 1339–1352. [CrossRef] [PubMed]
- Dai, J.-M.; Yu, M.-X.; Shen, Z.-Y.; Guo, C.-Y.; Zhuang, S.-Q.; Qiu, X.-S. Leucine Promotes Proliferation and Differentiation of Primary Preterm Rat Satellite Cells in Part through mTORC1 Signaling Pathway. *Nutrients* 2015, 7, 3387–3400. [CrossRef] [PubMed]
- 87. Yue, T.; Yin, J.; Li, F.; Li, D.; Du, M. High Glucose Induces Differentiation and Adipogenesis in Porcine Muscle Satellite Cells via mTOR. *BMB Rep.* **2010**, *43*, 140–145. [CrossRef] [PubMed]
- Aspnes, L.E.; Lee, C.M.; Weindruch, R.; Chung, S.S.; Roecker, E.B.; Aiken, J.M. Caloric Restriction Reduces Fiber Loss and Mitochondrial Abnormalities in Aged Rat Muscle. *FASEB J.* 1997, 11, 573–581. [CrossRef] [PubMed]
- 89. Phillips, T.; Leeuwenburgh, C. Muscle Fiber Specific Apoptosis and TNF-Alpha Signaling in Sarcopenia Are Attenuated By Life-Long Calorie Restriction. *FASEB J.* **2005**, *19*, 668–670. [CrossRef] [PubMed]
- Hepple, R.T.; Qin, M.; Nakamoto, H.; Goto, S. Caloric Restriction Optimizes the Proteasome Pathway with Aging in Rat Plantaris Muscle: Implications for Sarcopenia. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 2008, 295, R1231–R1237. [CrossRef] [PubMed]
- Mckiernan, S.H.; Colman, R.J.; Lopez, M.; Beasley, T.M.; Aiken, J.M.; Anderson, R.M.; Weindruch, R. Caloric Restriction Delays Aging-Induced Cellular Phenotypes in Rhesus Monkey Skeletal Muscle. *Exp. Gerontol.* 2011, 46, 23–29. [CrossRef] [PubMed]
- 92. Inoki, K.; Zhu, T.; Guan, K.-L. TSC2 Mediates Cellular Energy Response to Control Cell Growth and Survival. *Cell* **2003**, *115*, 577–590. [CrossRef]
- 93. Lee, C.K.; Klopp, R.G.; Weindruch, R.; Prolla, T.A. Gene Expression Profile of Aging and Its Retardation by Caloric Restriction. *Science* **1999**, *285*, 1390–1393. [CrossRef] [PubMed]
- 94. Rodríguez-Bies, E.; Navas, P.; López-Lluch, G. Age-Dependent Effect of Every-Other-Day Feeding and Aerobic Exercise in Ubiquinone Levels and Related Antioxidant Activities in Mice Muscle. *J. Gerontol. A Biol. Sci. Med. Sci.* **2015**, *70*, 33–43. [CrossRef] [PubMed]
- Boldrin, L.; Ross, J.A.; Whitmore, C.; Doreste, B.; Beaver, C.; Eddaoudi, A.; Pearce, D.J.; Morgan, J.E. The Effect of Calorie Restriction on Mouse Skeletal Muscle Is Sex, Strain and Time-Dependent. *Sci. Rep.* 2017, 7, 5160. [CrossRef] [PubMed]
- 96. Chung, H.Y.; Lee, E.K.; Choi, Y.J.; Kim, J.M.; Kim, D.H.; Zou, Y.; Kim, C.H.; Lee, J.; Kim, H.S.; Kim, N.D.; et al. Molecular Inflammation as an Underlying Mechanism of the Aging Process and Age-Related Diseases. J. Dent. Res. 2011, 90, 830–840. [CrossRef] [PubMed]
- Horrillo, D.; Sierra, J.; Arribas, C.; García-San Frutos, M.; Carrascosa, J.M.; Lauzurica, N.; Fernández-Agulló, T.; Ros, M. Age-Associated Development of Inflammation in Wistar Rats: Effects of Caloric Restriction. *Arch. Physiol. Biochem.* 2011, *117*, 140–150. [CrossRef] [PubMed]
- 98. Wåhlin-Larsson, B.; Carnac, G.; Kadi, F. The Influence of Systemic Inflammation on Skeletal Muscle in Physically Active Elderly Women. *Age (Dordr.)* **2014**, *36*, 9718. [CrossRef] [PubMed]
- 99. Fontana, L.; Villareal, D.T.; Das, S.K.; Smith, S.R.; Meydani, S.N.; Pittas, A.G.; Klein, S.; Bhapkar, M.; Rochon, J.; Ravussin, E.; et al. Effects of 2-Year Calorie Restriction on Circulating Levels of IGF-1, IGF-Binding Proteins and Cortisol in Nonobese Men and Women: A Randomized Clinical Trial. *Aging Cell* 2016, *15*, 22–27. [CrossRef] [PubMed]
- 100. Park, S.-J.; Gavrilova, O.; Brown, A.L.; Soto, J.E.; Bremner, S.; Kim, J.; Xu, X.; Yang, S.; Um, J.-H.; Koch, L.G.; et al. DNA-PK Promotes The Mitochondrial, Metabolic, and Physical Decline that Occurs during Aging. *Cell Metab.* 2017, 25, 1135–1146.E7. [CrossRef] [PubMed]
- Hord, J.M.; Botchlett, R.; Lawler, J.M. Age-Related Alterations in the Sarcolemmal Environment Are Attenuated by Lifelong Caloric Restriction and Voluntary Exercise. *Exp. Gerontol.* 2016, *83*, 148–157. [CrossRef] [PubMed]

- 102. Wang, H.; Sharma, N.; Arias, E.B.; Cartee, G.D. Insulin Signaling and Glucose Uptake in the Soleus Muscle of 30-Month-Old Rats after Calorie Restriction with or without Acute Exercise. J. Gerontol. Ser. A Biol. Sci. Med. Sci. 2016, 71, 323–332. [CrossRef] [PubMed]
- 103. Wang, H.; Arias, E.B.; Yu, C.S.; Verkerke, A.R.P.; Cartee, G.D. Effects of Calorie Restriction and Fiber Type on Glucose Uptake and Abundance of Electron Transport Chain and Oxidative Phosphorylation Proteins in Single Fibers from Old Rats. J. Gerontol. A Biol. Sci. Med. Sci. 2017, 72, 1638–1646. [CrossRef] [PubMed]
- 104. Ohanna, M.; Sobering, A.K.; Lapointe, T.; Lorenzo, L.; Praud, C.; Petroulakis, E.; Sonenberg, N.; Kelly, P.A.; Sotiropoulos, A.; Pende, M. Atrophy of S6k1(-/-) Skeletal Muscle Cells Reveals Distinct mTOR Effectors for Cell Cycle and Size Control. *Nat. Cell Biol.* 2005, *7*, 286–294. [CrossRef] [PubMed]
- 105. Risson, V.; Mazelin, L.; Roceri, M.; Sanchez, H.; Moncollin, V.; Corneloup, C.; Richard-Bulteau, H.; Vignaud, A.; Baas, D.; Defour, A.; et al. Muscle Inactivation of mTOR Causes Metabolic and Dystrophin Defects Leading to Severe Myopathy. J. Cell Biol. 2009, 187, 859–874. [CrossRef] [PubMed]
- 106. Mounier, R.; Lantier, L.; Leclerc, J.; Sotiropoulos, A.; Foretz, M.; Viollet, B. Antagonistic Control of Muscle Cell Size by AMPK and mTORC1. *Cell Cycle* 2011, 10, 2640–2646. [CrossRef] [PubMed]
- 107. Terada, N.; Patel, H.R.; Takase, K.; Kohno, K.; Nairn, A.C.; Gelfand, E.W. Rapamycin Selectively Inhibits Translation of mRNAs Encoding Elongation Factors and Ribosomal Proteins. *Proc. Natl. Acad. Sci. USA* 1994, 91, 11477–11481. [CrossRef] [PubMed]
- 108. Welsh, G.I.; Stokes, C.M.; Wang, X.; Sakaue, H.; Ogawa, W.; Kasuga, M.; Proud, C.G. Activation of Translation Initiation Factor eIF2B By Insulin Requires Phosphatidyl Inositol 3-Kinase. FEBS Lett. 1997, 410, 418–422. [CrossRef]
- 109. Pallafacchina, G.; Calabria, E.; Serrano, A.L.; Kalhovde, J.M.; Schiaffino, S. A Protein Kinase B-Dependent and Rapamycin-Sensitive Pathway Controls Skeletal Muscle Growth But Not Fiber Type Specification. *Proc. Natl. Acad. Sci. USA* 2002, 99, 9213–9218. [CrossRef] [PubMed]
- Léger, B.; Derave, W.; De Bock, K.; Hespel, P.; Russell, A.P. Human Sarcopenia Reveals an Increase in SOCS-3 and Myostatin and a Reduced Efficiency of AKT Phosphorylation. *Rejuvenation Res.* 2008, 11, 163–175b. [CrossRef] [PubMed]
- 111. Sharples, A.P.; Hughes, D.C.; Deane, C.S.; Saini, A.; Selman, C.; Stewart, C.E. Longevity and Skeletal Muscle Mass: The Role of IGF Signalling, the Sirtuins, Dietary Restriction and Protein Intake. *Aging Cell* 2015, 14, 511–523. [CrossRef] [PubMed]
- 112. Pardo, P.S.; Mohamed, J.S.; Lopez, M.A.; Boriek, A.M. Induction of SIRT1 by Mechanical Stretch of Skeletal Muscle through the Early Response Factor EGR1 Triggers an Antioxidative Response. *J. Biol. Chem.* 2011, 286, 2559–2566. [CrossRef] [PubMed]
- Schenk, S.; Mccurdy, C.E.; Philp, A.; Chen, M.Z.; Holliday, M.J.; Bandyopadhyay, G.K.; Osborn, O.; Baar, K.; Olefsky, J.M. SIRT1 Enhances Skeletal Muscle Insulin Sensitivity in Mice during Caloric Restriction. *J. Clin. Investig.* 2011, 121, 4281–4288. [CrossRef] [PubMed]
- 114. Fulco, M.; Schiltz, R.L.; Iezzi, S.; King, M.T.; Zhao, P.; Kashiwaya, Y.; Hoffman, E.; Veech, R.L.; Sartorelli, V. SIR2 Regulates Skeletal Muscle Differentiation as a Potential Sensor of the Redox State. *Mol. Cell* 2003, 12, 51–62. [CrossRef]
- 115. Fulco, M.; Cen, Y.; Zhao, P.; Hoffman, E.P.; Mcburney, M.W.; Sauve, A.A.; Sartorelli, V. Glucose Restriction Inhibits Skeletal Myoblast Differentiation by Activating SIRT1 Through AMPK-Mediated Regulation of Nampt. *Dev. Cell* 2008, 14, 661–673. [CrossRef] [PubMed]
- 116. Gerhart-Hines, Z.; Rodgers, J.T.; Bare, O.; Lerin, C.; Kim, S.-H.; Mostoslavsky, R.; Alt, F.W.; Wu, Z.; Puigserver, P. Metabolic Control of Muscle Mitochondrial Function and Fatty Acid Oxidation through SIRT1/PGC-1alpha. *Embo J.* 2007, 26, 1913–1923. [CrossRef] [PubMed]
- 117. Nemoto, S.; Fergusson, M.M.; Finkel, T. Nutrient Availability Regulates SIRT1 through a forkhead-Dependent Pathway. *Science* 2004, *306*, 2105–2108. [CrossRef] [PubMed]
- 118. Nemoto, S.; Fergusson, M.M.; Finkel, T. SIRT1 Functionally Interacts with the Metabolic Regulator and Transcriptional Coactivator PGC-1{Alpha}. *J. Biol. Chem.* **2005**, *280*, 16456–16460. [CrossRef] [PubMed]
- Rodgers, J.T.; Lerin, C.; Haas, W.; Gygi, S.P.; Spiegelman, B.M.; Puigserver, P. Nutrient Control of Glucose Homeostasis through a Complex of PGC-1α and SIRT1. *Nature* 2005, 434, 113–118. [CrossRef] [PubMed]
- 120. Kume, S.; Haneda, M.; Kanasaki, K.; Sugimoto, T.; Araki, S.; Isshiki, K.; Isono, M.; Uzu, T.; Guarente, L.; Kashiwagi, A.; et al. SIRT1 Inhibits Transforming Growth Factor Beta-Induced Apoptosis in Glomerular Mesangial Cells Via Smad7 Deacetylation. *J. Biol. Chem.* 2007, 282, 151–158. [CrossRef] [PubMed]

- 121. Defour, A.; Dessalle, K.; Castro Perez, A.; Poyot, T.; Castells, J.; Gallot, Y.S.; Durand, C.; Euthine, V.; Gu, Y.; Béchet, D.; et al. Sirtuin 1 Regulates SREBP-1c Expression in a LXR-Dependent Manner in Skeletal Muscle. *PLoS ONE* 2012, 7, E43490. [CrossRef] [PubMed]
- 122. Glass, D.J. A Signaling Role for Dystrophin: Inhibiting Skeletal Muscle Atrophy Pathways. *Cancer Cell* 2005, *8*, 351–352. [CrossRef] [PubMed]
- 123. Crosio, C.; Cermakian, N.; Allis, C.D.; Sassone-Corsi, P. Light Induces Chromatin Modification in Cells of the Mammalian Circadian Clock. *Nat. Neurosci.* **2000**, *3*, 1241–1247. [CrossRef] [PubMed]
- 124. Lees, S.J.; Rathbone, C.R.; Booth, F.W. Age-Associated Decrease in Muscle Precursor Cell Differentiation. *Am. J. Physiol. Physiol.* 2006, 290, C609–C615. [CrossRef] [PubMed]
- 125. Machida, S.; Booth, F.W. Increased Nuclear Proteins in Muscle Satellite Cells in Aged Animals As Compared to Young Growing Animals. *Exp. Gerontol.* **2004**, *39*, 1521–1525. [CrossRef] [PubMed]
- 126. Pardo, P.S.; Boriek, A.M. The Physiological Roles of SIRT1 in Skeletal Muscle. *Aging (Albany NY)* **2011**, *3*, 430–437. [CrossRef] [PubMed]
- 127. Brunet, A.; Sweeney, L.B.; Sturgill, J.F.; Chua, K.F.; Greer, P.L.; Lin, Y.; Tran, H.; Ross, S.E.; Mostoslavsky, R.; Cohen, H.Y.; et al. Stress-Dependent Regulation of Foxo Transcription Factors By the SIRT1 Deacetylase. *Science* 2004, 303, 2011–2015. [CrossRef] [PubMed]
- 128. Ghosh, H.S.; Mcburney, M.; Robbins, P.D. SIRT1 Negatively Regulates the Mammalian Target of Rapamycin. *PLoS ONE* **2010**, *5*, E9199. [CrossRef] [PubMed]
- 129. Samant, S.A.; Kanwal, A.; Pillai, V.B.; Bao, R.; Gupta, M.P. The Histone Deacetylase SIRT6 Blocks Myostatin Expression and Development of Muscle Atrophy. *Sci. Rep.* **2017**, *7*, 11877. [CrossRef] [PubMed]
- Yeung, F.; Hoberg, J.E.; Ramsey, C.S.; Keller, M.D.; Jones, D.R.; Frye, R.A.; Mayo, M.W. Modulation of NF-KappaB-Dependent Transcription and Cell Survival by the SIRT1 Deacetylase. *EMBO J.* 2004, 23, 2369–2380. [CrossRef] [PubMed]
- 131. Kanfi, Y.; Naiman, S.; Amir, G.; Peshti, V.; Zinman, G.; Nahum, L.; Bar-Joseph, Z.; Cohen, H.Y. The Sirtuin SIRT6 Regulates Lifespan in Male Mice. *Nature* **2012**, *483*, 218–221. [CrossRef] [PubMed]
- 132. Tilstra, J.S.; Robinson, A.R.; Wang, J.; Gregg, S.Q.; Clauson, C.L.; Reay, D.P.; Nasto, L.A.; St Croix, C.M.; Usas, A.; Vo, N.; et al. NF-KB Inhibition Delays DNA Damage-Induced Senescence and Aging in Mice. *J. Clin. Investig.* 2012, 122, 2601–2612. [CrossRef] [PubMed]
- 133. Belman, J.P.; Bian, R.R.; Habtemichael, E.N.; Li, D.T.; Jurczak, M.J.; Alcázar-Román, A.; Mcnally, L.J.; Shulman, G.I.; Bogan, J.S. Acetylation of Tug Protein Promotes the Accumulation of GLUT4 Glucose Transporters in an Insulin-Responsive Intracellular Compartment. *J. Biol. Chem.* 2015, 290, 4447–4463. [CrossRef] [PubMed]
- Arora, A.; Dey, C.S. SIRT2 Negatively Regulates Insulin Resistance in C2C12 Skeletal Muscle Cells. Biochim. Biophys. Acta 2014, 1842, 1372–1378. [CrossRef] [PubMed]
- 135. Xiong, Y.; Lei, Q.-Y.; Zhao, S.; Guan, K.-L. Regulation of Glycolysis and Gluconeogenesis by Acetylation of PKM and PEPCK. *Cold Spring Harb. Symp. Quant. Biol.* **2011**, *76*, 285–289. [CrossRef] [PubMed]
- 136. Palacios, O.M.; Carmona, J.J.; Michan, S.; Chen, K.Y.; Manabe, Y.; Ward, J.L.; Goodyear, L.J.; Tong, Q. Diet and Exercise Signals Regulate SIRT3 and Activate AMPK and PGC-1alpha in Skeletal Muscle. *Aging (Albany NY)* 2009, 1, 771–783. [CrossRef] [PubMed]
- 137. Seufert, C.D.; Graf, M.; Janson, G.; Kuhn, A.; Söling, H.D. Formation of Free Acetate by Isolated Perfused Livers from Normal, Starved and Diabetic Rats. *Biochem. Biophys. Res. Commun.* **1974**, *57*, 901–909. [CrossRef]
- Hallows, W.C.; Lee, S.; Denu, J.M. Sirtuins Deacetylate and Activate Mammalian Acetyl-Coa Synthetases. Proc. Natl. Acad. Sci. USA 2006, 103, 10230–10235. [CrossRef] [PubMed]
- Schwer, B.; Bunkenborg, J.; Verdin, R.O.; Andersen, J.S.; Verdin, E. Reversible Lysine Acetylation Controls the Activity of the Mitochondrial Enzyme Acetyl-Coa Synthetase 2. *Proc. Natl. Acad. Sci. USA* 2006, 103, 10224–10229. [CrossRef] [PubMed]
- Falcón, A.A.; Chen, S.; Wood, M.S.; Aris, J.P. Acetyl-Coenzyme a Synthetase 2 Is a Nuclear Protein Required for Replicative Longevity In Saccharomyces Cerevisiae. *Mol. Cell. Biochem.* 2010, 333, 99–108. [CrossRef] [PubMed]
- Dittenhafer-Reed, K.E.; Richards, A.L.; Fan, J.; Smallegan, M.J.; Fotuhi Siahpirani, A.; Kemmerer, Z.A.; Prolla, T.A.; Roy, S.; Coon, J.J.; Denu, J.M. SIRT3 Mediates Multi-Tissue Coupling for Metabolic Fuel Switching. *Cell Metab.* 2015, *21*, 637–646. [CrossRef] [PubMed]

- 142. Cantó, C.; Gerhart-Hines, Z.; Feige, J.N.; Lagouge, M.; Noriega, L.; Milne, J.C.; Elliott, P.J.; Puigserver, P.; Auwerx, J. AMPK Regulates Energy Expenditure By Modulating Nad+ Metabolism and SIRT1 Activity. *Nature* 2009, 458, 1056–1060. [CrossRef] [PubMed]
- 143. Wronska, A.; Lawniczak, A.; Wierzbicki, P.M.; Kmiec, Z. Age-Related Changes in Sirtuin 7 Expression in Calorie-Restricted and Refed Rats. *Gerontology* **2016**, *62*, 304–310. [CrossRef] [PubMed]
- 144. Margolis, L.M.; Rivas, D.A.; Berrone, M.; Ezzyat, Y.; Young, A.J.; Mcclung, J.P.; Fielding, R.A.; Pasiakos, S.M. Prolonged Calorie Restriction Downregulates Skeletal Muscle mTORC1 Signaling Independent of Dietary Protein Intake and Associated Microrna Expression. *Front. Physiol.* 2016, 7, 445. [CrossRef] [PubMed]
- 145. Treviño-Saldaña, N.; García-Rivas, G. Regulation of Sirtuin-Mediated Protein Deacetylation by Cardioprotective Phytochemicals. *Oxid. Med. Cell. Longev.* **2017**, 2017, 1750306. [CrossRef] [PubMed]
- 146. Winnik, S.; Auwerx, J.; Sinclair, D.A.; Matter, C.M. Protective Effects of Sirtuins in Cardiovascular Diseases: From Bench to Bedside. *Eur. Heart J.* **2015**, *36*, 3404–3412. [CrossRef] [PubMed]
- 147. Spadari, R.C.; Cavadas, C.; De Carvalho, A.E.T.S.; Ortolani, D.; De Moura, A.L.; Vassalo, P.F. Role of Beta-Adrenergic Receptors and Sirtuin Signaling in the Heart During Aging, Heart Failure, and Adaptation to Stress. *Cell. Mol. Neurobiol.* **2018**, *38*, 109–120. [CrossRef] [PubMed]
- Matsushima, S.; Sadoshima, J. The Role of Sirtuins in Cardiac Disease. Am. J. Physiol. Circ. Physiol. 2015, 309, H1375–H1389. [CrossRef] [PubMed]
- 149. Yu, W.; Zhou, H.-F.; Lin, R.-B.; Fu, Y.-C.; Wang, W. Short-Term Calorie Restriction Activates SIRT1-4 and -7 in Cardiomyocytes In Vivo and In Vitro. *Mol. Med. Rep.* **2014**, *9*, 1218–1224. [CrossRef] [PubMed]
- 150. Matsui, Y.; Takagi, H.; Qu, X.; Abdellatif, M.; Sakoda, H.; Asano, T.; Levine, B.; Sadoshima, J. Distinct Roles of Autophagy in the Heart during Ischemia and Reperfusion: Roles of AMP-Activated Protein Kinase and Beclin 1 In Mediating Autophagy. *Circ. Res.* **2007**, *100*, 914–922. [CrossRef] [PubMed]
- 151. Hariharan, N.; Zhai, P.; Sadoshima, J. Oxidative Stress Stimulates Autophagic Flux during Ischemia/Reperfusion. *Antioxid. Redox Signal.* **2011**, *14*, 2179–2190. [CrossRef] [PubMed]
- 152. Dolinsky, V.W.; Dyck, J.R.B. Calorie Restriction and Resveratrol in Cardiovascular Health and Disease. *Biochim. Biophys. Acta Mol. Basis Dis.* **2011**, *1812*, 1477–1489. [CrossRef] [PubMed]
- 153. Fujitsuka, N.; Asakawa, A.; Morinaga, A.; Amitani, M.S.; Amitani, H.; Katsuura, G.; Sawada, Y.; Sudo, Y.; Uezono, Y.; Mochiki, E.; et al. Increased Ghrelin Signaling Prolongs Survival in Mouse Models of Human Aging through Activation of Sirtuin1. *Mol. Psychiatry* **2016**, *21*, 1613–1623. [CrossRef] [PubMed]
- 154. Yamamoto, T.; Tamaki, K.; Shirakawa, K.; Ito, K.; Yan, X.; Katsumata, Y.; Anzai, A.; Matsuhashi, T.; Endo, J.; Inaba, T.; et al. Cardiac SIRT1 Mediates the Cardioprotective Effect of Caloric Restriction By Suppressing Local Complement System Activation after Ischemia-Reperfusion. *Am. J. Physiol. Heart Circ. Physiol.* 2016, 310, H1003-14. [CrossRef] [PubMed]
- Shinmura, K.; Tamaki, K.; Bolli, R. Impact of 6-Mo Caloric Restriction on Myocardial Ischemic tolerance: Possible Involvement of Nitric Oxide-Dependent Increase in Nuclear SIRT1. *Am. J. Physiol. Heart Circ. Physiol.* 2008, 295, H2348–H2355. [CrossRef] [PubMed]
- 156. De Cavanagh, E.M.V.; Inserra, F.; Ferder, L. Angiotensin Ii Blockade: How Its Molecular Targets May Signal to Mitochondria and Slow Aging. Coincidences with Calorie Restriction and mTOR Inhibition. *Am. J. Physiol. Heart Circ. Physiol.* **2015**, 309, H15–H44. [CrossRef] [PubMed]
- 157. Sheng, Y.; Lv, S.; Huang, M.; Lv, Y.; Yu, J.; Liu, J.; Tang, T.; Qi, H.; Di, W.; Ding, G. Opposing Effects on Cardiac Function by Calorie Restriction in Different-Aged Mice. *Aging Cell* **2017**, *16*, 1155–1167. [CrossRef] [PubMed]
- 158. Booth, S.A.; Charchar, F.J. Cardiac Telomere Length in Heart Development, Function, and Disease. *Physiol. Genomics* **2017**, *49*, 368–384. [CrossRef] [PubMed]
- 159. Zannolli, R.; Mohn, A.; Buoni, S.; Pietrobelli, A.; Messina, M.; Chiarelli, F.; Miracco, C. Telomere Length and Obesity. *Acta Paediatr.* 2008, *97*, 952–954. [CrossRef] [PubMed]
- Yang, Y.; Fu, W.; Chen, J.; Olashaw, N.; Zhang, X.; Nicosia, S.V.; Bhalla, K.; Bai, W. SIRT1 Sumoylation Regulates Its Deacetylase Activity and Cellular Response to Genotoxic Stress. *Nat. Cell Biol.* 2007, 9, 1253–1262. [CrossRef] [PubMed]
- 161. Conrad, E.; Polonio-Vallon, T.; Meister, M.; Matt, S.; Bitomsky, N.; Herbel, C.; Liebl, M.; Greiner, V.; Kriznik, B.; Schumacher, S.; et al. HIPK2 Restricts SIRT1 Activity Upon Severe DNA Damage By A Phosphorylation-Controlled Mechanism. *Cell Death Differ.* 2016, 23, 110–122. [CrossRef] [PubMed]

- 162. Alcendor, R.R.; Gao, S.; Zhai, P.; Zablocki, D.; Holle, E.; Yu, X.; Tian, B.; Wagner, T.; Vatner, S.F.; Sadoshima, J. SIRT1 Regulates Aging and Resistance to Oxidative Stress in the Heart. *Circ. Res.* 2007, 100, 1512–1521. [CrossRef] [PubMed]
- 163. Bindu, S.; Pillai, V.B.; Gupta, M.P. Role of Sirtuins in Regulating Pathophysiology of the Heart. *Trends Endocrinol. Metab.* **2016**, *27*, 563–573. [CrossRef] [PubMed]
- 164. Hsu, C.-P.; Zhai, P.; Yamamoto, T.; Maejima, Y.; Matsushima, S.; Hariharan, N.; Shao, D.; Takagi, H.; Oka, S.; Sadoshima, J. Silent Information Regulator 1 Protects the Heart from Ischemia/Reperfusion. *Circulation* 2010, 122, 2170–2182. [CrossRef] [PubMed]
- 165. Sundaresan, N.R.; Pillai, V.B.; Wolfgeher, D.; Samant, S.; Vasudevan, P.; Parekh, V.; Raghuraman, H.; Cunningham, J.M.; Gupta, M.P.M.; Gupta, M.P.M. The Deacetylase SIRT1 Promotes Membrane Localization and Activation of AKT and PDK1 during Tumorigenesis and Cardiac Hypertrophy. *Sci. Signal.* 2011, *4*, Ra46. [CrossRef] [PubMed]
- 166. Oka, S.; Alcendor, R.; Zhai, P.; Park, J.Y.; Shao, D.; Cho, J.; Yamamoto, T.; Tian, B.; Sadoshima, J. PPARα-SIRT1 Complex Mediates Cardiac Hypertrophy and Failure through Suppression of the ERR Transcriptional Pathway. *Cell Metab.* **2011**, *14*, 598–611. [CrossRef] [PubMed]
- 167. Sundaresan, N.R.; Gupta, M.; Kim, G.; Rajamohan, S.B.; Isbatan, A.; Gupta, M.P. SIRT3 Blocks the Cardiac Hypertrophic Response by Augmenting Foxo3a-Dependent Antioxidant Defense Mechanisms in Mice. *J. Clin. Investig.* 2009, 119, 2758–2771. [CrossRef] [PubMed]
- 168. Sadoshima, J. SIRT3 Targets mPTP and Prevents Aging in the Heart. *Aging (Albany NY)* **2011**, *3*, 12–13. [CrossRef] [PubMed]
- 169. Pillai, V.B.; Sundaresan, N.R.; Kim, G.; Gupta, M.; Rajamohan, S.B.; Pillai, J.B.; Samant, S.; Ravindra, P.V.; Isbatan, A.; Gupta, M.P. Exogenous Nad Blocks Cardiac Hypertrophic Response via Activation of the SIRT3-LKB1-AMP-Activated Kinase Pathway. J. Biol. Chem. 2010, 285, 3133–3144. [CrossRef] [PubMed]
- 170. Sundaresan, N.R.; Bindu, S.; Pillai, V.B.; Samant, S.; Pan, Y.; Huang, J.-Y.; Gupta, M.; Nagalingam, R.S.; Wolfgeher, D.; Verdin, E.; et al. Sirt3 Blocks Aging-Associated Tissue Fibrosis in Mice by Deacetylating and Activating Glycogen Synthase Kinase 3β. *Mol. Cell. Biol.* **2015**, *36*, 678–692. [CrossRef] [PubMed]
- 171. Liu, B.; Che, W.; Xue, J.; Zheng, C.; Tang, K.; Zhang, J.; Wen, J.; Xu, Y. SIRT4 Prevents Hypoxia-Induced Apoptosis in H9C2 Cardiomyoblast Cells. *Cell. Physiol. Biochem.* **2013**, *32*, 655–662. [CrossRef] [PubMed]
- 172. Liu, B.; Che, W.; Zheng, C.; Liu, W.; Wen, J.; Fu, H.; Tang, K.; Zhang, J.; Xu, Y. SIRT5: A Safeguard against Oxidative Stress-Induced Apoptosis in Cardiomyocytes. *Cell. Physiol. Biochem.* 2013, 32, 1050–1059. [CrossRef] [PubMed]
- 173. Rardin, M.J.; He, W.; Nishida, Y.; Newman, J.C.; Carrico, C.; Danielson, S.R.; Guo, A.; Gut, P.; Sahu, A.K.;
 Li, B.; et al. SIRT5 Regulates the Mitochondrial Lysine Succinylome and Metabolic Networks. *Cell Metab.*2013, 18, 920–933. [CrossRef] [PubMed]
- 174. Tang, X.; Chen, X.-F.; Chen, H.-Z.; Liu, D.-P. Mitochondrial Sirtuins in Cardiometabolic Diseases. *Clin. Sci.* **2017**, *131*, 2063–2078. [CrossRef] [PubMed]
- 175. Vitiello, M.; Zullo, A.; Servillo, L.; Mancini, F.P.; Borriello, A.; Giovane, A.; Della Ragione, F.; D'onofrio, N.; Balestrieri, M.L. Multiple Pathways of SIRT6 at the Crossroads in the Control of Longevity, Cancer, and Cardiovascular Diseases. *Ageing Res. Rev.* **2017**, *35*, 301–311. [CrossRef] [PubMed]
- 176. Khan, D.; Sarikhani, M.; Maniyadath, B.; Pandit, A.S.; Mishra, S.; Ahamed, F.; Dubey, A.; Fathma, N.; Atreya, H.S.; Kolthur-Seetharam, U.; et al. SIRT6 Deacetylase Transcriptionally Regulates Glucose Metabolism in Heart. J. Cell. Physiol. 2018. [CrossRef] [PubMed]
- 177. Vakhrusheva, O.; Smolka, C.; Gajawada, P.; Kostin, S.; Boettger, T.; Kubin, T.; Braun, T.; Bober, E. SIRT7 Increases Stress Resistance of Cardiomyocytes and Prevents Apoptosis and Inflammatory Cardiomyopathy in Mice. *Circ. Res.* **2008**, *102*, 703–710. [CrossRef] [PubMed]
- 178. Baird, M.F.; Grace, F.; Sculthorpe, N.; Graham, S.M.; Fleming, A.; Baker, J.S. Evidence of Direct Cardiac Damage Following High-Intensity Exercise in Chronic Energy Restriction: A Case Report and Literature Review. *Medicine (Baltimore)* 2017, 96, E7030. [CrossRef] [PubMed]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).