



# The Therapeutic Potential and Limitations of Ketones in Traumatic Brain Injury

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Traumatic brain injury (TBI) represents a significant health crisis. To date, no FDA approved pharmacotherapies are available to prevent the neurological deficits caused by TBI. As an alternative to pharmacotherapy treatment of TBI, ketones could be used as a metabolically based therapeutic strategy. Ketones can help combat post-traumatic cerebral energy deficits while also reducing inflammation, oxidative stress, and neurodegeneration. Experimental models of TBI suggest that administering ketones to TBI patients may provide significant benefits to improve recovery. However, studies evaluating the effectiveness of ketones in human TBI are limited. Unanswered questions remain about age- and sex-dependent factors, the optimal timing and duration of ketone supplementation, and the optimal levels of circulating and cerebral ketones. Further research and improvements in metabolic monitoring technology are also needed to determine if ketone supplementation can improve TBI recovery outcomes in humans.

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# INTRODUCTION

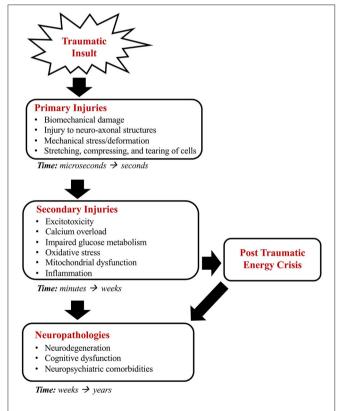
Traumatic brain injury (TBI) occurs when an insult to the head disrupts normal brain function. The Centers for Disease Control and Prevention reports that 2.87 million TBIs are diagnosed each year, with age adjusted TBI emergency room visits increasing 54% in recent years (1, 2). These numbers do not account for individuals who did not seek care, received outpatient or office-based care, or received care at federal facilities, such as those serving in the U.S. military or receiving care at a Veterans Affairs hospital (1). Because milder forms of TBI, including concussions, are often treated outside of emergency rooms, and because those who serve in the U.S. military are at significant risk for TBI, TBI incidence is underestimated (1).

TBIs are significant injuries that may result in long-term consequences, including neurodegeneration, cognitive dysfunction, and comorbid neuropsychiatric illnesses. It is estimated that 5.3 million individuals in the United States have a TBI-related disability, which places a sizable socioeconomic burden on patients, healthcare systems, and society (1, 3). Developing treatments that would help TBI patients recover faster and more completely would reduce healthcare costs, benefit society as a whole, and improve TBI patients' quality of life. However, TBIs are challenging to diagnose, treat, and study because TBIs present with a complex, multifaceted nature manifested through diverse clinical presentations, injury mechanisms, and factors specific to the individual (4). The path to developing better TBI treatments involves elucidating the underlying mechanisms of the brain's response to TBI in order to target specific pathophysiological responses.

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# PRIMARY AND SECONDARY MECHANISMS OF INJURY

The consequences of TBIs can be classified as either primary or secondary injuries. Primary injuries result directly from mechanical forces, including shearing and direct impact forces, at the moment of the traumatic event. Preventative measures, such as using protective equipment, can reduce the severity of the injury. However, brain tissue deformations from primary injuries are largely irreversible. Secondary injuries occur indirectly as a response to primary injuries and are characterized by interconnected cascades of biochemical, cellular, and molecular events that cause further damage but have the potential to be minimized and reversible (5, 6). Secondary injuries include an excessive release of excitatory neurotransmitters, calcium overload, glucose dysmetabolism, free radical overproduction, mitochondrial dysfunction, and neuroinflammation (7). These secondary injury responses deplete cerebral energy stores and can result in long-term neuropathological consequences. Figure 1 schematically illustrates the time-courses and relationship of primary and secondary injuries in TBI.



**FIGURE 1** | A schematic representation of injury processes following a TBI. The mechanical forces of the traumatic insult result in primary injuries characterized by deformations in brain tissue. Following the primary injury, secondary injuries ultimately result in cerebral energy deficiencies and long-term sequelae.

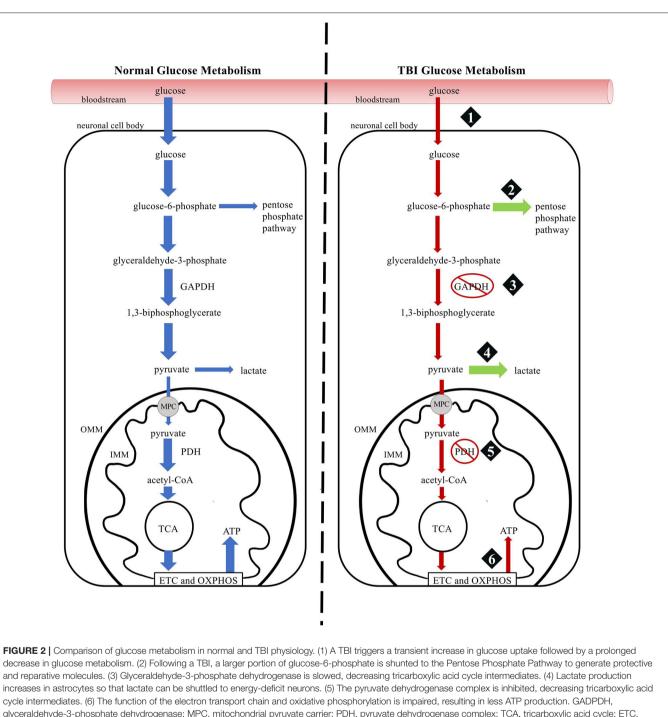
## **Cerebral Metabolism in Secondary Injuries**

The cascade of secondary injury mechanisms begins to occur immediately following the primary injury. The force of the primary injury triggers the release of glutamate in neurons and disrupts the integrity of the plasma membrane resulting in an efflux of potassium and influx of sodium and calcium (7, 8). Sustained glutamate release in damaged neurons activates ionotropic N-methyl-D-aspartate (NMDA) and α-amino-3-hydroxyl-5-methyl-4-isoxazole-propionate (AMPA) receptors (7, 9). The opening of these channels increases the ionic flux, further disrupting the ionic equilibrium and compounds the need for ion pumps to use adenosine triphosphate (ATP) to restore ionic and cellular homeostasis (8). The increased demand for ATP initially results in compensatory accelerated glucose metabolism, a state referred to as hyper-glycolysis (8). However, these neurochemical events quickly deplete cellular energy stores by creating a mismatch between energy supply and demand (8).

A hypometabolic stage begins around 6 hours post-injury with its duration being dependent upon injury severity (10). This stage is marked by glucose dysmetabolism and mitochondrial dysfunction. Failure of autoregulation mechanisms that maintain cerebral blood flow despite changes in cerebral perfusion pressure [cerebral perfusion pressure (CPP) = mean arterial pressure (MAP) - intracranial pressure (ICP)] results in decreased cerebral blood flow (11). Decreased cerebral blood flow reduces oxygen and glucose delivery to the brain, limiting glucose metabolism (7). Mitochondrial dysfunction results from excessive intracellular calcium accumulation (7). Excessive intracellular calcium is sequestered into the mitochondria where its accumulation opens the mitochondrial permeability transition pore (MPTP) and inhibits electron transport chain complexes, leading to a cerebral energy crisis and the initiation of apoptotic and necrotic cell death (12-14).

The impairment of oxidative phosphorylation leads to the production of reactive oxygen and nitrogen species that exceed the cell's antioxidant defenses (7, 15). As reactive oxygen and nitrogen species damage intracellular structures, repair enzymes are activated that use and deplete intracellular nicotinamide adenine dinucleotide (NAD+) stores (16). The decrease in NAD+ slows the ability of glyceraldehyde-3-phosphate dehydrogenase to catalyze glyceraldehyde 3-phosphate to 1,3-biphosphoglycerate in glycolysis, which disrupts glucose metabolism (16, 17). The ability of glucose to generate ATP is further disrupted as glucose is shunted to the pentose phosphate pathway at an increased rate in response to oxidative damage, as more pyruvate is converted to lactate in astrocytes to shuttle lactate to energy-deficit neurons, and as the pyruvate dehydrogenase complex is inhibited in response to oxidative stress (18-20). Figure 2 provides a schematic comparison of normal glucose metabolism and TBI glucose metabolism.

The observed impairments at numerous points in the glycolytic pathway and oxidative metabolism of glucose suggest that a more optimal substrate could be used as fuel after a traumatic brain injury. Considering that secondary injury processes are interconnected and have downstream consequences, interventions targeting specific secondary injury mechanisms could

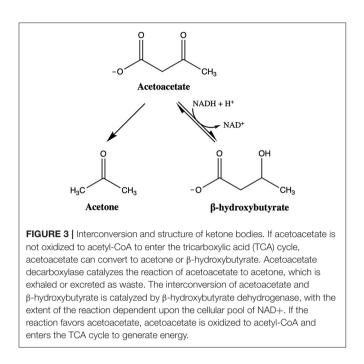


cycle intermediates. (6) The function of the electron transport chain and oxidative phosphorylation is impaired, resulting in less ATP production. GADPDH, glyceraldehyde-3-phosphate dehydrogenase; MPC, mitochondrial pyruvate carrier; PDH, pyruvate dehydrogenase complex; TCA, tricarboxylic acid cycle; ETC, electron transport chain; OXPHOS, oxidative phosphorylation; ATP, adenosine triphosphate; OMM, outer mitochondrial membrane; IMM, inner mitochondrial membrane.

improve TBI treatment outcomes. Because the reduction of cerebral energy has significant adverse consequences and is a strong, independent predictor of poor outcomes associated with neuropathology, cognitive deficits, and functional decline, establishing treatment methods to target cerebral energy deficits could help improve TBI recovery (21–23).

# **KETONES**

Ketone bodies are an alternative cerebral energy substrate that can target secondary injury mechanisms at various levels. Endogenous ketone bodies are synthesized in the body *via* lipolysis of free fatty acids in hepatic mitochondria and locally within



astrocytes (24, 25). There are three ketone bodies: acetoacetate,  $\beta$ -hydroxybutyrate, and acetone. **Figure 3** depicts the interconversion and structure of ketone bodies. When in the metabolic state of ketosis,  $\beta$ -hydroxybutyrate constitutes the majority of ketone bodies produced and is a reliable indicator of when the body is in ketosis.

Under normal dietary conditions with available carbohydrate stores, the production of ketone bodies is low. However, if the body has low carbohydrate stores, the body enters into a state of ketosis, where the production of ketone bodies is upregulated (26). Because carbohydrate metabolism is disrupted in TBIs, if the body can switch to ketosis, ketone bodies can function as an alternative energy source to glucose.

In terms of clinical use, the benefits of endogenously produced ketone bodies are limited due to the timeframe of inducing ketosis through a ketogenic diet containing low carbohydrates and high fats in the form of medium chain triglycerides. Endogenous ketone body production through a ketogenic diet and/or fasting takes 3–5 days to reach therapeutic blood ketone body levels (27). However, exogenous ketone supplementation can achieve therapeutic levels of blood ketone levels within 30 minutes of administration (28, 29). Exogenous ketones exist as liquids or powders that contain ketone bodies or ketone body precursors, including medium chain triglycerides, ketone mineral salts, and  $\beta$ -hydroxybutyrate esterified to precursor molecule. These compounds are clinically useful as they increase circulating levels of  $\beta$ -hydroxybutyrate without significantly lowering levels of insulin or glucose (27).

Medium chain triglycerides are comprised of saturated fatty acids containing 6–10 carbon atoms that are rapidly absorbed and converted to ketone bodies in the liver (30). A 30 g dose of medium chain triglycerides can raise circulating levels of  $\beta$ -hydroxybutyrate to 0.5 mM (31, 32). However, the dosage is limited due to poor gastrointestinal tolerability.

Ketone salts typically comprise  $\beta$ -hydroxybutyrate bound to a salt such as sodium, potassium, or calcium and can be administered orally or intravenously. They are widely available as a supplement or prescription. Oral administration of ketone salts can raise circulating levels of  $\beta$ -hydroxybutyrate to 1.0 mM, but limitations exist with gastrointestinal tolerability and the high mineral load (33–35). Intravenous ketone salts can achieve higher and more sustained levels of circulating  $\beta$ -hydroxybutyrate levels (up to 3.5 mM) than orally delivered ketone salts (36). However, intravenous ketone salts are still limited by concerns about the associated mineral load (35, 37).

Ketone esters are another exogenous ketone substrate and are comprised of  $\beta$ -hydroxybutyrate esterified to precursor molecule. The ketone ester most prominent in published literature is in the form of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate (38). It is highly tolerable and considered to be generally recognized as safe (GRAS) as a food ingredient by the FDA (38). After consumption, (R)-3-hydroxybutyl (R)-3-hydroxybutyrate is hydrolyzed to D-β-hydroxybutyrate and (R)-1,3-butanediol which is metabolized to D- $\beta$ -hydroxybutyrate and acetoacetate in the liver (39). Oral ingestion can raise circulating  $\beta$ -hydroxybutyrate levels up to 5 mM within 30 minutes (39, 40). The most significant limitation of ketone esters is that they are expensive to produce and not available on a prescription basis. Currently, only two companies manufacture ketone esters, and they are primarily sold for sports performance. Other limitations of oral ketone ester ingestion include its rapid metabolism, necessitating multiple doses to maintain  $\beta$ -hydroxybutyrate levels, and its poor taste. These concerns are less likely to be problematic for patients receiving treatment in an intensive care setting (27). Another possible limitation of oral ketone ester administration is that ketone esters are only available for oral, nasogastric, and orogastric administration. Nasogastric and orogastric administration of ketone esters may not be possible in some patients as placement of the gastric tube may increase intracranial pressure (41). No studies to date have examined intravenous administration of ketone esters. A better understanding of how orally administered ketone salts and esters behave in the body is needed before they may be administered intravenously. See Table 1 for a summary of exogenous ketone compounds.

## Metabolic Efficiency of Ketones

Several mechanisms of ketone body metabolism may improve recovery under the pathological conditions of a TBI. First, the ketone body,  $\beta$ -hydroxybutyrate, can be metabolized into acetyl-CoA with four enzymes compared to the ten enzymes required for glucose to enter the tricarboxylic acid (TCA) cycle. Second,  $\beta$ -hydroxybutyrate bypasses the need to use the pyruvate dehydrogenase complex to enter the TCA cycle. In TBIs, the pyruvate dehydrogenase complex is inhibited (42). The pyruvate dehydrogenase complex may be a critical factor leading to long-term energy deficits as its inhibition hinders the production NADH and FADH<sub>2</sub> in the TCA cycle, which disrupts the ability of the electron transport chain to generate ATP (42). Inhibition of the pyruvate dehydrogenase complex also decreases brain oxygen utilization and increases the ratio of lactate to pyruvate, both of which are indicative of impaired mitochondrial ATP production TABLE 1 | A summary of exogenous ketone compounds.

Exogenous ketone compound	Method of administration	Model(s)	Typical adult dosage	Plasma BHB concentration	Limitations	References
Medium chain triglycerides (MCT)	Oral	Rodent Human	15–30 g of MCT	0.5 mM	Poor GI tolerability	(31, 32)
					Low levels of ketosis	
					Multiple doses required to maintain BHB level	
Ketone salt	Oral	Rodent	10–20 g of BHB	1 mM	Poor GI tolerability	(33–35)
		Human			Low levels of ketosis	
					Multiple doses required to maintain BHB level	
					Associated mineral load	
Ketone salt	Intravenous infusion	Pig Human	20 µmol/kg/min	Up to 3.5 mM	Associated mineral load	(36, 37)
Ketone ester	Oral	Rodent Human	573 mg /kg body mass	Up to 5 mM	Poor taste	(27, 39, 40)
					Multiple doses required to maintain BHB level	

and decreased cellular phosphorylation energy (20). Because the metabolism of  $\beta$ -hydroxybutyrate does not utilize the pyruvate dehydrogenase complex, ketones may be advantageous to help restore deficits in cerebral energy.

Ketone bodies also consume less NAD+ than glucose to produce ATP. Both glucose and  $\beta$ -hydroxybutyrate pathways produce two molecules of acetyl-CoA, which subsequently enter the TCA cycle to generate ATP. Although glucose generates a higher final yield of ATP and reduces four molecules of NAD+,  $\beta$ -hydroxybutyrate reduces only one molecule of NAD+ during acetyl-CoA synthesis (43). The decreased use of NAD+ by ketones to form acetyl-CoA can increase the amount of NAD+ available to help maintain mitochondrial function, increase activation of antioxidant and pro-survival pathways, and decrease PARP-1-dependent cell death (44). By administering exogenous ketones, the metabolic efficiency of ketones can help spare cytosolic NAD+ pools, thereby increasing cell survival.

## **Neuroprotective Effects of Ketones**

In addition to being an efficient energy substrate for the injured brain, ketone bodies also have various neuroprotective effects. In TBIs, oxidative stress has been linked to many aspects of secondary injury mechanisms including lipid peroxidation, protein oxidation, DNA damage, electron transport chain inhibition, ionic dysregulation, and protease activation (7, 45). Ketone bodies have been shown to reduce oxidative stress by inhibiting the production of reactive oxygen species, preventing lipid peroxidation and protein oxidation, and increasing levels of antioxidation proteins (46). Because ketone bodies reduce oxidative stress, less NAD+ is depleted through enzymatic repair activity and

more NAD+ is available for other cellular processes. The attenuation of oxidative stress by ketone bodies has a significant neuroprotective effect by improving cellular function and reducing energy deficits.

Ketone bodies have also been shown to regulate inflammatory responses. In TBIs, inflammatory responses can be both detrimental and beneficial. Inflammatory processes are crucial to recovery because these processes help clear debris and facilitate regeneration mechanisms. However, excessive inflammation can be harmful and contribute to the development of cerebral edema, a breakdown of the blood-brain barrier, and neurodegeneration (47, 48). In TBIs, the mechanistic target of rapamycin (mTOR) signaling pathway and NLRP3 (NOD-, LRR-, and pyrin domain-containing protein 3) inflammasome have been shown to be particularly relevant in respect to inflammation and neurodegeneration (49, 50). The mTOR signaling pathway regulates many aspects of cell growth, metabolism, proliferation and survival, and is upregulated in TBIs (51). Ketone bodies have been shown to reduce mTOR expression through increased AMPactivated protein kinase activity (52). Various selective mTOR inhibitors, including rapamycin, have been shown to decrease neuroinflammation, neuronal death, astrogliosis, and neurodegeneration following TBI (49, 53). Targeting mTOR expression through ketones may represent an effective strategy to reduce inflammation and neurodegeneration.

Another way ketone bodies can reduce inflammation and neurodegeneration is through the inhibition of the NLRP3 inflammasome (54). In TBIs, the activation of the NLRP3 inflammasome leads to the release of caspase-1, which mediates secretion of pro-inflammatory cytokines and triggers pyroptotic cell death (50, 55). Ketone bodies have been shown to inhibit NLRP3mediated inflammatory responses and could help regulate excessive inflammation in TBI (54, 56). To effectively manage TBI inflammation, interventions need to limit acute proinflammatory responses to the level needed for beneficial recovery mechanisms and prevent the development of chronic inflammation. Because ketones have the ability to regulate inflammatory responses, ketones may be an effective treatment strategy to control inflammation.

Ketone bodies may also improve global and regional cerebral blood flow (52, 57). In TBIs, reductions in cerebral blood flow are seen in the acute phases and persist longer in patients with persistent TBI symptoms (58, 59). The decrease in cerebral blood flow is simultaneously accompanied by the increased demand on glycolysis and persists throughout the prolonged decrease in glucose metabolism (8). This results in a supply and demand mismatch with glucose not being brought to the brain, which contributes to the post-traumatic energy crisis (8). Ketones have been shown to improve cerebral blood flow through reduced mTOR expression and increased nitric oxide synthase protein expression (52). Because a decrease in cerebral blood flow inhibits the brain's ability to deliver oxygen and metabolites for proper brain function and recovery, ketone mediated improvements in cerebral blood flow have the potential to help to alleviate the mismatch in metabolic supply and demand.

Ketone bodies can also aid in closing the mitochondrial permeability transition pore (MPTP). The MPTP is a protein complex embedded in the inner and outer mitochondrial membrane that is linked to apoptotic and necrotic cell death. Following a TBI, the MPTP opens as a result of calcium overload and decrease in the ratio of ATP/ADP (14, 60). It has been established that the opening of the MPTP plays a pivotal role in the secondary injury cascade by destroying the integrity of the inner mitochondrial membrane (61, 62). This allows ions, cofactors, and water to enter mitochondria, leading to swelling, the eventual rupture of the outer mitochondrial membrane, and necrotic and apoptotic cell death (63). The metabolism of ketone bodies can help address cell death in three ways. First, when the MPTP opens, ions enter the mitochondrial matrix decreasing the free energy ( $\Delta G$ ) release of ATP hydrolysis (14). Because ketones can increase the  $\Delta G$  of ATP hydrolysis, ketones can help address energy deficits and improve cellular function (14). Second, ketones can increase the threshold of calcium-induced MPTP opening by decreasing mitochondrial reactive oxygen species levels (64, 65). Third, ketones can minimize apoptotic cell death by reducing cytochrome c release (66). Given the importance of mitochondria in cellular function, targeting the MPTP with ketones can help restore mitochondrial function to limit cell death.

## LIMITATIONS AND UNKNOWNS

Although evidence suggests ketones may be beneficial in TBI treatment, several aspects of treatment need to be investigated before exogenous ketones can become a valid therapeutic approach. More research is needed to understand differences in ketone metabolism due to age and sex, the optimal timing of ketone administration, and how to translate results from experimental models to human patients. See **Table 2** for a summary of the current state of knowledge regarding ketone therapy and TBI treatment.

## Age

Before exogenous ketones can be used in TBI treatment, differences in ketone metabolism across different ages must be considered. Studies with pediatric and adult rats fed a ketogenic diet following a TBI indicate that the efficiency of ketone metabolism is age dependent (67-69). Pediatric rats show a faster and more sustained increase in plasma ketone levels compared to adult rats (68). Pediatric rats also show improvements in motor and cognitive function and decreases in lesion volume and number of degenerating neurons while adult rats show no significant improvement (67, 68). The mechanisms underlying the observed differences between pediatric and adult TBI patients remains to be determined (70). However, it is hypothesized such differences may be due to the differences in monocarboxylate transporter expression between pediatric and adult TBI patients (16). Monocarboxylate transporters are membrane carrier proteins that transport ketone bodies across the plasma membrane, and they are more abundant in pediatric rats than in adult rats following a TBI (69, 71). Less monocarboxylate transporter expression may explain why the increase in plasma ketone concentration is delayed in adults, and therefore possibly not able to address ongoing secondary injury processes as efficiently.

Considering the rapid pathological progression following TBI, alternative approaches, such as incorporating a period of fasting to increase monocarboxylate transporter expression, could help resolve issues of age-dependent effects (72). However, a more comprehensive understanding of the metabolic changes associated with both pediatric and adult TBIs should be considered in order to develop ketone based pharmacological interventions. The current understanding of pediatric pathophysiology is limited due to the complexities of incomplete brain maturation and differences in cerebral metabolic requirements throughout development (70). Furthermore, our understanding of adult TBI is still improving (70). A more complete understanding of both pediatric and adult TBI pathophysiology is necessary to maximize the neuroprotective benefits of ketones.

## Sex

Our current understanding of TBI pathophysiology is also limited because it is currently uncertain whether TBI pathophysiology differs between males and females. Historically, in TBI studies, fewer females have been recruited to clinical trials, and male rodents have been the predominant experimental injury model (73). Because brain anatomy, cellular pathways, and drug pharmacokinetics can be affected by sex, our understanding of sex differences in TBI pathophysiology and recovery is lacking. The underrepresentation of females in both animal and human trials limits successful pharmacological developments and clinical care.

Recent studies suggest that neuropathological, physiological, social, and behavioral outcomes in TBIs are sex dependent

Known effects	Summary				
Metabolically efficient	Ketone body metabolism mechanisms are energetically favorable under the pathological conditions of a TBI and help reduce cerebral energy deficits.				
Reduce oxidative stress	Ketone bodies reduce oxidative stress by inhibiting the production of reactive oxygen species, preventing lipid peroxidation and oxidation, and increasing levels of antioxidant proteins.				
Regulate inflammation	Ketone bodies help regulate inflammatory processes that contribute to neurodegeneration and cerebral edema through the mTOR signaling pathway and NLRP3 inflammasome.				
Improve blood flow	Ketone bodies improve cerebral blood flow through reduced mTOR expression and increased nitric oxide synthase expression.				
Limit cell death	Ketone bodies can limit necrotic and apoptotic cell death by closing the MPTP protein complex and restoring mitochondrial func				
Limitations and unknowns	Summary				
Age	Pediatric animal models show a faster and more sustained increase in plasma ketone levels compared to adults. Ketone bodies may not be able to address ongoing secondary injury processes in adults as effectively.				
Sex	Aspects of mitochondrial function in human and animal studies are sex-dependent under normal physiological conditions. It is unknown how sex-dependent differences in TBI-induced mitochondrial dysfunction affect pathological responses.				
	Baseline ketone body levels are higher and rise significantly more following the administration of exogenous ketones in female TBI anima models. Further research is needed to understand why ketone metabolism differs between sexes and how those differences impact TBI pathophysiology.				
Timing and duration of ketone supplementation	Improvements in metabolic function following exogenous ketone administration may be dependent upon whether the patient is in a hypermetabolic or hypometabolic state.				
	The effects of long-term ketosis in TBI recovery are unknown.				
Experimental models	Animal models of TBI feature a homogenous type of injury with variables such as age, sex, genetic background, and injury parameters being well controlled. This may not accurately replicate the heterogenic nature of human TBI.				
Ketone levels	The optimal level of cerebral ketones and the most effective method of ketone supplementation to achieve such levels are unknown.				
Metabolic monitoring	Minimally invasive and accurate ways to monitor ketone metabolism in both intensive care and outpatient settings are needed.				

TABLE 2 | A summary of the current state of knowledge of ketone therapy in TBI treatment, including known effects, limitations, and unknowns.

(73). In a review examining 156 human TBI studies and 43 animal TBI studies, human studies report worse overall outcomes in females, whereas animal studies report better overall outcomes in females (73). While this difference may be due in part to injury severity, sample size, and the type of experimental injury model, several possible mechanisms might contribute to sex differences in TBI outcomes (73). These mechanisms include differences in how males and females acquire TBIs, how the brain responds at a cellular and molecular level to injury, and patterns of comorbidities and extracerebral injuries (73). At the cellular and molecular level, genetics and sex hormones have been shown to influence inflammation, edema, oxidative stress, excitotoxicity, and mitochondrial function (74-77). Because mitochondrial dysfunction and the resulting energy disruptions are central to TBI pathophysiology and ketones can help address energy disruptions, understanding sex differences in mitochondria are crucial. Multiple aspects of mitochondrial function in both human and animal studies are different between males and females under normal physiological conditions (73). These differences may be heightened during times of stress or injury (73). However, sex differences in TBIinduced mitochondrial dysfunction have only been explored to a limited extent. No studies to date have examined TBI-induced mitochondrial dysfunction in females to the same degree as TBI-induced mitochondrial dysfunction in males (78). Further research is required to understand how mitochondrial function may affect functional outcomes across both sexes in both humans and animals.

In respect to TBI and ketones, only one study to date has examined TBI sex differences in combination with ketone therapy (78). In this study, researchers found that baseline plasma concentrations of  $\beta$ -hydroxybutyrate were significantly lower in females compared to males (78). However, plasma concentrations of  $\beta$ -hydroxybutyrate increased by over 500% in females compared to a 100% increase in males following administration of  $\beta$ -hydroxybutyrate (78). This suggests the differences in male and female metabolism could potentially have negative effects on the brain as it is not yet understood how the brain differentiates between neuroprotective and neurotoxic levels of circulating ketones (79). Further research is needed to understand why ketone metabolism differs between the sexes and how those differences impact TBI pathophysiology.

# Timing

The timing and duration of exogenous ketone administration could also have significant consequences on TBI outcomes and warrants further investigation. Following a TBI, changes in cerebral energy metabolism occurs in two stages. The first stage is an acute period of hypermetabolism associated with excitotoxicity and increased glucose utilization (78). The second stage is a prolonged period of hypometabolism associated with the inhibition of glycolysis and mitochondrial dysfunction (78). Only one study to date has examined how the metabolic state during the time of ketone administration affects ketone metabolism in human TBI (78). In this study, researchers found that, in males, administering exogenous  $\beta$ -hydroxybutyrate in the hypometabolic state improved all measures of metabolic function, including mitochondrial function, reactive oxygen species production, and tricarboxylic acid cycle intermediate production (78). In contrast, β-hydroxybutyrate produced deleterious effects in females when administered during either the hypometabolic or hypermetabolic state (78). These results suggest that the neuroprotective properties of  $\beta$ -hydroxybutyrate depend upon both sex and metabolic state. However, the results of this study may be limited because metabolic functions were only analyzed at a single time point following induction of TBI. Additional research using rodent models has shown that other aspects of TBI pathophysiology, such as cytoskeletal damage and inflammatory patterns, peak at different times in males and females (76, 80). Because other aspects of TBI pathophysiology vary in timing between sexes, it is likely that variations in metabolic function across time also occur. Thus, future studies will need to examine both males at females at different times following TBI.

While research has shown that ketone metabolism can be beneficial during the acute phase of TBI, more research is needed to understand the effects of long-term ketosis in TBI recovery (35, 81). In healthy populations, studies have shown that longterm ketosis has no significant influence on long-term potentiation, synaptic function, and plasticity (82, 83). However, no studies to date have examined the long-term effects of ketosis specifically in TBI. It remains unclear whether long-term ketosis in TBI patients causes adverse effects. Future research will need to examine the optimal duration of exogenous ketone supplementation and any potential long-term effects.

Moderate-to-severe TBIs present an additional challenge in studying the timing of ketone administration due to the presence of additional complications. Although the secondary injury mechanisms of excitotoxicity, ionic disturbances, mitochondrial dysfunction, inflammation, and hypoglycemia still occur in moderate and severe TBIs, these issues are not the most pressing concern in early treatment. Early treatment of moderate-to-severe TBI primarily focuses on managing neurocritical care issues, such as multiple organ dysfunction syndrome, paroxysmal sympathetic hyperactivity, hemorrhagic progression, and coagulopathy (84). The administration of ketones in moderate-to-severe TBI with acute complications may not be beneficial and is largely unstudied in human TBI and animal models.

One barrier to studying the long-term effects of ketosis is that ketogenic diets are somewhat controversial, and high-quality clinical trials are lacking. Critics of the ketogenic diet, especially when it is employed primarily to lose weight, argue that the diet is a fad because conventional wisdom holds that high-fat diets are harmful, consumption of fiber and whole grains are essential, and ketogenic diets depend upon the consumption of animal products (85). Additionally, current research on ketogenic diets presents methodological challenges, such as attention to food quality, missing key micronutrients, and physical activity, that require studies to be interpreted with caution (85). However, evidence suggests that well-formulated ketogenic diets do not have any safety concerns for the general population (85). Although the ketogenic diet has a long history in clinical medicine and human evolution, high-quality clinical trials are needed to assess important questions about the long-term effects of ketogenic diets in healthy populations before the long-term effects of ketogenic diets can be studied in TBI.

# **Experimental Models**

To fully understand whether ketones are beneficial in treating TBIs, experimental models of TBI must be improved. Thus far, animal models of TBI have played an important role in characterizing TBI pathophysiology and in developing treatment strategies (15). However, animal models of TBI feature a homogenous type of injury with variables such as age, sex, genetic background, and injury parameters being well-controlled. In contrast, human TBI patients present a pathophysiological heterogeneity that can arise from the location, nature, and severity of the primary injury as well as preexisting factors and conditions such as age, health, sex, medication use, alcohol and drug use, and genetics (86). Furthermore, no studies to date have evaluated racial diversity in the pathophysiology of TBIs. As a result, no single animal TBI study can fully describe all possible pathophysiological consequences in human TBI. The heterogenic nature of TBIs may explain why the majority of pharmaceutical drugs identified to be effective in animal TBI models have failed in phase II and phase III clinical trials (87). The failure of translating preclinical animal studies into clinical settings underscores the need for new animal models of TBI to be developed and for existing models to be modified.

While early TBI research took place in rodent models and provided a fundamental understanding of TBI mechanisms, differences in rodent and human TBI pathophysiology exist. Because studying animal models with similar pathophysiological responses to humans is crucial to improving TBI treatments, the pig has become an increasingly prevalent TBI model (88). The pig is a more ideal model for TBI than rodents because its brain is closer in size, structure, composition, and development to the human brain and its immune system, inflammatory responses, and vasculature are more comparable to humans (88). Additionally, newly developed genetically engineered pigs with comorbidities and specific genetic factors associated with TBI may be helpful to test the effectiveness of novel therapeutic agents, including ketones, under conditions that increase the prevalence of TBI complications (88). Although the pig brain has numerous advantages that would help improve understanding human TBI, pig and human brains are not identical in pathological

responses to injury (88). Direct comparisons of human and pig brain TBI pathophysiological responses are lacking. Future studies that examine pathological differences and similarities between both species are needed to progress in developing more effective TBI treatments.

Thus far, the majority of TBI and ketone research has used rodent models. No studies to date have examined ketone administration in TBI pig models. However, a study examining mitochondrial function in response to cyclosporine administration in a TBI pig model showed that intravenous cyclosporine administration improved brain metabolism and mitochondrial function by closing the mitochondrial permeability transition pore (60). These results offer promise that ketones could also improve brain metabolism and mitochondrial function by providing an energy source to close the mitochondrial permeability transition pore in a large TBI model similar to humans.

The majority of TBI research in animals has used a controlled cortical impact (CCI) model, shock tube model, lateral fluid percussion injury (FPI) model, or weight drop model to replicate the mechanisms of TBI pathophysiology. The CCI model best replicates focal injuries with localized tissue deformation resulting from a blow to the head, such as in sports-related TBI or violent assaults (89, 90). The shock tube model best replicates diffuse injuries caused by acceleration and deceleration forces, such as in car accidents and blast-related military TBI (89). The FPI model replicates TBI with both focal and diffuse injury patterns, and the weight drop model can replicate focal or diffuse injuries depending on the technique used (89). Because each of these TBI models have different associated injury mechanisms and pathological consequences, utilizing multiple models in ketone and TBI research would allow the effectiveness of ketones in TBI treatment to be evaluated across various types of civilian and military TBI.

# Human Traumatic Brain Injuries and Ketones

Most evidence regarding the benefits of ketones in treating TBI has come from animal studies. Ketone therapy research in human TBI is only beginning to look at the safety and efficacy of ketone supplementation. The most recent research includes a phase II clinical trial that found an enterally administered ketogenic formulation can be safely administered to TBI patients while improving glucose control and an ongoing phase I clinical trial investigating the safety, feasibility, and effectiveness of an enteral ketogenic formulation for severe TBI (91, 92). Another recent study examined the ketogenic diet and medium chain triglyceride supplementation in post-concussion syndrome and reported improvements in visual memory and post-concussion syndrome symptomology (93). Together, these studies have established a physiological and symptomatic rationale for future clinical studies to examine ketone therapy in TBI.

Although not yet widely studied in human TBIs, ketones have been used more frequently in other health conditions with similar pathophysiological consequences. In type 2 diabetes, where cognitive impairment is associated with cerebral glucose hypometabolism, intravenous administration of ketone  $\ensuremath{\mathsf{TABLE 3}}\xspace$  ] A summary of how ketones have been used in health conditions relating to TBI pathophysiology.

Health condition	Study	Ketogenic agent	Findings
Type 2 diabetes	(94)	Ketone salt	Improved working memory performance
Aging	(95)	Ketone ester	Improved brain network stability
Alzheimer's	(96)	Ketogenic diet	Improved metabolism, reduced neuronal cell death
Parkinson's	(99)	Ketogenic diet	Improved motor and non-motor symptoms
Migraines	(101)	Ketogenic diet	Reduction in migraine frequency, severity, and medication use
Epilepsy	(105)	Ketogenic diet	Reduction in seizure frequency in children
	(106)	Ketogenic diet	Reduction in seizure frequency in adults

salts improves working memory performance (94). In the process of aging, where age-based cognitive impairment has been linked with cerebral glucose hypometabolism, ketosis achieved through both the ketogenic diet and ketone ester supplementation improves functional communication between brain regions (95). In Alzheimer's, where disease progression is linked to impaired glucose metabolism and neuronal cell death, the ketogenic diet mitigates neurodegeneration processes (96-98). In Parkinson's disease, where cells experience abnormal energy metabolism, oxidative stress, inflammation, and increased apoptosis, the ketogenic diet improves both motor and non-motor symptoms (99, 100). In migraines, where migraines are thought to be triggered by cerebral energy deficiencies or oxidative stress, the ketogenic diet reduces migraine frequency, severity, and medication use (101, 102). In epilepsy, where mitochondrial bioenergetics are impaired, the ketogenic diet has been used since the 1920s in children to reduce seizures and has been increasingly used in adults with drug resistant epilepsy (103, 104). Several clinical trials support the belief that ketogenic diets reduce seizures in both children and adults (105, 106). See Table 3 for a summary of how ketones have been used in health conditions relating to TBI pathophysiology.

The studies examining ketones in the treatment of various health conditions involving disruptions in cerebral metabolism focus on outcomes and symptoms that differ from TBI treatment measures. However, the underlying mechanisms behind why ketones help improve such outcomes provides support for why and how ketones can address similar mechanisms present in TBIs. Support for use of the ketogenic diet and forms of exogenous ketones in other health conditions involving disruptions in cerebral metabolism offer promise for future TBI and ketone studies in humans.

## **Metabolic Monitoring**

For ketones to be studied more extensively in clinical settings, methods of monitoring cerebral metabolism must be improved. Positron emission tomography (PET) is a common, non-invasive imaging approach used for various neurological problems, including TBI (107). In PET, <sup>18</sup>F is attached to deoxyglucose and can quantify the proportion of glucose uptake and phosphorylation, and <sup>11</sup>C is attached to acetoacetate to quantify the proportion of ketone body uptake (78, 107, 108). While PET imaging is informative, it has several limitations, including limited spatial resolution, the amount of radioisotope that can be delivered to injured tissue relative to normal tissue, procedural aspects that may not be feasible for every patient, and the inability to continuously monitor cerebral metabolism (78, 107). The most significant limitation of PET is that the absolute quantification of cerebral glucose and ketone uptake requires a complex and invasive procedure not suitable for clinical routine. The absolute quantification of metabolites requires an arterial input function that describes a non-metabolized compound's concentration as a function of time (109). The determination of an arterial input function is an invasive procedure and can disturb a patient's physiological status (109). While standardized uptake values can be used to provide semi-quantitative measures of cerebral metabolite uptake, standardized uptake values lack an absolute physiological scale that limits the ability to accurately assess potential TBI treatments (109, 110).

An image-derived input function is a non-invasive alternative to an arterial input function that can accurately quantify absolute physiological values. The most accurate method of calculating an image-derived input function uses a fully integrated positron emission tomography-magnetic resonance imaging (PET-MRI) system (111). Until recently, PET and MRI could not be performed simultaneously because the magnets in MRI scanners interfered with detectors in PET scanners, and merging separate PET and MRI images presented logistical challenges that made calculating an accurate image-derived input function a challenge (112). Recent advances in research have developed PET-MRI systems that can be used in combination with computational frameworks to generate an image-derived input function that accurately reflects arterial input function values (109, 111). Because the integrated PET-MRI system is a relatively new technology, it is primarily used in research settings and not yet available for routine clinical use. The first integrated PET-MRI systems in the United States were installed in 2010, and in 2017, there were 30 PET-MRI systems in the United States (113). As PET-MRI systems become more prevalent, the absolute quantification of physiological metabolite levels through image-derived input functions will allow more accurate evaluations of potential TBI treatments, including ketones.

Cerebral microdialysis (CMD) is an alternative approach to monitor cerebral metabolism. CMD allows semi-continuous bedside monitoring of cerebral metabolism by sampling parenchymal extracellular fluid with a probe to obtain concentrations of specific biomarkers (114). Various metabolites involved in cerebral glucose, ketone, and oxidative metabolism can be monitored *via* CMD (18, 29, 81). Advances in CMD use and interpretation have allowed CMD to become a routine neuromonitoring tool to help guide individualized treatment in neurointensive care TBI patients (114, 115). However, CMD is limited because metabolites can only be measured in a very narrow zone surrounding the CMD probe (116). While this narrow measurement zone can help understand the metabolism of specific areas in the brain, CMD does not provide a complete metabolic picture of the entire brain. Because cerebral energy metabolism is usually very different in different parts of the brain in TBI patients, treatments targeting specific aspects of cerebral metabolism based on CMD measurements may not prove effective for the entire brain (116). Another limitation of CMD is that CMD is generally only used in intensive care units with severe TBI patients and is not suitable for milder forms of TBI, such as concussions. While CMD has been used in animal studies to study milder forms of TBI, its practicality in clinical settings is limited.

Options for monitoring ketone metabolism in milder forms of TBI and outpatient settings are limited because cerebral ketone concentrations cannot be determined with the methods currently available. These options include urinalysis, breath analyzers, and blood meters. Urinalyses methods predominately use a urine ketone stick test, which can give a semi-quantitative and inexpensive measure of acetoacetate (117). However, the quantity of ketones in urine does not equate to plasma ketone concentrations because urine tests do not detect β-hydroxybutyrate, which is the predominate circulating ketone (117). Breath analyzers are a convenient and fast way to measure ketone levels. The main advantage of ketone breath analyzers is that they allow an unlimited number of ketone measurements to be easily conducted after the user purchases a breath analyzer device. However, ketone breath analyzers only measure the ketone body acetate, which is exhaled as a byproduct of fat metabolism. While acetate can be a reliable marker of endogenous ketosis and fat metabolism, levels of exogenous ketones may not be accurately detected (118). Furthermore, only one ketone breath analyzer (Biosense) on the market has been shown to have a high level of accuracy (119). Ketone blood meters measure capillary concentrations of β-hydroxybutyrate and are an accurate way of measuring circulating levels of ketones (117). The biggest limitation of ketone blood meters is that ketone test strips are relatively expensive, and finger prick tests may need to be performed multiple times throughout the day (117). One way to reduce the costs and improve the convivence of accurately measuring ketone levels is through developing a continuous ketone monitor, similar to that of continuous glucose monitoring systems. The first proof of concept of a continuous ketone monitor that can be placed within the subcutaneous tissue to measure ketone concentration for 14 days was recently completed (120). However, more research is needed to evaluate the performance of the device in various patient populations (120). Thus, to improve TBI treatment strategies in both intensive care and outpatient settings, it will be essential to establish technologies that can monitor ketone metabolism in accurate and minimally invasive ways. Improved metabolic monitoring technology will allow the development of quantitative measures to guide the ketone therapy methodology and assess the effectiveness of ketone therapy in TBI.

## **Ketone Levels**

To translate experimental data supporting the benefits of ketone metabolism in TBI into clinical practice, the optimal level of cerebral ketone bodies must be determined. Thus far, most ketone research has focused on plasma levels of ketone bodies, and limited data exists about ketone body levels in the brain. A study exploring cerebral ketone metabolism following TBI in humans indicates that plasma ketone bodies are the primary source of cerebral ketone bodies (81). However, the level of cerebral ketone bodies required to effectively address TBI mechanisms and the level of plasma ketone bodies required to achieve such cerebral ketone body levels are unknown (91).

Achieving optimal cerebral ketone body levels is complicated by ketone body transport into the brain that requires monocarboxylate transports to cross the blood brain barrier. Even if plasma ketone body levels are high enough to theoretically reach optimal cerebral ketone body levels, ketone transport into the brain is limited by monocarboxylate transporters. Further complicating this issue is that monocarboxylate transporters are expressed significantly more in pediatric TBIs compared to adult TBIs (121). Therefore, strategies such as fasting may be needed to increase ketone transport across the blood brain barrier to provide sufficient levels of cerebral ketone bodies in adult patients.

In addition to understanding the level of plasma and cerebral ketones needed to target TBI mechanisms, the optimal method of increasing cerebral ketone body levels must be determined. A ketogenic diet enriched with medium chain triglycerides provides only small increases in plasma ketone body levels and may not be sufficient to increase cerebral ketone body levels (81, 91). Other methods, such as intermittent feeding and exogenous ketone salts and esters, may be needed to raise ketone body concentrations high enough to serve as a cerebral metabolic substrate. Intermittent rather than continuous methods of feeding may be helpful to promote the metabolic switch from reliance on glucose to reliance upon ketone bodies and promote increased ketone body uptake by monocarboxylate transporters (122). Exogenous ketone delivery through ketone esters and ketone salts could be more effective in raising ketone body levels instead of using a medium chain triglyceride enriched diets to produce endogenous ketone bodies (81). Ketone esters may be the best approach to raise plasma ketone body levels to therapeutic concentrations as ketone esters result in a higher plasma ketone body level increase.

# REFERENCES

- Centers for Disease Control and Prevention. Report to Congress on Traumatic Brain Injury in the United States: Epidemiology and Rehabilitation. (2015). Available online at: https://www.cdc.gov/traumaticbraininjury/pdf/ TBI\_Report\_to\_Congress\_Epi\_and\_Rehab-a.pdf (accessed May 26, 2021).
- Centers for Disease Control and Prevention. Surveillance Report of Traumatic Brain Injury-Related Emergency Department Visits, Hospitalizations, and Deaths-United States, 2014. (2019). Available online at: https://www. cdc.gov/traumaticbraininjury/pdf/TBI-Surveillance-Report-FINAL\_508. pdf (accessed May 26, 2021).
- van Dijck JTJM, Dijkman MD, Ophuis RH, de Ruiter GCW, Peul WC, Polinder S. In-hospital costs after severe traumatic brain injury: a systematic review and quality assessment. *PLoS One.* (2019) 14:e0219529. doi: 10.1371/journal.pone.0219529

However, no studies to date have directly compared the effects of different ketogenic agents in TBIs. Future studies in both human and animal models are needed to determine the optimal level of plasma and cerebral ketone body levels and the most effective method to achieve such levels.

# DISCUSSION

Our understanding of TBI pathophysiology has drastically improved in the past 20 years. However, more than 30 large clinical trials have failed to identify targeted pharmacological therapies in TBI patients that improve recovery outcomes. The heterogenic nature of TBI presents challenges in both TBI research and treatment. Because disruptions in cerebral metabolism are a commonality across all forms of TBI, ketone therapy may be a valid therapeutic approach to improve recovery by boosting cerebral energy metabolism, mitigating secondary injury mechanisms, and exerting neuroprotective effects against acute and progressive neurodegenerative processes. The mechanisms of ketone therapy for TBI are beginning to be understood, however more research is needed in both animal models and humans to elucidate how to effectively utilize ketones and translate pre-clinical findings into clinical practice. Specifically, age-dependent differences in ketone metabolism, sex differences in the response to ketone administration, the timing and duration of ketone administration, and optimal level of cerebral ketone levels needs to be explored. Advances could also be made to improve TBI modeling techniques and methods of monitoring cerebral metabolism. Overall, ketone therapy holds significant promise as an option for TBI treatment, and preliminary evidence strongly warrants further investigation of ketones in TBI treatment.

# **AUTHOR CONTRIBUTIONS**

SD confirms sole responsibility for the conception and design and manuscript preparation of this review.

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- Hajiaghamemar M, Seidi M, Oeur AO, Margulies SS. Toward development of clinically translatable diagnostic and prognostic metrics of traumatic brain injury using animal models: a review and a look forward. *Exp Neurol.* (2019) 318:101–23. doi: 10.1016/j.expneurol.2019.04.019
- Lazaridis C, Rusin CG, Robertson CS. Secondary brain injury: predicting and preventing insults. *Neuropharmacology*. (2019) 145:145–52. doi: 10.1016/j.neuropharm.2018.06.005
- Mckee AC Daneshvar DH. The neuropathology of traumatic brain injury. *Handb Clin Neurol.* (2015) 127:45– 66. doi: 10.1016/B978-0-444-52892-6.00004-0
- Di Pietro V, Yakoub KM, Caruso G, Lazzarino G, Signoretti S, Barbey AK, et al. Antioxidant therapies in traumatic brain injury. *Antioxidants.* (2020) 9:260. doi: 10.3390/antiox9030260
- Giza CC Hovda DA. The new neurometabolic cascade of concussion. *Neuro-surgery*. (2014) 75:S24–S33. doi: 10.1227/NEU.00000000000505

- Ng YS, Lee AYW. Traumatic brain injuries: pathophysiology and potential therapeutic targets. *Front Cell Neurosci.* (2019) 13:528. doi: 10.3389/fncel.2019.00528
- Maudsley AA, Govind V, Saigal G, Gold SG, Harris L, Sheriff S. Longitudinal MR spectroscopy shows altered metabolism in traumatic brain injury. J Neuroimaging. (2017) 27:562–9. doi: 10.1111/jon.12463
- Armstead WM. Cerebral blood flow autoregulation and dysautoregulation. *Anesthesiol Clin.* (2016) 34:465–77. doi: 10.1016/j.anclin.2016.04.002
- Belosludtsev KN, Dubinin MV, Belosludtseva NV, Mironova GD. Mitochondrial Ca<sup>2+</sup> transport: mechanisms, molecular structures, and role in cells. *Biochemistry (Mosc).* (2019) 84:593–607. doi: 10.1134/S0006297919060026
- Prins ML, Greco T, Alexander D, Giza CC. The pathophysiology of traumatic brain injury at a glance. *Dis Model Mech.* (2013) 6:1307–15. doi: 10.1242/dmm.011585
- Veech RL, Valeri RC, Vanltallie TB. The mitochondrial permeability transition pore provides a key to the diagnosis and treatment of traumatic brain injury. *IUMB Life.* (2012) 64:203–7. doi: 10.1002/iub.590
- Xiong Y, Mahmood A, Chopp M. Animal models of traumatic brain injury. Nat Rev Neurosci. (2013) 14:128–42. doi: 10.1038/nrn3407
- Prins ML, Matsumoto JH. The collective therapeutic potential of cerebral ketone metabolism in traumatic brain injury. J Lipid Res. (2014) 55:2450–7. doi: 10.1194/jlr.R046706
- Sheline CT, Behrens MM, Choi DW. Zinc-induced cortical neuronal death: contribution of energy failure attributable to loss of NAD(+) and inhibition of glycolysis. J Neurosci. (2000) 20:3139–46. doi: 10.1523/JNEU-ROSCI.20-09-03139.2000
- Carpenter KL, Jalloh I, Hutchinson PJ. Glycolysis and the significance of lactate in traumatic brain injury. *Front Neurosci.* (2015) 9:112. doi: 10.3389/fnins.2015.00112
- Jalloh I, Carpenter KL, Grice P, Howe DJ, Mason A, Gallagher CN, et al. Glycolysis and the pentose phosphate pathway after human traumatic brain injury: microdialysis studies using 1,2-(13)C2 glucose. J Cereb Blood Flow Metab. (2015) 35:111–20. doi: 10.1038/jcbfm.2014.177
- Sharma P, Benford B, Li ZZ, Ling GS. Role of pyruvate dehydrogenase complex in traumatic brain injury and measurement of pyruvate dehydrogenase enzyme by dipstick test. J Emerg Trauma Shock. (2009) 2:67–72. doi: 10.4103/0974-2700.50739
- Xu Y, McArthur DL, Alger JR, Etchepare M, Hovda DA, Glenn TC, et al. Early nonischemic oxidative metabolic dysfunction leads to chronic brain atrophy in traumatic brain injury. *J Cereb Blood Flow Metab.* (2010) 30:883– 94. doi: 10.1038/jcbfm.2009.263
- Wright MJ, McArthur DL, Alger JR, Van Horn J, Irimia A, Filippou M, et al. Early metabolic crisis-related brain atrophy and cognition in traumatic brain injury. *Brain Imaging Behav.* (2013) 7:307–15. doi: 10.1007/s11682-013-9231-6
- Glenn TC, Kelly DF, Boscardin WJ, McArthur DL, Vespa P, Oertel M, et al. Energy dysfunction as a predictor of outcome after moderate or severe head injury: indices of oxygen, glucose, and lactate metabolism. *J Cereb Blood Flow Metab.* (2003) 23:1239–50. doi: 10.1097/01.WCB.0000089833.23606.7F
- Fukao T, Lopaschuk GD, Mitchell GA. Pathways and control of ketone body metabolism: on the fringe of lipid biochemistry. *Prostaglandins Leukot Essent Fatty Acids*. (2004) 70:243–51. doi: 10.1016/j.plefa.2003.11.001
- Le Foll C, Levin BE. Fatty acid-induced astrocyte ketone production and the control of food intake. *Am J Physiol Regul Integr Comp Physiol.* (2016) 310:R1186–R92. doi: 10.1152/ajpregu.00113.2016
- Dhillon KK. Gupta S. Biochemistry, Ketogenesis. Florida: StatPearls Publishing (2021). Available online at: https://www.ncbi.nlm.nih.gov/books/ NBK493179/ (accessed May 26, 2021).
- Stubbs BJ, Cox PJ, Evans RD, Santer P, Miller JJ, Faull OK, et al. On the metabolism of exogenous ketones in humans. *Front Physiol.* (2017) 8:848. doi: 10.3389/fphys.2017.00848
- Hashim SA, VanItallie TB. Ketone body therapy: from the ketogenic diet to the oral administration of ketone ester. J Lipid Res. (2014) 55:1818–26. doi: 10.1194/jlr.R046599
- 29. White H, Venkatesh B. Clinical review: ketones and brain injury. *Crit Care.* (2011) 15:219. doi: 10.1186/cc10020
- Marten B, Pfeuffer M, Schrezenmeir J. Medium chain triglycerides. Int Dairy J. (2006) 16:1374–82. doi: 10.1016/j.idairyj.2006.06.015

- Henderson ST, Vogel JL, Barr LJ, Garvin F, Jones JJ, Costantini LC. Study of the ketogenic agent AC-1202 in mild to moderate Alzheimer's disease: a randomized, double-blind, placebo-controlled, multicenter trial. *Nutr Metab* (*Lond*). (2009) 6:31. doi: 10.1186/1743-7075-6-31
- 32. Page KA, Williamson A, Yu N, McNay EC, Dzuira J, McCrimmon RJ, et al. Medium-chain fatty acids improve cognitive function in intensively treated type 1 diabetic patients and support *in vitro* synaptic transmission during acute hypoglycemia. *Diabetes*. (2009) 58:1237–44. doi: 10.2337/db08-1557
- Evans M, Cogan KE, Egan B. Metabolism of ketone bodies during exercise and training: physiological basis for exogenous supplementation. J Physiol. (2017) 595:2857–71. doi: 10.1113/JP273185
- Fischer T, Och U, Klawon I, Och T, Grüneberg M, Fobker M, et al. Effect of a sodium and calcium DL-β-hydroxybutyrate salt in healthy adults. J Nutr Metab. (2018) 2018:9812806. doi: 10.1155/2018/9812806
- Oddo M, Vespa P, Menon DK. Boosting the injured brain with supplemental energy fuels. *Intensive Care Med.* (2019) 45:872–5. doi: 10.1007/s00134-018-05517-6
- 36. Thomsen, HH, Rittig N, Johannsen M, Møller AB, Jørgensen JO, et al. Effects of 3-hydroxybutyrate and free fatty acids on muscle protein kinetics and signaling during LPS-induced inflammation in humans: anticatabolic impact of ketone bodies. *Am J Clin Nutr.* (2018) 108:857–67. doi: 10.1093/ajcn/n qy170
- Müller MJ, Paschen U, Seitz HJ. Effect of ketone bodies on glucose production and utilization in the miniature pig. J Clin Invest. (1984) 74:249–61. doi: 10.1172/JCI111408
- Clarke K, Tchabanenko K, Pawlosky R, Carter E, Todd King M, Musa-Veloso K, et al. Kinetics, safety and tolerability of (R)-3-hydroxybutyl (R)-3hydroxybutyrate in healthy adult subjects. *Regul Toxicol Pharmacol.* (2012) 63:401–8. doi: 10.1016/j.yrtph.2012.04.008
- Shivva V, Cox PJ, Clarke K, Veech RL, Tucker IG, Duffull SB. The population pharmacokinetics of D-β-hydroxybutyrate following administration of (R)-3-hydroxybutyl (R)-3-hydroxybutyrate. *AAPS J.* (2016) 18:678–88. doi: 10.1208/s12248-016-9879-0
- Cox PJ, Kirk T, Ashmore T, Willerton K, Evans R, Smith A, et al. Nutritional ketosis alters fuel preference and thereby endurance performance in athletes. *Cell Metab.* (2016) 24:256–68. doi: 10.1016/j.cmet.2016.07.010
- Lyon R, Nimmo GR. Gastric tubes in patients with severe brain injury. Crit Care. (2008) 12:P132. doi: 10.1186/cc6353
- Lazzarino G, Amorini AM, Signoretti S, Musumeci G, Lazzarino G, Caruso G, et al. Pyruvate dehydrogenase and tricarboxylic acid cycle enzymes are sensitive targets of traumatic brain injury induced metabolic derangement. *Intl J Mol Sci.* (2019) 20:5774. doi: 10.3390/ijms20225774
- Lodish H, Berk A, Matsudaira P, Zipursky L, Baltimore D, Darnell J. "Oxidation of glucose fatty acids to CO<sub>2</sub>," In: *Molecular Cell Biology, 4th Edn.* New York, NY: W.H. Freeman (2000). Available online at: https://www.ncbi.nlm. nih.gov/books/NBK21475/ (accessed May 26, 2021).
- Wood TR, Stubbs BJ, Juul SE. Exogenous ketone bodies as promising neuroprotective agents for developmental brain injury. *Dev Neurosci.* (2018) 40:451–62. doi: 10.1159/000499563
- Cornelius C, Crupi R, Calabrese V, Graziano A, Milone P, Pennisi G, et al. Traumatic brain injury: oxidative stress and neuroprotection. *Antiox Redox Signal.* (2013) 19:836–53. doi: 10.1089/ars.2012.4981
- Yang H, Shan W, Zhu F, Whu J, Wang Q. Ketone bodies in neurological diseases: focus on neuroprotection and underlying mechanisms. *Front Neurol.* (2019) 10:585. doi: 10.3389/fneur.2019.00585
- Simon DW, McGeachy MJ, Bayir H, Clark R, Loane DJ, Kochanek PM. The far-reaching scope of neuroinflammation after traumatic brain injury. *Nat Rev Neurol.* (2017) 13:572. doi: 10.1038/nrneurol.2017.116
- Xiong Y, Mahmood A, Chopp M. Current understanding of neuroinflammation after traumatic brain injury and cell-based therapeutic opportunities. *Chin J Traumatol.* (2018) 21:137–51. doi: 10.1016/j.cjtee.2018.02.003
- 49. Ding K, Wang H, Xu J, Lu X, Zhang L, Zhu L. Melatonin reduced microglial activation and alleviated neuroinflammation induced neuron degeneration in experimental traumatic brain injury: possible involvement of mTOR pathway. *Neurochem Int.* (2015) 76:23–31. doi: 10.1016/j.neuint.2014.06.015
- Mortezaee K, Khanlarkhani N, Beyer C, Zendedel A. Inflammasome: its role in traumatic brain and spinal cord injury. J Cell Physiol. (2018) 233:5160–9. doi: 10.1002/jcp.26287

- Laplante M, Sabatini DM. mTOR signaling at a glance. J Cell Sci. (2009) 122:3589–94. doi: 10.1242/jcs.051011
- Ma D, Wang AC, Parikh I, Green SJ, Hoffman JD, Chlipala M, et al. Ketogenic diet enhances neurovascular function with altered gut microbiome in young healthy mice. *Sci Rep.* (2018) 8:6670. doi: 10.1038/s41598-018-2 5190-5
- Saxton RA, Sabatini DM. mTOR signaling in growth, metabolism, and disease. *Cell*. (2017) 168:960–76. doi: 10.1016/j.cell.2017.02.004
- 54. Youm YH, Nguyen KY, Grant RW, Goldberg EL, Bodogai M, Kim D, et al. The ketone metabolite  $\beta$ -hydroxybutyrate blocks NLRP3 inflammasome-mediated inflammatory disease. *Nat Med.* (2015) 21:263–9. doi: 10.1038/nm.3804
- O'Brien WT, Pham L, Symons GF, Monif M, Shultz SR, McDonald SJ. The NLRP3 inflammasome in traumatic brain injury: potential as a biomarker and therapeutic target. J Neuroinflammation. (2020) 17:104. doi: 10.1186/s12974-020-01778-5
- 56. Kong G, Liu J, Li R, Lin J, Huang Z, Yang Z, et al. Ketone metabolite  $\beta$ -hydroxybutyrate ameliorates inflammation after spinal cord injury by inhibiting the NLRP3 inflammasome. *Neurochem Res.* (2021) 46:213–29. doi: 10.1007/s11064-020-03156-2
- 57. Svart M, Gormsen LC, Hansen J, Zeidler D, Gejl M, Vang K, et al. Regional cerebral effects of ketone body infusion with 3-hydroxybutyrate in humans: reduced glucose uptake, unchanged oxygen consumption and increased blood flow by positron emission tomography. A randomized, controlled trial. *PLoS One.* (2018) 13:e0190556. doi: 10.1371/journal.pone.0190556
- Kim J, Whyte J, Patel S, Avants B, Europa E, Wang J, et al. Resting cerebral blood flow alterations in chronic traumatic brain injury: an arterial spin labeling perfusion FMRI study. *J Neurotrauma*. (2010) 27:1399–411. doi: 10.1089/neu.2009.1215
- Soustiel JF, Glenn TC, Shik V, Boscardin J, Mahamid E, Zaaroor M. Monitoring of cerebral blood flow and metabolism in traumatic brain injury. J Neurotrauma. (2005) 22:955–65. doi: 10.1089/neu.2005.22.955
- Karlsson M, Pukenas B, Chawla S, Ehinger JK, Plyler R, Stolow M, et al. Neuroprotective effects of cyclosporine in a porcine pre-clinical trial of focal traumatic brain injury. *J Neurotrauma*. (2019) 36:14–24. doi: 10.1089/neu.2018.5706
- Lemasters, JJ, Nieminen AL, Qian T, Trost LC, Elmore SP, et al. The mitochondrial permeability transition in cell death: a common mechanism in necrosis, apoptosis and autophagy. *Biochim Biophys Acta*. (1998) 1366:177– 96. doi: 10.1016/S0005-2728(98)00112-1
- Sullivan PG, Rabchevsky AG, Waldmeier PC, Springer JE. Mitochondrial permeability transition in CNS trauma: cause or effect of neuronal cell death? *J Neurosci Res.* (2005) 79:231–9. doi: 10.1002/jnr.20292
- Bonora M, Pinton P. The mitochondrial permeability transition pore and cancer: molecular mechanisms involved in cell death. *Front Oncol.* (2014) 4:302. doi: 10.3389/fonc.2014.00302
- Kim DY, Simeone KA, Simeone TA, Pandya JD, Wilke JC, Ahn Y, et al. Ketone bodies mediate antiseizure effects through mitochondrial permeability transition. *Ann Neurol.* (2015) 78:77–87. doi: 10.1002/ana.24424
- Miller VJ, Villamena FA, Volek JS. Nutritional ketosis and mitohormesis: potential implications for mitochondrial function and human health. J Nutr Metab. (2018) 2018:5157645. doi: 10.1155/2018/5157645
- Hu ZG, Wang HD, Jin W, Yin HX. Ketogenic diet reduces cytochrome c release and cellular apoptosis following traumatic brain injury in juvenile rats. *Ann Clin Lab Sci.* (2009) 39:76–83. Available online at: http://www. annclinlabsci.org/content/39/1/76.long
- Appelberg KS, Hovda DA, Prins ML. The effects of a ketogenic diet on behavioral outcome after controlled cortical impact injury in the juvenile and adult rat. J Neurotrauma. (2009) 25:497–506. doi: 10.1089/neu.2008.0664
- Prins ML, Fujima LS, Hovda DA. Age-dependent reduction of cortical contusion volume by ketones after traumatic brain injury. *J Neurosci Res.* (2005) 82:413–20. doi: 10.1002/jnr.20633
- Prins ML, Giza CC. Induction of monocarboxylate transporter 2 expression and ketone transport following traumatic brain injury in juvenile and adult rats. *Dev Neurosci.* (2006) 28:447–56. doi: 10.1159/000094170
- Bowman CE, Scafidi J, Scafidi, S. Metabolic perturbations after pediatric TBI: it's not just about glucose. *Exp Neurol.* (2019) 316:74–84. doi: 10.1016/j.expneurol.2019.03.018

- Halestrap AP. The monocarboxylate transporter family– structure and functional characterization. *IUBMB Life*. (2012) 64:1–9. doi: 10.1002/iub.573
- 72. Matsuyama S, Ohkura S, Iwata K, Uenoyama Y, Tsukamura, H, et al. Food deprivation induces monocarboxylate transporter 2 expression in brainstem of female rat. *J Reprod Dev.* (2009) 55:256–61. doi: 10.1262/jrd.20214
- Gupte R, Brooks W, Vukas R, Pierce J, Harris J. Sex differences in traumatic brain injuries: what we know and what we should know. *J Neurotrauma*. (2019) 36:3063–91. doi: 10.1089/neu.2018.6171
- Demarest TG, McCarthy MM. Sex differences in mitochondrial (dys)function: implications for neuroprotection. J Bioenerg Biomembr. (2015) 47:173–88. doi: 10.1007/s10863-014-9583-7
- Günther M, Plantman S, Davidsson J, Angéria M, Mathiesen T, Risling M. COX-2 regulation and TUNEL-positive cell death differ between genders in the secondary inflammatory response following experimental penetrating focal brain injury in rats. *Acta Neurochir.* (2015) 157:649–59. doi: 10.1007/s00701-014-2331-2
- Villapol S, Loane DJ, Burns MP. Sexual dimorphism in the inflammatory response to traumatic brain injury. *Glia.* (2017) 65:1423–38. doi: 10.1002/glia.23171
- 77. Wagner AK, Bayir H, Ren D, Puccio A, Zafonte RD, Kochanek PM. Relationships between cerebrospinal fluid markers of excitotoxicity, ischemia, and oxidative damage after severe TBI: the impact of gender, age, and hypothermia. J Neurotrauma. (2004) 21:125–36. doi: 10.1089/089771504322778596
- Greco T, Vespa PM, Prins ML. Alternative substrate metabolism depends on cerebral metabolic state following traumatic brain injury. *Exp Neurol.* (2020) 329:113289. doi: 10.1016/j.expneurol.2020.113289
- Fedorovich SV, Voronina PP, Waseem TV. Ketogenic diet versus ketoacidosis: what determines the influence of ketone bodies on neurons? *Neural Regen Res.* (2018) 13:2060–3. doi: 10.4103/1673-5374.241442
- Kupina NC, Detloff MR, Bobrowski WF, Snyder BJ, Hall ED. Cytoskeletal protein degradation and neurodegeneration evolves differently in males and females following experimental head injury. *Exp Neurol.* (2003) 180:55–73. doi: 10.1016/S0014-4886(02)00048-1
- Bernini A, Masoodi M, Solari D, Miroz JP, Carteron L, Chrisinat N, et al. Modulation of cerebral ketone metabolism following traumatic brain injury in humans. J Cereb Blood Flow Metab. (2020) 40:177–86. doi: 10.1177/0271678X18808947
- Blaise JH, Ruskin DN, Koranda JL, Masino SA. Effects of a ketogenic diet on hippocampal plasticity in freely moving juvenile rats. *Physiol Rep.* (2015) 3:e12411. doi: 10.14814/phy2.12411
- Zarnowska IM. Therapeutic use of the ketogenic diet in refractory epilepsy: what we know and what still needs to be learned. *Nutrients*. (2020) 12:2616. doi: 10.3390/nu12092616
- Sheriff FG, Hinson HE. Pathophysiology and clinical management of moderate and severe traumatic brain injury in the ICU. Semin Neurol. (2015) 35:42–9. doi: 10.1055/s-0035-1544238
- Ludwig DS. The ketogenic diet: evidence for optimism but high-quality research needed. J Nutr. (2020) 150:1354–59. doi: 10.1093/jn/nxz308
- Margulies S, Hicks R. Combination therapies for traumatic brain injury: prospective considerations. J Neurotrauma. (2009) 26:925–39. doi: 10.1089/neu.2008.0794
- Briones TL. Animal models of traumatic brain injury: is there an optimal model that parallels human brain injury? *Annu Rev Nurs Res.* (2015) 33:31–73. doi: 10.1891/0739-6686.33.31
- Kinder HA, Baker EW, West FD. The pig as a preclinical traumatic brain injury model: current models, functional outcome measures, and translational detection strategies. *Neural Regen Res.* (2019) 14:413–24. doi: 10.4103/1673-5374.245334
- Ma X, Aravind A, Pfister, BJ, Chandra N, Haorah J. Animal models of traumatic brain injury and assessment of injury severity. *Mol Neurobiol.* (2019) 56:5332–45. doi: 10.1007/s12035-018-1454-5
- Osier ND, Dixon CE. The controlled cortical impact model: applications, considerations for researchers, and future directions. *Front Neurol.* (2016) 7:134. doi: 10.3389/fneur.2016.00134
- 91. White H, Venkatesh B, Jones M, Kruger PS, Walsham J, Fuentes H. Inducing ketogenesis *via* an enteral formulation in patients with acute brain injury: a phase II study. *Neurol Res.* (2020) 42:275–85. doi: 10.1080/01616412.2019.1709743

- Arora N. Ketogenic Diet for Traumatic Brain Injury (KETI). (2021). Available online at: https://clinicaltrials.gov/ct2/show/NCT03982602 (accessed May 27, 2021).
- Rippee MA, Chen J, Taylor MK. The ketogenic diet in the treatment of post-concussion syndrome-a feasibility study. *Front Nutr.* (2020) 7:160. doi: 10.3389/fnut.2020.00160
- Jensen NJ, Nilsson M, Ingerslev JS, Olsen DA, Fenger M, Svart M, et al. Effects of β-hydroxybutyrate on cognition in patients with type 2 diabetes. *Eur J Endocrinol.* (2020) 182:233–42. doi: 10.1530/EJE-19-0710
- 95. Mujica-Parodi LR, Amgalan A, Sultan SF, Antal B, Sun X, Skiena S, et al. Diet modulates brain network stability, a biomarker for brain aging, in young adults. *Proc Natl Acad Sci U S A.* (2020) 117:6170–7. doi: 10.1073/pnas.1913042117
- Broom GM, Shaw IC, Rucklidge JJ. The ketogenic diet as a potential treatment and prevention strategy for Alzheimer's disease. *Nutrition*. (2019) 60:118–21. doi: 10.1016/j.nut.2018.10.003
- Cunnane S, Nugent S, Roy M, Courchesne-Loyer A, Croteau E, Tremblay S, et al. Brain fuel metabolism, aging, and Alzheimer's disease. *Nutrition*. (2011) 27:3–20. doi: 10.1016/j.nut.2010.07.021
- de la Monte SM. Brain insulin resistance and deficiency as therapeutic targets in Alzheimer's disease. *Curr Alzheimers Res.* (2012) 9:35–66. doi: 10.2174/156720512799015037
- Phillips M, Murtagh D, Gilbertson LJ, Asztely F, Lynch C. Low-fat versus ketogenic diet in Parkinson's disease: a pilot randomized controlled trial. *Mov Disord*. (2018) 33:1306–14. doi: 10.1002/mds.27390
- 100. Norwitz NG, Hu MT, Clarke K. The mechanisms by which the ketone body d-β-hydroxybutyrate may improve the multiple cellular pathologies of Parkinson's disease. *Front Nutr.* (2019) 6:63. doi: 10.3389/fnut.2019.00063
- 101. Di Lorenzo C, Coppola G, Sirianni G, Bracaglia M, Di Lenola D, Siracusano A, et al. Migraine improvement during short lasting ketogenesis: a proof-ofconcept study. *Eur J Neurol.* (2015) 22:170–7. doi: 10.1111/ene.12550
- 102. Gross EC, Klement RJ, Schoenen J, D'Agostino DP, Fischer D. Potential protective mechanisms of ketone bodies in migraine prevention. *Nutrients*. (2019) 11:811. doi: 10.3390/nu11040811
- Gavrilovici C, Rho JM. Metabolic epilepsies amenable to ketogenic therapies: indications, contra-indications and underlying mechanisms. J Inherit Metab Dis. (2020) 44:42–53. doi: 10.1002/jimd.12283
- Husari KS, Cervenka MC. The ketogenic diet all grown up ketogenic diet therapies for adults. *Epilepsy Res.* (2020) 162:106319. doi: 10.1016/j.eplepsyres.2020.106319
- 105. Kverneland M, Molteberg E, Iversen PO, Veierød MB, Taubøll E, Selmer KK, et al. Effect of modified Atkins diet in adults with drug-resistant focal epilepsy: a randomized clinical trial. *Epilepsia*. (2018) 59:1567–76. doi: 10.1111/epi.14457
- 106. Zare M, Okhovat AA, Esmaillzadeh A, Mehvari J, Najafi MR, Saadatnia M. Modified Atkins diet in adult with refractory epilepsy: a controlled randomized clinical trial. *Iran J Neurol.* (2017) 16:72–7.
- 107. Byrnes KR, Wilson CM, Brabazon F, von Leden, Jurgens JS, Oakes TR, et al. FDG-PET imaging in mild traumatic brain injury: a critical review. Front Neuroenergetics. (2014) 5:13. doi: 10.3389/fnene.2013.00013
- 108. Courchesne-Loyer A, Croteau E, Castellano CA, St-Pierre V, Hennebelle M, Cunnane SC. Inverse relationship between brain glucose and ketone metabolism in adults during short-term moderate dietary ketosis: a dual tracer quantitative positron emission tomography study. J Cereb Blood Flow Metab. (2017) 37:2485–93. doi: 10.1177/0271678X16669366
- 109. Sundar LK, Muzik O, Rischka L, Hahn A, Lanzenberger R, Hienert M, et al. Towards quantitative (18F)FDG-PET/MRI of the brain: automated MRdriven calculation of an image-derived input function for the non-invasive

determination of cerebral glucose metabolic rates. *J Cereb Blood Flow Metab.* (2019) 39:1516–30. doi: 10.1177/0271678X18776820

- Berti V, Vanzi E, Polito C, Pupi A. Back to the future: the absolute quantification of cerebral metabolic rate of glucose. *Clin Transl Imaging*. (2013) 1:289–96. doi: 10.1007/s40336-013-0030-2
- 111. Sundar LK, Muzik O, Rischka L, Hahn A, Lanzenberger R, Hienert M, et al. Promise of fully integrated PET/MRI: noninvasive clinical quantification of cerebral glucose metabolism. J Nucl Med. (2020) 61:276–84. doi: 10.2967/jnumed.119.229567
- Vandenberghe S, Marsden PK. PET-MRI: a review of challenges and solutions in the development of integrated multimodality imaging. *Phys Med Biol.* (2015) 60:R115–R54. doi: 10.1088/0031-9155/60/4/R115
- 113. Ehman EC, Johnson GB, Villanueva-Meyer JE, Cha S, Leynes AP, Larson P, et al. PET/MRI: where might it replace PET/CT? J Magn Reson Imaging. (2017) 46:1247–62. doi: 10.1002/jmri.25711
- 114. Kurtz P, Rocha E. Nutrition therapy, glucose control, and brain metabolism in traumatic brain injury: a multimodal monitoring approach. *Front Neurosci.* (2020) 14:190. doi: 10.3389/fnins.2020.00190
- 115. Oddo M, Hutchinson PJ. Understanding and monitoring brain injury: the role of cerebral microdialysis. *Intensive Care Med.* (2018) 44:1945–48. doi: 10.1007/s00134-017-5031-6
- Nordström C. Cerebral microdialysis in TBI-limitations and possibilities. Acta Neurochir (Wein). (2017) 159:2275–7. doi: 10.1007/s00701-017-3339-1
- Dhatariya K. Blood ketones: measurement, interpretation, limitations, and utility in the management of diabetic ketoacidosis. *Rev Diabet Stud.* (2016) 13:217–25. doi: 10.1900/RDS.2016.13.217
- Anderson JC. Measuring breath acetone for monitoring fat loss: review. Obesity (Silver Spring). (2015) 23:2327–34. doi: 10.1002/oby.21242
- Suntrup Iii DJ, Ratto TV, Ratto M, McCarter JP. Characterization of a high-resolution breath acetone meter for ketosis monitoring. *PeerJ.* (2020) 8:e9969. doi: 10.7717/peerj.9969
- Alva S, Castorino K, Cho H, Ou J. Feasibility of continuous ketone monitoring in subcutaneous tissue using a ketone sensor. J Diabetes Sci Technol. (2021) 15:768–74. doi: 10.1177/19322968211008185
- 121. Prins ML. Cerebral metabolic adaptation and ketone metabolism after brain injury. J Cereb Blood Flow Metab. (2008) 28:1–16. doi: 10.1038/sj.jcbfm.9600543
- 122. Bear DE, Hart N, Puthucheary Z. Continuous or intermittent feeding: pros and cons. *Curr Opin Crit Care.* (2018) 24:256–61. doi: 10.1097/MCC.000000000000513

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