Increased oxidative stress and coenzyme Q10 deficiency in centenarians

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Aging populations are expanding worldwide, and the increasing requirement for nursing care has become a serious problem. Furthermore, successful aging is one of the highest priorities for individuals and societies. Centenarians are an informative cohort to study and inflammation has been found to be a key factor in predicting cognition and physical capabilities. Inflammation scores have been determined based on the levels of cytokines and Creactive protein, however, serum antioxidants and lipid profiles have not been carefully examined. We found that the redox balance of coenzyme Q10 significantly shifted to the oxidized form and levels of strong antioxidants, such as ascorbic acid and unconjugated bilirubin, decreased significantly compared to 76year-old controls, indicating an increased oxidative stress in centenarians. Levels of uric acid, an endogenous peroxynitrite scavenger, remained unchanged, suggesting that centenarians were experiencing moderate, chronic inflammatory conditions. Centenarians exhibited a hypocholesterolemic condition, while an increase in the ratio of free cholesterol to cholesterol esters suggests some impairment of liver function. Serum free fatty acids and monoenoic acid composition, markers of tissue oxidative damage, were significantly decreased in centenarians, indicating an impairment in the tissue repair system. Despite an elevation of the coenzyme Q10 binding protein Psap, serum total coenzyme Q10 levels decreased in centenarians. This suggests a serious deficiency of coenzyme Q10 in tissues, since tissue levels of coenzyme Q10 significantly decrease with age. Therefore, coenzyme Q10 supplementation could be beneficial for centenarians.

Key Words: coenzyme Q10, cholesterol metabolism, free fatty acids and their composition, centenarians, prosaposin

A ging populations are expanding worldwide, and the increasing requirement for nursing care has become a serious problem. Successful aging without cognition loss and physical deficiencies is one of the highest priorities for individuals and societies. Arai *et al.*⁽¹⁾ found that only 20% of centenarians enjoyed physical and cognitive independence at the age of 100 years, although most remained independent in daily living into their 90s. Those who maintained physical independence at 100 years of age were highly likely to become semi-supercentenarians (over 105 years) or supercentenarians (beyond 110 years).⁽¹⁾

To identify key factors in successful aging, Arai *et al.*⁽²⁾ focused on the characteristics of centenarians, semi-supercentenarians, and supercentenarians. They found that inflammation predicted cognition and physical capabilities in (semi-) supercentenarians better than chronologic age or gender. Interestingly, the inflammation score was lower in centenarian offspring compared to age-matched controls.⁽²⁾ They concluded that inflammation is an important malleable driver of aging up to extreme old age in humans.⁽²⁾ Other reviews also emphasize that the suppression of chronic inflammation is an important driver of successful aging at extreme old age. $^{(\mathrm{I},3,4)}$

In the above study, an inflammation score was estimated using cytomegalovirus immunoglobulin G antibody titers and plasma levels of interleukin- 6, tumor necrosis factor- α , and C-reactive protein.⁽²⁾ Acute inflammation, such as sepsis, is characterized by the formation of reactive oxygen and nitrogen species such as superoxide and nitric oxide.⁽⁵⁻⁷⁾ Therefore, peroxynitrite is also an important reactive molecule since it is produced from the combination of superoxide and nitric oxide.⁽⁵⁻⁷⁾ In fact, we observed a decline in plasma antioxidants, namely vitamin E (VE), ubiquinol-10 (CoQ10H₂), vitamin C (VC), and uric acid (UA), in patients with sepsis.⁽⁸⁾ However, no comprehensive study has been reported for centenarians. In this study, we compared serum levels of antioxidants in centenarians and 76-year-old controls. We found a significant decrease in VC and unconjugated bilirubin (BR) and a significant increase in the percentage (%CoQ10) of the oxidized form of coenzyme Q10 (CoQ10) to total coenzyme Q10 (TQ10) in centenarians, suggesting an increase of oxidative stress.

The plasma levels of high density lipoprotein (HDL) were reported to be decreased in centenarians.⁽²⁾ We, therefore, measured serum free cholesterol (FC) and cholesterol esters (CE) because their ratio (FC/CE) is determined by the activity of lecithin-cholesterol acyltransferase (LCAT) secreted with HDL from liver.^(9,10) We confirmed a significant decrease in CE and total cholesterol (TC) and a significant increase in FC/CE ratio in centenarians.

We also measured serum free fatty acids (FFA) and the content of oxidatively vulnerable polyunsaturated fatty acids in total FFA (%PUFA) as markers of tissue oxidative damage.⁽¹¹⁾ It is common that stearoyl-CoA desaturase is activated to compensate for the loss of PUFA; therefore, the percentages of palmitoleic acid and oleic acid in total FFA (%16:1 and %18:1, respectively) are also appropriate markers of tissue oxidative damage.⁽¹¹⁾

It is well known that human tissue levels of TQ10 decrease with age after the age of $20.^{(12)}$ For example, decreases in TQ10 of >30% and 50% in human heart were observed at ages 40 and 80, respectively.⁽¹²⁾ However, such a decline of TQ10 was not observed in human plasma within the range of 20 to $60.^{(13)}$ In this study, we found a significant decrease in serum TQ10 levels in centenarians as compared with 76-year-old controls. On the other hand, a significant increase of the coenzyme Q10 binding and transfer protein prosaposin (Psap)⁽¹⁴⁻¹⁶⁾ was observed. The possible role of Psap will be discussed.

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Materials and Methods

Study design. This study comprised 99 Japanese centenarians (25 males aged 100.8 ± 1.3 years and 74 females aged 100.9 ± 1.5 years) and 62 Japanese controls (25 males aged 74.6 ± 8.5 years and 37 females aged 76.2 ± 8.0 years). The above 4 groups are abbreviated as 101M, 101F, 75M and 76F, respectively. Written informed consent to participate was obtained either from the participants or a proxy when individuals lacked the capacity to consent. The protocol of this study was approved by the Ethical Committee of the Keio University School of Medicine. Non-fasting venous blood was sampled. Serum was collected and stored at -80° C until analysis. We also compared with previously reported 60-year-old controls⁽¹⁷⁾ (consisting of 38 males and 17 females, 60.1 ± 9.3 years).

Analytical procedures. Serum levels of VE, CoQ10H₂, CoQ10, FC and CE were determined as previously described with some modifications.⁽¹⁸⁾ In brief, serum was extracted with 19 volumes of 2-propanol and the extract was analyzed by HPLC using two analytical columns (Supelcosil ABZ+, 3 μ m, 3.3 cm × 4.6 mm i.d. and Ascentis LC-8, 5 μ m, 25 cm × 4.6 mm i.d.; Supelco Japan, Tokyo, Japan) connected in tandem, a reduction column (RC-10-1; Irica, Kyoto, Japan) and an amperometric electrochemical detector (Model Σ 985; Irica) with an oxidation potential of +600 mV (vs Ag/AgCl) on a glass carbon electrode. The mobile phase consisted of 50 mM sodium perchlorate in methanol/2-propanol (78/22, v/v), delivered at a flow rate of 0.8 ml/min. The analytical columns were cooled to 25°C.

Serum levels of VC, UA and BR were determined by HPLC on a bonded-phase aminopropylsilyl column (Supelcosil LC-NH₂, $5 \,\mu$ m, $25 \,\text{cm} \times 4.6 \,\text{mm}$ i.d.; Supelco Japan) with UV/VIS detection (265 nm for 0–15 min and 460 nm for 15–22 min), as described previously.⁽¹⁹⁾

Serum FFA were derivatized with monodansylcadaverine for analysis by HPLC.⁽²⁰⁾ Briefly, serum samples (50 µl) were mixed with 200 µl of methanol and then centrifuged at $13,000 \times g$ for 5 min. Aliquots (50 µl) of supernatants were mixed with 20 µl of methanol containing 25 µM tridecanoic acid (internal standard) and dried under a stream of nitrogen gas, and the residue was admixed with diethyl phosphorocyanidate (1 µl) and N,Ndimethylformamide (50 µl) containing monodansylcadaverine (2 mg/ml) and kept at room temperature in the dark for 20 min. A 5-µl sample was injected onto an octadecylsilyl column (3 µm, 3.3 cm × 4.6 mm i.d.; Supelco Japan) and a pKb-100 column $(5 \,\mu\text{m}, 25 \,\text{cm} \times 4.6 \,\text{mm i.d.}; \text{Supelco Japan})$ connected in tandem. The FFA components were measured by fluorescence detection (Model 821-FP; Japan Spectroscopic, Tokyo, Japan) with excitation at 320 nm and emission at 520 nm. The mobile phase consisted of acetonitrile/methanol/water (17.5/65.0/17.5, v/v/v)delivered at a flow rate of 1.5 ml/min. The analytical columns were heated to 40°C.

Serum levels of Psap were measured by a sandwich ELISA using monoclonal and polyclonal antibodies against human saposin B.⁽¹⁴⁾ Plasma was diluted 100 times with a phosphate-buffer saline containing 0.1% Triton X-100, 1 g/L NaN₃, 10 g/L BSA, and 1 mM EDTA. Purified saposin B was used as a standard.⁽¹⁴⁾

Statistical analysis. Data are presented as mean values with standard deviations. Statistical analysis was performed using one-way ANOVA followed by the Scheffe's multiple comparisons test. P<0.05 was considered statistically significant.

Results and Discussion

Serum antioxidants and oxidative stress. Figure 1 shows serum %CoQ10 and serum levels of VC, BR, and UA among male and female centenarians (101M and 101F, respectively) and 75-year-old male and 76-year-old female controls (75M and 76F, respectively). There were no significant differences between the

male and female groups in each age category. However, a significant increase in %CoQ10 was observed in centenarians compared with 76-year-old controls, indicating that the redox balance of coenzyme Q10 shifted to the oxidized form. This confirms an increase in oxidative stress in centenarians and agrees with a significant decrease in serum antioxidants such VC⁽¹¹⁾ and BR⁽²¹⁾ in centenarians. These results are also consistent with the observation that chronic inflammation is present in centenarians.⁽²⁾

Under acute inflammatory conditions like sepsis, the substantial formation of superoxide and nitric oxide, and their product peroxynitrite, is expected. In fact plasma UA levels declined significantly in patients with sepsis during a stay at an intensive care unit⁽⁸⁾ because UA is a good inhibitor of peroxynitrite.⁽²²⁻²⁴⁾ However, serum UA levels remained constant in centenarians and 76-year-old controls, suggesting that inflammation in centenarians is moderate and chronic.

Since there were no significant differences in %CoQ10, VC, BR, and UA between the male and female groups of centenarians and 76-year-old controls, the data were combined into centenarians and 76-year-old controls (abbreviated as 101 and 76) and compared with 60-year-old controls⁽¹⁷⁾ as shown in Fig. 2. %CoQ10 consistently increased with age, while levels of VC and BR in centenarians were significantly lower than 60- or 76-year-old controls. These data confirm that centenarians are under oxidative stress. However, levels of UA remained unchanged, suggesting that the formation of peroxynitrite is not very significant in centenarians and they are under moderate, chronic inflammatory conditions.

Serum levels of cholesterols. Figure 3 shows serum levels of FC, CE and TC, as well as the FC/CE ratio. A slight decrease in FC was observed in centenarians, however the difference was not significant. A significant decrease in CE and TC was observed, resulting in an increase in the FC/CE ratio. There were no significant differences in the levels of FC, CE and TC, and the FC/CE ratio between male and female groups in each age category. Thus, we plotted the pooled data against age (Fig. 4). FC, CE and TC were all observed to decrease with age. Since the decline of CE was more profound than FC, the FC/CE ratio increased with age. The FC/CE ratio is determined by the activity of LCAT secreted with HDL from liver.^(9,10) Therefore, these data indicate a degree of impairment in the secretion of LCAT with HDL and liver function. This is consistent with the previous observation that serum levels of HDL are low in centenarians.⁽²⁾

Serum FFA composition and tissue oxidative damage. Figure 5 shows serum levels of FFA, %PUFA, %18:1 and %16:1. A significant decrease in FFA was observed in centenarians. Under oxidative stress, plasma levels of FFA have been observed to increase in many cases, such as in newborn babies⁽²⁵⁾ and patients with hepatitis,⁽¹¹⁾ cirrhosis,⁽¹¹⁾ hepatoma, ⁽¹¹⁾ juvenile fibromyalgia,⁽²⁶⁾ and post-cardiac arrest syndrome.⁽²⁷⁾ Elevated plasma FFA was observed in the rat middle cerebral artery occlusion model of stroke.⁽²⁸⁾ It is of interest that repeated administration of the antioxidant edaravone significantly improved the neurological symptoms and impairment of motor function induced by a middle cerebral artery occlusion, and reduced the levels of FFA to those of a sham operation.⁽²⁸⁾ Formation of FFA under oxidative stress is assumed to be a result of phospholipase activity, (29-32) therefore, a significant decrease in serum FFA in centenarians must be ascribed to impairment of the repair system that counteracts increases in oxidative stress.

Under conditions of elevated oxidative stress, oxidatively vulnerable PUFA is selectively damaged which results in decreased membrane fluidity.⁽¹¹⁾ To compensate for the loss of PUFA, stearoyl-CoA desaturase is activated and converts stearic and palmitic acids to 18:1 and 16:1, respectively.⁽³³⁾ Accordingly, a decrease in %PUFA and an increase in %18:1 and %16:1 have been observed under oxidative stress.^(11,25-28)

Since there were no significant differences in FFA levels,



Fig. 1. Comparison of the percentage of oxidized coenzyme Q10 to total coenzyme Q10 (%CoQ10) in serum, and serum levels of ascorbic acid (VC), unconjugated bilirubin (BR), and uric acid (UA) among male centenarians (101M), male 75-year-old controls (75M), female 76-year-old controls (76M), and female centenarians (101F). Data are presented as mean + SD. *p<0.01 and **p<0.001, significant differences as determined by Scheffe's multiple comparison test.



Fig. 2. Changes in the percentage of oxidized coenzyme Q10 to total coenzyme Q10 (%CoQ10) in serum, and serum levels of ascorbic acid (VC), unconjugated bilirubin (BR), and uric acid (UA) with age. Data are presented as mean \pm SD. *p<0.05 and **p<0.001, significant differences as determined by Scheffe's multiple comparison test.



Fig. 3. Comparison of serum free cholesterol (FC), cholesterol esters (CE), total cholesterol (TC), and the FC/CE ratio among male centenarians (101M), male 75-year-old controls (75M), female 76-year-old controls (76M), and female centenarians (101F). Data are presented as mean + SD. *p<0.01 and **p<0.001, significant differences as determined by Scheffe's multiple comparison test.



Fig. 4. Changes in serum free cholesterol (FC), cholesterol esters (CE), total cholesterol (TC), and the FC/CE ratio with age. Data are presented as mean \pm SD. *p<0.01 and **p<0.001, significant differences as determined by Scheffe's multiple comparison test.



Fig. 5. Comparison of serum free fatty acids (FFA), the percentage of polyunsaturated fatty acids in total FFA (%PUFA), the percentage of palmitoleic acid in total FFA (%16:1), and the percentage of oleic acid in total FFA (%18:1) among male centenarians (101M), male 75-year-old controls (75M), female 76-year-old controls (76M), and female centenarians (101F). Data are presented as mean + SD. *p<0.001, significant differences as determined by Scheffe's multiple comparison test.



Fig. 6. Changes in serum free fatty acids (FFA), the percentage of polyunsaturated fatty acids in total FFA (%PUFA), the percentage of palmitoleic acid in total FFA (%16:1), and the percentage of oleic acid in total FFA (%18:1) with age. Data are presented as mean \pm SD. **p*<0.05 and ***p*<0.001, significant differences as determined by Scheffe's multiple comparison test.



Fig. 7. Comparison of serum total coenzyme Q10 (TQ10), the ratio of vitamin E to total cholesterol (VE/TC), the ratio of TQ10/TC, and serum prosaposin (Psap) among male centenarians (101M), male 75-year-old controls (75M), female 76-year-old controls (76M), and female centenarians (101F). Data are presented as mean + SD. *p<0.01 and **p<0.001, significant differences as determined by Scheffe's multiple comparison test.

%PUFA, %18:1, and %16:1 between the male and female groups in each age category, we plotted the pooled data against age (Fig. 6). No significant changes in %PUFA were observed in centenarians. In contrast, significant decreases in %18:1 and %16:1 were observed in centenarians, indicating impairment in the oxidative repair system. However, this hypothesis should be investigated further.

Serum TQ10 and Psap. Figure 7 shows serum levels of TQ10 and Psap, as well as the ratio of VE/TC and TQ10/TC in centenarians and 76-year-old controls. A significant decrease in TQ10 was observed in centenarians compared with 76-year-old controls, suggesting a coenzyme Q10 deficiency in centenarians. This is also the case in male centenarians even if TQ10 was normalized to TC, however, female centenarians were not significantly different. A similar trend was observed in VE/TC values.

On the other hand, a significant increase in coenzyme Q10 binding and transfer protein (Psap)(14-16) was observed in female centenarians compared with 76-year-old female controls, and male centenarians showed a similar tendency. Figure 8 shows the combined male and female data of 60- and 76-year-old controls, and centenarians. Psap levels increased progressively and significantly with age while TQ10 levels and the TQ10/TC ratio reached a maximum at 76 years and subsequently decreased. It is well known that tissue TQ10 levels decrease with age; for example >30% and 50% decreases in TQ10 were observed at the ages of 40 and 80, respectively, in human heart.⁽¹²⁾ Furthermore, the rate of coenzyme Q biosynthesis in rat heart is much less than that in rat kidney.⁽³⁴⁾ This should be also the case in human. These observations suggest that the human heart in octogenarians has a serious requirement for exogenous TQ10. Therefore, coenzyme Q10 should be transferred from its pool (most likely to be kidney) to heart using Psap. This is a likely explanation for the observed increase in serum Psap levels in 76-year-olds, and the consequent increase in serum TQ10 levels, compared with 60-year-old controls.

In centenarians, it is reasonable for Psap levels to increase in order to compensate for the loss of coenzyme Q10. Despite further elevation of Psap in centenarians, their TQ10 levels were decreased, indicating their tissue TQ10 levels were likely to be critically low.

Coenzyme Q10 is essential for ATP production in the mitochondria and is an important antioxidant in every cellular membrane and lipoprotein.⁽³⁵⁾ Its importance is also suggested by the observation that serum coenzyme Q10 levels were inversely associated with the risk of disabling dementia.⁽³⁶⁾ Furthermore, a mutation in the coenzyme Q10 synthesis enzyme was identified in patients with familial multiple system atrophy (MSA).⁽³⁷⁾ Plasma levels of coenzyme Q10 in patients with MSA were significantly lower than controls.^(38,39) Now the phase 2 clinical study of coenzyme Q10 supplementation to patients with MSA is on going. Therefore, coenzyme Q10 supplementation would be beneficial for centenarians, although this hypothesis requires further serious investigation.

Conclusion

Oxidative stress in centenarians was demonstrated as an increase in serum %CoQ10 and a decrease in VC compared with 76-year-old controls. Centenarians are suggested to exist in a moderate, chronic inflammatory condition because serum levels of UA were similar to those in 76-year-old controls. Centenarians exhibit a hypocholesterolemic condition and the observed increase in the FC/CE ratio suggests some impairment of liver function. A significant decrease in serum FFA, %18:1 and %16:1 also indicates impairment of the tissue repair system in centenarians. Despite an elevation of the coenzyme Q10 binding protein Psap, serum TQ10 levels decreased in centenarians, suggesting a serious TQ10 deficiency in tissues. Therefore, coenzyme Q10 supplementation is likely to be beneficial for centenarians.



Fig. 8. Changes in serum total coenzyme Q10 (TQ10), the ratio of vitamin E to total cholesterol (VE/TC), the ratio of TQ10/TC, and serum prosaposin (Psap) with age. Data are presented as mean \pm SD. *p<0.05, **p<0.01 and ***p<0.001, significant differences as determined by Scheffe's multiple comparison test.

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Abbreviations

percentage of palmitoleic acid in total FFA
percentage of oleic acid in total FFA
percentage of oxidized form of coenzyme Q10 in TQ10
percentage of polyunsaturated fatty acids in total FFA
unconjugated bilirubin
cholesterol esters
oxidized form of coenzyme Q10
ubiquinol-10, reduced form of coenzyme Q10
free cholesterol
free fatty acids

References

- Arai Y, Sasaki T, Hirose N. Demographic, phenotypic, and genetic characteristics of centenarians in Okinawa and Honshu, Japan. Part 2 Honshu, Japan. *Mech Ageing Dev* 2017; 165(Pt B): 80–85.
- 2 Arai Y, Martin-Ruiz CM, Takayama M, et al. Inflammation, but not telomere length, predicts successful ageing at extreme old age: a longitudinal study of semi-supercentenarians. *EBioMedicine* 2015; 2: 1549–1558.
- 3 Franceschi C, Capri M, Monti D, et al. Inflammaging and anti-inflammaging: a systemic perspective on aging and longevity emerged from studies in humans. Mech Ageing Dev 2007; 128: 92–105.
- 4 Tchkonia T, Zhu Y, van Deursen J, Campisi J, Kirkland JL. Cellular senes-

HDL LCAT MSA Psap TC TQ10 UA VC	high density lipoprotein lecithin-cholesterol acyltransferase multiple system atrophy prosaposin total cholesterol total coenzyme Q10 uric acid vitamin C
VC	vitamin C
VE	vitamin E

Conflict of Interest

We have not received any financial support or other benefits from commercial sources for the work reported in the manuscript. None of the authors have financial interests that could create a potential conflict of interest or the appearance of a conflict of interest with regard to this work.

cence and the senescent secretory phenotype: therapeutic opportunities. *J Clin Invest* 2013; **123**: 966–972.

- 5 Berg RM, Moller K, Bailey DM. Neuro-oxidative-nitrosative stress in sepsis. J Cereb Blood Flow Metab 2011; 31: 1532–1544.
- 6 Galley HF. Oxidative stress and mitochondrial dysfunction in sepsis. Br J Anaestesia 2011; 107: 57–64.
- 7 Schmoch T, Uhle F, Siegler BH, et al. The glyoxalase system and methylglyoxal-derived carbonyl stress in sepsis: Glycotoxic aspects of sepsis pathophysiology. Int J Mol Sci 2017; 18. pii: E657.
- 8 Yamaguchi J, Nagase M, Yamamoto Y, et al. Increased oxidative stress and

renal injury in patients with sepsis. J Clin Biochem Nutr 2018; 63: 137-143.

- 9 Florén CH, Chen CH, Franzén J, Albers JJ. Lecithin: cholesterol acyltransferase in liver disease. *Scand J Clin Lab Invest* 1987; **47**: 613–617.
- 10 Yamamoto Y, Yamashita S, Fujisawa A, Kokura S, Yoshikawa T. Oxidative stress in patients with hepatitis, cirrhosis, and hepatoma evaluated by plasma antioxidants. *Biochem Biophys Res Commun* 1998; 247: 166–170.
- 11 Yamamoto Y. Plasma marker of tissue oxidative damage and edaravone as a scavenger drug against peroxyl radicals and peroxynitrite. *J Clin Biochem Nutr* 2017; **60**: 49–54.
- 12 Kalén A, Appelkvist EL, Dallner G. Age-related changes in the lipidcompositions of rat and human tissues. *Lipids* 1989; 24: 579–584.
- 13 Wada H, Goto H, Hagiwara S, Yamamoto Y. Redox status of coenzyme Q10 is associated to chronological age. J Am Geriatrics Soc 2007; 55: 1141–1142.
- 14 Jin G, Kubo H, Kashiba M, et al. Saposin B is a human coenzyme Q10binding/transfer protein. J Clin Biochem Nutr 2008; 42: 167–174.
- 15 Kashiba M, Oizumi M, Suzuki M, et al. Prosaposin regulates coenzyme Q10 levels in HepG2 cells, especially those in mitochondria. J Clin Biochem Nutr 2014; 55: 85–89.
- 16 Kashiba M, Terashima M, Sagawa T, Yoshimura T, Yamamoto Y. Prosaposin knockdown in Caco-2 cells decreases cellular levels of coenzyme Q10 and ATP, and results in the loss of tight junction barriers. *J Clin Biochem Nutr* 2017; 60: 81–85.
- 17 Nagase M, Yamamoto Y, Miyazaki Y, Yoshino H. Increased oxidative stress in patients with amyotrophic lateral sclerosis and the effect of edaravone administration. *Redox Rep* 2016; 21: 104–112.
- 18 Yamashita S, Yamamoto Y. Simultaneous detection of ubiquinol and ubiquinone in human plasma as a marker of oxidative stress. *Anal Biochem* 1997; 250: 66–73.
- 19 Yamamoto Y, Ames BN. Detection of lipid hydroperoxides and hydrogen peroxide at picomole levels by an HPLC and isoluminol chemiluminescence assay. *Free Radic Biol Med* 1987; 3: 359–361.
- 20 Yamamoto Y, Nagata Y, Katsurada M, Sato S, Ohori Y. Changes in rat plasma-free fatty acid composition under oxidative stress induced by carbon tetrachloride: decrease of polyunsaturated fatty acids and increase of palmitoleic acid. *Redox Rep* 1996; 2: 121–125.
- 21 Stocker R, Yamamoto Y, McDonagh AF, Glazer AN, Ames BN. Bilirubin is an antioxidant of possible physiological importance. *Science* 1987; 235: 1043–1046.
- 22 Santos CX, Anjos EI, Augusto O. Uric acid oxidation by peroxynitrite: multiple reactions, free radical formation, and amplification of lipid oxidation. *Arch Biochem Biophys* 1999; **372**: 285–294.
- 23 Tsukada K, Hasegawa T, Tsutsumi S, et al. Effect of uric acid on liver injury during hemorrhagic shock. Surgery 2000; 127: 439–446.

- 24 Scott GS, Cuzzocrea S, Genovese T, Koprowski H, Hooper DC. Uric acid protects against secondary damage after spinal cord injury. *Proc Natl Acad Sci U S A* 2005; **102**: 3483–3488.
- 25 Hara K, Yamashita S, Fujisawa A, Ishiwa S, Ogawa T, Yamamoto Y. Oxidative stress in newborn infants with and without asphyxia as measured by plasma antioxidants and free fatty acids. *Biochem Biophys Res Commun* 1999; 257: 244–248.
- 26 Miyamae T, Seki M, Naga T, et al. Increased oxidative stress and coenzyme Q10 deficiency in juvenile fibromyalgia: amelioration of hypercholesterolemia and fatigue by ubiquinol-10 supplementation. *Redox Rep* 2013; 18: 12–19.
- 27 Nagase M, Sakurai A, Sugita A, et al. Oxidative stress and abnormal cholesterol metabolism in patients with post-cardiac arrest syndrome. J Clin Biochem Nutr 2017; 61: 108–117.
- 28 Yamamoto Y, Yanagisawa M, Tak NW, *et al.* Repeated edaravone treatment reduces oxidative cell damage in rat brain induced by middle cerebral artery occlusion. *Redox Rep* 2009; 14: 251–258.
- 29 Yasuda M, Fujita T. Effect of lipid peroxidation on phospholipase A₂ activity of rat liver mitochondria. Jpn J Pharmacol 1977; 27: 429–435.
- 30 Weglicki WB, Dickens BF, Mak IT. Enhanced lysosomal phospholipid degradation and lysophospholipid production due to free radicals. *Biochem Biophys Res Commun* 1984; 124: 229–235.
- 31 Beckman JK, Borowitz SM, Burr IM. The role of phospholipase A activity in rat liver microsomal lipid peroxidation. J Biol Chem 1987; 262: 1479–1481.
- 32 Gutteridge JMC, Quinlan GJ, Yamamoto Y. Hypothesis: are fatty acid patterns characteristic of essential fatty acid deficiency indicative of oxidative stress? *Free Radic Res* 1998; 28: 109–114.
- 33 Ntambi JM. Regulation of stearoyl-CoA desaturase by polyunsaturated fatty acids and cholesterol. *J Lipid Res* 1999; 40: 1549–1558.
- 34 Elmberger PG, Kalèn A, Appelkvist EL, Dallner G. In vitro and in vivo synthesis of dolichol and other main mevalonate products in various organs of the rat. Eur J Biochem 1987; 168: 1–11.
- 35 Turunen M, Olsson J, Dallner G. Metabolism and function of coenzyme Q. Biochim Biophys Acta 2004; 1660: 171–199.
- 36 Yamagishi K, Ikeda A, Moriyama Y, et al. Serum coenzyme Q10 and risk of disabling dementia: the Circulatory Risk in Communities Study (CIRCS). *Atherosclerosis* 2014; 237: 400–403.
- 37 Multiple-System Atrophy Research Collaboration. Mutations in COQ2 in familial and sporadic multiple-system atrophy. *New Engl J Med* 2013; 369: 233–244.
- 38 Kasai T, Tokuda T, Ohmichi T, et al. Serum levels of coenzyme Q10 in patients with multiple system atrophy. PLoS One 2016; 11: e0147574.
- 39 Mitsui J, Tsuji S. Plasma Coenzyme Q10 levels and multiple system atrophyreply. JAMA Neurol 2016; 73: 1499–1500.