



Current and Emerging Therapy for Malignant Pleural Mesothelioma: Focus on CD26/Dipeptidyl Peptidase IV as a Therapeutic Target



Bently P. Doonan¹, Kei Ohnuma², Long H. Dang¹, Chikao Morimoto² and Nam H. Dang^{1,*}

¹University of Florida, Gainesville, FL, USA; ²Juntendo University, Tokyo, Japan

Abstract: Background: Malignant mesothelioma is a largely incurable disease that is refractory to current therapies. CD26 is a multifunctional cell surface protein involved in autoimmune disease, diabetes, and cancer. It has a role in T cell function, extracellular protein modification, as a prognostic factor for cancer, and as a therapeutic target for malignant mesothelioma. New treatment strategies are urgently needed for malignant pleural mesothelioma (MPM), and CD26-targeted therapy represents a novel approach.

Outline: In this review, the most current and up-to-date literature available was reviewed and the current state of malignant mesothelioma treatment is described. Throughout the review the need for new therapeutic approaches is highlighted in the shortcomings of current therapy. CD26 is a target that is fit to take on these shortcomings. In this review we discuss the structure and function of CD26, its role in malignant mesothelioma and the future of anti-CD26 therapy as a versatile immunotherapeutic option.

Conclusion: This review highlights the areas of most promise in treating MPM, these include immune checkpoint blockade, passive immunization, and based on our recently published data, targeting of CD26 with its specific mAb. Finally we describe how the anti-CD26 mAb YS110 was recently evaluated in the first-in-human phase I clinical trial, showing prolonged disease stabilization and a favorable side effect profile. Through better understanding of CD26, new pathways to treating and potentially curing malignant mesothelioma may be discovered.

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1. INTRODUCTION

The human leukocyte surface antigen CD26 is an active cell surface peptidase that is structurally identical to dipeptidyl peptidase IV (DPPIV), able to cleave N-terminal dipeptides from peptides with

terminal L-alanine or L-proline residues (1-4). It is composed of 766 amino acids, the majority of which comprise the extracellular domain of the protein where a peptidase catalytic site is found and where important ligand binding sites for adenosine deaminase (ADA) and fibronectin are located [1-4]. The remainder of the protein structure includes a short 6-peptide cytoplasmic domain and a 23-peptide transmembrane region [4]. Through this peptidase activity CD26/DPPIV has significant effects on enhancing cellular response to external stimuli, effects on glucose homeostasis,

*Address correspondence to this author at the Division of Hematology/Oncology, University of Florida, P.O. Box 100278, Gainesville FL 326100278, USA; Tel: (352) 273-7832; Fax: (352) 273-5006; E-mail: nam.dang@medicine.ufl.edu

T cell stimulation and activation, and the biological behavior of selected human neoplasms. CD26 has relatively widespread expression on leukocytes, fibroblasts, mesothelium, endothelial, epithelial cells, and can be found in kidney, intestine, prostate, pancreas, and liver cells [5]. Since its discovery in 1966 by Hopsu-Havu and Glenner, CD26/DPPIV has been the focus of vigorous study in its pluripotent role in glucose homeostasis, inflammation, and more recently in tumorigenesis and as a therapeutic target in cancer [6]. The various immunomodulatory effects of CD26 have been previously summarized by our group and recently revisited and expanded by Klemann *et al.* [7, 8]. These works summarize the numerous substrates for DPPIV/CD26 and their far-reaching roles in autoimmune diseases such as multiple sclerosis, asthma, arthritis, and inflammatory bowel disease [7-9]. Likewise, CD26 involvement in malignancy has been extensively reviewed and characterized, including its potential role in terms of its role as a tumor suppressor, cancer biomarker, and therapeutic target [6, 10-14]. Additionally CD26 has been described as a marker for so-called cancer stem cells (CSCs) which have been a highly sought after targets in chemotherapeutic approaches [15]. Given the preponderance of evidence for CD26 involvement in various malignancies, as well as its role in immune activation and the biology of cancer stem cells, CD26 represents an ideal immunotherapeutic target; including for the aggressive, almost always fatal cancer malignant pleural mesothelioma (MPM).

Malignant pleural mesothelioma (MPM) is an aggressive and fatal disease. Over the past 60 years, since its acceptance as an independent oncological process, the incidence of MPM has continued to rise [16]. MPM is almost exclusively a direct result of exposure to asbestos [17]. Chronic pleural inflammation, ionizing radiation, and SV40 virus have been proposed as alternative exposures that can result in MPM, but these account for less than 20% of all cases [17]. Asbestos, a term for naturally occurring families of minerals that separate into thin fibers, has been used for greater than 5000 years for its high tensile strength and fire resistant properties [18]. It wasn't until the 1960s that the direct correlation between asbestos exposure and cancer development was validated and accepted [19]. Since that time, the WHO and International Agency for research in cancer have

defined asbestos as a class I carcinogen responsible for both lung cancer and malignant pleural mesothelioma [18]. Inhaled asbestos fibers end up in the pleura, induce cytotoxic effects, and cause DNA damage and chronic inflammation [20]. This process is constant and smoldering for the next 20-60 years prior to the development of MPM. This long period of latency and protracted asymptomatic period explains the delayed peak in MPM cases and the increasing incidence over the past 40 years [21]. For example, in the US the peak in asbestos consumption occurred in the early 1970s, and its manufacturing was banned in the late 1980s, with total consumption and exposure risk being significantly reduced by the late 1990s; but the peak in MPM diagnoses of roughly 2,500-3,000 cases did not occur until about 2002 [22]. The expected plateau effect for MPM diagnoses for most industrialized nations that have banned the use of asbestos are expected to occur between 2015 and 2030, but countries like Russia, China, Brazil, and India continue to both mine and use asbestos at an alarming rate [23]. China has become the world's largest asbestos-consuming country, has little to no reporting mechanism of its MPM rates, and will likely experience a surge in MPM diagnoses in the future [24]. With this predictable man-made epidemic looming on the horizon, new strategies are required for treating this aggressive disease as current strategies show limited efficacy, poor survival benefit, and have significant morbidity associated with them [18]. In this review, we will discuss the current state of malignant mesothelioma treatment and some of the burgeoning therapies currently in clinical trials. We will also highlight the work done on CD26 expression in MPM, its potential as a biomarker, and its functional role in MPM survival, invasion, and migration. Finally, we will review the ongoing clinical development of an anti-CD26 monoclonal antibody in malignant mesothelioma and its potential far-reaching implications as a novel immunotherapeutic agent.

2. CURRENT THERAPIES

If left untreated, MPM has an average life expectancy of 8 months and a 5-year mortality of greater than 95% [24]. Our best efforts with multimodal therapy may extend this outcome by mere months, further emphasizing the extreme need for improved therapies. Current therapeutic strategies

Table 1. Principal characteristics and benefit of current therapies for MPM.

First Line Therapy	Overall Survival (Months)	Improvement vs. SOC	Study Features/ Limitations	References
None	8	n/a	Na/	Zhang <i>et al.</i> Ann Transl Med 2015
Cisplatin + pemetrexed (SOC)	16.1	0	225 pts, newly diagnosed MPM, ECOG 0-2	Zalcman <i>et al.</i> Lancet 2016
SOC + bevacizumab	18.8	2.7	223 pts, newly diagnosed MPM, ECOG 0-2	Zalcman <i>et al.</i> Lancet 2016
SOC + EPP	21.9	5.8	54 pts, stage I to III MPM, otherwise healthy, ECOG 0-1	Krug <i>et al.</i> J clin oncol 2009
EPP + IMRT	14.2	None	63 pts, able to tolerate EPP, minimal comorbidities	Rice <i>et al.</i> Ann thorac surg 2007
EPP + SOC + hemithoracic RT	29.1	13	42 pts, ECOG 0-1, T1-3 N0-2 (33 pts who entered study could not tolerate all phases of therapy)	Krug <i>et al.</i> J clin oncol 2009
PD + SOC + IMPRINT	20.2	4.4	70 pts, retrospective study over 30 years, high karnofsky score	Shaikh <i>et al.</i> J of thoracic oncol 2017
CRS-207 + SOC	8.5*	N/A	38 pts, treatment naïve, ECOG 0-1	Jahan <i>et al.</i> J of thoracic oncol 2016

Second Line Therapy	Overall Survival (Months)	Improvement vs. Historic Chemo	Study Features/ Limitations	Reference
Tremelimumab + SOC	10.7	2.0	29 pts, ECOG 0-1, primarily epithelioid histology	Calabro <i>et al.</i> Lancet Respir Med 2015 MESOT-TREM-2012

Abbreviations: MPM (Malignant Pleural Mesothelioma), ECOG (Eastern Cooperative Oncology Group Performance Status), SOC (Standard of Care), pts (patients), EPP (Extrapleural Pneumonectomy), PD (Pleurectomy with Decortication), IMRT (Intensity-modulated Radiation Therapy), CRS-207 (live, attenuated, double-deleted listeria monocytogenes engineered to express tumor-associated antigen mesothelin). * (progression free survival, overall survival goal not met to date).

for MPM include surgery, radiation, chemotherapy, and more recently targeted therapy and immunotherapy [25-31]. Table 1 summarizes the mainstays of treatment and the benefit of traditional therapies over systemic chemotherapy. The roles of surgery and radiation as part of MPM treatment are rife with controversy, with these modalities showing limited benefit in patients with advanced disease.

2.1. Multimodal (Surgical Resection, Radiotherapy, and Chemotherapy)

Surgical intervention is difficult to perform, since achieving negative margins when extracting

thin areas of pleura is difficult to accomplish and is associated with a significant level of risk, requiring a high level of familiarity and expertise with the procedure [32]. Two main surgical interventions are currently in use, pleurectomy/decortication (P/D) and Extrapleural Pneumonectomy (EPP) [29]. As illustrated in the MARS trial, EPP, an invasive aggressive debulking procedure showed no benefit when added to chemotherapy/radiation, being associated with a worse median survival when compared with no surgical intervention, and may actually cause harm [33, 34]. However, it is important to note that this study, like many involving MPM, is extremely underpowered and involved only 19 individuals re-

ceiving EPP [34]. With the high morbidity and required technical expertise, this procedure has therefore fallen out of favor except in those patient fortunate enough to be diagnosed at an early-localized stage.

P/D is being evaluated currently in the MARS2 trial to evaluate the role of surgery in MPM therapy, aside from its value in diagnostic biopsy [32]. In a similar vein, radiotherapy has been used in combination with chemotherapy and surgery as part of a trimodal therapeutic approach, or following surgery to prevent tumor seeding of thoracoscopy or thoracotomy scars. Trimodal therapy involves chemotherapy followed by EPP or P/D and intensity modulated radiotherapy as combination treatment for MPM. Results from small trimodal clinical trials suggest that, in certain patient populations, this aggressive multipronged attack may improve overall survival by up to 6 months vs. standard of care (SOC) [35]. However, this treatment option is only valuable to those MPM patients with limited burden of disease, excellent performance status, and epithelioid histology [36]. While surgery and radiotherapy may have some role to play in specific MPM cases, the mainstay of treatment for the past 20 years has been systemic chemotherapy.

2.2. Systemic Chemotherapy in Malignant Mesothelioma (SOC)

For a prolonged period of time, there was no consensus as to the optimum systemic chemotherapy used for MPM due to the limited randomized clinical trial data to support one strategy over another. It is important to mention that systemic chemotherapy for MPM is palliative in nature and has been the only intervention to show modest improvement in overall survival [27, 37]. This situation changed in 2003 following the availability of the results of the EMPHACIS phase III trial, which showed the superiority of the combination of cisplatin and pemetrexed over cisplatin alone, which was the most commonly used first line chemotherapy at the time [38, 39]. In this study, median survival was improved from 9.3 to 12.1 months [39]. These data led to the formal approval of the pemetrexed/cisplatin combination as the new SOC for MPM, a development which has not changed in the decade plus since it was first described [37]. Additional studies have validated the use of a different antifolate, raltitrexed, with

cisplatin as an appropriate alternative regimen if pemetrexed is not well-tolerated [40]. Many clinicians also substitute carboplatin for cisplatin to reduce toxicity with little clinical difference in outcomes and without formal FDA approval [41]. Meanwhile, the recently published MAPS study demonstrated the clinical benefit of adding bevacizumab to SOC which resulted in an additional 2.7 month survival benefit though not without risks as the bevacizumab arm had higher reported adverse events across multiple subgroups and significant increases in grade 3-4 arterial and venous thromboembolic events [42]. However, even with this increase in median survival from the addition of bevacizumab plus SOC, MPM typically recurs as an incurable disease, necessitating the development of effective second line therapeutic options. Unlike the case with first line therapy, there is currently no established SOC therapy for disease recurrence or progression following initial management. The most common second line chemotherapy options include the vinca alkaloid vinorelbine, the anti-nucleoside analog gemcitabine, and the re-administration of single agent pemetrexed, which have shown the most promise in terms of tolerability but have failed to improve overall survival [27, 43]. In view of these shortcomings of currently available therapies, novel treatment strategies including targeted strategies and immunotherapy have been explored as both adjunctive and independent options for systemic therapy of MPM.

2.3. Targeted Therapies and Immunotherapy in Malignant Mesothelioma

Given the overwhelming lack of second line options in MPM and the large percentage of patients diagnosed with advanced disease that is not amenable to aggressive multimodal approaches, there has been a focus on targeted therapies with biological agents over the last 10 years, albeit with mostly disappointing results. Targeting various tyrosine kinases and the process of angiogenesis, as well as representing various forms of immunotherapy, these therapies can be broadly subcategorized into small molecule inhibitors, angiogenesis inhibitors, histone deacetylase (HDAC) inhibitors, and gene mutation targeting. The small molecule inhibitors include multitargeting receptor tyrosine kinase inhibitors (mTKIs), selective tyrosine kinase inhibitors (sTKIs), and

proteasome inhibitors. These molecules have garnered much focus in the oncologic world with broad applications in both solid and liquid tumors [44-47]. Unfortunately, phase I and phase II clinical trials involving the mTKIs sorafenib, sunitinib, pazopanib, and dasatinib showed either limited anti-mesothelioma activity, the inability to induce remission, and/or unacceptable toxicity [48-52]. These receptor tyrosine kinase inhibitors broadly target EGFR, VEGFR, PDGFR, and C-kit to exert their anticancer effects. While these self-signaling molecules are upregulated in MPM, they do not appear vital to its propagation and are not MPM specific, which likely contributes to the limited efficacy of these drugs. Interestingly, the process of angiogenesis has only been successfully targeted by the monoclonal antibody bevacizumab and not by the above mentioned mTKIs which target VEGFR, or by the biologic agent thalidomide which primarily works through angiogenesis inhibition [53, 54]. Thalidomide was tested in clinical trials as both adjunct to standard of care treatment and as maintenance therapy for MPM patients previously treated with platinum based chemotherapy. The results of these studies showed no benefit to thalidomide as adjunct and no improvement in overall survival (OS) *vs* supportive care alone as maintenance therapy [27, 54]. Guazelli *et al.*, recently summarized active phase I and phase II clinical trials and highlight that many studies have looked at targeting the EGFR or VEGFR pathway with little success to show for it up to this point, this review expertly highlights the current clinical trials that are ongoing from Clinicaltrials.gov [53]. Since EGFR expression is upregulated in the majority of MPM, selective tyrosine kinase inhibitors like erlotinib and gefitinib should theoretically exhibit increased activity against MPM. Unfortunately, similar to the mTKIs, results from phase II clinical trials were disappointing. Limited efficacy and marked resistance to these sTKIs was observed even in the presence of detectable EGFR expression on MPM tumors [55-57]. Similar results to these were seen with the proteasome inhibitor bortezomib, which has been approved for use in multiple myeloma and is currently in clinical trials for multiple other cancers including non-small cell lung cancer and metastatic breast cancer [58]. In two different clinical trials bortezomib failed to show objective response as monotherapy and failed to provide significant OS survival or

disease progression benefit when combined with SOC [59].

HDAC inhibitors, specifically vorinostat, which modify and limit deacetylation of histone and block access genes that are overused by cancer cells for progression and division, showed promising results in early clinical trials [60]. These results prompted the large-scale VANTAGE-014 phase III double blind, randomized placebo control study using vorinostat monotherapy as either second- or third-line therapy for MPM [61]. Results reported in the Lancet in 2015 of this large well designed and well executed study (660 patients enrolled) showed no benefit to vorinostat over placebo in terms of overall survival [61]. Gene mutations have been a hallmark of targeted cancer therapy, but few highly conserved gene mutations in MPM have been identified, and studies involving those found and targeted have failed to result in clinically significant efficacy. The most common mutations observed through molecular genetic analysis of patient MPM samples include BAP1, PTEN/PI3K, CDKN2A/ARF, and NF2 [62-65]. Of these, NF2 has been identified in 40% of MPM and results in inactivation of a protein called Merlin which is involved in cell adhesion and motility. Of potential therapeutic value is the fact that Merlin loss increases cell sensitivity to focal adhesion kinase (FAK) inhibitors [66]. Initial research showed FAK inhibitors, specifically defactinib, along with their MPM cytotoxic effect reduced so called cancer stem cell populations in MPM with potential for more durable prolonged response *vs* SOC [66]. These results prompted a large phase II COMMAND study which enrolled 372 patients to receive defactinib plus SOC *vs* placebo plus SOC control arm as first line therapy for MPM [67]. Unfortunately as is often the case in MPM clinical trials, the study was stopped during recruitment when no difference in defactinib *vs* placebo were observed, even when subdivided to those patients with identifiable merlin loss [68].

Immunotherapy has been on the forefront of cancer therapy for the past 20 years and recent successes in immune checkpoint inhibition, tumor escape mechanism targeting, passive immunotherapy, and dendritic cell vaccines have pushed the field further with ever broadening application [69-71]. Recent advances in immunotherapy and current clinical trials in MPM are well summarized by

Thapa *et al.*, [72]. In their review of current immunologic strategies, the failures of targeting MPM with single agent immunotherapy warrants the use of combination strategies to improve efficacy [72]. One of the more successful pathways of immunotherapy described in their work and others is the use of immune checkpoint inhibition as a novel target in MPM [72]. An example of immune checkpoint inhibition is the strategy of targeting and blocking CTLA-4. CTLA-4 is a cell surface co-factor expressed on the surface of T cells that acts as an inhibitory cofactor for CD80 and CD86 [73]. CTLA-4 competes with CD28 for binding with CD80/86 and when bound sends an inhibitor signal to antigen presenting cells to decrease the inflammatory response and diminish cell activation. This process allows CTLA-4 to protect surrounding cells and the system as a whole from uncontrolled immune stimulation [74]. Tumor cells, including malignant mesothelioma, express increased level of CTLA-4 as a means of blocking anti-tumor immune responses [74]. Inhibition of CTLA-4, or so-called checkpoint inhibition, therefore can restore the anti-tumor immune response, resulting in T cells recognition and attack of tumors that had previously been undetected. In a recent review, Guazelli *et al.* have summarized the role of CTLA-4 targeting in MPM [75]. They and others point out monoclonal antibodies (mAb) directed against CTLA-4 have shown impressive results in melanoma and have been tested in early clinical trials with MPM [75]. The CTLA-4 mAb Tremelimumab has been investigated in the MESOT-TREM-2008, and MESOT-TREM-2012 phase II clinical trials in patients with chemotherapy-resistant MPM [28, 75, 76]. In these studies, Tremelimumab treatment resulted in a disease control rate of 31% when administered every 3 months, and a control rate of 52% when given every 4 weeks, a regimen that led to improved efficacy in other cancers [75-77]. Furthermore, with the shorter dosing time, median OS was improved to 10.7 months compared to historical averages of 8.7 months with second line chemo [77]. These early results have led to an ongoing study comparing Tremelimumab monotherapy *vs* placebo control. Additionally, combination immunotherapies, CTLA-4 blockade combined with anti-PD-L1 therapy is under active investigation and was recently presented at ASCO 2016 with the combination of Tremelimumab and durvalumab

[75]. Like CTLA-4, programmed death ligand 1 (PD-L1) is overexpressed in MPM, particularly the sarcomatoid type, and exerts an inhibitory effect on T cells to suppress the anti-tumor immune response [77]. Binding PD-L1 with ligand specific mAbs therefore blocks this tumor escape mechanism, potentially resulting in greater susceptibility of MPM to immune destruction. In the recent KEYNOTE-028 phase I clinical trial, the PD-1 mAb pembrolizumab was tested in a 25 patient cohort and showed an impressive overall disease control rate of 76% [73, 78]. This has prompted larger phase II investigation to further explore the anti-MPM activity of pembrolizumab [73].

Additional passive immunotherapy strategies in MPM have focused on targeting the tumor-associated antigen Mesothelin. Mesothelin expression is increased in MPM and likely plays a role in cell adhesion and invasion [79, 80]. Strategies focusing on mesothelin targeting have involved the mAb Amatuximab, anti-mesothelin vaccine CRS-207, and the mAb-toxin fusion protein called SS1P [79]. Amatuximab is a chimeric anti-mesothelin mAb that was tested in a phase II multicenter trial of 89 MPM patients in combination with SOC (pemetrexed/cisplatin) *vs* SOC alone, with improved OS in the Amatuximab-containing arm [81]. The anti-mesothelin vaccine CRS-207 is a live, attenuated, derivative of *Listeria monocytogenes* that expressed the mesothelin Ag and activates both innate and adaptive immunity [82]. The synergistic effects of CRS-207 and SOC chemotherapy was tested in a phase I trial of 38 MPM patients which showed encouraging anti-tumor immunity, with a progression free survival of 8.5 months [82]. Encouraging results from this study has led to a large multicenter phase III clinical trial that is currently recruiting. Similarly, the recombinant anti-mesothelin and truncated *Pseudomonas* exotoxin SS1P has shown significant anti-tumor activity and tolerability in a phase I clinical trial [83]. Finally, adoptive transfer of *ex vivo* stimulated dendritic cells, so called dendritic cell vaccination has recently shown activity in MPM. In a trial of 10 patients with MPM, cyclophosphamide was given to reduce activity of T regulatory cells prior to infusion of dendritic cells pulsed with autologous tumor lysate [84]. Results of this small study showed radiographic disease control in 8/10 patients and an impressive OS of greater than 2

years in 7/10 patients [84]. This technique will likely be expanded and further investigated in future years.

3. POTENTIAL APPROACH TO TARGETING CD26 IN MPM

As discussed above, treatment for MPM beyond first line therapy is still unsatisfactory due to limited efficacy, and novel therapeutic approaches are urgently needed for this patient population. While immunotherapy approaches appear promising with encouraging efficacy in multiple small studies, the observed benefit is still relatively short lasting and typically limited to only a few months. Additionally some concerns that arise with all studies involving immunotherapy include utilizing progression free survival (PFS) vs OS as an overall marker of drug efficacy or combining drugs that lack single-agent activity. In a recent study published in the *Journal of Clinical Oncology*, Tan *et al.* performed a meta-analysis of trials with results posted on ClinicalTrials.gov where they found a majority of studies showing significant benefit on PFS but not on OS [85]. They highlight the need for studies to look at both PFS and OS and to not use one as a surrogate marker for the other [85]. In separate commentary, Gyawali and Prasad illustrate that multiple trials over the past decade where multiple drugs without proven single-agent efficacy were added together as a novel treatment and showed PFS but rarely showed an effect on OS [86]. They pose in additional manuscript that by combining multiple therapies to extent PFS we run the risk of subjecting patients to unnecessary side effects and risks for little benefit [87]. These compelling examples shed light on the grain of salt mentality that is needed when evaluating the efficacy of immunotherapy. Unfortunately, as is usually the case, most of these trials and analysis did not include studies involving MPM. Given the small treatment arms and aggressive nature of MPM small gains in PFS may indicate improvement in OS but the low power of these studies makes statistical extrapolation difficult and with such a short OS window for these patients even small gains should be looked at as possible treatment strategies. There studies confirm and support the need for improved targeting in diseases like MPM and stresses the importance of single-agent efficacy in therapy design before a drug is pushed forward through clinical trials and

towards FDA approval. Thus, an ideal target in MPM would be one that is highly expressed by malignant mesothelioma cells but absent from normal mesothelioma, and plays a role in tumor proliferation or invasiveness. In addition, therapy specifically targeting this antigen should work in concert with chemotherapy to enhance treatment efficacy without increasing side effect burden, have strong activity as a single agent, and improve both PFS and OS in MPM patients. All these characteristics can be found potentially in the surface antigen CD26.

3.1. CD26 Expression and Function in MPM

CD26 is a multifunctional cell surface receptor with roles in immune regulation, T cell activation, and malignant potential of various cancers. CD26 is highly expressed in MPM and was originally identified as a potential target by our group [88, 89]. Our initial work demonstrated that targeting CD26 with anti-CD26 mAb resulted in *in vitro* growth inhibition of MPM cell line [89]. Subsequently, we showed high CD26 expression on various human MPM types including localized MPM, well-differentiated papillary MPM, and diffuse MPM but not on adenomatoid tumors or reactive mesothelioma cells [88]. Complementing our earlier *in vitro* work, we also demonstrated for the first time the potential for targeting CD26 in MPM with anti-CD26 mAb in an *in vivo* NOD-SCID mouse model of MPM. This initial evaluation for CD26 expression on a small sample population of MPM tissues was then further expanded in a follow-up paper which demonstrated overexpression of CD26 in MPM in more than 120 different MPM surgical samples including epithelioid mesothelioma, biphasic mesothelioma, and sarcomatoid mesothelioma as well as 8 different mesothelioma cell lines [90]. Interestingly, the more aggressive sarcomatoid mesothelioma samples had reduced cell surface CD26 expression but retained cytoplasmic CD26 expression. This work suggested that the morphology of the mesothelioma cell type, which has been used as a prognostic factor in the disease, correlated with CD26 expression and that loss of membranous CD26 was associated with the more aggressive spindle shaped sarcomatoid mesothelioma, raising the possibility that the loss of CD26 in these cells represented an epithelial to mesenchymal transition for MPM potentially responsible for the difficulty in treating and poorer

outcome associated with sarcomatoid MPM [90]. We subsequently showed that CD26 surface expression was associated with improved survival in MPM patients that received chemotherapy and potentially contributed to mesothelioma chemosensitivity [91]. We also demonstrated through *in vitro* investigation that CD26 was associated with enhanced proliferative activity of MPM and through downstream gene activation, CD26 upregulated mechanisms that increased chemotherapy sensitivity [91]. Given the increased surface expression of CD26 in favorable MPM phenotypes, soluble CD26 (sCD26) was investigated as potential biomarker for MPM. We showed in a recent preclinical model of MPM that sCD26 levels were higher in patients with the favorable epithelial phenotype than those with the aggressive sarcomatoid subtype and through therapeutic monitoring, reduction in sCD26 may indicate progression of disease [92]. Whether sCD26 can be used in high-risk patients, those with previous significant asbestos exposure or those residents of asbestos mining communities, as a screening tool for early detection of disease remains to be determined. Additionally, CD26 expression was identified and characterized as a marker for so called cancer stem cells (CSC) in MPM and was found to be co-expressed with the CSC marker CD24 [93]. In this study we used shRNA knockdown of CD24 and CD26 to determine the effects of gene silencing in the MPM cell line Meso-1 [93]. We showed that through silencing of CD26 but not CD24 MPM cellular invasion was reduced and MPM showed reduced proliferation [93]. Gene knockout of CD26 additionally caused a reduction in the cancer cell signaling molecules IGFBP7, IGFBP3, Wnt5A, and IL7R [93]. These studies also showed CD26 played a role in asymmetric cell division and invasion potential in MPM cell lines [93]. These collective works clearly validated CD26 as a surface receptor upregulated in MPM with various roles in cell proliferation, invasion, and chemosensitivity, while loss of expression correlated with progression of MPM to a more aggressive treatment-resistant phenotype.

These studies on CD26 expression in MPM were subsequently followed by our work evaluating molecular mechanisms for CD26 role in cellular invasiveness. We showed that CD26 promotes invasiveness through the formation of CD26- $\alpha 5\beta 1$ integrin molecular complexes that interact with various cell surface and endothelium receptors and

stimulate extra-cellular matrix metalloproteinases [94]. In addition, the short 6 amino acid cytoplasmic region of CD26 plays a crucial role in MPM migration and invasion through upregulation of periostin [95]. We demonstrated that the lipid raft platforms clustered around CD26, indirect activation and phosphorylation of the proto-oncogene Src occurred resulting in nuclear translocation of the transcription factor Twist1 [95]. Through this mechanism, periostin production is increased, associated with an enhancement in the migratory potential and invasiveness of MPM, contributing to both progression of disease and metastases. CD26 therefore represents a targetable MPM specific molecule with a direct role in the invasiveness and metastatic potential of MPM.

3.2. CD26 Targeting in Malignant Mesothelioma

As mentioned above, proof-of-concept studies using preclinical MPM models demonstrate that targeting CD26 with its specific mAbs is a viable anti-cancer therapeutic approach. Treatment of MPM cell lines with the humanized anti-CD26 mAB, YS110 resulted in p27^{kip1} accumulation leading to cytotoxicity in both *in vitro* and *in vivo* models [88]. Binding of YS110 also had a direct role in regulating binding to extra-cellular matrix proteins, biphasic antitumor immunity through immune activation and direct cytotoxicity, as well as inhibition of distant metastases [88]. In addition, anti-CD26 mAB enhanced nuclear translocation of CD26 with downstream effects resulting in mesothelioma growth suppression [96, 97]. Cellular localization analysis revealed that YS110 caused an increase in transport *via* caveolin-dependent endocytosis and accumulation of CD26 to the nucleus of MPM resulting in suppression of POLR2 gene expression and subsequent growth suppression of MPM cells, while highlighting a secondary anti-tumor mechanism of anti-CD26 mAbs [96, 97]. Meanwhile, a recently published paper demonstrated that YS110 caused retarded G2/M cell cycle progression through inhibition of phosphorylation of cdc2 and cdc25C and activation of ERK1/2 [98]. Importantly, a synergistic effect between YS110 and the first line anti-MPM chemotherapeutic agent pemetrexed was observed, as the combination of YS110 and pemetrexed showed superior anti-tumor activity associated with combinatorial G1/S and G2/M cell cycle tran-

sition inhibition than either agent alone in a mouse xenograft model of MPM [98].

These encouraging preclinical results along with our increased understanding of the novel molecular mechanisms involved in CD26 targeting in cancer cells led to the first-in-human phase I study of YS110 in CD26 expressing cancer cells. This recently published study involved 33 patients with CD26+ tumors (22 of whom had heavily pre-treated MPM) treated with a standard 3+3 escalation scheme with escalating doses of YS110 [99]. YS110 was generally well tolerated even to doses of 6mg/kg weekly, with maximal tolerated dose not reached and only 2 patients reporting grade 3 or higher anaphylactic or allergic reactions, which entirely resolved with supportive treatment and dose omission [99]. When the study was subsequently amended to add clinically relevant allergies as a new exclusion criterion and to allow for the administration of a systemic steroid prophylaxis prior to each infusion to better control infusions reactions, the safety profile was even further improved with treatment doses being escalated to 6mg/kg without dose limiting toxicities. While there was a transient decrease in total peripheral lymphocyte counts and CD26+ lymphocyte subsets following antibody administration, there was no observed autoimmune or infectious disease occurrences [99]. Furthermore, in this first-in-human phase I study, prolonged disease stabilization was observed in a significant number of patients that received YS110 [99, 100]. Thirteen of 26 evaluable patients treated with YS110 had stable disease as the best response for an overall median PFS of 43 days, while 7 patients (including five cases of mesothelioma) experienced prolonged PFS of 184-399 days [99, 100]. Taken together, data from both preclinical studies as well as the recently completed first-in-human phase I clinical trial indicate that additional testing of YS110, which may represent a major breakthrough in the treatment of MPM, in future clinical trials as either single agent therapy or as part of combination therapies with other anti-neoplastic agents, would be warranted.

CONCLUSION

MPM is an ongoing oncological concern. With continued mining and use of asbestos globally in developing countries and major industrialized

nations like China and Russia, MPM cases will predictably increase in the coming decades. When found, treatment strategies include combination surgery, radiation, and chemotherapy; although few patients are diagnosed in time to benefit from definitive surgery, with systemic chemotherapy being thus left for many patients as their only option. While beneficial at reducing symptoms, systemic chemotherapy with pemetrexed and cisplatin (SOC) fails to provide curative benefit for most MPM patients and only extends overall survival by a matter of months. The addition of novel biologic agents such as bevacizumab to standard of care will likely provide modest survival benefit but has yet to become the approved standard. For most patients with MPM, following initial systemic therapy, disease commonly relapses with progression and metastases. Over the past 20 years, many chemo- and immunotherapeutics have been evaluated as potential treatments for MPM with limited benefit. Novel immunotherapy strategies including passive immunotherapy, mesothelin targeting, and checkpoint inhibition targeting have recently shown promise in small phase I and II studies. In this review, we highlight the need for novel therapeutic approaches in MPM and discuss the potential of CD26 as a new molecular target. CD26 is highly expressed in MPM with limited expression in normal mesothelial cells. Unlike other immunotherapeutic targets, CD26 has a direct role in progression, invasion, metastasis, and cancer-stem cell proliferation in MPM. Our recently completed first-in-human phase I clinical trial involving the anti-CD26 mAb YS110 suggests that successful targeting of CD26 may lead potentially to improved disease control and prolonged overall survival with limited toxic side effects in malignant mesothelioma. The successful development of a CD26-targeted approach will enhance the treatment armamentarium available against MPM as we prepare for this predictive epidemic.

LIST OF ABBREVIATIONS

DPPIV	=	Dipeptidyl Peptidase IV
MPM	=	Malignant Pleural Mesothelioma
mAb	=	Monoclonal Antibody
SOC	=	Standard of Care

CSC = Cancer Stem Cells

OS = Overall Survival

CONSENT FOR PUBLICATION

Not applicable.

CONFLICT OF INTEREST

Nam H. Dang, Chikao Morimoto, and Kei Ohnuma are stockholders of Y's AC Co, Ltd.

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REFERENCES

- [1] Ohnuma K, Dang NH, Morimoto C. Revisiting an old acquaintance: CD26 and its molecular mechanisms in T cell function. *Trends Immunol* 2008; 29(6): 295-301.
- [2] Boonacker E, Van Noorden CJ. The multifunctional or moonlighting protein CD26/DPPIV. *Eur J Cell Biol* 2003; 82(2): 53-73.
- [3] Hosono O, Ohnuma K, Dang NH, Morimoto C. CD26: A key molecule in immune regulation and autoimmune diseases. *Mod Rheumatol* 2003; 13(3): 199-204.
- [4] Matteucci E, Giampietro O. Dipeptidyl peptidase-4 (CD26): Knowing the function before inhibiting the enzyme. *Curr Med Chem* 2009; 16(23): 2943-51.
- [5] Mortier A, Gouwy M, Van Damme J, Proost P, Struyf S. CD26/dipeptidylpeptidase IV-chemokine interactions: Double-edged regulation of inflammation and tumor biology. *J Leukoc Biol* 2016; 99(6): 955-69.
- [6] Cordero OJ, Salgado FJ, Nogueira M. On the origin of serum CD26 and its altered concentration in cancer patients. *Cancer Immunol Immunother* 2009; 58(11): 1723-47.
- [7] Ohnuma K, Takahashi N, Yamochi T, Hosono O, Dang NH, Morimoto C. Role of CD26/dipeptidyl peptidase IV in human T cell activation and function. *Front Biosci* 2008; 13: 2299-310.
- [8] Klemann C, Wagner L, Stephan M, Von Hörsten S. Cut to the chase: A review of CD26/dipeptidyl peptidase-4 (DPP4)'S entanglement in the immune system. *Clin Exp Immunol* 2016; 185(1): 1-21.
- [9] Ohnuma K, Inoue H, Uchiyama M, *et al.* T-cell activation *via* CD26 and caveolin-1 in rheumatoid synovium. *Mod Rheumatol* 2006; 16(1): 3-13.
- [10] Dang NH, Morimoto C. CD26: An expanding role in immune regulation and cancer. *Histol Histopathol* 2002; 17(4): 1213-26.
- [11] Dang NH, Aytac U, Sato K, *et al.* T-large granular lymphocyte lymphoproliferative disorder: Expression of CD26 as a marker of clinically aggressive disease and characterization of marrow inhibition. *Br J Haematol* 2003; 121(6): 857-65.
- [12] Havre PA, Abe M, Urasaki Y, Ohnuma K, Morimoto C, Dang NH. The role of CD26/dipeptidyl peptidase IV in cancer. *Front Biosci* 2008; 13: 1634-45.
- [13] Lam CS, Cheung AH, Wong SK, *et al.* Prognostic significance of CD26 in patients with colorectal cancer. *PLoS One* 2014; 9(5): e98582.
- [14] Ho L, Aytac U, Stephens LC, *et al.* *In vitro* and *in vivo* antitumor effect of the anti-CD26 monoclonal antibody 1F7 on human CD30+ anaplastic large cell T-cell lymphoma Karpas 299. *Clin Cancer Res* 2001; 7(7): 2031-40.
- [15] Davies S, Beckenkamp A, Buffon A. CD26 a cancer stem cell marker and therapeutic target. *Biomed Pharmacother* 2015; 71: 135-8.
- [16] Ai J, Stevenson JP. Current issues in malignant pleural mesothelioma evaluation and management. *Oncologist* 2014; 19(9): 975-84.
- [17] Robinson BM. Malignant pleural mesothelioma: An epidemiological perspective. *Ann Cardiothorac Surg* 2012; 1(4): 491-6.
- [18] Røe OD, Stella GM. Malignant pleural mesothelioma: History, controversy and future of a manmade epidemic. *Eur Respir Rev* 2015; 24(135): 115-31.
- [19] Riva MA, Carnevale F, Sironi VA, De Vito G, Cesana G. Mesothelioma and asbestos, fifty years of evidence: Chris Wagner and the contribution of the Italian occupational medicine community. *Med Lav* 2010; 101(6): 409-15.
- [20] Bononi A, Napolitano A, Pass HI, Yang H, Carbone M. Latest developments in our understanding of the pathogenesis of mesothelioma and the design of targeted therapies. *Expert Rev Respir Med* 2015; 9(5): 633-54.
- [21] Pasdar EA, Smits M, Stapelberg M, *et al.* Characterisation of mesothelioma-initiating cells and their susceptibility to anti-cancer agents. *PLoS One* 2015; 10(5): e0119549.
- [22] Price B, Ware A. Time trend of mesothelioma incidence in the United States and projection of future cases: An update based on SEER data for 1973 through 2005. *Crit Rev Toxicol* 2009; 39(7): 576-88.
- [23] Markowitz S. Asbestos-related lung cancer and malignant mesothelioma of the pleura: Selected current issues. *Semin Respir Crit Care Med* 2015; 36(3): 334-46.
- [24] Zhang W, Wu X, Wu L, Zhao X. Advances in the diagnosis, treatment and prognosis of malignant pleural mesothelioma. *Ann Transl Med* 2015; 3(13): 182.
- [25] Adusumilli PS. Translational immunotherapeutics: Chemoimmunotherapy for malignant pleural mesothelioma. *Cancer* 2014; 120(21): 3268-71.
- [26] Blomberg C, Nilsson J, Holgersson G, *et al.* Randomized trials of systemic medically-treated malignant

- mesothelioma: A systematic review. *Anticancer Res* 2015; 35(5): 2493-501.
- [27] Buikhuisen WA, Hiddinga BI, Baas P, van Meerbeeck JP. Second line therapy in malignant pleural mesothelioma: A systematic review. *Lung Cancer* 2015; 89(3): 223-31.
- [28] Calabrò L, Maio M. Immune checkpoint blockade in malignant mesothelioma. *Semin Oncol* 2015; 42(3): 418-22.
- [29] Cao C, Akhunjy Z, Fu B, *et al.* Surgical management of malignant pleural mesothelioma: An update of clinical evidence. *Semin Thorac Cardiovasc Surg* 2015; 27(1): 6-8.
- [30] Christoph DC, Eberhardt WE. Systemic treatment of malignant pleural mesothelioma: New agents in clinical trials raise hope of relevant improvements. *Curr Opin Oncol* 2014; 26(2): 171-81.
- [31] Jing XQ, Zhou L, Sun XD, Yu JM, Meng X. Pemetrexed maintenance therapy following bevacizumab-containing first-line chemotherapy in advanced malignant pleural mesothelioma: A case report and literatures review. *Medicine (Baltimore)* 2016; 95(14): e3351.
- [32] Bertoglio P, Waller DA. The role of thoracic surgery in the management of mesothelioma: An expert opinion on the limited evidence. *Expert Rev Respir Med* 2016; 10(6): 663-72.
- [33] Treasure T, Waller D, Tan C, *et al.* The mesothelioma and radical surgery randomized controlled trial: The mars feasibility study. *J Thorac Oncol* 2009; 4(10): 1254-8.
- [34] Treasure T, Lang-Lazdunski L, Waller D, *et al.* Extrapleural pneumonectomy versus no extra-pleural pneumonectomy for patients with malignant pleural mesothelioma: Clinical outcomes of the Mesothelioma and Radical Surgery (MARS) randomised feasibility study. *Lancet Oncol* 2011; 12(8): 763-72.
- [35] Thieke C, Nicolay NH, Sterzing F, *et al.* Long-term results in malignant pleural mesothelioma treated with neoadjuvant chemotherapy, extrapleural pneumonectomy and intensity-modulated radiotherapy. *Radiat Oncol* 2015; 10: 267.
- [36] Federico R, Adolfo F, Giuseppe M, *et al.* Phase II trial of neoadjuvant pemetrexed plus cisplatin followed by surgery and radiation in the treatment of pleural mesothelioma. *BMC Cancer* 2013; 13: 22.
- [37] Kondola S, Manners D, Nowak AK. Malignant pleural mesothelioma: An update on diagnosis and treatment options. *Ther Adv Respir Dis* 2016; 10(3): 275-88.
- [38] Reck M, Gatzemeier U. Pemetrexed-cisplatin combination in mesothelioma. *Expert Rev Anticancer Ther* 2005; 5(2): 231-7.
- [39] Vogelzang NJ, Rusthoven JJ, Symanowski J, *et al.* Phase III study of pemetrexed in combination with cisplatin versus cisplatin alone in patients with malignant pleural mesothelioma. *J Clin Oncol* 2003; 21(14): 2636-44.
- [40] van Meerbeeck JP, Gaafar R, Manegold C, *et al.* Randomized phase III study of cisplatin with or without raltitrexed in patients with malignant pleural mesothelioma: An intergroup study of the European Organisation for Research and Treatment of Cancer Lung Cancer Group and the National Cancer Institute of Canada. *J Clin Oncol* 2005; 23(28): 6881-9.
- [41] Habib EE, Fahmy ES. Chemotherapy management of malignant pleural mesothelioma: A phase II study comparing two popular chemotherapy regimens. *Clin Transl Oncol* 2013; 15(11): 965-8.
- [42] Zalcman G, Mazieres J, Margery J, *et al.* Bevacizumab for newly diagnosed pleural mesothelioma in the Mesothelioma Avastin Cisplatin Pemetrexed Study (MAPS): A randomised, controlled, open-label, phase 3 trial. *Lancet* 2016; 387(10026): 1405-14.
- [43] Abdel-Rahman O, Kelany M. Systemic therapy options for malignant pleural mesothelioma beyond first-line therapy: A systematic review. *Expert Rev Respir Med* 2015; 9(5): 533-49.
- [44] Hall RD, Le TM, Haggstrom DE, Gentzler RD. Angiogenesis inhibition as a therapeutic strategy in Non-Small Cell Lung Cancer (NSCLC). *Transl Lung Cancer Res* 2015; 4(5): 515-23.
- [45] Kim SH, Park WS, Joung JY, Seo HK, Lee KH, Chung J. Systemic treatments for metastatic renal cell carcinoma: 10-year experience of immunotherapy and targeted therapy. *Cancer Res Treat* 2016; 48(3): 1092-101.
- [46] Cella D, Beaumont JL. Pazopanib in the treatment of advanced renal cell carcinoma. *Ther Adv Urol* 2016; 8(1): 61-9.
- [47] Siddiqi T, Rosen ST. Novel biologic agents for non-Hodgkin lymphoma and chronic lymphocytic leukemia-part 2: Adoptive cellular immunotherapy, small-molecule inhibitors, and immunomodulation. *Oncology (Williston Park)* 2015; 29(4): 299-308.
- [48] Dubey S, Jänne PA, Krug L, *et al.* A phase II study of sorafenib in malignant mesothelioma: Results of cancer and leukemia group B 30307. *J Thorac Oncol* 2010; 5(10): 1655-61.
- [49] Camidge DR, Blais N, Jonker DJ, *et al.* Sunitinib combined with pemetrexed and cisplatin: Results of a phase I dose-escalation and pharmacokinetic study in patients with advanced solid malignancies, with an expanded cohort in non-small cell lung cancer and mesothelioma. *Cancer Chemother Pharmacol* 2013; 71(2): 307-19.
- [50] Laurie SA, Gupta A, Chu Q, *et al.* Brief report: A phase II study of sunitinib in malignant pleural mesothelioma. The NCIC Clinical Trials Group. *J Thorac Oncol* 2011; 6(11): 1950-4.
- [51] Dudek AZ, Pang H, Kratzke RA, *et al.* Phase II study of dasatinib in patients with previously treated malignant mesothelioma (cancer and leukemia group B 30601): A brief report. *J Thorac Oncol* 2012; 7(4): 755-9.
- [52] Mathy A, Baas P, Dalesio O, van Zandwijk N. Limited efficacy of imatinib mesylate in malignant meso-

- thelioma: A phase II trial. *Lung Cancer* 2005; 50(1): 83-6.
- [53] Guazzelli A, Bakker E, Tian K, Demonacos C, Krstic-Demonacos M, Mutti L. Promising investigational drug candidates in phase I and phase II clinical trials for mesothelioma. *Expert Opin Investig Drugs* 2017; 26(8): 933-944.
- [54] Buikhuisen WA, Burgers JA, Vincent AD, *et al.* Thalidomide versus active supportive care for maintenance in patients with malignant mesothelioma after first-line chemotherapy (NVALT 5): An open-label, multicentre, randomised phase 3 study. *Lancet Oncol* 2013; 14(6): 543-51.
- [55] Govindan R, Kratzke RA, Herndon JE, *et al.* Gefitinib in patients with malignant mesothelioma: A phase II study by the cancer and leukemia group B. *Clin Cancer Res* 2005; 11(6): 2300-4.
- [56] Garland LL, Rankin C, Gandara DR, *et al.* Phase II study of erlotinib in patients with malignant pleural mesothelioma: A southwest oncology group study. *J Clin Oncol* 2007; 25(17): 2406-13.
- [57] Jackman DM, Kindler HL, Yeap BY, *et al.* Erlotinib plus bevacizumab in previously treated patients with malignant pleural mesothelioma. *Cancer* 2008; 113(4): 808-14.
- [58] O'Brien ME, Gaafar RM, Popat S, *et al.* Phase II study of first-line bortezomib and cisplatin in malignant pleural mesothelioma and prospective validation of progression free survival rate as a primary endpoint for mesothelioma clinical trials (European Organisation for research and treatment of cancer 08052). *Eur J Cancer* 2013; 49(13): 2815-22.
- [59] Tada Y, Suzuki T, Shimada H, Hiroshima K, Tatsumi K, Tagawa M. Molecular-targeted therapy for malignant mesothelioma. *Pleura* 2015; 2: 1-11.
- [60] Paik PK, Krug LM. Histone deacetylase inhibitors in malignant pleural mesothelioma: Preclinical rationale and clinical trials. *J Thorac Oncol* 2010; 5(2): 275-9.
- [61] Krug LM, Kindler HL, Calvert H, *et al.* Vorinostat in patients with advanced malignant pleural mesothelioma who have progressed on previous chemotherapy (VANTAGE-014): A phase 3, double-blind, randomised, placebo-controlled trial. *Lancet Oncol* 2015; 16(4): 447-56.
- [62] McMillan R, Zauderer M, Bott M, Ladanyi M. Important recent insights into the genetics and biology of malignant pleural mesothelioma. *Ann Cardiothorac Surg* 2012; 1(4): 462-5.
- [63] de Assis LV, Locatelli J, Isoldi MC. The role of key genes and pathways involved in the tumorigenesis of malignant mesothelioma. *Biochim Biophys Acta* 2014; 1845(2): 232-47.
- [64] Lo Iacono M, Monica V, Righi L, *et al.* Targeted next-generation sequencing of cancer genes in advanced stage malignant pleural mesothelioma: A retrospective study. *J Thorac Oncol* 2015; 10(3): 492-9.
- [65] Jean D, Daubriac J, Le Pimpec-Barthes F, Galateau-Salle F, Jaurand MC. Molecular changes in mesothelioma with an impact on prognosis and treatment. *Arch Pathol Lab Med* 2012; 136(3): 277-93.
- [66] Shapiro IM, Kolev VN, Vidal CM, *et al.* Merlin deficiency predicts FAK inhibitor sensitivity: A synthetic lethal relationship. *Sci Transl Med* 2014; 6(237): 237ra68.
- [67] Fennell DA, Baas P, Kindler HL, *et al.* COMMAND: A phase II randomized, double-blind, placebo-controlled, multicenter study of defactinib as maintenance therapy in subjects with malignant pleural mesothelioma that has not progressed on at least four cycles of pemetrexed/platinum therapy. *J Clin Oncol* 2014; 32: 15.
- [68] Verastem Stops Enrollment Due to Futility in the COMMAND Study of VS-6063 for the Treatment of Malignant Pleural Mesothelioma [press release]. 2015; <http://www.businesswire.com/news/home/20150928005465/en/Verastem-Stops-Enrollment-Due-Futility-COMMAND-Study>
- [69] Maggioni C, Barletta G, Rijavec E, Biello F, Gualco E, Grossi F. Advances in treatment of mesothelioma. *Expert Opin Pharmacother* 2016; 17(9): 1197-205.
- [70] Hassan R, Schweizer C, Lu KF, *et al.* Inhibition of mesothelin-CA-125 interaction in patients with mesothelioma by the anti-mesothelin monoclonal antibody MORAb-009: Implications for cancer therapy. *Lung Cancer* 2010; 68(3): 455-9.
- [71] Calabrò L, Maio M. Immune checkpoint blockade in malignant mesothelioma: A novel therapeutic strategy against a deadly disease? *Oncoimmunology* 2014; 3(1): e27482.
- [72] Thapa B, Watkins DN, John T. Immunotherapy for malignant mesothelioma: Reality check. *Expert Rev Anticancer Ther* 2016; 6: 1-10.
- [73] Ceresoli GL, Bonomi M, Sauta MG. Immune checkpoint inhibitors in malignant pleural mesothelioma: Promises and challenges. *Expert Rev Anticancer Ther* 2016; 16(7): 673-5.
- [74] Roncella S, Laurent S, Fontana V, *et al.* CTLA-4 in mesothelioma patients: Tissue expression, body fluid levels and possible relevance as a prognostic factor. *Cancer Immunol Immunother* 2016; 65(8): 909-17.
- [75] Guazzelli A, Bakker E, Krstic-Demonacos M, Lisanti MP, Sotgia F, Mutti L. Anti-CTLA-4 therapy for malignant mesothelioma. *Immunotherapy* 2017; 9(3): 273-80.
- [76] Calabrò L, Morra A, Fonsatti E, *et al.* Efficacy and safety of an intensified schedule of tremelimumab for chemotherapy-resistant malignant mesothelioma: An open-label, single-arm, phase 2 study. *Lancet Respir Med* 2015; 3(4): 301-9.
- [77] Marcq E, Pauwels P, van Meerbeeck JP, Smits EL. Targeting immune checkpoints: New opportunity for mesothelioma treatment? *Cancer Treat Rev* 2015; 41(10): 914-24.
- [78] Alley EW, Molife LR, Santoro A, *et al.* Abstract CT103: Clinical safety and efficacy of pembrolizumab (MK-3475) in patients with malignant pleural

- mesothelioma: Preliminary results from KEYNOTE-028. *Cancer Res* 2015; 75(15 Suppl): CT103.
- [79] Remon J, Reguart N, Corral J, Lianes P. Malignant pleural mesothelioma: New hope in the horizon with novel therapeutic strategies. *Cancer Treat Rev* 2015; 41(1): 27-34.
- [80] Servais EL, Colovos C, Rodriguez L, *et al.* Mesothelin overexpression promotes mesothelioma cell invasion and MMP-9 secretion in an orthotopic mouse model and in epithelioid pleural mesothelioma patients. *Clin Cancer Res* 2012; 18(9): 2478-89.
- [81] Hassan R, Kindler HL, Jahan T, *et al.* Phase II clinical trial of amatuximab, a chimeric antimesothelin antibody with pemetrexed and cisplatin in advanced unresectable pleural mesothelioma. *Clin Cancer Res* 2014; 20(23): 5927-36.
- [82] Jahan T, Hassan R, Alley E, *et al.* 208O_PR: CRS-207 with chemotherapy (chemo) in malignant pleural mesothelioma (MPM): Results from a phase 1b trial. *J Thorac Oncol* 2016; 11(4 Suppl): S156.
- [83] Hassan R, Sharon E, Thomas A, *et al.* Phase 1 study of the antimesothelin immunotoxin SS1P in combination with pemetrexed and cisplatin for front-line therapy of pleural mesothelioma and correlation of tumor response with serum mesothelin, megakaryocyte potentiating factor, and cancer antigen 125. *Cancer* 2014; 120(21): 3311-9.
- [84] Cornelissen R, Hegmans JP, Maat AP, *et al.* Extended tumor control after dendritic cell vaccination with low-dose cyclophosphamide as adjuvant treatment in patients with malignant pleural mesothelioma. *Am J Respir Crit Care Med* 2016; 193(9): 1023-31.
- [85] Tan A, Porcher R, Crequit P, Ravaud P, Dechartres A. Differences in treatment effect size between overall survival and progression-free survival in immunotherapy trials: A meta-epidemiologic study of trials with results posted at ClinicalTrials.gov. *J Clin Oncol* 2017; 35(15): 1686-94.
- [86] Gyawali B, Prasad V. Drugs that lack single-agent activity: Are they worth pursuing in combination? *Nat Rev Clin Oncol* 2017; 14(4): 193-4.
- [87] Gyawali B, Prasad V. Combining drugs and extending treatment: A PFS end point is not sufficient. *Nat Rev Clin Oncol* 2017; 14(9): 521-522.
- [88] Inamoto T, Yamada T, Ohnuma K, *et al.* Humanized anti-CD26 monoclonal antibody as a treatment for malignant mesothelioma tumors. *Clin Cancer Res* 2007; 13(14): 4191-200.
- [89] Dang NH, Torimoto Y, Schlossman SF, Morimoto C. Human CD4 helper T cell activation: Functional involvement of two distinct collagen receptors, 1F7 and VLA integrin family. *J Exp Med* 1990; 172(2): 649-52.
- [90] Amatya VJ, Takeshima Y, Kushitani K, Yamada T, Morimoto C, Inai K. Overexpression of CD26/DPPIV in mesothelioma tissue and mesothelioma cell lines. *Oncol Rep* 2011; 26(6): 1369-75.
- [91] Aoe K, Amatya VJ, Fujimoto N, *et al.* CD26 overexpression is associated with prolonged survival and enhanced chemosensitivity in malignant pleural mesothelioma. *Clin Cancer Res* 2012; 18(5): 1447-56.
- [92] Fujimoto N, Ohnuma K, Aoe K, *et al.* Clinical significance of soluble CD26 in malignant pleural mesothelioma. *PLoS One* 2014; 9(12): e115647.
- [93] Yamazaki H, Naito M, Ghani FI, Dang NH, Iwata S, Morimoto C. Characterization of cancer stem cell properties of CD24 and CD26-positive human malignant mesothelioma cells. *Biochem Biophys Res Commun* 2012; 419(3): 529-36.
- [94] Okamoto T, Iwata S, Yamazaki H, *et al.* CD9 negatively regulates CD26 expression and inhibits CD26-mediated enhancement of invasive potential of malignant mesothelioma cells. *PLoS One* 2014; 9(1): e86671.
- [95] Komiya E, Ohnuma K, Yamazaki H, *et al.* CD26-mediated regulation of periostin expression contributes to migration and invasion of malignant pleural mesothelioma cells. *Biochem Biophys Res Commun* 2014; 447(4): 609-15.
- [96] Yamada K, Hayashi M, Du W, *et al.* Localization of CD26/DPPIV in nucleus and its nuclear translocation enhanced by anti-CD26 monoclonal antibody with anti-tumor effect. *Cancer Cell Int* 2009; 9: 17.
- [97] Yamada K, Hayashi M, Madokoro H, *et al.* Nuclear localization of CD26 induced by a humanized monoclonal antibody inhibits tumor cell growth by modulating of POLR2A transcription. *PLoS One* 2013; 8(4): e62304.
- [98] Hayashi M, Madokoro H, Yamada K, *et al.* A humanized anti-CD26 monoclonal antibody inhibits cell growth of malignant mesothelioma *via* retarded G2/M cell cycle transition. *Cancer Cell Int* 2016; 16: 35.
- [99] Angevin E, Isambert N, Trillet-Lenoir V, *et al.* First-in-human phase I of YS110, a monoclonal antibody directed against CD26 in advanced CD26-expressing cancers. *Br J Cancer* 2017; 116: 1926.
- [100] Nakano T, Kuribayashi K, Mikami K. Possible new therapeutic agents for malignant pleural mesothelioma: Anti-CD26 monoclonal antibody and naftopidil. *Expert Rev Anticancer Ther* 2016; 8: 1-3.