

Assessment of heavy metals and microbial contamination of *gari* from Liberia

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Abstract

Cassava is a staple mostly eaten in the form of *gari*, after rice in Liberia. The local method of *gari* processing often leads to product contamination, thus, a study was done to assess the heavy metals and microbial contamination of *gari* in eight counties of the country. A total of sixty-one *gari* samples were collected and packaged in an airtight polyethylene bag for analyses, using standard methods. Results depict that the mean of the heavy metals in the *gari* samples is iron (Fe) 43.87 ppm, copper (Cu) 0.94 ppm, zinc (Zn) 5.49 ppm and aluminum (Al) 257.45 ppm. Yellow *gari* had the highest Fe (64.90 ppm), Cu (1.25 ppm) and Zn (7.85 ppm) content, but with the least Al content (87.15 ppm). The Fe content was lower in groundnut-fortified *gari* (42.93 ppm), and the Cu (0.70 ppm) and Zn (3.50 ppm) content were lower in groundnut-moringa-fortified *gari*. The samples and counties have no significant statistical effect ($p > .05$) on the heavy metals composition of the products. No microbial growth was observed in groundnut-fortified and groundnut-moringa-fortified *gari* but with coconut-fortified *gari* having the highest total fungi count of 800 CFU/g. The major fungi identified in the *gari* samples are *Penicillium* and *Aspergillus* spp., but with their counts within the regulated level. Therefore, the *gari* consumed in Liberia are safe except for the high Fe and Al content, which needs to be addressed with the use of unpainted stainless steel materials as food contact surfaces.

KEYWORDS

Cassava, contamination, *gari*, heavy metals, microbial, safety

1 | INTRODUCTION

Cassava is the second most consumed staple food crop in Liberia after rice. It is produced by over 60% of farming households, and an important contributor to the gross domestic product. According to the Food and Agricultural Organization, 534,810 metric tons of cassava was produced in 2014 in Liberia (FAO, 2017). Fresh cassava root contains high moisture content (75%–80%), thus, undergoes rapid spoilage within 48 to 72 hr, if not immediately processed (Ashaye, Adegbulugbe,

& Dawodu, 2005; Oluwole, Olatunji, & Odunfa, 2004; Oyewole & Asagbra, 2003). Processing the roots into various products increases the shelf life, and makes transportation to urban markets less expensive (Taiwo, 2006). Coulibaly, Arinloye, Faye, and Abdoulaye (2014) reported that cassava is processed into different forms in Liberia; fresh roots are processed into *dumbo*, *wet fufu*, starch, *depah*, and *gari* among others. The leaves are also made into soup in the fresh or dried form.

Gari is a dry, crispy, creamy-white and granular product, which is produced by crushing the cassava root into a mash, fermented,

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dewatered, and sieved into grits. The grits are then roasted to make the *gari*. *Gari* is a popular cassava food in Liberia and most parts of West Africa and some countries of Central Africa. It is commonly consumed directly or soaked in cold water with sugar, coconut, roasted peanut, or boiled cowpea as compliments, or as a stiff gel made with hot water and eaten with soup or stew. The acceptance and popularity of *gari* in urban and rural areas of West and Central Africa are attributed to its ability to store well, its convenience and ready-to-eat form (Flach, 1990). Most of the *gari* produced in Liberia is by low-level traditionally techniques due to a lack of modern processing equipment. The traditional cassava grater used is made of a flat iron sheet perforated with nails and fastened onto a wooden board (Coulibaly et al., 2014). The grating is done by rubbing the peeled roots against the sharp perforated surface of the iron sheet which grates the root into the mash. Bruising or injuries of the hands of the processor is common, leading to blood stains in the grated cassava. Stones and or tied woods are used to press out the excess moisture from the grated mash, however, pressers are used in some places. The roasting process is then done, using pans made from iron or earthen pots, and fire woods as a source of energy (Coulibaly et al., 2014). Additionally, partially roasted *gari* is completely dried in the sun without putting into consideration environmental pollution from moving vehicles.

This common traditional method of processing cassava roots in Liberia could result in poor quality products that may be contaminated by foreign matter and disease-causing agents (Bolade, 2016). However, there is insufficient information on the safety of *gari* produced in Liberia. Therefore, this study is aimed at assessing the heavy metals and microbial contamination of *gari* produced and consumed in Liberia.

2 | MATERIALS AND METHODS

2.1 | Collection of *gari* samples in Liberia

Sixty-one *gari* samples (white *gari*-45, yellow *gari*-1, Coconut-fortified *gari*-4, Groundnut-fortified *gari*-10, and Groundnut-moringa-fortified *gari*-1) were collected from the processors and marketers in eight counties; Rivercess, Grand Bassa, Bomi, Margibi, Sinoe, Gbarpolu, Montserrado, and Grand Capemount, for assessment. The yellow *gari* and groundnut-moringa-fortified *gari* were collected from just a point in Montserrado County. Each of the *gari* samples collected is a representative of the sampling frame, thus, the unequal sampling size. Samples were packaged in hermetically sealed polyethylene bags for laboratory analyses. The processing methods for the different types of *gari* are described in Table 1.

2.2 | Analysis of heavy metal contamination

The iron, zinc, copper, and aluminum content of the samples were determined, using the method described by Jones, Benton, and Vernon (1990). The samples were ashed at 550°C. The ash was dissolved in 5 ml water and 15 ml HNO₃/HCl (1:3) for heavy metal determination, using Atomic Absorption Spectrophotometer (Buck 205 model; Back Scientific, USA).

TABLE 1 Processing methods for *gari* from fresh cassava roots in Liberia

Type of <i>gari</i>	Processing method
White <i>gari</i>	Peeling, washing, grating, bagging and dewatering, granulation, and roasting in earthenware pots.
Yellow <i>gari</i>	Same processing steps as above with mixing of palm oil to the granules before roasting
Coconut-fortified <i>gari</i>	Grating and roasting of matured coconut pulp before mixing with white <i>gari</i>
Groundnut-fortified <i>gari</i>	Roasting and milling of groundnuts before mixing with white <i>gari</i>
Groundnut-moringa-fortified <i>gari</i>	Drying of fresh moringa leaves, milling, and mixing with groundnut-fortified <i>gari</i>

2.3 | Analysis of microbial contamination

Analysis of microbial contamination by total plate count of fungi was done following the method described by Amankwah, Barimah, Acheampong, Addai, and Nnaji (2009). Fungal isolates were identified and characterized under a light microscope (Leica Galen III) based on morphological and cultural features as described by Harrigan and McCance (1976).

2.4 | Statistical analysis

Analysis of variance (ANOVA), separation of the mean values (using Duncan's Multiple Range Test at $p < .05$), and frequency distributions were calculated, using Statistical Package for Social Scientists (SPSS) software (version 21.0).

3 | RESULTS AND DISCUSSIONS

3.1 | Heavy metal composition of *gari* products in Liberia

Dix (1981) reported that human exposure to heavy metals causes serious adverse health effects, including reduced growth and development, cancer, organ damage, and in extreme cases—death. Nevertheless, iron (Fe), copper (Cu), and zinc (Zn) are also referred to as trace metals, which are naturally present in foodstuff and confer some nutritional benefits to human, but toxic when consumed in excess (Magomya, Yebpella, Udiba, Amos, & Latayo, 2013). The means of the heavy metal composition of the *gari* samples were 43.87 ppm, 0.94 ppm, 5.49 ppm and 257.45 ppm for Fe, Cu, Zn, and Al, respectively (Table 2).

Although there were no statistically significant differences ($p > 0.05$) in the heavy metal composition of the products (Table 2), yellow *gari* had the highest Fe (64.90 ppm), Cu (1.25 ppm), and Zn (7.85 ppm) content, but the least Al content (87.15 ppm). The Fe content was lower in groundnut-fortified *gari* (42.93 ppm), and the Cu (0.70 ppm) and Zn (3.50 ppm) content were lower in groundnut-moringa-fortified *gari*. Additionally, groundnut-fortified *gari* had the highest Al content (180.52 ppm).

TABLE 2 Heavy metal composition of *gari* products in Liberia

Samples	N	Fe (ppm)	Cu (ppm)	Zn (ppm)	Al (ppm)
White <i>gari</i>	90	45.00 ± 52.86a	0.94 ± 1.00a	5.47 ± 6.49a	136.59 ± 165.13a
Coconut-fortified <i>gari</i>	8	60.75 ± 66.58a	0.93 ± 1.00a	4.64 ± 5.38a	104.16 ± 124.83a
Yellow <i>gari</i>	2	64.90 ± 91.78a	1.25 ± 1.77a	7.85 ± 11.10a	87.15 ± 123.25a
Groundnut- fortified <i>gari</i>	20	42.93 ± 51.14a	0.93 ± 0.99a	6.04 ± 6.86a	180.52 ± 221.25a
Groundnut-moringa- fortified <i>gari</i>	2	44.35 ± 62.72a	0.70 ± 0.99a	3.50 ± 4.95a	118.75 ± 167.94a
Counties					
Montserrado	52	51.89 ± 61.46a	0.92 ± 0.98a	4.92 ± 5.32ab	105.01 ± 179.45b
Bomi	6	25.62 ± 29.31a	0.90 ± 1.01a	4.07 ± 4.67b	1112.85 ± 128.02a
Gbarpolu	6	25.80 ± 29.40a	1.00 ± ± 1.16a	10.88 ± 14.85a	138.68 ± 154.91b
Grand Bassa	20	25.36 ± 29.74a	0.91 ± 0.96a	4.76 ± 5.65ab	145.60 ± 151.80b
Rivercess	4	50.20 ± 58.68a	1.00 ± 1.15a	5.80 ± 7.07ab	141.92 ± 165.13b
Margibi	20	55.65 ± 59.92a	0.88 ± 0.93a	6.10 ± 6.65ab	110.00 ± 143.34b
Grand Cape mount	12	47.29 ± 51.79a	0.87 ± 0.93a	5.49 ± 6.11ab	177.98 ± 274.55b
Sinoe	20	37.36 ± 42.29a	1.10 ± ± 1.17a	5.85 ± 6.21ab	127.53 ± 131.09b
Mean		43.87	0.94	5.49	257.45
P Samples		NS	NS	NS	NS
P Counties		NS	NS	NS	NS
P Samples × Counties		NS	NS	NS	NS

P, phosphorus; Fe, iron; Cu, copper; Zn, zinc; Al, aluminum; NS, not significant.

Means with different letters along the same column are significantly different ($p < .05$).

Considering the Counties, there were no statistically significant differences ($p > .05$) in the heavy metal compositions of the products apart from Zn (Table 2). Products from Margibi had the highest Fe (55.65 ppm) content, Bomi had the highest Al (1,112.85 ppm) and the lowest Zn (4.07 ppm) content and Gbarpolu and Sinoe had the highest Zn (10.88 ppm) and Cu (1.10 ppm) content, respectively. The least Fe (25.36 ppm) and Cu (0.87 ppm) content were found in Grand Bassa and Grand Capemount counties, respectively. Montserrado County has the lowest Al (105.01 ppm).

Metals such as Fe, Cu, and Zn have been observed to be essential components of many alloys, wires, and vehicle tyres and, which are usually released into the roadside environment because of mechanical abrasion and normal wear and tear (Harrison, Laxen, & Wilson, 1981). This implied that yellow *gari* with the highest values of these metals might have been roasted closer to a roadside or a market where heavy vehicular movement exist. However, the values for Zn and Cu in the yellow *gari* were below the recommended maximum limit of 10 mg/100 g and 7.3 mg/100 g respectively, stipulated by the Food and Agricultural Organization (FAO), and World Health Organization (WHO), while the Fe content of the sample was higher than the FAO/WHO standard of 42.5 mg/100 g (FAO/WHO, 2001). Thus, it will be important for the processors in Margibi counties with the highest Fe contained *gari*, to replace the local graters and or roaster made with galvanized or mild steel with stainless steel machines to reduce Fe contamination. Additionally, *gari* roasting should be completed on the roasting pan and not further dried under the sun closer to heavy vehicular movement (Bolade, 2016). Furthermore, the higher Al content in relation to the

lower Fe and Zn content in the *gari* samples may be attributed to contamination caused by the water used during processing or migration from the paints used on the food contact surfaces/processing machines (Nanda, Biswal, Acharya, Rao, & Pujari, 2014; Sato et al., 2014; Stahl et al., 2017). The Al content of the *gari* samples is very high compared to the stipulated provisional tolerable weekly intake of 7 mg/kg body weight reported by the Joint FAO/WHO Expert Committee on Food Additives (JECFA) and the Scientific Committee for Food (SCF) (JECFA, 1989). Thus, food contact surfaces/machines should not be painted with metallic polish to reduce Al contamination, as accumulation of Al has been reported to be potentially cyto- and neurotoxic to humans (Austrian Department of Health, 2014).

3.2 | Microbial identification in *gari* products

The total fungi count (TFC) of the *gari* samples revealed that no microbial growth was observed in groundnut-fortified and groundnut-moringa-fortified *gari*, but with coconut-fortified *gari* having the highest TFC of 800 CFU/g (Table 3). Additionally, *gari* from Bomi and Grand Cape mount counties have no fungi growth, but with *gari* from Rivercess county having the highest TFC (1500 CFU/g) (Table 3). The TFC of the *gari* samples in the present study is lower compared to the result of Obadina, Oyewole, and Odusami (2009) and Olopade, Oranusi, Ajala, and Olorunsola (2014). Obadina et al. (2009) reported 1.79×10^4 CFU/g for the TFC of *gari* collected from markets in Abeokuta Ogun state, while Olopade et al. (2014) reported the TFC of *gari* from different markets in Ota, Ogun state Nigeria to be, from no growth to 3,000 CFU/g. The International Commission on

TABLE 3 Microbial counts and identification in gari products

Samples	Total fungi count (CFU/g)	Organism identified
White gari	525.00 ± 823.46a	<i>Penicillium</i> sp., <i>Macrophomina</i> sp., <i>Aspergillus</i> sp.
Coconut-fortified gari	800.00 ± 663.32a	<i>Penicillium</i> sp., <i>Aspergillus</i> sp.
Yellow gari	342.00 ± 310.55a	<i>Aspergillus flavus</i> , <i>Penicillium</i> sp., <i>Rhizopus</i> sp.
Groundnut-fortified gari	NG	NI
Groundnut-moringa-fortified gari	NG	NI
Counties		
Montserrado	479.31 ± 794.80ab	<i>Penicillium</i> sp., <i>Macrophomina</i> sp.
Bomi	NG	NI
Gbarpolu	166.67 ± 288.68b	<i>Aspergillus flavus</i> , <i>Penicillium</i> sp.
Grand Bassa	980.00 ± 1,137.05ab	<i>Rhizopus</i> sp.
Rivercess	1,500.00 ± 707.11ab	<i>Penicillium</i> sp.
Margibi	866.67 ± 837.02ab	<i>Aspergillus niger</i> , <i>Aspergillus flavus</i> , <i>Penicillium</i> sp.
Grand Cape Mount	NG	NI
Sinoe	208.33 ± 2,429.30a	<i>Aspergillus niger</i> , <i>Aspergillus flavus</i> , <i>Penicillium</i> sp.
Mean	812.99	
P Samples	NS	
P Counties	NS	
P Samples × Counties	NS	

NG, no fungi growth; NI, no fungi identified; NS, not significant.

Means with different letters along the same column are significantly different ($p < .05$).

Microbiological Specifications for Foods (ICMSF) stated that ready to eat foods with counts of $\leq 10^3$ CFU/g are acceptable, counts of 10^4 – 10^5 CFU/g are tolerable while counts $\geq 10^6$ CFU/g are unacceptable (ICMSF, 1996). Thus, since the average TFC value of the gari samples is $< 10^3$ CFU/g, they may be safe for consumption.

Fungi such as *Penicillium* and *Aspergillus* spp. were found in white, yellow, and coconut-fortified gari, with *Macrophomina* sp. detected in only white gari (Table 3). *Rhizopus* spp. was found in gari from Grand Bassa county only. However, there is need to reduce the presence of *Aspergillus* and *Penicillium* spp. in the gari samples by following strictly good manufacturing and hygienic practices as these microorganisms are known to produce harmful mycotoxins (Kabak, Dobson, & Var, 2006; Oranusi, Wesley, & Oguoma, 2013). The products have no statistically significant effect ($p > .05$) on the TFC.

4 | CONCLUSION

The gari produced in Liberia may be safe for consumption since the zinc and copper content were below the recommended maximum limit stipulated by the FAO/WHO. However, there is a need for caution in the use of mild or galvanized steel instead of stainless steel materials as food contact surfaces, as well as painting of processing machine surfaces with metallic polish because of the high iron and

aluminum content in the gari. Although the average total fungi count value of the gari samples is less than the International Commission on Microbiological Specifications for Foods levels, there is need to reduce the presence of *Aspergillus* and *Penicillium* spp. in the gari samples by following strictly good manufacturing/hygienic practices, as these microorganisms are known to produce harmful mycotoxins under favorable conditions.

CONFLICT OF INTEREST

No conflict of interest.

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REFERENCES

- Amankwah, E. A., Barimah, J., Acheampong, R., Addai, L. O., & Nnaji, C. O. (2009). Effect of fermentation and malting on the viscosity of maize-soybean weaning blends. *Pakistan Journal of Nutrition*, 8, 1671–1675.
- Ashaye, O. A., Adegbulugbe, T. A., & Dawodu, O. J. (2005). Evaluation of the processing technologies of cassava chips and flour in Oyo and Ogun States of Nigeria. *World Journal of Agricultural Sciences*, 1(1), 56–58.
- Austrian Department of Health (2014). Aluminium-toxikologie und gesundheitliche Aspekte körpurnaher Anwendungen. Herausgeber, Medieninhaber und Hersteller: Bundesministerium für Gesundheit, Sektion IIRadetzkystraße 2, 1031 Wien, Österreich. ISBN 978-3-902611-79-6.
- Bolade, M. K. (2016). An investigation into the level of metallic pollutants in roadside-sundried food products from selected areas of Ondo and Osun states Nigeria. *Cogent Food and Agriculture*, 2, 1179161. <https://doi.org/10.1080/23311932.2016.1179161>
- Coulibaly, O., Arinloye, A.D., Faye, M., & Abdoulaye, T. (2014). Regional cassava value chains analysis in West Africa: Case study of Liberia. pp. 30.
- Dix, H. M. (1981). *Environmental pollution atmosphere, land, water and noise*. Chichester: John Wiley and Sons.
- FAO (2017). Food and Agricultural Organization Crop Statistics.
- FAO/WHO. (2001). *Food additive and contaminants*. Joint FAO/WHO Food Standard Program, ALINORM 01/12A, Codex Alimentarius Commission (pp. 1–289). The Hague: FAO/WHO
- Flach M. (1990). Gari processing in North West province of Cameroun. Working Document No. 5, FAO project.
- Harrigan, W. F., & McCance, E. M. (1976). *Laboratory methods in food and dairy microbiology* (p. 31). London U.K: Academic Press.
- Harrison, R. M., Laxen, D. P. H., & Wilson, S. J. (1981). Chemical associations of lead, cadmium, copper, and zinc in street dust and roadside soils. *Environmental Science and Technology*, 15, 1378–1383.
- ICMSF (1996). *International Commission on microbiological specifications for foods. Microorganisms in foods 5: Microbiological specifications of pathogens*. London: Blackie Academic and Professional; pp. 20–35.
- JECFA (1989). Joint FAO/WHO Expert Committee on Food Additives Toxicological evaluation of certain food additives and contaminants. *WHO Food Additives Series*, 24, 113–154.
- Jones, J. R., Benton, J., & Vernon, C. W.. (1990). Sampling, handling and analyzing plant tissue samples. In R. L. Westermann (Ed.), *Soil testing and plant analysis* (pp. 389–428). 3rd edn. Madison WI: Soil Sci. Soc. Am. Inc.
- Kabak, B., Dobson, A. D. W., & Var, I. (2006). Strategies to prevent mycotoxin contamination of food and animal feed: A review. *Critical Reviews in Food Science and Nutrition*, 46, 593–619.
- Magomya, A. M., Yebpella, G. G., Udiba, U. U., Amos, H. S., & Latayo, M. S. (2013). Potassium bromate and heavy metal content of selected bread samples produced in Zaria, Nigeria. *International Journal of Science and Technology*, 2, 232–237.
- Nanda, B. B., Biswal, R. R., Acharya, R., Rao, J. S. B., & Pujari, P. K. (2014). Determination of aluminum contents in selected food samples by instrumental neutron activation analysis. *Journal of Radioanalytical and Nuclear Chemistry*, 302, 1471–1474.
- Obadina, A.O., Oyewole, O.B., & Odusami, A.O. (2009). Microbiological safety and quality assessment of some fermented cassava products *lafun, fufu, gari*. *Scientific Research and Essay*, 4(5), 432–435.
- Olopade, B. K., Oranusi, S., Ajala, R., & Olorunsola, S. J. (2014). Microbiological quality of fermented Cassava (Gari) sold in Ota Ogun State Nigeria. *International Journal of Current Microbiology and Applied Sciences*, 3(3), 888–895.
- Oluwole, O. B., Olatunji, O. O., & Odufa, S. F. (2004). A process technology for conversion of dried cassava chips into *gari*. *Nigeria Food Journal*, 22, 65–73.
- Oranusi, S., Wesley, B., & Oguoma, O. I. (2013). Antifungal properties of lactic acid bacteria isolated from *Ricinus communis*, *Pentaclethra macrophylla* and Yoghurts. *Global Advanced Research Journal of Food Science and Technology*, 21, 01–06.
- Oyewole, O. B., & Asagbra, Y. (2003). Improving traditional cassava processing for nutritional enhancement. In Proceedings of the International Workshop on Food-based approaches for a Healthy Nutrition, Ouagadougou, pp. 23–28.
- Sato, K., Suzuki, I., Kubota, H., Furusho, N., Inoue, T., Yasukouchi, Y., & Akiyama, H. (2014). Estimation of daily aluminum intake in Japan based on food consumption inspection results: Impact of food additives. *Food Science and Nutrition*, 2, 389–397.
- Stahl, T., Falk, S., Rohrbeck, A., Georgii, S., Herzog, C., Wiegand, A., ... Brunn, H. (2017). Migration of aluminum from food contact materials to food—a health risk for consumers? Part I of III: Exposure to aluminum, release of aluminum, tolerable weekly intake (TWI), toxicological effects of aluminum, study design, and methods. *Environmental Sciences Europe*, 29, 19.
- Taiwo, K. A. (2006). Utilization potentials of cassava in Nigeria: The domestic and industrial products. *Food Reviews International*, 22, 29–42.

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