



# *Syzygium cumini* (L.) skeels: a prominent source of bioactive molecules against cardiometabolic diseases

Vinicyus Teles Chagas<sup>1,2</sup>, Lucas Martins França<sup>1,2</sup>, Sonia Malik<sup>2</sup> and Antonio Marcus de Andrade Paes<sup>1,2\*</sup>

<sup>1</sup> Laboratory of Experimental Physiology, Department of Physiological Sciences, Federal University of Maranhão, São Luís, Brazil, <sup>2</sup> Graduate Program in Health Sciences, Biological and Health Sciences Center, Federal University of Maranhão, São Luís, Brazil

Syzygium cumini (Myrtaceae) is a worldwide medicinal plant traditionally used in herbal medicines due to its vaunted properties against cardiometabolic disorders, which include: antihyperglycemic, hypolipemiant, antiinflammatory, cardioprotective, and antioxidant activities. These properties have been attributed to the presence of bioactive compounds such as phenols, flavonoids, and tannins in different parts of the plant, albeit the knowledge on their mechanisms of action is scarce. This mini-review highlights the cardiometabolic properties of *S. cumini* by correlating its already identified phytochemicals with their described mechanisms of action. Data herein compiled show that some compounds target multiple metabolic pathways; thereby, becoming potential pharmacological tools. Moreover, the lack of clinical trials on *S. cumini* usage makes it a fruitful field of interest for both scientific community and pharmaceutical industry.

Keywords: black plum, jamun, myrtaceae, phenolic compounds, metabolic syndrome, ethnopharmacology, medicinal plants, complementary and alternative medicine

# INTRODUCTION

Cardiometabolic syndrome is associated with multiple risk factors including insulin resistance, dyslipidemia, hypertension, and obesity (Alberti et al., 2009). According to World Health Organization, every year about 2.8 million people die worldwide due to overweight or obesity (Lim et al., 2012). Prevalence of diabetes appears with projections to affect about 439 million adults by 2030, whereas cardiovascular diseases account for 30% of deaths annually, including both developed and developing countries (Shaw et al., 2010). Because of their chronic degenerative nature, cardiometabolic-related disorders have long-lasting treatments, costly for both the patient and the health services, in addition to potentially harmful side effects caused by polytherapeutic regimens (Yanovski and Yanovski, 2014). In this context, herbal medicines have become the major source of bioactive molecules and emerged as potential therapeutic tools to fulfill a multiple-target strategy, especially because of their inherented large-scale structural diversity as compared with synthetic compounds (Prabhakar and Doble, 2008; Vasudeva et al., 2012).

Myrtaceae family comprises about 121 genera with 3800–5800 species of shrubs and trees distributed mainly in tropical and subtropical areas of the world (Stefanello et al., 2011). The genus *Syzygium*, a leading member of this family, embrace 1100 species with deserving attention to *Syzygium cumini* (L.) Skeels (syn.: *Eugenia jambolana, Syzygium jambolanum*), which has been

## **OPEN ACCESS**

#### Edited by:

Akio Inui, Kagoshima University, Japan

#### Reviewed by:

Subhalakshmi Ghosh, Jadavpur University, India Marie Amitani, Kagoshima University, Japan Masahiro Ohsawa, Nagoya City University, Japan

#### \*Correspondence:

Antonio Marcus de A. Paes marcuspaes@ufma.br

#### Specialty section:

This article was submitted to Ethnopharmacology, a section of the journal Frontiers in Pharmacology

Received: 10 August 2015 Accepted: 20 October 2015 Published: 03 November 2015

#### Citation:

Chagas VT, França LM, Malik S and Paes AMA (2015) Syzygium cumini (L.) skeels: a prominent source of bioactive molecules against cardiometabolic diseases. Front. Pharmacol. 6:259. doi: 10.3389/fphar.2015.00259

1

used in the treatment of numerous diseases, especially diabetes (Ayyanar and Subash-Babu, 2012). The use of *S. cumini* was introduced in western medicine in the mid-nineteenth century, when the first reports on the investigation of its antidiabetic properties were published (Helmstadter, 2008). *S. cumini* is a large tree native from Indian subcontinent, but widely cultivated in many countries in Asia, Africa, and South America (Srivastava and Chandra, 2013). It is popularly known as jamun in India, black plum in Europe, jambolan in Spanish-spoken countries and jambolão in Brazil (Corrêa, 1974).

S. cumini is known to possess wide range of medicinal properties, which have been attributed to the presence of bioactive compounds in different parts of the plant. The leaves are used in dermopathies, gastropathies, constipation, leucorrhea, and diabetes; fruits are used in the treatment of pharyngitis and splenic diseases; whereas barks are used as astringents, anthelmintic, and carminative. Furthermore, seeds are used as astringents, diuretic, and especially in the treatment of diabetes (Warrier, 1997; Helmstadter, 2008; Baliga et al., 2011). Pharmacological studies have expanded the biological activities of S. cumini, which include antihyperglycemic, antiinflammatory, antibacterial, cardioprotective, and antioxidant (Kumar et al., 2008a; Rekha et al., 2008; Sharma et al., 2008b, 2011; Mastan et al., 2009; Arun et al., 2011; Tanwar et al., 2011). Likewise, some remarkable and well-conducted literature surveys on pharmacological properties, phytochemical constituents as well as nutritive value of S. cumini have been published along the last decade (Helmstadter, 2008; Avyanar and Subash-Babu, 2012; Baliga et al., 2013; Srivastava and Chandra, 2013).

Notwithstanding those findings, however, there is scarcity of data on the relationship of its secondary metabolites with the described biological effects, especially on the mechanisms of action of these compounds. Therefore, this mini-review is aimed to specifically study the cardiometabolic properties of secondary metabolites already identified in this plant species, correlating them with their potential mechanisms of action. For this purpose, a detailed literature survey has been carried out using both preclinical and clinical studies as an attempt to identify the molecular mechanisms of action for the compounds identified in the various parts of *S. cumini*, even considering they may not have been necessarily isolated from this plant species, but had the same chemical identity of those found in it.

# PHYTOCHEMICAL CONSTITUENTS

Various secondary metabolites viz., flavonoids, phenolic acids, tannins, and terpenes have been reported in different parts of *S. cumini* (**Table 1**). For instance, leaves of this plant species contain high levels of flavonoids, especially quercetin, myricetin, myricitrin, kaempferol, and their glucoside derivatives, in addition to simple phenols like ellagic acid, ferulic acid, chlorogenic acid, and gallic acid (Mahmoud et al., 2001; Timbola et al., 2002; Ruan et al., 2008). The essential oil of leaves is prevalent in terpenes such as  $\alpha$ -pinene,  $\beta$ -pinene,  $\alpha$ -limonene,  $\alpha$ -cadinol, pinocarvone, pinocarveol (Shafi et al., 2002; Mohamed et al., 2013). The seeds are the most studied part of the plant,

being especially high in hexahydroxydiphenic (HDDP) acidderivated hydrolyzable tannins, terpenes like as  $\alpha$ -terpineol, eugenol, betulinic acid, and the same abovementioned phenolic acids (Bhatia and Bajaj, 1975; Ramya et al., 2012). Flowers have been found to show a very similar chemical composition to seeds, although pharmacological and chemical studies on this part are scarce (Baliga et al., 2011; Gordon et al., 2011). In addition, fruits also contain anthocynins, like as cyanidin, delphinidin, and petudinine, which give them a bright violet color (Ramya et al., 2012). Stem bark has essentially the same phenolic acids, flavonoids, and terpenes described already for other parts of *S. cumini* (Bhatia and Bajaj, 1975; Baliga et al., 2011).

# CARDIOMETABOLIC PROPERTIES AND POTENTIAL MECHANISMS OF ACTION

# **Antihyperglycemic Activity**

Use of S. cumini in the fight against diabetes has been studied by western medicine since more than 130 years (Helmstadter, 2008). In recent years, numerous preclinical studies have evaluated extracts of various parts, especially seeds, of this plant species for anti-hyperglycemic activity (Ravi et al., 2003; Schossler et al., 2004; Anandharajan et al., 2006; Sharma et al., 2008a,b; Ramya et al., 2012; Silva et al., 2012). Blood and urine glucose levels of streptozotocin-induced diabetic rats were decreased upon 30-days treatment with ethanolic extract of seed at doses of 100 mg/kg/day (Ravi et al., 2003). In addition to blood glucose lowering effect, flavonoid-rich extract of seed was also shown to recover peripheral glucose tolerance in streptozotocininduced diabetic rats (500 mg/kg/day, 21 days) (Sharma et al., 2008a) and mice (300 mg/kg/day, 15 days) (Sharma et al., 2008b). Those effects were ascribed to increased activity of peroxisome proliferator-activated receptors (PPAR) alpha and gamma, which was assessed in 3T3-L1 preadipocytes incubated for 24 h with increasing concentrations (1-100 mg/mL) of flavonoid-rich extract of seed. In this same study, seed extract was shown to possess insulinotropic activity, which may be involved in the abovementioned effects (Sharma et al., 2008a). Considering flavonoids already identified in S. cumini seed (Table 1), antihyperglycemic effect of rutin was attributed to inhibition of glucose metabolism enzymes hexokinase and glucose-6-phosphatase (Kamalakkannan and Prince, 2006). As highlighted in Figure 1, rutin (0.5-8 mM) also increased insulin secretion in streptozotocin-treated pancreatic islets of rats (Esmaeili et al., 2009). On the other hand, 20  $\mu$ M quercetin stimulated insulin secretion by activation of L-type calcium channels (Figure 1) in isolated rat beta cells (Bardy et al., 2013). Nevertheless, quercetin has also been described to increase insulin sensitivity by improving the production of the adipocyte-derived factors, like adiponectin and leptin (Wein et al., 2010).

S. cumini has also been described to promote other insulinrelated effects. Incubation of methanolic extract of leaf (1 ng/mL-10  $\mu$ g/mL) with cultured L6 myotubes for 0.5 and 24 h was found to increase mRNA expression of glucose transporter (GLUT)-4

Plant part	Metabolite class	Identified compounds	Reference
Leaf	Flavonoids	Catechin, kaempferol, myricetin, myricetin 3-O-β-D-glucuronopyranoside, myricetin-4'-methyl ether 3-O-α-rhamnopyranoside, myricetrin 4''-O-acetate, myricetrin 4''-O-acetyl-2-O-gallate, myricitrin, quercetin -3-O-α-rhamno_ pyranoside	Mahmoud et al., 2001
	Phenolic acids	Caffeic acid, chlorogenic acid, ellagic acid, ferulic acid, gallic acid	Mahmoud et al., 2001; Timbola et al., 2002; Ruan et al., 2008
	Tannins	Nilocetin	Mahmoud et al., 2001
	Terpenes	$\alpha$ -pinene, $\alpha$ -cadinol, pinocarvone, pinocarveol, $\alpha$ -terpeneol, myrtenol, eucarvone, muurolol, myrtenal, cineole, geranylacetone	Shafi et al., 2002; Mohamed et al., 2013
Seed	Flavonoids	Quercetin, rutin, 3,5,7,4-tetrahydroxy flavanone	Bhatia and Bajaj, 1975; Karthic et al., 2008
	Phenolic acids	Caffeic acid, ellagic acid, ferulic acid, gallic acid	Bhatia and Bajaj, 1975
	Tannins	Corilagin, 3,6-HHDP glucose, 4,6-HHDP glucose, 1-galloyl glucose, 3-galloyl glucose	Bhatia and Bajaj, 1975
	Terpenes	$\alpha\text{-terpineol},\beta\text{-pinene},\beta\text{-terpinene},betulinic acid, eugenol$	Williamson et al., 2002; Karthic et al., 2008
Fruit	Flavonoids	Myricetin, myricetin deoxyhexoside	Gordon et al., 2011
	Phenolic acids	Ellagic acid, gallic acid	Reynertson et al., 2008 Gordon et al., 2011
	Tannins	HHDP-galloyl glucose, trigalloylglucose	Gordon et al., 2011
	Terpenes	Citronellol, geraniol, hotrienol, nerol, $\beta$ -phenylethanol, phenylpropanal	Vernin et al., 1991
	Anthocyanins	Cyanidin, delfinidin, petudinin	Veigas et al., 2007
Flower	Flavonoids	Kaempferol, myricetin, dihydromyricetin, myricetin-3-L-arabinoside, isoquercetin, quercetin, quercetin-3-D-galactoside	Subramanian and Nair, 1972
	Phenolic acids	Ellagic acid	Baliga et al., 2011
	Terpenes	Eugenol, oleanolic acid	Ramya et al., 2012
Stem bark	Flavonoids	Myricetin, quercetin, kaempferol	Baliga et al., 2011
	Phenolic acids	3,3'-di-O-methyl ellagic acid, 3,3', 4-tri-O-methyl ellagic acid, gallic acid	Bhatia and Bajaj, 1975; Ramya et al., 2012
	Terpenes	β-siterol, friedelin, betulinic acid	Ramya et al., 2012

#### TABLE 1 | Phytochemical compounds identified in different parts of Syzygium cumini.

HHDP, hexahydroxydiphenic acid.

and phosphatidylinositol 3-kinase (PI3 kinase), both important mediators of insulin action in adipocytes and skeletal muscle (Anandharajan et al., 2006). Myricetin (1 mg/kg, 3× per day, 14 days), one of the most prevalent flavonoids in the leaf, has been shown to improve GLUT-4 expression in both adipose tissue and skeletal muscle as well (Figure 1; Liu et al., 2007). Flavonoidrich extract of S. cumini leaf (300 mg/kg/day, 15 days) reduced by 50% the expression of aldose reductase in renal tissue of diabetic rats (Sharma et al., 2008b), an effect previously described for phenolic compounds, such as myricetin, quercetin, kampferol, and ellagic acid (Haraguchi et al., 1998). The use of S. cumini (300 mg/kg/day, 15 days) elevated the concentrations of glycogen in liver and skeletal muscle, suggesting a stimulatory effect on glycogen synthase or glycogenolysis inhibition (Sharma et al., 2008b). These effects have been described for oleanolic acid (5-100 µM) (Ha et al., 2009) and caffeic acid (200 g/kg/day, 5 weeks) (Jung et al., 2006), compounds found in flowers and seeds, respectively. In vitro studies showed that betulinic acid and 3,5,7,4'-tetrahydroxy flavanone isolated from S. cumini seed inhibited pancreatic a-amylase, which may cause diminished intestinal absorption of carbohydrates (Karthic et al., 2008). Nevertheless, bark extract (1 g/kg/day, 30 days) was shown to regenerate cells from pancreatic duct (Schossler et al., 2004).

# **Antihyperlipidemic Activity**

Flavonoid-rich extract of S. cumini seed (300 mg/kg/day, 15 days) was described to reduce total cholesterol, LDL-cholesterol, and triacylglycerol as well as raise HDL-cholesterol levels (Sharma et al., 2008b). Similar results were found for aqueous extract of fruit at doses of 100 and 200 mg/kg (Rekha et al., 2008), for hydroalcoholic extract of seed kernel (100 mg/kg/day, 30 days) (Ravi et al., 2005) and for administration of Dihar (100 mg/kg/day, 6 weeks), an Indian mixture of herbs containing S. cumini (Patel et al., 2009). In all the abovementioned studies, dyslipidemia secondary to streptozotocin-induced diabetes was used as animal model to assess S. cumini antihyperlipidemic activity, which have been mainly ascribed to the inhibition of 3-hydroxy-3-methyl-glutaryl-CoA (HMG-CoA) reductase (Figure 1), the enzyme responsible for cholesterol synthesis (Ravi et al., 2005; Sharma et al., 2008b, 2011). Flavonoids found in S. cumini (Table 1) are expected to account for this activity, since it has been described that this class of compounds increases the expression of cAMP-dependent phosphokinase, enzyme responsible for HMG-CoA reductase inhibition (Havsteen, 2002). However, such effects might also be due reduction of intestinal absorption of cholesterol, as well as, increased free fatty acid and triacylglycerol clearances subsequent to insulin



action improvement. (Ravi et al., 2005; Birari and Bhutani, 2007; Sharma et al., 2012). In fact, preliminary data from our group has pointed out that hydroethanolic extract of *S. cumini* leaf inhibits both activity and expression of hepatic microsomal triglyceride-transfer protein, which is further controlled by insulin signaling pathways. Nevertheless, quercetin has been recently shown to prevent OP9 mouse stromal cells differentiation into adipocytes

by downregulation of adipogenic genes (Seo et al., 2015).

# **Cardioprotective Activity**

*S. cumini* has also been reported to promote hypotensive and antihypertensive effects. Chronic administration of hydroalcoholic extract of *S. cumini* leaf (100 and 250 mg/kg/day, 20 weeks) reduced blood pressure in normotensive rats, an effect further corroborated by decreased reactivity of vascular smooth muscle observed upon incubation of its chloroform (0.25 and 0.5 mg/mL) and aqueous fractions (0.1, 0.25, and 0.5 mg/mL; Ribeiro, 2007). More recently, the same group showed a dose-dependent reduction of blood pressure and heart rate on spontaneously hypertensive rats submitted to 8-weeks treatment with the same extract (0.5 g/kg/day) (Ribeiro et al., 2014). In both studies, authors speculated the extract might contain some compound able to non-competitively blockade L-type calcium channel. As depicted in **Figure 1**, myricetin has been reported to promote potent vasodilation by impairment of calcium influx (Herrera et al., 1996), similarly to quercetin, whose vasospasmolytic effect has been ascribed to blockage of calcium influx through L-type calcium channels (Hou et al., 2014).

Besides its effects on vasomotricity, S. cumini seems to improve hemodynamics as well. In a recent work, platelets

collected from diabetic patients were incubated with aqueous extract of S. cumini leaf (100 and 200 µg/mL) resulting in lower platelet aggregation and decreased oxidative damage, as assessed by measurement of lipoperoxide and nitric oxide levels and superoxide dismutase activity. The extract was still found to increase platelet cell membrane fluidity and to stimulate Na<sup>+</sup>/K<sup>+</sup>-ATPase activity (Raffaelli et al., 2014). In earlier work, De Bona et al. (2010) had already reported that same concentrations of aqueous extract of S. cumini leaf decreased adenosine deaminase and 5'-nucleotidase activities, important thrombogenic enzymes, when incubated with platelets from diabetic patients. Phenolic compounds identified in the extract, such as gallic acid, chlorogenic acid, and rutin, were suggested as possible candidates for the abovementioned activities. Moreover, administration of methanolic extract of seed (200 mg/kg) previously to isoproterenol-induced infaction in rats, reduced serum levels of myocardial necrosis biomarkers, specifically aspartate aminotransferase, alanine aminotransferase, uric acid, creatine phosphokinase, and lactate dehydrogenase (Mastan et al., 2009). Notwithstanding, administration of  $\alpha$ -hydroxy cinnamic acid (10, 15, and 20 mg/kg/day, 30 days), isolated from aqueous extract of S. cumini fruit, to high fatfed diabetic rats promoted antiatherosclerotic effects mainly characterized by decreased oxidized LDL levels, modulation of endothelial nitric oxide synthase, lower expression of soluble vascular cell adhesion molecule-1 and significant reduction of atherogenic lipoprotein apolipoprotein B100 along with an increase of apolipoprotein A1 (Tanwar et al., 2011).

# **Antiinflammatory Activity**

Inflammatory processes are directly involved in the development of cardiometabolic diseases including atherosclerosis, type 2 diabetes, and cancer (Hotamisligil, 2006). Methanolic and ethyl acetate extracts of S. cumini leaf (200 and 400 mg/kg) reduced carrageenan-induced paw edema in rats (Jain et al., 2011). Methanolic extract of seed (250 and 500 mg/kg, 21 days) also reduced paw edema volume and leukocyte migration in rats with adjuvant-induced arthritis (Kumar et al., 2008b). Ethanolic extract of the bark (100, 300, and 1000 mg/kg) reduced the production of prostaglandin E2, serotonin, and histamine (Muruganandan et al., 2002). Similarly, pre-treatment of mice with ethanolic extract of leaf (25, 50, and 100 mg/kg) reduced systemic production of pro-inflammatory factors, such as interleukin-5 (Brito et al., 2007), whereas TNF-α and NO' production was decreased at doses of 5 or 50 mg/kg administered 6-h previous to inflammation induction (Maciel et al., 2008).

# **Antioxidant Activity**

Polyphenolic and related antioxidant compounds are recognized as important cardiometabolic agents since they scavenge reactive oxygen/nitrogen species and stimulate antioxidant defenses (Valko et al., 2007), which may be involved in all the aforementioned activities of *S. cumini* (Figure 1). Oral administration of aqueous extract of seed (500, 1000, and 1500 mg/kg/day, 5 days) to mice treated with urethane 7,12dimethyl benzanthracene resulted in reduced chromosomal damage, significantly inhibited hepatic lipid peroxidation, which was associated with significantly increased activity of glutathione S-transferase, superoxide dismutase, and catalase (Arun et al., 2011). Pre-treatment of cyclophosphamideadministered rats with methanolic extract of S. cumini fruit (100 and 200 mg/kg/day, 14 days) resulted in reduced formation of hepatic malondialdehyde, lower frequency of aberrant metaphases, reduced formation of micronuclei, decreased cytotoxicity against marrow cells and increased activity of antioxidant enzymes, like as superoxide dismutase and catalase, besides increased GSH levels (Tripathi et al., 2013).

Considering in vitro studies, methanolic extract of S. cumini leaf and branch (1-15 µg/ml) has been shown to strongly act against OH' and DPPH' radicals, and decreased Fe<sup>3+</sup> to Fe<sup>+2</sup> reduction in the FRAP assay. Such activities showed a strong correlation with the high content of polyphenols and flavonoids present in the extract (Eshwarappa et al., 2014). In another report, antioxidant activity of ethyl acetate fraction (15.6–125 µg/mL) prepared from methanolic extract of S. cumini leaf was also correlated with its polyphenolic composition, especially to ferulic acid and catechins present in the extract (Ruan et al., 2008). Aqil et al. (2012) reported that ethanolic extracts of fruit and seed, at different concentrations, showed strong antioxidant activity assessed by mean of distinct assays, such as ABTS<sup>+</sup>, DPPH<sup>+</sup>, FRAP, and ORAC. These extracts also showed antiproliferative activity on lung cancer cells, making them potential sources of anticarcinogenic agents. Anthocyanins (delphinidin, cyanidin, petunidin, peonidin, and malvidin), besides ellagic acid and its derivatives (ellagitannins) identified in both extracts were suggested to be responsible for the antioxidant and antiproliferative activities (Aqil et al., 2012). Antioxidant activity of S. cumini fruit has also been shown to be due to gallic acid, as well as, derivatives of tetragalloyl glucose and myricetin (Gordon et al., 2011).

# TOXICITY

Several studies have shown that S. cumini do not produce acute or chronic toxicity when given by oral route. General toxicological screenings including behavioral, histomorphological as well as blood hematological and biochemical parameters have been conducted for extracts from seed (Chaturvedi et al., 2007), fruit (Kumar et al., 2008a), and leaf (Silva et al., 2012) of S. cumini, with no toxic effect noticed. Silva et al. (2012) demonstrated that acute administration of hydroalcoholic extract of leaf at doses as high as 2 g/kg produced no toxic effects in rodents. The same study demonstrated that chronic treatment for up to 180 days at doses of 50, 100, and 250 mg/kg did not cause hematological, biochemical, or histological alterations in target organs (Silva et al., 2012). These results are further corroborated by findings of potential hepatoprotective effect of ethanolic extract of S. cumini fruit, without any harmful side effects in animals (Bilal et al., 2011).

# **CLINICAL STUDIES**

Notwithstanding all the accumulated knowledge on the pharmacological properties of S. cumini, very limited data has come from clinical trials (Helmstadter, 2008). A study conducted by Srivastava et al. (1983) showed that acute administration of capsuled seed powder (4-24 g) to patients with severe diabetes notably reduced blood glucose levels in both fasting and post-prandial condictions. Similarly, Kohli and Singh (1993) reinforced the effects of seed powder (12 g/day, 3 months) on type 2 diabetes mellitus patients by showing that besides a 30% reduction of serum glucose levels, treatment also improved other classical symptoms of diabetes, like polyphagia, polyuria, and polidisia. On the other hand, a single administration of decocted dried powdered leaves of S. cumini (2 g in 250 mL water) to young normoglycemic patients had no effect on serum glucose levels (Teixeira et al., 2000). In a second approach, Teixeira et al. (2006) administered the same decoction to type 2 diabetic patients for 28 days observing no effect on glucose levels. Albeit contradictory results showed above, the plethora of preclinical studies on the cardiometabolic properties of S. cumini supports the necessity of well-designed trials that allow an efficient assessment of its therapeutic potentials in humans. In addition, standardization of extraction method and characterization of phytochemicals present in the extracts are fundamental to the success of such trials.

# REFERENCES

- Alberti, K. G., Eckel, R. H., Grundy, S. M., Zimmet, P. Z., Cleeman, J. I., Donato, K. A., et al. (2009). Harmonizing the metabolic syndrome: a joint interim statement of the international diabetes federation task force on epidemiology and prevention; national heart, lung, and blood institute; american heart association; world heart federation; international atherosclerosis society; and international association for the study of obesity. *Circulation* 120, 1640–1645. doi: 10.1161/CIRCULATIONAHA.109.192644
- Anandharajan, R., Jaiganesh, S., Shankernarayanan, N., Viswakarma, R., and Balakrishnan, A. (2006). In vitro glucose uptake activity of *Aegles marmelos* and *Syzygium cumini* by activation of Glut-4, PI3 kinase and PPAR  $\gamma$  in L6 myotubes. *Phytomedicine* 13, 434–441. doi: 10.1016/j.phymed.2005. 03.008
- Aqil, F., Gupta, A., Munagala, R., Jeyabalan, J., Kausar, H., Sharma, R. J., et al. (2012). Antioxidant and antiproliferative activities of anthocyanin/ellagitanninenriched extracts from *Syzygium cumini* L. (Jamun, the Indian Blackberry). *Nutr. Cancer* 64, 428–438. doi: 10.1080/01635581.2012.657766
- Arun, R., Prakash, M. V., Abraham, S. K., and Premkumar, K. (2011). Role of *Syzygium cumini* seed extract in the chemoprevention of in vivo genomic damage and oxidative stress. *J. Ethnopharmacol.* 134, 329–333. doi: 10.1016/j.jep.2010.12.014
- Ayyanar, M., and Subash-Babu, P. (2012). Syzygium cumini (L.) Skeels: a review of its phytochemical constituents and traditional uses. Asian Pac. J. Trop. Biomed. 2, 240–246. doi: 10.1016/S2221-1691(12)60050-51
- Baliga, M. S., Bhat, H. P., Baliga, B. R. V., Wilson, R., and Palatty, P. L. (2011). Phytochemistry, traditional uses and pharmacology of *Eugenia jambolana* Lam.(black plum): a review. *Food Res. Int.* 44, 1776–1789. doi: 10.1016/j.foodres.2011.02.007
- Baliga, M. S., Fernandes, S., Thilakchand, K. R., D'souza, P., and Rao, S. (2013). Scientific validation of the antidiabetic effects of *Syzygium jambolanum* DC (black plum), a traditional medicinal plant of India. *J. Altern. Complement. Med.* 19, 191–197. doi: 10.1089/acm.2011.0752
- Bardy, G., Virsolvy, A., Quignard, J., Ravier, M., Bertrand, G., Dalle, S., et al. (2013). Quercetin induces insulin secretion by direct activation of L-type

## CONCLUSION

The present work enlightens cardiometabolic properties described for *S. cumini*, which have been attributed to a limited amount of phytochemicals, particularly flavonoids, phenolic acids, and tannins (**Table 1**). As summarized in **Figure 1**, some compounds like myricetin, quercetin, rutin, ellagic, and gallic acids seem to be able to act on distinct pathways of cardiometabolic disorders, thus emerging as potential multi-targeted drugs. Nevertheless, the knowledge on their precise mechanisms of action is scanty and still deserves in-depth scientific research, especially concerning their jointed action as a phytocomplex. Finally, though toxicological studies have shown the species safety, clinical trials are barely inexistent pointing out a golden Eldorado for pharmaceutical companies.

# ACKNOWLEDGMENTS

Authors are thankful to Foundation for the Support of Research, Scientific, and Technological Development of the State of Maranhão – FAPEMA, which has importantly funded their research on pharmacological properties of *S. cumini* through the grants #APP01128/10 and #APP00280/12. We are also thankful to Victor Cardoso for his support on the figure drawing.

calcium channels in pancreatic beta cells. Br. J. Pharmacol. 169, 1102–1113. doi: 10.1111/bph.12194

- Bhatia, I., and Bajaj, K. (1975). Chemical constituents of the seeds and bark of *Syzygium cumini*. *Planta Med.* 28, 346–352. doi: 10.1055/s-0028-1097868
- Bilal, R., Zakaria, M., Usman, A., and Zia, A. (2011). Comparison of simvastatin with *Eugenia jambolana* fruit pulp in their effects on Alanine Transferase, Aspartate Aminotransferase and Creatinine Phosphokinase levels of hyperlipidaemic rats. J. Pak. Med. Assoc. 61, 1190.
- Birari, R. B., and Bhutani, K. K. (2007). Pancreatic lipase inhibitors from natural sources: unexplored potential. *Drug Discov. Today* 12, 879–889. doi: 10.1016/j.drudis.2007.07.024
- Brito, F. A., Lima, L. A., Ramos, M. F., Nakamura, M. J., Cavalher-Machado, S. C., Siani, A. C., et al. (2007). Pharmacological study of anti-allergic activity of *Syzygium cumini* (L.) Skeels. *Braz. J. Med. Biol. Res.* 40, 105–115. doi: 10.1590/S0100-879X2007000100014
- Chaturvedi, A., Kumar, M. M., Bhawani, G., Chaturvedi, H., Kumar, M., and Goel, R. (2007). Effect of ethanolic extract of *Eugenia jambolana* seeds on gastric ulceration and secretion in rats. *Indian J. Physiol. Pharmacol.* 51, 131–140.
- Corrêa, M. P., and Penna, L. D. A. (1974). *Dicionário das Plantas Úteis do Brasil e das Exóticas Cultivadas*, Vol. 5. Rio de Janeiro: Instituto Brasiliero de Desenvolvimento Florestal, 687.
- De Bona, K. S., Bellé, L. P., Sari, M. H., Thomé, G., Schetinger, M. R., Morsch, V. M., et al. (2010). Syzygium cumini extract decrease adenosine deaminase, 5' nucleotidase activities and oxidative damage in platelets of diabetic patients. Cell Physiol. Biochem. 26, 729–738. doi: 10.1159/00032 2340
- Eshwarappa, R. S. B., Iyer, R. S., Subbaramaiah, S. R., Richard, S. A., and Dhananjaya, B. L. (2014). Antioxidant activity of *Syzygium cumini* leaf gall extracts. *BioImpacts* 4, 101–107. doi: 10.5681/bi.201 4.018
- Esmaeili, M. A., Zohari, F., and Sadeghi, H. (2009). Antioxidant and protective effects of major flavonoids from *Teucrium polium* on beta-cell destruction in a model of streptozotocin-induced diabetes. *Planta Med.* 75, 1418–1420. doi: 10.1055/s-0029-1185704

- Gordon, A., Jungfer, E., Da Silva, B. A., Maia, J. G., and Marx, F. (2011). Phenolic constituents and antioxidant capacity of four underutilized fruits from the Amazon region. J. Agric. Food Chem. 59, 7688–7699. doi: 10.1021/jf201039r
- Ha, D. T., Tuan, D. T., Thu, N. B., Nhiem, N. X., Ngoc, T. M., Yim, N., et al. (2009). Palbinone and triterpenes from moutan cortex (*Paeonia suffruticosa*, Paeoniaceae) stimulate glucose uptake and glycogen synthesis via activation of AMPK in insulin-resistant human HepG2 Cells. *Bioorg. Med. Chem. Lett.* 19, 5556–5559. doi: 10.1016/j.bmcl.2009.08.048
- Haraguchi, H., Kanada, M., Fukuda, A., Naruse, K., Okamura, N., and Yagi, A. (1998). An inhibitor of aldose reductase and sorbitol accumulation from Anthocepharus chinensis. *Planta Med.* 64, 68–69.
- Havsteen, B. H. (2002). The biochemistry and medical significance of the flavonoids. *Pharmacol. Ther.* 96, 67–202. doi: 10.1016/S0163-7258(02)00 298-X
- Helmstadter, A. (2008). *Syzygium cumini* (L.) SKEELS (Myrtaceae) against diabetes-125 years of research. *Pharmazie* 63, 91-101.
- Herrera, M. D., Zarzuelo, A., Jiménez, J., Marhuenda, E., and Duarte, J. (1996). Effects of flavonoids on rat aortic smooth muscle contractility: structureactivity relationships. *Gen. Pharmacol.* 27, 273–277. doi: 10.1016/0306-3623(95) 02010-1
- Hotamisligil, G. S. (2006). Inflammation and metabolic disorders. *Nature* 444, 860-867. doi: 10.1038/nature05485
- Hou, X., Liu, Y., Niu, L., Cui, L., and Zhang, M. (2014). Enhancement of voltagegated K+ channels and depression of voltage-gated Ca2+ channels are involved in quercetin-induced vasorelaxation in rat coronary artery. *Planta Med.* 80, 465–472. doi: 10.1055/s-0034-1368320
- Jain, A., Sharma, S., Goyal, M., Dubey, S., Jain, S., Sahu, J., et al. (2011). Antiinflammatory activity of *Syzygium cumini* leaves. *Int. J. Phytomed.* 2, 124–126. doi: 10.3109/13880209.2013.768675
- Jung, U. J., Lee, M.-K., Park, Y. B., Jeon, S.-M., and Choi, M.-S. (2006). Antihyperglycemic and antioxidant properties of caffeic acid in db/db mice. *J. Pharmacol. Exp. Ther.* 318, 476–483. doi: 10.1124/jpet.106.105163
- Kamalakkannan, N., and Prince, P. S. M. (2006). Antihyperglycaemic and antioxidant effect of rutin, a polyphenolic flavonoid, in streptozotocininduced diabetic wistar rats. *Basic Clin. Pharmacol. Toxicol.* 98, 97–103. doi: 10.1111/j.1742-7843.2006.pto\_241.x
- Karthic, K., Kirthiram, K. S., Sadasivam, S., and Thayumanavan, B. (2008). Identification of alpha amylase inhibitors from *Syzygium cumini* linn seeds. *Indian J. Exp. Biol.* 46, 677–680.
- Kohli, K., and Singh, R. (1993). A clinical trial of jambu (Eugenia jambolana) in non-insulin dependant diabetes mellitus. J. Res. Ayurveda Siddha 13, 89–97.
- Kumar, A., Ilavarasan, R., Jayachandran, T., Deecaraman, M., Aravindan, P., Padmanabhan, N., et al. (2008a). Anti-diabetic activity of Syzygium cumini and its isolated compound against streptozotocin-induced diabetic rats. J. Med. Plants Res. 2, 246–249.
- Kumar, E., Mastan, S., Reddy, K. R., Reddy, G. A., Raghunandan, N., and Chaitanya, G. (2008b). Anti-arthritic property of the methanolic extract of *Syzygium cumini* seeds. *Int. J. Integr. Biol.* 4, 55–61.
- Lim, S. S., Vos, T., Flaxman, A. D., Danaei, G., Shibuya, K., Adair-Rohani, H., et al. (2012). A comparative risk assessment of burden of disease and injury attributable to 67 risk factors and risk factor clusters in 21 regions, 1990-2010: a systematic analysis for the Global Burden of Disease Study 2010. *Lancet* 380, 2224–2260. doi: 10.1016/S0140-6736(12)61766-8
- Liu, I.-M., Tzeng, T.-F., Liou, S.-S., and Lan, T.-W. (2007). Myricetin, a naturally occurring flavonol, ameliorates insulin resistance induced by a high-fructose diet in rats. *Life Sci.* 81, 1479–1488. doi: 10.1016/j.lfs.2007.08.045
- Maciel, M. C., Farias, J. C., Maluf, M. J., Gomes, E. A., Pereira, P. V., Aragão-Filho, W. C., et al. (2008). Syzygium jambolanum treatment improves survival in lethal sepsis induced in mice. BMC Complement Altern. Med. 8:57. doi: 10.1186/1472-6882-8-57
- Mahmoud, Ii Marzouk, M. S., Moharram, F. A., El-Gindi, M. R., and Hassan, A. M. (2001). Acylated flavonol glycosides from *Eugenia jambolana* leaves. *Phytochemistry* 58, 1239–1244. doi: 10.1016/S0031-9422(01)00365-X
- Mastan, S., Chaitanya, G., Bhavya Latha, T., Srikanth, A., Sumalatha, G., and Eswar Kumar, K. (2009). Cardioprotective effect of methanolic extract of *Syzygium cumini* seeds on isoproterenol-induced myocardial infarction in rats. *Der Pharm. Lett.* 1, 143–149.

- Mohamed, A. A., Ali, S. I., and El-Baz, F. K. (2013). Antioxidant and antibacterial activities of crude extracts and essential oils of Syzygium cumini leaves. PLoS ONE 8:e60269. doi: 10.1371/journal.pone.0060269
- Muruganandan, S., Pant, S., Srinivasan, K., Chandra, S., Tandan, S. K., Lal, J., et al. (2002). Inhibitory role of Syzygium cumini on autacoid-induced inflammation in rats. *Indian J. Physiol. Pharmacol.* 46, 482–486.
- Patel, S. S., Shah, R. S., and Goyal, R. K. (2009). Antihyperglycemic, antihyperlipidemic and antioxidant effects of Dihar, a polyherbal ayurvedic formulation in streptozotocin induced diabetic rats. *Indian J. Exp. Biol.* 47, 564–570.
- Prabhakar, P. K., and Doble, M. (2008). A target based therapeutic approach towards diabetes mellitus using medicinal plants. *Curr. Diabetes Rev.* 4, 291–308. doi: 10.2174/157339908786241124
- Raffaelli, F., Borroni, F., Alidori, A., Tirabassi, G., Faloia, E., Rabini, R. A., et al. (2014). Effects of in vitro supplementation with *Syzygium cumini* (L.) on platelets from subjects affected by diabetes mellitus. *Platelets* 26, 720–725. doi: 10.3109/09537104.2014.980797
- Ramya, S., Neethirajan, K., and Jayakumararaj, R. (2012). Profile of bioactive compounds in Syzygium cumini-a review. J. Pharm. Res. 5, 4548–4553.
- Ravi, K., Rajasekaran, S., and Subramanian, S. (2003). Hypoglycemic effect of Eugenia jambolana seed kernels on streptozotocin-induced diabetes in rats. Pharm. Biol. 41, 598–603. doi: 10.1080/13880200390501929
- Ravi, K., Rajasekaran, S., and Subramanian, S. (2005). Antihyperlipidemic effect of Eugenia jambolana seed kernel on streptozotocin-induced diabetes in rats. Food Chem. Toxicol. 43, 1433–1439. doi: 10.1016/j.fct.2005.04.004
- Rekha, N., Balaji, R., and Deecaraman, M. (2008). Effect of aqueous extract of Syzygium cumini Pulp on antioxidant defense system in streptozotocin induced diabetic rats. Iran. J. Pharmacol. Ther. 7, 137–145.
- Reynertson, K. A., Yang, H., Jiang, B., Basile, M. J., and Kennelly, E. J. (2008). Quantitative analysis of antiradical phenolic constituents from fourteen edible Myrtaceae fruits. *Food Chem.* 109, 883–890. doi: 10.1016/j.foodchem.2008.01.021
- Ribeiro, R. M. (2007). Estudo da Atividade Hipotensora das Folhas de Syzygium Jambolanum DC (Jambolão). Mestrado Dissertação (Mestrado em Saúde e Ambiente), Universidade Federal do Maranhão, São Luís.
- Ribeiro, R. M., Pinheiro Neto, V. F., Ribeiro, K. S., Vieira, D. A., Abreu, I. C., Silva Sdo, N., et al. (2014). Antihypertensive effect of Syzygium cumini in spontaneously hypertensive rats. Evid Based Complement. Alternat. Med. 2014:605452. doi: 10.1155/2014/605452
- Ruan, Z. P., Zhang, L. L., and Lin, Y. M. (2008). Evaluation of the antioxidant activity of *Syzygium cumini* leaves. *Molecules* 13, 2545–2556. doi: 10.3390/molecules13102545
- Schossler, D. R. C., Mazzanti, C. M., Luz, S. C. A. D., Filappi, A., Prestes, D., Silveira, A. F. D., et al. (2004). *Syzygium cumini* and the regeneration of insulin positive cells from the pancreatic duct. *Braz. J. Vet. Res. Anim. Sci.* 41, 236–239. doi: 10.1590/S1413-95962004000400003
- Seo, Y. S., Kang, O. H., Kim, S. B., Mun, S. H., Kang, D. H., Yang, D. W., et al. (2015). Quercetin prevents adipogenesis by regulation of transcriptional factors and lipases in OP9 cells. *Int. J. Mol. Med.* 35, 1779–1785. doi: 10.3892/ijmm.2015.2185
- Shafi, P. M., Rosamma, M. K., Jamil, K., and Reddy, P. S. (2002). Antibacterial activity of Syzygium cumini and Syzygium travancoricum leaf essential oils. *Fitoterapia* 73, 414–416. doi: 10.1016/S0367-326X(02)00131-4
- Sharma, A. K., Bharti, S., Kumar, R., Krishnamurthy, B., Bhatia, J., Kumari, S., et al. (2011). *Syzygium cumini* ameliorates insulin resistance and β-cell dysfunction via modulation of PPAR, dyslipidemia, oxidative stress, and TNFα in type 2 diabetic rats. *J. Pharmacol. Sci.* 119, 205–213. doi: 10.1254/jphs. 11184FP
- Sharma, A. K., Bharti, S., Kumar, R., Krishnamurthy, B., Bhatia, J., Kumari, S., et al. (2012). *Syzygium cumini* ameliorates insulin resistance and beta-cell dysfunction via modulation of PPAR, dyslipidemia, oxidative stress, and TNF-alpha in type 2 diabetic rats. *J. Pharmacol. Sci.* 119, 205–213. doi: 10.1254/jphs.11184FP
- Sharma, B., Balomajumder, C., and Roy, P. (2008a). Hypoglycemic and hypolipidemic effects of flavonoid rich extract from *Eugenia jambolana* seeds on streptozotocin induced diabetic rats. *Food Chem. Toxicol.* 46, 2376–2383. doi: 10.1016/j.fct.2008.03.020

- Sharma, B., Viswanath, G., Salunke, R., and Roy, P. (2008b). Effects of flavonoid-rich extract from seeds of *Eugenia jambolana* (L.) on carbohydrate and lipid metabolism in diabetic mice. *Food Chem.* 110, 697–705. doi: 10.1016/j.foodchem.2008.02.068
- Shaw, J. E., Sicree, R. A., and Zimmet, P. Z. (2010). Global estimates of the prevalence of diabetes for 2010 and 2030. *Diabetes Res. Clin. Pract.* 87, 4–14. doi: 10.1016/j.diabres.2009.10.007
- Silva, S. D. N., Abreu, I. C., Silva, G. F. C., Ribeiro, R. M., Lopes, A. D. S., Cartágenes, M. D. S. D. S., et al. (2012). The toxicity evaluation of *Syzygium cumini* leaves in rodents. *Rev. Bras. Farm.* 22, 102–108. doi: 10.1590/S0102-695X2011005000181
- Srivastava, S., and Chandra, D. (2013). Pharmacological potentials of *Syzygium cumini*: a review. J. Sci. Food Agric. 93, 2084–2093. doi: 10.1002/jsfa.6111
- Srivastava, Y., Bhatt, H., Gupta, O., and Gupta, P. (1983). Hypoglycemia induced by *Syzygium cumini* Linn. seeds in diabetes mellitus. *Asian Med. J.* 26, 489–492.
- Stefanello, M. E., Pascoal, A. C., and Salvador, M. J. (2011). Essential oils from neotropical Myrtaceae: chemical diversity and biological properties. *Chem. Biodivers.* 8, 73–94. doi: 10.1002/cbdv.201000098
- Subramanian, S. S., and Nair, A. G. R. (1972). Flavonoids of the flowers of Eugenia jambolana. Curr. Sci. 41, 703–704.
- Tanwar, R. S., Sharma, S. B., Singh, U. R., and Prabhu, K. M. (2011). Antiatherosclerotic potential of active principle isolated from *Eugenia jambolana* in streptozotocin-induced diabetic rats. *Evid. Based Complement. Alternat. Med.* 2011:127641. doi: 10.1155/2011/127641
- Teixeira, C., Fuchs, F., Weinert, L., and Esteves, J. (2006). The efficacy of folk medicines in the management of type 2 diabetes mellitus: results of a randomized controlled trial of *Syzygium cumini* (L.) Skeels. J. Clin. Pharm. Ther. 31, 1–5. doi: 10.1111/j.1365-2710.2006.00700.x
- Teixeira, C. C., Rava, C. A., Da Silva, P. M., Melchior, R., Argenta, R., Anselmi, F., et al. (2000). Absence of antihyperglycemic effect of jambolan in experimental and clinical models. *J. Ethnopharmacol.* 71, 343–347. doi: 10.1016/S0378-8741(00)00185-9
- Timbola, A. K., Szpoganicz, B., Branco, A., Monache, F. D., and Pizzolatti, M. G. (2002). A new flavonol from leaves of *Eugenia jambolana*. *Fitoterapia* 73, 174–176. doi: 10.1016/S0367-326X(02)00009-6
- Tripathi, P., Tripathi, R., Patel, R. K., and Pancholi, S. S. (2013). Investigation of antimutagenic potential of *Foeniculum vulgare* essential oil on

cyclophosphamide induced genotoxicity and oxidative stress in mice. Drug Chem. Toxicol. 36, 35–41. doi: 10.3109/01480545.2011.648328

- Valko, M., Leibfritz, D., Moncol, J., Cronin, M. T., Mazur, M., and Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* 39, 44–84. doi: 10.1016/j.biocel.2006. 07.001
- Vasudeva, N., Yadav, N., and Sharma, S. K. (2012). Natural products: a safest approach for obesity. *Chin. J. Integr. Med.* 18, 473–480. doi: 10.1007/s11655-012-1120-0
- Veigas, J. M., Narayan, M. S., Laxman, P. M., and Neelwarne, B. (2007). Chemical nature, stability and bioefficacies of anthocyanins from fruit peel of Syzygium cumini Skeels. Food Chem. 105, 619–627. doi: 10.1016/j.foodchem.2007.04.022
- Vernin, G., Vernin, G., Metzger, J., Roque, C., and Pieribattesti, J.-C. (1991). Volatile constituents of the jamrosa aroma *Syzygium jambos* L. aston from Reunion Island. *J. Essent. Oil Res.* 3, 83–97. doi: 10.1080/10412905.1991.9697916
- Warrier, P. K. (1997). Indian Medicinal Plants: A Compendium of 500 Species. Hyderabad: Orient Blackswan.
- Wein, S., Behm, N., Petersen, R. K., Kristiansen, K., and Wolffram, S. (2010). Quercetin enhances adiponectin secretion by a PPAR-gamma independent mechanism. *Eur. J. Pharm. Sci.* 41, 16–22. doi: 10.1016/j.ejps.2010.05.004
- Williamson, E. M., Foundation, D. R., and Limited, D. A. (2002). *Major Herbs of Ayurveda*. London: Churchill Livingstone.
- Yanovski, S. Z., and Yanovski, J. A. (2014). Long-term drug treatment for obesity: a systematic and clinical review. JAMA 311, 74–86. doi: 10.1001/jama.2013.281361

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Chagas, França, Malik and Paes. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.