

Identification of Novel 2,4,5-Trisubstituted Pyrimidines as Potent Dual Inhibitors of Plasmodial *PfGSK3/PfPK6* with Activity against Blood Stage Parasites In Vitro

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Cite This: *J. Med. Chem.* 2022, 65, 13172–13197



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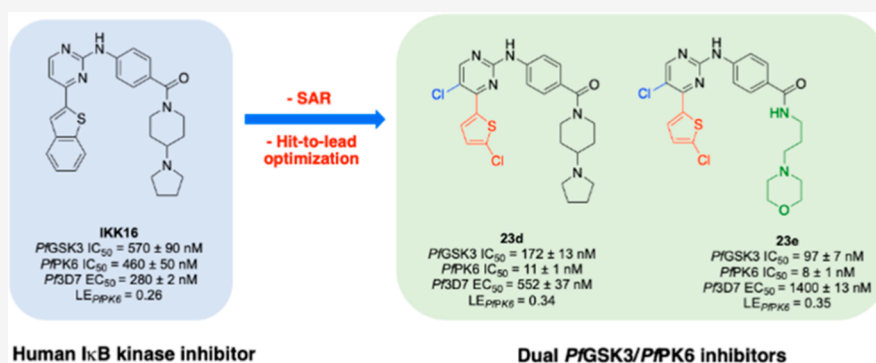
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ABSTRACT: Essential plasmodial kinases *PfGSK3* and *PfPK6* are considered novel drug targets to combat rising resistance to traditional antimalarial therapy. Herein, we report the discovery of **IKK16** as a dual *PfGSK3/PfPK6* inhibitor active against blood stage *Pf3D7* parasites. To establish structure–activity relationships for *PfPK6* and *PfGSK3*, 52 analogues were synthesized and assessed for the inhibition of *PfGSK3* and *PfPK6*, with potent inhibitors further assessed for activity against blood and liver stage parasites. This culminated in the discovery of dual *PfGSK3/PfPK6* inhibitors **23d** (*PfGSK3/PfPK6* IC₅₀ = 172/11 nM) and **23e** (*PfGSK3/PfPK6* IC₅₀ = 97/8 nM) with antiplasmodial activity (**23d** *Pf3D7* EC₅₀ = 552 ± 37 nM and **23e** *Pf3D7* EC₅₀ = 1400 ± 13 nM). However, both compounds exhibited significant promiscuity when tested in a panel of human kinase targets. Our results demonstrate that dual *PfPK6/PfGSK3* inhibitors with antiplasmodial activity can be identified and can set the stage for further optimization efforts.

INTRODUCTION

Malaria is an infectious disease caused by the protozoan parasite known as *Plasmodium*, which is transmitted to humans through the bite of an infected female *Anopheles* mosquito. A human host is infected when a mosquito transfers the parasite form, termed sporozoites, into the blood stream during a blood meal. These sporozoites invade the liver where they multiply via asexual reproduction to form parasites capable of erythrocyte invasion, termed merozoites. The merozoites egress from liver cells to reach the blood stream, where they replicate within the red blood cells and rapidly multiply. Following that, a fraction of merozoites will mature into male and female gametocytes, which are ingested by the female mosquito where sexual reproduction results in a new cycle of infection.¹ The World Health Organization (WHO) estimates that in 2020, there were 241 million malaria cases in 85 malaria endemic countries.² Deaths from malaria have been estimated at 627,000 in 2020.² There are five human-infective species of

Plasmodium parasite, with *Plasmodium falciparum* (*Pf*) being responsible for the majority of deaths worldwide.³ Encouragingly, the mortality rate of malaria (as measured by deaths per 100,000 population at risk) has been reduced from 25 in the year 2000 to 10 in 2019. This is the result of the global effort to combat malaria where the total funding for malaria control and elimination in 2020 was estimated at \$3.3 billion.² Although much progress has been achieved in attenuating the debilitating effects of malaria, there is still a need to identify

Received: June 23, 2022

Published: September 27, 2022



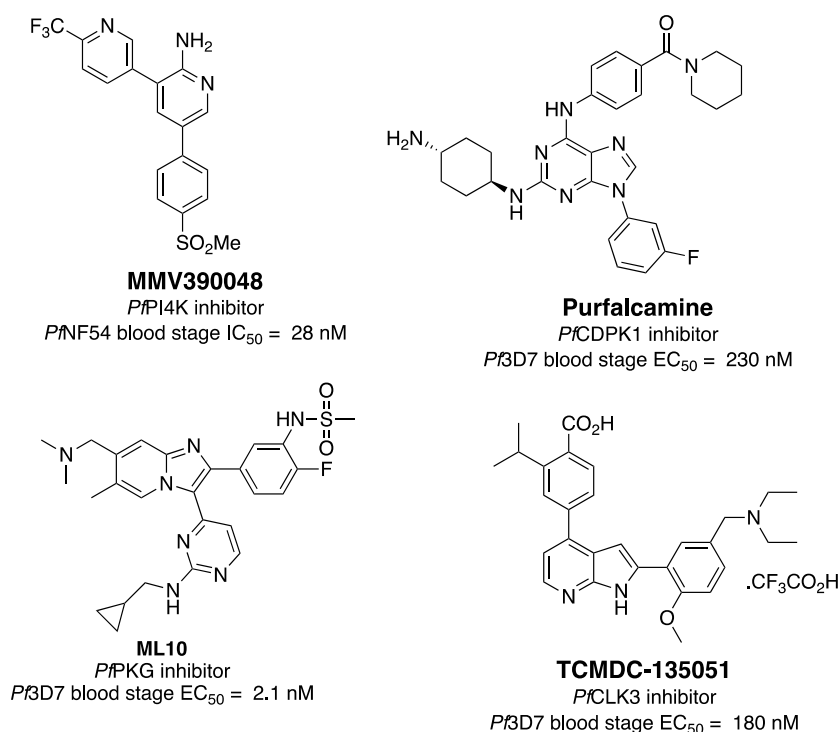


Figure 1. Structures of reported *Pf* kinase inhibitors with antiplasmodial activity.

novel antimalarial agents due to the risk of the increase of drug-resistant parasites.

Antimalarial drug development studies usually target two stages of the parasite's life cycle in the human host: (1) the erythrocytic stage, which is responsible for malaria pathogenesis and mortality, and (2) the hepatic stage, which is a clinically silent stage that is obligatory for the erythrocytic stage. Drugs that target the hepatic stage are considered prophylactic treatment. The parasites also have a transmission stage in the mosquito. Targeting this stage can block the disease from spreading.⁴ The WHO recommends artemisinin combination therapy (ACT) and vector control measures as the primary means to relieve the burden of malaria. However, artemisinin resistance has been detected in some parts of Asia, which has been linked to mutation in the *PfKelch13* gene.⁵ Moreover, a recent study from Rwanda found that *P. falciparum* samples isolated from patients were resistant to artemisinin in vitro, the first report of resistance in Africa.^{5,6}

Recently, plasmodial kinases have emerged as an attractive target for malaria treatment. Several reviews have highlighted the potential utility of plasmodial kinases as targets for antiplasmodial drug discovery.^{3,7–9} Recent studies have shown that many kinases of the *P. falciparum* kinome are indispensable for the survival of the parasite in humans and hence can be prioritized as drug targets.^{10,11} While the expression of kinases and their importance to viability can vary in the different life stages, several are essential throughout the life cycle.¹² Thus, targeting the essential kinases using potent and selective inhibitors may provide a new avenue in the fight against malaria since none of the approved antiplasmodial agents have been shown to target plasmodial kinases. Additionally, if these inhibitors can be developed into drugs, they could complement the current treatment regimen and lower the selection pressure for parasite drug resistance. However, only a few members of the plasmodial kinase family

have been pharmacologically validated as targets for antiplasmodial therapy. For instance, in 2017, Paquet et al. reported the discovery of a plasmodial phosphoinositol kinase inhibitor from a phenotypic screen, **MMV390048** (Figure 1). **MMV390048** targets *Pf* phosphatidylinositol 4-kinase (*Pf*PI4K) and could block multiple malaria life cycle stages in the human host, which prompted its advancement to clinical validation.¹³

Kato et al. demonstrated that *Pf*CDPK1 may be involved in regulating parasite motor-dependent processes that take place in the late schizont stage.¹⁴ Using an in vitro biochemical screen, they identified several structurally related 2,6,9-trisubstituted purines that were inhibitors of *Pf*CDPK1. From this series, purfalcamine was the most potent inhibitor, showing an IC₅₀ value of 17 nM against *Pf*CDPK1 (Figure 1). Purfalcamine was also active against blood stage parasites, inhibiting the growth of five different strains with EC₅₀ values in the nanomolar range. *Pf*PKG has also emerged as a kinase that could be targeted for antiplasmodial activity. Recent reports described a series of imidazopyridine inhibitors of *Pf*PKG, where the most potent compound of the series, **ML10** (Figure 1), inhibited blood stage proliferation with an EC₅₀ of 2.1 nM.¹⁵ This inhibitor not only targets merozoite invasion and egress but also prevents transmission of gametocytes to the mosquitos. *Pf*CLK3 has been recently identified as a validated protein kinase drug target for antiplasmodial drug discovery. Studies have shown that *Pf*CLK3 plays a role in the processing of parasite RNA, and thus, its inhibition could result in antiparasitic activity.¹⁶ This was confirmed with the identification of the potent *Pf*CLK3 inhibitor **TCMDC-135051** (Figure 1), which displayed nanomolar potency against the kinase, as well as showing 180 nM EC₅₀ against asexual blood stage *P. falciparum* parasites.^{17,18} Further studies showed that targeting *Pf*CLK3 can be used as a method for transmission blocking, as well as prophylaxis.¹⁸

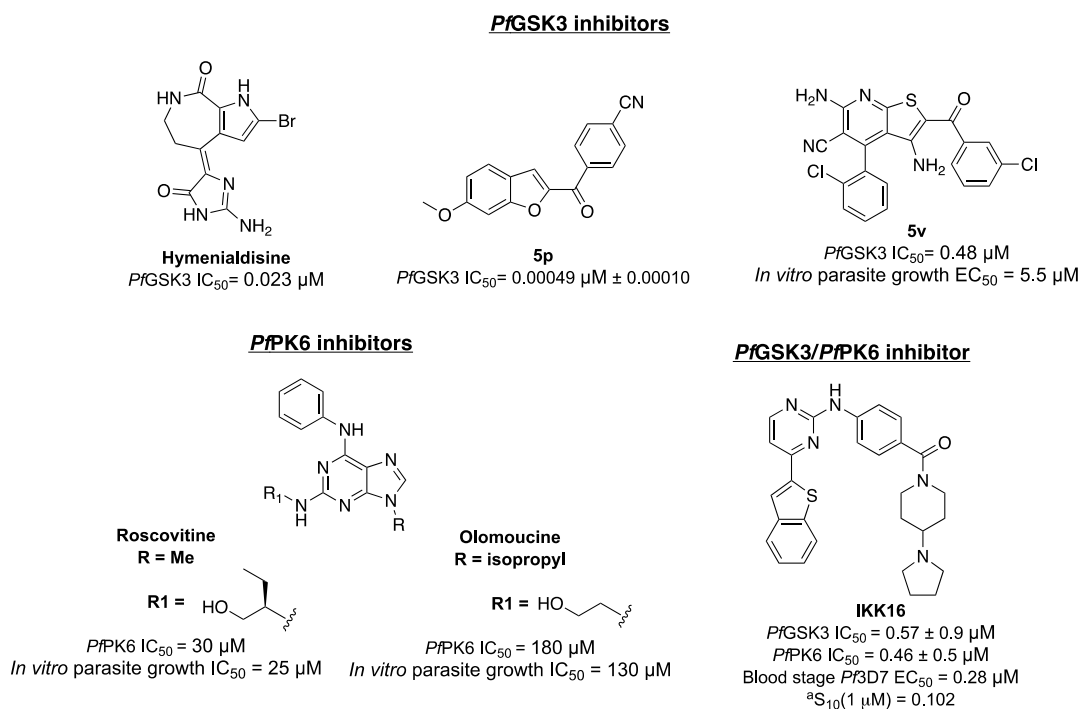


Figure 2. Exemplars of reported *Pf*GSK3 and *Pf*PK6 inhibitors^{25,27–29} and **IKK16**, a dual *Pf*GSK3 and *Pf*PK6 ligand identified as the starting point for this work. ^aSelectivity metric definition: S₁₀ (1 μM) = number of kinases having an experimental value percent of control (PoC) ≤10% at 1 μM divided by the total number of distinct kinases tested. This metric quantifies the percentage of kinases in a panel that a compound binds to at a chosen threshold at a particular screening concentration.

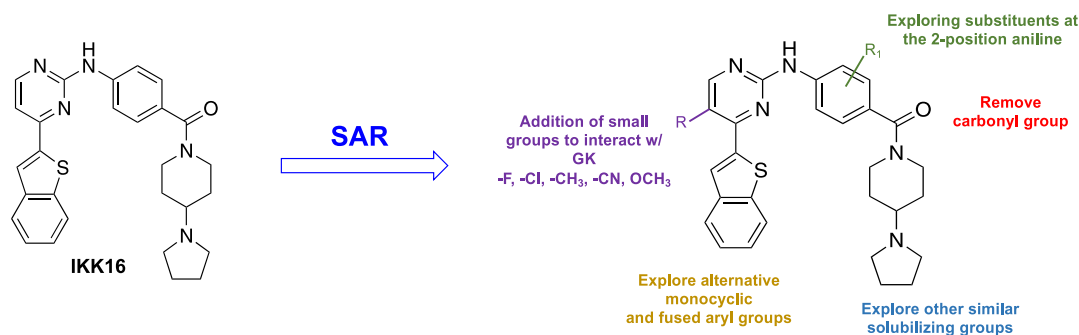


Figure 3. SAR study plan for the *Pf*GSK3/*Pf*PK6 dual inhibitor **IKK16**. (GK = gatekeeper residue of the kinase).

Using an *in vitro* split luciferase assay,¹⁹ which utilizes the proprietary KinaseSeeker technology, Luceome Biotechnologies developed assays against 11 plasmodial kinases, most of which have been shown to be essential for parasite survival.^{10,20} A pilot screen of these 11 *Pf* kinases utilizing a 110-member kinase inhibitor library composed of research stage, clinical stage, and marketed inhibitors identified several compounds with sub-micromolar activity against one or more plasmodial kinases (Luceome internal data). **IKK16**, a 2,4-disubstituted pyrimidine compound, was one of the inhibitors identified from the 110-compound screen (Figure 2). **IKK16** inhibited *Pf*GSK3 and *Pf*PK6, two of the 11 plasmodial kinases assayed, with IC₅₀ values of 570 ± 90 and 460 ± 50 nM, respectively.^{21,22} *Pf*GSK3 is a serine/threonine kinase that shares 75/76% sequence similarity with the kinase domains of the human GSK3α/GSK3β. The exact functions of *Pf*GSK3 are still unclear. However, one hypothesis implicates *Pf*GSK3 in regulating the strong circadian rhythm of the parasite.²³ Other reports outline the importance of *Pf*GSK3 for parasite invasion into the host erythrocytes.²⁴

*Pf*PK6 is also a serine/threonine kinase that shares structural similarity with both human CDK2 and p38 mitogen-activated protein kinase (MAPK). It contains 14 of the 15 conserved amino acid residues that are usually associated with the kinase domains of eukaryotes.²⁵ *Pf*PK6 is predominantly expressed during trophozoite and schizont stages of the intraerythrocytic cycle.^{9,25}

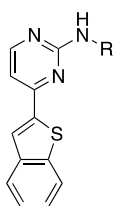
In addition to the aforementioned findings, independent studies using reverse genetics and saturation mutagenesis have deemed these two kinases essential for *P. falciparum* asexual blood stage proliferation.^{10,20} Accordingly, *Pf*GSK3 and *Pf*PK6 may have utility as novel targets for antimalarial pharmacotherapy.

Several inhibitors of *Pf*GSK3 and *Pf*PK6 have been reported in the literature; however, most of them lack the optimum biological profile in terms of enzymatic IC₅₀ or cellular EC₅₀ to be considered as antiplasmodial agents (Figure 2). **IKK16**, on the other hand, possessed a favorable activity profile, displayed low toxicity in a resazurin-based cell viability assay, and is orally bioavailable in rats.^{21,22,26} For these reasons, **IKK16**

appeared attractive as a starting point for antimalarial drug discovery efforts.

Our enthusiasm for **IKK16** stems from the fact that it inhibits not one but two plasmodial kinases thought to be essential for the survival of the malaria parasite. To our knowledge, no other compounds have been reported to target both kinases simultaneously with nanomolar potency. This multitargeted inhibition could be advantageous for antimalarial therapy, as it can reduce the likelihood of developing resistance. **IKK16** was originally reported as a human IKK inhibitor; however, no data regarding its broad kinome selectivity has been reported to date.²¹ Accordingly, we decided to test it against a panel of 468 human kinase targets at 1 μM using the DiscoverX KINOMEScan platform. **IKK16** showed an S_{10} (1 μM) value of 0.10, hitting 41 kinase targets with >90% inhibition (Figure 2 and Table 7). Although this

Table 1. In Vitro Enzymatic Activity of Simplified Analogues 4a–h, 5, and 6 on *PfGSK3* and *PfPK6*



compound	R	<i>PfGSK3</i> PoC@1 μM ^a	<i>PfPK6</i> PoC@1 μM
4a	Ph	100	100
4b	4-Cl-Ph	89	92
4c	3-Cl-Ph	89	100
4d	2-Cl-Ph	100	100
4e	4-OMe-Ph	97	88
4f	3-OMe-Ph	97	100
4g	2-OMe-Ph	100	100
4h	CH ₂ -Ph	100	100
5	4-CO ₂ Me-Ph	71	78
6	4-CO ₂ H-Ph	33	100

^a% enzymatic activity remaining after incubation with 1 μM of test compound.

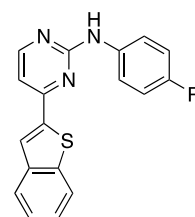
level of promiscuity against human kinases is not ideal, the literature report of low toxicity encouraged us to proceed to build an understanding of structure–activity relationships (SARs) for *PfGSK3* and *PfPK6*.

With this in mind, we pursued the 2-anilino-4-arylpyrimidine scaffold of **IKK16** hoping to achieve three goals: (1) establish SARs for activity on *PfPK6* and *PfGSK3*, (2) identify potent and selective dual inhibitors of plasmodial *PfPK6* and *PfGSK3*, and (3) screen the most potent kinase inhibitors identified from aim 2 for multistage antiplasmodial activity. The long-term goal of our campaign is to identify compounds that could serve as lead molecules for the future discovery of novel antiplasmodial medicines working through kinase inhibition. Our SAR study and optimization plan are summarized in Figure 3.

RESULTS AND DISCUSSION

Inhibition of Plasmodial *PfGSK3* and *PfPK6*. The synthesized compounds were tested for on target potency for inhibition of plasmodial *PfPK6* and *PfGSK3* using the proprietary KinaseSeeker split luciferase assay.¹⁹ Compounds were initially screened at 1 μM and only those showing $\geq 70\%$

Table 2. In Vitro Enzymatic Activity of Amide Analogues of *IKK16* on *PfGSK3* and *PfPK6*

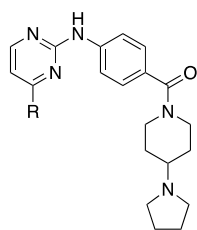


Compound	R	<i>PfGSK3</i> ^a IC ₅₀ (nM)	<i>PfPK6</i> IC ₅₀ (nM)
IKK16		570 \pm 90	460 \pm 50
8a		^b ND	181 \pm 4
8b		ND	243 \pm 15
8c		ND	264 \pm 24
9a		752 \pm 97	216 \pm 19
9b		695 \pm 88	115 \pm 11
9c		ND	398 \pm 35
9d		810 \pm 91	339 \pm 22
9e		308 \pm 27	340 \pm 25
9f		226 \pm 14	153 \pm 12
9g		243 \pm 20	156 \pm 7
9h		328 \pm 40	319 \pm 28
9i		223 \pm 33	215 \pm 21
9j		272 \pm 48	407 \pm 71
9k		ND	ND
12		ND	173 \pm 14

^aIC₅₀ values were determined using the KinaseSeeker assay with 5-fold dilutions, presented as mean values of two experiments performed in duplicate. ^bND: not determined; the compound did not reach the threshold of inhibiting $\geq 70\%$ kinase activity at 1 μM .

inhibition were further tested in a dose–response assay to determine their IC₅₀ values.

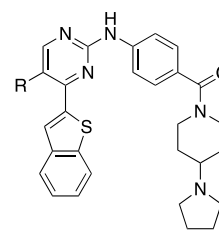
We began our SAR study by analyzing the role of the right-hand side of **IKK16** (Table 1). Typically, for aminopyrimidine kinase scaffolds, this 2-amino group makes a key hydrogen bond with a backbone carbonyl in the hinge region of the

Table 3. In Vitro Enzymatic Activity of 4-Substituted Pyrimidine Analogues on *Pf*GSK3 and *Pf*PK6

Compound	R	<i>Pf</i> GSK3 ^a IC ₅₀ (nM)	<i>Pf</i> PK6 IC ₅₀ (nM)
18a		1040 ± 148	317 ± 34
18b		^b ND	359 ± 40
18c		ND	236 ± 22
18d		ND	182 ± 22
18e		ND	768 ± 98
18f		ND	645 ± 79
18g		ND	329 ± 64
18h		698 ± 66	382 ± 37
18i		1141 ± 249	222 ± 9
18j		550 ± 60	130 ± 16
18k		ND	415 ± 35
18l		ND	563 ± 31
18m		490 ± 34	292 ± 22

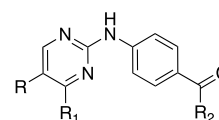
^aIC₅₀ values were determined using the KinaseSeeker assay with 5-fold dilutions, presented as mean values of two experiments performed in duplicate. ^bND: not determined; compound did not reach the threshold of inhibiting ≥70% kinase activity at 1 μM.

kinase and the substituents on the phenyl ring project toward the solvent interface.³⁰ Our first set of analogues investigated truncation of the 4-(pyrrolidinyl)piperidine moiety of **IKK16** together with the amide group to observe how these alterations would affect activity at *Pf*GSK3 and *Pf*PK6. This modification proved to be detrimental for activity, as compound **4a**, bearing only a simple aniline at the 2-position, failed to inhibit both kinases. We also evaluated various substituted anilines as 2-position substituents on the pyrimidine core. These anilines carried electron-donating as well as electron-withdrawing groups (**4b–g**). However, all these simplified analogues lacked

Table 4. In Vitro Enzymatic Activity of 5-Substituted Pyrimidine Analogues on *Pf*GSK3 and *Pf*PK6

compound	R	<i>Pf</i> GSK3 IC ₅₀ (nM) ^a	<i>Pf</i> PK6 IC ₅₀ (nM)
18n	CH ₃	710 ± 97	45 ± 2
18o	cyclopropyl	^b ND	114 ± 14
18p	OCH ₃	ND	294 ± 27
18q	F	456 ± 43	66 ± 2
18r	Cl	174 ± 16	19 ± 3
18s	Br	165 ± 18	57 ± 2
18t	CN	ND	77 ± 9

^aIC₅₀ values were determined using the KinaseSeeker assay with 5-fold dilutions, presented as mean values of two experiments performed in duplicate. ^bND: not determined; compound did not reach the threshold of inhibiting ≥70% kinase activity at 1 μM.

Table 5. In Vitro Enzymatic Activity of Hybrid Analogues on *Pf*GSK3 and *Pf*PK6 with Their Associated Ligand Efficiency Values

Compound	R	R ₁	R ₂	<i>Pf</i> GSK3 ^a IC ₅₀ (nM)	<i>Pf</i> PK6 IC ₅₀ (nM)	^b LE _{<i>Pf</i>PK6}
23a	CH ₃			^c ND	28 ± 3	0.32
23b	Cl			124 ± 5	14 ± 1	0.33
23c	CH ₃			1250 ± 155	14 ± 1	0.33
23d	Cl			172 ± 13	11 ± 1	0.34
23e	Cl			97 ± 7	8 ± 1	0.35
23f	Cl			79 ± 8	16 ± 2	0.34
23g	CH ₃			ND	12 ± 2	0.35

^aIC₅₀ values were determined using the KinaseSeeker assay with 5-fold dilutions, presented as mean values of two experiments performed in duplicate. ^bLigand efficiency was calculated as follows: *Pf*PK6 pIC₅₀/number of heavy atoms in the molecule. ^cND: not determined; compound did not reach the threshold of inhibiting ≥70% kinase activity at 1 μM.

any significant activity on both kinases. The benzylamine analogue **4h** was also inactive. Reinstalling the carbonyl group through a methyl ester (**5**)²¹ or a carboxylic acid (**6**)²¹ resulted in some binding, but the activity did not reach the threshold we set for IC₅₀ determination. Taken together, these results indicated the importance of an amide linker, as is present in **IKK16**, for activity on *Pf*GSK3 and *Pf*PK6.

Table 6. Activity of IKK16 Analogues against *P. falciparum* Blood Stage Proliferation with the Associated Cytotoxicity against HepG2 Cells

compound	<i>Pf</i> 3D7 EC ₅₀ (nM ± SEM) ^a	% viability@200 nM ^b	% viability@2 μM ^b
IKK16	280 ± 2	110	103
12	60 ± 5	100	97
18b	541 ± 29	105	82
18m	810 ± 19	110	87
18n	760 ± 61	87	66
18o	646 ± 10	83	55
18q	607 ± 16	98	79
18r	991 ± 65	94	61
18s	300 ± 9	83	47
18t	322 ± 7	95	84
23c	576 ± 28	87	65
23d	552 ± 37	113	34
23e	1400 ± 13	102	65
23f	2600 ± 35	99	54

^aEC₅₀ values were determined using the SYBR green I-based assay with 2-fold dilutions, presented as mean values performed in triplicate. ^b% of viable cells after incubation with 200 nM and 2 μM of the test compounds, presented as an average of two experiments performed in quadruplicate.

Table 7. Evaluation of Kinase Selectivity Scores

compound	S ₁₀ (1 μM) ^a	no. of kinase hits
IKK16	0.10	41
9g	0.08	32
18n	0.25	100
18r	0.22	88
23d	0.26	103
23e	0.16	64

^aSelectivity metric definition: S₁₀ (1 μM) = number of kinases having an experimental value PoC ≤ 10% at 1 μM divided by the total number of distinct kinases tested. This metric quantifies the percentage of kinases in a panel that a compound binds to at a chosen threshold at a particular screening concentration.

Next, we investigated the SAR around the amide group itself to assess the effect of various modifications on kinase activity (Table 2). We initially focused on the pyrrolidine ring of the 4-(pyrrolidinyl)piperidine side chain. The first modification we carried out was to open the pyrrolidine ring. Compound **8a** lost significant activity on *Pf*GSK3; however, it still retained activity at *Pf*PK6 and in fact showed an improvement in potency relative to **IKK16** (**8a** *Pf*PK6 IC₅₀ = 181 ± 4 nM). Other modifications to the pyrrolidine ring included replacing it with (*R*)-pyrrolidin-3-ol (**8b**), (*S*)-pyrrolidin-3-ol (**8c**), dimethyl amine (**9a**),²¹ as well as ring expansion to a six-membered ring (**9b**). The changes in **8b**, **8c**, and **9a** were all well tolerated by *Pf*PK6, with some improvement in potency and IC₅₀ values ranging from 216 to 274 nM. The same compounds lacked significant activity on *Pf*GSK3. Expanding the pyrrolidine ring to a six-membered piperidine ring (**9b**) provided roughly a 4-fold potency increase on *Pf*PK6. This ring expansion was somewhat tolerated on *Pf*GSK3, with **9b** having a *Pf*GSK3 IC₅₀ of 695 ± 88 nM. These findings suggest that an unaltered pyrrolidinyl ring is important for activity on *Pf*GSK3 since all modifications tried at this site, except for the subtle difference provided by ring expansion, resulted in significant loss of activity on *Pf*GSK3.

Next, we aimed to substitute the entire pyrrolidinyl-piperidine ring system of **IKK16** with simpler basic and non-basic functionalities that may mimic important elements of the original moiety. Substitution with a piperazine ring in **9c** saw a loss of activity on *Pf*GSK3, but activity on *Pf*PK6 was retained

with an IC₅₀ value of 398 ± 35 nM. Methylation of the piperazine-4-*N* atom resulted in **9d**, which retained activity on *Pf*PK6 but was less potent on *Pf*GSK3. Compound **9e** saw placing the amide nitrogen outside of the six-membered ring, and this modification restored activity on *Pf*GSK3 (*Pf*GSK3 IC₅₀ = 308 ± 27 nM). This pointed to the potential importance of maintaining a three-carbon linker between the amide nitrogen and the basic ring nitrogen for *Pf*GSK3 activity. In contrast, no difference in *Pf*PK6 IC₅₀ was observed for **9e** compared to the *N*-methyl piperazine analogue, **9d**.

Building upon this finding, we synthesized additional amides with substituents bearing a terminal basic nitrogen while maintaining the three-carbon linker between the amide nitrogen and the basic nitrogen, which was found to be important for *Pf*GSK3 activity. Accordingly, we used 3-(4-methylpiperazin-1-yl)propan-1-amine (**9f**), 3-morpholinopropan-1-amine (**9g**), and *N*1,*N*1-dimethylpropane-1,3-diamine (**9h**) as amide substituents. The piperazine analogue **9f** demonstrated an almost 3-fold increase in potency on *Pf*PK6 and a 2-fold potency increase on *Pf*GSK3 compared to **IKK16** (**9f** *Pf*PK6 IC₅₀ = 153 ± 12 nM and *Pf*GSK3 IC₅₀ = 226 ± 14 nM). The morpholine analogue **9g** exhibited comparable activity to **9f**, indicating that the *N*-methyl of the piperazine ring is not an important element for the observed potency enhancement. The simplified dimethyl analogue **9h** was less potent than the morpholine and piperazine analogues and demonstrated an IC₅₀ value of 328 ± 40 nM on *Pf*GSK3, while activity on *Pf*PK6 remained comparable to that of **IKK16**.

We also explored several non-basic amide substituents. The 3-(methylsulfonyl)propan-1-amine substituent led to a small increase in potency of inhibition on both kinases (**9i** *Pf*GSK3 IC₅₀ = 223 ± 33 nM and *Pf*PK6 IC₅₀ = 215 ± 21 nM), indicating that the presence of a basic moiety on the solvent-exposed tail was not a strict requirement for binding to *Pf*PK6 or *Pf*GSK3. The cyclopropylamine analogue **9j** showed a roughly 2-fold improvement in potency on *Pf*GSK3, while activity on *Pf*PK6 was largely unaffected in comparison to the parent, **IKK16**. Finally, compound **9k** with the 1-methyl-1*H*-pyrazole substituent showed no activity against either *Pf* kinase, which implied that aromaticity may not be well tolerated at this position.

Removing the carbonyl functionality from the amide group of **IKK16** resulted in the dibasic compound **12**. Although this modification abolished activity on *Pf*GSK3, the compound remained active on *Pf*PK6, demonstrating an IC_{50} value of 173 ± 14 nM, a nearly 3-fold increase in potency from **IKK16**.

Results from the SAR analysis of the right-hand side of **IKK16** demonstrate that several types of structural changes can be tolerated by *Pf*PK6, while *Pf*GSK3 appears to be more sensitive to changes, with several of the analogues losing *Pf*GSK3 activity with subtle structural changes. We also learned that binding can be retained without having an ionizable basic functionality at the distal end of the solvent-exposed tail.

We designed the next set of analogues to explore the 4-position of the pyrimidine ring. In addition to utilizing the moieties of similar size as benzothiophene, we investigated replacement of the benzothiophene of **IKK16** with smaller and less lipophilic aryl rings (Table 3). Incorporating the bioisosteric benzofuran to afford compound **18a** led to the decrease of activity on *Pf*GSK3, while activity on *Pf*PK6 remained almost the same as **IKK16** (**18a** *Pf*PK6 $IC_{50} = 317 \pm 34$ nM). A benzothiophene to naphthalene swap (**18b**) led to a significant loss of activity on *Pf*GSK3, while *Pf*PK6 activity remained comparable to that of the parent, **IKK16**. One possible explanation for this could be that the sulfur of the benzothiophene is somehow an important element for binding in the *Pf*GSK3 catalytic site while being less important for *Pf*PK6 binding. Next, we synthesized and tested a set of monocyclic and substituted monocyclic aromatic systems in the 4-position of the pyrimidine to probe ligand–kinase interactions at this position. Truncation of the fused phenyl ring of the benzothiophene of **IKK16** afforded compound **18c**, with an unsubstituted thiophene occupying the 4-position of the pyrimidine. Since this substitution restored a sulfur atom at this position, we were expecting to see some resurgence of *Pf*GSK3 activity. However, this truncation led to inactivity on *Pf*GSK3, but the IC_{50} on *Pf*PK6 was found to be 236 ± 22 nM, a 2-fold increase in potency from **IKK16**. This indicated that the fused benzene ring could be essential for *Pf*GSK3 activity. The other 4-position monocyclic aromatic rings included thiazole (**18d**), furan (**18e**), 1-methyl-1*H*-pyrazole (**18f**), and phenyl (**18g**). The thiazole analogue demonstrated a comparable activity profile to that of the thiophene counterpart, pointing to the minimal effect of the extra ring nitrogen on activity. Replacing the sulfur atom of the thiophene ring of **18c** for an oxygen to afford the furan isostere resulted in more than 3-fold decrease in potency on *Pf*PK6 (**18e** *Pf*PK6 $IC_{50} = 768 \pm 98$ nM) while remaining inactive on *Pf*GSK3. The more polar 1-methyl-1*H*-pyrazole derivative **18f** showed comparable activity to **18e** on both kinases. Results from **18e** and **18f** indicate that having polar groups on the aryl moiety of the pyrimidine-4-position may not be favored by *Pf*PK6. This finding was augmented by the observation that the more lipophilic phenyl analogue, **18g**, also displayed diminished activity on *Pf*GSK3 but showed a slightly better IC_{50} of 329 ± 64 nM on *Pf*PK6.

We attempted to mimic the benzothiophene by filling the space of the fused phenyl ring utilizing a 3,4-dichlorophenyl substituent. To our delight, compound **18h** saw a reinstatement of *Pf*GSK3 inhibitory activity (*Pf*GSK3 $IC_{50} = 698 \pm 66$ nM), while the compound remained equipotent to **18g** on *Pf*PK6.

So far, these results indicated that the benzothiophene was the optimum substituent on the pyrimidine 4-position for activity on *Pf*GSK3, while *Pf*PK6 tolerates some variability, such as thiophene **18c** with *Pf*PK6 $IC_{50} = 236 \pm 22$ nM. Building on this, and starting from **18c**, we attempted further structural exploration to restore activity on *Pf*GSK3 and/or improve *Pf*PK6 potency. We began by substituting the 5-position of the thiophene ring of **18c** with different lipophilic groups such as a methyl (**18i**), chloro (**18j**), and phenyl (**18k**). While the 5-methylthiophene substituent (**18i**) resulted in some reinstatement of *Pf*GSK3 activity ($IC_{50} = 1141 \pm 249$ nM), its IC_{50} on *Pf*PK6 remained roughly the same as that of the unsubstituted thiophene (**18i** *Pf*PK6 $IC_{50} = 222 \pm 9$ nM). Compound **18j**, with a 5-chlorothiophene substituent, saw a 2-fold increase in *Pf*GSK3 activity, showing an IC_{50} value of 550 ± 60 nM. Interestingly, **18j** saw some increase in potency against *Pf*PK6 also (**18j** *Pf*PK6 $IC_{50} = 130 \pm 16$ nM). The 5-phenylthiophene substituent (**18k**) afforded a compound that was inactive on *Pf*GSK3 and showed comparable activity to **IKK16** on *Pf*PK6.

If this pyrimidine series binds to *Pf* kinases in the same binding mode as is typically observed for similar pyrimidine kinase inhibitors, then the aryl group at the pyrimidine-4-position is likely directed toward the catalytic lysine of the kinase active site.³⁰ Consequently, we postulated that incorporation of a 4-position functional group capable of forming a hydrogen bond with this lysine residue could lead to better kinase inhibition. Previous studies have shown that such interaction is achievable (Lys208 in *Pf*GSK3).^{27,28} Accordingly, we synthesized compound **18l** with a (5-methylthiophen-2-yl)methanol group at the pyrimidine-4-position. This single analogue proved unsuccessful, as the activity on *Pf*GSK3 remained unchanged compared to the unsubstituted thiophene while also leading to decreased activity on *Pf*PK6. Further derivatives will need to be explored to see if engagement of the active site lysine is a viable strategy for potency enhancement. The trifluoroethyl-ether analogue **18m** was obtained as a byproduct in the synthesis of **18l**. Fortunately, this ether analogue showed activity on *Pf*GSK3 comparable to **IKK16** (**18m** *Pf*GSK3 $IC_{50} = 490 \pm 34$ nM) while showing an ~2-fold increase in potency on *Pf*PK6 compared to its alcohol counterpart (**18m** *Pf*PK6 $IC_{50} = 292 \pm 22$ nM). Taken together, these results establish that a range of hydrophobic groups can be utilized in the 4-position of the pyrimidine core. Further exploration of additional polar functional groups will be needed to identify any polar moieties that can maintain or improve potency.

In conclusion to the SAR analysis performed on the aryl substituent of the pyrimidine-4-position, we have identified the 5-chlorothiophene substituent of **18j** as a possible bioisostere of the benzothiophene with equipotent activity on *Pf*GSK3 and with enhanced potency on *Pf*PK6 in comparison to the parent, **IKK16**. Based on the latter result, **18j** was considered a better lead compound, demonstrating enhanced *Pf*PK6 ligand efficiency compared to **IKK16** (**18j** $LE_{PfPK6} = 0.30$ and **IKK16** $LE_{PfPK6} = 0.26$, Table S1).

The final position to be investigated in our SAR study was the 5-position of the pyrimidine (Table 4). Based on the putative binding mode of this pyrimidine inhibitor scaffold, inferred from observing different crystal structures (e.g., PDB: 2P33, 6VNE, and 2JKK), we expect this position to be proximal to the kinase gatekeeper residue. This prompted us to investigate the prospect of engaging the gatekeeper residues of

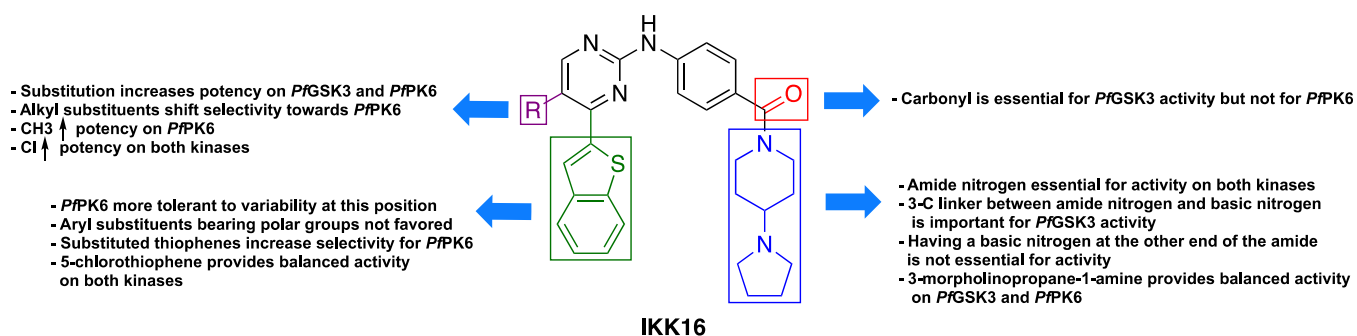


Figure 4. Summary of SAR study on IKK16.

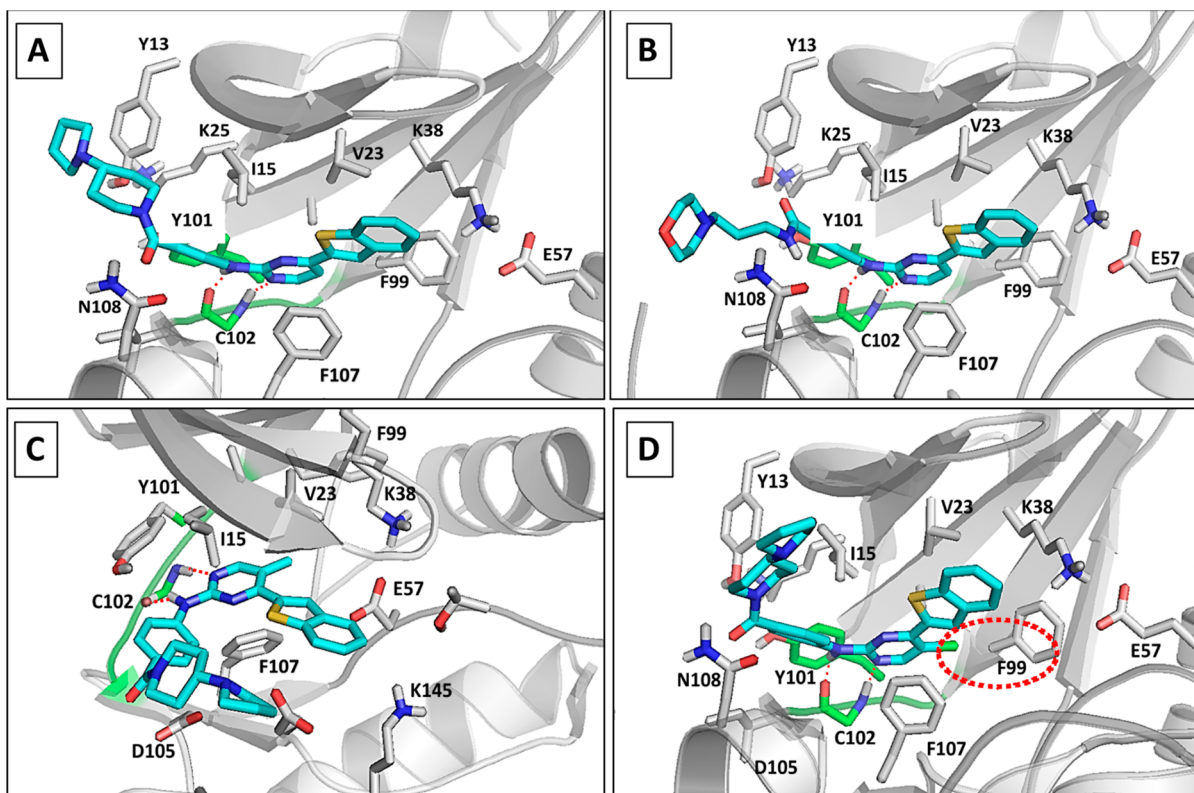


Figure 5. Homology model and docking studies of *PfPK6* inhibitors. Proposed binding modes of **IKK16** (A), **9g** (B), **18n** (C), and **18r** (D) in the developed homology model of *PfPK6*. Protein backbone is shown as a gray cartoon and the important residues in the binding site are displayed as gray sticks. Ligands are shown as cyan sticks with important polar contacts displayed as red dashed lines. The hinge region is colored green, and the hydrophobic contact between the 4-chloropyrimidine of **18r** and F99 is indicated by a red dashed circle (D).

PfGSK3 and *PfPK6* (Met157 and Phe99, respectively) through various substituents while observing the effect on binding. We decided to add electron-donating, electron-withdrawing, as well as neutral substituents to this position. **IKK16** analogues were synthesized bearing a methyl, cyclopropyl, methoxy, fluoro, chloro, bromo, and nitrile substituents at the 5-position of the pyrimidine core (**18n–t**, respectively). To our delight, all the synthesized analogues showed increased activity on *PfPK6* with IC_{50} values between 19 and 294 nM. The most potent compounds were **18n**, with a methyl substituent, and **18r**, with a chloro substituent, showing a 9- and 20-fold increase in potency compared to the parent **IKK16**, respectively (**18n** *PfPK6* IC_{50} = 45 ± 2 nM and **18r** *PfPK6* IC_{50} = 19 ± 3 nM). These 5-position substituents led to substantial variation in *PfGSK3* activity. Whereas all the alkyl-containing substituents displayed low activity on *PfGSK3*, all the halogenated analogues showed considerable

activity. The chloro-substituted **18r** and the bromo-substituted **18s** analogues were more active than the fluoro analogue **18q**, with IC_{50} values of 174 ± 16 and 165 ± 18 nM for chloro and bromo, respectively, and 456 ± 43 nM for fluoro. One can speculate that this ~3-fold enhancement arises from a putative halogen bonding interaction between the halogen atoms of these compounds and the sulfur atom of the methionine gatekeeper residue of *PfGSK3*, where several examples of this type of interaction have been reported in the literature.^{31–33}

Taken together, SAR analysis of the pyrimidine-5-position indicated that substitution in this region can be productive for modulating kinase activity and that there are differences in how *PfGSK3* and *PfPK6* respond to these changes. These results suggest that the *PfGSK3* catalytic site poorly tolerates alkyl substituents and prefers halogens, especially chloro and bromo. On the other hand, both alkyl-based substituents and halogens

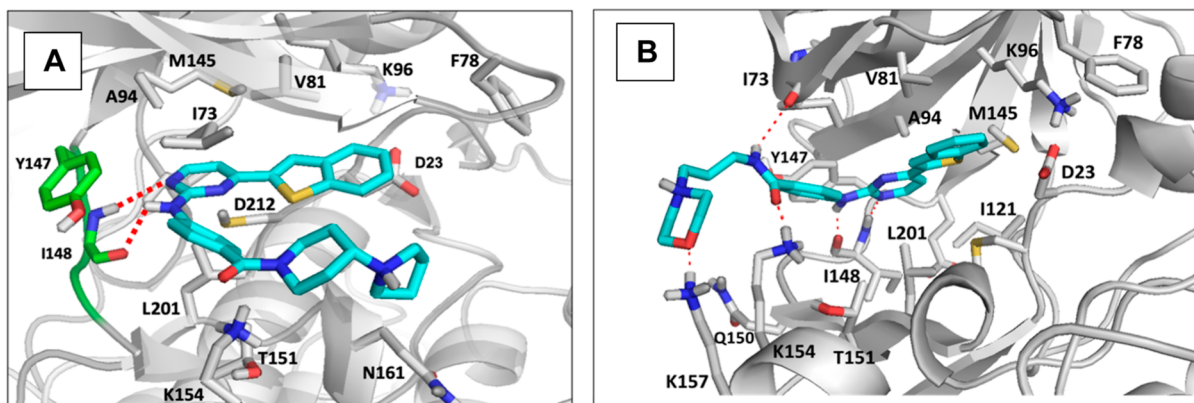
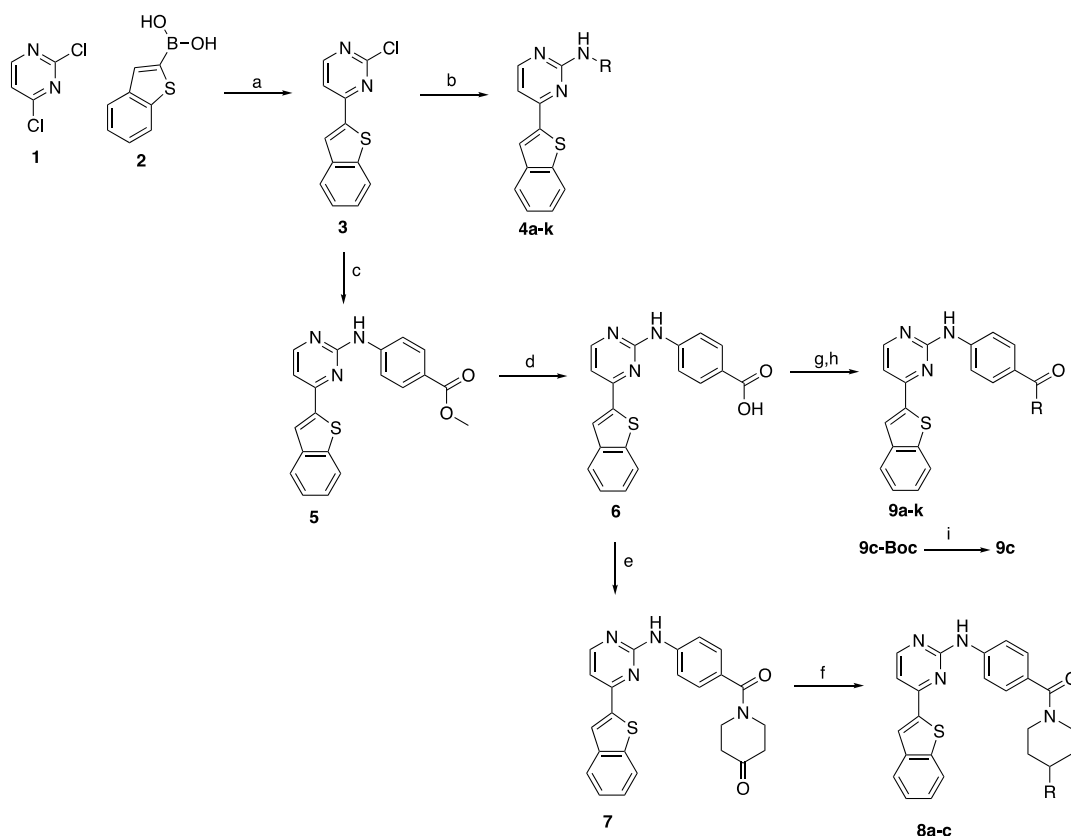


Figure 6. Proposed binding modes of IKK16 (A) and **9g** (B) in the developed homology model of *PfGSK3*. Protein backbone is shown as a gray cartoon and the important residues in the binding site are displayed as gray sticks. Ligands are shown as cyan sticks with important polar contacts displayed as red dashed lines.

Scheme 1. Synthesis of 2-Anilino-4-aryl Pyrimidine Analogues of IKK16^a

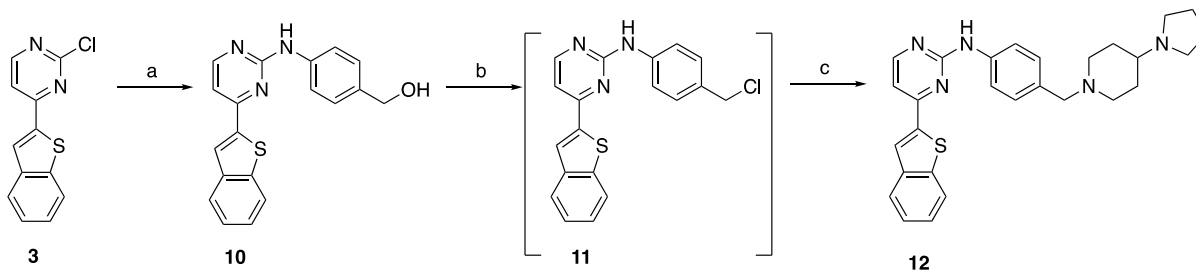


^aReagents and conditions: (a) $\text{Pd}(\text{PPh}_3)_4$, Na_2CO_3 , toluene/EtOH/water, 70 °C, and 79%; (b) aniline/amine, 1 N HCl, EtOH, μW , 160 °C, and 8–72%; (c) methyl-4-aminobenzoate, neat, 200 °C, and 40%; (d) 1 M LiOH, MeOH/THF, reflux, and 62%; (e) piperidine-4-one hydrochloride, EDCl, HOBT, DMF, TEA, rt, and 31%; (f) amine, $\text{NaBH}(\text{OAc})_3$, THF, rt, and 10–20%; (g) amines, EDCl, HOBT, DMF, TEA, rt (or 80 °C), and 5–58%; (h) HATU, TEA, DMF, 80 °C, and 4%; and (i) 4 M HCl/dioxane, DCM/MeOH, rt, and 42%.

enhanced the potency on *PfPK6*. The findings of our SAR study are summarized in Figure 4.

Evaluation of Inhibitors' Binding Mode in *PfPK6* and *PfGSK3* Catalytic Sites Using Homology Modeling and Docking Studies. To gain insights into the binding mode of the developed series of compounds in *PfPK6* and *PfGSK3*, we performed a molecular docking simulation into their ATP binding site. Starting with *PfPK6*, and since its crystal structure is not available, we built a homology model of the plasmodial

enzyme based on the human cyclin-dependent kinase-2 (CDK2).³⁴ The latter is an extensively studied kinase with several crystal structures in both active and inactive states. *PfPK6* shows a high degree of sequence similarity to CDK2, with greatly conserved structural features including their ATP-binding sites (Table S5). The ATP binding site of *PfPK6* is relatively narrower due to the presence of two tyrosine residues at its entry, which are not present in CDK2.³⁴ The plasmodial

Scheme 2. Synthesis of **12**^a

^aReagents and conditions: (a) 4-aminophenylmethanol, Pd₂(dba)₃, xantphos, Cs₂CO₃, dioxane, 90 °C, and 26%; (b) SOCl₂, DIPEA, THF, and rt; (c) 4-(pyrrolidin-1-yl)piperidine, K₂CO₃, DMF, rt, and 13%.

kinase has a U-shaped mostly hydrophobic adenine binding site limited on both sides with polar residues.

We first docked the parent compound **IKK16** into the ATP binding site of the *Pf*PK6 model, followed by docking the most potent analogues from Tables 2–4, **9g**, **18n**, and **18r**. All compounds bound in the expected orientation with their 2-anilino-pyrimidine group occupying the putative adenine binding site and making the canonical H-bond interactions with the backbone carbonyl and NH groups of Cys102 in the hinge region (Figure 5A–D). The pyrimidine core is anchored into this orientation by a hydrophobic clamp formed by Ile15 and Phe107. The benzothiazole group extends toward the hydrophobic back pocket and is near the polar residues involved in catalysis, Lys38 and Glu57. Extending from the pyrimidine 2-position of the molecules, the flexible pyrrolidino-piperidine or morpholino-propane chain protrude from the binding site toward the solvent-exposed region, making polar contacts with Asp105 and Asn108. The superior activities of **18n** and **18r** may be attributable to the presence of the 5-methyl or 5-chloro group on the pyrimidine nucleus, which is able to make favorable hydrophobic contacts with the gatekeeper residue Phe99 (referred to using a red dashed circle in Figure 5D).

Then, we turned our attention to studying the molecular interactions of these compounds in the catalytic site of *Pf*GSK3. Again, the crystal structure of this enzyme is not available, so we constructed a homology model based on the human GSK-3 β enzyme which shows a great degree of sequence similarity and a conserved ATP-binding site³⁵ (Table S5). The model was built using multiple templates of the human enzyme to obtain a highly accurate sequence alignment. The parent compound, **IKK16**, was docked into the putative ATP-binding site of the developed model (Figure 6A). The 2-anilino-pyrimidine nucleus of **IKK16** docks in the expected orientation of anilino-pyrimidines, forming the critical H-bonds with the backbone of Ile148 in the hinge region. It is also anchored in its place via hydrophobic interactions from both sides with Ala94 and Leu201. The benzothiazole ring fits into the hydrophobic back pocket stabilized by the interaction with Val81. Extending from the pyrimidine 2-position, the pyrrolidine ring extends toward the solvent, approaching Lys154 and Gln198 but does not appear to make significant interactions with these residues.

We followed by docking **9g**, which had also shown high activity against *Pf*GSK3, into our homology model (Figure 6B). The 2-anilino-pyrimidine nucleus of **9g** docks in the same orientation as that of **IKK16**, forming the critical H-bonds with the backbone of Ile148 in the hinge region. It is also anchored in its place via hydrophobic interactions with Ala94 and

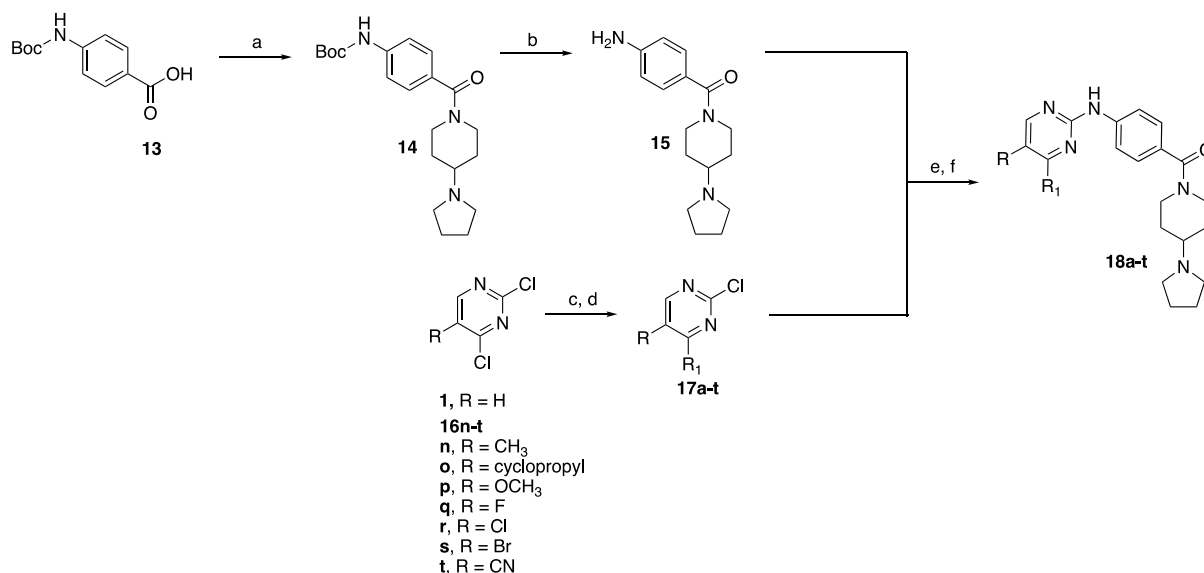
Leu201. Again, the benzothiazole ring fits into the hydrophobic back pocket stabilized by the interaction with Val81. Interestingly, the major difference from **IKK16** is that the morpholine ring and its linker extend toward the solvent, making an extensive network of H-bonds with Ile73, Lys154, and Lys157, which appears to contribute to the binding pose stability. These observations could potentially explain the greater activity of this compound compared to the parent molecule **IKK16** and warrants further SAR exploration.

Chemistry. The first step in the synthesis of **IKK16** analogues involved a Suzuki reaction utilizing 2,4-dichloropyrimidine (**1**) and benzo[*b*]thiophen-2-ylboronic acid (**2**) to afford the aryl halide intermediate **3** in very good yield (79%) (Scheme 1). From **3**, various analogues were synthesized using different anilines and amines as nucleophiles in a nucleophilic aromatic substitution (S_NAr) reaction with catalytic amounts of 1 M HCl solution to afford the corresponding 2,4-disubstituted pyrimidines **4a–h** in low to good yield (8–72%).

Compounds **8a–c** and **9a–k** were prepared starting with the aryl halide **3**, which was first subjected to neat S_NAr conditions using methyl-4-aminobenzoate as the nucleophile at 200 °C to afford the corresponding ester **5** in 40% yield²¹ (Scheme 1). Saponification of **5** using aqueous LiOH in refluxing MeOH/THF as a solvent generated the carboxylic acid **6** in 62% yield. Carboxylic acid **6** was used as an intermediate to generate amide analogues of **IKK16** in which the pyrrolidine ring was modified, or in which the 4-(pyrrolidin-1-yl)piperidine moiety was replaced all together. This was achieved using two different approaches. First, compounds **8a–c** were generated by coupling **6** with piperidine-4-one to generate the ketone **7**, followed by a reductive amination reaction using the desired amines with NaBH(OAc)₃ as the reducing agent (10–20% yield). Compounds **9a–k** were obtained via direct coupling of various cyclic, linear, aliphatic, or aromatic amines with the acid **6**, utilizing either EDCI/HOBt or HATU as the coupling reagents. The corresponding amides were obtained in low to good yield (5–58%).

Compound **9c**, bearing a piperazine ring at the amide position, was generated via deprotection of the Boc-protected precursor **9c-Boc** under acidic conditions.

Compound **12**, an analogue of **IKK16** lacking the amide carbonyl moiety, was also synthesized from the common intermediate **3** (Scheme 2). A Buchwald–Hartwig amination between intermediate **3** and 4-aminophenylmethanol using Pd₂(dba)₃ and xantphos afforded the corresponding alcohol **10** in 26% yield.³⁶ Afterward, SOCl₂-mediated chlorination of the alcohol resulted in the alkyl halide **11**, which was carried to the next step without isolation. Finally, addition of 4-(pyrrolidin-1-

Scheme 3. Convergent Synthesis Leading to 4- and 5-Substituted Pyrimidine Analogues of IKK16^a

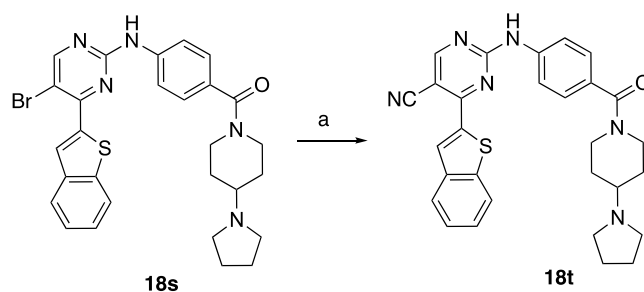
^aReagents and conditions: (a) 4-(pyrrolidin-1-yl)piperidine, HATU, TEA, DMF, rt, and 80%; (b) 4 M HCl/dioxane, DCM, rt, and 67%; (c) Pd(PPh₃)₄, boronic acid, Na₂CO₃, toluene/EtOH/water (or dioxane/water), 70 °C, and 8–66%; (d) 2-(tributylstannyl)thiazole, Pd(Ph₃P)₄, LiCl, DMF, 70 °C, and 10%; (e) TFA, 2,2,2-trifluoroethanol, μW, 140 °C, and 8–64%; and (f) Pd(OAc)₂, BINAP, Cs₂CO₃, dioxane, 80 °C, and 9–19%.

yl)piperidine to **11** in an S_N2 reaction using K₂CO₃ as the base resulted in the desired diamine **12**³⁷ (13% yield).

The next series of compounds were synthesized to probe substitutions at the 4- and 5- positions of the core pyrimidine ring. We employed a convergent synthesis in which pyrimidines **17a–t**, bearing the desired substituents, were prepared and then reacted with (4-aminophenyl)(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**15**) in a microwave-assisted reaction (Scheme 3). First, 4-((*tert*-butoxycarbonyl)-amino)benzoic acid (**13**) was coupled with 4-(pyrrolidin-1-yl)piperidine using HATU to afford the Boc-protected amide **14** in 80% yield (Scheme 3). Acid-catalyzed deprotection of the Boc group followed by basic work up afforded the aniline **15** in very good yield (67%). The substituted 2-chloropyrimidine intermediates **17a–t** were prepared via a Suzuki reaction using either 5-substituted or unsubstituted 2,4-dichloropyrimidines with the appropriate boronic acids to afford the desired products in 8–66% yield. The thiazole intermediate, **17d**, was prepared using Stille conditions utilizing 2-(tributylstannyl)thiazole and 2,4-dichloropyrimidine to afford the desired product in low yield (10%). Finally, reacting aniline **15** with **17a–t** (except **17l** and **17p**) using TFA in 2,2,2-trifluoroethanol resulted in analogues **18a–t** in 8–64% yield.³⁸ For **18l** and **18p**, aniline **15** was reacted with **17l** and **17p** under Buchwald–Hartwig conditions using Pd(OAc)₂ as the catalyst and *rac*-BINAP as the ligand to afford the desired analogues in 9–19% yield.

Compound **18t**, bearing a nitrile group at the 5-position of the pyrimidine, was synthesized in a different manner from the rest of its congeners (Scheme 4). Briefly, the brominated analogue, **18s**, was treated with Zn(CN)₂ as a cyanide source, Pd(dppf)₂Cl₂ as a catalyst, and diisopropylethylamine as a base and then heated to 170 °C in DMF under microwave conditions. This procedure afforded **18t** in 20% yield.³⁹

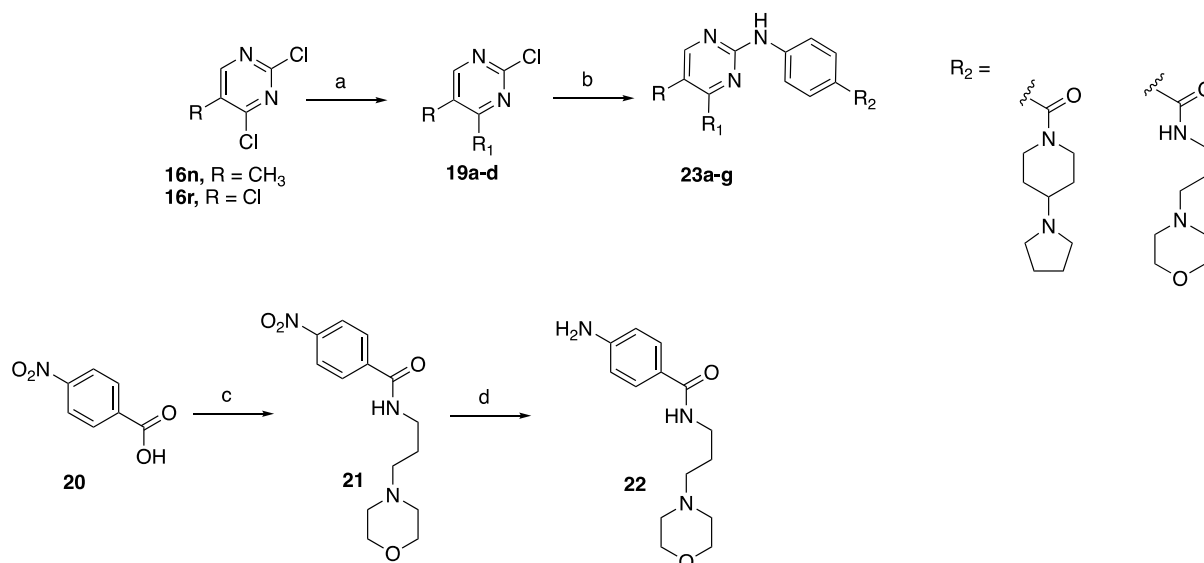
Design of Hybrid Analogues. The initial SAR study on IKK16 (Tables 1–4) led to identification of analogues with enhanced enzymatic potency on both PfGSK3 and PfPK6. To

Scheme 4. Conversion of 5-Bromo to 5-Nitrile^a

^aReagents and conditions: (a) Zn(CN)₂, Pd(dppf)₂Cl₂, DIPEA, DMF, 170 °C, μW, and 20%.

drive the enzymatic potency even higher, we envisioned designing a second set of analogues based on combining the structural features of the best performing derivatives from Tables 1–4. Compounds chosen were **9g** (Table 2), **18i** and **18j** (Table 3), and **18n** and **18r** (Table 4). The hybrid analogues would combine substitutions on the pyrimidine-4-position with those on the 5-position, as well as the amide region, in hope that this would lead to a synergistic effect, resulting in greater inhibition of the target kinases. The synthesized compounds were tested for in vitro enzymatic activity against PfGSK3 and PfPK6. In addition, we calculated the ligand efficiency values of these new hybrid analogues based on their PfPK6 potencies (Tables 5 and S1).

We first combined the (5-methylthiopheno) substituent from the 4-position pyrimidine core with the 5-methyl and 5-chloro pyrimidine substituents, keeping the 4-(pyrrolidin-1-yl)piperidine tail from the pyrimidine 2-position as it is, culminating in **23a** and **23b**, respectively (Table 5). Consistent with previous findings, we observed that a methyl at the 5-position of the pyrimidine increases preference for PfPK6 over PfGSK3. This can be observed in **23a**, which demonstrated 28 ± 3 nM IC₅₀ at PfPK6, while activity at PfGSK3 in a screen at 1 μM did not reach our threshold for determining an IC₅₀

Scheme 5. Synthesis of Hybrid Analogues^a

^aReagents and conditions: (a) boronic acid, Pd(PPh₃)₄, Na₂CO₃, toluene/EtOH/water, 70 °C, and 38–50%; (b) **15** or **22**, TFA, 2,2,2-trifluoroethanol, μ W, 140 °C, and 6–15%; (c) 3-morpholinopropan-1-amine, HATU, DIPEA, 40 °C, and 29%; and (d) Fe, NH₄Cl, EtOH/water, and reflux.

value. On the other hand, the chloro analogue **23b** was merely 9-fold selective for PfPK6. The activity of **23b** on both kinases was comparable to that of most potent analogue from the first set, **18r**.

Next, we combined the pyrimidine 5-position methyl and chloro substituents with (5-chlorothiophene) at the 4-position of the pyrimidine forming **23c** and **23d** (Table 5). Again, the same pattern of selectivity appears. Compound **23c** showed micromolar activity at PfGSK3 while showing a 14 ± 1 nM IC₅₀ at PfPK6. Compound **26d**, on the other hand, was less selective, showing IC₅₀ values of 172 ± 13 nM at PfGSK3 and 11 ± 1 nM at PfPK6.

Encouraged by these findings, we followed up by making the 3-morpholinopropan-1-amine analogues of **23b–d**. We chose to make these because this side chain had shown enhancement of enzymatic activity on both kinases in our first analogue set (Table 2). The resulting new analogues **23e–g** demonstrated excellent potency on PfPK6, with **23e** displaying an IC₅₀ value of 8 ± 1 nM. As for PfGSK3 activity, **23e** and **23f** are the first analogues reported in this study to show less than 100 nM IC₅₀ against this kinase. Analogue **23g** did not show significant activity on PfGSK3.

In addition to their high potency, all the compounds from Table 5 showed PfPK6 LE values greater than the recommended minimum of 0.3.⁴⁰ Additionally, **23a–g** also exhibited PfPK6 LE values greater than those of their parent congeners, as well as IKK16. This provided an indication (beyond potency alone) that our medicinal chemistry optimization campaign was heading in the appropriate direction.

Synthesis of Hybrid Analogues. The hybrid compounds **23a–g** were synthesized using a similar method to the one used for the first set of analogues (Scheme 5). Briefly, commercially available pyrimidines **16n** and **16r** were subjected to Suzuki conditions with appropriate boronic acids to afford **19a–d** in 38–50% yield. Concurrently, 4-nitrobenzoic acid (**20**) was coupled with 3-morpholinopropan-1-amine using HATU, giving amide **21** in 29% yield.

Compound **21** was reduced to the aniline intermediate using Fe and NH₄Cl under refluxing conditions. The resulting aniline **22** was carried to the next step without purification. The final analogues were obtained using microwave-assisted reactions, coupling **19a–d** with either **15** or **22** to afford **23a–g** in low yield (6–15%).

Activity on Blood Stage Parasites. All the synthesized compounds were evaluated for inhibition of *P. falciparum* 3D7 load in erythrocytes utilizing a SYBR green assay.⁴¹ Compounds of interest were initially screened at 1 μM concentration, and only compounds that inhibited >75% parasite load were considered active and subsequently evaluated in a dose–response study to determine their EC₅₀ values (Table 6). Concurrently, we evaluated the analogues for cytotoxicity against HepG2 cells, a human hepatoma cell line, using a commercially available CellTiter-Glo assay (Promega). This assay is a commonly used proxy for evaluating cytotoxicity against human cells and has been previously used to evaluate cytotoxicity among the antimalarial hits from TCAMS.⁴² The cytotoxicity data is presented as % viability after incubation with 200 nM or 2 μM of the test compounds.

Compounds which demonstrated >75% activity against *P. falciparum* blood stage parasites in the primary screen at 1 μM were **12**, **18b**, **18m–o**, **18q–t**, and **23c–f**. Follow-up dose–response experiments for these compounds resulted in a wide range of EC₅₀ values ranging from 60 to 2600 nM (Table 6). Compound **12** (EC₅₀ = 60 ± 8 nM) showed the greatest antiparasitic effect, surpassing IKK16 (EC₅₀ = 280 ± 2 nM). Analogues **18s** and **18t** demonstrated comparable activity to IKK16 despite showing improved kinase inhibition. However, these compounds also showed some cytotoxic effect at 2 μM concentration.

All the screened compounds did not show significant toxicity at 200 nM, with all of them resulting in >80% viability after incubation with the cells (Table 6). However, at 2 μM, we started to observe a trend where analogues which had a 5-position substituent on the pyrimidine ring tend to exhibit greater cytotoxicity (**18n–s**). This can be exemplified by

observing the difference in cytotoxicity between **IKK16** and **18n** (which only differs by a methyl group at the pyrimidine-5-position), in which **18n** shows ~40% more cytotoxicity than **IKK16**. Compound **23d** showed the greatest cytotoxic effect (34% viable cells) at 2 μM , indicating that it may have a small window between its antiparasmodial EC_{50} and CC_{50} .

This cellular activity data indicated that the improvement in potency we observed in the in vitro enzymatic assay for these analogues did not necessarily translate into a significant improvement in the antiparasitic activity compared to the reference, **IKK16**. This could be attributed to multiple factors, such as differences related to kinase activity in an in vitro biochemical assay versus kinase activity in live parasite isolates, differing off-target profiles, as well as differences in physicochemical properties that may hamper access to parasites, and/or intracellular stability.

Given these results, we were interested in assessing some of the physicochemical properties of these compounds and how they compare to the parent **IKK16**, hoping that this could offer some insight. To do this, we chose our lead kinase inhibitors, **23b–f**, and subjected them to a preliminary characterization of their kinetic solubility and effective permeability in the PAMPA assay (Table S2). The compounds showed a wide range of solubility values. Analogues **23b** and **23d** exhibited greater solubility than **IKK16** with values of 56.3 and 55.3 $\mu\text{g}/\text{mL}$ versus 19.4 $\mu\text{g}/\text{mL}$, respectively. Interestingly, **23e** and **23d** were almost completely insoluble using this assay format. This may be due to the lower basicity of the morpholine ring which reduces the ionized fraction of the compounds at physiological pH, thus lowering their overall aqueous solubility.

In the membrane permeability assay, all the analogues tested demonstrated much lower permeability than **IKK16**, which indicated that the physicochemical properties of these compounds need to be further optimized (Table S2).

Activity on Liver Stage Parasites. We next checked to see if any of our novel kinase inhibitors displayed activity against the liver stage parasites. We chose compounds that had shown <20 nM IC_{50} at either *PfGSK3* or *PfPK6* in hopes that this potency would translate to noticeable antiparasitic activity in liver cells. We utilized the common rodent malaria model *Plasmodium berghei* ANKA to infect human HepG2 liver cells. For this assay, the hepatocyte viability in the presence of inhibitors was evaluated in parallel with assessing their effect on the *P. berghei* parasite load (Table S4).

Based on their superior potency against *PfGSK3/PfPK6*, the compounds chosen for this assay were **23b–e**. Unfortunately, none of the compounds exhibited a *P. berghei* EC_{50} that was 5-fold greater than the HepG2 CC_{50} (Table S4). Due to this low activity window, antiparasitic effects could not be distinguished from hepatotoxic action. Accordingly, the compounds were considered inactive against liver stage parasites.

Evaluation of Kinase Selectivity Scores. Compounds with the best kinase inhibitory profile from our first set of analogues, **9g**, **18n**, and **18r**, together with the lead hybrid analogues **23d** and **23e** were screened against 468 human kinases using the DiscoverX KINOMEScan platform to determine the selectivity for their kinase targets (Table 7 and Supporting Information). Compounds were screened at 1 μM concentration, and their selectivity scores S_{10} (1 μM) were determined. Examining the selectivity scores, we noticed that substitution on the 5-position of the pyrimidine led to an

increase in the number of human kinase targets inhibited by the analogues, compared to the parent molecule, **IKK16**. This can be observed in the high level of promiscuity of **18n** and **18r**, both of which have 5-position substituents and showing S_{10} (1 μM) values of 0.25 and 0.23, respectively, an almost 2-fold reduction in selectivity. Interestingly, replacing the pyrrolidino-piperidine tail group of **IKK16** with 3-morpholinopropan-1-amine resulted in fewer kinase target hits and accordingly a smaller S_{10} (1 μM) value for **9g** (S_{10} (1 μM) = 0.08). The hybrid compound **23d** showed the highest level of promiscuity, hitting 103 kinase targets of the 468 kinases screened in the assay. On the other hand, replacing the pyrrolidino-piperidine tail group of **23d** with 3-morpholinopropan-1-amine (**23e**) reduced the promiscuity by almost 40%, going from 103 to 64 kinase hits. The same pattern was observed when comparing **IKK16** with **9g**. Taken together, these results indicated that the substituent at the 5-position of the pyrimidine could be generating a common pharmacophore that is recognized by more human kinases than the unsubstituted pyrimidine, while the 3-morpholinopropan-1-amine tail leads to analogues with reduced promiscuity against human kinases screened in this panel. This underscores the need for further exploration and optimization of these analogues by utilizing the information we gained from the SAR study to increase the selectivity for their kinase targets.

CONCLUSIONS

With the noticeable rise in resistance to traditional first-line antimalarial therapy, we need new methods to combat the deadly malaria parasite. The emergence of plasmodial kinases as viable targets for antimalarial drugs has prompted the scientific community to pursue plasmodial kinase inhibition as an avenue to discover medicines with new modes of action for combating plasmodial infections. By screening a library of known kinase inhibitors against 11 plasmodial kinases, we identified **IKK16**, a sub-micromolar inhibitor of *PfGSK3* and *PfPK6*, which was also active against blood stage parasites. Given the beneficial prospect of dual kinase inhibition, we conducted an extensive medicinal chemistry campaign focusing on establishing SAR around the **IKK16** scaffold as well as identifying analogues with enhanced *PfGSK3/PfPK6* inhibitor activity and reduced promiscuity. Our SAR study focused on exploring and modifying three different positions of the **IKK16** scaffold. We observed that activity against *PfGSK3* was typically more dramatically affected by these modifications, suggesting that, for this scaffold, its catalytic pocket is perhaps less tolerant of changes than that of *PfPK6*, which tolerated many of the structural modifications that were applied. In general, many of the analogues synthesized showed some degree of preference for *PfPK6* over *PfGSK3*. We were able to identify the 5-chlorothiophene group as a suitable bioisostere of the bulky benzothiophene, having lower molecular weight and leading to better ligand efficiency. We noticed that the preference for *PfPK6* may be modulated with certain substitutions, while the same could not be achieved for *PfGSK3*. This was most noticeable with the 5-position substituents, where a 5- CH_3 resulted in almost a 16-fold preference for *PfPK6*, while the 5-chloro substituent was only 9-fold selective (**18n** vs **18r**, respectively). The greatest enhancement of kinase inhibition potency was also achieved by substitution on the pyrimidine-5-position. Building upon this finding, we identified multiple potent inhibitors of *PfGSK3* and *PfPK6*, most of which demonstrated greater

kinase inhibition than the hit compound, **IKK16**. However, the 5-position substituent on the pyrimidine core also appears to be favored by many other kinase targets, as evidenced by the increased promiscuity of analogues having this substitution. In addition to their kinase activity, many of the analogues reported herein inhibited parasitemia, although with varying potencies ranging from micromolar to low nanomolar. Screening of the most potent kinase inhibitors identified, **23b–e**, against *P. berghei* liver stage parasites, showed that their antiparasitic activity may not be *Pf* kinase dependent but rather a result of non-specific hepatotoxicity. By assessing some of the physicochemical properties of these compounds, we found that **23b–d** showed acceptable kinetic solubility, while the morpholine analogues **23d** and **23e** were highly insoluble. On the other hand, all these analogues demonstrated poor apparent permeability compared to **IKK16**. This may, in part, contribute to poor compound accumulation in the erythrocytes and thus poor target engagement in the parasite. Analogues **23d** and **23e** were the most promising analogues identified in terms of dual kinase inhibition, ligand efficiency, and their modest cellular activity against blood stage parasites. To the best of our knowledge, **23d** is among the few dual *Pf*GSK3/*Pf*PK6 inhibitors with nanomolar potency against both kinases as well as nanomolar cellular activity against blood stage parasites (**23d** *Pf*GSK3 IC₅₀ = 172 ± 13 nM, *Pf*PK6 IC₅₀ = 11 ± 1 nM, and blood stage EC₅₀ = 552 ± 37 nM).

Although they have several favorable features as described above, **23d** and **23e** suffer from high levels of kinase promiscuity, originating in part from the 5-Cl substituent on the pyrimidine core. This promiscuity may also be a contributing factor to the modest cytotoxicity observed for these compounds. Our future work will include further modifications of the structure of these two hits that explore additional functional groups based on the knowledge from our SAR study, with the aim of reducing their kinase promiscuity while increasing the on-target potency. We also plan to work on improving their physicochemical properties, especially membrane permeability and solubility, to enhance the drug-likeness of these novel compounds and allow us to move them to more advanced malaria efficacy models.

EXPERIMENTAL SECTION

Chemistry. All reagents and solvents used were purchased from commercial sources and were used without further purification. NMR spectra were obtained using a Bruker 850 MHz or INOVA 400 MHz spectrometers at room temperature; chemical shifts are expressed in parts per million (ppm, δ units) and are referenced to the residual protons in the deuterated solvent used. Coupling constants are given in units of hertz (Hz). Splitting patterns describe apparent multiplicities and are designated as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), and br s (broad singlet), dd (doublet of doublets), ddd (double double doublet), and tt (triplet of triplets). The purity of compounds submitted for biological screening was determined to be $\geq 95\%$ as measured by high-performance liquid chromatography (HPLC). Analytical thin-layer chromatography (TLC) was performed on silica gel plates, 200 μm , with an F254 indicator. Column chromatography was performed using RediSep Rf preloaded silica gel cartridges on Isolera one Biotage automated purification systems. Samples for high-resolution mass spectrometry were analyzed with a Thermo Fisher Q Exactive HF-X (Thermo Fisher, Bremen, Germany) mass spectrometer coupled with a Waters Acquity H-class liquid chromatograph system. Samples were introduced via a heated electrospray source (HESI) at a flow rate of 0.3 mL/min. Electrospray source conditions were set as follows: spray voltage 3.0 kV, sheath gas (nitrogen) 60 arb, auxiliary gas

(nitrogen) 20 arb, sweep gas (nitrogen) 0 arb, nebulizer temperature 375 °C, capillary temperature 380 °C, and RF funnel 45 V. The mass range was set to 150–2000 *m/z*. All measurements were recorded at a resolution setting of 120,000. Separations were conducted on a Waters Acquity UPLC BEH C18 column (2.1 × 50 mm, 1.7 μm particle size). LC conditions were set at 95% water with 0.1% formic acid (A) ramped linearly over 5.0 min to 100% acetonitrile with 0.1% formic acid (B) and held until 6.0 min. At 7.0 min, the gradient was switched back to 95% (A) and allowed to re-equilibrate until 9.0 min. The injection volume for all samples was 3 μL . Analytical LC/MS data was obtained using a Waters Acquity ultrahigh-performance liquid chromatography (UPLC) system equipped with a photodiode array (PDA) detector using the following method: solvent A = water + 0.2% FA, solvent B = ACN + 0.1% FA, and flow rate = 1 mL/min. The gradient started at 95% A for 0.05 min. Afterward, it was ramped up to 100% B over 2 min and held for an additional minute at this concentration, before returning to the initial gradient. Compounds were purified on prep HPLC using an Agilent 1100 equipped with a Phenomenex column (phenyl–hexyl, 75 × 30 mm, 5 μm) using the following method: solvent A: water + 0.05% TFA; solvent B: MeOH; and flow rate: 70.00 mL/min. LC conditions were set at 90% (A) ramped linearly over 8.0 min to 100% (B) and held until 10.0 min at 100% B. At 10.0 min, the gradient was switched back to 90% (A).

General Procedure A: Suzuki Reaction. 4-(Benzo[*b*]thiophen-2-yl)-2-chloropyrimidine (**3**). 2,4-Dichloropyrimidine (**1**) (1.30 g, 1.0 equiv, 8.40 mmol), benzo[*b*]thiophen-2-ylboronic acid (1.50 g, 1.0 equiv, 8.4 mmol) (**2**), and Na₂CO₃ (2.70 g, 3 equiv, 25.00 mmol) were dissolved in a mixture of toluene (37.00 mL), EtOH (9.250 mL), and water (9.250 mL). The solvent was degassed thoroughly by bubbling argon gas through it for 5 min. Pd(PPh₃)₄ (0.78 g, 0.08 equiv, 0.67 mmol) was added to the reaction mixture, and it was heated to 70 °C and stirred overnight. The reaction mixture was cooled to room temperature and then poured onto water. The layers were separated, and the aqueous layer was then extracted with ethyl acetate (×3). The combined organic extracts were washed with brine, dried with Na₂SO₄, and filtered. The solvent was removed in vacuo. The crude compound was purified using silica gel chromatography using a gradient of hexanes and DCM 100:0 to 50:50 to afford the title material as a white solid (1.65 g, 79%). ¹H NMR (400 MHz, CDCl₃): δ 8.54 (d, *J* = 5.2 Hz, 1H), 8.06 (s, 1H), 7.86–7.74 (m, 2H), 7.53 (d, *J* = 5.2 Hz, 1H), 7.42–7.26 (m, 2H). ¹³C NMR (101 MHz, CDCl₃): δ 162.32, 161.90, 159.69, 141.79, 140.11, 139.80, 126.93, 126.29, 125.30, 125.26, 122.95, 114.54. LCMS (ESI+) *m/z*: 247 [M + H]⁺.

General Procedure B: Microwave-Assisted S_NAr. 4-(Benzo[*b*]thiophen-2-yl)-*N*-phenylpyrimidin-2-amine (**4a**). A mixture of 4-(benzo[*b*]thiophen-2-yl)-2-chloropyrimidine (0.05 g, 1 equiv, 0.20 mmol), aniline (0.02 g, 0.02 mL, 1.2 equiv, 0.24 mmol), and a catalytic amount of 1 N HCl solution was heated in EtOH (3.00 mL) for 1 h at 160 °C under microwave conditions. The solvent was removed in vacuo. The crude compound was purified by silica gel chromatography using a gradient of hexanes and ethyl acetate to afford the title material as a yellow solid (0.04 g, 60%). ¹H NMR (400 MHz, DMSO): δ 9.74 (s, 1H), 8.56 (d, *J* = 5.1 Hz, 1H), 8.38 (s, 1H), 8.09–8.04 (m, 1H), 7.97–7.91 (m, 1H), 7.90–7.84 (m, 2H), 7.51 (d, *J* = 5.2 Hz, 1H), 7.49–7.42 (m, 2H), 7.38–7.31 (m, 2H), 6.99 (tt, *J* = 7.3, 1.1 Hz, 1H). ¹³C NMR (101 MHz, DMSO): δ 159.91, 158.89, 158.84, 142.77, 140.39, 140.24, 139.85, 128.53, 126.16, 125.06, 124.97, 124.81, 122.89, 121.54, 118.93, 107.01. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₁₈H₁₄N₃S, 304.0903; found, 304.0892.

4-(Benzo[*b*]thiophen-2-yl)-*N*-(4-chlorophenyl)pyrimidin-2-amine (**4b**). This compound was synthesized according to general procedure B starting from **3** and 4-chloroaniline (0.06 g, 0.486 mmol) to afford the title material as a yellowish-white solid (0.05 g, 36%). ¹H NMR (400 MHz, DMSO): δ 9.91 (s, 1H), 8.58 (d, *J* = 5.2 Hz, 1H), 8.39 (s, 1H), 8.08–8.03 (m, 1H), 7.97–7.92 (m, 1H), 7.92–7.86 (m, 2H), 7.54 (d, *J* = 5.2 Hz, 1H), 7.50–7.42 (m, 2H), 7.41–7.36 (m, 2H). ¹³C NMR (101 MHz, DMSO): δ 159.66, 158.94, 158.91, 142.56, 140.25, 139.84, 139.39, 128.37, 126.23, 125.23, 125.00,

124.85, 122.87, 120.39, 107.38. HRMS-ESI+ (m/z): $[M + H]^+$ calcd for $C_{18}H_{14}ClN_3S$, 338.0513; found, 338.0502.

4-(Benzo[*b*]thiophen-2-yl)-*N*-(3-chlorophenyl)pyrimidin-2-amine (4c). This compound was synthesized according to general procedure B starting from **3** and 3-chloroaniline (0.04 g, 0.290 mmol) to afford the title material as a yellowish-white solid (0.05 g, 55%). 1H NMR (400 MHz, DMSO): δ 9.98 (s, 1H), 8.60 (d, $J = 5.2$ Hz, 1H), 8.45–8.32 (m, 1H), 8.13 (t, $J = 2.1$ Hz, 1H), 8.08–8.02 (m, 1H), 7.98–7.90 (m, 1H), 7.75 (dd, $J = 8.4, 1.2$ Hz, 1H), 7.57 (d, $J = 5.2$ Hz, 1H), 7.51–7.42 (m, 2H), 7.35 (t, $J = 8.1$ Hz, 1H), 7.03 (dd, $J = 8.0, 1.2$ Hz, 1H). ^{13}C NMR (101 MHz, DMSO): δ 159.63, 159.00, 158.94, 142.50, 141.99, 140.27, 139.85, 133.09, 130.16, 126.32, 125.40, 125.06, 124.91, 122.89, 121.02, 118.13, 117.19, 107.67. HRMS-ESI+ (m/z): $[M + H]^+$ calcd for $C_{18}H_{14}ClN_3S$, 338.0513; found, 338.0503.

4-(Benzo[*b*]thiophen-2-yl)-*N*-(2-chlorophenyl)pyrimidin-2-amine (4d). 4-(Benzo[*b*]thiophen-2-yl)-2-chloropyrimidine (0.10 g, 1 equiv, 0.405 mmol) (**3**), 2-chloroaniline (0.05 g, 0.043 mL, 1 equiv, 0.405 mmol), *rac*-BINAP (0.03 g, 0.1 equiv, 0.041 mmol), and CS_2CO_3 (0.33 g, 2.5 equiv, 1.01 mmol) were stirred in dioxane (3.00 mL). The solvent was degassed by bubbling argon gas for 5 min. $Pd(OAc)_2$ (0.005 g, 0.05 equiv, 0.020 mmol) was added, and the reaction mixture was heated to 90 °C and stirred for 24 h. Afterward, the reaction mixture was cooled to room temperature and then filtered through Celite. The crude compound was purified using silica chromatography using a gradient of hexanes and ethyl acetate to afford the title material as a yellow solid (0.14 g, 61%). 1H NMR (850 MHz, DMSO): δ 8.91 (s, 1H), 8.52 (d, $J = 5.1$ Hz, 1H), 8.36 (s, 1H), 8.07–8.01 (m, 1H), 7.96–7.88 (m, 2H), 7.54 (dd, $J = 8.0, 1.5$ Hz, 1H), 7.52 (d, $J = 5.1$ Hz, 1H), 7.47–7.43 (m, 2H), 7.40 (td, $J = 7.6, 1.5$ Hz, 1H), 7.22–7.18 (m, 1H). ^{13}C NMR (214 MHz, DMSO): δ 160.19, 159.09, 158.97, 142.50, 140.31, 139.78, 136.28, 129.45, 127.38, 127.09, 126.14, 125.62, 125.29, 125.13, 124.93, 124.79, 122.85, 107.46. HRMS-ESI+ (m/z): $[M + H]^+$ calcd for $C_{18}H_{14}ClN_3S$, 338.0513; found, 338.0501.

4-(Benzo[*b*]thiophen-2-yl)-*N*-(4-methoxyphenyl)pyrimidin-2-amine (4e). This compound was synthesized according to general procedure B starting from **3** and 4-methoxyaniline (0.06 g, 0.486 mmol) to afford the title material as a yellow solid (0.10 g, 74%). 1H NMR (400 MHz, DMSO): δ 9.56 (s, 1H), 8.51 (d, $J = 5.1$ Hz, 1H), 8.35 (s, 1H), 8.09–8.00 (m, 1H), 7.96–7.88 (m, 1H), 7.75 (d, $J = 9.0$ Hz, 2H), 7.53–7.32 (m, 3H), 7.02–6.79 (m, 2H), 3.75 (s, 3H). ^{13}C NMR (101 MHz, DMSO): δ 160.00, 158.84, 158.78, 154.29, 142.93, 140.20, 139.86, 133.51, 126.10, 124.93, 124.86, 124.77, 122.86, 120.61, 113.75, 106.42, 55.19. HRMS-ESI+ (m/z): $[M + H]^+$ calcd for $C_{19}H_{16}N_3OS$, 334.1009; found, 334.0999.

4-(Benzo[*b*]thiophen-2-yl)-*N*-(3-methoxyphenyl)pyrimidin-2-amine (4f). This compound was synthesized according to general procedure B starting from **3** and 3-methoxyaniline (0.06 g, 0.486 mmol) to afford the title material as a yellow solid (0.02 g, 15%). 1H NMR (400 MHz, DMSO): δ 9.73 (s, 1H), 8.57 (d, $J = 5.2$ Hz, 1H), 8.39 (s, 1H), 8.12–8.02 (m, 1H), 8.00–7.89 (m, 1H), 7.61 (t, $J = 2.2$ Hz, 1H), 7.52 (d, $J = 5.2$ Hz, 1H), 7.49–7.37 (m, 3H), 7.23 (t, $J = 8.2$ Hz, 1H), 6.58 (ddd, $J = 8.2, 2.5, 0.9$ Hz, 1H), 3.81 (s, 3H). ^{13}C NMR (101 MHz, DMSO): δ 159.87, 159.60, 158.89, 158.83, 142.65, 141.59, 140.20, 139.83, 129.25, 126.18, 125.12, 125.00, 124.81, 122.90, 111.34, 107.12, 104.58, 55.07. HRMS-ESI+ (m/z): $[M + H]^+$ calcd for $C_{19}H_{16}N_3OS$, 334.1009; found, 334.0998.

4-(Benzo[*b*]thiophen-2-yl)-*N*-(2-methoxyphenyl)pyrimidin-2-amine (4g). This compound was synthesized according to general procedure B starting from **3** and 2-methoxyaniline (0.06 g, 0.486 mmol) to afford the title material as a yellow solid (0.06 g, 41%). 1H NMR (400 MHz, DMSO): δ 8.54 (d, $J = 5.2$ Hz, 1H), 8.38 (d, $J = 0.8$ Hz, 1H), 8.27 (dd, $J = 7.3, 1.7$ Hz, 1H), 8.17 (s, 1H), 8.09–8.02 (m, 1H), 7.98–7.90 (m, 1H), 7.52 (d, $J = 5.2$ Hz, 1H), 7.50–7.40 (m, 2H), 7.12–6.98 (m, 3H), 3.88 (s, 3H). ^{13}C NMR (101 MHz, DMSO): δ 159.78, 158.99, 158.98, 149.13, 142.53, 140.26, 139.83, 128.40, 126.19, 125.18, 124.97, 124.82, 122.91, 122.88, 120.43, 120.21, 110.90, 107.25, 55.80. HRMS-ESI+ (m/z): $[M + H]^+$ calcd for $C_{19}H_{16}N_3OS$, 334.1009; found, 334.0996.

4-(Benzo[*b*]thiophen-2-yl)-*N*-benzylpyrimidin-2-amine (4h). A mixture of **3** (0.10 g, 1 equiv, 0.41 mmol), phenylmethanamine (0.07 g, 0.07 mL, 1.5 equiv, 0.61 mmol), and TEA (0.08 g, 0.11 mL, 2.0 equiv, 0.81 mmol) in EtOH (5.00 mL) was heated for 30 min at 120 °C under microwave conditions. The solvent was removed in vacuo. The crude compound was purified by silica gel chromatography using a gradient of hexanes and ethyl acetate to afford the title material as a white solid (0.01 g, 8%). 1H NMR (400 MHz, DMSO): δ 8.35 (d, $J = 5.1$ Hz, 1H), 8.27 (s, 1H), 8.04–7.98 (m, 1H), 7.94–7.85 (m, 2H), 7.49–7.34 (m, 4H), 7.34–7.28 (m, 2H), 7.25 (d, $J = 5.2$ Hz, 1H), 7.23–7.18 (m, 1H), 4.56 (d, $J = 6.3$ Hz, 2H). ^{13}C NMR (101 MHz, DMSO): δ 162.19, 158.88, 140.41, 140.15, 139.81, 128.18, 127.41, 126.58, 125.91, 124.83, 124.65, 124.47, 122.81, 104.95, 44.04. HRMS-ESI+ (m/z): $[M + H]^+$ calcd for $C_{19}H_{16}N_3S$, 318.1059; found, 318.1049.

4-((4-(Benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)benzoate (5). A neat mixture of **3** (0.97 g, 1 equiv, 3.93 mmol) and methyl-4-aminobenzoate (0.71 g, 1.2 equiv, 4.72 mmol) was heated at 200 °C for 2 h. The resulting crude compound was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 80:20 to afford the title material as a yellow solid (0.57 g, 40%). 1H NMR (400 MHz, DMSO): δ 10.23 (s, 1H), 8.64 (d, $J = 5.2$ Hz, 1H), 8.42 (s, 1H), 8.12–8.06 (m, 1H), 8.03 (d, $J = 8.8$ Hz, 2H), 7.98–7.89 (m, 3H), 7.62 (d, $J = 5.2$ Hz, 1H), 7.53–7.37 (m, 2H), 3.84 (s, 3H). ^{13}C NMR (101 MHz, DMSO): δ 166.02, 159.50, 159.04, 158.96, 145.05, 142.42, 140.34, 139.85, 130.17, 126.27, 125.42, 125.02, 124.89, 122.96, 121.90, 117.90, 108.06, 51.74. HRMS-ESI+ (m/z): $[M + H]^+$ calcd for $C_{20}H_{16}N_3O_2S$, 362.0958; found, 362.0945.

4-((4-(Benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)benzoic acid (6). To a solution of **5** (1.60 g, 1 equiv, 4.43 mmol) in THF/MeOH (1:1, 70 mL) was added 1 M LiOH solution (13.30 mL, 3 equiv, 13.30 mmol), and the reaction mixture was heated to reflux and stirred overnight. The solvent was evaporated, and the resulting solid was dissolved in ice water and stirred vigorously. After acidification with 2 N HCl solution until reaching a pH of 2, the precipitated solid was collected by filtration. The solid was washed with ice water and then dried in vacuo to afford the title material as a yellow solid (0.96 g, 62%). 1H NMR (400 MHz, DMSO): δ 10.15 (s, 1H), 8.63 (d, $J = 5.2$ Hz, 1H), 8.42 (s, 1H), 8.13–8.05 (m, 1H), 8.02–7.86 (m, 5H), 7.60 (d, $J = 5.2$ Hz, 1H), 7.51–7.37 (m, 2H). ^{13}C NMR (101 MHz, DMSO): δ 167.26, 159.57, 159.02, 158.95, 144.47, 142.49, 140.33, 139.85, 130.25, 126.25, 125.37, 125.02, 124.88, 123.39, 122.96, 117.83, 107.90. HRMS-ESI+ (m/z): $[M + H]^+$ calcd for $C_{19}H_{14}N_3O_3S$, 348.0801; found, 348.0792.

1-((4-(Benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)benzoyl-piperidin-4-one (7). To a stirring solution of **6** (0.44 g, 1 equiv, 1.28 mmol), 1-hydroxybenzotriazole-hydrate (0.328 g, 80% wt, 1.20 equiv, 1.53 mmol), and TEA (0.39 g, 0.53 mL, 3.00 equiv, 3.83 mmol) in DMF (8.00 mL) was added 3-(((ethylimino)methylene)amino)-*N,N*-dimethylpropan-1-amine hydrochloride (0.29 g, 1.20 equiv, 1.53 mmol). The reaction mixture was stirred at room temperature for 15 min, followed by addition of 4-oxopiperidin-1-ium chloride (0.21 g, 1.20 equiv, 1.53 mmol), and the reaction was left to stir at room temperature overnight. It was then poured onto water, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with water, brine, and dried with Na_2SO_4 , and the solvent was removed in vacuo. The crude compound was purified by silica gel chromatography using a gradient of hexanes and ethyl acetate to afford the title material as a yellow solid (0.17 g, 31%). 1H NMR (850 MHz, DMSO): δ 8.60 (d, $J = 5.1$ Hz, 1H), 8.41 (d, $J = 0.8$ Hz, 1H), 8.09–8.03 (m, 1H), 7.98–7.93 (m, 3H), 7.57 (d, $J = 5.1$ Hz, 1H), 7.53–7.49 (m, 2H), 7.48–7.43 (m, 2H), 3.95–3.52 (m, 4H), 2.52–2.41 (m, 4H). LCMS (ESI+) m/z : 429 $[M + H]^+$.

4-((4-(Benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-methyl(propyl)amino)piperidin-1-yl)methanone (8a). To a stirring solution of **7** (0.050 g, 1 equiv, 0.12 mmol) and two drops of glacial acetic acid in THF (1.50 mL) was added *N*-methylpropan-1-amine (0.01 g, 0.02 mL, 1.5 equiv, 0.18 mmol). The reaction mixture was stirred at room temperature for 15 min after which was added $NaBH(OAc)_3$ (0.07 g, 3.0 equiv, 0.35 mmol), and then, it was left to

stir at room temperature overnight. MeOH was carefully added to the reaction mixture, and the solvents were removed in vacuo. The crude compound was purified using silica gel chromatography using a system of DCM/MeOH 100:0 to 80:20 to afford the title material as a white solid (0.008 g, 10%). ¹H NMR (400 MHz, MeOD): δ 8.48 (d, *J* = 5.3 Hz, 1H), 8.17 (s, 1H), 7.97–7.86 (m, 4H), 7.48–7.36 (m, 5H), 4.78–3.78 (m, 2H), 3.25–2.81 (m, 2H), 2.79–2.67 (m, 1H), 2.52–2.42 (m, 2H), 2.30 (s, 3H), 2.10–1.69 (m, 2H), 1.63–1.45 (m, 4H), 0.92 (t, *J* = 7.3 Hz, 3H). ¹³C NMR (214 MHz, MeOD): δ 172.64, 161.40, 161.28, 159.66, 144.16, 143.70, 142.48, 141.58, 129.59, 129.07, 127.22, 125.91, 125.86, 125.84, 123.60, 119.64, 108.59, 61.90, 56.75, 49.01, 43.14, 38.05, 28.94, 21.10, 12.16. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₈H₃₂N₅O₅, 486.2322; found, 486.2310.

(*R*)-4-((4-(Benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)phenyl-(4-(3-hydroxypyrrolidin-1-yl)piperidin-1-yl)methanone (**8b**). To a stirring solution of **7** (0.050 g, 1 equiv, 0.12 mmol) in THF (1.50 mL) was added (*R*)-pyrrolidin-3-ol (0.01 g, 0.01 mL, 1.20 equiv, 0.14 mmol), and the solution was stirred at room temperature for 1 h. Afterward, NaBH(OAc)₃ (0.07 g, 3.0 equiv, 0.35 mmol) was added, and the reaction was stirred at room temperature overnight. The solvent was removed in vacuo, and then water and ethyl acetate were added. The layers were separated, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were dried with Na₂SO₄, and the solvent was removed in vacuo. The crude compound was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 80:20 to afford the title material as a yellow solid (0.008 g, 10%). ¹H NMR (400 MHz, MeOD): δ 8.47 (d, *J* = 5.2 Hz, 1H), 8.16 (s, 1H), 7.99–7.83 (m, 4H), 7.49–7.35 (m, 5H), 4.72–4.44 (m, 1H), 4.45–4.33 (m, 1H), 4.25–3.73 (m, 1H), 3.25–2.87 (m, 4H), 2.84–2.65 (m, 2H), 2.65–2.51 (m, 1H), 2.22–1.89 (m, 3H), 1.87–1.72 (m, 1H), 1.64–1.42 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.65, 161.37, 161.26, 159.66, 144.14, 143.74, 142.46, 141.57, 129.49, 129.06, 127.23, 125.91, 125.87, 125.85, 123.58, 119.64, 108.62, 70.84, 63.22, 60.99, 51.11, 42.31, 34.67, 31.84. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₈H₃₀N₅O₅S, 500.2115; found, 500.2104.

(*S*)-4-((4-(Benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)phenyl-(4-(3-hydroxypyrrolidin-1-yl)piperidin-1-yl)methanone (**8c**). This compound was synthesized using the same procedure for **8b**, starting with **7** and (*S*)-pyrrolidin-3-ol (0.01 g, 0.11 mmol) to afford the title material as a yellow solid (0.008 g, 20%). ¹H NMR (850 MHz, MeOD): δ 8.47 (d, *J* = 5.1 Hz, 1H), 8.17 (s, 1H), 7.95–7.91 (m, 3H), 7.91–7.85 (m, 1H), 7.46–7.37 (m, 5H), 4.66–4.46 (m, 1H), 4.41–4.32 (m, 1H), 4.08–3.81 (m, 1H), 3.25–3.04 (m, 1H), 2.93 (dd, *J* = 10.3, 6.2 Hz, 2H), 2.88–2.79 (m, 1H), 2.69–2.64 (m, 1H), 2.60 (dd, *J* = 10.4, 3.3 Hz, 1H), 2.46–2.40 (m, 1H), 2.15–2.09 (m, 1H), 2.09–1.81 (m, 2H), 1.77–1.71 (m, 1H), 1.59–1.37 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.63, 161.40, 161.27, 159.66, 144.16, 143.69, 142.48, 141.58, 129.62, 129.02, 127.23, 125.91, 125.86, 125.84, 123.60, 119.65, 108.60, 71.05, 63.11, 61.10, 51.05, 42.64, 34.84, 32.41. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₈H₃₀N₅O₅S, 500.2115; found, 500.2101.

General Procedure C: Amide Coupling. 4-((4-(Benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-(dimethylamino)piperidin-1-yl)methanone (**9a**). To a stirring solution of **6** (0.15 g, 1 equiv, 0.43 mmol), *N,N*-dimethylpiperidin-4-amine (0.06 g, 1.2 equiv, 0.52 mmol), 1-hydroxybenzotriazole-hydrate (0.11 g, 80% wt, 1.2 equiv, 0.52 mmol), and TEA (0.13 g, 0.18 mL, 3 equiv, 1.30 mmol) in DMF (6 mL) was added 3-(((ethylimino)methylene)amino)-*N,N*-dimethylpropan-1-amine hydrochloride (0.10 g, 1.2 equiv, 0.52 mmol), and the reaction was stirred at room temperature for 24 h. The reaction mixture was poured onto water, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with water and brine and dried with Na₂SO₄, and the solvent was removed in vacuo. The crude product was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 90:10 to afford the title material as a yellow solid (0.03 g, 13%). ¹H NMR (400 MHz, DMSO): δ 9.98 (s, 1H), 8.59 (d, *J* = 5.1 Hz, 1H), 8.40 (s, 1H), 8.11–7.99 (m, 1H), 8.00–7.85 (m, 3H), 7.56 (d, *J* = 5.2 Hz, 1H), 7.49–7.42 (m, 2H), 7.42–7.37 (m, 2H), 3.14–2.72 (m, 4H), 2.49–

2.40 (m, 1H), 2.25 (s, 6H), 1.88–1.71 (m, 2H), 1.37 (qd, *J* = 12.0, 4.1 Hz, 2H). ¹³C NMR (101 MHz, DMSO): δ 169.07, 159.75, 158.99, 142.59, 141.67, 140.30, 139.88, 128.75, 127.87, 126.28, 125.30, 125.05, 124.91, 122.93, 118.15, 107.57, 61.49, 41.16, 27.94. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₆H₂₈N₅O₅, 458.2009; found, 458.1995.

[1,4'-Bipiperidin]-1'-yl(4-((4-(benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)phenyl)methanone (**9b**). To a stirring solution of **6** (0.080 g, 1 equiv, 0.23 mmol), 1-hydroxybenzotriazole-hydrate (0.06 g, 80% wt, 1.2 equiv, 0.28 mmol), and TEA (0.070 g, 0.096 mL, 3.0 equiv, 0.69 mmol) in DMF (2.00 mL) was added 3-(((ethylimino)methylene)amino)-*N,N*-dimethylpropan-1-amine hydrochloride (0.05 g, 1.2 equiv, 0.28 mmol). The reaction was stirred at room temperature for 15 min, followed by addition of 1,4'-bipiperidine (0.05 g, 1.2 equiv, 0.28 mmol) after which the reaction was stirred at room temperature overnight. The reaction mixture was poured onto water, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with water and brine and dried with Na₂SO₄, and the solvent was removed in vacuo. The crude compound was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 90:10 to afford the title material as a yellow solid (0.04 g, 35%). ¹H NMR (850 MHz, DMSO): δ 9.98 (s, 1H), 8.60 (d, *J* = 5.1 Hz, 1H), 8.40 (s, 1H), 8.11–8.04 (m, 1H), 7.96–7.94 (m, 1H), 7.94–7.90 (m, 2H), 7.56 (d, *J* = 5.1 Hz, 1H), 7.48–7.43 (m, 2H), 7.41–7.37 (m, 2H), 4.66–3.58 (m, 2H), 3.11–2.63 (m, 2H), 2.49–2.40 (m, 5H), 1.83–1.59 (m, 2H), 1.52–1.44 (m, 4H), 1.44–1.32 (m, 4H). ¹³C NMR (214 MHz, DMSO): δ 168.96, 159.72, 158.94, 142.56, 141.58, 140.27, 139.84, 128.82, 127.79, 126.21, 125.24, 124.99, 124.85, 122.90, 118.10, 107.51, 61.76, 49.66, 27.74, 26.03, 24.51. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₉H₃₂N₅O₅S, 498.2322; found, 498.2308.

tert-Butyl 4-((4-(benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)benzoyl)piperazine-1-carboxylate (**9c-Boc**). This compound was synthesized according to general procedure C starting with **6** and *tert*-butylpiperazine-1-carboxylate (0.24 g, 1.29 mmol) to afford the title material as a yellow solid (0.26 g, 58%). ¹H NMR (850 MHz, CDCl₃): δ 8.47 (d, *J* = 5.1 Hz, 1H), 7.99 (s, 1H), 7.90 (d, *J* = 7.7 Hz, 1H), 7.85 (dd, *J* = 7.3, 1.5 Hz, 1H), 7.83–7.78 (m, 2H), 7.50–7.46 (m, 3H), 7.44–7.37 (m, 2H), 7.22 (d, *J* = 5.1 Hz, 1H), 3.89–3.28 (m, 8H), 1.48 (s, 9H). MS ESI+ *m/z*: 516 [M + H]⁺.

4-((4-(Benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)phenyl-(piperazin-1-yl)methanone (**9c**). To a stirring solution of **9c-Boc** (0.07 g, 1 equiv, 0.13 mmol) in DCM/MeOH (2:1, 0.5 mL) was added 4 M HCl in dioxane (16 equiv, 0.50 mL), and the reaction was left to stir at room temperature overnight. The solvent was removed in vacuo, and the residue was triturated with DCM and then filtered. The residue was washed with DCM and then purified using silica gel chromatography using a system of DCM/MeOH 100:0 to 85:15 to afford the title material as white solid (0.02 g, 42%). ¹H NMR (400 MHz, DMSO): δ 10.04 (s, 1H), 8.61 (d, *J* = 5.1 Hz, 1H), 8.42 (s, 1H), 8.10–8.03 (m, 1H), 8.01–7.87 (m, 3H), 7.58 (d, *J* = 5.1 Hz, 1H), 7.52–7.39 (m, 4H), 3.79–3.64 (m, 4H), 3.19–3.03 (m, 4H). ¹³C NMR (214 MHz, DMSO): δ 169.47, 159.69, 158.99, 142.51, 142.21, 140.27, 139.86, 128.37, 127.20, 126.29, 125.35, 125.05, 124.91, 122.89, 118.09, 107.68, 42.66. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₃H₂₂N₅O₂S, 416.1540; found, 416.1528.

4-((4-(Benzo[*b*]thiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-methylpiperazin-1-yl)methanone (**9d**). This compound was synthesized according to general procedure C starting from **6** and 1-methylpiperazine (0.05 g, 0.06 mL, 0.52 mmol) to afford the title material as a yellow solid (0.06 g, 32%). ¹H NMR (400 MHz, DMSO): δ 9.99 (s, 1H), 8.60 (d, *J* = 5.2 Hz, 1H), 8.41 (s, 1H), 8.10–8.04 (m, 1H), 7.98–7.88 (m, 3H), 7.57 (d, *J* = 5.1 Hz, 1H), 7.51–7.42 (m, 2H), 7.42–7.36 (m, 2H), 3.59–3.42 (m, 4H), 2.36–2.27 (m, 4H), 2.20 (s, 3H). ¹³C NMR (214 MHz, DMSO): δ 169.06, 159.69, 158.93, 142.53, 141.70, 140.26, 139.83, 128.40, 127.97, 126.20, 125.24, 124.98, 124.84, 122.88, 118.09, 107.53, 54.56, 45.62. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₄H₂₄N₅O₅, 430.1696; found 430.1684.

4-((4-(Benzo[b]thiophen-2-yl)pyrimidin-2-yl)amino)-N-(1-methylpiperidin-4-yl)benzamide (**9e**). This compound was synthesized according to general procedure C starting from **6** and 1-methylpiperidin-4-amine (0.06 g, 0.48 mmol) to afford the title material as a yellow solid (0.02 g, 12%). ¹H NMR (400 MHz, DMSO): δ 10.04 (s, 1H), 8.61 (d, *J* = 5.2 Hz, 1H), 8.42 (s, 1H), 8.14 (d, *J* = 7.7 Hz, 1H), 8.11–8.05 (m, 1H), 7.99–7.93 (m, 3H), 7.91–7.85 (m, 2H), 7.59 (d, *J* = 5.2 Hz, 1H), 7.51–7.42 (m, 2H), 3.87–3.70 (m, 1H), 2.96–2.78 (m, 2H), 2.26 (s, 3H), 2.21–2.04 (m, 2H), 1.86–1.74 (m, 2H), 1.74–1.53 (m, 2H). ¹³C NMR (214 MHz, DMSO): δ 165.31, 159.66, 159.08, 158.86, 143.06, 142.59, 140.27, 139.90, 128.10, 127.21, 126.30, 125.31, 125.06, 124.91, 122.91, 117.63, 107.63, 54.20, 45.99, 45.38, 31.04. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₅H₂₆N₅OS, 222.5963; found, 222.5958.

4-((4-(Benzo[b]thiophen-2-yl)pyrimidin-2-yl)amino)-N-(3-(4-methylpiperazin-1-yl)propyl)benzamide (**9f**). This compound was synthesized according to general procedure C starting with **6** and 3-(4-methylpiperazin-1-yl)propan-1-amine (0.04 g, 0.27 mmol) to afford the title material as a yellow solid (0.05 g, 47%). ¹H NMR (400 MHz, DMSO): δ 10.03 (s, 1H), 8.61 (d, *J* = 5.2 Hz, 1H), 8.41 (s, 1H), 8.38 (t, *J* = 5.5 Hz, 1H), 8.09–8.05 (m, 1H), 7.98–7.93 (m, 3H), 7.88–7.82 (m, 2H), 7.58 (d, *J* = 5.2 Hz, 1H), 7.50–7.43 (m, 2H), 3.32–3.24 (m, 3H), 2.47–2.22 (m, 9H), 2.14 (s, 3H), 1.75–1.62 (m, 2H). ¹³C NMR (214 MHz, DMSO): δ 165.70, 159.66, 159.04, 158.87, 143.01, 142.57, 140.26, 139.88, 127.87, 127.26, 126.29, 125.30, 125.04, 124.89, 122.88, 117.74, 107.63, 55.93, 54.80, 52.74, 45.76, 38.02, 26.26. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₇H₃₁N₅OS, 487.2275; found, 487.2263.

4-((4-(Benzo[b]thiophen-2-yl)pyrimidin-2-yl)amino)-N-(3-morpholinopropyl)benzamide (**9g**). To a stirring solution of **6** (0.08 g, 1 equiv, 0.23 mmol), 1-hydroxybenzotriazole-hydrate (0.059 g, 80% wt, 1.2 equiv, 0.28 mmol), and TEA (0.070 g, 0.10 mL, 3.0 equiv, 0.69 mmol) in DMF (2.00 mL) was added 3-((ethylimino)methylene)-amino)-N,N-dimethylpropan-1-amine hydrochloride (0.05 g, 1.2 equiv, 0.28 mmol). The reaction was stirred at room temperature for 15 min, then 3-morpholinopropan-1-amine (0.04 g, 0.04 mL, 1.2 equiv, 0.28 mmol) was added, and the reaction was heated to 80 °C and stirred for 18 h. Afterward, the reaction mixture was poured onto water, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with water and brine and dried with Na₂SO₄, and the solvent was removed in vacuo. The crude product was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 90:10 to afford the title material as a white solid (0.04 g, 37%). ¹H NMR (400 MHz, DMSO): δ 10.04 (s, 1H), 8.61 (d, *J* = 5.2 Hz, 1H), 8.41 (s, 1H), 8.38 (t, *J* = 5.8 Hz, 1H), 8.09–8.04 (m, 1H), 7.99–7.93 (m, 3H), 7.88–7.83 (m, 2H), 7.58 (d, *J* = 5.2 Hz, 1H), 7.50–7.42 (m, 2H), 3.59 (t, *J* = 4.6 Hz, 4H), 3.32–3.27 (m, 2H), 2.43–2.29 (m, 6H), 1.70 (p, *J* = 7.0 Hz, 2H). ¹³C NMR (214 MHz, DMSO): δ 165.72, 159.65, 159.05, 158.86, 143.02, 142.60, 140.28, 139.88, 127.86, 127.25, 126.29, 125.30, 125.04, 124.88, 122.89, 117.74, 107.61, 66.23, 56.32, 53.38, 37.90, 25.91. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₆H₂₈N₅O₂S, 474.1958; found, 474.1944.

4-((4-(Benzo[b]thiophen-2-yl)pyrimidin-2-yl)amino)-N-(3-(dimethylamino)propyl)benzamide (**9h**). This compound was synthesized according to general procedure C starting from **6** and N1,N1-dimethylpropane-1,3-diamine (0.05 g, 0.07 mL, 0.52 mmol) to afford the title material as a yellow solid (0.02 g, 10%). ¹H NMR (400 MHz, DMSO): δ 10.03 (s, 1H), 8.61 (d, *J* = 5.2 Hz, 1H), 8.44–8.33 (m, 2H), 8.10–8.04 (m, 1H), 7.98–7.89 (m, 3H), 7.88–7.79 (m, 2H), 7.58 (d, *J* = 5.2 Hz, 1H), 7.51–7.41 (m, 2H), 3.31–3.24 (m, 2H), 2.26 (t, *J* = 7.1 Hz, 2H), 2.14 (s, 6H), 1.66 (p, *J* = 7.1 Hz, 2H). ¹³C NMR (214 MHz, DMSO): δ 166.07, 159.81, 159.24, 159.06, 143.18, 142.68, 140.42, 140.04, 128.05, 127.39, 126.53, 125.48, 125.26, 125.12, 123.05, 118.00, 107.84, 57.14, 45.33, 37.90, 27.32. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₄H₂₆N₅OS, 432.1853; found 432.1842.

4-((4-(Benzo[b]thiophen-2-yl)pyrimidin-2-yl)amino)-N-(3-(methylsulfonyl)propyl)benzamide (**9i**). This compound was synthesized according to general procedure C starting with **6** and 3-

(methylsulfonyl)propan-1-aminium chloride (0.11 g, 0.61 mmol) to afford the title material as a yellow solid (0.05 g, 21%). ¹H NMR (400 MHz, DMSO): δ 10.05 (s, 1H), 8.62 (d, *J* = 5.2 Hz, 1H), 8.44 (t, *J* = 5.8 Hz, 1H), 8.42 (s, 1H), 8.10–8.06 (m, 1H), 8.00–7.94 (m, 3H), 7.91–7.83 (m, 2H), 7.59 (d, *J* = 5.2 Hz, 1H), 7.51–7.41 (m, 2H), 3.44–3.35 (m, 2H), 3.21–3.15 (m, 2H), 2.99 (s, 3H), 2.02–1.89 (m, 2H). ¹³C NMR (214 MHz, DMSO): δ 166.01, 159.65, 159.06, 158.88, 143.16, 142.57, 140.27, 139.88, 128.00, 126.97, 126.30, 125.33, 125.06, 124.90, 122.90, 117.73, 107.67, 51.58, 40.13, 37.79, 22.42. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₃H₂₃N₄O₃S₂, 467.1206; found, 467.1194.

4-((4-(Benzo[b]thiophen-2-yl)pyrimidin-2-yl)amino)-N-cyclopropylbenzamide (**9j**). This compound was synthesized according to general procedure C starting from **6** and cyclopropanamine to afford the title material as a yellow solid (0.01 g, 6%). The N.B. Product was purified using a gradient of hexanes and ethyl acetate. ¹H NMR (400 MHz, DMSO): δ 10.03 (s, 1H), 8.61 (d, *J* = 5.2 Hz, 1H), 8.41 (s, 1H), 8.29 (d, *J* = 4.2 Hz, 1H), 8.10–8.04 (m, 1H), 7.99–7.90 (m, 3H), 7.87–7.78 (m, 2H), 7.58 (d, *J* = 5.2 Hz, 1H), 7.52–7.41 (m, 2H), 2.89–2.80 (m, 1H), 0.73–0.66 (m, 2H), 0.61–0.54 (m, 2H). ¹³C NMR (214 MHz, DMSO): δ 167.13, 159.66, 159.08, 158.88, 143.08, 142.59, 140.28, 139.90, 127.96, 127.03, 126.32, 125.33, 125.07, 124.92, 122.91, 117.69, 107.65, 23.03, 5.80. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₂H₁₉N₅OS, 387.1274; found, 387.1264.

4-((4-(Benzo[b]thiophen-2-yl)pyrimidin-2-yl)amino)-N-(1-methyl-1H-pyrazol-4-yl)benzamide (**9k**). To a stirring solution of **6** (0.175 g, 1 equiv, 0.504 mmol), 1-methyl-1H-pyrazol-4-amine (0.06 g, 1.20 equiv, 0.61 mmol), and TEA (0.153 g, 0.21 mL, 3.00 equiv, 1.51 mmol) in DMF (4 mL) was added HATU (0.23 g, 1.20 equiv, 0.61 mmol). The reaction was heated to 80 °C and stirred overnight. The reaction mixture was poured onto water, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with water and brine and dried with Na₂SO₄, and the solvent was removed in vacuo. The crude compound was purified by silica gel chromatography using a gradient of hexanes and ethyl acetate to afford the title material as a greenish-white solid (0.01 g, 4%). ¹H NMR (400 MHz, DMSO): δ 10.27 (s, 1H), 10.11 (s, 1H), 8.63 (d, *J* = 5.2 Hz, 1H), 8.43 (s, 1H), 8.11–8.05 (m, 1H), 8.04–8.00 (m, 3H), 7.99–7.93 (m, 3H), 7.60 (d, *J* = 5.2 Hz, 1H), 7.58 (d, *J* = 0.8 Hz, 1H), 7.51–7.43 (m, 2H), 3.83 (s, 3H). ¹³C NMR (214 MHz, DMSO): δ 163.05, 159.64, 159.10, 158.89, 143.38, 142.55, 140.27, 139.89, 130.13, 128.19, 126.65, 126.32, 125.36, 125.07, 124.91, 122.90, 122.06, 121.52, 117.81, 107.74, 38.69. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₃H₁₈N₆OS, 427.1336; found, 427.1325.

4-((4-(Benzo[b]thiophen-2-yl)pyrimidin-2-yl)amino)phenyl)-methanol (**10**). A mixture of **3** (0.340 g, 1 equiv, 1.38 mmol), (4-aminophenyl)methanol (0.19 g, 1.10 equiv, 1.52 mmol), and Cs₂CO₃ (0.898 g, 2.00 equiv, 2.76 mmol) was stirred in dioxane (12 mL), and the solvent was degassed by bubbling argon gas for 5 minutes. Afterward, xantphos (0.08 g, 0.10 equiv, 0.14 mmol) and Pd₂(dba)₃ (0.13 g, 0.10 equiv, 0.14 mmol) were added to the reaction mixture, which was heated to 90 °C and stirred overnight. The mixture was cooled to room temperature and filtered through Celite. The filtrate was concentrated in vacuo, and the crude product was purified by silica gel chromatography using a gradient of DCM/ethyl acetate to afford the title material as a yellow solid (0.12 g, 26%). ¹H NMR (400 MHz, DMSO): δ 9.71 (s, 1H), 8.55 (d, *J* = 5.2 Hz, 1H), 8.37 (s, 1H), 8.11–8.04 (m, 1H), 7.97–7.90 (m, 1H), 7.84–7.76 (m, 2H), 7.49 (d, *J* = 5.2 Hz, 1H), 7.48–7.40 (m, 2H), 7.33–7.21 (m, 2H), 5.07 (t, *J* = 5.7 Hz, 1H), 4.47 (d, *J* = 5.8 Hz, 2H). LCMS (ESI+) *m/z*: 334 [M + H]⁺.

4-(Benzo[b]thiophen-2-yl)-N-(4-(chloromethyl)phenyl)pyrimidin-2-amine (**11**). To a stirring suspension of **10** (0.80 g, 1 equiv, 2.4 mmol) and DIPEA (0.62 g, 0.84 mL, 2 equiv, 4.8 mmol) in THF (23 mL) was added SOCl₂ (0.86 g, 0.53 mL, 3 equiv, 7.2 mmol) dropwise, and the reaction mixture was stirred at room temperature overnight. Excess thionyl chloride and the reaction solvent were removed under reduced pressure. Afterward, water and DCM were added to the reaction mixture, and the layers were separated. The organic layer was washed with water and brine and then dried with

Na_2SO_4 . The solvent was removed in vacuo, and the crude product was used for the next step without further purification.

4-(Benzob[thiophen-2-yl]-N-(4-((4-(pyrrolidin-1-yl)piperidin-1-yl)methyl)phenyl)pyrimidin-2-amine (12). To a stirring solution of **11** (0.10 g, 1 equiv, 0.28 mmol) and K_2CO_3 (0.12 g, 3.01 equiv, 0.85 mmol) in DMF (2.00 mL) was added 4-(pyrrolidin-1-yl)piperidine (0.09 g, 2.00 equiv, 0.57 mmol), and the reaction mixture was stirred at room temperature overnight. The reaction was quenched by adding water, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with brine, dried with Na_2SO_4 , and concentrated in vacuo. The crude product was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 40:60. The isolated product was re-purified using prep HPLC to afford the TFA salt of title material as a yellow solid (0.03 g, 13%). ^1H NMR (400 MHz, MeOD): δ 8.48 (d, J = 5.3 Hz, 1H), 8.19 (s, 1H), 8.03–7.95 (m, 2H), 7.93–7.86 (m, 2H), 7.52–7.47 (m, 2H), 7.46–7.36 (m, 3H), 4.34 (s, 2H), 3.75–3.60 (m, 4H), 3.51–3.39 (m, 1H), 3.22–2.97 (m, 4H), 2.53–2.37 (m, 2H), 2.21–1.90 (m, 6H). ^{13}C NMR (151 MHz, MeOD): δ 162.80, 162.34, 160.52, 145.26, 144.93, 143.77, 142.84, 134.25, 128.64, 127.43, 127.26, 127.23, 124.76, 121.81, 109.90, 62.60, 61.53, 54.55, 52.38, 28.71, 25.13. HRMS–ESI+ (m/z): $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{28}\text{H}_{33}\text{N}_5\text{S}$, 470.2373; found, 470.2360.

tert-Butyl 4-(4-(pyrrolidin-1-yl)piperidine-1-carbonyl)phenylcarbamate (14). To a stirring solution of 4-(tert-butoxycarbonyl)amino)benzoic acid **13** (0.50 g, 1.0 equiv, 2.11 mmol), 4-(pyrrolidin-1-yl)piperidine (0.39 g, 1.20 equiv, 2.53 mmol), and TEA (0.64 g, 0.88 mL, 3.00 equiv, 6.32 mmol) in DMF (10.00 mL) was added HATU (0.88 g, 1.10 equiv, 2.32 mmol), and the reaction was allowed to stir at room temperature for 24 h. Afterward, the reaction mixture was poured onto water, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with water and brine and dried with Na_2SO_4 , and the solvent was removed in vacuo. The crude compound was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 90:10 to afford the title material as a white solid (0.63 g, 80%). ^1H NMR (400 MHz, DMSO): δ 9.53 (s, 1H), 7.55–7.39 (m, 2H), 7.31–7.15 (m, 2H), 4.41–3.50 (m, 2H), 3.11–2.82 (m, 2H), 2.54–2.44 (m, 4H), 2.34–2.19 (m, 1H), 1.90–1.76 (m, 2H), 1.72–1.62 (m, 4H), 1.48 (s, 9H), 1.41–1.28 (m, 2H). ^{13}C NMR (101 MHz, DMSO): δ 169.28, 153.12, 141.09, 129.94, 128.18, 117.86, 79.78, 61.04, 51.19, 46.12, 31.48, 28.51, 23.36. LCMS (ESI+) m/z : 374 $[\text{M} + \text{H}]^+$.

(4-Aminophenyl)(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (15). To a stirring solution of **14** (0.63 g, 1 equiv, 1.69 mmol) in DCM (2.00 mL) was added 4 M HCl in dioxane (5 equiv, 2.00 mL), and the reaction solution was stirred at room temperature for 48 h. The solvent was evaporated, and the residue was triturated with DCM and then filtered. The filtered residue was washed with DCM, collected, and then dried in vacuo. Afterward, it was dissolved in water, and the pH was adjusted to 12 using a 1 M NaOH solution. The aqueous layer was extracted with DCM ($\times 3$). The combined organic extracts were washed with brine and dried with Na_2SO_4 , and the solvent was removed in vacuo to afford the title material as a white solid (0.31 g, 67%). ^1H NMR (400 MHz, MeOD): δ 7.22–7.14 (m, 2H), 6.77–6.58 (m, 2H), 4.65–3.82 (m, 2H), 3.13–2.82 (m, 2H), 2.70–2.56 (m, 4H), 2.34 (tt, J = 11.0, 4.0 Hz, 1H), 2.08–1.93 (m, 2H), 1.88–1.74 (m, 4H), 1.53–1.38 (m, 2H). LCMS (ESI+) m/z : 274 $[\text{M} + \text{H}]^+$.

4-(Benzofuran-2-yl)-2-chloropyrimidine (17a). This compound was synthesized according to general procedure A starting from **1** and benzofuran-2-ylboronic acid (0.30 g, 1.85 mmol) to afford the title material as a white solid (0.03 g, 8%). ^1H NMR (400 MHz, CDCl_3): δ 8.68 (d, J = 5.1 Hz, 1H), 7.77 (d, J = 1.0 Hz, 1H), 7.74 (d, J = 5.2 Hz, 1H), 7.72–7.67 (m, 1H), 7.58 (dq, J = 8.4, 0.9 Hz, 1H), 7.44 (ddd, J = 8.4, 7.2, 1.3 Hz, 1H), 7.32 (ddd, J = 8.0, 7.2, 1.0 Hz, 1H). LCMS (ESI+) m/z : 231 $[\text{M} + \text{H}]^+$.

2-Chloro-4-(naphthalen-2-yl)pyrimidine (17b). 2,4-Dichloropyrimidine **1** (0.22 g, 1 equiv, 1.45 mmol) was dissolved in a mixture of dioxane (10.00 mL) and water (2.50 mL). The solution was degassed by bubbling nitrogen gas for 5 minutes. Naphthalen-2-ylboronic acid (0.25 g, 1 equiv, 1.45 mmol), Cs_2CO_3 (1.42 g, 3 equiv, 4.36 mmol),

and $\text{Pd}(\text{PPh}_3)_4$ (0.13 g, 0.08 equiv, 0.116 mmol) were added to the reaction mixture, and the reaction mixture was heated to 70 °C and stirred for 20 h. Afterward, the reaction mixture was poured into water, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with water and brine, dried with Na_2SO_4 , filtered, and then concentrated. The crude compound was purified by silica gel chromatography using a gradient of hexanes and ethyl acetate to afford the title material as a yellow solid (0.23 g, 66%). ^1H NMR (400 MHz, DMSO): δ 8.86 (d, J = 5.3 Hz, 1H), 8.85–8.83 (m, 1H), 8.28 (d, J = 5.3 Hz, 1H), 8.27–8.21 (m, 1H), 8.15–8.11 (m, 1H), 8.09 (d, J = 8.7 Hz, 1H), 8.03–7.94 (m, 1H), 7.69–7.57 (m, 2H). LCMS (ESI+) m/z : 241 $[\text{M} + \text{H}]^+$.

2-Chloro-4-(thiophen-2-yl)pyrimidine (17c). This compound was synthesized according to general procedure A starting from **1** and thiophen-2-ylboronic acid (0.30 g, 2.34 mmol) to afford the title material as a white solid (0.27 g, 59%). ^1H NMR (400 MHz, CDCl_3): δ 8.54 (d, J = 5.3 Hz, 1H), 7.83 (dd, J = 3.8, 1.1 Hz, 1H), 7.60 (dd, J = 5.0, 1.1 Hz, 1H), 7.47 (d, J = 5.3 Hz, 1H), 7.18 (dd, J = 5.0, 3.8 Hz, 1H). LCMS (ESI+) m/z : 196 $[\text{M} + \text{H}]^+$.

2-(2-Chloropyrimidin-4-yl)thiazole (17d). To a solution of $\text{Pd}(\text{PPh}_3)_4$ (0.12 g, 0.05 equiv, 0.10 mmol) and LiCl (0.10 g, 1.20 equiv, 2.42 mmol) in anhydrous DMF (3.00 mL) were added **1** (0.30 g, 1 equiv, 2.01 mmol) and 2-(tributylstannyl)thiazole (0.90 g, 0.76 mL, 1.20 equiv, 2.42 mmol) under argon pressure, and the reaction mixture was stirred at 70 °C for 16 h. The reaction mixture was cooled to room temperature, followed by addition of a saturated KF solution. The formed precipitate was filtered through a Celite pad and washed with ethyl acetate. The filtrate was extracted with ethyl acetate, and the combined organic extracts were washed with brine and dried with Na_2SO_4 . The solvent was removed in vacuo, and the crude product was purified by silica gel chromatography using a gradient of hexanes and ethyl acetate to afford the title material as a white solid (0.04 g, 10%). ^1H NMR (400 MHz, CDCl_3): δ 8.66 (d, J = 5.1 Hz, 1H), 7.99 (d, J = 5.1 Hz, 1H), 7.97 (d, J = 3.1 Hz, 1H), 7.57 (d, J = 3.1 Hz, 1H). LCMS (ESI+) m/z : 198 $[\text{M} + \text{H}]^+$.

2-Chloro-4-(furan-2-yl)pyrimidine (17e). This compound was synthesized according to general procedure A starting from **1** and (furan-2-ylboronic acid (0.19 g, 1.68 mmol)) to afford the title material as a white solid (0.20 g, 66%). ^1H NMR (400 MHz, CDCl_3): δ 8.58 (d, J = 5.2 Hz, 1H), 7.63 (dd, J = 1.8, 0.8 Hz, 1H), 7.52 (d, J = 5.2 Hz, 1H), 7.39 (dd, J = 3.6, 0.8 Hz, 1H), 6.61 (dd, J = 3.6, 1.7 Hz, 1H). LCMS (ESI+) m/z : 181 $[\text{M} + \text{H}]^+$.

2-Chloro-4-(1-methyl-1H-pyrazol-4-yl)pyrimidine (17f). This compound was synthesized according to general procedure A starting from **1** and (1-methyl-1H-pyrazol-4-yl)boronic acid (0.25 g, 1.99 mmol) to afford the title material as a pink solid (0.10 g, 26%). ^1H NMR (400 MHz, CDCl_3): δ 8.49 (d, J = 5.3 Hz, 1H), 8.10 (s, 1H), 8.01 (s, 1H), 7.29 (d, J = 5.3 Hz, 1H), 3.97 (s, 3H). LCMS (ESI+) m/z : 195 $[\text{M} + \text{H}]^+$.

2-Chloro-4-phenylpyrimidine (17g). This compound was synthesized according to general procedure A starting from **1** and phenylboronic acid (0.25 g, 2.01 mmol) to afford the title material as a white solid (0.25 g, 65%). ^1H NMR (400 MHz, CDCl_3): δ 8.64 (d, J = 5.2 Hz, 1H), 8.13–8.03 (m, 2H), 7.65 (d, J = 5.2 Hz, 1H), 7.58–7.46 (m, 3H). LCMS (ESI+) m/z : 191 $[\text{M} + \text{H}]^+$.

2-Bromo-4-(3,4-dichlorophenyl)pyrimidine (17h). This compound was synthesized according to general procedure A starting from 2,4-dibromopyrimidine (0.30 g, 1.26 mmol) and (3,4-dichlorophenyl)boronic acid (0.24 g, 1.26 mmol) to afford the title material as a white solid (0.18 g, 47%). ^1H NMR (400 MHz, CDCl_3): δ 8.61 (d, J = 5.2 Hz, 1H), 8.21 (d, J = 2.2 Hz, 1H), 7.91 (dd, J = 8.4, 2.2 Hz, 1H), 7.65 (d, J = 5.2 Hz, 1H), 7.59 (d, J = 8.4 Hz, 1H). LCMS (ESI+) m/z : 303 $[\text{M} + \text{H}]^+$.

2-Chloro-4-(5-methylthiophen-2-yl)pyrimidine (17i). This compound was synthesized according to general procedure A starting from **1** and (5-methylthiophen-2-yl)boronic acid (0.29 g, 2.01 mmol) to afford the title material as a yellow solid (0.18 g, 41%). ^1H NMR (400 MHz, CDCl_3): δ 8.48 (d, J = 5.4 Hz, 1H), 7.65 (d, J = 3.7 Hz, 1H), 7.38 (d, J = 5.4 Hz, 1H), 6.88–6.75 (m, 1H), 2.56 (d, J = 0.5 Hz, 3H). LCMS (ESI+) m/z : 211 $[\text{M} + \text{H}]^+$.

2-Chloro-4-(5-chlorothiophen-2-yl)pyrimidine (17j). This compound was synthesized according to general procedure A starting from **1** and (5-chlorothiophen-2-yl)boronic acid (0.33 g, 2.01 mmol) to afford the title material as a green solid (0.26 g, 56%). ¹H NMR (400 MHz, CDCl₃): δ 8.54 (d, *J* = 5.3 Hz, 1H), 7.59 (d, *J* = 4.1 Hz, 1H), 7.39 (d, *J* = 5.3 Hz, 1H), 6.99 (d, *J* = 4.1 Hz, 1H). LCMS (ESI+) *m/z*: 230 [M + H]⁺.

2-Chloro-4-(5-phenylthiophen-2-yl)pyrimidine (17k). This compound was synthesized according to general procedure A starting from **1** and (5-phenylthiophen-2-yl)boronic acid (0.34 g, 1.68 mmol) to afford the title material as a brown solid (0.25 g, 54%). ¹H NMR (400 MHz, CDCl₃): δ 8.51 (d, *J* = 5.5 Hz, 1H), 7.78 (d, *J* = 4.0 Hz, 1H), 7.69–7.61 (m, 2H), 7.47–7.39 (m, 3H), 7.39–7.33 (m, 2H). LCMS (ESI+) *m/z*: 272 [M + H]⁺.

(5-(2-Chloropyrimidin-4-yl)thiophen-2-yl)methanol (17l). This compound was synthesized according to general procedure A starting from **1** and (5-(hydroxymethyl)thiophen-2-yl)boronic acid (0.25 g, 1.58 mmol) to afford the title material as a yellow solid (0.22 g, 60%). ¹H NMR (400 MHz, CDCl₃): δ 8.53 (d, *J* = 5.3 Hz, 1H), 7.72 (d, *J* = 3.8 Hz, 1H), 7.45–7.40 (m, 1H), 7.09–7.04 (m, 1H), 4.89 (br s, 2H). LCMS (ESI+) *m/z*: 227 [M + H]⁺.

4-(Benzo[b]thiophen-2-yl)-2-chloro-5-methylpyrimidine (17n). This compound was synthesized according to general procedure A starting from **2** and 4-dichloro-5-methylpyrimidine (**16n**) (0.23 g, 1.40 mmol) to afford the title material as a pink solid (0.09 g, 24%). ¹H NMR (400 MHz, CDCl₃): δ 8.47 (d, *J* = 1.0 Hz, 1H), 8.00 (d, *J* = 1.9 Hz, 1H), 7.89 (m, 2H), 7.48–7.35 (m, 2H), 2.64 (d, *J* = 1.7 Hz, 3H). LCMS (ESI+) *m/z*: 261 [M + H]⁺.

4-(Benzo[b]thiophen-2-yl)-2-chloro-5-cyclopropylpyrimidine (17o). This compound was synthesized according to general procedure A starting from **2** and 2,4-dichloro-5-cyclopropylpyrimidine (**16o**) (0.50 g, 2.81 mmol) to afford the title material as a white solid (0.20 g, 49%). ¹H NMR (400 MHz, DMSO): δ 8.63 (d, *J* = 0.8 Hz, 1H), 8.56 (s, 1H), 8.08–7.97 (m, 2H), 7.53–7.40 (m, 2H), 2.32–2.19 (m, 1H), 1.27–1.13 (m, 2H), 0.96–0.80 (m, 2H). LCMS (ESI+) *m/z*: 287 [M + H]⁺.

4-(Benzo[b]thiophen-2-yl)-2-chloro-5-methoxyypyrimidine (17p). This compound was synthesized according to general procedure A starting from **2** and 2,4-dichloro-5-methoxyypyrimidine (**16p**) (0.25 g, 1.40 mmol) to afford the title material as a white solid (0.18 g, 49%). ¹H NMR (400 MHz, DMSO): δ 8.71 (s, 1H), 8.50 (s, 1H), 8.10–7.94 (m, 2H), 7.53–7.35 (m, 2H), 4.16 (s, 3H). LCMS (ESI+) *m/z*: 277 [M + H]⁺.

4-(Benzo[b]thiophen-2-yl)-2-chloro-5-fluoropyrimidine (17q). This compound was synthesized according to general procedure A starting from **2** and 2,4-dichloro-5-fluoropyrimidine (**16q**) (0.23 g, 1.40 mmol) to afford the title material as a white solid (0.20 g, 55%). ¹H NMR (400 MHz, CDCl₃): δ 8.51 (d, *J* = 2.7 Hz, 1H), 8.29–8.23 (m, 1H), 7.95–7.84 (m, 2H), 7.50–7.37 (m, 2H). LCMS (ESI+) *m/z*: 264 [M + H]⁺.

4-(Benzo[b]thiophen-2-yl)-2,5-dichloropyrimidine (17r). This compound was synthesized according to general procedure A starting from **2** and 2,4,5-trichloropyrimidine (**16r**) (0.26 g, 1.40 mmol) to afford the title material as a white solid (0.20 g, 50%). ¹H NMR (400 MHz, CDCl₃): δ 8.63 (d, *J* = 0.9 Hz, 1H), 8.61 (s, 1H), 7.94–7.83 (m, 2H), 7.50–7.38 (m, 2H). LCMS (ESI+) *m/z*: 280 [M + H]⁺.

4-(Benzo[b]thiophen-2-yl)-5-bromo-2-chloropyrimidine (17s). This compound was synthesized according to general procedure A starting from **2** and 5-bromo-2,4-dichloropyrimidine (**16s**) (0.30 g, 1.32 mmol) to afford the title material as a white solid (0.17 g, 39%). ¹H NMR (400 MHz, CDCl₃): δ 8.74 (s, 1H), 8.73 (s, 1H), 7.93–7.82 (m, 2H), 7.51–7.36 (m, 2H).

General Procedure D: S_NA, Using TFA/TFE. 4-(((4-(Benzofuran-2-yl)pyrimidin-2-yl)amino)phenyl)(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**18a**). To a stirring suspension of **17a** (0.03 g, 1 equiv, 0.12 mmol) and **15** (0.04 g, 1.20 equiv, 0.15 mmol) in 2,2,2-trifluoroethanol (1.15 mL) was added TFA (0.04 g, 0.02 mL, 2.52 equiv, 0.31 mmol) slowly under an argon atmosphere. The vial was then sealed and heated to 140 °C under microwave conditions for 1 h. The solvent was removed in vacuo. The crude product was purified by

silica gel chromatography using a system of DCM/MeOH 100:0 to 80:20 and then re-purified by prep HPLC to afford the trifluoroacetate salt of the title material as a yellow solid (0.02 g, 30%). ¹H NMR (400 MHz, MeOD): δ 8.58 (d, *J* = 5.1 Hz, 1H), 7.99–7.88 (m, 2H), 7.74 (d, *J* = 7.8 Hz, 1H), 7.71 (s, 1H), 7.61 (d, *J* = 8.3 Hz, 1H), 7.49–7.41 (m, 3H), 7.38 (d, *J* = 5.1 Hz, 1H), 7.32 (t, *J* = 7.6 Hz, 1H), 4.81–3.90 (m, 2H), 3.75–3.60 (m, 2H), 3.55–3.40 (m, 1H), 3.26–2.80 (m, 4H), 2.32–2.13 (m, 4H), 2.10–1.94 (m, 2H), 1.76–1.57 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 174.12, 162.71, 161.66, 158.84, 158.39, 156.03, 145.37, 130.85, 130.53, 130.12, 128.92, 126.05, 124.60, 120.87, 113.84, 110.37, 109.98, 64.46, 54.16, 31.76, 25.12. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₈H₃₀N₅O₂, 468.2394; found, 468.23935.

4-(((4-(Naphthalen-2-yl)pyrimidin-2-yl)amino)phenyl)(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**18b**). This compound was synthesized according to general procedure D from **15** and **17b** (0.05 g, 0.21 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.02 g, 15%). ¹H NMR (400 MHz, MeOD): δ 8.71 (s, 1H), 8.55 (d, *J* = 5.3 Hz, 1H), 8.28 (dd, *J* = 8.7, 1.8 Hz, 1H), 8.07–7.90 (m, 5H), 7.63–7.54 (m, 3H), 7.52–7.45 (m, 2H), 4.80–3.84 (m, 2H), 3.75–3.63 (m, 2H), 3.54–3.40 (m, 1H), 3.24–2.73 (m, 4H), 2.34–2.11 (m, 4H), 2.13–1.92 (m, 2H), 1.81–1.59 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.89, 166.27, 161.67, 159.84, 144.31, 136.17, 135.64, 134.71, 130.05, 129.60, 129.26, 128.78, 128.74, 128.55, 128.50, 127.72, 125.05, 119.68, 110.03, 63.20, 52.90, 30.06, 23.85. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₃₀H₃₂N₅O, 478.2601; found, 478.2598.

4-(Pyrrolidin-1-yl)piperidin-1-yl(4-((4-(thiophen-2-yl)pyrimidin-2-yl)amino)phenyl)methanone (**18c**). This compound was synthesized according to general procedure D from **15** and **17c** (0.04 g, 0.20 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (yield not determined). ¹H NMR (400 MHz, MeOD): δ 8.41 (d, *J* = 5.5 Hz, 1H), 7.94 (dd, *J* = 3.8, 1.1 Hz, 1H), 7.92–7.86 (m, 2H), 7.72 (d, *J* = 4.8 Hz, 1H), 7.54–7.42 (m, 2H), 7.32 (d, *J* = 5.5 Hz, 1H), 7.22 (dd, *J* = 5.0, 3.8 Hz, 1H), 4.80–3.89 (m, 2H), 3.76–3.59 (m, 2H), 3.54–3.38 (m, 1H), 3.26–2.74 (m, 4H), 2.30–2.13 (m, 4H), 2.12–1.89 (m, 2H), 1.80–1.56 (m, 2H). ¹³C NMR (101 MHz, MeOD): δ 172.71, 162.06, 159.99, 157.93, 143.71, 143.50, 132.07, 129.86, 129.72, 129.40, 129.23, 120.11, 107.94, 63.18, 52.88, 23.85. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₄H₂₈N₅OS, 434.2009; found, 434.2008.

4-(Pyrrolidin-1-yl)piperidin-1-yl(4-((4-(thiazol-2-yl)pyrimidin-2-yl)amino)phenyl)methanone (**18d**). This compound was synthesized according to general procedure D from **15** and **17d** (0.05 g, 0.18 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.01 g, 14%). ¹H NMR (850 MHz, MeOD): δ 8.62 (d, *J* = 5.0 Hz, 1H), 8.03 (d, *J* = 3.1 Hz, 1H), 7.95–7.92 (m, 2H), 7.83 (d, *J* = 3.1 Hz, 1H), 7.57 (d, *J* = 5.0 Hz, 1H), 7.47–7.43 (m, 2H), 4.82–3.84 (m, 2H), 3.76–3.60 (m, 2H), 3.45 (tt, *J* = 11.8, 4.1 Hz, 1H), 3.28–2.74 (m, 4H), 2.38–2.11 (m, 4H), 2.10–1.93 (m, 2H), 1.82–1.53 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.78, 168.93, 161.39, 160.86, 159.34, 145.79, 143.86, 129.23, 129.10, 124.62, 119.76, 108.22, 63.20, 52.91, 30.09, 23.85. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₃H₂₇N₅OS, 435.1962; found, 435.1950.

4-(((4-(Furan-2-yl)pyrimidin-2-yl)amino)phenyl)(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**18e**). This compound was synthesized according to general procedure D from **15** and **17e** (0.04 g, 0.19 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.05 g, 64%). ¹H NMR (400 MHz, MeOD): δ 8.47 (d, *J* = 5.3 Hz, 1H), 7.96–7.88 (m, 2H), 7.77 (dd, *J* = 1.8, 0.8 Hz, 1H), 7.51–7.41 (m, 2H), 7.35 (dd, *J* = 3.6, 0.8 Hz, 1H), 7.20 (d, *J* = 5.4 Hz, 1H), 6.67 (dd, *J* = 3.5, 1.8 Hz, 1H), 4.80–3.86 (m, 2H), 3.80–3.60 (m, 2H), 3.51–3.39 (m, 1H), 3.27–2.72 (m, 4H), 2.33–2.10 (m, 4H), 2.08–1.90 (m, 2H), 1.78–1.49 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.86, 161.35, 159.85, 157.46, 153.38, 146.67, 144.19, 129.22, 128.72, 119.53, 113.46, 113.33, 107.64, 63.20, 52.89, 29.94, 23.85. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₄H₂₈N₅O₂, 418.2238; found, 418.2235.

4-(((4-(1-Methyl-1H-pyrazol-4-yl)pyrimidin-2-yl)amino)phenyl)(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**18f**). To a stirred

suspension of **17f** (0.04 g, 1 equiv, 0.21 mmol) and **15** (0.06 g, 1.1 equiv, 0.23 mmol) in 2,2,2-trifluoroethanol (1.75 mL) was added TFA (0.06 g, 0.04 mL, 2.5 equiv, 0.52 mmol) slowly under argon pressure. The vial was sealed and heated to 140 °C under microwave conditions for 1 h. The solvent was removed in vacuo. The crude compound was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 80:20. Afterward, the resulting product was re-purified using reversed-phase chromatography using a gradient of water (0.5% TFA) and MeOH to afford the trifluoroacetate salt of the title material as a yellow solid (0.02 g, 20%). ¹H NMR (400 MHz, MeOD): δ 8.39–8.28 (m, 2H), 8.14 (d, *J* = 0.7 Hz, 1H), 7.90–7.79 (m, 2H), 7.52–7.43 (m, 2H), 7.17 (d, *J* = 5.7 Hz, 1H), 4.82–4.10 (m, 2H), 3.98 (s, 3H), 3.77–3.60 (m, 2H), 3.45 (tt, *J* = 11.8, 3.9 Hz, 1H), 3.28–2.79 (m, 4H), 2.33–2.11 (m, 4H), 2.10–1.97 (m, 2H), 1.75–1.59 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.88, 161.41, 161.13, 159.08, 144.26, 139.55, 132.38, 129.25, 128.65, 123.10, 119.57, 109.18, 63.20, 52.89, 39.29, 29.96, 23.85. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₄H₃₀N₇O, 432.2506; found, 432.2504.

4-((4-Phenylpyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (18g). This compound was synthesized according to general procedure D from **15** and **17g** (0.06 g, 0.31 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.022 g, 16%). ¹H NMR (850 MHz, MeOD): δ 8.51 (d, *J* = 5.3 Hz, 1H), 8.20–8.12 (m, 2H), 7.96–7.90 (m, 2H), 7.56–7.50 (m, 3H), 7.48–7.44 (m, 2H), 7.40 (d, *J* = 5.3 Hz, 1H), 4.85–3.88 (m, 2H), 3.74–3.59 (m, 2H), 3.45 (tt, *J* = 11.8, 4.0 Hz, 1H), 3.26–2.79 (m, 4H), 2.33–2.14 (m, 4H), 2.11–1.90 (m, 2H), 1.73–1.57 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.80, 166.87, 161.10, 159.06, 143.95, 138.16, 132.25, 129.97, 129.26, 129.07, 128.34, 119.91, 109.72, 63.20, 52.90, 30.06, 23.85. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₆H₃₀N₅O, 428.2445; found, 428.2431.

4-((4-(3,4-Dichlorophenyl)pyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (18h). This compound was synthesized according to general procedure D from **15** and **17h** (0.06 g, 0.20 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.025 g, 26%). ¹H NMR (850 MHz, MeOD): δ 8.55 (d, *J* = 5.2 Hz, 1H), 8.41–8.34 (m, 1H), 8.12–8.03 (m, 1H), 7.94–7.86 (m, 2H), 7.69 (d, *J* = 8.3 Hz, 1H), 7.47–7.43 (m, 2H), 7.39 (dd, *J* = 5.2, 1.2 Hz, 1H), 4.81–3.94 (m, 2H), 3.73–3.59 (m, 2H), 3.45 (tt, *J* = 11.8, 4.1 Hz, 1H), 3.27–2.84 (m, 4H), 2.38–2.12 (m, 4H), 2.10–1.94 (m, 2H), 1.81–1.55 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.80, 163.76, 161.58, 160.30, 144.01, 138.70, 135.90, 134.13, 132.11, 130.13, 129.22, 129.01, 127.79, 119.84, 109.60, 63.20, 52.91, 30.15, 23.85. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₆H₂₈Cl₂N₅O, 496.1665; found, 496.1656.

4-((4-(5-Methylthiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (18i). This compound was synthesized according to general procedure D from **15** and **17i** (0.04 g, 0.19 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.02 g, 24%). ¹H NMR (400 MHz, MeOD): δ 8.35 (d, *J* = 5.6 Hz, 1H), 7.95–7.86 (m, 2H), 7.76 (d, *J* = 3.8 Hz, 1H), 7.50–7.43 (m, 2H), 7.25 (d, *J* = 5.6 Hz, 1H), 6.96–6.84 (m, 1H), 4.79–3.83 (m, 2H), 3.74–3.62 (m, 2H), 3.52–3.39 (m, 1H), 3.24–2.80 (m, 4H), 2.56 (s, 3H), 2.34–2.09 (m, 4H), 2.08–1.93 (m, 2H), 1.78–1.48 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.88, 161.32, 161.15, 159.15, 146.66, 144.17, 141.52, 129.45, 129.20, 128.68, 128.14, 119.55, 107.57, 63.20, 52.88, 29.95, 23.85, 15.64. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₅H₃₀N₅OS, 448.2166; found, 448.2163.

4-((4-(5-Chlorothiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (18j). This compound was synthesized according to general procedure D from **15** and **17j** (0.04 g, 0.17 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.02 g, 27%). ¹H NMR (400 MHz, MeOD): δ 8.44 (d, *J* = 5.3 Hz, 1H), 7.93–7.85 (m, 2H), 7.73 (d, *J* = 4.0 Hz, 1H), 7.50–7.41 (m, 2H), 7.24 (d, *J* = 5.3 Hz, 1H), 7.09 (d, *J* = 4.0 Hz, 1H), 4.78–3.90 (m, 2H), 3.76–3.59 (m, 2H), 3.45 (tt, *J* = 11.8, 4.0 Hz, 1H), 3.25–2.76 (m, 4H), 2.33–2.11 (m, 4H), 2.10–1.96 (m, 2H), 1.81–1.59 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 174.09, 162.47, 161.43, 161.10, 145.23, 144.29, 136.83, 130.49,

130.46, 130.18, 129.87, 120.95, 108.54, 64.45, 54.16, 31.18, 25.11. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₄H₂₇ClN₅OS, 468.1619; found, 468.1618.

4-((4-(5-Phenylthiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (18k). This compound was synthesized according to general procedure D from **15** and **17k** (0.05 g, 0.18 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.04 g, 43%). ¹H NMR (400 MHz, MeOD): δ 8.41 (d, *J* = 5.4 Hz, 1H), 7.97–7.86 (m, 3H), 7.78–7.66 (m, 2H), 7.51–7.42 (m, 5H), 7.41–7.33 (m, 1H), 7.30 (d, *J* = 5.5 Hz, 1H), 4.83–3.83 (m, 2H), 3.78–3.59 (m, 2H), 3.53–3.38 (m, 1H), 3.26–2.81 (m, 4H), 2.33–2.13 (m, 4H), 2.09–1.96 (m, 2H), 1.80–1.55 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 174.15, 162.50, 162.24, 160.71, 151.12, 145.39, 144.24, 136.51, 131.47, 130.79, 130.49, 130.04, 128.20, 126.95, 120.91, 109.05, 64.46, 54.15, 31.48, 25.18. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₃₀H₃₂N₅OS, 510.2322; found, 510.2324.

4-((4-(5-(Hydroxymethyl)thiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (18l). **(5-(2-Chloropyrimidin-4-yl)thiophen-2-yl)methanone (17l)** (0.07 g, 1 equiv, 0.31 mmol), **15** (0.10 g, 1.20 equiv, 0.37 mmol), and Cs₂CO₃ (0.30 g, 3 equiv, 0.93 mmol) were added to 1,4-dioxane (4.00 mL). The resulting suspension was degassed by bubbling argon gas through the solvent for 5 min. Afterward, BINAP (0.06 g, 0.30 equiv, 0.09 mmol) and Pd(OAc)₂ (0.01 g, 0.14 equiv, 0.05 mmol) were added to the reaction mixture, which was heated to 90 °C and stirred for 3 h. The mixture was cooled to room temperature and filtered. The filtrate was concentrated in vacuo, and the crude product was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 80:20. The isolated product was re-purified using prep HPLC to afford the trifluoroacetate salt of the title material as a yellow solid (0.03 g, 19%). ¹H NMR (400 MHz, MeOD): δ 8.39 (d, *J* = 5.5 Hz, 1H), 7.94–7.88 (m, 2H), 7.81 (d, *J* = 3.8 Hz, 1H), 7.50–7.42 (m, 2H), 7.28 (d, *J* = 5.5 Hz, 1H), 7.12–7.04 (m, 1H), 4.81 (s, 2H), 4.75–3.76 (m, 2H), 3.76–3.58 (m, 2H), 3.53–3.39 (m, 1H), 3.25–2.77 (m, 4H), 2.33–2.13 (m, 4H), 2.10–1.95 (m, 2H), 1.78–1.53 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.88, 161.22, 159.40, 151.67, 144.15, 143.17, 129.21, 128.92, 128.72, 126.97, 119.61, 107.74, 63.20, 60.35, 52.90, 30.22, 23.85. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₅H₃₀N₅O₂S, 464.2115; found, 464.2102.

4-(Pyrrolidin-1-yl)piperidin-1-yl(4-((4-(5-((2,2,2-trifluoroethoxy)methyl)thiophen-2-yl)pyrimidin-2-yl)amino)phenyl)methanone (18m). This compound was synthesized according to general procedure D from **15** and **17l** (0.04 g, 0.18 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.02 g, 19%). ¹H NMR (400 MHz, MeOD): δ 8.43 (d, *J* = 5.3 Hz, 1H), 7.95–7.88 (m, 2H), 7.80 (d, *J* = 3.8 Hz, 1H), 7.49–7.40 (m, 2H), 7.28 (d, *J* = 5.3 Hz, 1H), 7.19–7.13 (m, 1H), 4.89 (s, 2H), 4.82–4.10 (m, 2H), 4.00 (q, *J* = 8.9 Hz, 2H), 3.73–3.61 (m, 2H), 3.51–3.40 (m, 1H), 3.27–2.89 (m, 4H), 2.29–2.12 (m, 4H), 2.09–1.96 (m, 2H), 1.75–1.56 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.87, 161.28, 160.90, 159.65, 145.58, 144.88, 144.12, 129.48, 129.21, 128.75, 128.72, 125.69, 119.60, 107.86, 69.54, 67.86 (q, *J* = 34.7 Hz), 63.20, 52.90, 29.89, 23.84. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₇H₃₁F₃N₅O₂S, 546.2145 [M + H]⁺; found, 546.2141.

4-((4-(Benzo[b]thiophen-2-yl)-5-methylpyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (18n). This compound was synthesized according to general procedure D from **15** and **17n** (0.05 g, 0.19 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.01 g, 8%). ¹H NMR (400 MHz, MeOD): δ 8.38 (s, 1H), 8.05 (s, 1H), 7.99–7.87 (m, 4H), 7.49–7.44 (m, 2H), 7.44–7.37 (m, 2H), 4.82–3.89 (m, 2H), 3.73–3.61 (m, 2H), 3.50–3.40 (m, 1H), 3.27–2.91 (m, 4H), 2.57 (s, 3H), 2.33–2.10 (m, 4H), 2.08–1.92 (m, 2H), 1.77–1.58 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.96, 161.96, 159.45, 158.67, 145.16, 144.35, 142.07, 141.83, 129.29, 128.40, 128.29, 127.24, 126.08, 125.74, 123.08, 119.43, 119.23, 63.21, 52.90, 30.01, 23.85, 17.97. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₉H₃₂N₅OS, 498.2322; found, 498.23193.

4-((4-(Benzo[b]thiophen-2-yl)-5-cyclopropylpyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**18o**). To a stirred suspension of 4-(benzo[b]thiophen-2-yl)-2-chloro-5-cyclopropylpyrimidine (**17o**) (0.035 g, 1 equiv, 0.12 mmol) and **15** (0.04 g, 1.2 equiv, 0.15 mmol) in 2,2,2-trifluoroethanol (1.25 mL) was added TFA (0.04 g, 0.02 mL, 2.5 equiv, 0.31 mmol) slowly under argon pressure. The vial was sealed and heated to 140 °C under microwave conditions for 1 h. The solvent was removed in vacuo. The crude product was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 93:7 to afford the title material as a yellow solid (0.01 g, 19%). ¹H NMR (400 MHz, MeOD): δ 8.42 (s, 1H), 8.38 (s, 1H), 7.98–7.83 (m, 4H), 7.47–7.43 (m, 2H), 7.43–7.33 (m, 2H), 4.75–3.71 (m, 2H), 3.58–3.31 (m, 3H), 3.22–2.77 (m, 4H), 2.34–1.98 (m, 7H), 1.74–1.56 (m, 2H), 1.22–1.06 (m, 2H), 0.86–0.64 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.92, 160.63, 160.35, 159.21, 144.67, 144.25, 142.10, 142.04, 129.37, 129.28, 128.55, 127.24, 126.12, 125.70, 124.21, 123.11, 119.32, 63.22, 52.88, 30.16, 23.87, 13.14, 7.87. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₃₁H₃₄N₅OS, 524.2479; found, 524.2477.

4-((4-(Benzo[b]thiophen-2-yl)-5-methoxy-pyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**18p**). 4-(Benzo[b]thiophen-2-yl)-2-chloro-5-methoxy-pyrimidine (**17p**) (0.060 g, 1 equiv, 0.22 mmol), **15** (0.065 g, 1.1 equiv, 0.24 mmol), and Cs₂CO₃ (0.21 g, 3 equiv, 0.65 mmol) were added to 1,4-dioxane (3.00 mL). The resulting suspension was degassed by bubbling argon gas for 5 min. Afterward, BINAP (0.041 g, 0.30 equiv, 0.066 mmol) and Pd(OAc)₂ (0.007 g, 0.15 equiv, 0.03 mmol) were added to the reaction mixture, which was heated to 90 °C and stirred for 3 h. The mixture was cooled to room temperature and then filtered. The filtrate was concentrated in vacuo, and the crude product was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 80:20. The isolated product was re-purified using prep HPLC to afford the trifluoroacetate salt of the title material as a yellow solid (0.01 g, 9%). ¹H NMR (400 MHz, MeOD): δ 8.42 (s, 2H), 7.96–7.82 (m, 4H), 7.49–7.42 (m, 2H), 7.44–7.35 (m, 2H), 4.80–4.16 (m, 2H), 4.11 (s, 3H), 3.75–3.60 (m, 2H), 3.53–3.39 (m, 1H), 3.26–2.67 (m, 4H), 2.36–2.11 (m, 4H), 2.11–1.91 (m, 2H), 1.81–1.55 (m, 2H). ¹³C NMR (151 MHz, MeOD): δ 173.05, 155.39, 149.11, 146.29, 144.81, 144.46, 142.11, 141.81, 130.00, 129.38, 127.86, 127.20, 126.04, 125.72, 123.13, 118.54, 63.24, 57.42, 52.92, 30.03, 23.85. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₉H₃₂N₅O₂S, 514.2271; found, 514.2269.

4-((4-(Benzo[b]thiophen-2-yl)-5-fluoropyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**18q**). This compound was synthesized according to general procedure D from **15** and **17p** (0.04 g, 0.13 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.007 g, 10%). ¹H NMR (400 MHz, MeOD): δ 8.45 (d, *J* = 3.4 Hz, 1H), 8.19 (dd, *J* = 1.6, 0.8 Hz, 1H), 7.96–7.84 (m, 4H), 7.50–7.37 (m, 4H), 4.78–3.80 (m, 2H), 3.77–3.60 (m, 2H), 3.52–3.39 (m, 1H), 3.26–2.75 (m, 4H), 2.33–2.11 (m, 4H), 2.10–1.92 (m, 2H), 1.79–1.55 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.86, 157.53 (d, *J*_{C–F} = 2.7 Hz), 150.92 (d, *J*_{C–F} = 2.56 Hz), 147.99 (d, *J*_{C–F} = 2.5 Hz), 147.86 (d, *J*_{C–F} = 10 Hz), 144.14, 142.12, 141.90, 139.41 (d, *J*_{C–F} = 7.2 Hz), 129.79 (d, *J*_{C–F} = 14 Hz), 129.29, 128.74, 127.72, 126.29, 126.05, 123.34, 119.12, 63.21, 52.90, 30.01, 23.86. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₈H₂₉N₅OSF, 502.207; found, 502.2069.

4-((4-(Benzo[b]thiophen-2-yl)-5-chloropyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**18r**). This compound was synthesized according to general procedure D from **15** and **17r** (0.04 g, 0.12 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.01 g, 10%). ¹H NMR (400 MHz, MeOD): δ 8.60 (d, *J* = 0.9 Hz, 1H), 8.50 (s, 1H), 7.99–7.79 (m, 4H), 7.53–7.32 (m, 4H), 4.80–3.92 (m, 2H), 3.67 (s, 2H), 3.57–3.41 (m, 1H), 3.28–2.84 (m, 4H), 2.34–2.11 (m, 4H), 2.10–1.91 (m, 2H), 1.83–1.52 (m, 2H). ¹³C NMR (101 MHz, MeOD): δ 171.34, 159.09, 157.56, 154.48, 142.23, 140.87, 140.82, 140.29, 128.71, 127.83, 127.74, 126.36, 125.00, 124.52, 121.69, 118.25, 116.51, 61.76, 51.47, 28.55, 22.43. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₈H₂₉ClN₅OS, 518.1776; found, 518.1775.

4-((4-(Benzo[b]thiophen-2-yl)-5-bromopyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (**18s**). This compound was synthesized according to general procedure D from **15** and **17s** (0.06 g, 0.18 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.008 g, 8%). ¹H NMR (400 MHz, MeOD): δ 8.73 (s, 1H), 8.63 (s, 1H), 8.01–7.88 (m, 4H), 7.54–7.34 (m, 4H), 4.79–3.85 (m, 2H), 3.74–3.59 (m, 2H), 3.53–3.37 (m, 1H), 3.26–2.75 (m, 4H), 2.32–2.11 (m, 4H), 2.09–1.94 (m, 2H), 1.74–1.54 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.77, 163.28, 159.33, 157.31, 143.60, 143.09, 142.45, 141.51, 129.89, 129.26, 129.24, 127.76, 126.41, 125.96, 123.13, 119.78, 105.82, 63.19, 52.90, 29.57, 23.86. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₈H₂₉N₅OSBr, 562.127; found, 562.1268.

4-(Benzo[b]thiophen-2-yl)-2-((4-(pyrrolidin-1-yl)piperidine-1-carbonyl)phenyl)amino)pyrimidine-5-carbonitrile (**18t**). A mixture of **18s** (0.025 g, 1 equiv, 0.037 mmol), Pd(dppf)₂Cl₂ (0.008 g, 0.30 equiv, 0.01 mmol), Zn(CN)₂ (0.026 g, 0.014 mL, 6.0 equiv, 0.22 mmol), and DIPEA (0.019 g, 0.026 mL, 4.0 equiv, 0.15 mmol) in DMF (1.00 mL) was stirred at 170 °C under microwave conditions for 30 min. Water and NaHCO₃ solution were added to the crude reaction mixture, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with water and brine and dried with Na₂SO₄, and then the solvent was removed in vacuo. The crude product was purified by silica gel chromatography using a system of DCM/MeOH 100:0 to 80:20. The isolated product was re-purified by prep HPLC to afford the trifluoroacetate salt of the title material as a yellow solid (0.004 g, 20%). ¹H NMR (850 MHz, MeOD): δ 8.77 (s, 1H), 8.65 (s, 1H), 7.98–7.87 (m, 4H), 7.53–7.50 (m, 2H), 7.49 (ddd, *J* = 8.0, 7.0, 1.2 Hz, 1H), 7.44 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H), 4.84–4.51 (m, 1H), 4.26–3.89 (m, 1H), 3.75–3.63 (m, 2H), 3.48 (tt, *J* = 11.7, 4.0 Hz, 1H), 3.29–2.74 (m, 4H), 2.39–2.12 (m, 4H), 2.08–1.99 (m, 2H), 1.81–1.66 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.43, 164.69, 161.69, 160.93, 142.86, 142.32, 141.58, 141.41, 130.83, 129.47, 129.18, 128.42, 126.71, 126.29, 123.41, 121.06, 118.54, 63.18, 52.87, 30.38, 23.89. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₉H₂₉N₆OS, 509.2118; found, 509.2105.

2-Chloro-5-methyl-4-(5-methylthiophen-2-yl)pyrimidine (**19a**). This compound was synthesized according to general procedure A starting from (5-methylthiophen-2-yl)boronic acid (0.26 g, 1.84 mmol) and 2,4-dichloro-5-methylpyrimidine (**16n**) (0.30 g, 1.84 mmol) to afford the title material as a yellow solid (0.20 g, 50%). ¹H NMR (400 MHz, CDCl₃): δ 8.34 (d, *J* = 0.9 Hz, 1H), 7.59 (d, *J* = 3.8 Hz, 1H), 6.91–6.80 (m, 1H), 2.55 (d, *J* = 1.0 Hz, 3H), 2.49 (s, 3H). LCMS (ESI+) *m/z*: 225 [M + H]⁺.

2,5-Dichloro-4-(5-methylthiophen-2-yl)pyrimidine (**19b**). This compound was synthesized according to general procedure A starting from (5-methylthiophen-2-yl)boronic acid (0.23 g, 1.64 mmol) and 2,4,5-trichloropyrimidine (**16r**) (0.30 g, 1.64 mmol) to afford the title material as a white solid (0.17 g, 44%). ¹H NMR (400 MHz, DMSO): δ 8.85 (s, 1H), 8.18 (d, *J* = 3.9 Hz, 1H), 7.08–6.98 (m, 1H), 2.55 (d, *J* = 0.5 Hz, 3H). LCMS (ESI+) *m/z*: 245 [M + H]⁺.

2-Chloro-4-(5-chlorothiophen-2-yl)-5-methylpyrimidine (**19c**). This compound was synthesized according to general procedure A starting from (5-chlorothiophen-2-yl)boronic acid (0.30 g, 1.84 mmol) and 2,4-dichloro-5-methylpyrimidine (**16n**) (0.30 g, 1.84 mmol) to afford the title material as a white solid (0.22 g, 48%). ¹H NMR (400 MHz, CDCl₃): δ 8.40 (q, *J* = 0.8 Hz, 1H), 7.54 (d, *J* = 4.2 Hz, 1H), 7.01 (d, *J* = 4.1 Hz, 1H), 2.50 (d, *J* = 0.8 Hz, 3H). LCMS (ESI+) *m/z*: 245 [M + H]⁺.

2,5-Dichloro-4-(5-chlorothiophen-2-yl)pyrimidine (**19d**). This compound was synthesized according to general procedure A starting from (5-chlorothiophen-2-yl)boronic acid (0.27 g, 1.64 mmol) and 2,4,5-trichloropyrimidine (**16r**) (0.30 g, 1.64 mmol) to afford the title material as a white solid (0.16 g, 38%). ¹H NMR (400 MHz, dms): δ 8.93 (s, 1H), 8.21 (d, *J* = 4.3 Hz, 1H), 7.37 (d, *J* = 4.3 Hz, 1H). LCMS (ESI+) *m/z*: 265 [M + H]⁺.

N-(3-Morpholinopropyl)-4-nitrobenzamide (**21**). To a stirring solution of 4-nitrobenzoic acid (**20**) (0.30 g, 1.00 equiv, 1.80 mmol) and DIPEA (0.46 g, 0.63 mL, 2.00 equiv, 3.59 mmol) in DMF (5.00 mL) was added HATU (1.37 g, 2.00 equiv, 3.59 mmol), and the

reaction was allowed to stir at room temperature for 15 min. Afterward, 3-morpholinopropan-1-amine (0.29 g, 0.29 mL, 1.10 equiv, 1.98 mmol) was added, and the reaction was heated to 40 °C and stirred for 4 h. Upon completion of the reaction, the mixture was poured onto ice water, and the aqueous layer was extracted with ethyl acetate. The combined organic extracts were washed with water and brine and then dried with Na₂SO₄. After filtration, the solvent was removed in vacuo, and the residue was purified using silica gel chromatography using a system of DCM/MeOH–NH₃ to afford the title material as a yellow solid (0.15 g, 29%). ¹H NMR (400 MHz, DMSO): δ 8.82 (t, *J* = 5.6 Hz, 1H), 8.34–8.28 (m, 2H), 8.11–8.01 (m, 2H), 3.61–3.55 (m, 4H), 3.35–3.28 (m, 2H), 2.47–2.37 (m, 6H), 1.72 (p, *J* = 7.1 Hz, 2H). LCMS (ESI+) *m/z*: 294 [M + H]⁺.

4-Amino-*N*-(3-morpholinopropyl)benzamide (22). To a stirring suspension of **21** (0.150 g, 1 equiv, 0.511 mmol) in EtOH/water (2.70 mL, 3:1) were added iron (0.09 g, 3.00 equiv, 1.53 mmol) and NH₄Cl (0.14 g, 5.01 equiv, 2.56 mmol), and the reaction mixture was heated to reflux and stirred for 4 h. Upon completion, the reaction mixture was filtered while hot, and the filtering agent was rinsed with ethyl acetate. The filtrate was concentrated in vacuo, and the resulting crude product was used as such for the next step without further purification.

4-((5-Methyl-4-(5-methylthiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (23a). This compound was synthesized according to general procedure D from **15** and **19a** (0.10 g, 0.45 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.037 g, 14%). ¹H NMR (400 MHz, MeOD): δ 8.24 (s, 1H), 7.91–7.82 (m, 2H), 7.65 (d, *J* = 3.9 Hz, 1H), 7.48–7.40 (m, 2H), 6.97–6.86 (m, 1H), 4.82–3.91 (m, 2H), 3.71–3.57 (m, 2H), 3.44 (tt, *J* = 11.7, 7.8, 4.1 Hz, 1H), 3.25–2.80 (m, 4H), 2.56 (s, 3H), 2.44 (s, 3H), 2.28–2.11 (m, 4H), 2.07–1.92 (m, 2H), 1.77–1.54 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.78, 160.05, 158.94, 157.86, 147.48, 143.69, 142.31, 132.74, 129.26, 129.03, 128.53, 119.71, 118.13, 63.19, 52.88, 30.00, 23.85, 17.95, 15.49. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₆H₃₂N₅O₅, 462.2322; found, 462.2321.

4-((5-Chloro-4-(5-methylthiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (23b). This compound was synthesized according to general procedure D from **15** and **19b** (0.10 g, 0.41 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.026 g, 11%). ¹H NMR (400 MHz, MeOD): δ 8.42 (s, 1H), 8.14 (d, *J* = 3.9 Hz, 1H), 7.93–7.84 (m, 2H), 7.48–7.42 (m, 2H), 6.97–6.86 (m, 1H), 4.79–3.93 (m, 2H), 3.77–3.58 (m, 2H), 3.54–3.38 (m, 1H), 3.24–2.82 (m, 4H), 2.57 (s, 3H), 2.35–2.13 (m, 4H), 2.10–1.92 (m, 2H), 1.77–1.52 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.81, 160.08, 158.88, 155.95, 147.87, 143.84, 139.69, 133.60, 129.22, 128.96, 128.27, 119.53, 116.70, 63.19, 52.89, 29.91, 23.85, 15.47. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₅H₂₉ClN₅O₅, 482.1776; found, 482.1775.

4-((4-(5-Chlorothiophen-2-yl)-5-methylpyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (23c). This compound was synthesized according to general procedure D from **15** and **19c** (0.10 g, 0.408 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.03 g, 12%). ¹H NMR (400 MHz, MeOD): δ 8.33 (s, 1H), 7.91–7.81 (m, 2H), 7.60 (d, *J* = 4.2 Hz, 1H), 7.47–7.37 (m, 2H), 7.10 (d, *J* = 4.2 Hz, 1H), 4.79–3.98 (m, 2H), 3.74–3.61 (m, 2H), 3.51–3.40 (m, 1H), 3.25–2.92 (m, 4H), 2.45 (s, 3H), 2.30–2.09 (m, 4H), 2.07–1.93 (m, 2H), 1.75–1.59 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.90, 161.94, 159.26, 157.51, 144.41, 144.21, 135.58, 131.08, 129.25, 129.24, 128.54, 119.31, 118.18, 63.21, 52.90, 30.14, 23.85, 17.58. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₅H₂₉ClN₅O₅, 482.1776; found, 472.1775.

4-((5-Chloro-4-(5-chlorothiophen-2-yl)pyrimidin-2-yl)amino)phenyl(4-(pyrrolidin-1-yl)piperidin-1-yl)methanone (23d). This compound was synthesized according to general procedure D from **15** and **19d** (0.10 g, 0.37 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.022 g, 10%). ¹H NMR (400 MHz, MeOD): δ 8.46 (s, 1H), 8.15 (d, *J* = 4.2 Hz, 1H), 7.93–7.74 (m, 2H), 7.53–7.33 (m, 2H), 7.12 (d, *J* = 4.2 Hz, 1H), 4.79–3.83 (m, 2H), 3.75–3.59 (m, 2H), 3.45 (tt, *J* = 11.7, 4.0 Hz, 1H), 3.26–2.82

(m, 4H), 2.34–2.11 (m, 4H), 2.06–1.93 (m, 2H), 1.74–1.54 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 172.72, 160.61, 158.89, 154.68, 143.56, 141.24, 136.97, 132.88, 129.31, 129.29, 129.21, 119.74, 116.70, 63.18, 52.90, 29.97, 23.86. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₄H₂₆Cl₂N₅O₅, 502.1230; found, 502.1229.

4-((5-Chloro-4-(5-chlorothiophen-2-yl)pyrimidin-2-yl)amino)-*N*-(3-morpholinopropyl)benzamide (23e). This compound was synthesized according to general procedure D from **22** (0.04 g, 0.16 mmol) and **19d** (0.05 g, 0.20 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.006 g, 6%). ¹H NMR (850 MHz, MeOD): δ 8.50 (s, 1H), 8.17 (d, *J* = 4.1 Hz, 1H), 7.89–7.87 (m, 2H), 7.87–7.83 (m, 2H), 7.14 (d, *J* = 4.1 Hz, 1H), 4.13–4.07 (m, 2H), 3.85–3.76 (m, 2H), 3.55–3.49 (m, 4H), 3.26–3.21 (m, 2H), 3.20–3.13 (m, 2H), 2.12–2.05 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 170.72, 160.70, 158.82, 154.66, 145.03, 141.32, 136.99, 132.93, 129.37, 129.33, 127.66, 119.40, 116.88, 65.21, 55.95, 53.17, 37.44, 25.45. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₂H₂₄Cl₂N₅O₂S, 492.1022; found, 492.1021.

4-((5-Chloro-4-(5-methylthiophen-2-yl)pyrimidin-2-yl)amino)-*N*-(3-morpholinopropyl)benzamide (23f). This compound was synthesized according to general procedure D from **22** (0.04 g, 0.16 mmol) and **19b** (0.04 g, 0.14 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.010 g, 10%). ¹H NMR (850 MHz, MeOD): δ 8.44 (s, 1H), 8.16 (d, *J* = 3.8 Hz, 1H), 7.91–7.89 (m, 2H), 7.88–7.86 (m, 2H), 7.05–6.85 (m, 1H), 4.12–4.05 (m, 2H), 3.84–3.77 (m, 2H), 3.55–3.49 (m, 4H), 3.27–3.22 (m, 2H), 3.17 (td, *J* = 12.4, 3.7 Hz, 2H), 2.57 (d, *J* = 1.1 Hz, 3H), 2.11–2.05 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 170.80, 160.15, 158.79, 155.91, 147.88, 145.31, 139.76, 133.63, 129.32, 128.32, 127.33, 119.21, 116.86, 65.21, 55.93, 53.17, 37.43, 25.46, 15.46. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₃H₂₇ClN₅O₂S, 472.1568; found, 472.1567.

4-((4-(5-Chlorothiophen-2-yl)-5-methylpyrimidin-2-yl)amino)-*N*-(3-morpholinopropyl)benzamide (23g). This compound was synthesized according to general procedure D from **22** (0.04 g, 0.16 mmol) and **19c** (0.04 g, 0.14 mmol) to afford the trifluoroacetate salt of the title material as a yellow solid (0.010 g, 15%). ¹H NMR (850 MHz, MeOD): δ 8.35 (d, *J* = 0.8 Hz, 1H), 7.88–7.86 (m, 4H), 7.61 (d, *J* = 4.1 Hz, 1H), 7.11 (d, *J* = 4.1 Hz, 1H), 4.13–4.04 (m, 2H), 3.85–3.77 (m, 2H), 3.55–3.48 (m, 4H), 3.26–3.21 (m, 2H), 3.17 (td, *J* = 12.4, 3.7 Hz, 2H), 2.46 (s, 3H), 2.13–2.01 (m, 2H). ¹³C NMR (214 MHz, MeOD): δ 170.84, 161.84, 159.04, 157.56, 145.61, 144.43, 135.69, 131.20, 129.33, 129.31, 126.97, 119.02, 118.36, 65.21, 55.93, 53.16, 37.41, 25.46, 17.58. HRMS–ESI+ (*m/z*): [M + H]⁺ calcd for C₂₃H₂₇ClN₅O₂S, 472.1568; found, 472.1568.

Biology. KinaseSeeker Assay. Stock solutions (10 mM) of test compounds were serially diluted in DMSO to make assay stocks. Prior to initiating screening or IC₅₀ determination, the test compounds were evaluated for false positive against split luciferase. The test compound was screened against *Pf* kinases at a minimum of eight different concentrations in duplicate. For kinase assays, a 24 mL aliquot of lysate containing Cfluc-kinase and Fos-Nfluc was incubated with either 1 μL of DMSO (for no-inhibitor control) or compound solution in DMSO for 2 h in the presence of a kinase-specific probe. Luciferin assay reagent (80 μL) was added to each solution, and luminescence was immediately measured on a luminometer. The % inhibition was calculated using the following equation: % inhibition = [ALU (control) – ALU (sample)]/ALU (control) × 100. For IC₅₀ determinations, each compound was tested at a minimum of eight different concentrations. The % inhibition was plotted against compound concentration, and the IC₅₀ value was determined for each compound using an eight-point curve.

***P. falciparum* Blood Stage Culture and Parasite Load Assays.** *P. falciparum* 3D7 parasites were continuously cultured in vitro in complete medium [10.44 g/L RPMI 1640 (Thermo Fisher Scientific), 25 mM HEPES, pH 7.2 (Thermo Fisher Scientific), 0.37 mM hypoxanthine (Sigma), 24 mM sodium bicarbonate (Sigma), 0.5% (wt/vol) AlbuMAX II (Thermo Fisher Scientific), and 25 μg/mL gentamicin (Sigma)] supplemented with freshly washed human erythrocytes (Gulf Coast Regional Blood Center, Houston, TX) approximately every 48 h. The parasite cultures were maintained at

2–10% parasitemia with 1% hematocrit at 37 °C in a 3% O₂, 5% CO₂, and 92% N₂ atmosphere. Highly synchronized cultures were generated by treatment with 25 volumes of 5% (wt/vol) D-sorbitol (Sigma) at 37 °C for 10 min during the early ring stage. Prior to the assays, *P. falciparum* 3D7 parasites were synchronized as described above and adjusted to 2% parasitemia and 2% hematocrit. Compounds were initially assayed for parasite inhibition at 1 μM. Dose–response curves were generated for select compounds by dispensing 100 μL of the culture into each well of a 96-well black plate (Corning), followed by administration of compounds (0–5 μM) in triplicate (HP D300 Digital Dispenser). Quinacrine at 500 nM was employed as the positive control and 0.5% DMSO as the negative control. Plates were incubated at 37 °C in a 3% O₂, 5% CO₂, and 92% N₂ atmosphere after drug administration. At 34 h post-reinvasion (i.e., 72 h after drug administration), 40 μL of lysis solution [20 mM Tris–HCl, pH 7.5 (Fisher Chemical), 5 mM EDTA dipotassium salt dihydrate (Fisher Chemical), 0.16% (wt/vol) saponin (Sigma), and 1.6% (vol/vol) Triton X-100 (Fisher Chemical)] containing fresh 10x SYBR Green I (Thermo Fisher Scientific) was added to each well and incubated in the dark at room temperature for 24 h. The fluorescent signals were measured at 535 nm with excitation at 485 nm using an EnVision 2105 multimode plate reader (PerkinElmer). Data was normalized to the negative and positive controls to obtain the relative percent parasite load. EC₅₀ values were determined by fitting data to a standard dose–response equation (GraphPad Prism). The Z-factor ranged from 0.5 to 0.9.

HepG2 CellTiter-Glo Cytotoxicity Assay. HepG2 cells were maintained in Dulbecco's modified Eagle's medium (DMEM, Gibco) supplemented with 10% FBS, 1% NEAA, and 1% L-glutamine. No antibiotics were used. Cells were plated at 4000 cells/well in a 384-well plate (Costar) and incubated overnight (37 °C, 5% CO₂) before adding compounds. Compounds were added in quadruplicate and incubated for 48 h. The DMSO percentage was constant across all concentrations of compound. The cell viability was measured using CellTiter-Glo2 (Promega), and the luminescence signal was read on a GloMax plate reader (Promega).

***P. berghei* Liver Stage Assays.** HepG2 cells were maintained in DMEM with L-glutamine (Gibco) supplemented with 10% heat-inactivated fetal bovine serum (HI-FBS) (v/v) (Sigma-Aldrich) and 1% antibiotic-antimycotic (Thermo Fisher Scientific) in a standard tissue culture incubator (37 °C, 5% CO₂). *P. berghei* ANKA sporozoites used for liver stage experiments were isolated from freshly dissected salivary glands of infected mosquitoes (University of Georgia Sporocore). Dose–response curves were generated for select compounds by assessing the *P. berghei* parasite load in hepatocytes as previously described (PMID: 22586124). Briefly, HepG2 (8,000 cells/well) were seeded into 384-well white microplates (Corning). After 24 h, compounds (0–100 μM) were added (HP D300 Digital Dispenser) before infection with *P. berghei* ANKA sporozoites (4000 spz/well). DMSO (1% v/v) was added as the negative control. All samples were evaluated in triplicate and had a final DMSO concentration of 1%. After 44 h post-infection, the HepG2 cell viability and parasite load were assessed using CellTiter-Fluor (Promega) and Bright-Glo (Promega) reagents, respectively, according to manufacturer's protocols. The relative fluorescence and luminescence signal was measured using an EnVision plate reader (PerkinElmer). The signal intensity of each well was normalized to the negative control (1% DMSO) to assess the relative viability. Dose–response analysis was performed with GraphPad Prism.

Molecular Modeling. The sequences of the target proteins, PfPK6 and PfGSK-3, were obtained from Uniprot (accession numbers: AF091845 and O77344, respectively). The homology models were generated using Modeller v10.1., where five different templates were used in building the target models of each protein. This technique helps in attaining the desired flexibility of the ATP-binding site of the target protein while, at the same time, providing a high degree of confidence about the sequence alignment. The X-ray crystal structures of the templates with their co-crystallized ligands were downloaded from RCSB in PDB format (sequence identity with the target proteins and resolution of each template are illustrated in Table S5, Supporting

Information). The sequences of the target proteins were aligned with the sequences of the related templates using Clustal Omega online tool and, when needed, the adjustment was done by BioEdit. Modeller software was then used to build seven loop-refined models for each target protein, and the model with the lowest PDF violation was chosen for subsequent processing.⁴³ The addition of hydrogen atoms and energy minimization of the crude models were performed using the Molecular Operating Environment (MOE) software, v2019. The minimization step was carried out to a gradient of 0.001 in three sequential steps, where the positional restraints on heavy atoms, backbone, and the entire protein were gradually reduced in each step, respectively. The final homology models were evaluated by different tools. The backbone RMSD of the model was measured relative to that of its template crystal structure individually, and fortunately, the deviation in all cases was less than 0.5 Å, suggesting an appropriate preservation of the template's structural information. In addition, the overall stereochemical quality of the obtained models was also assessed using Procheck software. The analysis of the Ramachandran plot revealed that over 90% of the residues were found in the favorable regions, while more than 8% of the residues lie in the additional allowed region, with no steric clashes, distorted geometry, or outliers. The minimized proteins were then subjected to further refinement using molecular dynamics simulations to ensure better quality of the generated models. Initially, each protein was solvated in a rectangular TIP3P water box, with 10 Å edge distance, using CHARMM-GUI online tool. The protein–water system was then equilibrated in GROMACS software 2021 for 500 ps using the NVT ensemble, followed by an MD production stage for 10 ns.⁴⁴ The final structure after simulation was first re-minimized using MOE (as per the protocol previously described) before being used in the molecular docking study. The Dock module in MOE software was utilized to dock the compounds into the putative binding site of the PfPK6 and PfGSK3 homology models. The active site was predicted using a site finder guided by the amino acids of the hinge region in both proteins, where dummy atoms were added to determine this binding pocket. Docking was performed using the induced fit protocol while other parameters were maintained as default. A maximum of 30 poses were generated for each ligand, and the best poses were selected based on the provided score as well as visual examination. The top scoring pose showing H-bonds with the hinge region was selected, minimized, and used for the analysis. Docking poses were analyzed using the PyMOL Molecular Graphics software 2.5.2 (Schrödinger, LLC., New York, NY, USA).

■ ASSOCIATED CONTENT

SI Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jmedchem.2c00996>.

¹H NMR and ¹³C NMR spectra of the final compounds and HRMS spectra and HPLC traces of compounds submitted for biological screening (PDF)

Kinase profiling data for IKK16, 9g, 18n, 18r, 23d, and 23e (CSV)

Molecular formula strings (XLSX)

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Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

This work was supported in part by the National Institutes of Health under grant no. 1R44AI150237-01 and the National Science Foundation under grant nos. 1644868 and CHE-1726291. We thank the University of North Carolina's Department of Chemistry Mass Spectrometry Core Laboratory, especially Diane Weatherspoon, for their assistance with mass spectrometry analysis. We are grateful for the support by

the Structural Genomics Consortium (SGC), a registered charity (no. 1097737) that receives funds from Bayer AG, Boehringer Ingelheim, Bristol Myers Squibb, Genentech, Genome Canada through Ontario Genomics Institute [OGI-196], EU/EFPIA/OICR/McGill/KTH/Diamond Innovative Medicines Initiative 2 Joint Undertaking [EUBOPEN grant 875510], Janssen, Merck KGaA (aka EMD in Canada and USA), Pfizer, and Takeda.

ABBREVIATIONS USED

EDCI, *N*-ethyl-*N'*-(3-dimethylaminopropyl)carbodiimide; HATU, 1-[bis(dimethylamino)methylene]-1*H*-1,2,3-triazolo-[4,5-*b*]pyridinium 3-oxid hexafluorophosphate; LE, ligand efficiency; SAR, structure–activity relationship; S_N, nucleophilic aromatic substitution; S_N2, nucleophilic substitution (bimolecular); TCAMS, Tres Cantos antimalarial set; TEA, triethylamine; WHO, World Health Organization

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