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Brief Communication

Genetic studies of the susceptibility of classical and wild-derived inbred mouse strains to monkeypox virus



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ABSTRACT

Previously, we screened 38 inbred mouse strains for susceptibility to monkeypox virus (MPXV) and focused on wild-derived CAST mice because of their extreme vulnerability. Here, we provide further analysis of inbred mouse strains. NZW/Lac and C58 mice exhibited more weight loss than other classical inbred strains but all survived intranasal challenges with 10^4 to 10^6 PFU of MPXV. Mice from three wild derived strains, in addition to CAST, exhibited severe weight loss and died or were euthanized. LD₅₀ values for CASA, MOLF and PERA were 100, 6800 and $> 10^5$ PFU, respectively. CASA was inbred independently from the same founders as CAST, whereas MOLF and PERA are genetically and geographically distinct. The MPXV susceptibility of the F1 progeny of CAST and either C57BL/6 or BALB/c indicated that resistance is dominant. Back-crossing the F1 progeny of C57BL/6 and CAST to CAST suggested more than one independent resistant locus.

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Introduction

Monkeypox virus (MPXV) is the most severe poxvirus infection of humans, excluding variola virus, and has been designated as a Select Agent by the United States government because of the potential to exploit MPXV for bioterrorism. MPXV primarily infects rodents in Africa but can be transmitted to other animals as well as humans. Human monkeypox clinically resembles smallpox except for lower mortality and fewer human-to-human transmissions (McCullum and Damon, 2014; Parker et al., 2007). A virulent strain of MPXV is prevalent in the rain forests of central Africa, particularly in the Democratic Republic of the Congo, whereas a milder strain is present in West Africa. The latter was imported to the United States with infected dormice, rope squirrels and giant pouched rats in 2003 and spread to closely housed North American prairie dogs and ultimately to humans, resulting in 47 laboratory confirmed and additional clinically diagnosed human cases (Hutson et al., 2007; Reynolds and Damon, 2012). The ability to infect prairie dogs and other wild rodents and the occurrence of sporadic human MPXV infections in countries neighboring the Democratic Republic of the Congo, contribute to concerns that monkeypox may be an emerging disease.

Several small animal models including the American black-tailed prairie dog, the thirteen-lined ground squirrel, and the African dormouse have been used for studies of MPXV pathogenicity,

antivirals and vaccines (Hutson and Damon, 2010; Parker and Buller, 2013). However, except for the African dormouse these animals are not readily raised in captivity and there are no commercial sources of the latter. Moreover, immunological reagents are not available for these rodents. Although the commonly used classical inbred mouse strains are relatively resistant to MPXV, a few wild-derived inbred strains are susceptible (Americo et al., 2010) and one of these, the CAST/Eij mouse, has been further studied (Americo et al., 2014; Earl et al., 2012). The susceptibility to MPXV varied by age and route and was greater by the intraperitoneal route (LD₅₀ = 14 PFU) compared to the intranasal route (LD₅₀ = 680 PFU) for 6-week old female mice (Americo et al., 2010). Scarification and footpad inoculation only caused local lesions. The low interferon γ response of CAST mice to infection with MPXV and the protection afforded by exogenous interferon γ may be clues to the nature of their susceptibility (Earl et al., 2012). Moreover, the sensitivity of CAST mice extends to other orthopoxviruses including vaccinia virus and cowpox virus (Americo et al., 2014). The primary purpose of the present study was to analyze the susceptibility to MPXV of mouse strains that showed less severe symptoms than CAST mice in the initial screen and to gain insight into the genetics of resistance by cross breeding sensitive and resistant strains.

Results

Resistance of classical inbred mouse strains to MPXV

We previously screened 38 mouse strains, of which 32 were classically inbred, from the Jackson Laboratory Phenome Project

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for sensitivity to an intranasal (i.n.) dose of 2×10^4 PFU of the virulent MPXV-Z79-CB2 virus (Americo et al., 2010). NZW/LacJ and C58/J exhibited an average maximum 14% weight loss, which was greater than any of the other classical inbred strains. In that screen, C57BL/6j mice lost 4% of their weight and BALB/cj mice lost no weight. The resistance of BALB/c mice was confirmed by the absence of mortality after infection with doses up to 10^7 PFU. We considered, however, that NZW/Lac and C58 mice might be more susceptible to MPXV at higher doses than the 2×10^4 PFU used in the screen. To further evaluate their susceptibility, NZW/Lac and C58 mice were infected with several doses of MPXV. The animals were monitored for signs of disease including hunched posture, ruffled fur, and lethargy for up to 18 days. Weight loss was recorded daily and is shown as percent of the pre-infection weight (Fig. 1A and B). Both strains infected with 10^6 PFU displayed signs of disease including maximal weight loss of 20–23%. Loss of weight was first observed between days 3–5 post-infection and continued until days 6–10, after which animals showed improved health, increased weight and recovery from disease. As reported in the original screen (Americo et al., 2010), both NZW/Lac and C58 mice (Fig. 1A and B) exhibited greater weight loss than C57BL/6 (Fig. 2A,B and C). With the 10^6 PFU dose, the difference in weight loss of NZW/Lac mice relative to C57BL/6 was highly significant ($p < 0.004$) each day from 8 onwards; for C58 mice the difference from C57BL/6 was significant each day from 4 through 12 ($p = 0.001–0.04$). With all three strains, weight loss was delayed

and less severe at 10^5 PFU than at the higher dose and at 10^4 PFU there was only minor weight loss and minimal disease. In summary, no deaths were observed in any of the classical inbred strains even with an inoculum of 10^6 PFU, indicating a high degree of resistance to MPXV.

Susceptibility of wild-derived strains of mice to MPXV

In our initial multi-strain screen (Americo et al., 2010), three wild-derived strains showed signs of morbidity and mortality at the input dose of 2×10^4 PFU. CAST, MOLF/Eij and PERA/Eij mice exhibited greater than 20% weight loss and 100%, 75%, and 40% died or were euthanized, respectively. The sensitivity of CAST mice was further investigated and an LD_{50} of 680 PFU was determined (Americo et al., 2010). To more closely analyze the susceptibility of the MOLF, and PERA mice, we infected them i.n. with doses of MPXV ranging from 10^3 to 10^6 PFU. We also challenged CASA/Rk mice, which were inbred independently from the same founder mice used to derive CAST mice, with 10^2 to 10^5 PFU of MPXV. The CASA mice lost substantial weight even at the lowest dose of 10^2 PFU (Fig. 1C), whereas MOLF (Fig. 1D) and PERA (Fig. 1E) mice lost substantial weight with 10^5 and 10^4 PFU but had only mild weight loss at 10^3 PFU. All CASA mice died or were euthanized after infection with 10^3 PFU or more (Fig. 1F) and a substantial number of MOLF mice succumbed at doses of 10^4 PFU (Fig. 1G) or

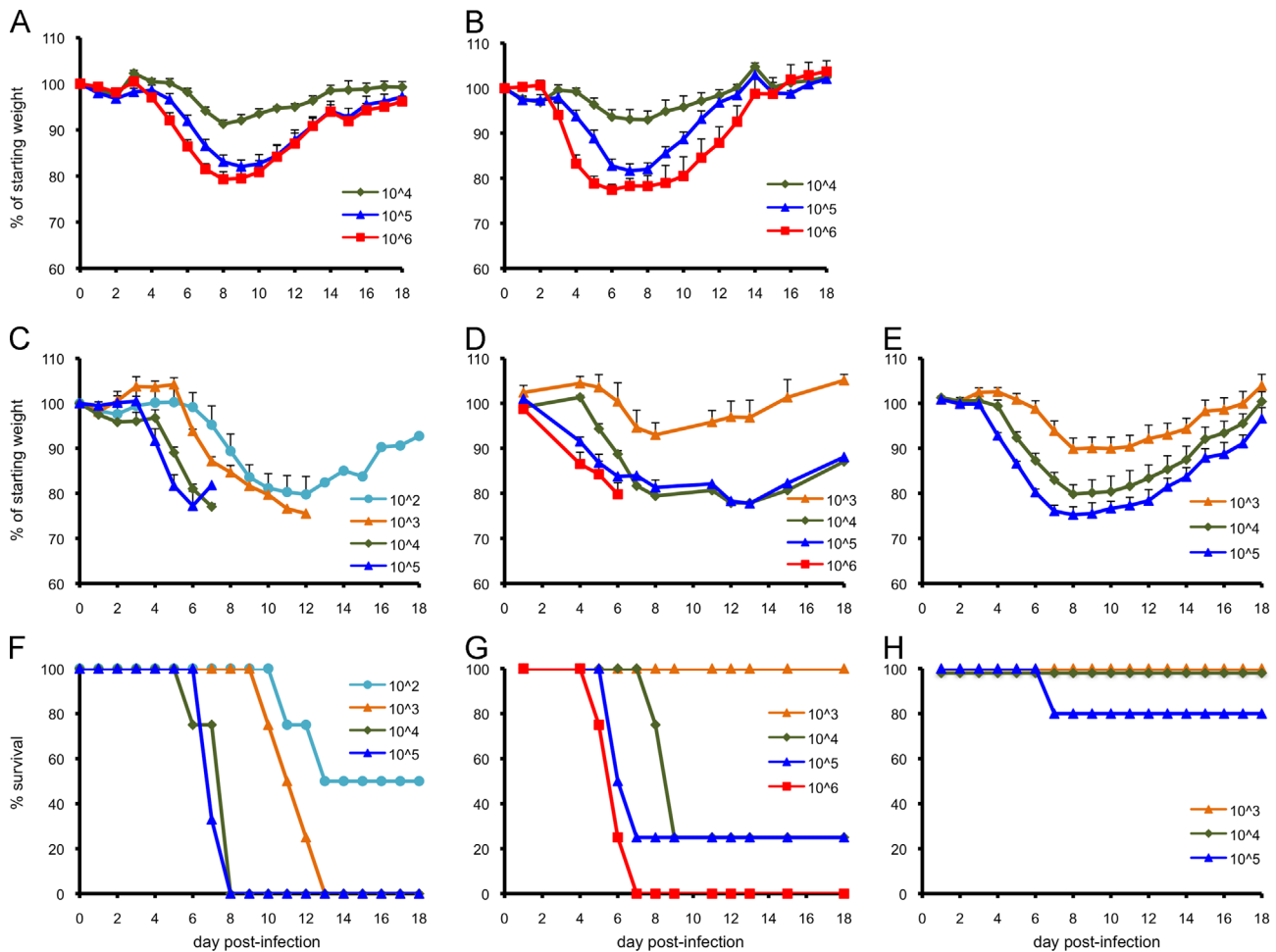


Fig. 1. Weight loss and survival of classical and wild derived inbred mouse strains infected i.n. with MPXV. Weight loss of groups ($n=5$) of female NZW/lac (A) and C58 (B) classical inbred mice infected with 10^4 – 10^6 PFU of MPXV are shown. Weight loss and survival of groups ($n=3–5$) of female CASA (C, F), MOLF (D, G) and PERA (E, H) wild-derived inbred mice infected with 10^2 – 10^6 PFU of MPX are shown. Doses of MPXV are indicated by color.

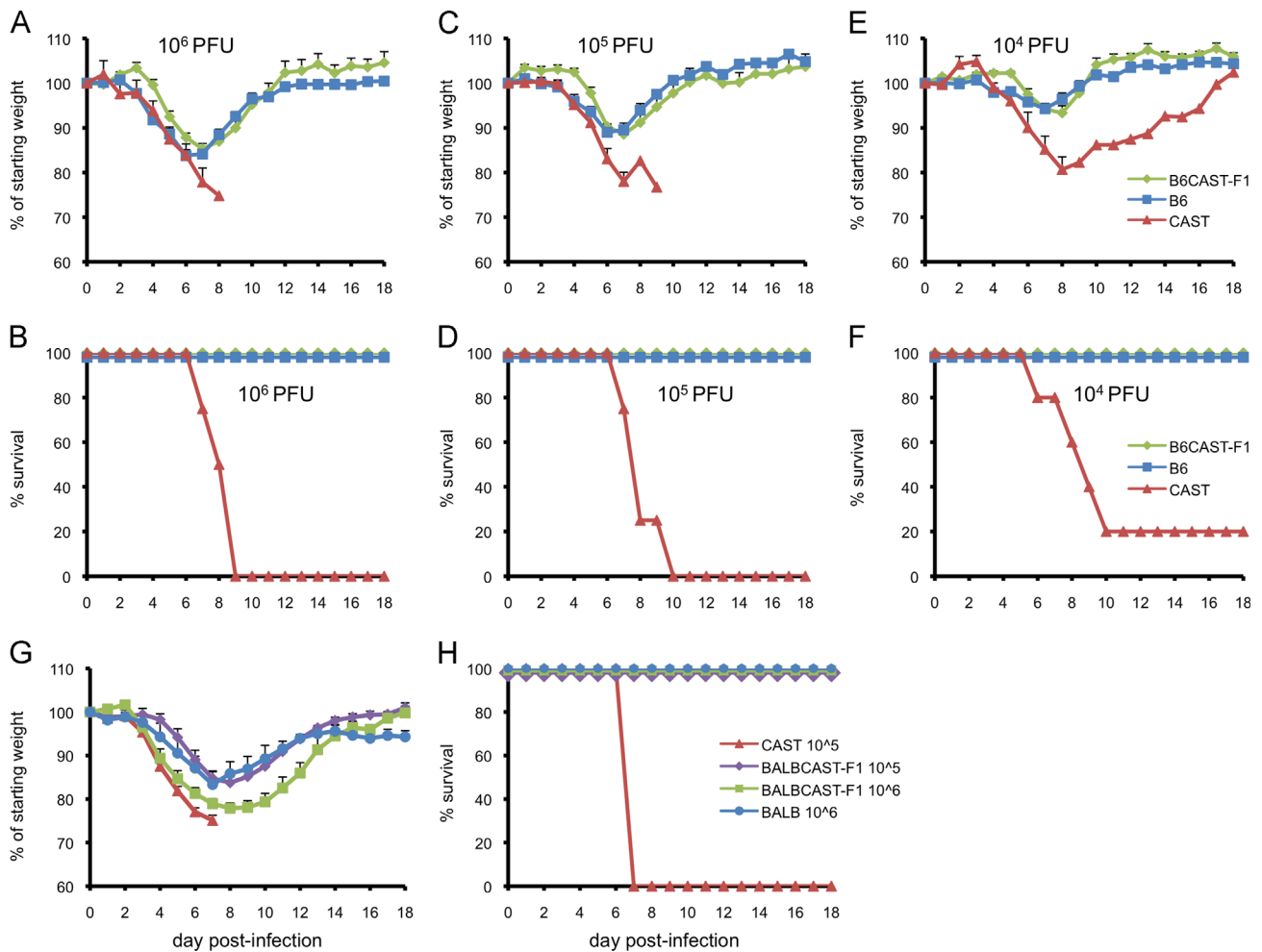


Fig. 2. MPXV infection of the F1 generation of CAST mice crossed with C57BL/6 or BALB/c mice. Percent of starting weight (A, C and E) and survival (B, D and F) of parental CAST and C57BL/6 and F1 progeny infected i.n. with MPXV. BALB/c: 10^6 ($n=5$ male, 5 female), 10^5 ($n=4$ male, 4 female), 10^4 PFU ($n=5$ male, 5 female). CAST: 10^6 PFU ($n=4$ male), 10^5 PFU ($n=4$ male), 10^4 PFU ($n=5$ male). F1: 10^6 PFU ($n=3$ male, 3 female); 10^5 PFU ($n=3$ male, 3 female); 10^4 PFU ($n=3$ male, 3 female). Percent of starting weight (G) and survival (H) of parental CAST and BALB/c and F1 progeny infected i.n. with MPXV. BALB/c: 10^6 PFU ($n=5$ female). CAST: 10^5 PFU ($n=4$ female). F1: 10^6 PFU ($n=2$ male, 4 female), 10^5 PFU ($n=2$ male, 3 female). Mouse strains are color-coded.

more, whereas only 1 PERA mouse succumbed at 10^5 PFU (Fig. 1H). Based on these data, the LD_{50} values for CASA, MOLF and PERA were 100, 6,800 and $>100,000$ PFU, respectively. The LD_{50} of CASA was slightly lower than that previously determined for the closely related CAST strain (Americo et al., 2010). However, we continue to use CAST mice because of their greater availability than CASA.

Evidence for dominance of resistance over sensitivity to MPXV

In order to determine whether resistance of C57BL/6 mice to MPXV is a dominant or recessive trait, C57BL/6 female mice were crossed with CAST mice to produce F1 progeny. Groups of F1 mice (3 male and 3 female) as well as parental C57BL/6 and CAST mice were infected i.n. with 10^4 , 10^5 , or 10^6 PFU of MPXV. As shown in Fig. 2A–F, weight loss and survival of the F1 mice were indistinguishable from that of the resistant C57BL/6 parent regardless of sex. In the same experiment, parental CAST mice suffered severe weight loss and succumbed by day 10 post-infection with survival of only one mouse at the lowest dose (Fig. 2A–F). We also compared the F1 progeny with parental C57BL/6 and CAST mice infected with 10^3 PFU by the intraperitoneal route. Again, the F1 progeny were resistant to MPXV compared to CAST mice (data not

shown). Thus, resistance to MPXV is a dominant characteristic of C57BL/6.

We also crossed female BALB/c mice with male CAST mice and infected the F1 progeny as well as the parental strains with 10^5 or 10^6 PFU of MPXV. The F1 progeny survived; whereas all CAST mice succumbed by day seven (Fig. 2G and H) indicating that resistance is dominant. However, at 10^6 PFU the F1 mice lost more weight than the parental BALB/c mice ($p=0.03$ to 0.004 from days 9 through 12) (Fig. 2G).

Evidence for multiple resistance loci to MPXV in C57BL/6 mice

To investigate whether dominance is due to more than a single genetic locus, the female F1 generation of parental C57BL/6 and CAST mice were backcrossed with male CAST mice. Nineteen progeny, 8 males and 11 females, were infected with 2×10^4 PFU of MPXV. Heterogeneity of weight loss was observed ranging from minimal C57BL/6-like to severe CAST-like for both male (Fig. 3A) and female (Fig. 3B) backcross mice. All 11 female backcross mice survived, whereas 3 of 8 male backcross mice (38%) did not (Fig. 3C). If resistance of C57BL/6 mice were determined by a single genetic locus, then 50% of the backcross progeny would be fully resistant and 50% fully sensitive. If resistance were determined by two independent loci, then 75% of the backcross progeny

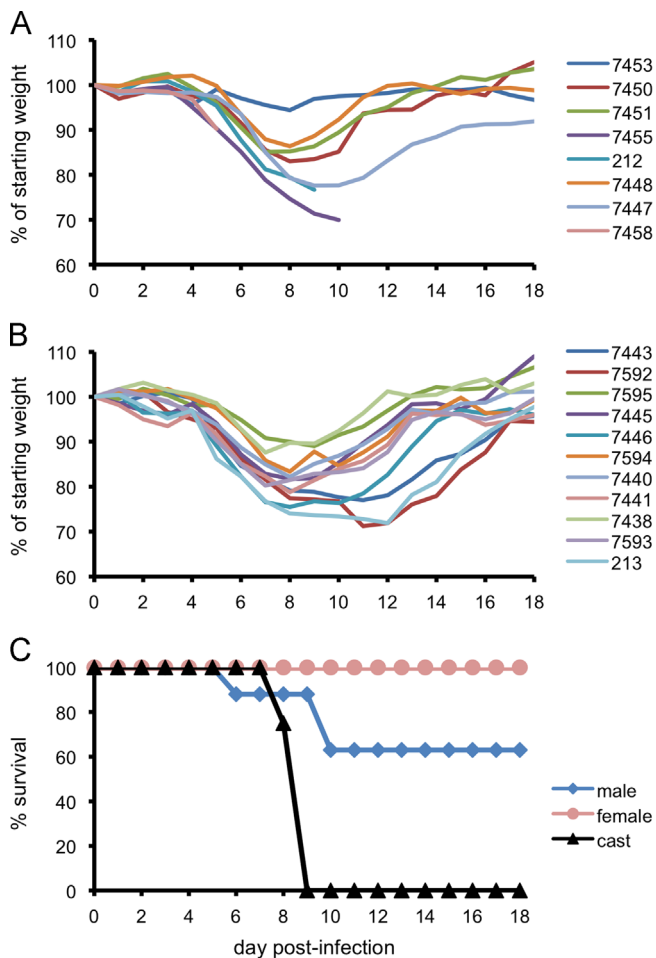


Fig. 3. MPXV infection of progeny from backcross of the F1 generation of parental CAST and C57BL/6 mice with CAST mice. Mice were infected i.n. with 2×10^4 PFU of MPXV. (A) Weight loss of male backcross progeny ($n=8$). (B) Weight loss of female backcross progeny ($n=11$). (C) Percent survival of mice. Mice strains are color coded according to key.

should be resistant and 25% sensitive. The survival of all female backcross mice strongly suggests multiple loci. A large number of backcross mice would be needed for single nucleotide polymorphism (SNP) analysis to identify resistant loci and evaluate the apparently greater sensitivity of male compared to female backcross mice.

Discussion

Common classical inbred mice have mosaic genomes derived predominantly from the Western European *Mus musculus domesticus* with additional sequences mainly from the Japanese *M. m. molossinus* and exhibit limited diversity (Takada et al., 2013). Our previous (Americo et al., 2010) and present data demonstrating the relative resistance to MPXV infection displayed by more than 30 classical inbred strains, likely represent conserved genetic sequences. The two most sensitive classical inbred strains of 32 tested are NZW/Lac and C58, although all survived doses up to 10^6 PFU. In contrast, we found that genetically diverse wild-derived strains exhibit a broad range of susceptibilities to MPXV. CAST and CASA are the most sensitive with LD_{50} of less than 10^3 , MOLF has intermediate sensitivity with a LD_{50} of less than 10^4 and PERA has a LD_{50} of greater than 10^5 . CAST and CASA are species of *M. m. castaneus* that were derived from a small population of

founder mice originally trapped in a grain storage facility in Thailand (JAX[®] NOTES Issue 456, Winter 1994). However, as CAST and CASA mice were inbred separately in different laboratories, the founder mice may also have been susceptible to MPXV. MOLF is an inbred species of *M. m. molossinus* that was derived from mice trapped in Japan and PERA is an inbred species of *M. m. domesticus* trapped in Peru. Not all wild-derived mice are highly susceptible to MPXV; however, since in our initial screen we found that SPRET/Eij and CZECHII/Eij lost no weight at all after infection with 2×10^4 PFU and PWK/Phj lost less than 8%.

Crossbreeding of CAST with C57BL/6 and with BALB/c was carried out to investigate whether resistance or sensitivity to MPXV was dominant. In both cases the F1 progeny were relatively resistant to MPXV. Based on the number of survivors, a backcross of F1 females derived from CAST X C57BL/6 with male CAST mice indicated the likely presence of more than one resistance locus. However, there was a sex difference: 3 of 8 male backcross mice succumbed whereas the 11 female backcross mice survived. A much larger number of backcross mice than the 19 used here would be necessary to confidently map resistant loci by SNP analysis. We also challenged the F1 generation between a cross of CAST and MOLF mice and found that they were more resistant to MPXV than the CAST parent (our unpublished data).

There is evidence from serial backcross experiments that multiple genes contribute to the resistance of C57BL/6 mice to ectromelia virus (Browstein et al., 1992). However, unlike MPXV, ectromelia is pathogenic in many classical inbred mouse strains. Increased severity of male compared to female mice has also been reported for infection of inbred mice with ectromelia virus (Browstein et al., 1992; Wallace et al., 1985). The Collaborative Cross panel (Threadgill and Churchill, 2012), which was derived by interbreeding eight different mouse strains, could provide an alternative method of mapping MPXV-resistance loci. Although the CAST mouse was included among the founders, the other seven strains in the panel all exhibit resistance to MPXV, possibly making it difficult to identify individual resistance genes.

For several reasons, we believe that the greater sensitivity of CAST mice compared to classical inbred strains is due to an inadequate immune response. Lung titers of MPXV in BALB/c mice following i.n. infection approach that of CAST mice but, in contrast to CAST mice, the virus is rapidly cleared. BALB/c mice make a more rapid and greater interferon γ response than CAST mice (Earl et al., 2014; Earl et al., 2012). In addition, interferon γ - and interferon γ receptor-knock-out C57BL/6 mice are less resistant to MPXV than parental C57BL/6 mice and exogenous interferon γ protects CAST mice against MPXV infection (Earl et al., 2012). The transcription factor STAT1 is involved in up regulating host response gene expression due to signaling by types I, II or III interferons and STAT1-deficient mice are even more susceptible to MPXV than interferon γ - and interferon γ receptor-deficient mice (Stabenow et al., 2010).

Materials and methods

Cells and Viruses

BS-C-1 cells were maintained at 37 °C and 5% CO₂ in modified Eagle minimal essential medium (EMEM; Quality Biologicals, Inc., Gaithersburg, MD) supplemented with 8% heat-inactivated fetal bovine serum, 2 mM L-glutamine, 10 U of penicillin/ml, and 10 μ g of streptomycin/ml. MPXV-Z79-CB2 (Americo, 2010), a clonal isolate derived from MPXV-Z79-005, was used in all experiments. Purified virus was prepared as described previously (Americo et al., 2010).

Mice

The following inbred mouse strains were obtained from Jackson Laboratories (Bar Harbor, ME): C57BL/6J, CAST/Eij, MOLF/Eij, C58/J, NZW/LacJ, CASA/RKJ, and F1 and back-cross progeny. BALB/c mice were obtained from Taconic Biotechnology, Germantown, NY). Mice were maintained in small, ventilated microisolator cages.

Inoculation of mice

Animal experiments were performed in an ABSL-3 facility with approval of the NIAID Animal Care and Use Committee and the Centers for Disease Control. On the day of infection, MPXV was thawed, sonicated, and diluted in phosphate buffered saline containing 0.05% bovine serum albumin. The titer of each dose was verified by plaque assay on BS-C-1 cells. Infections were performed by instillation of 10 μ l of virus into one nostril. All wild-derived mice and progeny of wild-derived mice were lightly anaesthetized with isoflurane prior to infection. Mock-infected animals were inoculated with an equivalent volume of diluent. Animals were observed and weighed daily for up to 18 days. Animals that lost 30% of their starting weight were humanely euthanized in accordance with NIAID Animal Care and Use Guidelines

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References

- Americo, J.L., Moss, B., Earl, P.L., 2010. Identification of wild-derived inbred mouse strains highly susceptible to monkeypox virus infection for use as small animal models. *J. Virol.* 84, 8172–8180.
- Americo, J.L., Sood, C.L., Cotter, C.A., Vogel, J.L., Kristie, T.M., Moss, B., Earl, P.L., 2014. Susceptibility of the wild-derived inbred CAST/Ei mouse to infection by orthopoxviruses analyzed by live bioluminescence imaging. *Virology* 449, 120–132.
- Browstein, D.G., Bhatt, P.N., Gras, L., Budris, T., 1992. Serial backcross analysis of genetic resistance to mousepox, using marker loci for rmp-2 and rmp-3. *J. Virol.* 66, 7073–7079.
- Earl, P.L., Americo, J.L., Cotter, C.A., Moss, B., 2014. Comparative live bioluminescence imaging of monkeypox virus dissemination in a wild-derived inbred mouse (*Mus musculus castaneus*) and outbred African dormouse (*Graphiurus kelleni*). *Virology* 475C, 150–158.
- Earl, P.L., Americo, J.L., Moss, B., 2012. Lethal monkeypox virus infection of CAST/Eij mice is associated with a deficient interferon-gamma response. *J. Virol.* 86, 9105–9112.
- Hutson, C.L., Damon, I.K., 2010. Monkeypox virus infections in small animal models for evaluation of anti-poxvirus agents. *Viruses* 2, 2763–2776.
- Hutson, C.L., Lee, K.N., Abel, J., Carroll, D.S., Montgomery, J.M., Olson, V.A., Li, Y., Davidson, W., Hughes, C., Dillon, M., Spurlock, P., Kazmierczak, J.J., Austin, C., Miser, L., Sorhage, F.E., Howell, J., Davis, J.P., Reynolds, M.G., Braden, Z., Karem, K.L., Damon, I.K., Regnery, R.L., 2007. Monkeypox zoonotic associations: insights from laboratory evaluation of animals associated with the multi-state US outbreak. *Am. J. Trop. Med. Hyg.* 76, 757–768.
- McCollum, A.M., Damon, I.K., 2014. Human monkeypox. *Clin. Infect. Dis.* 58, 260–267.
- Parker, S., Buller, R.M., 2013. A review of experimental and natural infections of animals with monkeypox virus between 1958 and 2012. *Futur. Virol.* 8, 129–157.
- Parker, S., Nuara, A., Buller, R.M.L., Schultz, D.A., 2007. Human monkeypox: an emerging zoonotic disease. *Futur. Microbiol.* 2, 17–34.
- Reynolds, M.G., Damon, I.K., 2012. Outbreaks of human monkeypox after cessation of smallpox vaccination. *Trends Microbiol.* 20, 80–87.
- Stabenow, J., Buller, R.M., Schriewer, J., West, C., Sagartz, J.E., Parker, S., 2010. A mouse model of lethal infection for evaluating prophylactics and therapeutics against monkeypox virus. *J. Virol.* 84, 3909–3920.
- Takada, T., Ebata, T., Noguchi, H., Keane, T.M., Adams, D.J., Narita, T., Shin, I.T., Fujisawa, H., Toyoda, A., Abe, K., Obata, Y., Sakaki, Y., Moriwaki, K., Fujiyama, A., Kohara, Y., Shiroishi, T., 2013. The ancestor of extant Japanese fancy mice contributed to the mosaic genomes of classical inbred strains. *Genome Res.* 23, 1329–1338.
- Threadgill, D.W., Churchill, G.A., 2012. Ten years of the collaborative cross. *Genetics* 190, 291–294.
- Wallace, G.D., Buller, R.M., Morse 3rd, H.C., 1985. Genetic determinants of resistance to ectromelia (mousepox) virus-induced mortality. *J. Virol.* 55, 890–891.