

Genetic Variants Associated With Plasma Lipids Are Associated With the Lipid Response to Niacin

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Background—Niacin is a broad-spectrum lipid-modulating drug, but its mechanism of action is unclear. Genome-wide association studies have identified multiple loci associated with blood lipid levels and lipoprotein (a). It is unknown whether these loci modulate response to niacin.

Methods and Results—Using data from the AIM-HIGH (Atherothrombosis Intervention in Metabolic Syndrome with Low HDL/High Triglycerides and Impact on Global Health Outcomes) trial (n=2054 genotyped participants), we determined whether genetic variations at validated loci were associated with a differential change in plasma lipids and lipoprotein (a) 1 year after randomization to either statin+placebo or statin+niacin in a variant-treatment interaction model. Nominally significant interactions ($P<0.05$) were found for genetic variants in *MVK*, *LIPC*, *PABPC4*, *AMPD3* with change in high-density lipoprotein cholesterol; *SPTLC3* with change in low-density lipoprotein cholesterol; *TOM1* with change in total cholesterol; *PDXDC1* and *CYP26A1* with change in triglycerides; and none for lipoprotein (a). We also investigated whether these loci were associated with cardiovascular events. The risk of coronary disease related death was higher in the minor allele carriers at the *LIPC* locus in the placebo group (odds ratio 2.08, 95% confidence interval 1.11-3.90, $P=0.02$) but not observed in the niacin group (odds ratio 0.89, 95% confidence interval 0.48-1.65, $P=0.7$); P -interaction =0.02. There was a greater risk for acute coronary syndrome (odds ratio 1.85, 95% confidence interval 1.16-2.77, $P=0.02$) and revascularization events (odds ratio 1.64, 95% confidence interval 1.2-2.22, $P=0.002$) in major allele carriers at the *CYP26A1* locus in the placebo group not seen in the niacin group.

Conclusions—Genetic variation at loci previously associated with steady-state lipid levels displays evidence for lipid response to niacin treatment.

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Niacin has been used in the treatment of dyslipidemias for over 50 years.^{1,2} Niacin reduces total cholesterol (TC), low-density lipoprotein cholesterol (LDL-C), triglycerides

(TG), and increases high density lipoprotein cholesterol (HDL-C).² Niacin also lowers lipoprotein (a) [Lp(a)], an independent risk factor for coronary disease.^{3,4} Niacin was one of the first pharmacologic agents shown to reduce the incidence of nonfatal myocardial infarction and cardiac death.¹ Niacin has also demonstrated beneficial effects on arterial plaque regression in combination with statin therapy.^{5,6} However, in recent clinical outcome trials, the addition of extended-release niacin to intensive LDL-C-lowering therapy did not further reduce atherothrombotic events compared with intensive LDL-C-lowering therapy alone.^{7,8}

The mechanisms underlying the lipid-lowering effects of niacin are still unresolved. Niacin has been shown to increase HDL-C via reduction of HDL-apolipoprotein A-I catabolism and possibly reduction in the expression of cholesteryl ester transfer protein.^{9,10} In vitro studies have suggested that niacin may directly inhibit TG synthesis by inhibition of diacylglycerol acyltransferase in the liver, a key enzyme catalyzing the final step of TG synthesis.⁹ Niacin stimulates

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An accompanying Table S1 is available at <https://www.ahajournals.org/doi/suppl/10.1161/JAHA.117.008461>

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Clinical Perspective

What Is New?

- Niacin has been used to modulate the lipid profile, but its mechanism of action is still unclear.
- Using a candidate gene approach, we examined whether genetic loci associated with basal lipid traits were associated with the change in plasma lipid levels in response to niacin.
- We identified common variants in *MVK*, *LIPC*, *PABPC4*, *AMPD3*, *SPTLC3*, *PDXDC1*, and *CYP26A1* genes that were suggestive of treatment-related changes in lipid traits.

What Are the Clinical Implications?

- These findings suggest that genetic variation at loci previously associated with steady-state lipid levels displays evidence for lipid response to niacin treatment.
- After replication of these signals in other larger, independent studies, clinicians may use this information to identify patients who may benefit from niacin therapy.

the hydroxyl-carboxylic acid receptor 2 (also known as the niacin receptor) on adipocytes, resulting in a reduction in free fatty acids returning to the liver and decreased assembly of very low-density lipoproteins.² We previously reported that the coding variant *HCAR2* M317I was not associated with the change in LDL-C (percentage change -3.7 ± 39.1 , -2.6 ± 37.4 , -3.5 ± 35.2 , $P=0.58$, in the Met-Met, Met-Ile, and Ile-Ile carriers, respectively), HDL-C (28.2 ± 25.4 , 27.6 ± 23.5 , 26.1 ± 22.8 , $P=0.62$), and TG (-20.9 ± 38.4 , -23.0 ± 38.6 , -22.5 ± 36.2 , $P=0.50$) after 1 year of niacin+statin treatment.¹¹ This variant was, however, associated with Lp(a) lowering secondary to niacin (-22.7 ± 35.2 , -15.2 ± 40.1 , -15.8 ± 37.3 , $P=0.005$).¹¹

Recent large-scale genome-wide association studies (GWAS) have identified 157 loci to be significantly associated with basal fasting lipid traits.¹² Plasma Lp(a) concentrations are highly genetically regulated with the majority of genetic variation being attributable to the *LPA* locus.^{4,13} It is unknown whether genetic variation at these loci also mediates the effects of lipid-lowering medications. Individual response to niacin is highly heterogeneous suggesting a potential influence of genetic variation on the pharmacologic response. Although genetic predictors of lipid response to other lipid-altering drugs (statins,^{14,15} fibrates^{16,17}) have been reported, the pharmacogenetics of niacin has not been fully examined.

Here we investigated whether genetic loci associated with basal lipid traits and Lp(a) are associated with the change in plasma lipids and Lp(a) on treatment with ER Niacin in the AIM-HIGH (Atherothrombosis Intervention in Metabolic Syndrome with Low HDL/High Triglycerides and Impact on Global

Health Outcomes) study. We also examined whether these loci were associated with atherothrombotic events in the trial by treatment group.

Methods

A deidentified data set from the AIM-HIGH study is available through BioLINCC at the National Heart, Lung and Blood Institute.¹⁸ Consent language and DNA or patient-level DNA results will not be made available.

Ethics Statement

Participants provided written informed consent, and all research was conducted according to the principles outlined in the Declaration of Helsinki. The protocol was approved by the institutional review boards at all participating clinical sites. The genetic substudy was approved by the institutional review board at the University of Pennsylvania.

AIM-HIGH Cohort

The AIM-HIGH study design and baseline characteristics of the participants have been previously published.⁷ Briefly, the trial tested whether extended-release niacin added to intensive LDL-lowering therapy including a statin, as compared with intensive, matched LDL-lowering therapy alone, would reduce the risk of cardiovascular events when LDL was equalized between groups, in an attempt to test whether raising HDL would confer a benefit. The trials enrolled patients with established atherosclerotic cardiovascular disease and atherogenic dyslipidemia (low levels of HDL-C, elevated TG, and small dense particles of LDL-C). Of the total 3414 AIM-HIGH participants, 2054 had complete genetic and phenotype data for the current analysis.

Genotyping

Participants in the trial were genotyped using the Cardio-MetaboChip (Illumina, San Diego, CA). The MetaboChip is a gene-centric array containing $\approx 200\,000$ single nucleotide polymorphisms (SNPs), which were identified through genome-wide meta-analyses for metabolic and cardiovascular diseases and phenotypes.¹⁹ Genotyping was performed on Illumina's iScan System at the Center for Advanced Genomics at the Children's Hospital of Philadelphia. Of the total 3414 AIM-HIGH participants, 2432 provided DNA for genetic investigation. After initial DNA quality control, 2317 of the 2432 samples were genotyped with a $>95\%$ call rate. Cryptic relatedness was estimated by pairwise identity-by-descent analysis using PLINK (Shaun Purcell, Harvard, Boston, MA),²⁰ resulting in 5 duplicate pairs. The sample of the duplicate pair with lower genotyping call rate was removed. Among 196 725

SNPs on the chip, 19 229 SNPs were monomorphic, and they were removed in subsequent quality control analyses. Multi-dimensional scaling analysis was used to infer genetic race. Among the 2312 remaining samples, 79 were inferred to have African ancestry, 2101 to have European ancestry, and the rest to represent other races. For the purpose of this study, we performed analyses in participants with European ancestry only.

The lead SNPs primarily associated with each lipid trait were selected from the meta-analysis published by the Global Lipids Genetics Consortium (Center for Statistical Genetics, Ann Arbor, MI).¹² If a locus was associated with multiple lipid traits, we only examined the lipid trait primarily associated with that locus. Of the 157 loci validated as blood lipid concentration predictors,¹² 20 were not found on the MetaboChip, and proxies were selected for 18 of them based on linkage disequilibrium in the 1000 Genomes Project pilot data using the Broad Institute SNAP tool employing an r^2 threshold of 0.8 (see Table S1).²¹ We were unable to find proxies for rs964184 in *APOA1* or rs11649653 in *CTF1*. Of the 48 SNPs in the *LPA* and 1 SNP in the *APOE* gene region that are independently associated with Lp(a) concentrations,¹³ 3 SNPs were directly genotyped by MetaboChip, and proxies were found for 5 (Table S1).

Participants were further excluded from the analysis if they were missing lipid data (baseline or year-1 lipids), yielding 2054 and 1877 for the baseline and 1-year lipid analyses, respectively. The primary outcome was the percentage change in plasma concentrations of the 4 lipid traits and Lp(a) from baseline to 1 year after treatment with statin+niacin or statin+placebo. We employed a linear model with a SNP-treatment interaction term to test the additive effect of genotype on the percentage change in lipid traits and Lp(a) at 1 year after the randomization, adjusted for age, sex, body mass index, and treatment arm. We also examined the baseline prerandomization plasma concentrations of HDL-C, LDL-C, TC, TG, and Lp(a). Log transformations were carried out on nonnormally distributed variables. Logistic regression models were fit to evaluate the effect of genotype on coronary artery disease (CAD) outcomes in an interaction model. For the single-marker lipid analysis, P values were adjusted for multiple testing using the Bonferroni approach based on 320 hypotheses (58, 30, 37, 27, 8 SNPs for HDL-C, LDL-C, TC, TG, and Lp(a), respectively, interrogated at 2 time points), yielding a statistical significance threshold of 0.0002. Because no SNPs achieved this P value in the interaction analysis, top hits with a P value of <0.05 were reported because these SNPs have previously been associated with lipid traits at genome-wide levels of significance, and our analyses represent further characterization of each of these established loci.

A poststudy power analysis was performed for the association of *LIPC* SNP rs1532085 and change in HDL-C using a bootstrap method, resampling with replacement 10 000 times using the linear regression function in R v3.4.4.²² At α thresholds of $P=0.05$ and 0.0002, the study had a power of 87% and 26%, respectively, to detect this association.

Results

Study Population and Lipoprotein Changes

The clinical and demographic characteristics for the AIM-HIGH population that provided DNA during the course of the study as compared with the whole cohort are provided in Table 1. In the whole cohort as previously published,⁷ treatment with niacin resulted in a significant increase in HDL-C and a significant decrease in triglyceride concentrations as compared with the placebo group. The change in lipids we observed in the genetic subgroup was similar to that in the whole AIM-HIGH cohort. In a separate analysis of AIM-HIGH, the addition of niacin resulted in a significant reduction in Lp(a) levels by 21% in the statin+ER niacin group compared with 5.9% in the statin+placebo group ($P<0.05$).²³

Baseline Associations

The SNP genotypes, their chromosomal locations, and the nearest genes and their allele frequencies in the AIM-HIGH trial are shown in Table S1. Allele frequencies were comparable to those previously reported by the Global Lipids Genetics Consortium¹² and in a recent GWAS for Lp(a).¹³ We tested associations of these SNPs with baseline HDL-C, LDL-C, TC, TG, and Lp(a). All ($P<0.05$) associations are shown in Table 2. The threshold for Bonferroni correction for multiple comparisons was an adjusted α level of 0.0002. At this level we replicated associations with HDL-C at 1 locus and LDL-C at 1 locus. We also replicated the association of 8 variants within the *LPA* locus with Lp(a) levels. Given the size of our genotyped cohort, this is consistent with expectations based on power. Furthermore, more than 90% of the AIM-HIGH cohort was taking a statin at baseline and had a median LDL-C of 74 mg/dL, which may have obscured additional baseline associations.

Association With Lipid Traits at 1 Year

We tested each SNP for interaction with niacin in modulating the change in lipid traits at 1 year. None of the interaction results reached our adjusted P value corrected for multiple testing. Nominally significant SNP-treatment interactions were found for *MVK*, *LIPC*, *PABPC4*, and *AMPD3* with change

Table 1. Clinical and Demographic Characteristics of the Study Participants

Mean±SD, n (%)	Genetic Subgroup		Total AIM-HIGH Study	
	Statin+Placebo (n=1020)	Statin+ER Niacin (n=1034)	Statin+Placebo (n=1696)	Statin+ER Niacin (n=1718)
Age, y	64.0±8.7	64.6±8.7	63.7±8.7	63.7±8.8
Sex, female	164 (16.1%)	158 (15.3%)	251 (14.8%)	253 (14.7%)
Body mass index, kg/m ²	31.2±5.3	31.6±5.7	30.9±5.2	31.5±5.5*
History of myocardial infarction	556 (54.5)	554 (53.6)	955 (56.3)	968 (56.3)
History of stroke	240 (23.5)	244 (23.6)	362 (21.3)	358 (20.8)
History of hypertension	741 (72.6)	787 (76.1)	1189 (70.1)	1250 (72.8)
History of diabetes mellitus	341 (33.4)	351 (34.0)	570 (33.6)	588 (34.2)
Baseline HDL-C, mg/dL	35.1±5.6	34.6±5.6*	35.3±5.9	34.8±5.9*
Baseline LDL-C, mg/dL	74.6±22.2	73.5±22.0	75.8±24.3	76.2±25.7
Baseline TC, mg/dL	146.0±26.8	144.8±26.9	145.2±26.6	145.4±28.2
Baseline TG, (mg/dL), median (IQR)	162 (133-215)	166 (131-217)	162 (128-218)	164 (127-218)
Baseline lipoprotein (a) (nmol/L), median (IQR)	32 (13-118)	36 (14-132)	32.7 (13.1-122.6)	36.1 (13.5-126.6)
Change in LDL-C	−4.5 (−20.5, 13.9)	−9.5 (−28.0, 12.3)*	−4.25 (−20.57, 15.70)	−10.00 (−28.00, 12.68)**
Change in HDL-C	9.4 (0, 18.8)	25.0 (11.4 to 39.5)**	9.09 (0.00, 18.92)	23.33 (10.34, 39.29)**
Change in TC	0 (−12.0, 11.1)	−5.0 (−16.4, 8.4)**	−0.55 (−11.81, 11.59)	−5.19 (−16.17, 8.00)**
Change in TG	−4.4 (−24.6, 20.9)	−29.3 (−48.0, −6.4)**	−5.03 (−25.61, 20.77)	−28.24 (−46.61, −3.13)**
Change in Lp(a)	−7.5 (−25.9, 11.3)	−19.7 (−38.5, −0.6)	−7.0 (−25, 13.0)	−20.0 (−39.0, 1.0)

Change in lipid traits reported as median percentage change from baseline to 1 year (IQR). AIM-HIGH indicates Atherothrombosis Intervention in Metabolic Syndrome with Low HDL/High Triglycerides and Impact on Global Health Outcomes; ER, extended-release; HDL-C, high-density lipoprotein cholesterol; IQR, interquartile range; LDL-C, low-density lipoprotein cholesterol; Lp(a), lipoprotein (a); TC, total cholesterol; TG, triglycerides.

* $P < 0.05$ compared with placebo group.

** $P < 0.0001$.

in HDL-C; *SPTLC3* with change in LDL-C; *TOM1* with change in TC; *PDXDC1* and *CYP26A1* with change in TG at 1 year (Table 3). No significant SNP-treatment interactions were found for Lp(a).

Lipid levels by genotype are reported in Table 4. For the *MVK* gene, there was no change by genotype in HDL-C levels in the placebo group, but there was a nominally significant genotype effect in the niacin group. At the *LIPC* locus, a change in HDL-C by genotype was observed in the placebo group, but this effect was diminished in the niacin group. In the AIM-HIGH trial the placebo group achieved an overall 9% increase in HDL-C at the end of the first year, possibly because they received a small dose of immediate-release niacin to mask the treatment assignment, a dose previously shown to significantly raise HDL-C.²⁴ At the *PABPC4* locus a change in HDL-C by genotype was observed in the placebo group that was not observed in the niacin group. The change in HDL-C was nominally significant in the interaction model for *AMPD3* but no longer significant within each treatment strata.

The change in LDL-C was greater in minor allele carriers at the *SPTLC3* locus in the niacin group but not in the placebo group. Major allele carriers at the *PDXDC1* locus had a greater decrease in TG if they were treated with niacin but not with

placebo. Last, minor allele carriers at the *CYP26A1* locus had the largest decrease in TG levels in the niacin group but not in the placebo group. The minor allele for *CYP26A1* was associated with lower TG at baseline but a larger decrease in TG at 1 year in the niacin group only ($P < 0.0001$) (Table 4). The placebo group saw a nonsignificant change in TG from baseline regardless of the *CYP26A1* genotype.

Cardiovascular Outcomes

We also tested whether the SNPs nominally significant in the interaction model for lipid traits were associated with the risk of developing atherosclerotic events during the 3-year follow-up in the AIM-HIGH trial. The interactions of treatment and SNP on the primary cardiovascular end point (defined in the AIM-HIGH trial as the composite of death from CAD, nonfatal myocardial infarction, ischemic stroke, hospitalization for acute coronary syndrome, and symptom-driven revascularization) and the individual components of the primary end point are reported in Table 5.

A SNP-treatment interaction was found for *LIPC* and cardiovascular death. Homozygous minor allele carriers in the placebo group experienced the highest rate of CAD death (4%) versus heterozygous carriers (2.3%) and homozygous major

Table 2. Significant Genotypic Associations With Lipid Traits at Baseline

SNP	Trait	Locus	Chr	MAF	N	β	SE	P Value
rs3764261	HDL-C	<i>CETP</i>	16	0.30	2054	0.178	0.032	2.4×10^{-8}
rs1532085	HDL-C	<i>LIPC</i>	15	0.37	2054	0.106	0.023	0.0004
rs3136441	HDL-C	<i>LRP4</i>	11	0.12	2054	0.130	0.045	0.0037
rs581080	HDL-C	<i>TTC39B</i>	9	0.19	2054	-0.087	0.037	0.018
rs838880	HDL-C	<i>SCARB1</i>	12	0.31	2054	0.063	0.031	0.042
rs7239867	HDL-C	<i>LIPG</i>	18	0.17	2054	-0.077	0.038	0.046
rs6450176	HDL-C	<i>ARL15</i>	5	0.26	2054	-0.064	0.032	0.048
rs629301	LDL-C	<i>SORT1</i>	1	0.20	2054	-0.153	0.039	8.05×10^{-5}
rs4299376	LDL-C	<i>ABCG 5/8</i>	2	0.31	2054	0.082	0.034	0.015
rs10490626	LDL-C	<i>INSIG2</i>	2	0.072	2054	-0.141	0.060	0.018
rs364585	LDL-C	<i>SPTLC3</i>	20	0.39	2054	-0.063	0.032	0.046
rs4253772	TC	<i>PPARA</i>	22	0.11	2054	0.138	0.050	0.0059
rs1169288	TC	<i>HNF1A</i>	12	0.32	2054	0.090	0.033	0.0072
rs11065987	TC	<i>BRAP</i>	12	0.44	2054	-0.082	0.031	0.0090
rs2642442	TC	<i>MOSC1</i>	1	0.31	2054	-0.071	0.033	0.032
rs2954029	TG	<i>TRIB1</i>	8	0.44	2054	-0.113	0.031	0.0003
rs1260326	TG	<i>GCKR</i>	2	0.43	2054	0.112	0.031	0.0003
rs2131925	TG	<i>ANGPTL3</i>	1	0.33	2054	-0.089	0.033	0.0074
rs12678919	TG	<i>LPL</i>	8	0.073	2054	-0.131	0.060	0.028
rs174546	TG	<i>FADS 1-2-3</i>	11	0.34	2054	0.072	0.033	0.029
rs7769879	Lp(a)	<i>SLC22A3</i>	6	0.39	2054	0.351	0.032	7.03×10^{-28}
rs539298	Lp(a)	<i>SLC22A3</i>	6	0.47	2054	-0.273	0.031	1.99×10^{-18}
rs4252109	Lp(a)	<i>PLG</i>	6	0.29	2054	-0.290	0.034	1.32×10^{-17}
rs2504927	Lp(a)	<i>SLC22A3</i>	6	0.43	2054	-0.250	0.032	4.59×10^{-15}
rs394352	Lp(a)	<i>SLC22A3</i>	6	0.29	2054	-0.245	0.034	7.30×10^{-13}
rs3798221	Lp(a)	<i>LPA</i>	6	0.19	2054	-0.282	0.039	1.00×10^{-12}
rs986666	Lp(a)	<i>SLC22A3</i>	6	0.20	2054	-0.155	0.039	8.10×10^{-5}
rs2457561	Lp(a)	<i>SLC22A3</i>	6	0.19	2054	-0.154	0.041	0.00015

Chr indicates chromosome; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; Lp(a), lipoprotein (a); MAF, minor allele frequency; SE, standard error; SNP, single nucleotide polymorphism; TC, total cholesterol; TG, triglycerides.

allele carriers (1.0%) (odds ratio [OR]=2.08; 95% confidence interval 1.11-3.90; $P=0.02$) (Table 6). There was no difference in the rate of CAD death by *LIPC* genotype among the subjects receiving niacin (OR=0.89, 95% confidence interval 0.48-1.65, $P=0.71$). There was also a SNP-treatment interaction for *AMPD3* and CAD death. Minor allele carriers of the *AMPD3* had the lowest HDL-C levels at baseline and saw the smallest change in HDL-C levels at 1 year in both the niacin and placebo groups. However, the subjects homozygous for the minor allele in the placebo group had the highest rate of CAD death (9.1%) versus heterozygous carriers (2.5%) and homozygous major allele carriers (1.4%) (OR=2.16, 95% confidence interval 1.09-4.2, $P=0.03$). There was no

difference in CAD death by *AMPD3* genotype in the niacin group (OR=1.8, 95% 0.96-3.45, $P=0.07$).

A nominally significant SNP-treatment interaction was found for *CYP26A1* and acute coronary syndrome and symptom-driven revascularization. As mentioned above, the minor allele for *CYP26A1* was associated with lower TG at baseline but a larger decrease in TG at 1 year in the niacin group only (Table 4), whereas the placebo group saw no change in TG from baseline regardless of *CYP26A1* genotype. Major allele carriers had a higher rate of cardiac events—both acute coronary syndrome and symptom-driven revascularization—in the placebo group only (Table 6). There were no differences in cardiac events by *CYP26A1* genotype in the niacin group.

Table 3. Nominally Significant Gene-Treatment Interactions in Association With the Change in Lipid Traits From Baseline to 1 Year

SNP	Trait	Locus	Chr	MAF	N	β	SE	P Value_Interaction
rs10850443	HDL-C	<i>MVK</i>	12	0.47	1877	-0.170	0.059	0.0039
rs1532085	HDL-C	<i>LIPC</i>	15	0.36	1877	0.176	0.061	0.0040
rs4660293	HDL-C	<i>PABPC4</i>	1	0.23	1877	0.178	0.071	0.013
rs2923084	HDL-C	<i>AMPD3</i>	11	0.18	1877	-0.156	0.077	0.043
rs364585	LDL-C	<i>SPTLC3</i>	20	0.39	1877	-0.139	0.067	0.039
rs138777	TC	<i>TOM1</i>	22	0.34	1877	0.133	0.066	0.044
rs3198697	TG	<i>PDXDC1</i>	16	0.39	1877	0.175	0.062	0.0047
rs2068888	TG	<i>CYP26A1</i>	10	0.440	1877	-0.120	0.061	0.049

Chr indicates chromosome; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; MAF, minor allele frequency; SE, standard error; SNP, single nucleotide polymorphism; TC, total cholesterol; TG, triglycerides.

Discussion

In this study we evaluated whether genetic loci associated with plasma lipid levels and Lp(a) could also be pharmacogenetic markers of lipid response to niacin. We present data that common variants in *MVK*, *LIPC*, *PABPC4*, *AMPD3*, *SPTLC3*, *PDXDC1*, and *CYP26A1* genes were associated with treatment-related changes in lipid traits as observed 1 year following randomization in the AIM-HIGH trial. Additionally, *LIPC*, *AMPD3*, and *CYP26A1* variants were associated with cardiovascular events in the placebo-treated patients but not in the group receiving niacin.

In the AIM-HIGH trial, subjects in the placebo group received a small dose of immediate-release niacin to mask the treatment assignment; accordingly, the placebo group achieved an overall 9% increase in HDL-C at the end of the first year compared with 23% in the statin+niacin group. At the *LIPC* locus, rs1532085 was significantly associated with the change in HDL-C in the group receiving statin+placebo treatment, but treatment with niacin appeared to overcome the genotype effect. Most intriguing is that homozygous minor allele carriers in the statin+placebo group who saw the smallest change in HDL-C had the highest frequency of cardiovascular-related death (OR=2.08, $P=0.02$), which was not observed in the statin+niacin group (OR=0.89, $P=0.7$; P -interaction=0.02).

Hepatic lipase, encoded by *LIPC*, is a plasma lipolytic enzyme that hydrolyzes triglycerides and phospholipids in chylomicron remnants, intermediate-density lipoproteins, and HDL.²⁵ Hepatic lipase is an important determinant of plasma HDL-C, converting the large, buoyant, phospholipid-rich HDL₂ to small, dense HDL₃.²⁶ The presence of the C-allele in a common promoter polymorphism in *LIPC* (-514 C>T) is associated with greater hepatic lipase activity, small, dense LDL-C particles, and lower levels of the atheroprotective HDL₂ levels.^{27,28} Lipid-lowering therapies, including niacin,

have been shown to decrease hepatic lipase activity, increase HDL₂ and coronary disease regression, with a significantly greater effect in the CC (carrying two copies of the C allele at -514 C>T) subjects.²⁸ Although the rs1532085 is not in linkage disequilibrium with the -514 C>T variant, there is previous evidence for differential lipid-lowering responses by genotype at the *LIPC* locus.²⁸ In the genome-wide analysis published by the Global Lipids Consortium, the minor allele of rs1532085 was associated with higher HDL-C plasma concentrations and decreased transcript expression in liver tissue.²⁹ In the current study the minor allele of rs1532085 was also associated with higher HDL-C concentration at baseline, but at 1 year, the change in HDL-C was smaller in the minor allele carriers randomized to the statin+placebo arm. It may seem paradoxical that a variant associated with higher HDL-C may be associated with an increased risk for CAD. However, it has been previously demonstrated that a loss-of-function variant in *SCARB1* (P376L), coding for the scavenger receptor B1, the major receptor for HDL-C, was associated with significantly increased plasma HDL-C and an increased risk of CAD.³⁰ The growing consensus surrounding HDL biology indicates that HDL function and cholesterol flux may be more important than steady-state concentrations of HDL-C.³¹ Although rs1532085 has not previously been associated with CAD, 2 other variants in *LIPC*, rs588136 ($P=3.7 \times 10^{-4}$) and rs1800588 ($P=4.7 \times 10^{-4}$) have been nominally associated with CAD.^{32,33}

Previous GWAS studies have shown that the minor allele carriers of *CYP26A1*, rs2068888, have significantly lower TG levels.²⁹ In our study baseline TG was not different at this SNP because of the sample size of our study, but the change in TG levels was different by genotype in the niacin-treated subjects but not in the subjects receiving statin alone. This SNP has also been nominally associated with CAD ($P=5.4 \times 10^{-5}$)^{32,34} and atrial fibrillation ($P=0.0064$).^{34,35} Cytochrome P450 26A1 is an endoplasmic reticulum protein with high expression in

Table 4. Levels of Quantitative Traits at 1 Year by Genotype and Treatment Arm at Loci With a Nominally Significant Interaction

Gene/ Chr/SNP	Alleles			Placebo+Statin				Niacin+Statin				Absolute Change			
	MAF	Minor/Major	Trait	Baseline		Year 1		% Change		Absolute Change		Year 1		% Change	
				N	Mean (SD)*	N	Mean (SD)	Mean (SD)	Mean (SD)	N	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)	Mean (SD)
<i>MIK</i>	0.47	C/T	HDL	280	34.9 (5.4)	256	38.1 (7.9)	9.1 (15.9)	3.1 (5.7)	304	34.8 (5.7)	277	45.4 (12.1)	30.8 (25.3)	10.7 (9.3)
12				474	35.4 (5.5)	435	39.1 (7.4)	10.6 (15.3)	3.6 (5.3)	517	34.5 (5.4)	471	43.5 (10.6)	26.4 (23.1)	9.1 (8.1)
rs10850443				265	34.7 (5.4)	245	38.2 (7.2)	10.5 (16.0)	3.5 (5.3)	211	34.5 (5.8)	189	43.4 (11.2)	26.4 (24.1)	9.0 (8.8)
		<i>P</i> Value by GT						0.22	0.26					0.012	0.007
<i>LIPC</i>	0.36	A/G	HDL	404	34.6 (5.7)	376	38.2 (7.5)	11.7 (15.9)	3.9 (5.3)	429	34.5 (5.8)	387	43.2 (11.5)	25.8 (23.3)	8.9 (8.6)
15				465	35.1 (5.3)	428	38.4 (7.1)	9.4 (15.3)	3.2 (5.3)	472	34.5 (5.5)	430	44.5 (11.2)	29.0 (24.7)	10.0 (8.6)
rs1532085				151	36.1 (5.3)	133	39.6 (8.2)	8.1 (15.4)	3.0 (6.0)	132	35.1 (5.3)	121	44.9 (9.7)	29.2 (23.6)	10.0 (8.2)
		<i>P</i> Value by GT						0.006	0.026					0.037	0.031
<i>PABPC4</i>	0.23	G/A	HDL	617	35.1 (5.4)	567	38.8 (7.3)	11.0 (5.6)	3.7 (5.4)	580	34.9 (5.8)	530	44.4 (11.2)	27.2 (23.5)	9.4 (8.6)
1				361	35.3 (5.5)	332	38.3 (7.7)	9.1 (15.6)	3.1 (5.5)	390	34.3 (5.4)	354	43.8 (11.3)	28.4 (24.6)	9.7 (8.8)
rs4660293				42	32.9 (5.8)	38	34.7 (6.3)	7.3 (16.4)	2.1 (5.6)	64	34.0 (5.5)	55	43.8 (10.2)	28.7 (25.0)	9.7 (8.5)
		<i>P</i> Value by GT						0.029	0.031					0.33	0.41
<i>AMPD3</i>	0.18	G/A	HDL	664	35.1 (5.4)	617	38.4 (7.6)	9.8 (15.9)	3.3 (5.5)	698	34.6 (5.7)	633	44.2 (11.3)	28.2 (23.5)	9.7 (8.6)
11				323	35.1 (5.7)	292	38.8 (7.3)	10.9 (15.2)	3.7 (5.3)	300	34.6 (5.5)	275	43.9 (11.1)	27.7 (25.5)	9.5 (8.7)
rs2923084				33	33.4 (5.1)	28	36.8 (6.7)	9.9 (15.9)	3.2 (5.0)	36	34.9 (5.3)	31	41.2 (9.2)	18.2 (18.5)	6.3 (6.9)
		<i>P</i> Value by GT						0.28	0.30					0.26	0.2
<i>SPTLC3</i>	0.39	A/G	LDL	376	76.1 (22.8)	342	71.3 (19.1)	-2.7 (28.6)	-5.6 (24.1)	406	74.2 (21.8)	361	68.9 (20.7)	-1.1 (37.4)	-5.4 (25.5)
20				482	72.3 (21.0)	448	70.6 (18.5)	1.2 (33.1)	-3.3 (23.9)	479	73.1 (22.5)	443	65.6 (19.8)	-4.5 (38.1)	-8.5 (26.3)
rs364585				162	72.4 (22.4)	144	69.2 (7.3)	1.2 (34.0)	-3.6 (24.5)	149	73.2 (22.5)	134	65.1 (15.1)	-5.2 (37.3)	-9.0 (23.8)
		<i>P</i> Value by GT						0.08	0.22					0.11	0.043
<i>TOM1</i>	0.34	A/G	TC	456	144.4 (24.6)	413	143.9 (24.6)	1.5 (19.6)	-0.7 (29.5)	462	146.2 (27.6)	422	138.8 (27.5)	-3.5 (21.2)	-8.3 (32.0)
22				441	145.4 (26.4)	408	143.7 (24.4)	0.8 (19.3)	-1.9 (28.5)	437	142 (25.7)	392	136.5 (26.2)	-1.5 (22.2)	-5.2 (31.4)
rs138777				123	149.9 (30.2)	117	144.2 (8.4)	-0.08 (25.7)	-5.5 (38.1)	135	147.3 (36.2)	128	141 (26.4)	-2.2 (21.3)	-6.5 (33.9)
		<i>P</i> Value by GT						0.35	0.38					0.14	0.12
<i>PDXDC1</i>	0.39	T/C	TG	392	156 (128, 212)	365	156 (119, 207)	-4 (-23, 21)	-7 (-36, 32)	359	178 (140, 235)	331	117 (85, 160)	-32.0 (-49, -13)	-54 (-97, -21)
16				471	165 (134, 216)	431	158 (123, 206)	-3 (-26, 24)	-5 (-43, 35)	516	162 (130, 213)	466	120 (83, 171)	-29.4 (-48, -11.5)	-46 (-80, -2)
rs3198697				157	173 (136, 222)	142	154 (113, 207)	-9 (-29, 11)	-16 (-47, 20)	159	156 (127, 203)	145	124 (85, 167)	-24.5 (-46, -3)	-38 (-77, -5)
		<i>P</i> Value by GT						0.13	0.077					0.034	0.015
<i>CYP26A1</i>	0.44	A/G	TG	331	165 (135, 224)	306	161 (121, 213)	-4 (27, 20)	-7 (-44, 30)	311	165 (129, 225)	288	131 (90, 187)	-23 (-42, 2)	-36 (-76, 3)
10				493	159 (131, 203)	451	155 (118, 206)	-4 (-23, 22)	-7 (-44, 33)	525	171 (135, 217)	473	114 (82, 161)	-32 (-51, -11)	-54 (-92, -18)
rs2068888				196	161 (133, 222)	181	157 (123, 204)	-7 (-27, 21)	-11 (-45, 32)	198	161 (126, 210)	181	115 (83, 157)	-32 (-48, -10)	-48 (-85, -16)
		<i>P</i> Value by GT						0.40	0.59					<0.0001	0.001

Chr indicates chromosome; GT, genotype; HDL, high-density lipoprotein cholesterol; IQR, interquartile range; LDL, low-density lipoprotein cholesterol; MAF, minor allele frequency; SNP, single nucleotide polymorphism; TC, total cholesterol; TG, triglycerides. *TG reported as median (IQR).

Table 5. Nominally Significant Lipid Traits Associated With Coronary Artery Disease Events

Gene	<i>MVK</i>	<i>LIPC</i>	<i>PABPC4</i>	<i>AMPD3</i>	<i>SPTLC3</i>	<i>TOM1</i>	<i>PDXDC1</i>	<i>CYP26A1</i>
SNP	rs10850443	rs1532085	rs4660293	rs2923084	rs364585	rs138777	rs3198697	rs2068888
Lipid trait	HDL-C	HDL-C	HDL-C	HDL-C	LDL-C	TC	TG	TG
N	2054	2054	2054	2054	2054	2054	2054	2054
Composite end point								
OR	1.06	1.17	0.88	0.93	0.94	1.23	1.05	0.82
SE	0.12	0.12	0.15	0.16	0.12	0.12	0.12	0.12
<i>P</i> Value×int	0.63	0.20	0.41	0.64	0.62	0.10	0.67	0.11
Death from CHD								
OR	1.29	2.09	0.88	2.17	0.48	1.51	1.09	1.63
SE	0.32	0.32	0.41	0.35	0.37	0.32	0.32	0.33
<i>P</i> Value×int	0.43	0.021*	0.76	0.028*	0.05	0.20	0.78	0.13
Overall death								
OR	0.87	1.14	1.20	1.32	0.75	1.11	0.82	1.15
SE	0.21	0.21	0.25	0.25	0.22	0.22	0.22	0.21
<i>P</i> Value×int	0.51	0.53	0.46	0.27	0.19	0.63	0.35	0.50
Myocardial infarction								
OR	1.06	1.17	0.87	0.93	0.99	1.26	0.97	0.87
SE	0.19	0.19	0.24	0.25	0.20	0.19	0.20	0.20
<i>P</i> Value×int	0.74	0.42	0.58	0.76	0.94	0.23	0.88	0.49
Ischemic events								
OR	1.07	0.80	1.36	0.62	1.80	0.77	0.50	0.93
SE	0.42	0.45	0.50	0.64	0.43	0.48	0.50	0.44
<i>P</i> Value×int	0.87	0.62	0.55	0.45	0.17	0.59	0.17	0.86
Hospitalization from ACS								
OR	1.14	0.92	1.13	0.88	0.80	0.93	1.13	0.56
SE	0.22	0.23	0.27	0.29	0.23	0.24	0.22	0.24
<i>P</i> Value×int	0.55	0.73	0.65	0.67	0.34	0.76	0.60	0.017*
Symptom driven revascularization								
OR	1.22	1.11	0.79	0.71	1.04	1.27	1.10	0.60
SE	0.15	0.15	0.19	0.21	0.15	0.15	0.15	0.16
<i>P</i> Value×int	0.18	0.50	0.23	0.10	0.80	0.11	0.54	0.0013*

ACS indicates acute coronary syndrome; CHD, congenital heart disease; HDL, high-density lipoprotein cholesterol; LDL, low-density lipoprotein cholesterol; OR, odds ratio; *P*-value×int, interaction *P*-value; SE, standard error; SNP, single nucleotide polymorphism; TC, total cholesterol; TG, triglycerides.

**P* < 0.05.

the liver that metabolizes all-*trans*-retinoic acid, thereby regulating cellular levels of retinoic acid.³⁶ Retinoic acid binds the retinoid x receptor,³⁷ which plays an important role in lipid metabolism by itself and also by heterodimerizing with other well-known nuclear receptors such as peroxisome proliferator-activated receptors, farnesoid x receptor, and liver x receptor.^{38,39} Retinoic acid treatment has been shown to reduce TG levels in mice, whereas retinoid x receptor deletion induced the synthesis of TG.³⁹ In patients receiving oral

retinoids for the treatment of dermatological conditions, 44% experienced elevations in plasma triglycerides.⁴⁰ Treatment with bexarotene, a third-generation retinoid used in the treatment of T-cell lymphoma, results in hyperlipidemia in most patients, and fatal cases of cholestasis and pancreatitis have been reported.⁴¹ Recent GWAS studies investigating the polygenic genetic signal for the basis of CAD and several cardiovascular disease risk factors have shown that the liver x receptor/retinoid x receptor and farnesoid x receptor/

Table 6. Coronary Events by Genotype and Treatment Arm at Loci With a Nominally Significant Interaction

Gene/Chr/ SNP	Alleles			Placebo+Statin				Niacin+Statin		
	MAF	Minor/Major	Trait	GT	N	Frequency of Events	Odds Ratio (95% CI)	N	Frequency of Events	Odds Ratio (95% CI)
<i>LIPC</i>	0.36	A*/G	CAD Death	GG	404	4 (1%)	2.08 (1.11, 3.90)	429	9 (2.1%)	0.89 (0.48, 1.65)
15				AG	465	10 (2.2%)		472	13 (2.8%)	
rs1532085				AA	151	6 (4.0%)		132	2 (1.5%)	
			<i>P</i> Value by GT				0.02			0.71
<i>AMPD3</i>	0.18	G*/A	CAD Death	AA	664	9 (1.4%)	2.16 (1.09, 4.29)	698	11 (1.6%)	1.82 (0.96, 3.45)
11				AG	323	8 (2.5%)		300	12 (4.0%)	
rs2923084				GG	33	3 (9.1%)		36	1 (2.8%)	
			<i>P</i> Value by GT				0.03			0.07
<i>CYP26A1</i>	0.44	A/G*	ACS	GG	331	21 (6.3%)	1.85 (1.16, 2.77)	311	15 (4.8%)	1.20 (0.79, 1.85)
10				GA	493	17 (3.5%)		525	25 (4.8%)	
rs2068888				AA	196	4 (2.0%)		198	6 (3.0%)	
			<i>P</i> Value by GT				0.02			0.39
<i>CYP26A1</i>	0.44	A/G*	Revascularization	GG	331	41 (12.4%)	1.64 (1.20, 2.22)	311	29 (9.3%)	0.98 (0.71, 1.32)
10				GA	493	50 (10.1%)		525	49 (9.3%)	
rs2068888				AA	196	7 (3.6%)		198	19 (9.6%)	
			<i>P</i> value by GT				0.002			0.83

P-value within each treatment group determined by logistic regression adjusted for age, sex, BMI. ACS indicates acute coronary syndrome; BMI, body mass index; CAD, coronary artery disease; Chr, chromosome; CI, confidence interval; GT, genotype; MAF, minor allele frequency; SNP, single nucleotide polymorphism.

*Risk allele.

retinoid x receptor activation pathways are the top pathways enriched by CAD SNPs.⁴²

We were able to replicate the association with known variants at the *LPA* locus with baseline Lp(a) concentrations in AIM-HIGH. It was disappointing that none of these variants was associated with the change in Lp(a) in response to niacin. Thus, the mechanism by which niacin lowers Lp(a) is still unclear.

Previous GWAS studies have identified loci associated with statin response.^{14,15} A meta-analysis has identified 4 genetic loci, *APOE* (rs445925), *LPA* (rs10455872), *SORT1* (rs646776), and *SLCO1B1* (rs2900478), associated with percentage LDL-C reduction following statin therapy at a genome-wide level.¹⁴ Only *CETP* was identified with the change in HDL-C in response to statins.¹⁵ We did not find *APOE*, *LPA*, *SORT1*, or *CETP* to be associated with niacin response in our study. A candidate gene study found a SNP at the *APOA1* (rs964184) locus associated with fenofibrate response.¹⁶ This SNP was not genotyped in our cohort, so we were unable to determine whether it also mediated response to niacin. A pharmacogenetic analysis using a genome-wide approach in the ACCORD (Action to Control Cardiovascular Risk in Diabetes) trial identified *HSD17B3*, *SMAD3*, and *IPO11* as genetic markers of fenofibrate response.¹⁷

There are several limitations to this study. First, the sample size in our study is small. To detect the association of *LIPC* SNP rs1532085 and change in HDL-C, the study was powered at 87% and 26% for α thresholds of 0.05 and 0.0002, respectively. The latter threshold using the Bonferroni method is likely too conservative because it does not acknowledge any prior information. In our study we were informing our analysis using prior knowledge about variants that are known to be associated with steady-state plasma lipid values to look for a differential effect in response to niacin. Therefore, the true power of the study is likely between these 2 threshold values. Second, we did not have access to another large cohort on chronic niacin treatment to replicate our findings, and our findings would require replication. It would be interesting to determine if our findings would replicate in the HPS2-THRIVE study, which used a similar trial design.⁸ Third, we only evaluated the role of niacin on lipid-dependent mechanisms on coronary disease risk, and we did not evaluate known lipid-independent genes, as niacin has been shown to display anti-inflammatory and antioxidant effects.⁴³⁻⁴⁶ Last, there were a small number of black participants in the AIM-HIGH study, so we were unable to examine genetic predictors of niacin response in this ethnic group. The lower participation

is unfortunate in light of evidence that blacks are already known to have a significantly different response to niacin, at least in terms of triglyceride lowering⁴⁷ and adverse effects.⁴⁸

In conclusion, we have identified several genetic variants that are associated with the lipid response to niacin treatment. The association with genetic variation at *LIPC* encoding hepatic lipase is particularly interesting in that niacin has been previously suggested to modulate hepatic lipase activity. Although our results require replication, they represent the first pharmacogenetic study of lipid response to niacin and implicate *LIPC* and *CYP26A1* as potential mediators of niacin's effects on lipids and cardiovascular events.

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Disclosures

None.

References

- Canner PL, Berge KG, Wenger NK, Stamler J, Friedman L, Prineas RJ, Friedewald W. Fifteen year mortality in coronary drug project patients: long-term benefit with niacin. *J Am Coll Cardiol*. 1986;8:1245–1255.
- Carlson LA. Nicotinic acid: the broad-spectrum lipid drug. A 50th anniversary review. *J Intern Med*. 2005;258:94–114.
- Kamstrup PR, Tybjaerg-Hansen A, Steffensen R, Nordestgaard BG. Genetically elevated lipoprotein(a) and increased risk of myocardial infarction. *JAMA*. 2009;301:2331–2339.
- Clarke R, Peden JF, Hopewell JC, Kyriakou T, Goel A, Heath SC, Parish S, Barlera S, Franzosi MG, Rust S, Bennett D, Silveira A, Malarstig A, Green FR, Lathrop M, Gigante B, Leander K, de Faire U, Seedorf U, Hamsten A, Collins R, Watkins H, Farrall M. Genetic variants associated with lipoprotein level and coronary disease. *N Engl J Med*. 2009;361:2518–2528.
- Taylor AJ, Sullenberger LE, Lee HJ, Lee JK, Grace KA. Arterial biology for the investigation of the treatment effects of reducing cholesterol (arbitrator 2): a double-blind, placebo-controlled study of extended-release niacin on atherosclerosis progression in secondary prevention patients treated with statins. *Circulation*. 2004;110:3512–3517.
- Taylor AJ, Villines TC, Stanek EJ, Devine PJ, Griffen L, Miller M, Weissman NJ, Turco M. Extended-release niacin or ezetimibe and carotid intima-media thickness. *N Engl J Med*. 2009;361:2113–2122.
- Boden WE, Probstfield JL, Anderson T, Chaitman BR, Desvignes-Nickens P, Koprowicz K, McBride R, Teo K, Weintraub W. Niacin in patients with low HDL cholesterol levels receiving intensive statin therapy. *N Engl J Med*. 2011;365:2255–2267.
- Landray MJ, Haynes R, Hopewell JC, Parish S, Aung T, Tomson J, Wallendszus K, Craig M, Jiang L, Collins R, Armitage J. Effects of extended-release niacin with laropiprant in high-risk patients. *N Engl J Med*. 2014;371:203–212.
- Kamanna VS, Ganji SH, Kashyap ML. Recent advances in niacin and lipid metabolism. *Curr Opin Lipidol*. 2013;24:239–245.
- vander Hoorn JW, de Haan W, Berbee JF, Havekes LM, Jukema JW, Rensen PC, Princen HM. Niacin increases HDL by reducing hepatic expression and plasma levels of cholesteryl ester transfer protein in APOE*3Leiden.CETP mice. *Arterioscler Thromb Vasc Biol*. 2008;28:2016–2022.
- Tuteja S, Wang L, Dunbar RL, Chen J, DerOhannessian S, Marcovina SM, Elam M, Lader E, Rader DJ. Genetic coding variants in the niacin receptor, hydroxyl-carboxylic acid receptor 2, and response to niacin therapy. *Pharmacogenet Genomics*. 2017;27:285–293.
- Willer CJ, Schmidt EM, Sengupta S, Peloso GM, Gustafsson S, Kanoni S, Ganna A, Chen J, Buchkovich ML, Mora S, Beckmann JS, Bragg-Gresham JL, Chang HY, Demirkan A, Den Hertog HM, Do R, Donnelly LA, Ehret GB, Esko T, Feitosa MF, Ferreira T, Fischer K, Fontanillas P, Fraser RM, Freitag DF, Gudrasani D, Heikkilä K, Hyppönen E, Isaacs A, Jackson AU, Johansson A, Johnson T, Kaakinen M, Kettunen J, Kleber ME, Li X, Luan J, Lyttikainen LP, Magnusson PK, Mangino M, Mihailov E, Montasser ME, Muller-Nurasyid M, Nolte IM, O'Connell JR, Palmer CD, Perola M, Petersen AK, Sanna S, Saxena R, Service SK, Shah S, Shungin D, Sidore C, Song C, Strawbridge RJ, Surakka I, Tanaka T, Teslovich TM, Thorleifsson G, Van den Herik EG, Voight BF, Volcik KA, Waite LL, Wong A, Wu Y, Zhang W, Absher D, Asiki G, Barroso I, Been LF, Bolton JL, Bonnycastle LL, Brambilla P, Burnett MS, Cesana G, Dimitriou M, Doney AS, Doring A, Elliott P, Epstein SE, Eyjolfsson GI, Gigante B, Goodarzi MO, Grallert H, Gravito ML, Groves CJ, Hallmans G, Hartikainen AL, Hayward C, Hernandez D, Hicks AA, Holm H, Hung YJ, Illig T, Jones MR, Kaleebu P, Kastelein JJ, Khaw KT, Kim E, Klopp N, Komulainen P, Kumari M, Langenberg C, Lehtimäki T, Lin SY, Lindstrom J, Loos RJ, Mach F, McArdle WL, Meisinger C, Mitchell BD, Muller G, Nagaraja R, Narisu N, Nieminen TV, Nsubuga RN, Olafsson I, Ong KK, Palotie A, Papamarkou T, Pomilla C, Pouta A, Rader DJ, Reilly MP, Ridker PM, Rivadeneira F, Rudan I, Ruukonen A, Samani N, Scharnagl H, Seeley J, Silander K, Stancakova A, Stirrups K, Swift AJ, Tiret L, Uitterlinden AG, van Pelt LJ, Vedantam S, Wainwright N, Wijmenga C, Wild SH, Willemsen G, Wilsgaard T, Wilson JF, Young EH, Zhao JH, Adair LS, Arveiler D, Assimes TL, Bandinelli S, Bennett F, Bochud M, Boehm BO, Boomsma DI, Borecki IB, Bornstein SR, Bovet P, Burnier M, Campbell H, Chakravarti A, Chambers JC, Chen YD, Collins FS, Cooper RS, Danesh J, Dedoussis G, de Faire U, Feranil AB, Ferrières J, Ferrucci L, Freimer NB, Gieger C, Groop LC, Gudnason V, Gyllenstein U, Hamsten A, Harris TB, Hingorani A, Hirschhorn JN, Hofman A, Hovingh GK, Hsiung CA, Humphries SE, Hunt SC, Hveem K, Iribarren C, Jarvelin MR, Jula A, Kahonen M, Kaprio J, Kesaniemi A, Kivimäki M, Kooner JS, Koudstaal PJ, Krauss RM, Kuh D, Kuusisto J, Kyvik KO, Laakso M, Lakka TA, Lind L, Lindgren CM, Martin NG, Marz W, McCarthy MI, McKenzie CA, Meneton P, Metspalu A, Moilanen L, Morris AD, Munroe PB, Njolstad I, Pedersen NL, Power C, Pramstaller PP, Price JF, Psaty BM, Quertermous T, Rauramaa R, Saleheen D, Salomaa V, Sanghera DK, Saramies J, Schwarz PE, Sheu WH, Shuldiner AR, Siegbahn A, Spector TD, Stefansson K, Strachan DP, Tayo BO, Tremoli E, Tuomilehto J, Uusitupa M, van Duijn CM, Vollenweider P, Wallentin L, Wareham NJ, Whitfield JB, Wolfenbutter BH, Ordovas JM, Boerwinkle E, Palmer CN, Thorsteinsdottir U, Chasman DI, Rotter JJ, Franks PW, Ripatti S, Cupples LA, Sandhu MS, Rich SS, Boehnke M, Deloukas P, Kathiresan S, Mohlke KL, Ingelsson E, Abecasis GR. Discovery and refinement of loci associated with lipid levels. *Nat Genet*. 2013;45:1274–1283.
- Mack S, Coassin S, Rueedi R, Youssi NA, Seppala I, Gieger C, Schonherr S, Forer L, Erhart G, Marques-Vidal P, Ried JS, Waeber G, Bergmann S, Dahnhardt D, Stockl A, Raitakari OT, Kahonen M, Peters A, Meitinger T, Strauch K, Kedenko L, Paulweber B, Lehtimäki T, Hunt SC, Vollenweider P, Lamina C, Kronenberg F. A genome-wide association meta-analysis on lipoprotein (a) concentrations adjusted for apolipoprotein (a) isoforms. *J Lipid Res*. 2017;58:1834–1844.
- Postmus I, Trompet S, Deshmukh HA, Barnes MR, Li X, Warren HR, Chasman DI, Zhou K, Arsenault BJ, Donnelly LA, Wiggins KL, Avery CL, Griffin P, Feng Q, Taylor KD, Li G, Evans DS, Smith AV, de Keyser CE, Johnson AD, de Craen AJ, Stott DJ, Buckley BM, Ford I, Westendorp RG, Slagboom PE, Sattar N, Munroe PB, Sever P, Poulter N, Stanton A, Shields DC, O'Brien E, Shaw-Hawkins S, Chen YD, Nickerson DA, Smith JD, Dube MP, Boekholdt SM, Hovingh GK, Kastelein JJ, McKeigue PM, Betteridge J, Neil A, Durrington PN, Doney A, Carr F, Morris A, McCarthy MI, Groop L, Ahlqvist E, Bis JC, Rice K, Smith NL, Lumley T, Whitsel EA, Sturmer T, Boerwinkle E, Ngwa JS, O'Donnell CJ, Vasan RS, Wei WQ, Wilke RA, Liu CT, Sun F, Guo X, Heckbert SR, Post W, Sotoodehnia N, Arnold AM, Stafford JM, Ding J, Herrington DM, Kritchevsky SB, Eiriksdottir G, Launer LJ, Harris TB, Chu AY, Giulianini F, MacFadyen JG, Barratt BJ, Nyberg F, Stricker BH, Uitterlinden AG, Hofman A, Rivadeneira F, Emilsson V, Franco OH, Ridker PM, Gudnason V, Liu Y, Denny JC, Ballantyne CM, Rotter JJ, Adhikari Cupples L, Psaty BM, Palmer CN, Tardif JC, Colhoun HM, Hitman G, Krauss RM, Wouter Jukema J, Caulfield MJ. Pharmacogenetic meta-analysis of genome-wide association studies of LDL cholesterol response to statins. *Nat Commun*. 2014;5:5068.
- Postmus I, Warren HR, Trompet S, Arsenault BJ, Avery CL, Bis JC, Chasman DI, de Keyser CE, Deshmukh HA, Evans DS, Feng Q, Li X, Smit RA, Smith AV, Sun F, Taylor KD, Arnold AM, Barnes MR, Barratt BJ, Betteridge J, Boekholdt SM, Boerwinkle E, Buckley BM, Chen YI, de Craen AJ, Cummings SR, Denny JC,

- Dube MP, Durrington PN, Eiriksdottir G, Ford I, Guo X, Harris TB, Heckbert SR, Hofman A, Hovingh GK, Kastelein JJ, Launer LJ, Liu CT, Liu Y, Lumley T, McKeigue PM, Munroe PB, Neil A, Nickerson DA, Nyberg F, O'Brien E, O'Donnell CJ, Post W, Poulter N, Vasan RS, Rice K, Rich SS, Rivadeneira F, Sattar N, Sever P, Shaw-Hawkins S, Shields DC, Slagboom PE, Smith NL, Smith JD, Sotoodehnia N, Stanton A, Stott DJ, Stricker BH, Sturmer T, Uitterlinden AG, Wei WQ, Westendorp RG, Whitless EA, Wiggins KL, Wilke RA, Ballantyne CM, Colhoun HM, Cupples LA, Franco OH, Gudnason V, Hitman G, Palmer CN, Psaty BM, Ridker PM, Stafford JM, Stein CM, Tardif JC, Caulfield MJ, Jukema JW, Rotter JJ, Krauss RM. Meta-analysis of genome-wide association studies of HDL cholesterol response to statins. *J Med Genet*. 2016;53:835–845.
16. Aslibekyan S, Goodarzi MO, Frazier-Wood AC, Yan X, Irvin MR, Kim E, Tiwari HK, Guo X, Straka RJ, Taylor KD, Tsai MY, Hopkins PN, Korenman SG, Borecki IB, Chen YD, Ordovas JM, Rotter JJ, Arnett DK. Variants identified in a GWAS meta-analysis for blood lipids are associated with the lipid response to fenofibrate. *PLoS One*. 2012;7:e48663.
 17. Rotroff DM, Pijut SS, Marvel SW, Jack JR, Havener TM, Pujol A, Schluter A, Graf GA, Ginsberg HN, Shah HS, Gao H, Morieri ML, Doria A, Mychaleckyi JC, McLeod HL, Buse JB, Wagner MJ, Moutsier-Reif AA. Genetic variants in HSD17B3, SMAD3, and IPO11 impact circulating lipids in response to fenofibrate in individuals with type 2 diabetes. *Clin Pharmacol Ther*. 2018;103:712–721.
 18. Biological specimen and data repository information coordinating center (BioLINCC). National heart lung and blood institute. Available at: <https://biolincc.nhlbi.nih.gov/home/>. Accessed February 15, 2015.
 19. Voight BF, Kang HM, Ding J, Palmer CD, Sidore C, Chines PS, Burt NP, Fuchsberger C, Li Y, Erdmann J, Frayling TM, Heid IM, Jackson AU, Johnson T, Kilpelainen TO, Lindgren CM, Morris AP, Prokopenko I, Randall JC, Saxena R, Soranzo N, Speliotes EK, Teslovich TM, Wheeler E, Maguire J, Parkin M, Pottter S, Rayner NW, Robertson N, Stirrups K, Winckler W, Sanna S, Mulas A, Nagaraja R, Cucca F, Barroso I, Deloukas P, Loos RJ, Kathiresan S, Munroe PB, Newton-Cheh C, Pfeufer A, Samani NJ, Schunkert H, Hirschhorn JN, Altshuler D, McCarthy MI, Abecasis GR, Boehnke M. The metabochip, a custom genotyping array for genetic studies of metabolic, cardiovascular, and anthropometric traits. *PLoS Genet*. 2012;8:e1002793.
 20. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, Maller J, Sklar P, de Bakker PI, Daly MJ, Sham PC. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet*. 2007;81:559–575.
 21. Johnson AD, Handsaker RE, Pulit SL, Nizzari MM, O'Donnell CJ, de Bakker PIW. Snap: a web-based tool for identification and annotation of proxy SNPs using hapmap. *Bioinformatics*. 2008;24:2938–2939.
 22. R Core Team. R, a language and environment for statistical computing, R foundation for statistical computing. Vienna, Austria. 2017. Available at: <https://r-project.org>. Accessed November 14, 2017.
 23. Albers JJ, Slee A, O'Brien KD, Robinson JG, Kashyap ML, Kwiterovich PO Jr, Xu P, Marcovina SM. Relationship of apolipoproteins a-1 and b, and lipoprotein(a) to cardiovascular outcomes: the AIM-HIGH trial (Atherosclerosis Intervention in Metabolic Syndrome Leth Low HDL/High Triglyceride and Impact on Global Health Outcomes). *J Am Coll Cardiol*. 2013;62:1575–1579.
 24. Wink J, Giacoppe G, King J. Effect of very-low-dose niacin on high-density lipoprotein in patients undergoing long-term statin therapy. *Am Heart J*. 2002;143:514–518.
 25. Santamarina-Fojo S, González-Navarro H, Freeman L, Wagner E, Nong Z. Hepatic lipase, lipoprotein metabolism, and atherogenesis. *Arterioscler Thromb Vasc Biol*. 2004;24:1750.
 26. Kuusi T, Saarinen P, Nikkila EA. Evidence for the role of hepatic endothelial lipase in the metabolism of plasma high density lipoprotein2 in man. *Atherosclerosis*. 1980;36:589–593.
 27. Zambon A, Deeb SS, Hokanson JE, Brown BG, Brunzell JD. Common variants in the promoter of the hepatic lipase gene are associated with lower levels of hepatic lipase activity, buoyant LDL, and higher HDL₂ cholesterol. *Arterioscler Thromb Vasc Biol*. 1998;18:1723–1729.
 28. Zambon A, Deeb SS, Brown BG, Hokanson JE, Brunzell JD. Common hepatic lipase gene promoter variant determines clinical response to intensive lipid-lowering treatment. *Circulation*. 2001;103:792–798.
 29. Teslovich TM, Musunuru K, Smith AV, Edmondson AC, Stylianou IM, Koseki M, Pirruccello JP, Ripatti S, Chasman DI, Willer CJ, Johansen CT, Fouchier SW, Isaacs A, Peloso GM, Barbalic M, Ricketts SL, Bis JC, Aulchenko YS, Thorleifsson G, Feitosa MF, Chambers J, Orho-Melander M, Melander O, Johnson T, Li X, Guo X, Li M, Shin Cho Y, Jin Go M, Jin Kim Y, Lee JY, Park T, Kim K, Sim X, Twee-Hee Ong R, Croteau-Chonka DC, Lange LA, Smith JD, Song K, Hua Zhao J, Yuan X, Luan J, Lamina C, Ziegler A, Zhang W, Zee RY, Wright AF, Witteman JC, Wilson JF, Willemssen G, Wichmann HE, Whitfield JB, Waterworth DM, Wareham NJ, Waeber G, Vollenweider P, Voight BF, Vitart V, Uitterlinden AG, Uda M, Tuomilehto J, Thompson JR, Tanaka T, Surakka I, Stringham HM, Spector TD, Soranzo N, Smit JH, Sinisalo J, Silander K, Sijbrands EJ, Scuteri A, Scott J, Schlessinger D, Sanna S, Salomaa V, Saharinen J, Sabatti C, Ruukonen A, Rudan I, Rose LM, Roberts R, Rieder M, Psaty BM, Pramstaller PP, Pichler I, Perola M, Penninx BW, Pedersen NL, Pattaro C, Parker AN, Pare G, Oostra BA, O'Donnell CJ, Nieminen MS, Nickerson DA, Montgomery GW, Meitinger T, McPherson R, McCarthy MI, McArdle W, Masson D, Martin NG, Marroni F, Mangino M, Magnusson PK, Lucas G, Luben R, Loos RJ, Lokki ML, Lettre G, Langenberg C, Launer LJ, Lakatta EG, Laaksonen R, Kyvik KO, Kronenberg F, König IR, Khaw KT, Kaprio J, Kaplan LM, Johansson A, Jarvelin MR, Janssens AC, Ingelsson E, Igl W, Kees Hovingh G, Hottenga JJ, Hofman A, Hicks AA, Hengstenberg C, Heid IM, Hayward C, Havulinna AS, Hastie ND, Harris TB, Haritunians T, Hall AS, Gyllenstein U, Guiducci C, Groop LC, Gonzalez E, Gieger C, Freimer NB, Ferrucci L, Erdmann J, Elliott P, Ejebe KG, Doring A, Dominiczak AF, Demissie S, Deloukas P, de Geus EJ, de Faire U, Crawford G, Collins FS, Chen YD, Caulfield MJ, Campbell H, Burt NP, Bonnycastle LL, Boomsma DI, Boekholdt SM, Bergman RN, Barroso I, Bandinelli S, Ballantyne CM, Assimes TL, Quertermous T, Altshuler D, Seielstad M, Wong TY, Tai ES, Feranil AB, Kuzawa CW, Adair LS, Taylor HA Jr, Borecki IB, Gabriel SB, Wilson JG, Holm H, Thorsteinsdottir U, Gudnason V, Krauss RM, Mohlke KL, Ordovas JM, Munroe PB, Kooner JS, Tall AR, Hegele RA, Kastelein JJ, Schadt EE, Rotter JJ, Boerwinkle E, Strachan DP, Mooser V, Stefansson K, Reilly MP, Samani NJ, Schunkert H, Cupples LA, Sandhu MS, Ridker PM, Rader DJ, van Duijn CM, Peltonen L, Abecasis GR, Boehnke M, Kathiresan S. Biological, clinical and population relevance of 95 loci for blood lipids. *Nature*. 2010;466:707–713.
 30. Zanon P, Khetarpal SA, Larach DB, Hancock-Cerutti WF, Millar JS, Cuchel M, DerOhannessian S, Kontush A, Surendran P, Saleheen D, Trompet S, Jukema JW, De Craen A, Deloukas P, Sattar N, Ford I, Packard C, Majumder A, Alam DS, Di Angelantonio E, Abecasis G, Chowdhury R, Erdmann J, Nordestgaard BG, Nielsen SF, Tybjaerg-Hansen A, Schmidt RF, Kuulasmaa K, Liu DJ, Perola M, Blankenbreg S, Salomaa V, Mannisto S, Amouyel P, Arveiler D, Ferrières J, Muller-Nurasyid M, Ferrario M, Kee F, Willer CJ, Samani N, Schunkert H, Butterworth AS, Howson JM, Peloso GM, Stitzel NO, Danesh J, Kathiresan S, Rader DJ. Rare variant in scavenger receptor BI raises HDL cholesterol and increases risk of coronary heart disease. *Science*. 2016;351:1166–1171.
 31. Tuteja S, Rader DJ. High-density lipoproteins in the prevention of cardiovascular disease: changing the paradigm. *Clin Pharmacol Ther*. 2014;96:48–56.
 32. Schunkert H, König IR, Kathiresan S, Reilly MP, Assimes TL, Holm H, Preuss M, Stewart AFR, Barbalic M, Gieger C, Absher D, Aherrahrou Z, Allayee H, Altshuler D, Anand SS, Andersen K, Anderson JL, Ardisson D, Ball SG, Balmforth AJ, Barnes TA, Becker DM, Becker LC, Berger K, Bis JC, Boekholdt SM, Boerwinkle E, Braund PS, Brown MJ, Burnett MS, Buyschaert I, Carlquist JF, Chen L, Cichon S, Codd V, Davies RW, Dedoussis G, Dehghan A, Demissie S, Devaney JM, Diemert P, Do R, Doering A, Eifer S, Mokhtari NEE, Ellis SG, Elosua R, Engert JC, Epstein SE, de Faire U, Fischer M, Folsom AR, Freyer J, Gigante B, Giarelli D, Gretarsdottir S, Gudnason V, Gulcher JR, Halperin E, Hammond N, Hazen SL, Hofman A, Horne BD, Illig T, Iribarren C, Jones GT, Jukema JW, Kaiser MA, Kaplan LM, Kastelein JJP, Kaw K-T, Knowles JW, Kolovou G, Kong A, Laaksonen R, Lambrechts D, Leander K, Lettre G, Li M, Lieb W, Loley C, Lotery AJ, Mannucci PM, Maouche S, Martinelli N, McKeown PP, Meisinger C, Meitinger T, Melander O, Merlini PA, Mooser V, Morgan T, Muhleisen TW, Muhlestein JB, Munzel T, Musunuru K, Nahrstaedt J, Nelson CP, Nothen MM, Olivieri O, Patel RS, Patterson CC, Peters A, Peyvandi F, Ou L, Quyyumi AA, Rader DJ, Rallidis LS, Rice C, Rosendaal FR, Rubin D, Salomaa V, Sampietro ML, Sandhu MS, Schadt E, Schafer A, Schillert A, Schreiber S, Schrezenmeier J, Schwartz SM, Siscovick DS, Sivananthan M, Sivapalratnam S, Smith A, Smith TB, Snoop JD, Soranzo N, Spertus JA, Stark K, Stirrups K, Stoll M, Tang WHW, Tenstedt S, Thorleifsson G, Thorleifsson G, Tomaszewski M, Uitterlinden AG, van Rij AM, Voight BF, Wareham NJ, Wells GA, Wichmann HE, Wild PS, Willenborg C, Witteman JCM, Wright BJ, Ye S, Zeller T, Ziegler A, Cambien F, Goodall AH, Cupples LA, Quertermous T, Marz W, Hengstenberg C, Blankenbreg S, Ouwehand WH, Hall AS, Deloukas P, Thompson JR, Stefansson K, Roberts R, Thorsteinsdottir U, O'Donnell CJ, McPherson R, Erdmann J. Large-scale association analysis identifies 13 new susceptibility loci for coronary artery disease. *Nat Genet*. 2011;43:333–338.
 33. Broad Institute of MIT and Harvard. Cardiovascular disease knowledge portal. Available at: <http://broadcvdi.org/gene/geneinfo/lipc>. Accessed April 6, 2018.
 34. Broad Institute of MIT and Harvard. Cardiovascular disease knowledge portal. Available at: <http://broadcvdi.org/gene/geneinfo/cyp26a1>. Accessed April 6, 2018.
 35. Ellinor PT, Lunetta KL, Albert CM, Glazer NL, Ritchie MD, Smith AV, Arking DE, Muller-Nurasyid M, Krijthe BP, Lubitz SA, Bis JC, Chung MK, Dorr M, Ozaki K, Roberts JD, Smith JG, Pfeufer A, Sinner MF, Lohman K, Ding J, Smith NL, Smith JD, Rienstra M, Rice KM, Van Wagoner DR, Magnani JW, Wakili R, Clauss S, Rotter JJ, Steinbeck G, Launer LJ, Davies RW, Borkovich M, Harris TB, Lin H, Volker U, Volzke H, Milan DJ, Hofman A, Boerwinkle E, Chen LY, Soliman EZ, Voight BF, Li G, Chakravarti A, Kubo M, Tedrow UB, Rose LM, Ridker PM, Conen D, Tsunoda T, Furukawa T, Sotoodehnia N, Xu S, Kamatani N, Levy D, Nakamura Y, Parvez B, Mahida S, Furie KL, Rosand J, Muhammad R, Psaty BM, Meitinger T, Perz S, Wichmann HE, Witteman JC, Kao WH, Kathiresan S, Roden DM, Uitterlinden AG, Rivadeneira F, McKnight B, Sjogren M, Newman AB, Liu Y,

- Gollob MH, Melander O, Tanaka T, Stricker BH, Felix SB, Alonso A, Darbar D, Barnard J, Chasman DI, Heckbert SR, Benjamin EJ, Gudnason V, Kaab S. Meta-analysis identifies six new susceptibility loci for atrial fibrillation. *Nat Genet*. 2012;44:670–675.
36. Thatcher JE, Zelter A, Isoherranen N. The relative importance of CYP26A1 in hepatic clearance of all-*trans* retinoic acid. *Biochem Pharmacol*. 2010;80:903–912.
37. Bushue N, Wan YJ. Retinoid pathway and cancer therapeutics. *Adv Drug Deliv Rev*. 2010;62:1285–1298.
38. Mukherjee R, Strasser J, Jow L, Hoener P, Paterniti JR Jr, Heyman RA. RXR agonists activate PPAR α -inducible genes, lower triglycerides, and raise HDL levels in vivo. *Arterioscler Thromb Vasc Biol*. 1998;18:272–276.
39. He Y, Gong L, Fang Y, Zhan Q, Liu HX, Lu Y, Guo GL, Lehman-McKeeman L, Fang J, Wan YJ. The role of retinoic acid in hepatic lipid homeostasis defined by genomic binding and transcriptome profiling. *BMC Genom*. 2013;14:575.
40. Lilley JS, Linton MF, Fazio S. Oral retinoids and plasma lipids. *Dermatol Ther*. 2013;26:404–410.
41. Bexarotene (Targretin) [US prescribing information]. Available at: http://www.accessdata.fda.gov/drugsatfda_docs/label/1999/21055lbl.pdf. Accessed March 24, 2017.
42. LeBlanc M, Zuber V, Andreassen BK, Witoelar A, Zeng L, Bettella F, Wang Y, McEvoy LK, Thompson WK, Schork AJ, Reppe S, Barrett-Connor E, Ligthart S, Dehghan A, Gautvik KM, Nelson CP, Schunkert H, Samani NJ, Ridker PM, Chasman DI, Aukrust P, Djurovic S, Frigessi A, Desikan RS, Dale AM, Andreassen OA. Identifying novel gene variants in coronary artery disease and shared genes with several cardiovascular risk factors. *Circ Res*. 2016;118:83–94.
43. Digby JE, McNeill E, Dyar OJ, Lam V, Greaves DR, Choudhury RP. Anti-inflammatory effects of nicotinic acid in adipocytes demonstrated by suppression of fractalkine, RANTES, and MCP-1 and upregulation of adiponectin. *Atherosclerosis*. 2010;209:89–95.
44. Chai JT, Digby JE, Choudhury RP. GPR109A and vascular inflammation. *Curr Atheroscler Rep*. 2013;15:325.
45. Ganji SH, Kashyap ML, Kamanna VS. Niacin inhibits fat accumulation, oxidative stress, and inflammatory cytokine IL-8 in cultured hepatocytes: impact on non-alcoholic fatty liver disease. *Metabolism*. 2015;64:982–990.
46. Wu BJ, Yan L, Charlton F, Witting P, Barter PJ, Rye K-A. Evidence that niacin inhibits acute vascular inflammation and improves endothelial dysfunction independent of changes in plasma lipids. *Arterioscler Thromb Vasc Biol*. 2010;30:968–975.
47. Usman MH, Qamar A, Gadi R, Lilly S, Goel H, Hampson J, Mucksavage ML, Nathanson GA, Rader DJ, Dunbar RL. Extended-release niacin acutely suppresses postprandial triglyceridemia. *Am J Med*. 2012;125:1026–1035.
48. Tuteja S, Dunbar R, Qu L, Li M, Mucksavage ML, DerOhannessian SL, Reilly M, Rader D. *Pharmacogenomics of the flushing response to acute niacin administration*. Abstract. Indianapolis, IN: American Society of Clinical Pharmacology and Therapeutics; March 2013.

SUPPLEMENTAL MATERIAL

Table S1. SNPs tested and association with baseline lipid traits.

SNP	Trait	Locus	Chr	MAF	N	Effect of minor allele				Proxy	GLGC SNP	r2
						Beta	SE	Stat	Pvalue			
rs12748152	HDL	PIGV-NR0B2	1	0.08	2054	-0.006	0.053	-0.12	0.91			
rs12145743	HDL	HDGF-PMVK	1	0.34	2054	-0.018	0.030	-0.59	0.56			
rs1689800	HDL	ZNF648	1	0.37	2054	-0.015	0.030	-0.51	0.61			
rs4650994	HDL	ANGPTL1	1	0.47	2054	0.0004	0.028	0.01	0.99			
rs4660293	HDL	PABPC4	1	0.24	2054	-0.065	0.034	-1.89	0.06			
rs4846914	HDL	GALNT2	1	0.41	2054	-0.046	0.029	-1.58	0.11			
rs12328675	HDL	COBLL1	2	0.12	2054	-0.059	0.045	-1.30	0.19			
rs2972146	HDL	IRS1	2	0.34	2054	0.029	0.031	0.92	0.36			
rs13326165	HDL	STAB1	3	0.20	2054	-0.023	0.036	-0.63	0.53			
rs2013208	HDL	RBMS5	3	0.48	2054	0.037	0.029	1.28	0.20			
rs2290547	HDL	SETD2	3	0.18	2054	0.055	0.038	1.45	0.15			
rs2606736	HDL	ATG7	3	0.38	2054	-0.011	0.030	-0.36	0.72			
rs6805251	HDL	GSK3B	3	0.38	2054	-0.049	0.030	-1.66	0.10			
rs10019888	HDL	C4orf52	4	0.17	2054	0.010	0.038	0.28	0.78			
rs13107325	HDL	SLC39A8	4	0.08	2054	-0.018	0.053	-0.35	0.73			
rs2602836	HDL	ADH5	4	0.44	2054	-0.030	0.029	-1.03	0.31			
rs3822072	HDL	FAM13A	4	0.48	2054	0.002	0.029	0.07	0.94			
rs6450176	HDL	ARL15	5	0.26	2054	-0.064	0.032	-1.98	0.05			
rs1936800	HDL	RSPO3	6	0.48	2054	0.028	0.029	0.98	0.33			
rs634869	HDL	CITED2	6	0.43	2054	-0.005	0.029	-0.18	0.85	Y	rs605066	1.00
rs17173637	HDL	TMEM176A	7	0.10	2054	-0.094	0.049	-1.93	0.05			
rs4142995	HDL	SNX13	7	0.39	2054	0.009	0.029	0.31	0.76			
rs4731702	HDL	KLF14	7	0.48	2054	0.001	0.028	0.02	0.99			
rs4917014	HDL	IKZF1	7	0.32	2054	0.005	0.031	0.16	0.87			
rs702485	HDL	DAGLB	7	0.45	2054	-0.005	0.029	-0.16	0.87			
rs2293889	HDL	TRPS1	8	0.44	2054	0.006	0.029	0.20	0.84			
rs9987289	HDL	PPP1R3B	8	0.09	2054	-0.028	0.050	-0.55	0.58			
rs1883025	HDL	ABCA1	9	0.26	2054	-0.012	0.032	-0.36	0.72			
rs581080	HDL	TTC39B	9	0.19	2054	-0.087	0.037	-2.36	0.02			
rs970548	HDL	MARCH8-ALOX5	10	0.25	2054	-0.017	0.033	-0.50	0.61			
rs11246602	HDL	OR4C46	11	0.12	2054	0.042	0.045	0.93	0.35			
rs12801636	HDL	KAT5	11	0.22	2054	-0.021	0.034	-0.61	0.54			
rs2923084	HDL	AMPD3	11	0.19	2054	-0.025	0.037	-0.66	0.51			
rs3136441	HDL	LRP4	11	0.12	2054	0.130	0.045	2.90	0.0037			
rs499974	HDL	MOGAT2-DGAT2	11	0.17	2054	-0.056	0.038	-1.47	0.14			
rs10850443	HDL	MVK	12	0.47	2054	-0.001	0.028	-0.05	0.96	Y	rs7134594	0.97
rs10773003	HDL	SBNO1	12	0.09	2043	0.048	0.051	0.94	0.35	Y	rs4759375	1.00
rs11057408	HDL	ZNF664	12	0.33	2054	0.009	0.030	0.30	0.77	Y	rs4765127	1.00
rs7134375	HDL	PDE3A	12	0.42	2054	0.049	0.029	1.67	0.10			
rs838880	HDL	SCARB1	12	0.31	2054	0.063	0.031	2.04	0.04			
rs4983559	HDL	ZBTB42-AKT1	14	0.39	2054	0.037	0.029	1.25	0.21			
rs1532085	HDL	LIPC	15	0.37	2054	0.106	0.030	3.58	0.00035			
rs2652834	HDL	LACTB	15	0.20	2054	-0.002	0.036	-0.06	0.95			
rs1121980	HDL	FTO	16	0.44	2054	-0.037	0.029	-1.26	0.21			
rs16942887	HDL	LCAT	16	0.12	2054	0.087	0.045	1.94	0.05			
rs2925979	HDL	CMIP	16	0.31	2054	0.001	0.031	0.04	0.97			
rs3764261	HDL	CETP	16	0.28	2054	0.178	0.032	5.60	2.40E-08			
rs11869286	HDL	STARD3	17	0.35	2054	0.013	0.030	0.42	0.68			
rs4129767	HDL	PGS1	17	0.48	2054	0.068	0.029	2.37	0.02			
rs4148008	HDL	ABCA8	17	0.32	2054	-0.055	0.031	-1.76	0.08			
rs12967135	HDL	MC4R	18	0.24	2054	-0.010	0.033	-0.30	0.76			
rs7239867	HDL	LIPG	18	0.17	2054	-0.077	0.038	-2.00	0.05			
rs17695224	HDL	HAS1	19	0.29	2054	0.038	0.032	1.20	0.23			
rs386000	HDL	LILRA3	19	0.20	2054	0.061	0.036	1.68	0.09			
rs7254882	HDL	ANGPTL4	19	0.48	2054	0.011	0.029	0.37	0.71			
rs737337	HDL	ANGPTL8	19	0.09	2054	-0.043	0.051	-0.85	0.40			
rs1800961	HDL	HNF4A	20	0.03	2054	-0.060	0.085	-0.71	0.48			
rs6065906	HDL	PLTP	20	0.19	2054	0.027	0.036	0.73	0.47			
rs181362	HDL	UBE2L3	22	0.20	2054	-0.036	0.036	-1.02	0.31			
rs2479409	LDL	PCSK9	1	0.35	2054	-0.006	0.032	-0.18	0.85			
rs267733	LDL	ANXA9-CERS2	1	0.15	2054	-0.052	0.046	-1.13	0.26			
rs629301	LDL	SORT1	1	0.20	2054	-0.153	0.039	-3.95	8.05E-05			
rs10490626	LDL	INSIG2	2	0.07	2054	-0.141	0.060	-2.36	0.02			
rs1250229	LDL	FN1	2	0.27	2054	0.006	0.035	0.18	0.85			
rs1367117	LDL	APOB	2	0.30	2054	0.035	0.034	1.02	0.31			
rs2030746	LDL	LOC84931	2	0.42	2054	0.033	0.032	1.03	0.30			
rs2710642	LDL	EHBP1	2	0.32	2054	-0.023	0.033	-0.70	0.48			
rs4299376	LDL	ABCG5/8	2	0.31	2054	0.082	0.034	2.44	0.01			
rs17404153	LDL	ACAD11	3	0.12	2054	-0.017	0.048	-0.35	0.73			
rs7640978	LDL	CMTM6	3	0.08	2054	0.011	0.056	0.20	0.84			
rs4530754	LDL	CSNK1G3	5	0.44	2054	-0.020	0.032	-0.64	0.52			
rs1564348	LDL	LPA	6	0.16	2054	-0.025	0.043	-0.59	0.55			
rs1800562	LDL	HFE	6	0.06	2054	0.024	0.063	0.38	0.70			
rs3757354	LDL	MYLIP	6	0.21	2054	-0.055	0.038	-1.44	0.15			
rs4722551	LDL	MIR148A	7	0.17	2054	-0.010	0.041	-0.23	0.81			
rs10102164	LDL	SOX17	8	0.20	2054	-0.001	0.039	-0.02	0.98			
rs7832643	LDL	PLEC1	8	0.41	2054	0.008	0.032	0.25	0.80	Y	rs11136341	0.81
rs9411489	LDL	ABO	9	0.20	2054	0.058	0.038	1.53	0.13			
rs11220462	LDL	ST3GAL4	11	0.13	2054	0.019	0.046	0.42	0.68			
rs4942486	LDL	BRCA2	13	0.48	2054	0.039	0.031	1.25	0.21			
rs8017377	LDL	NYNRIN	14	0.47	2054	-0.056	0.031	-1.78	0.08			

rs1801689	LDL	APOH-PRXCA	17	0.03	2054	0.040	0.090	0.45	0.66			
rs6504872	LDL	OSBPL7	17	0.49	2054	0.001	0.031	0.02	0.99			
rs4420638	LDL	APOE	19	0.18	2054	0.022	0.041	0.54	0.59			
rs6511720	LDL	LDLR	19	0.11	2054	-0.087	0.051	-1.72	0.09			
rs2223745	LDL	TOP1	20	0.48	2054	0.050	0.032	1.58	0.11			
rs2328223	LDL	SNX5	20	0.19	2054	-0.071	0.040	-1.79	0.07			
rs364585	LDL	SPTLC3	20	0.39	2054	-0.063	0.032	-2.00	0.05			
rs5763662	LDL	MTMR3	22	0.02	2054	0.065	0.114	0.57	0.57			
rs1077514	Total Chol	ASAP3	1	0.14	2054	0.031	0.044	0.71	0.48			
rs1556562	Total Chol	EVI5	1	0.21	2054	-0.001	0.038	-0.02	0.99	Y	rs7515577	1.00
rs2642442	Total Chol	MOSC1	1	0.31	2054	-0.071	0.033	-2.15	0.03			
rs558971	Total Chol	IRF2BP2	1	0.47	2054	-0.039	0.031	-1.29	0.20	Y	rs514230	0.96
rs11563251	Total Chol	UGT1A1	2	0.11	2054	0.027	0.050	0.54	0.59			
rs11694172	Total Chol	FAM117B	2	0.24	2054	0.006	0.036	0.18	0.86			
rs2287623	Total Chol	ABCB11	2	0.40	2054	0.010	0.031	0.31	0.76			
rs7570971	Total Chol	RAB3GAP1	2	0.37	2054	-0.020	0.030	-0.65	0.51			
rs11709504	Total Chol	RAF1	3	0.19	2054	0.062	0.041	1.52	0.13	Y	rs2290159	0.81
rs13315871	Total Chol	PXK	3	0.09	2054	0.038	0.054	0.71	0.47			
rs12916	Total Chol	HMGCR	5	0.41	2054	0.045	0.031	1.46	0.14			
rs6882076	Total Chol	TIMD4	5	0.36	2054	0.005	0.032	0.14	0.89			
rs2758886	Total Chol	KCNK17	6	0.30	2054	0.034	0.033	1.02	0.31			
rs2814982	Total Chol	C6orf106	6	0.11	2054	-0.017	0.049	-0.35	0.73			
rs3177928	Total Chol	HLA	6	0.15	2054	-0.038	0.043	-0.88	0.38			
rs9376090	Total Chol	HBS1L	6	0.25	2054	0.012	0.034	0.36	0.72			
rs12670798	Total Chol	DNAH11	7	0.24	2054	0.053	0.036	1.48	0.14			
rs1997243	Total Chol	GPR146	7	0.16	2054	0.050	0.042	1.19	0.23			
rs2072183	Total Chol	NPC1L1	7	0.23	2054	0.068	0.037	1.86	0.06			
rs4738684	Total Chol	CYP7A1	8	0.34	2054	-0.010	0.033	-0.30	0.77	Y	rs2081687	0.91
rs3780181	Total Chol	VDLDR	9	0.07	2054	-0.069	0.061	-1.13	0.26			
rs10904908	Total Chol	VIM-CUBN	10	0.43	2054	0.051	0.031	1.63	0.10			
rs2255141	Total Chol	GPAM	10	0.31	2054	-0.032	0.034	-0.94	0.35			
rs10128711	Total Chol	SPTY2D1	11	0.27	2054	-0.030	0.034	-0.87	0.38			
rs11603023	Total Chol	PHLDB1	11	0.44	2054	-0.025	0.030	-0.81	0.42			
rs7941030	Total Chol	UBASH3B	11	0.39	2054	-0.002	0.031	-0.06	0.95			
rs11065987	Total Chol	BRAP	12	0.45	2054	-0.082	0.031	-2.62	0.0090			
rs1169288	Total Chol	HNF1A	12	0.31	2054	0.090	0.033	2.69	0.0072			
rs4883201	Total Chol	PHC1-A2ML1	12	0.11	2054	-0.015	0.049	-0.31	0.75			
rs2000999	Total Chol	HPR	16	0.22	2054	0.030	0.037	0.81	0.42			
rs314253	Total Chol	DLG4	17	0.34	2054	-0.044	0.033	-1.32	0.19			
rs10401969	Total Chol	CILP2	19	0.06	2054	0.007	0.064	0.10	0.92			
rs492602	Total Chol	FLJ36070	19	0.49	2054	0.007	0.031	0.22	0.82			
rs2277862	Total Chol	ERGIC3	20	0.15	2054	0.013	0.043	0.30	0.76			
rs2902940	Total Chol	MAFB	20	0.30	2054	-0.031	0.034	-0.91	0.37			
rs138777	Total Chol	TOM1	22	0.34	2054	0.021	0.032	0.66	0.51			
rs4253772	Total Chol	PPARA	22	0.11	2054	0.138	0.050	2.76	0.0059			
rs2131925	TG	ANGPTL3	1	0.33	2054	-0.089	0.033	-2.68	0.0074			
rs1260326	TG	GCKR	2	0.43	2054	0.112	0.031	3.59	0.0003			
rs645040	TG	MSL2L1	3	0.22	2054	-0.028	0.038	-0.74	0.46			
rs442177	TG	KLHL8	4	0.40	2054	-0.050	0.031	-1.59	0.11			
rs6831256	TG	LRPAP1	4	0.44	2054	0.053	0.031	1.70	0.09			
rs9686661	TG	MAP3K1	5	0.21	2054	0.021	0.038	0.54	0.59			
rs719726	TG	RSPO3	6	0.43	2054	-0.005	0.031	-0.16	0.87			
rs998584	TG	VEGFA	6	0.50	2054	-0.021	0.031	-0.70	0.49			
rs13238203	TG	TYW1B	7	0.03	2054	-0.008	0.093	-0.08	0.93			
rs17145738	TG	MLXIPL	7	0.11	2054	-0.037	0.050	-0.74	0.46			
rs38855	TG	MET	7	0.48	2054	0.033	0.031	1.05	0.30			
rs11776767	TG	PINX1	8	0.38	2054	0.032	0.032	0.99	0.32			
rs12678919	TG	LPL	8	0.07	2054	-0.131	0.060	-2.20	0.03			
rs2954029	TG	TRIB1	8	0.44	2054	-0.113	0.031	-3.61	0.00032			
rs10761741	TG	JMJD1C	10	0.44	2054	0.045	0.031	1.44	0.15	Y	rs10761731	0.99
rs1832007	TG	AKR1C4	10	0.15	2054	-0.081	0.044	-1.83	0.07			
rs2068888	TG	CYP26A1	10	0.44	2054	-0.030	0.031	-0.96	0.34			
rs174546	TG	FADS1-2-3	11	0.34	2054	0.072	0.033	2.19	0.03			
rs11613352	TG	LRP1	12	0.22	2054	0.000	0.038	-0.01	0.99			
rs2412710	TG	CAPN3	15	0.02	2054	-0.012	0.104	-0.12	0.91			
rs2929275	TG	FRMD5	15	0.05	2054	0.056	0.067	0.84	0.40	Y	rs2929282	0.97
rs3198697	TG	PDXDC1	16	0.39	2054	-0.043	0.032	-1.36	0.17			
rs9930333	TG	FTO	16	0.44	2054	0.077	0.031	2.46	0.01			
rs8077889	TG	MPP3	17	0.22	2054	0.067	0.038	1.75	0.08			
rs7248104	TG	INSR	19	0.41	2054	-0.016	0.032	-0.51	0.61			
rs731839	TG	PEPD	19	0.34	2054	0.046	0.033	1.39	0.16			
rs5756931	TG	PLA2G6	22	0.39	2054	-0.026	0.031	-0.84	0.40			

rs394352	Lp(a)	SLC22A3	6	0.29	2054	-0.245	0.034	-7.22	7.30E-13
rs2504927	Lp(a)	SLC22A3	6	0.43	2054	-0.250	0.032	-7.90	4.59E-15
rs4252109	Lp(a)	PLG	6	0.29	2054	-0.290	0.034	-8.62	1.32E-17
rs539298	Lp(a)	SLC22A3	6	0.47	2054	-0.273	0.031	-8.84	1.99E-18
rs7769879	Lp(a)	SLC22A3	6	0.39	2054	0.351	0.032	11.11	7.03E-28
rs986666	Lp(a)	SLC22A3	6	0.20	2054	-0.155	0.039	-3.95	8.10E-05
rs2457561	Lp(a)	SLC22A3	6	0.19	2054	-0.154	0.041	-3.81	0.00015
rs3798221	Lp(a)	LPA	6	0.19	2054	-0.282	0.039	-7.18	1.00E-12

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